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WALTER WYMAN, Surgeon-General

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M. J. ROSENAU, Director

APRIL, 1907

STUDIES UPON HYPERSUSCEPTIBILITY AND IMMUNITY

By

M. J. ROSENAU
and
JOHN F. ANDERSON



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CONTENTS.

| Introduction | Page. |
|---|-------|
| Part I: | |
| The sensitizing substance | 11 |
| Part II: | |
| The toxic principle | 15 |
| Influence of ferments. | 15 |
| Influence of alkaloids. | 16 |
| Influence of salts | 16 |
| Influence of miscellaneous substances | 17 |
| Influence of formaldehyd. | 17 |
| Influence of calcium chlorid | 18 |
| Influence of freezing. | 21 |
| PART III: | |
| Is the toxic principle specific? | 23 |
| Horse serum versus other proteid substances | 23 |
| Other proteid substances versus horse serum | 24 |
| PART IV: | |
| Other blood serums and other albuminous substances are also toxic | 25 |
| Other blood serums | 25 |
| Other albuminous substances | 25 |
| Part V: | |
| Hypersusceptibility and immunity produced by bacterial proteids | 31 |
| Part VI: | |
| Comparative toxicity of untreated and refined antitoxic serum | 39 |
| Part VII: | |
| Comparative toxicity of different horse serums | 43 |
| Toxicity of "serum antidiphterique" (Pasteur Institute) | 43 |
| Toxicity of normal horse serum (our roan) | 44 |
| PART VIII: | |
| The immunity to hypersusceptibility or "anti-anaphylaxis" | 45 |
| PART IX: | |
| Maternal transmission of hypersusceptibility and immunity | 47 |
| Group A, sensitized female, untreated male | 47 |
| Group B, sensitized female, sensitized male | 49 |
| Group C, immune female, immune male | 50 |
| Group D, immune female, untreated male | 52 |
| Group E, untreated female, sensitized male | 54 |
| Conclusions | 54 |

| | rage. |
|--|----------|
| Part X: | |
| The relations of hypersusceptibility to various influences | 55 55 |
| Aggressines | ~~ |
| Methemaglobin | 56 |
| Oxygen | 56 |
| Diphtheria toxine | 57 |
| Tetanus toxine | 58 |
| The spleen and the thyroid | 58 |
| Part XI: | 0.1 |
| Miscellaneous | |
| Feeding experiments with cooked meat | 61 |
| Feeding experiments with raw beef | 61 |
| Result of cardiac injections | 62 |
| The guinea pig remains susceptible a very long time | 62 |
| The effect of first injections of horse serum into guinea pigs | 64 |
| Part XII: | 0.11 |
| Summary and conclusions | 65 |

STUDIES UPON HYPERSUSCEPTIBILITY AND IMMUNITY.

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We have shown b that horse serum is apparently a bland and harmless substance when injected into a normal guinea pig, but this injection renders the guinea pig susceptible to a subsequent injection of horse serum. At least ten days must elapse between the first and the second injection for this hypersusceptibility to manifest itself.

The present bulletin gives the results of our further work upon this interesting phenomenon. We have endeavored to obtain a deeper insight into the cause and nature of hypersusceptibility and have attempted to localize the phenomenon in certain fluids, cells, or organs of the body.

We foresaw last year that the problem of hypersusceptibility has an important bearing upon the question of immunity and expressed the opinion that "resistance to disease may be largely gained through a process of hypersusceptibility. Whether this increased susceptibility is an essential element or only one stage in the process of resistance to disease, must now engage our attention." We can not escape the

^aManuscript submitted for publication April 27, 1907.

^bRosenau, M. J., and Anderson, John F.: A study of the cause of sudden death following the injection of horse serum. Pub. Health and Mar.-Hosp. Serv., Hyg. Lab. Bull. No. 29, 1906.

^{——:} A new toxic action of horse serum. Journ. med. research, Vol. 15, No. 1 (n. s., Vol. 10, No. 1), July, 1906, pp. 179–208.

Anderson, John F.: I. Maternal transmission of immunity to diphtheria toxine. II. Maternal transmission of immunity to diphtheria toxine and hypersusceptibility to horse serum in the same animal. Pub. Health and Mar.-Hosp. Serv., Hyg. Lab. Bull. No. 30, 1906, and Journ. med. research, Vol. 15, No. 2 (n. s., Vol. 10, No. 2), Sept., 1906, pp. 241–260.

^cRosenau, M. J., and Anderson, John F.: Hypersusceptibility. Journ. Am. Med. Assn., Vol. 42, No. 13, Sept. 29, 1906, pp. 1007–1010.

conviction that this phenomenon of hypersusceptibility has an important bearing on the prevention and cure of certain infectious processes. Our work this year upon the hypersusceptibility produced by the bacterial proteids strengthens this belief, for our results prove that the phenomenon of hypersusceptibility to certain proteid substances extracted from the bacterial cell is followed by a definite immunity against infection by the micro-organism.

Since our studies last year several papers have been published which, in the main, have corroborated our findings.

McClintock and King a gave ten guinea pigs from $\frac{1}{250}$ to 1 c. c. of horse serum by the stomach and thirteen days later 6 c. c. of serum, either subcutaneously or intraperitoneally, without causing symptoms in any of them. They conclude that the sensitizing action of horse serum given by the mouth is not nearly so great as when given subcutaneously or intraperitoneally. This is in confirmation of our reported experiments.

Currie^b has studied the effect of repeated injections of horse serum in persons admitted for treatment in the city of Glasgow Fever and Smallpox Hospital at Belvidere. He concludes that it is apparent from the facts detailed by him that repeated injections of horse serum induce symptoms of supersensitation in man, but it is also apparent that the same facts lend no countenance to the suggestion that the death of persons suffering from diphtheria is to be apprehended as the result of repeated injections of antidiphtheric serum.

Besredka and Steinhardt^c studied with much care certain features of hypersusceptibility to horse serum in guinea pigs; they note that the French serums are much less toxic than those used by Otto in Frankfurt and the serums used by us. Besredka and Steinhardt had a mortality of about 25 per cent when 5 c. c. of serum was given intraperitoneally at the second injection, whereas death was the rule in our experiments under similar conditions. Most of their work was done with doses of 0.05 to 0.25 c. c. given directly into the brain, which either killed or caused grave symptoms in susceptible guinea pigs. Besredka and Steinhardt lay stress upon the production of "anti-anaphylaxis," which we termed "immunity." They found that a single injection of serum given into the peritoneum of a sensitized guinea pig conferred immunity to a subsequent injection of 0.25 c. c. into the brain: in one case the anti-anaphylaxis was present one and a

^a McClintock, Charles T., and King, Walter E.: The oral administration of antitoxins for prevention of diphtheria, tetanus, and possibly sepsis. Journ.infec. diseases, Vol. 3, No. 5, Oct., 1906, pp. 700–720.

^bCurrie, J. R.: On the supersensitation of persons suffering from diphtheria by repeated injections of horse serum. Journ. hyg., Vol. 7, No. 1, Jan., 1907, pp. 35–60.

^c Besredka, A., and Steinhardt, Edna: De l'anaphylaxie et de l'anti-anaphylaxie vis-à-vis du sérum de cheval. Ann. de l'Inst. Pasteur, Vol. 21, No. 2, Feb. 25, 1907, pp. 117−127.

half hours after the injection into the abdominal cavity. They were unable to demonstrate any protective properties in various organs of immune guinea pigs, confirming our work along the same lines.

Nicolle ^a found that guinea pigs were not susceptible to the necrotic action induced by repeated injections of horse serum, as is the case in rabbits; this corresponds with our observations. He also found that daily injections or "spaced" injections, after the method of Arthus, did not induce a high degree of hypersusceptibility in guinea pigs.

Besredka^b questions whether we should not consider this toxic property of horse serum, as well as its antitoxic power. He suggests that a serum, 0.05 c. c. of which when given into the brain will kill or cause grave symptoms in a sensitive guinea pig, should be considered as above the average toxicity and ought to be excluded from use in man.

The work of Otto^c on the "Theobald Smith Phenomenon," and of von Pirquet and Schick dupon "the serum disease" has been previously referred to.

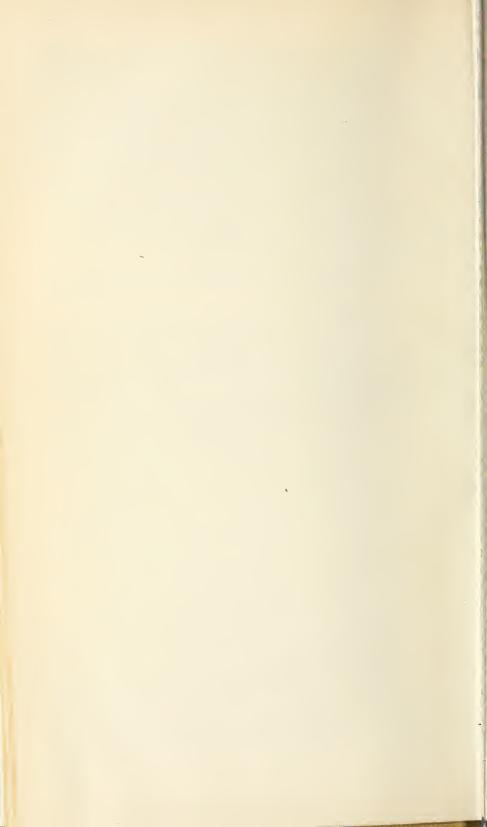
a Nicolle, Maurice: Contribution à l'étude du phénomène d'Arthus. Ann. de l'Inst. Pasteur, Vol. 21, No. 2, Feb. 25, 1907, pp. 128-136.

b Besredka, A.: De la toxicité des sérums thérapeutiques et dumoyen de la doser.
 Comp. rend. soc. biol., Vol. 62, No. 10, Mar. 22, 1907, pp. 477–478.

c Otto, R.: Das Theobald Smithschen Phänomen der Serum-Ueberempfindlichkeit. v. Leuthold-Gedenkschr., Bd. 1.

d von Pirquet, C. Frh., and Schick, B.: Die Serumkrankheit. Leipzig and Wien, 1905, 144 p. 8°.

³⁶⁸¹⁻No. 36-07-2



Part I.

THE SENSITIZING SUBSTANCE.

We ventured the suggestion in our former publication that the substance that sensitizes the guinea pig is the same as that which later poisons it; profound chemical changes perhaps in the central nerve cells, are probably produced by the first injection. Our subsequent work has produced nothing to alter this working hypothesis.

Vaughan a advances the theory that the first injection of the strange proteid is broken up into components, one of which is toxic, but that the animal is not poisoned because this breaking up takes place slowly. The cells, however, learn from this lesson how to break up the complex molecule, so that when more of the strange proteid is introduced at the second injection it is violently rent asunder, quickly liberating large quantities of the toxic principle of the complex molecule.

Vaughan and Wheeler^b have elaborated this explanation by further studies upon egg-white and bacterial proteids split into poisonous and nonpoisonous portions. These authors believe that when egg-white, or the nonpoisonous portion of egg-white, is injected into a fresh animal certain cells of the body are so influenced that they elaborate a new ferment, which, in the form of zymogen, remains in the cell until activated by the second injection, when it is set free and splits up the egg-white in a manner similar to that used by Vaughan in the laboratory. Vaughan and Wheeler believe that the effect induced in the animal is the same as that caused by the poisonous portions of egg-white as they have split it up in the retort.

Currie suggests that the first injection of serum results after an interval in the formation of an antibody. When the second injection of serum is given, after at least ten days from the first, the antibodyproducing substance of the second injection of serum and the antibody

35-60.

a Vaughan, V. C.: Discussion of "Hypersusceptibility," by M. J. Rosenau and J. F. Anderson. Journ. Am. Med. Assn., Vol. 47, No. 13, Sept. 29, 1906, p. 1009.

b Vaughan, Victor C., and Wheeler, May: Effects of egg-white and its split products upon animals. A study of susceptibility and immunity. Abstract of papers to be read at twenty-second annual meeting of Assn. Am. Physicians, Washington, May 7-9, 1907, p. 9.

^c Currie, J. R.: On the supersensitation of persons suffering from diphtheria by repeated injections of horse serum. Journ. Hygiene, Vol. 7, No. 1, Jan., 1907, pp.

produced by the first injection come in contact without delay; their union is rapid; the whole charge of the poisonous substance is quickly set free and the toxic symptoms are sudden and severe.

Besredka and Steinhardt^a had, as a working hypothesis, the following: The sensitized guinea pig which appears in good health has, in spite of its apparent well-being, perhaps a latent lesion of the brain. A second injection of serum, made into the peritoneal cavity twelve days later, is able to awaken this nervous lesion, resulting in grave symptoms or even death.

In view of these theoretical considerations it is important to make further studies upon the sensitizing substance in horse serum and other proteid substances.

The following experiments show that the filtrate from horse serum after precipitation with ammonium sulphate renders guinea pigs sensitive. The filtrate contains most of the serum albumen and very little of the globulins. It is exceedingly weak in antitoxic strength.

G. P. No. 400. Six c. c. antitoxic horse serum (Natl. VIII., 18), intraperitoneally. Marked symptoms.

[Previous treatment: 38 days prior, 5 c. c. filtrate of antitoxic serum, precipitated (NYBH). Subcutaneously.]

This filtrate was kindly furnished us by Dr. W. H. Park from some antidiphtheric serum undergoing the Gibson process of refining.

The following experiments show that formaldehyd does not destroy the sensitizing property of horse serum:

G. P. No. 390W. Six c. c. normal horse (roan) serum, intraperitoneally. Severe symptoms.

[Previous treatment: 47 days prior, 6 c. c. antitoxic horse serum (Natl. XVIII)+1 per cent formalin, 23 hours exposure. Subcutaneously.]

G. P. No. 500W. Six c. c. normal horse (roan) serum, intraperitoneally. Very severe symptoms.

[Previous treatment: 29 days prior, 3 c. c. normal horse (roan) serum+5 per cent formalin, 4 hours 30 minutes exposuure. Subcutaneously.]

The results with formaldehyd have a special significance in view of the fact that this active reducing agent is capable of destroying the poisonous properties of tetanus and diphtheria toxines.

We have shown before that the sensitizing and poisonous principles in horse serum are not dialyzable through parchment paper. From the following limited experiments it would seem that the sensitizing principle is not dialyzable through a collodion sac when placed in the peritoneal cavity of the animal.

G. P. No. B. Six c. c. normal horse (No. 15) serum, subcutaneously. No symptoms. [Previous treatment: 32 days prior, collodion sac containing about 3 c. c. normal horse (No. 15) serum placed in peritoneal cavity.]

 $^{^\}alpha$ Besredka, A., and Steinhardt, Edna: De l'anaphylaxie et de l'anti-anaphylaxie vis-à-vis du sérum de cheval. Ann. de l'Inst. Pasteur, Vol. 21, No. 2, Feb. 25, 1907, pp. 117–127.

G. P. No. Cx. Collodion sac containing about 3 c. c. normal horse (No. 15) serum placed in peritoneal cavity.

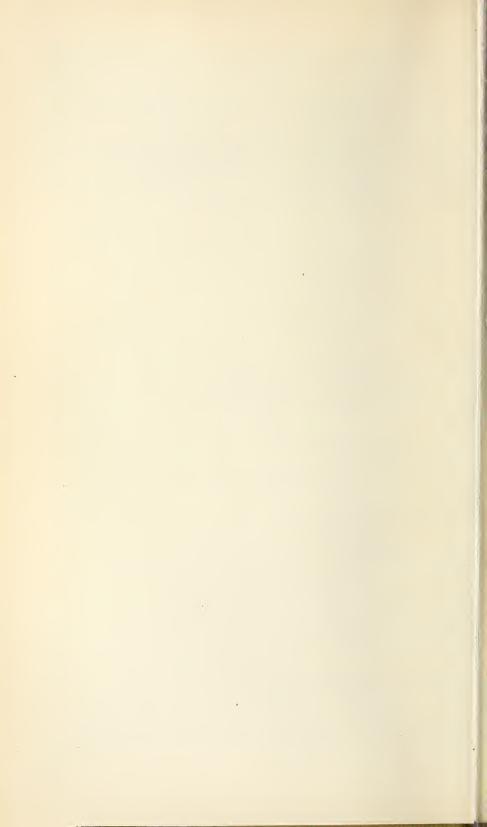
28 days later, 5 c. c. normal horse (roan) serum, subcutaneously. No symptoms.

1 day later, sac removed.

 $23~{\rm days}$ after removal of sac, 5 c. c. normal horse (roan) serum, subcutaneously. Mild symptoms.

By reference to page 62 it will be seen that guinea pigs may be sensitized by injecting horse serum directly into the heart.

Further work upon the action of the sensitizing substance is in progress and will be reported later.



Part II.

THE TOXIC PRINCIPLE.

We added a number of different ferments, alkaloids, and simpler chemical substances to horse serum in order to modify, destroy, or neutralize its toxic action. All these attempts have so far been found unavailing, as will be seen by the following experiments:

FERMENTS.

The ferments were added to the horse serum and allowed to stand at 15° C. over night.

Taka diastase.

G. P. No. 5417. Six c. c. antitoxic horse serum (Natl. IX., 19)+Taka diastase (PDCo.), subcutaneously. Dead, 90 minutes.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{500}$ c. c. antitoxic horse serum (A. A 208).]

Pancreatin.

G. P. No. 5413. Six c. c. same serum+pancreatin (PDCo.), subcutaneously. Dead, 40 minutes.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $9+\frac{1}{600}$ c. c. antitoxic horse serum (A. A 208).]

Rennin.

G. P. No. 5405. Six c. c. same serum+rennin (Hansen's junket tablet), subcutaneously. Dead, 50 minutes.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{500}$ c. c. antitoxic horse serum (A. A 208).]

Myrosin.

G. P. No. 5416. Six c. c. same serum+myrosin (from white mustard seed), subcutaneously. Dead, 25 minutes.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{700}$ c. c. antitoxic horse serum (A. A 208).]

Invertin.

G. P. No. 5375. Six c. c. same serum+invertin (yeast), subcutaneously. Dead, 80 minutes.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5 + \frac{1}{1000}$ c. c. antitoxic horse serum (NYBH 305)].

Emulsin.

G. P. No. 5373. Six c. c. same serum+emulsin (from almonds), subcutaneously. Severe symptoms.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{900}$ c. c. antitoxic horse serum (NYBH 305).]

Pepsin in acid solution.

G. P. No. 5409. Six c. c. same serum+pepsin (Wyeth's) rendered acid with HCl, subcutaneously. Severe symptoms.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{600}$ c. c. antitoxic

horse serum (A. A 208).

Pepsin in alkaline solution.

G. P. No. 5412. Six c. c. same serum+pepsin (Wyeth's) rendered alkaline, subcutaneously. Severe symptoms. (Died 18 hours later.)

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{500}$ c. c. antitoxic horse serum (A. A 208).]

Ingluvin.

G. P. No. 5418. Six c. c. same serum+ingluvin (Warren), subcutaneously. Severe symptoms. (Died 18 hours later.)

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{500}$ c. c. antitoxic horse serum A. A 208).]

Malt.

G. P. No. 5411. Six c. c. same serum+malt (from corn), subcutaneously. Severe symptoms.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{500}$ c. c. antitoxic horse serum (A. A 208).]

Papain.

G. P. No. 5414. Six c. c. same serum+papain (Merck's), subcutaneously. Severe symptoms. (Died 18 hours later.)

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{600}$ c. c. antitoxic horse serum (A. A 208).]

ALKALOIDS.

Atropin.

G. P. No. 5408. Six c. c. antitoxic horse serum (Natl. IX, 19)+0.002 atropin sulphate, subcutaneously. Dead, 27 minutes.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{570}$ c. c. antitoxic horse serum (A. A 208).]

Strychnin.

P. No. 5420. Six c. c. same serum+0.001 gm. strychnin sulphate, subcutaneously. Dead, 25 minutes.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{600}$ c. c. antitoxic horse serum (A. A 208).]

Morphin.

G. P. No. 5406. Six c. c. same serum+0.002 gm. morphin sulphate, subcutaneously. Dead, 35 minutes.

[Previous treatment: 27 days prior, 0.139 c. c. toxine No. $5+\frac{1}{500}$ c. c. antitoxic horse serum (A. A 208).]

Caffein.

G. P. No. 5377. Six c. c. antitoxic horse serum (Natl. IX, 19)+0.01 gm. caffein citrate, subcutaneously. Dead, 55 minutes.

[Previous treatment: 28 days prior, 0.139 c. c. toxine No. $5 + \frac{1}{1050}$ c. c. antitoxic horse serum (NYBH 305).]

SALTS.

Calcium chlorid.

G. P. No. 5342. Six c. c. same serum+0.5 c. c. calcium chlorid, subcutaneously. Severe symptoms.

[Previous treatment: 35 days prior, 0.139 c. c. toxine No. $5+\frac{1}{3\,2\,0}$ c. c. antitoxic horse serum (A. A 201).]

Sodium nitrate.

G. P. No. 5376. Six c. c. same serum+0.3 c. c. 1 per cent sodium nitrate, subcutaneously. Severe symptoms.

[Previous treatment: 28 days prior, 0.139 c. c. toxine No. $5+\frac{1}{3\,000}$ c. c. antitoxic horse serum (NYBH 305).]

Sodium chlorid.

G. P. No. 5341. Six c. c. same serum+0.5 gm. sodium chlorid, subcutaneously. Severe symptoms.

[Previous treatment: 35 days prior, 0.139 c. c. toxine No. $5+\frac{1}{240}$ c. c. antitoxic horse serum (A. A 201).]

Magnesium sulphate.

G. P. No. 5364. Six c. c. same serum+0.2 gm. magnesium sulphate, subcutaneously. Dead, 38 minutes.

[Previous treatment: 35 days prior, 0.139 c. c. toxine No. $5+\frac{1}{1000}$ c. c. antitoxic horse serum (NYBH 306).]

 $Ammonium\ sulphate.$

G. P. No. 5384. Six c. c. same serum+0.1 gm. ammonium sulphate, subcutaneously. Dead at once.

[Previous treatment: 28 days prior, 0.139 c. c. toxine No. $5+\frac{1}{1200}$ c. c. antitoxic horse serum (NYBH 305).]

G. P. No. 7847. Six c. c. normal horse (roan) serum+0.1 gm. ammonium sulphate, subcutaneously. Dead, 23 minutes.

[Previous treatment: 59 days prior, 0.24 c. c. toxine No. $9+\frac{1}{300}$ c. c. antitoxic horse serum (A 192).]

MISCELLANEOUS SUBSTANCES.

Ox bile.

G. P. No. 7616. Six c. c. normal horse (No. 15) serum+heated ox bile, equal parts, intraperitoneally 3 hours after mixing. Dead, 11 minutes.

[Previous treatment: 45 days prior, 0.142 c. c. toxine No. $5 + \frac{1}{360}$ c. c. antitoxic horse serum (A. A 248).]

Animal charcoal.

G. P. No. 7614. Six c. c. normal horse (No. 15) serum+animal charcoal; shaken up well, filtered, let stand for 3 hours; intraperitoneally. Dead, 20 minutes.

[Previous treatment: 45 days prior, 0.142 c. c. toxine No. $5 + \frac{1}{380}$ c. c. antitoxic horse serum (A. A 248).]

Yeast cells.

G. P. No. 7615. Six c.c. normal horse (No. 15) serum+ground yeast cells; let stand 3 hours; intraperitoneally. Dead, 35 minutes.

[Previous treatment: 45 days prior, 0.142 c. c. toxine No. $5+\frac{1}{380}$ c. c. antitoxic horse serum (A. A 245).]

FORMALDEHYD.

In view of the fact that formaldehyd has a destructive action upon such "haptin" substances as tetanus and diphtheria toxines, and in further view of the fact that the sensitizing and toxic principles of horse serum seem to belong to the haptin group of substances in the sense used by Ehrlich, it became interesting to determine what effect formaldehyd would have upon hypersusceptibility produced by horse serum.

Normal serum+5 per cent formalin:

G. P. No. 442. Six c. c. normal horse (roan) serum+5 per cent formalin, subcutaneously. Dead, 12 minutes.

[Previous treatment: 33 days prior, 0.0006 gm. tetanus toxine $A + \frac{1}{1000}$ c. c. antitoxic horse serum (Hoechst), subcutaneously.]

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G. P. No. 7542. Six c. c. same serum+5 per cent formalin; 21 hours exposure; subcutaneously. Dead, 60 minutes.

[Previous treatment: 35 days prior, 0.142 c. c. toxine No. $5+\frac{1}{170}$ c. c. anti-

toxic horse serum (Mul 2100).]

Control G. P. Six c. c. same serum+5 per cent formalin; 21 hours exposure; subcutaneously. Severe symptoms of formaldehyd poisoning.

Antitoxic serum+1 per cent formalin:

G. P. No. 7501. Six c. c. antitoxic horse serum (Natl. IX)+1 per cent formalin; 22 hours exposure; subcutaneously. Dead, 60 minutes.

[Previous treatment: 41 days prior, 0.24 c. c. toxine No. $9 + \frac{1}{560}$ c. c. antitoxic

horse serum (Led 4D).]

G. P. No. 7198. Five c. c. same mixture; 4 days exposure; subcutaneously. Marked symptoms.

[Previous treatment: 59 days prior, 0.142 c. c. toxine No. $5+\frac{1}{920}$ c. c. anti-

toxic horse serum (NYBH 305).]

G. P. No. 7589. Five c. c. same mixture; 4 days exposure; subcutaneously. Mild symptoms.

[Previous treatment: 37 days prior, 0.142 c. c. toxine No. $5+\frac{1}{1700}$ c. c. antitoxic horse serum (Park ppt. 305–306).]

Normal serum+5 per cent formalin:

G. P. No. 7776. Six c. c. normal horse (roan) serum+5 per cent formalin; 4 hours 20 minutes exposure; subcutaneously. Mild symptoms.

[Previous treatment: 46 days prior, 0.142 c. c. toxine No. $5+\frac{1}{340}$ c. c. antitoxic horse serum (S 1351).]

G. P. No. 7629. Six c. c. same mixture and exposure; subcutaneously. Slight symptoms.

[Previous treatment: 64 days prior, 0.142 c. c. toxine No. $5+\frac{1}{350}$ c. c. antitoxic horse serum (A. ppt. 31).]

G. P. No. 7689. Six c. c. same mixture; 4 hours exposure; subcutaneously. Slight symptoms.

[Previous treatment: 56 days prior, 0.142 c. c. toxine No. $5+\frac{1}{330}$ c. c. antitoxic horse serum (PDCo 080235).]

While the above three pigs showed slight and mild symptoms of hypersusceptibility, they all showed severe symptoms of formaldehyd poisoning.

These results plainly show that formaldehyd, in the strength and time stated, does not apparently appreciably influence the toxicity of horse serum. We have seen before that it also has no effect upon the sensitizing action, page 12.

CALCIUM CHLORID.

Netter^a has shown that when 1 gram of calcium chlorid is given on the day of injection and on the two following days the number of

a Netter, Arnold: Efficacité de l'ingestion de chlorure de calcium comme moyen préventif des éruptions consécutives aux injections de sérum. Compt. rend. soc. biol., tome 60, No. 6, Feb. 16, 1906, p. 279.

^{——:} Influence des quantités de sérum injectées et du nombre des injections sur les éruptions sériques. Nécessité d'augmenter les quantités de sels de chaux dans les cas d'injections répétées ou supérieures à quarante centimètres cubes. Idem, p. 281.

children showing eruption following the injection of serum is greatly reduced. We thought perhaps this salt might have some influence upon the phenomenon produced in guinea pigs by two injections of horse serum. Three series of experiments were made with this object in view.

The effect of 0.1 gram of CaCl₂ by mouth for three consecutive days before the second injection of horse scrum.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|-------------|---|---------------------------------------|
| 6065 | 0.14 c.c. toxine $5+\frac{1}{680}$ c. c. antitoxic horse serum (NYHD. 310), subcutaneously; then for 3 days before second injection, 0.1 gm. CaCl ₂ by mouth. | Days. 65 | 3 c. c. normal horse (roan) serum, intraperitoneally. | Marked symptoms. |
| 6063 6064 | do | | 6 c. c. normal horse (roan) serum, intraperitoneally. | Dead, 40 minutes. Dead, 3 minutes. |
| 6057 | do | 65 | 3 c. c. normal horse (roan) serum, in- traperitoneally. | Mild symptoms. |
| 6062 | 0.14 c. c. toxine $5+\frac{1}{720}$ c. c. antitoxic horse serum (NYHD 310), subcutaneously; then for 3 days before second injection, 0.1 gm. CaCl ₂ by mouth. | 65 | do | Dead, 20 minutes. |

Effect of 0.1 gram CaCl₂ duily for twenty days before the sensitizing inoculation was given, and every other day until the second injection of horse serum, fourteen days later.

| No. G. P. | , First injection. | Interval. | Second injection. | Result. |
|--------------|--|----------------------|---|--|
| ٠ | 0.1 gm. CaCl ₂ by mouth daily for 20 days; then 0.14 c. c. toxine $5+\frac{1}{250}$ c. c. antitoxic horse serum (S. 1500), subcutaneously; then CaCl ₂ every other day till second | Days. | 3 c. c. normal horse (roan) serum, in- traperitoneally. | Dead, 30 minutes. |
| | injection. dododododododo | 14 14 14 14 | do do do | Severe symptoms. Marked symptoms. Severe symptoms. Severe symptoms. |

Effect of 0.1 gram $CaCl_2$ for fifteen consecutive days before the second injection of horse serum, 104 days after the sensitizing injection.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|--------------|---|-----------------------|
| 5556 | 0.14 c. c. toxine $5+\frac{1}{600}$ c. c. antitoxic horse serum (NYBH. 310), subcutaneously; then daily for 15 days before second injection, 0.1 gm. CaCl ₂ by | Days. 104 | 6 c. c. normal horse (roan) serum, in- traperitoneally. | Dead, 35 minutes. |
| 5550 | mouth. 0.14 c. c. toxine $5+\frac{1}{300}$ c. c. antitoxic horse serum (NYBH. 310), subcutaneously; then daily for 15 days before second injection, 0.1 gm. CaCl ₂ by | 104 | do | Dead, 25 minutes. |
| 5557 | mouth. 0.14 c. c. toxine $5 + \frac{1}{500}$ c. c. antitoxic horse serum (NYBH. 310), subcutaneously; then daily for 15 days before second injection, 0.1 gm. CaCl ₂ by | 104 | do | Dead, 15 minutes. |
| 5553 | mouth. 0.14 c. c. toxine $5+\frac{1}{400}$ c. c. antitoxic horse serum (NYBH. 310), subcutaneously; then daily for 15 days before second injection, 0.1 gm. CaCl ₂ by mouth. | 104 | 3 c. c. normal horse (roan) serum, in- traperitoneally. | Dead, 18 minutes. |
| 5555 | 0.14 c. c. toxine $5+\frac{1}{300}$ c. c. antitoxic horse serum (NYBH. 310), subcutaneously; then daily for 15 days before second injection, 0.1 gm. CaCl ₂ by mouth. | 104 | do | Dead, 65 minutes. |
| 5552 | 0.14 c. c. toxine $5+\frac{1}{100}$ c. c. antitoxic horse serum (NYBH.310), subcutaneously; then daily for 15 days before second injection, 0.1 gm. CaCl ₂ by mouth. | 104 | do | Severe symptoms. |
| 5551 | 0.14 c. c. toxine $5+\frac{1}{300}$ c. c. antoxine horse serum (NYBH. 310), subcutaneously; then daily for 15 days before second injection, 0.1 gm. CaCl ₂ by mouth. | 104 | do | Very severe symptoms. |

Effect of 0.1 gram CaCl₂ for fifteen consecutive days before the second injection of horse serum, 104 days after the sensitizing injection—Continued.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|--------------|---|------------------|
| 5554 | 0.14 c. c. toxine $5 + \frac{1}{500}$ c. c. antitoxic horse serum (NYBH. 310), subcutaneously; then daily for 15 days before second injection, 0.1 gm. CaCl ₂ by mouth. | Days. 104 | 6 c. c. normal horse (roan) serum, intraperitoneally. | Marked symptoms. |

As will be seen from the above, the guinea pigs which received 0.1 gm. of CaCl₂ for three days previous to the second injection of serum reacted in the usual manner, 2 of them dying in a few minutes, and the other two had severe symptoms.

Of those which received 0.1 gm. CaCl₂ daily for twenty days before being given their sensitizing dose and then every other day for fourteen days before they were given the second dose of serum, none showed any marked resistance.

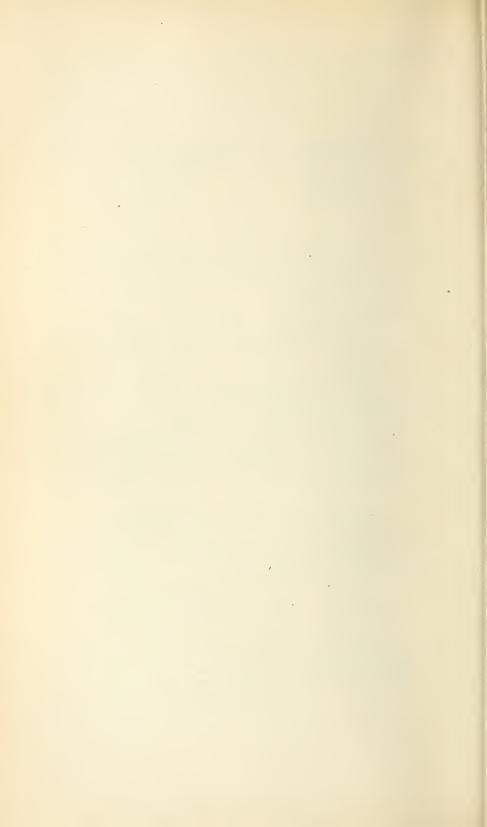
Of those that were sensitized first and then given 0.1 gm. CaCl₂ for the fourteen days previous to the second dose, 5 out of 8 died in spite of the fact that 4 received only 3 c. c. of serum.

It is, therefore, quite plain that, while CaCl₂ may modify the occurrence of rashes in children following a single injection of serum, it does not influence to any marked extent the toxic effect in guinea pigs of a second injection of serum given fourteen days after the first injection.

FREEZING DOES NOT DESTROY THE TOXIC PRINCIPLE.

Some normal horse serum (roan) was frozen hard at 15° F., then thawed at room temperature, and found toxic when injected into a sensitized guinea pig. For example:

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|-------------|--|------------------|
| 5329 | 0.22 c. c. toxine no. 7+ \[\frac{1}{100} \] c. c. antitoxic horse serum (Natl., XIV), subcutane- ously. | Days. 13 | 6 c. c. normal horse (roan) serum, frozen hard in brine, then thawed at room temperature, intraperitoneally. | Severe symptoms. |



Part III.

IS THE TOXIC PRINCIPLE SPECIFIC?

The toxic action is quantitatively specific so far as various blood serums are concerned. That is, a guinea pig sensitized with horse serum is more susceptible to a subsequent injection of horse serum than to a subsequent injection of the blood serum of cattle, sheep, cats, dogs, hogs, etc. The specific character of the hypersusceptibility is more apparent when proteid substances of quite different origin are used at the first and second injections. For example, guinea pigs sensitized with horse serum do not react at all to subsequent injections of peptone, vegetable proteid extracts, egg albumen or milk.

Horse serum versus other proteid substances.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|-----------|--|--------------|
| 7030 | 0.142 c. c. toxine 5+ $\frac{1}{130}$ c. c. antitoxic horse serum (Ld. 4B), | Days. 52 | 6 c. c. 3 per cent peptone, intraperitoneally. | No symptoms. |
| 7039 | subcutaneously. 0.142 c. c. toxine $5+\frac{1}{1060}$ c. c. antitoxic horse serum (Ld. 5 H), | 52 | do | No symptoms. |
| 7038 | subcutaneously. 0.142 c. c. toxine 5+ \$\frac{5}{5}\tau_0\$ c. c. antitoxic horse serum (Ld.5H), subcutaneously. | 52 | 6 c. c. watery extract of peas kept at 15° C. 24 hours, filtered through porcelain intraperitoneally. | No symptoms. |
| 7033 | 0.142 c. c. toxine 5+ \frac{1}{7\frac{1}{40}} c. c. antitoxic horse serum (Ld. 34), subcutaneously. | 52 | do | No symptoms. |
| 7036 | 0.142 c. c. toxine 5+ 10.142 c. c. toxine 5+ 10.142 c. c. antitoxic horse serum (Ld.5H), subcutaneously. | 52 | 6 c. c. egg albumen, saturated solution in salt water (not filtered), intraperi- toneally. | No symptoms. |
| 7041 | 0.142 c. c. toxine 5+ 7 60 c. c. antitoxic horse serum (Ld.5C), subcutaneously. | 52 | do | No symptoms. |
| 7032 | 0.142 c. c. toxine 5+ | 52 | 6 c. c. bottom milk unfiltered, intrape- ritoneally. | No symptoms. |
| 7035 | 0.142 c. c. toxine 5+ \$\frac{1}{300}\$ c. c. antitoxic horse serum (Ld.5H), subcutaneously. | 52 | do | No symptoms. |

It naturally occurred to us to determine whether guinea pigs sensitized with injections of albuminous substances, such as are contained in milk, the white of eggs, peas, etc., are sensitive to subsequent injections of horse serum.

The following experiments plainly indicate that animals sensitized with milk, egg albumen, peptone, or the albuminous substance extracted from peas, do not react when subsequently injected with horse serum.

Other proteid substances versus horse serum.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|-------------------|---|----------------|--|--|
| 705 | 0.1 c. c. fresh whole milk, subcutaneously. | Days. | 6 c. c. normal horse (No. 15) serum, intraperitoneally. | No symptoms. |
| 736 738 703 | do do $\frac{1}{250}$ gm. peptone, sub- | 31 31 31 | dododo | No symptoms. No symptoms. No symptoms. |
| 730 | cutaneously. 1 c. c. egg albumen in salt solution, subcutaneously. | 21 | do | No symptoms. |
| 731 742 | 1 c. c. watery extract peas, subcutaneously. | 21 21 | do 10 c. c. normal horse (No. 15) serum, intraperitoneally. | No symptoms. |
| 7 4 3 | 0.5 c. c. watery extract peas, subcutaneously. | 21 | do | No symptoms. |
| 744 | 0.1 c. c. watery extract peas, subcutaneously. | 21 | do | No symptoms. |
| 484 | 1 c. c. normal horse (roan) serum, subcutaneously. | 64 | 6 c. c. hemoglobin, horse (roan), in- traperitoneally. | Mild symptoms. |
| 485 | 6 c. c. normal horse (roan) serum, subcutaneously. | 64 | do | Mild symptoms. |
| 486 487 | dodo | | do | Slight symptoms. Slight symptoms. |

Part IV.

OTHER BLOOD SERUMS AND OTHER ALBUMINOUS SUBSTANCES ARE ALSO TOXIC.

So much of our work has been done with horse serum that we desire to record some further experiments with the blood serums of other animals. We confirm and extend our previous work that the same reactions may be induced in the guinea pig with the blood serums of various animals, such as the dog, ox, sheep, cat, and hog.

Other blood serums.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--|---|--|---|---|
| 461 462 463 465 466 467 468 469 470 471 472 473 474 475 476 477 480 481 482 483 484 485 | Subcutaneously, 2 3 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 | Days. 37 37 37 37 37 37 37 37 37 37 37 37 70 70 70 70 70 70 70 70 70 70 70 70 70 | Intraperitone- ally, 6 c. c. se- rum of— Oxdodododo Sheepdo | Dead, 120 minutes. Marked symptoms. Severe symptoms. Severe symptoms. Slight symptoms. Dead, 110 minutes. Severe symptoms. Dead, 12 hours. Mild symptoms. Dead, 12 hours. Dead, 12 hours. Dead, 12 minutes. Dead, 120 minutes. Dead, 20 minutes. Dead, 65 minutes. Dead, 70 minutes. Dead, 120 minutes. Dead, 120 minutes. Dead, 120 minutes. Dead, 100 minutes. Dead, 50 minutes. Dead, 50 minutes. Dead, 50 minutes. Dead, 50 minutes. Dead, 65 minutes. |

OTHER ALBUMINOUS SUBSTANCES.

As soon as we concluded that it is probably the proteid substance in horse serum that is chiefly concerned in sensitizing and poisoning the guinea pigs, we thought of other proteid substances obtained from widely different sources.

We have found that hemoglobin, egg albumen, milk, and extract of peas are quite as active as horse serum. Peptone seems to have slight sensitizing and poisonous properties; leucin and tyrosin none at all. The reaction following the second injection of proteid matter in the guinea pigs appears, then, to be common to all the higher forms of albuminous substances, no matter from what source. It occurs to us that this phenomenon of hypersusceptibility in the guinea pig may be used as a physiological test to distinguish true proteid substances from the lower forms of nitrogenous compounds. It would

be interesting to determine whether the synthetic peptids and polypeptids of Fisher sufficiently approach the true proteid molecular structure to induce hypersusceptibility in the guinea pig.

From our work with other proteid substances it was but a logical step to the albuminous content of the bacterial cell, which is dealt with in another part of this work.

Hemoglobin versus hemoglobin.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|-------------|--|-----------------------|
| 411 | 0.1 c. c. hemoglobin (washed 20 times) subcutaneously. | Days. 22 | 6 c. c. hemoglobin, intraperitoneally. | Marked symptoms. |
| 412 | 0.5 c. c. hemoglobin (washed 20 times) subcutaneously. | 22 | do | Severe symptoms. |
| 413 | 1 c. c. hemoglobin (washed 20 times) subcutaneously. | 22 | do | Dead, 5 minutes. |
| 414 | 3 c. c. hemoglobin (washed 20 times) subcutaneously. | 22 | do | Slight symptoms. |
| 415 | 5 c. c. hemoglobin (washed 20 times) subcutaneously. | 22 | do | Very severe symptoms. |

The hemoglobin was obtained by dissolving the washed red corpuscles of a normal horse in distilled water. The red corpuscles for the hemoglobin solution used at the first injection, in order to sensitize the guinea pigs, was washed and centrifuged 20 times in order to surely wash away all traces of serum, the smallest remaining quantities of which might have confused the results. The hemoglobin used at the second injection was dissolved from red corpuscles washed four times.

Egg albumen versus egg albumen.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|-------------|---|-------------------|
| 406 | 0.1 c. c. egg albumen, subcutaneously. | Days. 22 | 6 c. c. saturated solution of egg albumen in salt solution, intraperitoneally. | Dead, 30 minutes. |
| 407 | 0.5 c. c. egg albumen, subcutaneously. | 22 | dodo | Dead, 18 minutes. |
| 408 | 1 c. c. egg albumen, sub- cutaneously. | 22 | do | Dead, 25 minutes. |
| 409 | 3 c. c. egg albumen, sub- cutaneously. | 22 | do | Dead, 25 minutes. |
| 410 | 5 c. c. egg albumen, sub- cutaneously. | 22 | do | Dead, 20 minutes. |
| 732 | 1 c. c. egg albumen + salt solution, subcutaneously. | 21 | 6 c. c. egg albumen + salt solution equal quantities, intraperitoneally. | Severe symptoms. |

Milk versus milk.

| No. G. P. | First injection. | Interval. | Second injection, | Result. |
|--------------|--|-------------|---|----------------------------|
| 401 | 3 c. c. milk filtered through porcelain, subcutaneously. | Days, 26 | 10 c. c. bottom milk, intraperitoneally. | Slight symptoms. |
| 402 | 1 c. c. milk filtered through porcelain, subcutaneously. | 26 | do | Slight symptoms. |
| 403 | 0.5 c. c. milk filtered through por elain, subcutaneously. | 26 | do | Slight symptoms. |
| 404 | 0.1 c. c. milk filtered through porcelain, subcutaneously. | 26 | do | Slight symptoms. |
| 706 | 0.5 c. c. fresh whole milk, subcutaneously. | 31 | 10 c. c. fresh whole milk, intraperitoneally. | Dead, 20 minutes. |
| 707 | 1 c. c. fresh whole milk, subcutaneously. | 31 | do | Very severe symp- toms. |
| 708 | 3 c. c. fresh whole milk, subcutaneously. | 31 | do | |
| 709 | 5 c. c. fresh whole milk, subcutaneously. | 31 | do | Very severe symp- |
| 737 | 0.1 c.c. fresh whole milk, subcutaneously. | 31 | do | Very severe symp- toms. |
| Contro | ol: 6 c. c. fresh whole mill | k, intrape | ritoneally | No symptoms. |

Peas versus peas.

| | - | | | | |
|--|---|-------------|--|-------------------------------|--|
| No. G. P. | First injection. | Interval. | Second injection. | Result. | |
| 416 | 0.1 c. c. watery extract peas, 24 hours at 15° C. (acid), filtered through porcelain, subcutaneously. | Days. 26 | 10 c. c. watery extract of peas, 3 days at 15° C., filtered through porcelain intrape- | Marked symptoms. | |
| 417 | 0.5 c. c. watery extract peas, 24 hours at 15° C. (acid), filtered through porcelain, subcutaneously. | 26 | ritoneally. | Dead, 7 hours and 30 minutes. | |
| 418 | 1 c. c. watery extract peas, 24 hours at 15° C. (acid), filtered through porcelain, subcutaneously. | 26 | do | Marked symptoms, | |
| 419 | 3 c. c. watery extract peas, 24 hours at 15° C. (acid), filtered through porcelain, subcutaneously. | 26 | do | Dead, 4 hours. | |
| 420 | 5 c. c. watery extract peas, 24 hours at 15° C. (acid), filtered through porcelain, subcutaneously. | 26 | do | Dead, 2 hours. | |
| Control: 10 c. c. waterv extract of peas, 3 days at 15° C., fil- | | | | No symptoms. | |
| tered through porcelain, intraperitoneally. | | | 37 | | |
| Do | | | No symptoms. | | |

Peptone versus peptone.

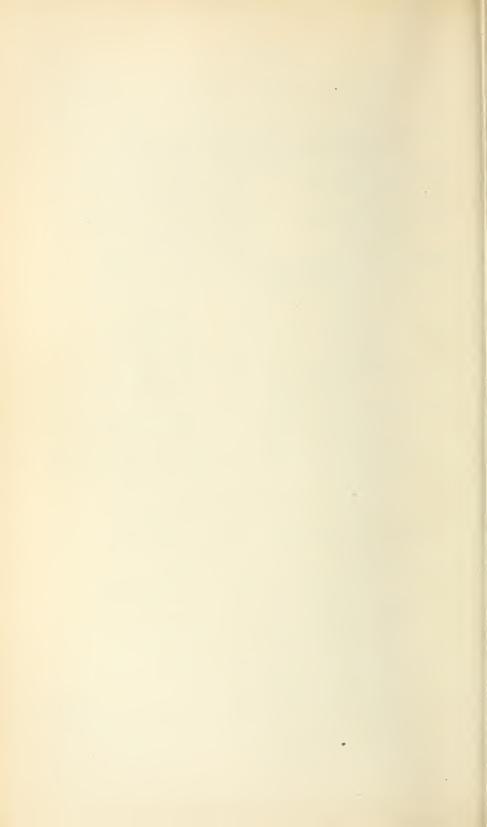
| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--|--|-------------|--|------------------|
| 492 | 0.5 c. c. half-saturated solution peptone, sub- | Days. 21 | 6 c. c. saturated solution of peptone, | Slight symptoms. |
| 490 | cutaneously. 1 c. c. half-saturated solution peptone, sub- | 21 | intraperitoneally. | Mild symptoms. |
| 491 | cutaneously. 3 c. c. half-saturated solution peptone, subcutaneously. | 21 | do | Marked symptoms. |
| 488 | 0.1 c. c. half-saturated solution peptone, subcutaneously. | 21 | do | Marked symptoms. |
| 748 | 0.1 c. c. heated, half-sat- urated solution pep- tone, subcutaneously. | 21 | 6c. c. half-saturated solution of peptone, intraperitoneally. | Marked symptoms. |
| 749 | 0.5 c. c. heated, half-sat- urated solution pep- tone, subcutaneously. | 21 | dodo | No symptoms. |
| 750 | 1 c. c. heated, half-sat- urated solution pep- tone, subcutaneously. | 21 | do | Marked symptoms. |
| 751 | 3 c. c. heated, half-sat- urated solution, pep- tone, subcutaneously. | 21 | 6c. c. half-saturated solution of peptone, intraperitoneally. | Marked symptoms. |
| 752 | 5 c. c. heated, half-sat- urated solution pep- tone, subcutaneously. | 21 | 6 c. c. half-saturated solution of pep- tone, subcutane- | Marked symptoms. |
| Control: 6 c. c. half-saturated solution of peptone, heated, subcutaneously. | | | | No symptoms. |

Tyrosin versus tyrosin.

| No. G. P. | First injection. | Interval. | Second injection. | Results. |
|--|---|-------------|--|--------------|
| 739 | 0.002 gm. watery solution of tyrosin, subcutaneously. | Days. 21 | 6 c. c. tyrosin (0.1 gm. + 50 c. c.) watery solution, in- | No symptoms. |
| 740 | 0.01 gm. watery solution of tyrosin, subcutaneously. | 21 | traperitoneally. 20 c. c. tyrosin (0.1 gm. + 50 c. c.) watery solution, intraperitoneally. | No symptoms. |
| 741 | 0.02 gm. watery solution of tyrosin, subcutaneously. | 21 | 10 c. c. tyrosin (0.1 gm. + 50 c. c.) watery solution, intraperitoneally. | No symptoms. |
| Control: 6 c. c. tyrosin (0.1 gm. + 50 c. c.) watery solution, intraperitoneally, into a fresh guinea pig. | | | | No symptoms. |

Leucin versus leucin.

| No. G. P. | First injection. | Interval. | Second injection. | Results. |
|-----------------|---|--------------|--|--------------|
| 745 | 0.02 gm. leucin in watery solution, subcutaneously. | Days. 17 | 20 c. c. (0.2 gm. + 50 c. c.) watery solution intraperitoneally. | No symptoms. |
| 746 | 0.02 gm. leucin in watery solution, subcutaneously. | 17 | | No symptoms. |
| 747 | 0.02 gm. leucin in watery solution, subcutaneously. | 17 | 10 c. c. (0.2 gm. + 50 c. c.) watery solution intraperitoneally. | No symptoms. |
| Contro trape | ol: 6 c. c. leucin (0.2 gm. eritoneally, into a fresh g | No symptoms. | | |



Part V.

HYPERSUSCEPTIBILITY AND IMMUNITY PRODUCED BY BACTERIAL PROTEIDS.

Experimental studies with the bacterial proteids are of the greatest importance on account of the practical uses to which results along this line may lead. Our conviction that the phenomenon of hypersusceptibility which we have been studying in the guinea pig has a deep significance in general pathology, especially in the problem of immunity, induced us to undertake an extensive series of experiments with proteid extracts obtained from bacterial cell masses. Some of this work is sufficiently advanced for us to record our results in part.

Hypersusceptibility may easily be induced in guinea pigs with proteid extracts obtained from the bacterial cell. The first injection of most of the extracts used by us seems comparatively harmless to the animal. A second injection of the same extract shows, however, that profound physiological changes have taken place. A definite period must elapse between the first and the second injection. The symptoms presented by the guinea pigs as a result of the second injection resemble those caused by horse serum.

The phenomenon induced by a second injection is followed (in certain cases) by an immunity to the corresponding infection.

These results strengthen our belief that the phenomenon of hypersusceptibility has a practical significance in the prevention and cure of certain infectious processes. It gives a possible explanation to the period of incubation of some of the communicable diseases. Is it a coincidence that the period of incubation of a number of infectious diseases is about ten to fourteen days, which corresponds significantly with the time required to sensitize animals with a strange proteid? In certain infectious diseases with short periods of incubation, such as pneumonia, the crisis which commonly appears about the tenth day may find a somewhat similar explanation. It is evident that disease processes produced by soluble toxines, such as diphtheria and tetanus, do not belong to the category now under consideration.

EXTRACT OF COLON BACILLUS.

The extract from the colon bacillus in the following experiments was obtained as follows:

A 2-day-old culture of *B. coli communis* in Dunham's solution was used to heavily inoculate the surface of 84 large agar plates. These

plates were grown at 37° C. for four days and the surface growth collected.

The bacterial mass was frozen forty-eight hours at about 15° F., thawed at room temperature, and then ground with sand by hand in a mortar and pestle for five hours, shaken vigorously half an hour, and again frozen eighteen hours. After again thawing, the fluid was diluted with salt solution and filtered through a Berkefeld filter. The clear filtrate gave a distinct coagulum with heat and acetic acid.

All the other extracts were obtained by a similar process. In the case of the tubercle bacillus the bacterial mass was first washed three days in running water to eliminate the soluble tuberculin as much as possible.

B. coli.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|-------------------|---|-----------|--|---|
| 817 819 818 | 5 c. c. colon extract, subcutaneously. 1 c. c. colon extract, subcutaneously. | Days. 35 | 6 c. c. colon extract, intraperitoneally. dodo | Marked symptoms. Mild symptoms. Marked symptoms |
| 820 821 | do | 35 35 | subcutaneously. do 6 c. c. colon extract, | Slight symptoms. Mild symptoms. |
| 822 | 0.5 c. c. colon extract, subcutaneously. | 35 | intraperitoneally. 6 c. c. colon extract, subcutaneously. | Mild symptoms. |
| 823 824 | 0.1 c. c. colon extract, subcutaneously. 0.01 c. c. colon extract, | 35 35 | 6 c. c. colon extract, intraperitoneally. 6 c. c. colon extract, | Slight symptoms. Marked symptoms. |
| 825 | subcutaneously. 0.005 c. c. colon extract, subcutaneously. | 35 | subcutaneously. 6 c. c. colon extract, intraperitoneally. | Severe symptoms. |

The hypersusceptibility induced by the colon extracts manifested itself by symptoms resembling those already described in the case of horse serum. The guinea pigs scratched at the mouth with their hind legs. Most of them showed evidences of respiratory embarrassment by quickened, labored, or irregular breathing. Many of the pigs lay over on their sides, which is a very common symptom. A few developed jerky movements, but in no case was convulsion noted. The pigs looked quite sick and ill at ease, but gradually recovered, so that by next morning they seemed normal.

Ten days following the second injection of the extract all the above pigs were given 5 c. c. of a heavy emulsion of colon bacillus from 24-hour old agar slants, but showed no symptoms, and remain in good condition. Three controls received the same injection and died in twelve hours.

YEAST.

The manifestations of hypersusceptibility produced by the proteid extract from yeast cells are restlessness, scratching, irregular respirations; the guinea pigs lie down and look sick; sometimes jerky movements are seen and, in one instance, convulsions.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|-------------------|--|----------------|--|---|
| 755 | 1 c. c. extract yeast cells, subcutaneously. | Days. 22 | 6 c. c. extract yeast cells, intraperitoneally. | Very severe symptoms. |
| 754 | 0.5 c. c. extract yeast cells, subcutaneously. | 22 | 6 c. c. extract yeast cells, subcutaneously. | Slight symptoms. |
| 864 | 0.1 c. c. extract yeast cells, subcutaneously. | 19 | 5 c. c. extract yeast cells, subcutaneously. | Slight symptoms. |
| 803 | 1 c. c. extract yeast cells, subcutaneously. | 27 | 6 c. c. extract yeast cells, intraperitoneally. | Mild symptoms. |
| 807 804 809 | dodododo | 27 27 27 | do | Mild symptoms. Marked symptoms. Dead, 2 hours 10 |
| 802 | do | 27 | 6 c. c. extract yeast cells, subcutaneously. | minutes. Slight symptoms. |
| 805 810 806 | do do do | 27 27 27 | do do | Slight symptoms. Marked symptoms. Very severe symptoms. |
| 815 | 0.005 c. c. extract yeast cells, subcutaneously. | 27 | 5 c. c. extract yeast cells, subcutane- ously. | No symptoms. |
| 814 | 0.01 c. c. extract yeast cells, subcutaneously. | 27 | do | Slight symptoms. |
| 813 | 0.02 c. c. extract yeast cells, subcutaneously. | 27 | do | Mild symptoms. |
| 812 | 0.1 c. c. extract yeast cells, subcutaneously. | 27 | do | Severe symptoms. |
| 811 | 0.5 c. c. extract yeast cells, subcutaneously. | 27 | do | Very severe symptoms. |
| 801 | 5 c. c. extract yeast cells, subcutaneously. | 27 | do | Mild symptoms. |
| 800 | 10 c. c. extract yeast cells, subcutaneously. | 27 | do | Very severe symptoms. |

HAY BACILLUS.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|-----------|--|------------------|
| 869 | 1 c. c. extract subtilis, subcutaneously. | Days. | 7 c. c. extract subtilis, intraperitoneally. | Slight symptoms. |
| 864 | 10 c. c. extract subtilis, subcutaneously. | 25 | 6 c. c. extract subtilis, intraperitoneally. | Marked symptoms. |

3681—No. 36—07——5

HAY BACILLUS-Continued.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|---|-------------|--|------------------|
| 865 | 8 c. c. extract subtilis, subcutaneously. | Days. 25 | 6 c. c. extract sub- tilis, intraperito- neally. | No symptoms. |
| 866 | 6 c. c. extract subtilis, subcutaneously. | 25 | do | Marked symptoms. |
| 868 | | 25 | do | Marked symptoms. |
| 870 | 0.5 c. c. extract subtilis, subcutaneously. | 25 | do | Marked symptoms. |
| 871 | 0.1 c. c. extract subtilis, subcutaneously. | 25 | do | Slight symptoms. |
| 872 | 0.01 c. c. extract subtilis, subcutaneously. | 25 | do | Slight symptoms. |
| 873 | 0.001 c. c. extract subtilis, subcutaneously. | 25 | do | Slight symptoms. |

ANTHRAX.

Indications of hypersusceptibility produced by anthrax are scratching, rapid respirations; pigs frequently fall over on their sides and look sick. None of the pigs coughed or had convulsions.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|------------------------------|--|-------------|--|--|
| 842 | 10 c. c. extract of anthrax, subcutane- | Days. 21 | thrax, subcuta- | Marked symptoms. |
| 843 | ously. 5 c. c. extract of anthrax, subcutaneously. | 21 | neously. do | Mild symptoms. |
| 844 | | 21 | do | Mild symptoms. |
| \$45 \$46 \$47 \$48 | do d | 21 21 | dodododododododododo | Slight symptoms. Severe symptoms. Mild symptoms. Mild symptoms. |
| 849 | ously. 0.1 c. c. extract of anthrax, subcutaneously. | 21 | do | Slight symptoms. |
| 850 | 0.01 c. c. extract of anthrax, subcutane- | 21 | do | Slight symptoms. |
| 851 | | 21 | do | Slight symptoms. |
| 852 | 1 c. c. extract of an- thrax, subcutaneous- | 11 | do | No symptoms. |
| 853 854 | dodo | 11 11 | 4 c. c. extract anthrax, subcutaneously. | No symptoms. Mild symptoms. |
| | do | 11 | | Slight symptoms. |
| 856 | do | 11 | do | No symptoms. |

All the above guinea pigs were subsequently inoculated with a virulent culture of anthrax. They all died in a few days with the usual lesions.

A number of guinea pigs were given the extract from anthrax bacilli before infection; some were given a single injection, some two injections, and others daily injections for twenty days. Other guinea pigs were given the extract used as a vaccine, both in single and repeated injections, after being infected with anthrax bacilli. The extract did not seem to have any influence on the course of the disease, whether given before or after the infection.

TUBERCULOSIS.

The indications of hypersusceptibility induced by extract of tubercle bacilli are restlessness, scratching, irregular respiration, tremor; most of the pigs lie down on their sides and look sick.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|---|-----------|--|------------------|
| 827 | 0.1 c. c. extract human tubercle bacilli, subcutaneously. | Days. | 6 c. c. extract human tubercle bacilli, subcutane- | Mild symptoms. |
| 828 | 1 c. c. extract human tubercle bacilli, subcutaneously. | 32 | ously. do | No symptoms. |
| 829 | 2 c. c. extract human tubercle bacilli, subcutaneously. | 32 | do | Mild symptoms. |
| 830 | 3 c. c. extract human tubercle bacilli, subcutaneously. | 32 | do | Mild symptoms. |
| 831 | 4 c. c. extract human tubercle bacilli, subcutaneously. | 32 | do | Mild symptoms. |
| 832 | 5 c. c. extract human tubercle bacilli, sub- cutaneously. | 32 | 6 c. c. extract human tubercle bacilli, intraperitoneally. | Severe symptoms. |
| 833 | 6 c. c. extract human tubercle bacilli, subcutaneously. | 32 | ·do | Slight symptoms. |
| 834 | 7 c. c. extract human tubercle bacilli, subcutaneously. | 32 | 10 c.c. extract human tubercle bacilli, intraperitoneally. | Slight symptoms. |
| 835 | 8 c. c. extract human tubercle bacilli, subcutaneously. | 32 | 6 c. c. extract human tubercle bacilli, intraperitoneally. | Mild symptoms. |
| 836 | 9 c. c. extract human tubercle bacilli, sub- cutaneously. | 32 | | No symptoms. |

The guinea pigs which have reacted to two injections of proteid extract obtained from the tubercle bacillus are now being tested for immunity to infection with tubercle cultures.

TYPHOID.

The indications of hypersusceptibility induced by two injections of typhoid extract manifest themselves by rapid respirations; most of the pigs lie down on their sides. The symptoms presented by this series of pigs were mild.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|-------------|---|------------------|
| 857 | 10 c. c. typhoid extract, subcutaneously. | Days. 34 | 10 c. c. typhoid extract, subcutaneously. | Slight symptoms. |
| 858 | 5 c. c. typhoid extract, subcutaneously. | 34 | do | Slight symptoms. |
| 859 | 1 c. c. typhoid extract, subcutaneously. | 34 | do | Slight symptoms. |
| 860 | do | 34 | 6 c. c. typhoid ex- tract, intraperi- toneally. | Slight symptoms. |
| 862 | 0.5 c. c. typhoid extract, subcutane- | 34 | 10 c. c. typhoid extract, subcutaneously. | Slight symptoms. |
| 863 | 0.1 c. c. typhoid extract, subcutaneously. | 34 | do | Slight symptoms. |

Nine days following the second injection of the extract, five pigs of the above series, which had received 10 c. c. of the typhoid extract at the second injection, resisted a large dose of a virulent typhoid culture. Two controls died in eighteen hours. One or two of the pigs which had received the extract were slightly sick the following day, but the next day had fully recovered and have remained so. A definite immunity was, therefore, conferred by the two injections of extract from the typhoid bacillus.

TYPHO-LYSIN.

Along somewhat the same lines efforts to obtain the phenomenon of hypersusceptibility with the dissolved typhoid bacilli (natural lysis) failed.

G. P. 480W and 550W:

September 11 to 20, fed dead typhoid culture daily.

September 26 to 28, fed live typhoid culture daily.

December 14 to 28, 1 c.c. bouillon culture of typhoid daily, subcutaneously.

January 15, 10 c.c. heavy old bouillon culture of typhoid, intraperitoneally. Killed 1 hour later. Peritoneal contents collected. Then peritoneal cavity washed with salt solution.

G. P. 100T and 109T:

September 11 to 20, fed dead typhoid bacilli daily.

September 28 to November 9, fed live typhoid bacilli daily.

December 17 to 28, 1 c.c. bouillon culture of typhoid daily, subcutaneously.

February 19, 6 c.c. heavy old bouillon culture of typhoid, intraperitoneally. Killed 1 hour later. Peritoneal contents collected. Then peritoneal cavity washed with salt solution.

Peritoneal contents and saline washings from the peritoneal cavities of the above pigs used as follows:

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|-------------|--|------------------------------|
| 114 | 0.5 c. c. peritoneal contents of G. P. 480 and 550 containing dissolved typhoid bacilli, subcutaneously. | Days. 35 | 6 c. c. saline washings from peritoneal cavities of 10 pigs (100T to 109T) containing dissolved typhoid bacilli intraperitoneally. | No symptoms. |
| 115 | 0.75 c. c. peritoneal contents of G. P. 480 and 550 containing dissolved typhoid bacilli, subcutaneously. | 35 | do | No symptoms. |
| 116 117 | 2 c. c. peritoneal contents of G. P. 480 and 550 containing dissolved typhoid bacilli, subcutaneously. | · 35 35 | dodo | No symptoms. No symptoms. |
| 118 | 0.5 c. c. saline washings from peritoneal cavities of G. P. 480 and 550, containing dissolved typhoid ba- | 35 - | do | No symptoms. |
| 119 | cilli, subcutaneously. 1 c. c. saline washings from peritoneal cavities of G. P. 480 and 550, containing dissolved typhoid ba- | 35 | do | No symptoms. |
| 120 | cilli, subcutaneously. 1.5 c. c. saline washings from peritoneal cavities of G. P. 480 and 550, containing dissolved typhoid ba- | 35 | do | No symptoms. |
| 121 | cilli, subcutaneously. 6 c. c. saline washings from peritoneal cavities of G. P. 480 and 550, containing dissolved typhoid ba- | 35 | do | No symptoms. |
| 111T | cilli, subcutaneously. 0.5 c. c. bouillon culture of typhoid bacilli, subcutaneously. | 35 | do | No symptoms. |
| 112T | 1 c. c. bouillon culture of typhoid bacilli, subcutaneously. | 35 | do | |
| 113T | dodo | 35 | do | No symptoms. |



Part VI.

COMPARATIVE TOXICITY OF UNTREATED AND REFINED ANTITOXIC SERUM.

It has long been known that diphtheria antitoxin is precipitated from the serum with the globulins, and many attempts have been made to separate the antitoxin from the nonantitoxic substances contained in the serum.

Gibson a has evolved a practical method of concentrating and refining diphtheria antitoxic serum. Part of the process consists in placing the one-half saturation of ammonium sulphate precipitate derived from the antitoxic serum in saturated sodium chlorid solution. This dissolves a portion of the globulins with all the antitoxin. In this way the nucleoproteids and insoluble globulins present in the first precipitate are eliminated. The soluble globulins are precipitated by acetic acid, filtered, partially dried, and finally placed in a sack of parchment membrane and dialyzed in running water. This antitoxic solution of soluble globulins is then rendered neutral and sufficient sodium chlorid added to make it isotonic.

Park and Throne b find, from a comparative study of 100 cases, that the removal of a considerable portion of the non-antitoxic globulins from the serum by the Gibson method eliminates much of the deleterious matter from the serum, so that severe rashes, joint complications, fever, and other constitutional disturbances are less likely to occur from the antitoxic globulins than from the antitoxic serum from which they were obtained.

We asked ourselves the question whether the precipitated and refined serum is less toxic to sensitized guinea pigs than the untreated serum from which it was made. Doctor Park kindly furnished us some of the precipitated serum and the corresponding untreated serum from which it was made in order to carry out these tests.

^a Gibson, R. B. Journ. biolog. chemistry. Vol. 1, Nos. 2 and 3, 1906. ^b Park, William H., and Throne, Binford: The results of the use of "refined diphtheria antitoxin," Gibson's "globulin preparation," in the treatment of diphtheria. Trans. Assn. Am. Physicians, Vol. 21, 1906, pp. 259-267.

Comparative toxicity of untreated and refined antitoxic serum.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|-------------|---|-----------------------|
| 7491 | 0.24 c. c. toxine $9+\frac{1}{180}$ c. c. antitoxic horse serum (PD. 08004), subcutaneously. | Days. 25 | 1 c.* c. antitoxic horse serum, Gib- son precipitated (XYBH, 305-6), intraperitoneally. | Dead, 25 minutes. |
| 7480 | 0.24 c. c. toxine $9+\frac{1}{320}$ c. c. antitoxic horse serum (S. 68D), subcutaneously. | 25 | dodo | Dead, 30 minutes. |
| 7493 | 0.24 c. c. toxine $9+\frac{1}{300}$ c. c. antitoxic horse serum (PD. 08516), | 25 | do | Dead, 40 minutes. |
| 7489 | subcutaneously. 0.24 c. c. toxine $9+\frac{1}{120}$ c. c. antitoxic horse serum (PD. 08004), subcutaneously. | 25 | do | Dead, 42 minutes. |
| 7478 | 0.24 c. c. toxine $9+\frac{1}{180}$ c. c. antitoxic horse serum (S. 69H), subcutaneously. | 25 | 1 c. c. antitoxic horse serum (NYBH. 305-6), untreated. | Dead, 12 minutes. |
| 7490 | 0.24 c. c. toxine $9+\frac{1}{150}$ c. c. antitoxic horse serum (PD. 08004), subcutaneously. | 25 | dodo | Very severe symptoms. |
| 7477 | 0.24 c. c. toxine $9 + \frac{1}{150}$ c. c. antitoxic horse serum (S. 69H), subcutaneously. | 25 | do | Very severe symptoms. |
| 7505 | 0.24 c. c. toxine $9+\frac{1}{170}$ c. c. antitoxic horse serum (Hb. 27), subcutaneously. | . 25 | 2 c. c. antitoxic horse serum, Gib- son precipitated (NYBH, 305-6), subcutaneously. | Marked symptoms. |
| 7503 | 0.24 c. c. toxine $9+\frac{1}{300}$ c. c. antitoxic horse serum (Hb. 27), subcutaneously. | 25 | do | Severe symptoms. |
| 7502 | 0.24 c. c. toxine $9 + \frac{1}{230}$ c. c. antitoxic horse serum (Hb. 26), subcutaneously. | 25 | do | Marked symptoms. |
| 7504 | 0.24 c. c. toxine $9+\frac{1}{370}$ c. c. antitoxic horse serum (Hb. 26), subcutaneously. | 25 | 2 c. c. antitoxic horse serum (NYBH. 305-6), untreated. | Marked symptoms. |
| 7506 | 0.24 c. c. toxine $9+\frac{1}{220}$ c. c. antitoxic horse serum (Hb. 27), sub- | 25 | do | Severe symptoms. |
| 7507 | cutaneously. 0.24 c. c. toxine $9+\frac{1}{270}$ c. c. antitoxic horse serum (Hb. 27), subcutaneously. | 25 | do | Severe symptoms. |

Comparative toxicity of untreated and refined antitoxic serum.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|-----------|---|-----------------------|
| 7195 | 0.142 c. c. toxine $5+\frac{1}{940}$ c. c. antitoxic horse serum (NYBH. 305), subcutaneously. | Days. 41 | 1 c. c. antitoxic horse serum, Gib- son precipitated (NYBH. 305-6), subcutaneously. | Severe symptoms. |
| 7190 | 0.142 c. c. toxine 5+ 10000 c. c. antitoxic horse serum (NYBH. 305), subcutaneously. | 41 | do | Severe symptoms. |
| 7194 | 0.142 c. c. toxine $5 + \frac{1}{9 \cdot 50}$ c. c. antitoxic horse serum (NYBH. 305), subcutaneously. | 41 | do | Very severe symptoms. |
| 7189 | 0.142 c. c. toxine 5+ TOTAL C. c. antitoxic horse serum (NYBH. 305), subcutaneously. | 41 | 1 c. c. antitoxic horse serum (NYBH. 305-6), untreated. | Very severe symptoms. |
| 7193 | 0.142 c. c. toxine $5 + \frac{1}{960}$ c. c. antitoxic horse serum (NYBH. 305), subcutaneously. | 41 | | Very severe symptoms. |
| 7196 | 0.142 c. c. toxine $5 + \frac{1}{540}$ c. c. antitoxic horse serum (NYBH. 305), subcutaneously. | 41 | do | Very severe symptoms. |

We think it evident from the above that refined antitoxic serum precipitated and dialyzed in accordance with the Gibson method is quite as toxic, bulk for bulk, as the untreated serum from which it has been obtained. We must, however, consider that the treatment to which the serum is subjected in accordance with the Gibson method concentrates its antitoxic power about twice. There is, therefore, a distinct advantage gained, so far as bulk is concerned, in giving a corresponding number of antitoxic units; for, the serum reaction in children depends partly upon the quantity of serum given.



Part VII.

COMPARATIVE TOXICITY OF DIFFERENT HORSE SERUMS.

Besredka and Steinhardt^a believe that the French horse serums are much less toxic than those used by Otto^b and the serums used by us. Besredka and Steinhardt had a mortality of about 25 per cent when 5 c. c. of serum was injected intraperitoneally at the second injection, whereas Otto's and our percentage under similar conditions was much higher. Besredka kindly sent us a quantity of "serum antidiphterique" prepared at the Pasteur Institute and this serum was injected into a series of guinea pigs in order to compare its toxicity with the normal horse serum of our roan horse that we have used so much in these experiments.

Toxicity of "serum antidiphterique" (Pasteur Institute).

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|---|-------------|--|-------------------|
| 7768 | 0.142 c. c. toxine $5 + \frac{1}{320}$ c. c. antitoxic horse serum (S. spl. 1351), | Days. 76 | 5 c. c. antitoxic horse serum (Pas- teur Institute), | Dead, 11 minutes. |
| 7774 | subcutaneously. $0.142 \mathrm{c.c.}$ toxine $5+\frac{1}{350}$ c. c. antitoxic horse serum (S. spl. 1351), | 76 | intraperitoneally. | Dead, 10 minutes. |
| 7723 | subcutaneously. 0.142 c. c. toxine $5 + \frac{1}{5 \text{ fo}}$ c. c. antitoxic horse serum (PD. 08022), | 83 | do | Dead, 15 minutes. |
| 7726 | subcutaneously. 0.142 c. c. toxine $5+\frac{1}{580}$ c. c. antitoxic horse serum (PD. 08022), | 83 | do | Dead, 9 minutes. |
| 7849 | subcutaneously. 0.142 c. c. toxine $5 + \frac{1}{290}$ c. c. antitoxic horse serum (A. 192), sub- | 58 | do | Dead, 19 minutes. |
| 440 | cutaneously. 0.0006 gm. tetanus toxine $A + \frac{1}{300}$ c. c. antitoxic horse serum (Hoechst), subcu- | 56 | do | Dead, 10 minutes. |
| 451 | taneously. 0.0006 gm. tetanus toxine $A + \frac{1}{500}$ c. c. antitoxic horse serum (Parke), subcutaneously. | 56 | do | Dead, 10 minutes. |

^aBesredka, A., and Steinhardt, Edna: De l'anaphylaxie et de l'anti-anaphylaxie vis-à-vis du sérum de cheval. Ann. de l'Inst. Pasteur, Vol. 21, No. 2, Feb. 25, 1907, pp. 117–127.

b Otto, R.: Das Theobald Smithschen Phänomen der Serum-Ueberempfindlichkeit. Leuthold-Gedenkschr., Bd. 1, 1905.

Toxicity of normal horse serum (our roan).

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|---|-------------|--|-----------------------|
| 7845 | 0.24 c. c. toxine $9 + \frac{1}{310}$ c. c. antitoxic horse serum (A. 192), sub- | Days. 58 | 5 c. c. normal horse (roan) serum in- traperitoneally. | Dead, 20 minutes. |
| 7850 | cutaneously. 0.24 c. c. toxine $9+\frac{1}{290}$ c. c. antitoxic horse serum (A. 142), sub- | 58 | do | Dead, 18 minutes. |
| 7725 | c. c. antitoxic horse serum (PD. 08022), | 80 | do | Dead, 35 minutes. |
| 444 | subcutaneously. 0.0006 gm. tetanus toxine $A + \frac{1}{1200}$ c. c. antitoxic horse serum (Hoechst), subcutaneously. | 56 | do | Very severe symptoms. |
| 447 | 0.0006 gm. tetanus toxine $A + \frac{1}{300}$ c. c. antitoxic horse serum $(M.2122)$, subcutaneously. | 56 | do | Very severe symptoms. |

It is perfectly evident from the above that our results upon the comparative toxicity of the French and American serums do not agree with those reported by Besredka and Steinhardt. With us, the French serums are perhaps somewhat more toxic than our own. We believe these contradictory results are due to other causes than the relative toxicity of the different serums. It is not likely that these differences are due to varying susceptibility of the different breeds of guinea pigs. We have found little difference between guinea pigs obtained from five or six different sources. Further, we have sometimes been struck with the fact that guinea pigs from our own stock and raised under precisely similar conditions show striking differences of degree in the reaction to the second injection. For instance, all the guinea pigs sensitized with toxine-antitoxin mixtures upon a certain date will subsequently prove exceedingly sensitive, and most of them will die at the second injection, whereas another lot of guinea pigs similarly sensitized at another time will prove much less susceptible at the second injection. So far as we are able to judge, this difference of toxicity depends upon something connected with the sensitizing action and not with the variety of horse serum given at the second injection.

Note.—When toxine is mentioned in the tables diphtheria toxine is meant unless otherwise stated.

Part VIII.

THE IMMUNITY TO HYPERSUSCEPTIBILITY OR "ANTI-ANAPHYLAXIS,"

The immunity produced against the toxic action by repeated injections of horse serum has been called anti-anaphylaxis by Besredka and Steinhardt.^a From our subsequent work we learn that this immunity is relatively not quite as lasting and definite as many instances of active immunity seen in the laboratory against bacterial infections. Guinea pigs that have received a number of prior injections of horse serum may again show symptoms when reinjected with large amounts. The symptoms in such cases are usually mild, and death has never occurred in an "immunized" guinea pig as a result of subsequent injections with horse serum.

G. P. No. 410:

January 15, 1906, 1 c. c. antitoxic horse serum (Natl. VIII, 17), subcutaneously. No symptoms.

January 23, 1906, 1 c. c. antitoxic horse serum (Natl. VIII, 17), subcutaneously. No symptoms.

February 8, 1906, 6 c. c. antitoxic horse serum (Natl. VIII, 17), subcutaneously. No symptoms.

February 14, 1906, 6 c. c. antitoxic horse serum (Natl. VIII, 17), subcutaneously. No symptoms.

February 23, 1906, 6 c. c. antitoxic horse serum (Natl. VIII, 17), subcutaneously.

No symptoms.

March 29, 1906, 6 c. c. antitoxic horse serum (Natl. VIII, 18), subcutaneously.

No symptoms.

April 18, 1906, 6 c. c. antitoxic horse serum (Natl. VIII, 19), subcutaneously. No symptoms.

May 16, 1906, 6 c. c. normal horse (roan) serum, subcutaneously. No symptoms. May 18 to June 27, 1906, normal horse (roan) serum, 1 c. c. daily, subcutaneously.

September 7, 1906, 6 c. c. normal horse (roan) serum, intraperitoneally. No symptoms.

G. P. No. 430:

May 18 to June 27, 1906, 1 c. c. normal horse (roan) serum, daily, subcutaneously. No symptoms.

February 27, 1907, 6 c. c. normal horse (roan) serum, intraperitoneally. Severe symptoms.

G. P. No. 427:

May 18 to June 27, 1906, 1 c. c. normal horse (roan) serum, daily, subcutaneously. No symptoms.

February 27, 1907, 6 c. c. normal horse (roan) serum, intraperitoneally. Severe symptoms.

G. P. No. 4426:

January 10, 1906, 0.002 c. c. toxine No. 5, subcutaneously.

January 18, 1906, 6 c. c. normal horse (No. 15) serum, intraperitoneally. No symptom.

February 14, 1906, 6 c. c. antitoxic horse serum (Natl. VIII, 17), intraperitoneally. Symptoms (?).

February 23, 1906, 6 c. c. antitoxic horse serum (Natl. VIII, 17), intraperitoneally. No symptoms.

March 29, 1906, 6 c. c. antitoxic horse serum (Natl. VIII, 17), intraperitoneally. Mild symptoms.

April 18, 1906, 6 c. c. antitoxic horse serum (Natl. VIII, 18), intraperitoneally. No symptoms.

May 16, 1906, 6 c. c. normal horse (roan) serum, intraperitoneally. Severe symptoms.

May 18 to June 27, 1906, 1 c. c. normal horse (roan) serum, intraperitoneally, daily, except Sunday; 22 injections. No symptoms.

January 25, 1907, 6 c. c. normal horse (No. 15) serum, intraperitoneally. No symptoms.

March 26, 1907, 6 c. c. normal horse (roan) serum, intraperitoneally. No symptoms.

For other instances of this variation of susceptibility see Part IX, pages 59 to 62, Bulletin 29, Hygienic Laboratory, United States Public Health and Marine-Hospital Service.

Part IX.

MATERNAL TRANSMISSION OF HYPERSUSCEPTIBILITY AND IMMUNITY.

In our previous work we showed that hypersusceptibility to the toxic effects of horse serum may be transmitted from the mother guinea pig to her young. Later, one of us (Anderson) showed that the female guinea pig may transmit hypersusceptibility to horse serum and immunity to diphtheria toxine at the same time. On account of certain analogies between the reaction to tuberculin and the toxic action of horse serum, we have made further studies along these lines. In this bulletin we shall refer only to our studies upon the transmission of hypersusceptibility and immunity to the toxic action of horse serum, leaving related studies with tuberculosis and tuberculin for a future publication.

Our present studies corroborate the fact that hypersusceptibility to the toxic action of horse serum is always transmitted from the mother guinea pig to her young. This function is solely maternal; the male takes no part whatever in the transmission of these acquired properties. Whether this maternal transmission is hereditary or congenital can not be definitely stated.

We are able to exclude the milk as a factor in transmitting the hypersusceptibility to the toxic action of horse serum by a series of

exchange experiments, which are given in detail below.

"Exchange" experiments consist in at once placing guinea pigs born of a susceptible mother to nurse with an untreated female and, in exchange, the young of the untreated female are at the same time placed to nurse with the susceptible female. From these "exchange" experiments we learn that the hypersusceptibility is not transmitted to the young in the milk.

We also learn from our experiments that hypersusceptibility may be transmitted from mother to young whether the mother is sensitized before or after conception. The fact that this influence may take place after conception might be taken to indicate that the transmission

is congenital and not hereditary.

GROUP A.

FAMILY NO. 1.

(Sensitized female; untreated male.)

Female (G. P. No. 610). October 20, 1906. Six c. c. antitoxic horse serum. (Natl. IX, 17) intraperitoneally. Dead, 30 minutes.

[Previous treatment: 151 days prior, 0.15 c. c. toxine No. $9+\frac{1}{250}$ c. c. antitoxic

horse serum (Natl. VIII, 18).]

- Male (G. P. No. 102). June 8, 1906. Untreated. Put in cage with above female after female was sensitized.
- Offspring. August 24, 1906. Four young born. Two tested as follows:
 - G. P. No. 610a. October 20. 59 days old. Three c. c. antitoxic horse serum (Natl. IX, 17) intraperitoneally. Dead. 4 minutes.
 - (G. P. No. 610b.) Ditto. Dead, 60 minutes.

FAMILY NO. 2.

(Sensitized female; untreated male.)

Female (G. P. No. 612). January 25, 1907. Six c. c. normal horse serum (horse No. 15) intraperitoneally. Very severe symptoms.

[Previous treatment: 245 days prior, 0.15 c. c. toxine $9+\frac{1}{230}$ c. c. antitoxic horse serum (Natl. VIII, 18).]

- Male (untreated). June 8, 1906. Placed in cage with above female after she was sensitized.
- Offspring. One young born January 10, 1907, and tested as follows:
 - (G. P. No. 612a.) January 25. 15 days old. Two c. c. horse serum (horse No. 15) intraperitoneally. Marked symptoms.

FAMILY NO. 3.

(Sensitized female; untreated male.)

Female (G. P. No. 611). September 7, 1906. Six c. c. normal horse serum (roan) intraperitoneally. Dead, 120 minutes.

[Previous treatment: 106 days prior, received 0.15 c. c. toxine $9+\frac{1}{250}$ c. c. antitoxic horse serum (Natl. VIII, 18).]

- Male (untreated). Paired 17 days after treatment of female.
- Offspring. One young. Tested as follows:
 - (G. P. No. 611a.) September 7, 1906. 2 days old. One c. c. normal horse serum (roan) intraperitoneally. Dead, 20 minutes.

FAMILY NO. 4.

(Sensitized female; untreated male. Exchange.)

Female (G. P. No. 613). February 27, 1907. Six c. c. normal horse (roan) serum intraperitoneally. Very severe symptoms.

[Previous treatment: 282 days prior 0.15 c. c. toxine $9 + \frac{1}{230}$ c. c. antitoxic horse serum (Natl. VIII, 18).]

- Male (G. P. No. 10x). June 8, 1906. Untreated. Put into cage 18 days after female was sensitized.
- Offspring. January 22, 1907. Two young born and at once put to nurse with untreated female. Tested as follows:
 - (G. P. No. 613a.) February 27, 1907. 36 days old. Two c. c. normal (roan) horse serum intraperitoneally. Very severe symptoms.
 - (G. P. No. 613b.) Ditto. Dead, 45 minutes.

Two young, born of untreated female, put to nurse with above sensitized female (No. 613). Tested as follows:

- (G. P. No. P-68a.) February 27, 1907. 36 days old. Two c. c. normal horse (roan) serum intraperitoneally. No symptoms.
 - (G. P. No. P-68b.) Ditto. No symptoms.

FAMILY NO. 5.

(Sensitized female; untreated male. Exchange.)

Female (G. P. No. 614). March 26, 1907. Six c. c. normal horse (roan) serum intraperitoneally. Dead, 38 minutes.

[Previous treatment; 309 days prior, 0.15 c. c. toxine No. $9+\frac{1}{250}$ c. c. antitoxic horse serum (Natl. VIII, 18).]

- Male (G. P. No. 10). June 8, 1906. Untreated. Put in cage after female was sensitized.
- Offspring. February 17, 1907. Two young born, at once put to nurse with untreated female, and tested as follows:
 - (G. P. No. 614a). March 26, 1907. 38 days old. Two c. c. normal horse (roan) serum intraperitoneally. Dead, 25 minutes.
 - (G. P. No. 614b). Ditto. Dead, 20 minutes.

Two young, born of untreated female and nursed with above sensitized female (614), tested as follows:

- (G. P. No. P-20). March 26, 1907. 38 days old. Two c. c. normal horse (roan) serum intraperitoneally. No symptoms.
 - (G. P. No. P-20a). Ditto. No symptoms.

GROUP B.

FAMILY NO. 6.

(Sensitized female; sensitized male. Exchange.)

Female (G. P. No. 601). October 23, 1906. Six c. c. antitoxic horse serum (Natl. IX, 18) intraperitoneally. Dead, 15 minutes.

[Previous treatment: 153 days prior, 0.15 c. c. toxine No. $9 + \frac{1}{250}$ c. c. antitoxic horse serum (Natl. VIII, 18).]

Male (G. P. No. 606x). Paired after both were sensitized. September 7, 1906. Six c. c. normal horse (roan) serum intraperitoneally. Dead, 17 minutes.

[Previous treatment: 106 days prior, 0.15 c. c. toxine No. $9 + \frac{1}{2\frac{1}{3}0}$ c. c. antitoxic horse serum (Natl. VIII, 18).]

- Offspring. Two young, born August 28, 1906. Immediately nursed with an untreated female.
 - (G. P. No. 601a.) October 23, 1906. 56 days old. Three c. c. antitoxic horse serum (Natl. IX, 18) intraperitoneally. Severe symptoms.
 - (G. P. No. 601b.) Ditto. Severe symptoms.
 - Three untreated young pigs placed to nurse with the above mother (No. 601).
 - (G. P. No. X.) October 23. 56 days old. Three c. c. antitoxic horse serum intraperitoneally. No symptoms.
 - (G. P. No. Y.) Ditto. No symptoms.
 - (G. P. No. Z.) Ditto. No symptoms.
 - (G. P. mother of X, Y, and Z.) October 23. Six c. c. same serum intraperitoneally. No symptoms.

FAMILY NO. 7.

(Sensitized female; sensitized male. Exchange.)

Female (G. P. No. 603). February 27, 1907. Six c. c. normal horse (roan) serum intraperitoneally. Dead, 45 minutes.

[Previous treatment: 282 days prior, 0.15 c. c. toxine No. $9 + \frac{1}{2\sqrt{30}}$ c. c. antitoxic horse serum (Natl. VIII, 18).]

Male (G. P. No. 606y). June 8, 1906. Placed in cage with above female after both were sensitized.

Offspring. January 17, 1907. Two young born, nursed with untreated female, and tested as follows:

(G. P. No. 603a.) February 27, 1907. 41 days old. Two c. c. normal horse (roan) serum intraperitoneally. Marked symptoms.

(G. P. No. 603b.) Ditto. Very severe symptoms.

Three young, born of untreated female and nursed with above sensitized female (603), tested as follows:

(G. P. No. 66a.) February 27, 1907. 41 days old. Two c. c. normal horse (roan) serum intraperiton cally. No symptoms.

(G. P. No. 66b.) Ditto. No symptoms.

(G. P. No. 66c.) Ditto. No symptoms.

GROUP C.

FAMILY NO. 8.

(Immune female; immune male.)

Female (G. P. No. 428). January 25, 1907. Six c. c. normal horse serum (horse No. 15) intraperitoneally. Slight symptoms.

[Previous treatment: May 18 to June 27, 1906, 1 c. c. normal horse (roan) serum daily (except Sunday), subcutaneously, 22 injections.]

Male (G. P. No. 4530). June 8, 1906. Immune. Placed in cage with above female. For history of this male see Family No. 14.

Offsping. December 28, 1906. Two young born and nursed with own mother. Tested as follows:

(G. P. No. 428a.) January 25, 1907. 28 days old. Two c. c. normal horse serum (horse No. 15) intraperitoneally. Marked symptoms.

(G. P. No. 428b.) Ditto. Marked symptoms.

FAMILY NO. 9.

(Immune female; immune male. Exchange.)

Female (G. P. No. 4426). January 10, 1906. 0.002 c. c. toxine 7. Severe reaction, slough.

January 18, 1906. Six c. c. normal horse serum (horse No. 15) intraperitoneally. No symptoms.

February 14, 1906. Six c. c. antitoxic horse serum (Natl. VIII, 17) intraperitoneally. Symptoms.

February 23, 1906. Ditto. No symptoms.

March 29, 1906. Ditto. Mild symptoms.

April 18, 1906. Ditto (Natl. VIII, 18). No symptoms.

May 16, 1906. Six c. c. normal (roan) horse serum intraperitoneally. Severe symptoms.

May 18 to June 27, 1906. One c. c. normal horse (roan) serum subcutaneously daily (except Sunday), 22 injections. No symptoms.

January 25, 1907. Six c. c. normal (No. 15). No symptoms.

March 26, 1907. Six c. c. normal (roan). No symptoms.

Male (G. P. No. 4530). June 8, 1906. Immune. Placed in cage with above female. Previous treatment:

January 12, 1906. 0.19 c. c. toxine 7+1 unit antitoxic horse serum (B27).

January 22, 1906. Six c. c. antitoxic horse serum (Natl. VIII, 17) intraperitoneally. Definite symptoms.

February 8, 1906. Ditto. Symptoms. February 8, 1906. Ditto. No symptoms.

February 14, 1906. Ditto. No symptoms.

February 23, 1906. Ditto. No symptoms.

March 25, 1906. Ditto (Natl. VIII, 18). No symptoms.

April 18, 1906. Ditto. Ditto. No symptoms.

May 16, 1906. Six c. c. normal horse (roan) serum intraperitoneally. No symptoms.

May 18 to June 27, 1906. One c. c. same serum subcutaneously daily (except

Sunday), 22 injections. No symptoms.

January 25, 1907. Six c. c. normal horse serum (horse No. 15). Very slight symptoms.

Offspring, first litter. One young born December 26, 1906. As soon as born, placed to nurse with untreated female (pen 120), whose young were placed to nurse with G. P. No. 4426.

(G. P. No. 4426a.) January 25, 1907. Two c. c. normal horse (No. 15) serum intraperitoneally, when 30 days old. No symptoms.

Young of untreated female P. 120.

(G. P. No. P. 120a.) January 25, 1907. 35 days old. Two c. c. normal horse (No. 15) serum intraperitoneally. No symptoms.

February 18, 1907. 24 days later. Six c. c. same serum. Very severe symptoms.

(G. P. No. P. 120b.) January 25, 1907. Same as G. P. No. P. 120a. No. symptoms.

February 18, 1907. Ditto. Dead, 70 minutes.

Second litter. Three young born March 1, 1907, and not exchanged.

(G. P. No. 4426b.) March 26, 1907. 25 days old. Two c. c. normal horse (roan) serum intraperitoneally. No symptoms.

(G. P. No. 4426c.) Same treatment. No symptoms.

(G. P. No. 4426d.) Same treatment. No symptoms.

These three pigs were again tested April 18 with 4 c. c. of normal horse (roan) serum into the peritoneal cavity. All three showed slight symptoms, thus proving that immunity, or "antianaphylaxis," was not transmitted from the mother.

FAMILY NO. 10.

(Immune female; immune male. Exchange.)

Female (G. P. No. 429). January 25, 1907. Six c. c. normal horse (No. 15) serum intraperitoneally. Slight symptoms.

[Previous treatment: May 18 to June 27, 1906, 1 c. c. normal horse (roan) serum

daily except Sunday, 22 injections.]

Male (G. P. No. 4530). June 8, 1906. Placed in cage with female. For history of this male, see Family No. 14.

Offspring. Three young, born December 29, 1906, and nursed with untreated G. P. No. P. 50. Tested as follows:

(G. P. No. 429a.) January 25, 1907. 27 days old. Two c. c. normal horse serum (horse No. 15) intraperitoneally. Marked symptoms.

Ditto. Marked symptoms. (G. P. No. 429b.)

Marked symptoms. (G. P. No. 429c.) Ditto.

Two young of untreated G. P. No. P. 50 nursed with immune female No. 429, and tested as follows:

(G. P. No. P. 50a.) January 25, 1907. 27 days old. Two c. c. normal horse serum (horse No. 15) intraperitoneally. No symptoms.

(G. P. No. P. 50b.) Ditto. No symptoms.

GROUP D.

FAMILY NO. 11.

(Immune female; untreated male.)

Female (G. P. No. 410). January 15, 1906. One c. c. antitoxic horse serum (Natl. VIII, 17). No symptoms.

January 23, 1906. Ditto. No symptoms.

February 8, 1906. Six c. c. same serum. No symptoms.

Ditto. February 14, 1906. No symptoms. Ditto. February 23, 1906. No symptoms.

March 29, 1906. Six c. c. antitoxic horse serum (Natl. VIII, 18). No symptoms.

April 18, 1906. Ditto. No symptoms.

May 16, 1906. Six c. c. normal horse (roan) serum. No symptoms.

May 18 to June 27, 1906, daily. One c. c. same serum. No symptoms.

September 7, 1906. Six c. c. normal horse (roan) serum intraperitoneally. No symptoms.

All injections subcutaneously.

Male (G. P. No. 410m). June 8, 1906. Untreated. Put in cage after female was immunized.

Offspring. July 20, 1906. Three young born, and tested as follows:

(G. P. No. 410a.) September 7, 1906. 18 days old. Two c. c. normal horse (roan) serum intraperitoneally. No symptoms.

(G. P. No. 410b.) Ditto. No symptoms. (G. P. No. 410c.) Ditto. No symptoms.

FAMILY NO. 12.

(Immune female; untreated male.)

Female (G. P. No. 430). February 27, 1907. Six c. c. normal horse (roan) serum intraperitoneally. Severe symptoms.

[Previous treatment: 245 days prior (May 18 to June 27). One c. c. normal horse (roan) serum subcutaneously daily.]

Male (G. P. No. -). June 8, 1906. Untreated. Put in cage during period of immunization.

Offspring. January 22, 1907. Two young born and nursed with mother. Tested as follows:

(G. P. No. 430a). February 27, 1907. 36 days old. Two c. c. normal horse (roan) serum intraperitoneally. Marked symptoms.

Ditto. (G. P. No. 430b.) Marked symptoms.

(G. P. No. 430c.) Mild symptoms.

FAMILY NO. 13.

(Immune female; untreated male.)

Female (G. P. No. 110). February 8 to 20, 1906. One c. c. Daily subcutaneous injections 1 c. c. antitoxic horse serum (Natl. IX, 19). No symptoms.

March 29, 1906. Six c. c. antitoxic horse serum (Natl. VIII, 18). Mild symptoms.

April 4 to 17, 1906. One c. c. daily injections antitoxic horse serum (Natl. IX. 17). No symptoms.

April 23, 1906. Six c. c. normal horse (roan) serum. Very slight symptoms.

May 15, 1906. Ditto. Very slight symptoms.

May 18 to June 27, 1906. One c. c. daily, same serum. No symptoms.

September 7, 1906. Six c. c. normal horse (roan) serum, intraperitoneally. Very slight symptoms.

Male (G. P. No. —). June 8, 1906. Untreated. Put in cage after immunization of female was well advanced.

Offspring. August 10, 1906. One young born, and tested as follows:

(G. P. No. 110a.) September 7, 1906. 27 days old. Two c. c. normal horse (roan) serum intraperitoneally. Very slight symptoms.

FAMILY NO. 14.

(Immune female; untreated male.)

Female (G. P. No. 426). September 7, 1906. Six c. c. normal horse (roan) serum, intraperitoneally. Slight symptoms.

[Previous treatment: May 19 to June 27, 1906, 1 c. c. daily subcutaneous injections of normal horse (roan) serum.]

Male (G. P. No. 426m). June 8, 1906. Untreated. Put in cage during period of immunization of female.

Offspring. July 24, 1906. Two young born, and tested as follows:

(G. P. No. 426a.) September 7, 1906. 41 days old. Two c. c. normal horse (roan) serum, intraperitoneally. Slight symptoms.

(G. P. No. 426b.) Ditto. Slight symptoms.

FAMILY NO. 15.

(Immune female; untreated male. Exchange.)

Female (G. P. No. 427). February 27, 1907. Six c. c. normal horse (roan) serum intraperitoneally. Severe symptoms.

[Previous treatment: 245 days prior (May 18 to June 26) 1 c. c. normal horse (roan) serum subcutaneously daily.] No symptoms.

Male (G. P. No. 42601). June 8, 1906. Normal. Put into cage during period of immunization of female.

Offspring. January 22, 1907. Two young born. At once placed to nurse with untreated female. Tested as follows:

(G. P. No. 427a.) February 27, 1907. 36 days old. Three c. c. normal horse (roan) serum intraperitoneally. Severe symptoms. (G. P. No. 427b.) Ditto. Dead, 62 minutes.

Two young born of untreated female, but nursed with above female No. 427, tested as follows:

(G. P. No. P. 108a.) February 27, 1907. 36 days old. Three c. c. normal horse (roan) serum intraperitoneally. No symptoms.

(G. P. No. P. 108b.) Ditto. No symptoms.

GROUP E.

FAMILY NO. 16.

(Untreated female; sensitized male.)

Female (G. P. No. 606). September 7, 1906. Six c. c. horse serum intraperitoneally. No prior treatment. No symptoms.

Male (G. P. No. 606x). September 9, 1906. Six c. c. horse serum (roan) intraperitoneally. Dead, 17 minutes.

[Previous treatment: 106 days prior received 0.15 c. c. tox. $9+\frac{1}{250}$ c. c. antitoxic horse serum (Natl. VIII, 18).]

Offspring. Two young born August 4, 1906, and tested as follows:

(G. P. No. 606a.) September 9, 1906. 1 month old. Two c. c. normal horse serum into peritoneum. No symptoms.

(G. P. No. 606b.) September 9, 1906. 1 month old. Two c. c. normal horse serum (roan) into peritoneum. No symptoms.

FAMILY NO. 17.

(Untreated female; sensitized male.)

Females (G. P. No. 609). January 25, 1907. Six c. c. normal horse serum (horse No. 15), intraperitoneally. No prior treatment. No symptoms.

(G. P. No. 607). Ditto. No prior treatment. No symptoms.

Male (G. P. No. 4527). January 25, 1907. Six c. c. normal horse serum (horse No. 15), intraperitoneally. Dead, 37 minutes.

[Previous treatment: 1 year 13 days prior, 0.19 c. c. toxine No. 7+1 unit B27.] June 8, 1906. Placed with untreated females Nos. 609 and 607.

Offspring. December 26, 1906. Three young born to G. P. No. 609. September 6, 1906. One young born to G. P. No. 607.

Young tested as follows:

(G. P. No. 609a). January 25, 1907. 30 days old. Two c. c. normal horse serum (horse No. 15) intraperitoneally. No symptoms.

(G. P. No. 609b). Ditto. No symptoms. (G. P. No. 609c). Ditto. No symptoms.

(G. P. No. 607a). Ditto (141 days old). No symptoms.

We conclude from these experiments that—

- 1. Hypersusceptibility to the toxic action of horse serum is transmitted through the female guinea pig; the male has no influence.
 - 2. The susceptibility is not transmitted through the milk.
- 3. Maternal transmission of hypersusceptibility succeeds, whether the female guinea pig is sensitized before or after conception.

Part X.

THE RELATION OF HYPERSUSCEPTIBILITY TO VARIOUS INFLUENCES.

We have already shown that hypersusceptibility to the action of horse serum in the guinea pig has no evident relation to hemolysis or precipitins. We offer the following experiments, planned with the object of correlating hypersusceptibility with other phenomena:

RELATION TO AGGRESSINES.

The work of Bail upon aggressine induced us to try whether a similar action may explain hypersusceptibility. The following experiments indicate that no relation exists between the two phenomena.

PERITONEAL FLUID FROM NORMAL GUINEA PIG INTO SENSITIZED GUINEA PIG.

Eight c. c. of normal horse (No. 15) serum was injected into the peritoneal cavity of a normal guinea pig. This produced no symptom. Two hours later the animal was chloroformed and about 6 c. c. of fluid was taken from the peritoneal cavity and injected into the following sensitized guinea pig:

G. P. No. 7050. Six c. c. of above fluid into peritoneal cavity. Dead, 20 minutes. [Previous treatment: 57 days prior, 0.142 c. c. toxine No. $5+\frac{1}{200}$ c. c. antitoxic horse serum (S. 063H), subcutaneously.]

PERITONEAL FLUID FROM SENSITIZED GUINEA PIG INTO NORMAL GUINEA PIG.

Six c. c. of normal horse (No. 15) serum was injected into the peritoneal cavity of G. P. No. 7051, which had been sensitized fifty-seven days previously with 0.142 c. c. toxine No. $5+\frac{1}{250}$ c. c. axtitoxic horse serum (S. 063H). The guinea pig developed typical symptoms and died in fifteen minutes as a result of the second injection. Fifteen minutes after the death of this guinea pig about 4 c. c. of the peritoneal contents were withdrawn and injected into the peritoneal cavity of a normal guinea pig. This caused no symptoms.

PERITONEAL FLUID FROM SENSITIZED GUINEA PIG INTO SENSITIZED GUINEA PIG.

Six c. c. normal horse (No. 15) serum was injected into the peritoneal cavity of a sensitized guinea pig (No. 7068) which had received fifty-five days previously a subcutaneous injection of 0.142 c. c. toxine No. $5 + \frac{1}{190}$ c. c. antitoxic horse serum (Natl. V, 7). As a result of the second injection of horse serum the guinea pig had characteristic symptoms and died in thirty minutes. Fifteen minutes after the

death of this guinea pig about 3 c. c. of the peritoneal contents were collected and injected into the following sensitized guinea pig:

G. P. No. 7026. Three c. c. of above fluid into peritoneal cavity. Very severe symptoms.

[Previous treatment: 57 days prior, 0.142 c. c. toxine No. $5+\frac{1}{290}$ c. c. antitoxic

horse serum (Ld. 8), subcutaneously.

PERITONEAL FLUID FROM SENSITIZED GUINEA PIGS INTO NORMAL GUINEA PIGS.

The peritoneal contents were collected immediately after the death from 9 sensitized guinea pigs that had received 6 c. c. normal horse serum (No. 15) each; 3.5 c. c. of this fluid was injected into the peritoneal cavity of a normal guinea pig, but produced no symptoms.

After withdrawing the fluid of the peritoneal cavities of the above 9 guinea pigs the peritoneal cavities were washed with sterile salt solution and this fluid injected into the following normal guinea pigs:

G. P. No. A.—6 c. c. saline washings from above 9 guinea pigs, intraperitoneally. No symptoms.

> B.—5 c. c. saline washings from above 9 guinea pigs, intraperitoneally. No symptoms.

C.—7.5 c. c. saline washings from above 9 guinea pigs, intraperitoneally. No symptoms.

D.—10 c. c. saline washings from above 9 guinea pigs, intraperitoneally. No symptoms.

E.—20 c. c. saline washings from above 9 guinea pigs, intraperitoneally. No symptoms.

F.—6 c. c. saline washings from above 9 guinea pigs, intraperitoneally. No symptoms.

G.—6 c. c. saline washings from above 9 guinea pigs, intraperitoneally. No symptoms.

RELATION TO METHEMAGLOBIN POISONING.

The symptoms in the guinea pig somewhat resemble methemaglobin poisoning. We are indebted to Assistant Surgeon A. M. Stimson for comparative spectroscopic studies of the blood of normal guinea pigs and of the blood of susceptible guinea pigs immediately after death caused by a second injection of horse serum. No methemaglobin was found. Only the bands corresponding to oxyhemaglobin were seen in the blood of the guinea pigs examined immediately after death.

OXYGEN HAS NO INFLUENCE UPON THE SYMPTOMS.

A sensitized guinea pig was inoculated with horse serum and at once placed in an almost pure atmosphere of oxygen. Another sensitized guinea pig (not reinoculated) was placed under the same bell jar as a control. The inoculated pig developed symptoms and was dead in thirty-five minutes. The control animal showed no unusual manifestations after thirty minutes in the atmosphere of oxygen. It was then given an injection of horse serum and immediately replaced under the

bell jar. It developed characteristic symptoms and died in fifteen minutes.

G. P. No. 7028. Six c. c. antitoxic horse serum (Natl. IX, 17), intraperitoneally; immediately placed in an atmosphere of almost pure oxygen. Dead, 35 minutes. [Previous treatment: 54 days prior, 0.142 c. c. toxine No. $5+\frac{1}{470}$ c. c. antitoxic horse serum (Led. 8).]

Control G. P. Kept 30 minutes in same atmosphere. No symptoms.

[Previous treatment: 52 days prior, 0.142 c. c. toxine No. $5+\frac{1}{240}$ c. c. antitoxic horse serum (PDCo. 07555A).]

At the end of 30 minutes in oxygen atmosphere, given 6 c. c. antitoxic horse serum (Natl. IX, 17), intraperitoneally. Dead, 15 minutes.

THE INFLUENCE OF DIPHTHERIA TOXINE UPON HYPERSUSCEPTIBILITY.

The question was raised by both Otto and us as to the influence of the diphtheria toxine in accentuating the phenomenon of hypersusceptibility.

While guinea pigs may be sensitized with fresh normal horse serum alone, it seemed to us and also to Otto that a greater degree of hypersusceptibility is produced when sensitized with a mixture of diphtheria toxine and antitoxic horse serum than when the horse serum alone is given at the first injection. It seems, however, that this is by no means always the case.

All the guinea pigs in the following series were sensitized with $_{250}$ c. c. of antitoxic horse serum (Alex. A., 228) precipitated and dialyzed according to Gibson's method. One-half the animals received, in addition, 0.2 c. c. toxine No. 7 (MLD = 0.006). After thirty-one days interval all the guinea pigs were tested with 3 c. c. normal horse serum (roan) into the peritoneal cavity.

[All tested by injecting 3 c. c. normal horse (No. 15) serum intraperitoneally.]

| No. G. P. | Previous treatment, 31 days prior. | Result. |
|---|---|--|
| 720 721 722 723 724 725 726 727 728 729 710 711 712 713 714 715 716 717 718 | 2150 c. c. precipitated antitoxic horse serum (Alex. A., 228), subcutaneously do d | Dead, 28 minutes. Dead, 12 minutes. Very severe symptoms. Dead, 44 minutes. Dead, 10 minutes. Very severe symptoms. Dead, 10 minutes. Dead, 10 minutes. Dead, 31 minutes. Dead, 31 minutes. Very severe symptoms. Dead, 27 minutes. Severe symptoms. Very severe symptoms. Dead, 27 minutes. Severe symptoms. Very severe symptoms. Dead, 92 minutes. |

THE INFLUENCE OF TETANUS TOXINE UPON HYPERSUSCEPTIBILITY.

Besredka and Steinhart^a intimate that guinea pigs sensitized with a mixture of tetanus toxine and antitetanic serum are not sensitive to subsequent injections of horse serum. These scientists, however, suggest that their failures on this point may have been due to the small amount of horse serum used at the first injection, viz, $\frac{1}{100000}$ and $\frac{1}{1000000}$ c. c.

We tested some of our used tetarus guinea pigs to determine this point and found that tetarus toxine does not apparently influence the phenomenon of hypersusceptibility to horse serum. The guinea pigs were sensitized with 0.0006 gm. of a dried tetarus toxine, which represents 100 minimal lethal doses, plus various amounts of antitetanic serum. All those tested reacted to a second injection of horse serum.

Tetanus toxine.

| No. G. P. | First injection. | Interval. | Second injection. | Result. • |
|--------------|---|-------------|---|-----------------------|
| 438 | 0.0006 gm. tetanus toxine $A + \frac{1}{600}$ c. c. antitoxic horse serum (Hoechst) subcuta- | Days. 17 | 6 c. c. normal horse (roan) serum, in- traperitoneally. | Dead, 30 minutes. |
| 439 | neously. 0.0006 gm. tetanus toxine $A + \frac{1}{700}$ c. c. antitoxic horse serum (Hoechst) subcuta- | 17 | do | Dead, 30 minutes. |
| 444 | neously. 0.0006 gm. tetanus toxine $A + \frac{1}{1200}$ c. c. antitoxic horse serum (Hoechst) subcutaneously. | 28 | do | Very severe symptoms. |
| 447 | 0.0006 gm. tetanus toxine $A + \frac{1}{500}$ (M 2122) c. c. antitoxic horse serum subcutaneously. | 28 | do | Very severe symptoms. |
| 451 | 0.0006 gm. tetanus tox- ine $A + \frac{1}{500}$ (Pk) c. c. antitoxic horse serum subcutaneously. | 28 | 6 c. c. normal horse (roan) serum, in- traperitoneally (Pasteur Insti- tute). | Dead, 10 minutes. |
| 440 | 0.0006 gm, tetanus toxine $A + \frac{1}{8\sqrt{10}}$ (Hoechst) c. c. antitoxic horse serum subcutaneously. | 28 | do | Dead, 10 minutes. |

THE RELATION OF THE SPLEEN AND THYROID TO HYPERSUSCEPTIBILITY.

While we believe that this reaction is probably localized in the central nervous system, several experiments were undertaken to determine what influence the spleen or the thyroid gland may have upon the hypersusceptibility produced by the injection of horse serum.

The removal of the thyroid gland or the spleen either before or after an injection of horse serum does not prevent the phenomenon of hypersusceptibility.

It is of interest to note that the guinea pigs upon which splenectomy was performed lost much hair and became reduced in weight, although the appetite seemed to remain good.

THYROIDECTOMY.

G. P. No. 7017. 0.142 c. c. toxine No. $5+\frac{1}{100}$ c. c. antitoxic horse serum (Mem. A1103), subcutaneously.

55 days later, thyroid removed.

80 days from sensitizing inoculation and 25 days after thyroidectomy, given 6 c. c. normal horse (No. 15) serum, subcutaneously. Dead, 10 minutes.

G. P. No. 7018. O. 142 c. c. toxine No. $5+\frac{1}{250}$ c. c. antitoxic horse serum (Mem. 1103).

55 days later, thyroid removed.

125 days from sensitizing inoculation and 70 days after thyroidectomy, given 6 c. c. normal horse (No. 15) serum, intraperitoneally. Dead, 30 minutes.

G. P. No. X. Untreated. Thyroid removed.

11 days after thyroidectomy, given 0.15 c. c. toxine No. $9 + \frac{1}{300}$ c. c. antitoxic horse serum (PD., 08516), subcutaneously.

17 days after sensitizing inoculation and 28 days after thyroidectomy, given 6 c. c. normal horse (No. 15) serum, intraperitoneally. Dead, 28 minutes.

G. P. No. 425W. Untreated. Thyroid removed.

11 days later, 0.15 c. c. toxine No. $9+\frac{1}{300}$ c. c. antitoxic horse serum (PD .08516).

69 days after sensitizing inoculation and 58 days after thyroidectomy, 6 c. c. normal horse (No. 15) serum, intraperitoneally. Dead, 5 minutes.

SPLENECTOMY.

G. P. No. 7044. 0.142 c. c. toxine No. $5+\frac{1}{520}$ c. c. antitoxic horse serum (Hb. 21A), subcutaneously.

55 days later, spleen removed.

80 days from sensitizing inoculation and 25 days after splenectomy, 6 c. c. normal horse (No. 15) serum, intraperitoneally. Very severe symptoms.

G. P. No. 7024. 0.142 c. c. toxine No. $5+\frac{1}{280}$ c. c. antitoxic horse serum (Mem 1103), subcutaneously.

55 days later, spleen removed.

80 days from sensitizing inoculation and 25 days after splenectomy, 6 c. c. normal horse (No. 15) serum, intraperitoneally. Very severe symptoms.

G. P. No. 380W. Spleen removed.

69 days later, 6 c. c. normal horse (No. 15) serum, intraperitoneally. No symptoms.

G. P. No. XY. Spleen removed.

11 days later, 0.15 c. c. toxine No. $9+\frac{1}{300}$ c. c. antitoxic horse serum (PD. 08516), subcutaneously.

28 days after splenectomy and 17 days after sensitizing, 6 c. c. normal horse (No. 15) serum, intraperitoneally. Severe symptoms.

G. P. No. 410W. Spleen removed.

11 days later, $0.\overline{15}$ c. c. toxine No. $9+\frac{1}{300}$ c. c. antitoxic horse serum (PD. 08516), subcutaneously.

58 days after sensitizing and 69 days after splenectomy, 6 c. c. normal horse (No. 15) serum, intraperitoneally. Marked symptoms.



Part XI.

MISCELLANEOUS.

FEEDING EXPERIMENTS WITH COOKED MEAT.

In our former work we showed that guinea pigs may be sensitized by feeding them blood serum or meat. We know that heating blood serum to 100° C. for fifteen minutes is sufficient to destroy its toxic action. We then asked ourselves the question, Would the heating of meat prevent the sensitizing action? The following experiments indicate that heat does destroy this property of meat, as far as the guinea pig is concerned.

In these experiments well-cooked horse meat was used. It was heated in the hot air sterilizer at 110° C. for thirty minutes. Two grams of it was fed to the animals daily from June 15 to June 30. Each guinea pig, therefore, received 32 gm. of the cooked meat. None of them showed symptoms when injected eighteen days later with 6 c. c. of normal horse serum.

| No. G. P. | First treatment. | Interval. | Second treatment. | Result. |
|--------------------------|---|----------------------|---|---|
| 496 | Fed 2 gm. horse meat, heated to 110° C. (hot air) 30 minutes, daily | Days. 18 | 6 c. c. normal horse (roan) serum, intraperitoneally. | No symptoms. |
| 497 498 499 500 | for 16 days. do | 18 18 18 18 | dododododododododododododo | No symptoms. No symptoms. No symptoms. No symptoms. |

FEEDING EXPERIMENTS WITH RAW BEEF.

We know that guinea pigs fed with horse serum or horse meat are susceptible to subsequent injections of horse serum, and we are now able to show, by the following series of experiments, that guinea pigs fed with beef are susceptible to subsequent injections of cattle serum:

| No. G. P. | First treatment. | Interval. | Second treatment. | Result. |
|--------------|---|-----------|--|--|
| 502 | Fed 2 gm. dried beef daily for 23 days do | 19 | 6 c. c. cattle serum, intraperitoneally. do do | Mild symptoms. Mild symptoms. Severe symptons. |

RESULT OF CARDIAC INJECTIONS.

We have shown that guinea pigs may be readily sensitized by subcutaneous or intraperitoneal inoculations, and that the second injection produces symptoms when the serum is injected either under the skin or into the peritoneal cavity.

Besredka and Steinhardt have shown that the injection of serum into the brain of a sensitized guinea pig is very poisonous. We are able to confirm this observation. We then asked ourselves the question, Can guinea pigs be sensitized by injecting the horse serum directly into the heart? And can sensitized guinea pigs be poisoned by such injections directly into the circulation? We are now enabled to answer these questions affirmatively, in view of the following experiments:

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|---------------|--|-------------------|---|-------------------------------|
| 7077 | 0.142 c. c. toxine $5+\frac{1}{210}$ c. c. antitoxic horse serum (NYBH. 17), subcutaneously. | Days. 131 | 1.5 c. c. normal horse (roan) se- rum, into heart. | Dead, 3 minutes. |
| 7692 | 0.142 c. c. toxine $5+\frac{1}{440}$ c. c. antitoxic horse serum (A. 247), subcutaneously. | 35 | 0.01 c. c. normal horse (roan) se- rum, into heart. | Dead, 55 minutes. |
| 7629 | 0.142 c. c. toxine $5+\frac{1}{300}$ c. c. antitoxic horse serum (A. ppt. 31), | 44 | 1 c. c. normal horse (roan) serum, into heart. | Dead, $3\frac{1}{2}$ minutes. |
| 7632 | subcutaneously. 0.142 c. c. toxine $5 + \frac{1}{250}$ c. c. antitoxic horse serum (A. ppt. 31), | 44 | do | Dead, 3 minutes. |
| 7691 | subcutaneously. 0.142 c. c. toxine $5+\frac{1}{440}$ c. c. antitoxic horse serum (A. 247), sub- | 35 | do | Dead, 3 minutes. |
| Con- trol. | cutaneously. 2 c. c. normal horse (roan) serum, into heart. | | | No symptoms. |
| | Same guinea pig | 20 days later. | 6 c. c. normal horse (roan) serum, in- traperitoneally. | Marked symptoms. |

These experiments indicate that the endothelial cells lining the peritoneal cavity or the connective cells of the subcutaneous tissue do not necessarily play a rôle in the phenomenon we are studying.

THE GUINEA PIG REMAINS SUSCEPTIBLE A VERY LONG TIME.

That the guinea pig remains susceptible to the toxic action of horse serum a very long time is indicated in the following experiments, in which 378 days elapsed between the first treatment and the second injection.

| No. G. P. | First injection. | Interval. | Second injection. | Result. |
|--------------|--|--------------|--|-------------------|
| 4515 | 0.19 c. c. toxine 7+1 unit antitoxic horse serum (B. 27), sub- | Days. 23S | 6 c.c. normal horse (roan) serum, in- traperitoneally. | Dead, 30 minutes. |
| 4527 | cutaneously. | 378 | 6 c. c. normal horse (No. 15) serum intraperitoneally. | Dead, 37 minutes. |
| 4495 | do | | Ditto, 2 days later | |
| | Two young, born of a follows: | | | |
| 4495a | | | | Dead, 13 minutes. |
| 4495b | | | | Dead, 12 minutes. |

The above guinea pig (No. 4495) showed no symptoms at all after receiving 6 c. c. of horse serum into the peritoneal cavity 365 days after the first injection. It was then given the same quantity of serum two days following and showed severe symptoms. We have had several such instances following intraperitoneal injections, and can only explain it by the fact that sometimes the serum enters the lumen of the intestine instead of the peritoneal cavity. We called attention to this probability in Hygienic Laboratory Bulletin No. 29, page 63.

THE EFFECT OF FIRST INJECTIONS OF HORSE SERUM INTO GUINEA PIGS.

Theobald Smith a stated that guinea pigs which have received no preliminary doses of serum may die of a first injection; of 58 guinea pigs receiving 3 to 5 c. c. of diphtheria antitoxin with no preliminary treatment, 9 died and 49 showed no effect, making 15.5 per cent of the guinea pigs reacting to the first injection of horse serum. In reply to a question, Smith stated that he did not know whether or not the animals were the young of guinea pigs that had been treated.

We have injected many guinea pigs with horse serum and have never noted symptoms of death to follow the first injection, and can not help but believe that the results obtained by Smith are explained by our studies upon the maternal transmission of hypersusceptibility.

^a Smith, Theobald: Discussion of Rosenau and Anderson's paper on "Hypersusceptibility." Journ. Am. Med. Assn., Vol. 27, No. 13, Sept. 29, 1906, p. 1010.



Part XII.

SUMMARY AND CONCLUSIONS.

Profound chemical changes, perhaps in the central nervous system, are probably produced by the first injection of a strange proteid.

Guinea pigs may be sensitized with horse serum by injections directly into the heart. From this it appears that the cells lining the peritoneal cavity or the connective tissue cells of the subcutaneous tissue do not necessarily play a part in the phenomenon of hypersusceptibility.

Guinea pigs may be sensitized with the filtrate obtained from horse serum after precipitation with ammonium sulphate.

Formaldehyd does not appreciably influence the toxicity of horse serum and has no effect upon the sensitizing action.

The sensitizing substance is not dialyzable through a collodion sac. The toxic principle is not altered by various ferments, such as takadiastase, pancreatin, rennin, myrosin, invertin, emulsin, pepsin, ingluvin, malt, or papain, nor by certain alkaloids, such as atropin, strychnin, morphin, or caffein; it is also not altered by calcium chlorid, sodium nitrate, sodium chlorid, magnesium sulphate, or ammonium sulphate.

Guinea pigs sensitized with horse serum do not react to the second injection of other proteid substances such as peptone, extract of peas, egg albumen, and milk. Conversely, guinea pigs sensitized with subcutaneous injections of these substances do not react to a subsequent injection of horse serum.

Guinea pigs show quite as high a degree to susceptibility to cattle, sheep, hog, dog, and cat serum as they do to horse serum.

Guinea pigs are quite susceptible to injections of hemoglobin, egg lbumen, milk, or the extract of peas when given two injections with n interval of at least ten days. Simpler albuminous substances, such speptone, seem to have slight sensitizing and poisonous properties, hile lower nitrogenous compounds such as leucin and tyrosin possess one at all.

The reaction following a second injection of proteid matter in the inea pig appears to be common to all the higher forms of albumins substances, no matter from what source.

Chis phenomenon of hypersusceptibility in the guinea pig may be ful as a physiological test to distinguish true proteid substances in the lower forms of nitrogenous compounds. The refined antitoxic serum, bulk for bulk, is quite as toxic to sensitized guinea pigs as the untreated serum from which it was precipitated and dialyzed. There is, however, a distinct advantage gained in using the concentrated serum, as the same number of units may be given in half the bulk, and it is well known that the serum reaction in man depends partly upon the quantity of serum given.

Serum from one horse appears quite as toxic as serum from other horses. The apparent differences seem to depend upon something

connected with the sensitizing action.

The immunity produced by repeated injections, termed "anti-anaphylaxis" by Besredka and Steinhardt, appears to be relatively not quite as lasting and definite as many instances of active immunity against bacterial infections.

Hypersusceptibility to the toxic action of horse serum is transmitted through the female guinea pig. The male has no influence.

The susceptibility is not transmitted through the milk.

Maternal transmission of hypersusceptibility succeeds whether the female guinea pig is sensitized before or after conception.

The phenomenon of hypersusceptibility appears to have no relation

to aggressins.

Methemaglobin is not present in the blood of guinea pigs dead of a second injection of horse serum.

Oxygen has no influence upon the symptoms.

Neither diphtheria toxine nor tetanus toxine appreciably influence the phenomenon of hypersusceptibility produced by horse serum.

The removal of the spleen or thyroid gland does not influence hypersusceptibility in the guinea pig.

Guinea pigs fed upon beef are susceptible to a subsequent injection of cattle serum.

Guinea pigs fed with cooked meat are not susceptible to subsequent injections of serum.

When a second injection of horse serum is given directly into the heart of a susceptible guinea pig the symptoms are manifested with promptness and virulence. Under these circumstances, 0.01 c. c. (injected directly into the heart) in one instance was sufficient to cause the death of a sensitized guinea pig.

Guinea pigs remain susceptible a very long time. There is no diminution in the susceptibility of a guinea pig to subsequent injections of horse serum for at least one year. The longest period we have observed is 480 days.

We have never seen symptoms resulting from the first injection of horse serum in a guinea pig born of an untreated mother.

The problem of hypersusceptibility has an important bearing upon the question of immunity. Our work indicates that hypersusceptibility is either an essential element or one stage in the process of

resistance to a certain class of diseases. We can not escape the conviction that further studies upon the phenomenon of hypersusceptibility will have an important bearing upon the prevention and cure of certain infectious processes. The hypersusceptibility obtained by bacterial proteids and the subsequent immunity furnishes data for this belief.

From our work upon the proteid substances of animal and vegetable origin, it was but a step to the albuminous content of the bacterial cell. Experimental studies upon the bacterial proteids are of the greatest importance on account of the practical uses to which results along this line may lead.

Hypersusceptibility may easily be induced in guinea pigs with proteid extracts obtained from the bacterial cell. The first injection of the extracts used by us seems comparatively harmless to the animal. second injection of the same extract shows, however, that profound physiological changes have taken place. A definite period must elapse between the first and the second injection. The symptoms presented by the guinea pig as a result of the second injection resemble those caused by horse serum.

The phenomenon induced by a second injection is followed, in cer-

tain cases, by an immunity to the corresponding infection.

These results give a possible explanation of the period of incubation in some of the communicable diseases. Is it a coincidence that the period of incubation in a number of infectious diseases is about ten to fourteen days, which corresponds significantly with the time required to sensitize animals with a strange proteid? In certain infectious diseases with a short period of incubation, such as pneumonia, the crisis, which commonly appears in about ten days, may find a somewhat similar explanation. It is evident that diseased processes produced by soluble toxines, such as diphtheria and tetanus, do not belong to the category now under consideration.

The phenomenon of hypersusceptibility has been produced in the guinea pig by extracts obtained from the colon bacillus, yeast, hay bacillus, anthrax, tubercle bacillus, and the typhoid bacillus. The hypersusceptibility produced by the colon and typhoid bacillus was followed by a definite immunity to the corresponding infections. the case of anthrax, however, immunity does not follow hypersusceptibility to the anthrax proteid. We are, therefore, not dealing with a general law applicable to all infections, but with certain limi-

tations as in the case of antitoxic immunity.

NOTICE TO LIBRARIANS AND BIBLIOGRAPHERS CONCERNING THE SERIAL PUBLICATIONS OF THIS LABORATORY.

The Hygienic Laboratory was established in New York, at the Marine Hospital on Staten Island, August, 1887. It was transferred to Washington, with quarters in the Butler Building, June 11, 1891, and a new laboratory building, located in Washington, was authorized by act of Congress March 3, 1901.

The following bulletins [Bulls. Nos. 1-7, 1900 to 1902, Hyg. Lab., U. S. Mar.-Hosp. Serv., Wash.] have been issued:

- No. 1.—Preliminary note on the viability of the *Bacillus pestis*. By M. J. Rosenau.
- No. 2.—Formalin disinfection of baggage without apparatus. By M. J. Rosenau.
- No. 3.—Sulphur dioxid as a germicidal agent. By H. D. Geddings.
- No. 4.—Viability of the Bacillus pestis. By M. J. Rosenau.
- No. 5.—An investigation of a pathogenic microbe (B. typhi murium Danyz) applied to the destruction of rats. By M. J. Rosenau.
- No. 6.—Disinfection against mosquitoes with formaldehyd and sulphur dioxid. By M. J. Rosenau.
- No. 7.—Laboratory technique: Ring test for indol, by S. B. Grubbs and Edward Francis; Collodium sacs, by S. B. Grubbs and Edward Francis; Microphotography with simple apparatus, by H. B. Parker.

By act of Congress approved July 1, 1902, the name of the "United States Marine-Hospital Service" was changed to the "Public Health and Marine-Hospital Service of the United States," and three new divisions were added to the Hygienic Laboratory.

Since the change of name of the Service the bulletins of the Hygienic Laboratory have been continued in the same numerical order, as follows:

- No. 8.—Laboratory course in pathology and bacteriology. By M. J. Rosenau. (Revised edition March, 1904.)
 - No. 9.—Presence of tetanus in commercial gelatin. By John F. Anderson.
- No. 10.—Report upon the prevalence and geographic distribution of hookworm disease (uncinariasis or anchylostomiasis) in the United States. By Ch. Wardell Stiles.
- No. 11.—An experimental investigation of *Trypanosoma lewisi*. By Edward Francis. No. 12.—The bacteriological impurities of vaccine virus; an experimental study. By M. J. Rosenau.
- No. 13.—A statistical study of the intestinal parasites of 500 white male patients at the United States Government Hospital for the Insane; by Philip E. Garrison, Brayton H. Ransom, and Earle C. Stevenson. A parasitic roundworm (*Agamomermis culicis* n. g., n. sp.) in American mosquitoes (*Culex sollicitans*); by Ch. Wardell Stiles. The type species of the cestode genus *Hymenolepis*; by Ch. Wardell Stiles.
- No. 14.—Spotted fever (tick fever) of the Rocky Mountains; a new disease. By John F. Anderson.
- No. 15.—Inefficiency of ferrous sulphate as an antiseptic and germicide. By Allan J. McLaughlin.
 - No. 16.—The antiseptic and germicidal properties of glycerin. By M. J. Rosenau.
 - No. 17.—Illustrated key to the trematode parasites of man. By Ch. Wardell Stiles.
- No. 18.—An account of the tapeworms of the genus *Hymenolepis* parasitic in man, including reports of several new cases of the dwarf tapeworm (*H. nana*) in the United States. By Brayton H. Ransom.

No. 19.—A method for inoculating animals with precise amounts. By M. J. Rosenau.

No. 20.—A zoological investigation into the cause, transmission, and source of Rocky Mountain "spotted fever." By Ch. Wardell Stiles.

No. 21.—The immunity unit for standardizing diphtheria antitoxin (based on Ehrlich's normal serum). Official standard prepared under the act approved July 1, 1902. By M. J. Rosenau.

No. 22.—Chloride of zinc as a deodorant, antiseptic, and germicide. By T. B. McClintic.

No. 23.—Changes in the Pharmacopæia of the United States of America. Eighth Decennial Revision. By Reid Hunt and Murray Galt Motter.

No. 24.—The International Code of Zoological Nomenclature as applied to medicine. By Ch. Wardell Stiles.

No. 25.—Illustrated key to the cestode parasites of man. By Ch. Wardell Stiles.

No. 26.—On the stability of the oxidases and their conduct toward various reagents. The conduct of phenolphthalein in the animal organism. A test for saccharin, and a simple method of distinguishing between cumarin and vanillin. The toxicity of ozone and other oxidizing agents to lipase. The influence of chemical constitution on the lipolytic hydrolysis of ethereal salts. By J. H. Kastle.

No. 27.—The limitations of formaldehyde gas as a disinfectant with special reference to car sanitation. By Thomas B. McClintic.

No. 28.—A statistical study of the prevalence of intestinal worms in man. By Ch. Wardell Stiles and Philip E. Garrison.

No. 29.—A study of the cause of sudden death following the injection of horse serum. By M. J. Rosenau and John F. Anderson.

No. 30.—I. Maternal transmission of immunity to diphtheria toxin. II. Maternal transmission of immunity to diphtheria toxin and hypersusceptibility to horse serum in the same animal. By John F. Anderson.

No. 31.—Variations in the peroxidase activity of the blood in health and disease. By Joseph H. Kastle and Harold L. Amoss.

No. 32.—A stomach lesion in guinea pigs caused by diphtheria toxine and its bearing upon experimental gastric ulcer. By M. J. Rosenau and John F. Anderson.

No. 33.—Studies in experimental alcoholism. By Reid Hunt.

No. 34.—I. Agamofilaria georgiana n. sp., an apparently new roundworm parasite from the ankle of a negress. II. The zoological characters of the roundworm genus Filaria Mueller, 1787. III. Three new American cases of infection of man with horse-hair worms (species Paragordius varius), with summary of all cases reported to date. By Ch. Wardell Stiles.

No. 35.—Report on the origin and prevalence of typhoid fever in the District of Columbia. By M. J. Rosenau, L. L. Lumsden, and Joseph H. Kastle. (Including articles contributed by Ch. Wardell Stiles, Joseph Goldberger, and A. M. Stimson.)

No. 36.—Studies upon hypersusceptibility and immunity. By M. J. Rosenau and John F. Anderson.

In citing these bulletins, beginning with No. 8, bibliographers and authors are requested to adopt the following abbreviations: Bull. No. ——, Hyg. Lab., U. S. Pub. Health & Mar.-Hosp. Serv., Wash., pp. ——.

MAILING LIST.

The Service will enter into exchange of publications with medical and scientific organizations, societies, laboratories, journals, and authors. Its publications will also be sent to nonpublishing societies and individuals in case sufficient reason can be shown why such societies or individuals should receive them. All applications for these publications should be addressed to the "Surgeon-General, U. S. Public Health and Marine-Hospital Service, Washington, D. C."



TREASURY DEPARTMENT

Public Health and Marine-Hospital Service of the United States
Walter Wyman, Surgeon-General

HYGIENIC LABORATORY.—BULLETIN No. 37

M. J. ROSENAU, Director June, 1908

INDEX-CATALOGUE OF

MEDICAL AND VETERINARY ZOOLOGY

SUBJECTS:
TREMATODA AND TREMATODE DISEASES

BY

CH. WARDELL STILES

AND

ALBERT HASSALL



WASHINGTON
GOVERNMENT PRINTING OFFICE
1908



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CONTENTS.

| | Page |
|---|------|
| Introduction | 7 |
| List of specific names. | 10 |
| Bibliography of diseases, supergeneric, generic, and specific names | |
| Addenda | 385 |
| Notice to Librarians and bibliographers concerning the serial publications of | |
| this laboratory | 399 |



INDEX-CATALOGUE OF MEDICAL AND VETERINARY ZOOLOGY.

SUBJECTS: TREMATODA AND TREMATODE DISEASES.

By CH. WARDELL STILES, Ph. D.,

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INTRODUCTION.

The Index-Catalogue of Medical and Veterinary Zoology represents the combined card-catalogues of the Zoological Division of the United States Bureau of Animal Industry and of the Zoological Division, Hygienic Laboratory, United States Public Health and Marine-Hospital Service. These catalogues are arranged in three sections: Authors, Subjects, and Hosts.

OBJECT OF INDEX CATALOGUE.—The object in publishing the catalogue is to place it in permanent form. There have been four fires in the two laboratory buildings, but fortunately none of these gained headway. Were either building to be destroyed, and any part of the catalogue to be lost, the inconvenience such loss would entail in the regular work would be very extensive. By having the catalogue in printed form, it is made more accessible, and accidents are guarded against. By placing it at the disposal of other workers it is hoped to render their work less arduous, and especially it is desired to reduce the confusion in nomenclature.

Suggestions for using the index.—It should be recalled that this is an index, not a treatise. Persons who consult it should therefore not expect to find in it the reasons for any given entry. Nor

does the entry of the synonymy in any given case mean that this synonymy is necessarily correct; it is simply quoting published synonymy. The same statement refers to the hosts cited.

In looking up the literature of any given species, the reader should consult the cross reference (pp. 10 to 83) for the various genera under which the species has been published, and the synonyms given in the general entries; by looking up these references in the general catalogue (pp. 84 to 383), all or practically all the references to the species are obtained.

SUGGESTIONS IN PUBLISHING NEW NAMES.—In indexing literature, one is impressed with the differences in arrangement of the material as published by different authors. Some writers arrange the systematic portions of their papers in such a manner that it is an easy matter to obtain the important data. Other authors are less careful in this respect, and their material is arranged, in some cases, in a style which almost defies indexing.

As a very convenient style we would suggest the following diagrams:

Genus FASCIOLA Linnæus, 1758.

1758: Fasciola Linn., 1758a, 644, 648-649 (hepatica [give here either the type species or all of the species in the order given by the author]). [Give origin of name, if desired.]

[Give here the synonymy in same style as foregoing entry.]

GENERIC DIAGNOSIS.—Fasciolidæ, Fasciolinæ [It is always very useful to give the name of the family and subfamily]: [Here give the generic diagnosis.]

Type species.—[If type has not been designated, state why the species in question is now selected; as by first species rule, tautonomy. elimination. etc.]

Species FASCIOLA HEPATICA Linnæus, 1758.

1758: Fasciola hepatica Linn., 1758a, 648-649 [add here type host and type locality]. [Add synonymy here, in same style as foregoing entry.]

Specific diagnosis,—Fasciola (p. —) [give page where generic diagnosis is printed]:
[Add here specific diagnosis.]

Habitat.—[Give hosts and geographic distribution here.]

Type specimen.—[Give museum number of type specimen.]

A list of hosts, together with the parasites found in them, is an exceedingly useful adjunct to any paper dealing with parasites. Several authors have published such lists, but most authors fail to do so. If the plan of giving a compendium of hosts with every paper were more generally adopted, the papers would be very much more valuable. In indexing the hosts, especially in papers by certain authors, we have met with considerable difficulty, for it is often necessary to read carefully an entire article in order to avoid overlooking the hosts. And even then it is, unfortunately, not exceptional that the reader can not be certain that he is indexing correctly.

ABBREVIATIONS.

As a general rule we do not favor the use of abbreviations. In a work of this kind, however, in which certain names are repeated from 50 to 2,000 times, the use of abbreviations becomes necessary, in order to reduce both the bulk and the expense.

Generic names.—The general rule is here followed of abbreviating all generic names ending in -stoma or -stomum to -st, as Amphist. for Amphistoma and Amphistomum, Dist. for Distoma and Distomum, etc.

All names ending in -cercaria are abbreviated to -cerc., as Cerc. for Cercaria, Eucerc. for Eucercaria, etc.

All other abbreviations can, we believe, be understood without explanation—Fasc. for Fasciola, Plan. for Planaria, etc.

AUTHORS' NAMES.—The following abbreviations refer to authors' names:

Abildg. for Abildgaard. Ben. for Beneden. Bettend. for Bettendorf. E. Bl. for E. Blanchard. R. Bl. for R. Blanchard. Blainy, for Blainville. Brand, for Brandes. Burm. for Burmeister. Cerf. for Cerfontaine. Crep. for Creplin. Cuv. for Cuvier. Day. for Dayaine. Dies. for Diesing. Duj. for Dujardin. Erc. for Ercolani. Fil. for Filippi. Fischder. for Fischæder. Fraip. for Fraipont. Gamb. for Gamble. Gery, for Gervais. Goldb. for Goldberg. Hass. for Hassall. Jægers. for Jægerskiæld. Kath. for Kathariner. Kholodk. for Kholodkowski.

Kowal, for Kowalewski.

Kuech, for Kuechenmeister.

Leuck, for Leuckart. Levin. for Levinsen. Linn. for Linnæus. Linst, for Linstow. Lint. for Linton. Lænnb. for Lænnberg. Mol. for Molin. Mont. for Monticelli. Moul. for Moulinié. Much. for Muchling. Nord, for Nordmann, Odhn, for Odhner, Olss. for Olsson. Pag. for Pagenstecher. Par. for Parona. Per. for Perugia. Perr. for Perroncito. Poir, for Poirier. St.-Remy for Saint-Remy. Sieb, for Siebold. Sons. for Sonsino. Staff. for Stafford. Stoss, for Stossich. Rail. for Railliet. Rud. for Rudolphi. Tasch. for Taschenberg. Will.-Suhm for Willemoes-Suhm.

OTHER ABBREVIATIONS.—All literature references are taken from the Author's Catalogue of this Index (Bull. 39, U. S. Bureau of Animal Industry). The following common abbreviations are also used:

f. for family.
g. for genus.
m. for monotypical.
t. h. for type host.

t. l. for type locality.

tld. for type by later designation. tod. for type by original designation. subf. for subfamily. subg. for subgenus.

ALPHABETICAL LIST OF SPECIFIC AND SUBSPECIFIC NAMES IN TREMATODA.

The following is a complete list of all the specific and subspecific names of *Trematoda*, so far as contained in our card catalogue at date of going to press, together with the generic names with which they have been used. The references are the earliest reference we have found for the names in question, together with the type host and type locality, so far as we have as yet traced these.

It has been found necessary to include in this list some species which are not *Trematoda*. (See addenda, p. 385.)

abbreviata Olss., 1876, 12, t. h. Acanthias vulgaris.—1876: Onchocotyle. 1899: Squalonchocotyle.

abbreviatum Brand., 1888a, 55, t. h. Crocodilus; Brazil.—1888: Diplost.

abdominalis Gœze, 1782a, 41, F. intestinalis Linn., 1758a, renamed, includes also Ligula piscium.—1782: Fasc. [Ligula, cestode.]

aberrans Goto, 1894a, 198, t. h. Belone schismatorhynchus; Hagi, Japan.—1894: Axine.

aberrans Looss, 1902m, 630, t. h. Chelone mydas; Egyptian coast.—1902: Microscaphidium.

absconditus Looss, 1901e, 631, t. h. Bagrus bayad, B. docmac.—1901: Acanthochasmus.

acalepharum Costa (1864), 90, larva in Acalephen.—1864: Macrurochæta (type) [1894: Cerc. setifera.]

acanthocephalum Stoss., 1887, 94, t. h. Belone acus; Triest.—1887: Dist. 1898: Echinost. 1899: Tergestia (probably type)

acanthoides Rud., 1819a, 114, t. h. Phoca vitulina; Berlin.—1819: Dist. (Echinost.). 1860: Echinost.

acanthurum Par. & Per., 1896, 2, t. h. Brama rayi; Genova.—1896: Microcotyle.
 acanthus Nicoll, 1906, 514, t. h. Larus argentatus.—1906: Zeugorchis, type.
 1907: Parorchis, type.

acarizum Looss, 1902m, 415, t. h. Thalassochelys corticata; Egyptian coast.—1902: Enodiotrema.

acceptum Looss, 1901b, 203, t. h. Crenilabrus pavo, C. griseus, bladder; Triest.— 1901: Phyllodist.

acerca Biehringer, 1884a, 3, t. h. Onchidium carpenteri Stearns.—1884: Cerc.

acerinæ cernuæ Claparède (1857), t. h. Acerina cernua.—1857: Tetracotyle.

acervo-calcaphorum Erc., 1881e, 21, for acervocalciferum (Dist.).

acervocalciferum Gastaldi, 1854, 6, t. h. Rana esculenta.—1854: Dist.

acervocalcoforum Linst., 1875, 193, for acervocalciferum (Dist.).

acervocalcophorum Erc., 1881e, 20, for acervocalciferum (Dist.).

acetabularis Braun, 1900, 388, for acetabulatus (Pycnoporus).

acetabulatus Looss, 1899b, 611, t. h. Vesperugo kuhli; apparently Egypt, January.—1899: Pycnoporus.

acquans Sons., 1891, 263, for æquans (Diplectanum).

acreatum Stoss., 1892, 18, for ocreatum (Monost.).

actxonis Pag., 1862, 306, t. h. Acteon viridis; Cette.—1862: Dist.

aculeata Erc., 1881, see Par., 1894, 163, t. h. Lymnæa auricularis; Bologna.—Cerc.

aculeatum Nitzsch, in Giebel, 1857, 266, t. h. Strix bubo.—1857: Dist.

aculeatum Linst., 1879b, 338, t. h. Testudo græca; loc. not given.—1879: Monost. [1890: Dist. linstowii.] 1901: Telorchis.

aculeatum Par. & Per., 1889, 745, t. h. Corvina nigra; Genova.—1889: Diplectanum.

aculeatum Couch, ——.—: Trist. [1898: T. molæ.] aculeatus Odhn., 1905, 297, t. h. Conger vulgaris; Mediterranean, Belgium, Sweden.—

1905: Prosorhynchus.

aculeatus Dies., 1858, 275, t. h. Planorbis marginatus.—1858: Bucephalus (Bucephalopsis).

acuta Goto, 1894a. 217, t. h. Thynnus sibi; Hagi, Osatsubé, Japan, August.—1894: Hexacotyle.

acutum Leuck., 1842, 33, t. h. Mustela putorius.—1842: Dist.

adhærens Braun, 1901, 942, t. h. Mycothera sp.; Brazil.—1901: Glaphyrost. (type).

adhærens Looss, 1901, 624 (= vallei 1899, type of Lophotaspis), t. h. Thalassochelys corticata; Egypt.—1901: Lophotaspis (type).

adolphi Stoss., 1902, 19, t. h. Grus cinerea, Ardea cinerea; Siberia.—1902: Cyclocælum. [adriaticum v. Dr. (Dist. [tunicate].)]

aduncum Lint., 1905, 327, t. h. Opsanus tau; Beaufort, N. C.—1905; Dist.

advena Duj., 1843, 338, t. h. Šorex: Europe.—1843: Brachylaima (type). [1845: Dist. (Brachylaimus, type).] [1847: Brachylæmus.] 1892: Dist. [Brachylaimus

æglefini Mueller, 1776, 224, t. h. Gadus æglefinus, int.—1776: Fasc. 1803: Dist. [1850: Dist. simplex.] [1904: Sinistroporus (type).]

æglefini Ben., 1870, 57 [see also æglefini Mueller, 1776].—1870: Dist.

ægrefini Bosc., 1802a, v. 1, for æglefini 1776 (Fasc.).

ægyptiaca Odhn., 1905, 370, Monost. verrucosum of Looss, renamed.—1905: Notocotyle. æguptiacum Cobbold, 1876t, 757, t. h. Equus; Egypt.—1876: Diplost. 1876: Hemist. 1893: Gastrodiscus.

xgyptiacum Looss, 1896b, 33, t. h. buffles, bœufs, moutons; Egypt.—1896; Dist. 1896: Dist. hepaticum. 1898: Fasc. hepatica. 1899: Fasc. [1902: F. gigantea.] xgyptiaca Cobbold, 1866, 6, heterophyes 1852, renamed.—1866: Heterophyes (type). xquptiacus Looss, 1898a, 459, apparently lapsus for gregarius.—1898: Gastrothylax.

æquale Duj., 1845a, 410, t. h. Strix flammea: Rennes.—1845: Dist. (Brachylaimus). 1892: Mesogonimus. [1899: Clinost. ("possibly").]

xqualis Looss, 1902, 888, t. h. dogs and cats; Egypt.—1902: Heterophyes.

xquans Wagener, 1857, 99, t. h. Labrax lupus.—1857: Dactylogyrus. 1858: Diplectanum (probably type).

xquans Looss, 1899b, 652, t. h. Gerbillus xgyptius; Egypt.—1899: Heterolope. 1900:

Harmost.

æquatus Staff., 1905, 691, t. h. Eutenia sirtalis; Canada.—1905: Zeugorchis (type). affine Dies., 1850a, 410, based on Schmitz, 1826, 15, figs. 1-13, t. h. Bombinator igneus; Berlin.—1850: Hexathyridium.

affine Dies., 1850a, 359 [not Rud., 1819a]. Monost. tenuicolle Rud., 1819a, renamed, t. h. Lampris guttatus; Groningæ.—1850: Dist.

affine Leidy, 1858a, 110, t. h. Fiber zibethicus; U. S. A.—1858: Monost.

affine Rud., 1819a. 110, t. h. Perca cirrosa; Arimini.—1819: Dist. 1845: Dist. (Apoblema). 1901: Derogenes.

affine Lint., 1898, 511, t. h. Paralichthys dentatus: Woods Hole.—1898: Octoplectanum. 1901: Diclidophora.

affinis Eichwald, 1829a, 247, t. h. Lymnæus stagnalis.—1829: Cerc.

affixus Looss, 1899b, 587, t. h. Corvus cornix, Falco tinnunculus, Circus æruginosus, Recurvirostra avocetta: Egypt.—1899: Pygorchis (type).

africanum Stiles, 1901, 594, bursicola Looss, 1899 [not Crep., 1837], renamed.—1901: Echinost.

africanum Stoss. in Galli-Valerio, 1906, 49, t. h. a fish; French Congo.—1906: Clinost. agamos Linst., 1872, 1, t. h. Gammarus pulex L.—1872: Dist.

agassizi Goto, 1899a, 286, t. h. Remora brachyptera; Newport, R. I.—1899: Dionchus (type).

agassizii Mont., 1905, 69, for agassizi (Dionchus).

agilis Fil., 1857c, 4, t. h. Lymnæus stagnalis; Turin.—1857: Cerc. 1858: C. (Gymnocephala).

agilis Leidy, 1858a, 110, free in Delaware River.—1859: Cerc.

alacre Looss, 1901, 401, t. h. Labrus maculatus, L. merula, Crenilabrus pavo, C. quin-quemaculatus, C. griseus.—1901: Dist.

alata Geze, 1782a, 176, t. h. Canis vulpes; Germany.—1782; Plan. [1788; Alaria vulpis, type]. 1790; Festuc. 1793; Fasc. 1800; Dist. 1819; Holost. 1850; Hemist. (type). 1889; Emist. 1894; Diplost. (Hemist.). 1896; Conchosomum (type).

alata Hemp. & Ehrenb., 1828a, in water; Berlin.—1828: Histrionella. 1856: Cerc.

alatus Linst., 1878, 227, t. h. Blicca bjærkna; Europe.—1878: Dactylogyrus.

alba Erc., 1881, 12, for Cerc. brunnea var. Dies.—1881: Cerc.

alba Cerf., 1899a, 377, t. h. Raja alba.—1899: Rajonchocotyle.

albicolle Rud., 1819a, 98, t. h. Falco pennatus; Mus. Vien.—1819: Dist. 1845: D. (Dicrocœlium). 1901: Dicrocœlium.

albidum Braun, 1893, 347, t. h. Felis catus domestica; Kœnigsberg i. Pr.—1893; Dist. (Dicrocœlium). 1896: Opisthorchis. 1899: Metorchis (type).

albidum Brand., in Ofenheim, 1900, 145, t. h. Aëtobatis narinari; Pacific.—1900: Anaporrhutum (type).

albocæruleum Stoss., 1889, 28. t. h. Sargus salviani; Triest.→1889: Dist.

album Kuhl & van Hasselt, 1822a; 1824a, 311 t. h. Chelonia midas; Iles des Gocotiers.—1822: Monost. 1899: Cricocephalus (type).

album Stoss., 1890, 42, t. h. Cantharus orbicularis; Triest.—1890: Dist. [1899; ? Creadium.] 1904: Lepocreadium (type). 1901: Allocreadium.

alcedinis Par. & Per., 1890, 7, t. h. Smaris alcedo; Genova.—1890: Microcotyle.

allostomum Dies., 1850a, 367. Dist. colubri murorum 1811 and 1819 renamed, t. h. Tropidonotus natrix: M. C. V.—1850: Dist.

alosæ Hermann, 1782, 180, t. h. Clupea alosa.—1782: Mazocraes (type).

alosæ Hermann, 1783a, 46. t. h. Maifisch im Rhein, Clupea alosa.—1783: Fasc. [1850: Dist. appendiculatum.]

alosæ Leuck. (1828) [see lanceolatum].—Octobothrium. [1845: O. lanceolatum.] alosæ Mont., 1888, 13.—1888: Ophycotyle.

alosa Kuhn 1829, 358, t. h. Clupea alosa.—1829: Octost. (type). [1852: Octobothrium lanceolatum.]

alosæ Mayer, 1841a, see lanceolatus alosæ.

alosæ Ben. & Hesse, 1863: 1864, 102, t. h. Alosa vulgaris.—1864: Glossocotyle (type). 1890: Octobothrium (Glossocotyle).

aloysiæ Stoss., 1885, 161, t. h. Corvina nigra; Triest.—1885: Dist. 1886: D. (Brachylaimus). [1899: Anoiktost., type.]

alpina Dana.—1891: Plan.

altemon Ben., 1870, 48, see atomon (Dist.).

aluconis intestinale Rud., 1819a, 119, t. h. Strix aluco.—1819: Dist. [1892: sub Echinost. apiculatum.]

aluconis thoracicum Rud., 1819a, 119 (Plan. pusilla Braun, 1790, pro parte renamed); t. h. Strix aluco.—1819: Dist.

alveatum Mehlis, in Crep., 1846, 142, t. h. Anas, Cygnus, see p. 323.—1845: Monost. 1892: Notocotyle.

alveatum Mehlis of Mueh., 1898, renamed alveiforme Cohn, 1904.

alveiforme Cohn, 1904, 230, alveatum Mehlis of Mueh., 1898, renamed.—1904: Monost.
americana Hass., 1891c, 359, t. h. Bos taurus: U. S. A.; carnosa Hass., 1891, renamed.—1891: Fasc. 1892: Dist. [Fasc. magna.]

americanus Staff., 1902, 719, t. h. Rana virescens; Ashbridge's Bay, Toronto.—1905: Cephalogonimus.

americanum Osborn, 1903, 252, t. h. Amblystoma punctatum; Minnesota.—1903: Phyllodist.

amiuri Staff., 1904. 402, t. h. Amiurus nebulosus; Canada.—1904: Monost.

amphibothrium Wagener, 1857, 57, t. h. Acerina cernua.—1857: Dactylogyrus.

amphielucus Looss, 1899b, 565, for amphilcucus (Metorchis).

amphileucum Looss, 1896b, 55, t. h. Naja haje; Alexandria, Egypt.—1896: Dist. 1898-

Opisthorchis. 1899: Metorchis.

amphiorchis Braun, 1899b, 719, t. h. Thalassochelys corticata at Triest, Chelone mydas and Podocnemis expansa.—1899: Dist. 1899: Anadasmus (type). [1901: Orchidasma (type).] 1901: Orchidasma (type).

amphistoma Nord., 1840, 616, probably lapsus (D[istoma]).

amphistomi subclavati Ben., 1858a, 82, in Cyclas cornea, etc.—1858: Cerc. [1859:

Diplodiscus subclavatus.]

amphistomoides Bojanus, 1817b, 270, t. h. Castor fiber.—1817: Dist. 1817: Dystoma. [1850: Amphist. subtriquetrum.] [1902: Cladorchis (Stichorchis) subtriquetrus.] amplicara Looss, 1899b, 606, D. cygnoides var. A. of Bensley, 1897, renamed.—1899:

Gorgodera.

ampullaceum Buttel-Reepen, 1900, 586, in Cetacean; Indian Ocean.—1900: Dist.

anarhichæ Jacoby, 1900, 13, for anarrhichæ.—1900: Dist.

anarrhichæ Rud., 1819a, 121; anarrhichælupi Rathke, pars (from intestine) renamed.— 1819: Dist. [1819: ? D. appendiculatum.]

an[arrhichæ] lup[i] Rathke, 1799, 70, t. h. Anarrhichas lupus.—1799: Dist. [1809: D. incisum.

anastrophus Fischder., 1901, 375, t. h. Cervus dichotomus; Brazil.—1901: Balanorchis (type).

anatinus Markow, 1902, 1903a, 287, t. h. Anas boschas dom.—[1902: Prosthogonimus.]

1902: Prostogonimus.

anatis Schrank, 1788, xvi, 164, t. h. ducks.—1788: Festucaria [type]. 1790: Fasc. 1800: Dist. [1815: Monostomeus, type.] [1850: Dist. echinatum.] [1892: Echinost. echinatum.] [1901: ? Echinost. echinatum.]

anatis Rud., 1819a, 793, for anatis querquedulæ (Amphist.).

anatis domesticæ Rud., 1809a, 431 (Hirudo fasciolaris Mueller, 1788, Fasc. anatis Bruguière, 1791, renamed).—1809: Dist. [1850: Dist. ovatum.]

anatis fuscæ Viborg, 1795, 243, t. h. Anas fusca; Copenhagen.—1795: Dist.

anatis nigræ Dies., 1858e, 322, t. h. Anas nigra.—1858: Holost.

anatis querquedulæ Rud., 1819a, 92, t. h. Anas querquedula; Cat. Ent. Vien.—1819: Amphist.

anatis tadornæ Viborg, 1795, 196, t. h. Anas tadorna.—1795: Strigea. 1809: Amphist. [1814: A. isostomum.] [1845: Holost. erraticum.]

anceolatum Braun, 1892a, 677, misprint for lanceolatum (Dist.).

anceps Mol., 1859, 845, t. h. Fulica atra; Padua.—1859: Dist. 1892: Echinost.

anceps Looss, 1899b, 629, t. h. Machetes pugnax; Marg, April.—1899: Prymnoprion. 1901: Prosthogonimus. [1902: Pros. cuneatus.]

anchoratus Duj., 1845a, 480, t. h. "la carpe;" Europe.—1845: Gyrodactylus. 1857: Dactylogyrus. [1879: D. auriculatus.]

ancyli lacustris Dies., 1855a, 400, based on Baer, 1827b, 656, t. h. Ancylus lacustris.— 1855: Dist. 1855: Cercariæum.

andersoni Cobbold, 1876, 46, t. h. Platanista gangetica.—1876: Dist. 1892: D. (Brachylaimus).

anellidicola Mont., 1888, 88, for annelidicola (Cyclatella).

anguillæ Gmelin, 1790a, 3056, t. h. Anguilla; refers to Leeuwenhoek, 316, fig. 6.—1790: Fasc. 1803: Dist. [1819: ? D. polymorphum.]

anguillulæ Dies., 1850a, 340, lapsus for anguillæ Abildg. and Zed.—1850: Dist.

anguis Linst., 1885, 250, t. h. Anguis fragilis.—1885: Dist. 1895: Agamodist.

angulata Mueller, 1774, 58, "in fundo arenoso oceani."—1774: Fasc. 1776: Planaria. ——: Amphiporus.

angulatum Duj., 1845a, 401 (not Fasc. angulata Mueller, 1776), t. h. Anguilla: Morbihan.—1845: Dist. (Podocotyle [type]). 1901: Podocotyle, type. [1905: see Podocotyle atomon.

angusta Rail., 1895, 338, see hepatica angusta.—1895: Fasc. hepatica. 1899: Fasc. [F. gigantica.]

[angusta Muenster, 1842, 98.—Hirudinella. Hirudella. Leach.]

angusticolle Hausmann, 1896a, 391, t. h. Cottus gobio; Basel, Switzerland.—1896: Dist. 1899: Creadium. 1901: Allocreadium.

angustum Staff., 1900, 407, t. h. Chrysemys picta.—1900: Dist. 1905: Telorchis.

angustum Schlotthauber, 1860, 129, t. h. Scolopax gallinago.—1860: Monost.

aniarum Leidy, 1891a, 414, t. h. Tropidonotus sipedon.—1891: Dist.

annelidicola Ben. & Hesse, 1863, 1864a, 82, t. h. ? Clymene.—1863: Cyclatella (type). [1869: Loxosoma.]

annulatum Dies., 1850a, 386, t. h. Gymnotus electricus; Brazil.—1850: Dist. 1860: Echinost. 1892: Dist. (Echinost.).

annulicauda Bory, 1825b, 253.—1825: Histrionella.

annuligerum Nord., 1832, 53, t. h. Flussbarsch, Perca fluviatilis.—1832: Dist. 1886: D. (Brachylaimus).

anodontæ Osborn, 1898, 416, t. h. Unio luteolus, Anodonta; New York.—1898: Platyaspis. [1899: Cotylaspis insignis.]

anonymum Dies., 1858e, 341, based on Bellingham, 1844, 428, t. h. Gadus æglefinus, G. euxinus, Merlangus carbonarius, M. vulgaris; Ireland.—1858: Dist.

anseris Gmelin, 1790a, 3055; Fasc. verrucosa Frœlich, 1789, renamed; t. h. Anser domesticus, rectum: Europe.—1790: Fasc. [1850: Notocotyle seriale.] [1861: Monost. verrucosum.] [1905: Catatropis verrucosa, type.]

anthos Braun, 1899b, 720, t. h. Cheloniæ; Yedo.—1899: Dist. 1901: Calycodes (type).

anticum Staff., 1905, Apr. 11, 693, t. h. Vespertilio subtilis Say; Canada.—1905: Lecithodendrium.

apertum Rud., 1819a, 108, t. h. Apogon ruber s. Mullus imberbis; Naples.—1819: Dist. 1845: D. (Apoblema).

apiculata Rud., 1803, 31; D. stridulæ Reich, 1801, renamed.—1803; Fasc. 1809; Dist. (Echinost.). 1860; Echinost.

apiculatum Olss., 1869, 4, t. h. Acanthias vulgaris; Skagerrack.—1869: Microbothrium (type). 1890: Pseudocotyle.

apodis Packard, 1882, 142, in egg-sacks of Apus.—1882: Dist.

apolaimum Heymann, 1905, 91, t. h. Kachuga tectum (Gray) small int.—1905: Dist.
appendiculata Rud., 1802, 78, t. h. Clupea alosa, stomach; Europe.—1802: Fasc.
1808: Dist. [1809: Hemiurus (type).] 1845: Dist. (Apoblema [type]). 1847:

Apoblema. 1896: Hemiurus (type).

appendiculata Frœlich, 1802. 56 (not Rud., 1802) t. h. Anas boschas; Europe: see papillatum Rud., 1814a, 105, renamed oxycephalum Rud., 1819a, 98, 375.—1802; Fasc. [1814: Dist. oxycephalum.] [1850: Dist. oxycephalum.]

appendiculata Leidy, 1877, 202, t. h. Helix arborea.—1877: Dist. [1891: D. centrap-

pendiculatum.

appendiculata Kuhn, (1829c), 460, t. h. Squalus catulus.—1829: Polyst. 1840: Hexabothrium (type). 1850: Onchocotyle (type). 1879: Hexathyridium. 1890: Octocotyle. 1899: Acanthonchocotyle (type).

apri Gmelin, 1790a, 3054, see hepatica apri.—1790: Fasc. hepatica. aquilæ Leidy, 1887, 24, t. h. Haliaëtos leucocephalus.—1887: Dist.

arcanum Nickerson, 1900, Oct., 811, in frogs.—1900: Dist. 1902: Pleurogenes. 1905: Loxogenes (type).

arcticus Odhn., 1905, 339, t. h. Phoca barbata; Spitzbergen.—1905: Orthosplanchnus (type).

arcuata Steenstrup, see Par., 1894, 164, t. h. Lymnæa obscura. L. stagnalis; Bologna.—Cerc.

arcuata Sons., 1890. 112, t. h. Lichia amia: Pisa.—1890: Octocotyle. 1890: Octobothrium. [1890: Octocotyle striata.]

arcuatum Brand., 1892b, 507, t. h. in aquatic birds [probably Anas clangula; Berlin].—1892: Monost. [1892: Cyclocœlum.] 1902: Cyclocœlum.

arcuatum Duj., 1845a, 410, t. h. Corvus glandarius; Rennes.—1845: Dist. (Brachylaimus).

arcuatum Lint., 1900, 277, t. h. Sarda sarda; Woods Hole.—1900: Gasterost.

ardeæ Gmelin, 1790a, 3055, t. h. Ardea stellaris. int.; Europe: Planaria teres pars Gœze, 1782a, renamed.—1790: Fasc. 1803: Dist. ——: Planaria. [1850: Dist. ferox.]

ardex minutx Pontallié, 1853.—Dist. (Cladocalium).

ardex nigrx Viborg (1795), see Rud., 1809a, index.—1795: Dist. [1809: D. hians.]

ardeæ stellaris Rud., 1809a, 432; Fasc. ardeæ Gmelin, 1790, renamed.—1809: Dist. [1850: D. ferox.]

arenula Crep., 1825a, 53, t. h. Fulica atra.—1825: Dist.

areolatum Rud., 1809a, 50, Fasc. platessæ Mueller, 1788, renamed; t. h. Pleuronectes platessa.—1809: Dist. 1886: D. (Echinost.).

aristotelis Stoss., 1892, 14, D. chilostomum Mehlis of Ben., 1873b, t. h. Rhinolophus hippocrepis, Vespertilio murinus, V. desycneme, V. daubentonii, V. emarginatus, V. mystacinus, Nannugo pipistrellus.—1892: Dist. 1892: D. (Brachylaimus).

armata Sieb., 1837, 187, based on Wagner, 1834, 131.—1837: Cerc. 1855: Cerc. (Xiphidiocerc.). 1858: C. (Acanthocephala). [1858: Dist. endolobum.]

armata Rud., 1793a, 26; Fasc. putorii Gmelin, 1790a, renamed.—1793: Fasc. 1803: Dist. [1850: Dist. trigonocephalum.] [?1892: Echinost. echinatum.]

armata minor Ben., 1858, 98, t. h. Lymnæus stagnalis; Belgium.—1858: Cerc. [1859: Dist. retusum.]

armatissimum Linst., 1903, 280, t. h. Iguana sp.—1903: Dist.

armatum MacCallum, 1895, 401, see isoporum armatum.—1895: Dist. isoporum. [See also Creadium.]

armatum Mayer, 1841, 4, Echinorhynchus renamed.—1841: Monost.

armatum Leuck., 1835, 88, t. h. Acipenser rostratus.—1835: Diclibothrium (type). 1835: Diclybothrium. 1842: Diplobothrium (type). 1845: Polyst. (Hexacotyle).

armatum Molin, 1858, 130 (not Rud. 1793), t. h. Phasianus gallus; Patavii.—1858: Dist. [1896: Echinost. echinatum.]

armatum Mol., 1859, 291, t. h. Conger conger; Batavii.—1859: Gasterost.

armatum Fuhrmann, 1904, 61, t. h. Rostrhamus sociabilis.—1904: Echinost.

armatum paludinæ impuræ Fil., 1857c, pl. 2, figs. 14-15.—1857: Dist.

arrectum Duj., 1845a, 403, t. h. "lézard vert;" Rennes.—1845: Dist. (Brachycœlium).

1895: D. (Dicrocœlium).

arrectum Duj., of Mol., 1859, 831, misdetermined.—1859: Dist. [1899: Telorchis.] 1904: Telorchis.

ascidia Rud., 1819a, 108, t. h. Sparus boops and S. pagrus; Arimini and Naples.—1819: Dist. 1903: D. (Brachylaimus).

ascidia Ben., 1873, 328, not Rud., 1819a; see lagena Brand., 1888, not Rud., 1819a.
 1873: Dist. 1892: D. (Dicrocœlium). [1896: Lecithodendrium.] 1899: Lecithodendrium (type).

ascidiæ Dies., 1858e, 373, t. h. Mentula marina.—1858: Aspidogaster.

ascidioides Ben., 1873, 332, t. h. Vesperugo noctula.—1873: Dist. 1892: D. (Brachylaimus). 1899: Lecithodendrium. 1902: D. (Lecithodendrium). [1905: Lec. chilostomum Mehlis.]

ascoidea Leidy, 1877, 201, t. h. Planorbis parvus.—1877: Dist. (Gymnocephala).

asper Looss, 1899b, 601, separated from variegatus.—1899: Hæmatolæchus. 1905: Pneumonœces.

asperum Dies., 1838a, 189, t. h. Tapirus americanus; 1839, 236, Matogrosso and Cacheira do Bananeira, South America.—1838: Amphist. 1901: Cladorchis.

asperum Nitzsch, in Crep., 1849a, 71, t. h. Anas fuligula.—1849: Monost.

asperum Vaillant, 1863, 347, t. h. Siren lacertina.—1863: Monost. 1892: Monostomulum.

asperum Wright, 1879, 57, t. h. Ardea minor.—1879: Dist. 1892: Echinost. 1903: Pegosomum.

asperus Stoss., 1904, 1, t. h. Plecotus auritus; Grisignana, Istria.—1904: Plagiorchis.

aspidophori Ben., 1870, 34, t. h. Aspidophorus europæus.—1870: Dist.

assula Duj., 1845a, 398, t. h. Coluber natrix; Toulouse.—1845: Dist. (Dicroccelium).

asymphyloporum Stoss., 1901, 96, t. h. Trutta trutta; Lake Plitvica, Croazia.—1901: Allocreadium.

atomata Bosc, 1802a, v. 1, 260.—1802: Planaria.

atomon Rud., 1802, 70, t. h. Pleuronectes flesus, stomach; Greifswald, Europe.—1802:
 Fasc. 1809: Dist. 1886: D. (Dicroccelium). 1901: Allocreadium. 1905: Podocotyle (type).

atriventre Weinland, 1856, 24, t. h. Physa heterostropha.—1856: Dist.

attenuata Staff., 1902, 418, t. h. Rana catesbiana, R. virescens; Canada.—1902: Gorgodera. 1905: Gorgoderina.

attenuatum Rud., 1809a, 328, t. h. Scolopax gallinago; Greifswald.—1809: Monost. (Monost.). [Notocotyle verrucosum.]

attenuatum Rud., 1814a, 103, Fasc. longicollis Abildg., renamed; later renamed Dist. naja Rud., 1819a, 377.—1814: Dist.

attenuatum Duj., 1845a, 392 [nec Rud.], t. h. Turdus merula; Rennes.—1845: Dist. (Dicrocœlium). 1900: Dicrocœlium. [1850: D. macrourum.]

attenuatum Bremser, MS., in Braun, 1901f, 563, t. h. Rhynchops nigra; Brazil.—1901: Dist. [1901: Microlistrum spinetum.]

attenuatum Dies., 1836d, 238, t. h. Salmo paccu; Caiçara, Brazil.—1836: Amphist.

attenuatum Linst., 1906, 11, t. h. Buteo vulgaris.—1906: Hemist.

augusticaudum Staff., 1904, 488, t. h. Lota maculosa, Stitzostedion vitreum; Canada.— 1904: Mimodist. (type).

augustus Staff., 1905, Apr. 11, 690, for angustum 1900.—1905: Telorchis.

aurantiaca Haswell, 1900, 431, t. h. Astacopsis sp.—1900: Temnocephala.

auricularis Wedl, 1857, for auriculatus (Gyrodactylus).

auriculata Bosc, 1802a, v. 1, 261.—1802: Planaria.

auriculatum Wedl, 1857, 242, t. h. Acipenser ruthenus.—1857: Dist. 1886: D. (Crossodera). 1903: Bunodera.

auriculatum Wedl of Lint., 1898, 521.—1898: Dist. [1904: Acrodactyla petalosa (type of A.).]

auriculatus Nord., 1832, 108, t. h. Cyprinus brama.—1832: Gyrodaetylus. 1850: Daetylogyrus (type).

auriflavum Mol., 1859, 287, t. h. Ardea nycticorax; Batavii.—1859: Diplost.

auritum Duj., 1845a, 370, t. h. Strix flammea; Rennes.—1845: Holost. 1850: Hemist. auxis Tasch., 1879, 613, t. h. Auxis rochei; Naples.—1879: Didymozoon.

axenos Mont., 1898; or 1899, 83, t. h. unknown; Brazil.—1899: Temnocephala.

baccigerum Rud., 1819a, 108, t. h. Atherina hepsetus; Naples.—1819: Dist.

bacillare Mol., 1859, 834, t. h. Centrolophus pompilius; Batavii.—1859: Dist. 1886: D. (Dicroccelium). [1899: Creadiinæ, ? Creadium.]

baculifer Braun., 1900, 28, t. h. Didelphys palmata; Brazil.—1900: Rhopalias.

baculum Lint., 1905, 327, t. h. Scomberomorus maculatus; Beaufort, N. C.—1905: Gasterost.

baculus Dies., 1850a, 391, D. mergi 1819 renamed, t. h. Mergus albellus.—1850: Dist. 1860: Echinost.

badia Rathke, 1799, 147.—1799: Plan.

baelzi Cobbold, 1884g, 976, D. pulmonale Baelz renamed, t. h. Homo.—1884: Dist. [Paragonimus, type].

bagri incapsulatum Wedl, 1861, 479, t. h. Bagrus sp.—1861: Dist.

baraldii Sons., 1892, 91, t. h. Zamenis viridiflavus Lacep.—1892: Dist. 1895: D. (Brachylaimus).

barbata Linn., 1761, 505, t. h. Loligo; Sweden.—1761: Fasc. [1779: Echinorhynchus.]
1809: Dist. [1809: Tetrabothriorhynchus migratorius, cestode.] [1819: Tetrarhynchus megabothrius.]
1853: Tetrabothriorhynchus. [1905: Tetrarhynchus.]

barbatum Cohn, 1902k, 47, t. h. Coryphæna.—1902; Lecithocladium. 1907: Dinurus.
baryurum Staff., 1903, 822, t. h. Necturus maculatus Raf.; Canada.—1903: Monocæcum (type).

bathycotyle Fischder., 1901, 368, t. h. Bos kerabau; Ceylon.—1901: Paramphist.

batis Cerf., 1899a, 376, t. h. Raja batis.—1899: Rajonchocotyle (type).

batryophoron Ben., 1870, 51, for botryophoron (Dist.).

beleocephalum Linst., 1873, 104, t. h. Ardea cinerea.—1873: Dist. 1892: Echinost.

[bellii, reptile (Bucephalus).]

bellinghamii Cobbold, 1860a, 45, falconum Dies., renamed, t. h. Falco nisus, F. rufus.—1860: Holost.

bellones Otto, 1823, 300, t. h. Bellone acus; Naples.—1823: Cyclocotyla (type). 1837: Cyclocotyle (type). 1840: Octobothrium.

belones Nord., 1840, 600, for bellones Otto (Octobothrium).

belones Crep., 1839, 291, for bellones Otto (Cyclocotyla).

belones Braun, 1893, 871, t. h. Belones vulgaris; based on Wedl, 1855, 382–383.—1893: Dist.

belones Abildg., 1794d, 59, t. h. Belone acus.—1794: Axine (type).

belones vulgaris Dies., 1855, 64, t. h. Belone vulgaris; Triest; based on Wedl, 1855, 382–383.—1855: Dist.

benedeni St.-Remy, 1898, 566, for van benedeni.—1898: Dactylogyrus.

benedeni Stoss., 1898, 51, for benedenii.—1898: Dist. 1902: Haploporus (type).

benedeni Mont., 1893, see benedenii.—1904: Nematobothrium (Didymozoon).

benedeni St.-Remy, 1898, 566, for van benedenii (Tetraonchus).

benedenii Stoss., 1887, 95, t. h. Mugil chelo.—1887: Dist. 1902: Haploporus (type).

benedenii Mont., 1893, 137, t. h. Orthagoriscus.—1893: Didymozoon.

bergense Olss., 1868, 43, t. h. Anguilla vulgaris.—1868: Dist. 1886: D. (Brachylaimus) [1905: Lecithaster gibbosus.]

bergi Par., 1900, 193, t. h. Raja planata; Montevideo.—1900: Fasc.

beroës Will, 1844, 343, t. h. Beroës rufescens; Triest.—1844: Dist. [1850: D. papillosum.]

betencourti Mont., 1892b, 127; 1893, 33, t. h. Scyllium; Boulogne.—1893: Dist. 1899: Pleurogenes.

bicornis Bosc., 1802a, v. 1, 257.—1802: Plan.

bicoronatum Stoss., 1883, 113, t. h. Ciconia nigra.—1883: Dist. 1886: D. (Echinost.). 1901: Stephanochasmus. 1902: Echinost.

bifasciata Haswell, see Mont., 1889, 2.—1889: Temnocephala.

bifurcatum Wedl, 1861, 477, t. h. Crocodilus vulgaris; Egypt.—1861: Dist. 1888: Diplost.

bifurcum Braun, 1899, 631, t. h. Flussschildkröten.—1899: Dist. 1901: Telorchis.

bijugum Braun, 1901g, 896, t. h. Himantopus melanopterus; Brazil.—1901: Stomylotrema.

bijugum Miescher, (1838a), pp. 28, 1 pl., t. h. Fringillidæ.—1838: Monost. [1840] Monost. faba.]

[bilamellatæ, see sub Fasc.]

bilharzii Herff, 1894, 415, for hæmatobium.—1894: Dist. [Schistosoma.]

bilineata Haldeman, 1840a, 3, t. h. Limnea catascopium; Camden on Delaware.— 1840: Cerc. 1850: Histrionella.

biliosum Leidy, 1858, 111, in a fish; America.—1858: Dist.

bilis Braun, 1790, 61, t. h. Falco melanaëtus; Germany.—1790: Plan. 1790: Fasc. 1803: Dist. [1809: Dist. crassiusculum.] 1898: Campula.

billis Bosc, 1802a, v. 1, 269, for bilis 1790 (Fasciola).

bilobum Rud., 1819a, 114, t. h. Tantalus falcinellus; Mus. Vien.—1819: Dist. (Echinost.) 1860: Echinost.

bilobus Looss, 1901, 569, t. h. Chelone mydas; Egypt.—1901: Pleurogonius.

binodis Mueller, 1776, 224, in intest. of fish.—1776: Fasc. 1803: Dist.

biparasiticum Goto, 1894a, 251, t. h. Parapetalus, Thynnus albacora; Japan.—1894: Trist.

bipartita Sons., 1897, 253, t. h. Limnæa palustris; Pisa.—1897: Cerc.

bipartitum Wedl, 1855, 378, t. h. Thynnus vulgaris.—1855: Monost. 1860: Wedlia (type). [1878: Didymozoon (type).] 1893: Dist. 1902: Didymost. (type).

birói Mont., 1905, 21, t. h. Sesarma gracilipes A. Edw.; Sattelberg, New Guinea.—1905: Craniocephala (type).

blainvillei Cobbold, 1860a, 39, M. delphini Blainv., renamed.—1860: Monost.

blanchardi, Stoss., 1898, 61, t. h. Labrax lupus; Triest.—1898: Gasterost.

blanchardi Tasch., 1878, 564, for blanchardii (Trist.).

blanchardi Haswell, 1893f, 153, corrigendum, t. h. Engæus fossor; Grippsland, Victoria, Australia.—[1893: Actinodactylus.] 1893: Actinodactylella.

blanchardii Cobbold, 1860a, 8, Brachylæmus erinacei Bl., 1847, renamed, t. h. Erinaceus europæus; Paris.—1860: Dist. [1892: D. linguæforme.]

8588—No. 37—08——2

blanchardii Dies., 1850a, 430, T. squali E. Bl., 1847, renamed, t. h. Squalus sp.; New Zealand.—1850; Trist.

blennii Mueller, 1776, 224, t. h. Blennius viviparus.—1776; Fasc. 1803; Dist. [1809; D. divergens.]

bliccæ Linst., 1877, 185, t. h. Blicca bjoerkna.-1877: Dist.

bolodes Braun. 1902b. 11, t. h. Fulica atra; Rossitten.—1902: Dist.

bombynæ Zed., 1800a, xvi, t. h. Rana bombyna, lungs; Europe.—1800; Monost. [1894; Dist. variegatum.]

bomfordi Montgomery, 1906, 143, t. h. Bos indicus; India.—1906: Schistosoma.

bonnieri Mont., 1893, 40, t. h. Trigla gurnardus; Wimereux.—1893: Dist. 1899: Liopyge (type). [1902: Liocerca (type).]

borcalis Sons., 1890, 177, misprint for borealis (Onchocotyle).

borealis Olss., 1893. 7, t. h. Thymallus vulgaris, Coregonus lavaretus; Scandinavia.—1893: Dactylodiscus (type). 1905: Tetraonchus.

borealis Ben., 1853l, 59, t. h. Scimnus glacialis.—1853: Onchocotyle, 1857: Polyst. 1899: Squalonchocotyle (type).

boschadis Schrank, 1790, 122.—1790: Festuc.

boschadis Schrank, 1803. 209, t. h. Anas boschas sylvestris, A. b. dom.—1803: Fasc.

bosci Cobbold, 1859d, 364, includes Fasc, colubri Bosc, Dist, colubri americani Rud., t. h. Coluber sp.—1859: Dist, 1895: D. (Dicrocœlium). [1905: Zeugorchis.]

boscii Cobbold, 1879, 455, for bosci.—1879: Dist., 1895: D. (Dicroccelium).

bothriophoron Braun, 1892f, 49, t. h. Bos indicus; Madagascar.—1892: Amphist. 1901: Parmaphist.

bothriophorum Stiles, 1898a, 24, for bothriophoron (Amphist.).

bothryophoron Braun, 1892a, 700, for botryophoron Dist. ..

bothryophorus Olss., of Looss, 1899b, 728, renamed confusus 1905.—1899: Hemiurus, 1901: Lecithaster (type).

botryophoron Olss., 1868, 42. t. h. Cyclopterus.—1868: Dist. 1886: D. (Brachylaimus), 1905: Lecithophyllum (type).

botryophorum Odhn., 1905, 359; for botryophoron Olss., 1868.—1905; Lecithophyllum (type).

botryophorus Odhn., 1905, 357, for bothryophorus of Looss, 1899 (Hemiurus).—1905: Lecithaster. [L. confusus.]

boum Gmelin, 1790a, 354.—1790: Fasc. hepatica.

boris Sons., 1876, 84, t. h. Bos taurus: Egypt.—1876: Bilharzia. 1893: Gynæcophorus. 1895: Schistosoma.

brachycolia Luehe. 1901d. 51, t. h. Box salpa: Genua.—1901: Mesometra.

brachycælium Cohn. 1903. 39. host not given.—1903: Amphist.

brachydelphium Heymann, 1905, 81, t. h. Dermatemys mavii.—1905: Patagium (type).
brachysoma Villot, 1878, 27, t. h. Anthura gracilis Leach.—1878: Cerc. [1888: Dist brachysoma.]

brachysomum Crep., 1837a, 314, t. h. Hæmatopus ostralegus.—1837: Dist. 1892: D. (Brachyccelium). 1899: Levinsenia (type). 1901: Levinseniella (type).

brachyura Fil., 1837a, 337, t. h. Planorbis submarginatus: Pavia. See next entry.—1837: Cerc.

brachyura Dies., 1850a. 296. Dist. polymorphum Fil.. 1837a. 337, renamed; t. h. Planorbis submarginatus; Ticini.—1850: Cerc. 1855: Cerc. (Eucerc.). 1858: C. Acanthocephala).

brachyura Lespés. 1857b. 117, t. h. Trochus cynereus.—1857: Cerc. [1858: C. (Acanthocephala pachycerca.]

brachyurum Linst., 1905, 418, 422, misprint for baryurum (Monocæcum).

bramæ Mueller. 1776, 224. t. h. Cyprinus brama.—1776; Fasc. 1803; Dist. [1809; Dist. globiporum.] [Sphærost.]

bramæ Par. & Perugia, 1896. 1, t. h. Brama rayi; Genova.—1896: Octobothrium. 1898: Octocotyle.

branchialis Vallot, 1840.—1840: Astacobdella (type).

branchialis Darr, 1902, 644. t. h. mackerel; German East Africa.—1902; Bathycotyle (type).

braunii Cobbold, 1860a, 43, M. murænulæ renamed, t. h. Coregonus murænula.—1860: Monost.

brasilianum Stoss., 1902b, 15, t. h. Scolopax flaviceps; Brazil.—1902: Cyclocœlum.
brevicaudata Piana, 1882, teste Parona, 1894, 161, t. h. Helix carthusiana; Reggio Em.—
1882: Cerc.

brevicaudatum Nord., 1832a, 52, t. h. Barbus communis; Berlin.—1832: Holost. 1850: Diplost.

brevicolle Crep., 1829, 54, t. h. Hæmatopus ostralegus; Greifswald.—1829: Dist. 1892: D. (Dicrocœlium). 1902: Psilost.

brevicornis Mont., 1889, 1, t. h. Hydromedusa maximiliani, Hydraspis radiolata; Brazil.—1889: Temnocephala.

breviplexus Staff., 1902, 901, t. h. Rana catesbiana, R. virescens; Canada.—1902: Hæmatolæchus. 1905: Pneumonæces.

brunnea Mueller, 1774, 54.—1774: Fasc. 1776: Planaria.

brunnea Dies., 1850a, 296, Cerc. III Baer, renamed; t. h. Limnæus stagnalis; Regiomontii.—1850: Cerc. 1855: C. (Eucerc.). 1858: C. (Gymnocephala.) [1858: Dist. echinata.]

brunnea var. Dies., see Erc., 1881e, 12.—[1881: Cerc. alba.]

brusinæ Stoss., 1889. 25, t. h. Oblata melanura; Triest.—1889: Dist. 1899: Pleurogenes. [1904: Diphterostomum (type).]

brusinai Looss, 1901, 399, for brusinæ 1889.—1901: Dist. 1903: D. (Brachycæcum). [1904: Diphterost. (type).]

buccatus Nicoll, 1907, 72, t. h. Hippoglossus vulgaris; Scotland.—1907: Stephanochasmus.

buccini Fil., 1855b, 23.—1855: Dist.

buccini mutabilis Fil., 1855b, 17, t. h. Buccinum mutabilis; Gulf of Genoa.—1855: Dist. 1855: Cerc. 1858: C. (Acanthocephala).

bucephalus Erc., 1881e, 40, t. h. Unio pictorum.—1881: Cerc.

buchholzii Braun, 1889a, 320, lapsus for "Fasc. hepatica, ovata plana, Buchholzii," see lanceatum.—1889: Fasc.

bufonis Linst., 1877, 185, t. h. Bufo vulgaris.—1877: Dist.

bulbosum Brand., 1888a, 67, t. h. Geronticus albicollis, Nauclerus furcatus; Mus. Vien.—1888: Holost.

bumpusii Lint., 1900a, 267, t. h. Dasyatis centrura; Woods Hole, Mass.—1900: Epibdella. 1903: Phylline.

bursæ fabricius Podwyssozki, 1890, 157, t. h. Gallus domesticus.—1890: Dist.

bursarium Nitzsch, in Giebel, 1857, 265, t. h. Falco peregrinus.—1857: Holost.

bursicola Crep., 1837, 310, t. h. Ardea cinerea.—1837: Dist. [Echinost. cloacinum.]

bursicola Odhn., 1900, 14, t. h. Somateria mollissima; Sweden.—1900: Gymnophallus.
 bursicola Looss, 1899b, 694, t. h. Milvus parasiticus, Falco tinnunculus; Africa. See also africanum 1901.—1899: Echinost.

bursigerum Brand., 1888, 65, t. h. Larus ridibundus; Mus. Vien.—1888: Holost.

buski B1., 1888a, 622, for buskii q. v.—1888: Dist. 1888: Dicroccelium. 1893: D.
 (Dicroccelium). 1895: Dist. 1895: Opisthorchis. 1902: Fasciolopsis.

buskii Lankester, 1857, 437, t. h. Homo; Asia.—1857: Dist. 1858: Dicroccelium. 1895: Opisthorchis. 1901: Fasciolopsis [type].

buteonis Gmelin, 1790a, 3054, t. h. Falco buteo; Europe.—1790: Fasc. [1782: Plan.]. 1803: Dist.

cacozelus Nicoll, 1907, 72, t. h. Pleuronectes limanda, Hippoglossus vulgaris.—1907: Derogenes.

caduceus Odhn., 1902, 26, t. h. Krokodil; Sudan.—1902: Oistosomum (type).

caducus Looss, 1901e, 603, t. h. Gadus minutus, Lophius piscatorius; apparently Triest.—1901: Stephanochasmus.

cæca Haswell, 1900b, 432, t. h. Phreatoicopsis n. sp.; Victoria.—1900: Temnocephala. [cæruleum Sluiter, 1898, 4, a tunicate (Dist.).]

cahirinum Looss, 1896, 119, t. h. Bagrus bayad; Cairo, Egypt.—1896: Dist. 1899: Haplorchis.

calceolus Mol., 1858, 129, t. h. Conger conger; Patavii.—1858: Dist. 1886: D. (Dicrocœlium).

calceostoma Wagener, 1857, 99, t. h. Sciæna aquila.—1857: Dactylogyrus. [1858: Calceost. elegans, type.] 1907: Calceost. type.

calicophorum Fischder., 1901, 370, t.h. Bos taurus; East Africa, Capland, Queensland, China.—1901: Paramphist.

calidris Rud., 1819a, 120, t. h. Scolopax calidris.—1819: Dist. [1850: Monost. mutabile.] [1886: D. brachysomum.]

caligarum Tasch., 1878, 564, for caligorum (Udonella).

caligi Kroyer, -—. Phylline. 1858: Udonella. [1858: Udonella caligarum.]

caligi Ben., 1858a, 189, for caligorum.

caligorum Johnston, 1835, 497, t. h. Caligus on Hippoglossus vulgaris.—1835: Udonella

callionymi Ben., 1870, 53, t. h. Callionymus dracunculus.—1870: Dist.

calyptrocotyle Mont., 1891, 110, t. h. Beroë ovata.—1891: Dist. 1893: D. (Accacœlium.) [1900: Accacœlium.] 1902: Orophocotyle.

campannlatum Rivolta, 1884, 27, for campanulatum (Dist.).

campanula Duj., 1845a, 435, t. h. Esox lucius; Rennes.—1845: Dist. (Crossodera). [1850: D. nodulosum.] 1860: Crossodera. [1858: Gasterost. fimbriatum.] 1886: D. (Echinost.).

campanula Linst., 1886a, 125, for campula Cobbold.—1886: Dist. [1892: D. oblongum.]

campanulatum Erc., 1874, 432, t. h. Canis familiaris.—1874: Dist.

campula Cobbold, 1876, 40, oblonga (Campula) renamed.—1876: Dist. 1899: Metorchis. 1899: Opisthorchis.

canaliculatum Rud., 1819a, 676, t. h. Sterna sp., galericulata teste Dies.; Brazil.—1819: 1902: Bilharziella.

canaliculatum Mehlis in Crep., 1846, 138, t. h. Colymbus cristatus.—1846: Dist.

cancri locustæ Rud., 1810a, 288.—1878: Trematodum.

candida Mueller (or Abildg.), 1806a, v. 4, 32.—1806: Strigea. [1850: Holost. erraticum.] candida Mueller, 1774, 71, in littore Grænlandiæ sub lapidibus.—1774: Fasc. Planaria. —: Tetrastemma.

canicula Cerf., 1899a, 374, Onchocotyle appendiculata Kuhn of Stoss., 1877, renamed; t. h. Scyllium canicula.—1899: Acanthonchocotyle.

canis Cerf., 1899a, 375, t. h. Galeus canis; Roscoff.—1899: Squalonchocotyle. 1900: Onchocotyle.

canthari Ben. & Hesse, 1863, 113; 1864, 113, t. h. Cantharus griseus.—1863: Microcotyle.

caouanæ Koll. in Braun, 1901b, 23, t. h. Thalassochelys caouana, int.—1901: Monost. [1901: Enodiotrema megachondrum.]

capense Looss, 1902m, 855, "Anaporrhutum ricchiardii Lopez" of Ofenh., renamed.— 1902: Probolitrema.

capense Harley, 1864a, 55, t. h. Homo; Cape of Good Hope.—1864: Dist. 1864: Bilharzia.

capitala Bosc, 1802a, v. 1, 261; Baltic Sea.—1802: Planaria.

capitata Mueller, 1774, 70.—1774: Fasc. 1776: Planaria.

capitellatum Rud., 1819a, 83, 343, t. h. Sparus salpa; Naples.—1819: Monost. (Monost.).

capitellatum Rud., 1819a, 99, t. h. Uranoscopus scaber; Arimini and Naples.—1819: Dist. 1900: Anisocœlium (type).

capriciosa Cuénot, 1892, in 1-23, t. h. Synapta inhærens teste Braun, 1893b, 183.—1893:

capsulare Dies., 1858e, 355, based on Wedl, 1857, v. 26, 247, pl. 1, fig. 8, t. h. Ardea purpurea, A. nycticorax, A. cinerea, Gallinula crex, Podiceps nigricollis.—1858: Dist. 1892: Agamodist.

capsularia Sons., 1892, 7 Oct., 144, t. h. Cleopatra bulimoides; Cairo, Egypt.—1892: Cerc.

capyristes Klein, 1905, 60, t. h. Rana hexadactyla.—1905: Pneumonœces.

carbonarii Cerf., 1895h, 929.—1895: Dactylocotyle.

carinariæ delle Chiaje, (1841a), 139.—1841: Dist.

carinatum Zed., 1803a, 217, cyprinaceum Zed., 1800, renamed.—1803: Dist. [1809: D. globiporum, pars D. inflexum.]

carnea Rathke, 1799, 83.—1799: Planaria.

carnosa Hass., 1891a, 208, in Bos taurus; U. S. A.—1891: Fasc. 1892: Dist. 1892: D. (Fasc.). [See americanum.]

carnosum Rud., 1819a, 93, t. h. Sparus dentex: Naples.—1819: Dist. 1886: D. (Brachylaimus).

carolinæ Stoss., 1889, 26, t. h. Alausa finta; Triest.—1889: Dist. [1891: Apoblema ocreatum.] 1899: Pronopyge. [1899: Pronopyge ocreata, type.]

caryocatactis Zed., 1800a, 163, t. h. Corvus caryocatactes; Europe.—1800: Dist. [1850: D. caudale.] [1902: Harmost. caudale.]

caryophyllacea Rud., 1810a, 353 for caryophyllina (Festucaria).

caryophyllata Bory de St. Vincent, 1823a, 354, in infusions de chènevis.—1823: Cerc.

caryophyllina Rud., 1802, 66, t. h. Gasterosteus aculeatus: Greifswald.—1802: Festuc. 1803: Monost. 1809: M. (Hypost.). 1828: Hypost. (probably type). 1845: Monost.

caryophyllum Dies., 1850a, 317, t. h. Falco pileatus; Rio Parana, Brazil.—1850: Eustemma (type). [1888: Holost. eustemma.]

caryophyllum Mont., 1892, 717, for caryophyllinum (Monost.).

catellina Mueller, 1786, 130, in aqua fossarum, ubi Lemna.—1786: Cerc. 1815: Furcocerca. 1827: Dicranophorus.

catellus Mueller, 1773, 65, in aqua [etc., see p. 124.].—1773: Cerc. 1815: Furcocerca. 1827: Dicranophorus.

catervarium Looss, 1896, 118, t. h. Alosa finta; Cairo, Egypt.—1896: Dist.

cattoi R. Bl., in Catto, 1905, 70; 1905, 11, t. h. Homo; China.—1905: Schistosoma. [S. japonicum.]

caudale Rud., 1809a, 382, caryocatactis Zed., 1800, renamed, t. h. Corvus caryocatactes.—1809: Dist. 1901: Harmost.

caudale from Pyrrhocorax alpinus.—1809: Dist. [1902: Harmost. mesostomum.]

caudale of Mueller, 1897, 16, in Coracias garrula.—1897: Dist. [1902: Urogonimus macrostomus.]

caudata Goto, 1894, 186, t. h. Sebastes sp.; Japan.—1894: Microcotyle.

caudata Bosc, 1802a, 271, t. h. dorade.—1802: Fasc. [1809: Dist. coryphænæ.] [1819, 1850: D. tornatum.] 1859: Dist.

caudata Mueller, 1774, 70.—1774: Fasc. 1787: Planaria.

caudatum Polonio (1859), t. h. Natrix torquata, Tropidonotus viperinus; Padua.—1859: Dist. 1896: D. (Opisthorchis).

caudatum Linst., 1873, 103, t. h. Erinaceus europæus.—1873: Dist. 1892: D. (Brachylaimus.) 1899: Heterolope. [1899: D. leptostomum.]

candiporum Rud., 1819a, 96, t. h. Zeus faber; Arimini.—1819: Dist. 1845: D. (Apoblema). 1901: Lecithochirium. 1907: Synaptobothrium.

cavatica Fries.—Planaria.

cavix Bosc, 1811, 269.—1811: Tetragulus. 1829: Linguatula.

caviæ Sons., 1890, 100, t. h. Cavia cobaya.—1890: Dist. 1893: Fasc. hepatica.

cellulosa Looss, 1896b, 227, t. h. Melania tuberculata Bourg.; near Alexandria, Egypt.—1896: Cerc.

centra appendiculatum Leidy, 1904a, 277, misprint for centrappendiculatum (Dist.). centrappendiculatum Leidy, 1891a, 416, appendiculatum Leidy, 1877, 202 [not Rud.] renamed.—1891: Dist.

centrodes Braun, 1901g, 941, t. h. Tinamus variegatus; Brazil.—1901: Harmost.

cephala Risso, 1826, 262, t. h. Tetraodon luna; Europe.—1826: Trist. [1898: T. molæ.]

cephalo Kroyer, 1852–53a, 745, for cephala (Trist.).

cercatum Ben., 1858a, 179 [possibly lapsus for ——? ——].—1858: Monost.

cercatum Mont., 1893, 40, host and loc. unknown.—1893: Urogonimus. 1893: Dist.

cercatus Mont., 1893, 162, see cercatum (Urogonimus).

cerco pitheci Cobbold, 1861e, 119, t. h. Cercopithecus fuliginosus.—1861: Cerc.

cerebrale Yamagiwa, 1890, 457, t. h. Homo; see westermanii.—1890: Dist. [Paragonimus westermanii.]

cervi Schrank, 1790, 123.—1790: Fasc. [1790: Festucaria.] 1893: Strigea. 1898:
 Amphist. 1901: Paramphist. (type).

cervi Zed., (1790), 65, t. h. Hirsch.—1790: Festucaria. See cervi Schrank.

cervi Gmelin, 1790a, 3054.—1790: Fasc. hepatica.

cervi Cobbold, 1861e, 119, t. h. Cervus axis.—1861: Cerc.

cesti veneris Vogt, —, 299, t. h. Cestum veneris.—Dist.

certicillus Mol., 1858, 131, t. h. Lophius piscatorius; Patavii.—1858: Dist. 1886: D. (Echinost.). 1893: Echinost. 1899: Anoiktost. 1899: Stephanost. (type). [1901: Stephanochasmus, type.]

cestoides E. Ben., 1870, 17, t. h. Raja batis.—1870: Dist.

eglindraceum Looss, 1894a, 37, misprint for cylindraceum (Dist.).

characis Stoss., 1886, 26, t. h. Charax puntazzo; Triest.—1886: Dist. (Brachylaimus).
 chefrenianum Looss, 1896, 73, t. h. Rhinopoma microphyllum; Ghizeh.—1896: Dist.
 1899: Lecithodendrium.

chefresianum Looss, 1896, pl. 5, misprint for chefrenianum (Dist.).

cheloniæ atræ Braun, 1899e, 629, see pachyderma.—1899: Dist.

cheloniæ imbricatæ Dies., 1858e, 358, based on Bellingham, 1844, 340, t. h. Chelonia imbricata; Ireland.—1858: Amphist.

chelydræ Staff., 1900, 406, t. h. Chelydra serpentina.—1900: Dist. 1905: Auridist. (type).

chelydræ MacCallum, 1902, 632, t. h. Chelydra serpentina; Dunnville, Ontario.—1902: Heronimus (type).

chiajæ Tasch., 1879, 251, for chiajei (Solenocotyle).

chiajeæ Mont., 1888, 89, for chiajei (Solenocotyle).

chiajei Dies., 1850a, 420, Polyst. loliginis Chiaje, 1823, renamed; t. h. Loligo vulgaris; Naples.—1850: Solenocotyle (type).

chilensis Gay, 1836, see next entry.—1836: Branchiobdella. 1850: Astacobdella.

chilensis Moquin-Tandon, teste E. Bl., 1849, 51, t. h. cangregos; Chili.—Branchiobdella. 1849; Temnocephala.

chili Osborn, 1903, 315, t. h. Micropterus dolomieu; Chautauqua, N. Y.—1903: Cryptogonimus (type).

chilinsis Mont., 1899, 113, for chilensis (Temnocephala).

chilostomum Mehlis, 1831, 186, t. h. Vespertilio murinus et al.—1831: Dist. 1892:
 D. (Dicrocœlium). 1900: Lecithodendrium.

chilostomum Ben., 1873, 27.—1873: Dist. [1892: D. aristotelis.]

chimæræ Ariola, 1899a 8, t. h. Chimæra monstrosa; Genova, Italy.—1899: Agamodist.
chimæræ Kroyer, 1852–53a, 813, t. h. Chimæra monstrosa L.—1853: Crobylophorus (type).

chinense Cobbold, 1876, 97, D. sinense renamed, t. h. Homo.—1876: Dist.

chiri Goto, 1894, 193, t. h. Chirus hexagrammus; Japan.—1894: Microcotyle.

chloropodis Zed., 1800a, xvii, 164, t. h. Fulica chloropus; Europe.—1800: Dist. [Dist. uncinatum.]

chlorotica Dies., 1850a, 296, Cerc. II Baer renamed; t. h. Paludina vivipara; Regio. montii.—1850: Cerc. 1855: C. (Eucerc.). 1858: C. (Acanthocephala).

choledochum Linst., 1883a, 306, t. h. Anas sp.—1883: Dist. 1892: D. (Dicrocœlium). 1898: Opisthorchis.

choledochus Odhn., 1900, 14, t. h. Vulpanser tadorna.—1900: Gymnophallus.

chordale Burckhardt, 1891a, 62, t. h. Protopterus annectens.—1891: Amphist. [1892: Tetracotyle, Holostomidæ.]

chryophrii Cerf., 1898a, 303, for chrysophryi (Choricotyle).

chrysophri St.-Remy, 1898, 555, for chrysophryi (Diclidophora).

chrysaëti Rud., 1819a, 119, t. h. Falco chrysaëtus.—1819: Dist. [1892: D. crassius-culum.]

chrysophris Mont., 1888, 11, 16, for chrysophryi (Choriocotyle).

chrysophryi Ben. & Hesse, 1863; 1864, 109, t. h. Chrysophrys aurata.—1864: Choricotyle (type). 1890: Octobothrium. 1890: Microcotyle. 1895: Diclidophora.
chrystallinum Hannover, 1864a, 3, for crystallinum 1819 (Dist.).

cignoides Mont., 1888, 40, 80, for cygnoides (Dist.).

ciliata Mueller, 1774, 55, free form.—1774: Fasc. 1776: Planaria.

cimbiforme Mont., 1896, 165 for cymbiforme (Dist.).

cincta Rud., 1803, 31, t. h. Tringa vanellus.—1803; Fasc. 1809; Dist. 1809; D. (Echinost.). 1860; Echinost.

cinctum Brand., 1888a, 67, t. h. Ardea sp.; Brazil.—1888: Holost.

[cinerea (anatomical term) Fasc.]

circularis Linst., 1904, 493; or 1894, 17, t. h. Acipenser ruthenus.—1904: Erpocotyle. [circumvallata Sluiter, 1900. Dist. (tunicate).]

cirrata Rud., 1802, 66, of Rud., 1808, xxv, for F. cirrhata.—1802: Fasc. [cirrhata].
 1809: Dist. 1845: D. (Brachylaimus). 1899: Lepoderma. 1899: Plagiorchis.
 cirrhata Rud., 1802, 66, t. h. Coryus frugilegus; Greifswald, June—1802: Fasc. 1855;

Dist. 1899: Lepoderma.

cirrhigerum Nord., 1840, 616, for cirrigerum (Dist.).

cirrigerum Baer, 1827, 553, t. h. Astacus fluviatilis.—1827: Dist. [1850: D. isostomum.]

cirrosum Rud., 1808a, 296, for cirratum (Dist.).

cladocalium Dies., 1858, 354, t. h. Ardea minuta; based on Pontallié, 1853, 103.—1858: Dist.

clathratum Deslongchamps, in Lamouroux, 1824, 563, t. h. Cypselus apus.—1824: Dist.
1845: D. (Dicrocœlium). 1899: Dicrocœlium. [1899: Lyperosomum.]

clathratum Deslongchamps, of Olsson, 1876, 24, renamed olssoni 1900.—1876; Dist. [1900: Dicrocœlium olssoni.]

clathratum Dies., 1850a, 308, t. h. Lutra brasiliensis; Matogrosso.—1850: Hemist.

clausi Mont., 1888, 79 for clausii (Cerc.).

clausii Mont., 1888, 72.—1888: Cerc.

clava Dies., 1850a, 356, hosts Eunectes scytale, Hydroscopus plumbeus, Coluber flaviventris, Clœlia fasciata; Brazil.—1850: Dist. 1899: Telorchis (type).

clavata Menzies, 1791, 187, t. h. Scomber pelamys; Pacific.—1791: Fasc. 1808: Dist. 1828: Hirudinella (type).

clavatum Steenstrup, 1842, 59, under skin of frogs, Rana temporaria, [for 1842: Amphist. subclavatum?].

clavatum Nord., 1832, 42, t. h. Perca cernua, P. fluviafilis, P. lucioperca.—1832: Diplost. 1850: Tylodelphys (type).

claviforme Brand., 1888, 247, t. h. Tringa alpina.—1888: Dist. 1899: Lecithoden-drium. 1892: D. (Brachycœlium). 1907: Spelotrema.

clavigerum Zed., 1803a, 199, Festuc. strigis 1788, renamed; in Strix.—1803: Amphist. [1809: A. macrocephalum.]

clavigerum Rud., 1819a, 103, t. h. Bufo viridis, at Berlin; B. cinereus, Rana temporaria, R. esculenta, Hyla arborea.—1819: Dist. 1845: D. (Brachycælium). 1899: Pleurogenes (type).

calvigerum Rud. of Duj., 1845a, renamed confusus, 1894.—1845: Dist. (Brachycœlium). 1899: Prosotocus, type.

clavus Mol., 1858, 128, t. h. Gadus merlucius; Patavii.—1858: Holost.

cloacinum Braun, 1901, 259, bursicola Crep., renamed.—1901: Echinost.

clupeæ Ben., 1870, 67, t. h. Clupea sprattus; Belgium.—1870: Gasterost.

clupeæ Schrank, 1788, 20, t. h. rheinische Mayfische.—1788: Fasc. 1803: Dist.

clupeæ rhenanæ Rud., 1809a, 437, includes Fasc. alosæ Hermann, 1783, F. clupeæ Schrank, 1788.—1809: Dist.

cobboldi Fischder., 1901, 372, for cobboldii (Gastrothylax).

cobboldi Montgomery, 1906, Feb. 12, 21, t. h. Equus caballus; India.—1906: Pseudodiscus.

cobboldii J. Poir., 1883, 77, t. h. Palonia frontalis; Java.—1883: Gastrothylax.

cobitidis Linst., 1890d, 179, t. h. Cobitis barbatula.—1890: Diplost.

coccineum Cuv., 1817, 42, t. h. la môle, le xiphias, etc.—1817: Trist. (type). 1820: Phylline. 1828: Capsala.

coccineum of Rud., 1819a, 123, in Orthagoriscus molæ, Xiphias gladius, renamed rudolphianum, 1850.—1819: Trist.

cochlea Wedl, 1857, 258, t. h. Esox lucius.—1857: Gyrodactylus. [1888: Tetraonchus monenteron.] 1890: Dactylogyrus.

cochlear Dies., 1858, 38, t. h. Scomber scombrus.—1858: Grubea (type).

cochlear Dies., 1850a, 357, D. cochleariforme sternæ Rud. renamed; t. h. Sterna cantiaca, S. minuta, int.; Brazil.—1850: Dist. 1901: Microlistrum.

cochlear Looss, 1899b, 667, t. h. Chelonia mydas; Egyptian coast.—1899: Pyelosomum (type).

cochleare Fischer, 1883, 1, t. h. Halicore dugong; Philippines.—1883: Opisthotrema (type).

cochleariforme Rud., 1809a, 326, includes Festuc. cyprinacea, t. h. Cyprinus barbus.— 1809: Monost. (Hypost.).

cochleariforme Rud., 1819a, 681, t. h. Pelecanus aquila, Sterna minuta; Brazil.—1819: Dist. 1901: Microlistrum (type).

cochleariforme sternæ Dies., 1850a, 357, see cochlear.—1850: Dist.

cochleariformis Dies., 1838a, 189, t. h. Cataphractus.—1838: Aspidocotylus. 1877: Cotylegaster. 1879: Aspidogaster. 1892: Aspidocotyle.

cocinea Tasch., 1878, 567, for coccinea (Phylline).

calebs Linst., 1875, 192, t. h. Fringilla calebs.—1875: Dist. 1892: Agamodist.

cœlomaticum Giard & Billet, 1892a, 613, t. h. cattle; Tonkin.—1892: Dist. 1896: Dicrocœlium. 1893: D. (Dicrocœlium). 1907: Eurytrema.

cœruleus Braun, 1902b, 11, t. h. Cairina moschata; Brazil.—1902: Metorchis.

coleostomum Looss, 1896, 101, t. h. Pélican; Egypt.—1896: Dist. 1899: Anoiktost. 1899: Ascocotyle (type).

collinsi Sons., 1895, 182, for collinsii, Amphist. (Pseudodiscus).

collinsii Cobbold, 18751, 741, t. h. Equus caballus; India.—1875: Amphist. [1895: A. (Pseudodiscus).]

collinsii var. stanleyi Cobbold, 1879b, 359, t. h. Equus.—1879: Amphist. [1900: A. hawkesi.] [See stanleyii 1875.]

collurionis Schrank, 1790, 123, t. h. Lanius collurio.—1790: Fasc. 1803: Dist.

colostomum Vaullegeard, 1901, 143, for coleostomum (Dist.).

colubri Bosc, 1802, 271, t. h. couleuvre d'Amérique.—1802: Fasc. 1809: Dist. [1859: D. bosci.]

colubri Linst., 1877, 192, t. h. Coluber natrix.—1877: Tetracotyle. [1889: Holost. variabile.]

colubri americani Rud., 1819a, 121, D. colubri 1809, renamed.—1819: Dist. [1859: D. bosci.]

colubri murorum Rud., 1819a, 121, t. h. Coluber murorum.—1819: Dist. [1850: D. allo-stomum.]

colubri natricis intestinale Rud., 1809a, 433, t. h. Coluber natrix.—1809: Dist. [1819: D. mentulatum.]

colubri natricis pulmonale Rud., 1809a, 434.—1809: Dist. [1819: D. naja.]

colubri tesselati Rud., 1819a, 121, t. h. Coluber tesselatus.—1819: Dist. [1850: D. mentulatum.]

columbæ Mazzanti, 1889a, 161, t. h. Columba livia; Pisa.—1889: Dist. [1896: Mesogonimus commutatus.]

columbæ liviæ Mueller, 1897, 26, t. h. Columba livia.—1897: Echinost.

columbellæ Pag., 1862, 306, t. h. Columbella rustica.—1862: Cerc.

colymbi immeris Viborg, 1795, 241.—1795: Fasc. s. Ligula.

comes Haswell, 1893e, 96, t. h. Astacopsis serratus; Australia.—1893: Temnocephala.

cometa Bory de St. Vincent, 1823a, 354, in infusions d'orge.—1823: Cerc.

commune Olss., 1876, 31, t. h. Labrus.—1876: Dist. 1886: D. (Dicrocœlium). 1901: Allocreadium.

communis Odhn., 1905, 348, in numerous Scandinavian marine fishes.—1905: Hemiurus.

commutatum Dies., 1858e, 339, Dist. dimorphum Wagener, 1852 [not Dies., 1850], renamed, t. h. Phasianus gallus; Pisa.—1858: Dist. 1889: Mesogonimus. [1899: Clinost.]

commutatum Dies., 1850a, 311, t. h. Sterna caspica; M. C. V.—1850: Hemist.

compactum Ccbbold, 1859d, 363, t. h. Viverra mungos.—1859: Dist. 1892: Mesogonimus. [1899: Paragonimus.] 1900: Paragonimus.

compactus Fischder., 1901, 370, t. h. Bos taurus; Africa.—1901: Stephanopharynx (type).

compascua Kowal., 1898, 72, see xanthosoma compascua.—1898: Opisthorchis. 1902: Metorchis.

complanatum Rud., 1814a, 103, t. h. Ardea cinerea; Berlin, Germany.—1814: Dist. 1845: Dist. (Dicroccelium). 1899: Clinost.

complanatum Erc. of Par., 1894, 144, for campanulatum (Dist.).

complexum Stiles & Hass., 1894e, 425, t. h. Felis catus dom.; U. S. A.—1894: Dist. (Dicrocœlium). 1896: Opisthorchis. 1899: Metorchis.

complexus Seely, 1906, 249, t. h. Rana pipiens; North Carolina, U. S. A.—1906: Pneumonœces.

complicatum Mehlis, 1846, 141, t. h. Haliaëtus carbo.—1846: Dist.

compressus Brand., 1898a, 27, t. h. Bos indicus; Vien. Mus.—1898: Gastrothylax.

concavum Crep., 1825a, 45, t. h. Colymbus rufogularis.—1825; Dist. 1892; D. (Dicrocœlium). 1899; Tocotrema. 1899; Cryptocotyle (type). 1899; Cotylogonimus (Cryptocotyle, type). 1903; Cryptocotyle.

conchicola Baer, 1826a, 124, t. h. Anadonta, Unio; Prussia.—1826: Aspidogaster (type).

1851: Aspidonotus (type).

conchicole Gronkowski, 1902a, 515, for conchicola (Aspidogaster).

conchiola Mont., 1888, 40, for conchicola (Aspidogaster).

conchycola Dies., 1850a, 414, for conchicola (Aspidogaster).

concinna Scott, 1904, 278, t. h. Trygon pastinaca; Scotland.—1904: Thaumatocotyle (type).

concinna Scott, 1902, 301; t. h. not given.—1902: Acanthocotyle.

concinnum Braun, 1901, 700, t. h. Viverra zibetha.—1901: Dicrocælium.

confusum Looss, 1894a, 2, 101, D. clavigerum of Duj., 1845a, renamed; t. h. Rana esculenta; Paris, Rennes.—1894: Dist. 1899: Prosotocus (type). 1905: Pleurogenes.

confusus Odhn., 1905, 357, bothryophorus of Looss, 1899, 728, renamed; t. h. Alosa finta in Mediterranean and Clupea harengus in North Sea.—1905: Lecithaster (type by Odhn.).

coni mediterranei Fil., 1857c, 14, t. h. Conus mediterraneus; Mediterranean.—1857: Cerc. 1858: Cercariæum.

conicum Polonio, 1859, teste Par., 1894, 627.—1859: Dist.

conicum Polonio, 1859, t. h. Natrix torquata; Padua.—1859: Diplodiscus.

conicum Zed., 1803a, 188, Festucaria cervi Zed., 1790, renamed; t. h. Cervus elaphus.—1803: Monost. 1809: Amphist. [1901: Paramphist. cervi, type.]

coniferum Mehlis in Creplin, 1846, 138, t. h. Colymbus cristatus.—1846: Holost.

conis Perroncito, 1886, 250, misprint for conus Crep., 1825.—1886: Dist.

coniunctum Rivolta, 1884, 26, for conjunctum, 1860 (Dist.).

conjunctum Cobbold, 1860a, 8, t. h. American Canis fulvus.—1860: Dist. 1893: D. (Brachylaimus). 1895: Opisthorchis. 1899: Metorchis.

conjunctum of Lewis & Cunningham, as a parasite of man, in India, see noverca.— 1872: Dist. 1895: Opisthorchis.

conoideus Bloch, 1782a, 35, t. h. Anas boschas dom.; Europe.—1782: Cuculanus. [1850: D. echinatum.] 1886: Dist. 1896: Echinost.

conostomum Olss., 1876, 17, t. h. Coregonus oxyrhynchus.—1876: Dist. 1886: D. (Brachylaimus). [1899: Spathidium.] 1902: Phyllodist.

constrictum Dies., 1850a, 322, t. h. Abramis brama.—1850: Monost. 1892: Diplosto-mulum.

constrictum Leared, 1862, 271, t. h. edible turtle; see mistroides.—1862: Dist. 1896:
 Mesogonimus. 1899: Hapalotrema (type).

constrictum Mehlis, 1846, 142, t. h. Anas mollissima, A. nigra.—1846: Dist.

constructum Staff., 1900, 407, misprint for constrictum Leared (Dist.).

continuum Ariola, 1899, 6, t. h. Carcharias rondeletti; Genova.—1899: Dist.

contortum Rud., 1819a, 118, t. h. Orthagoriscus mola; Naples.—1819: D. (Echinost.) 1893. D. (Accacœlium). 1898: Podocotyle. 1899: Accacœlium (type).

contractus Looss, 1902, 136, t. h. Mugil chelo.—1902: Dicrogaster.

contribulans Braun, 1901f, 568, t. h. Hirundo rustica.—1901: Eumegacetes.

conum Erc., 1881a. or 1882, see Par., 1894. 164, t. h. Bythinia tentaculata; Bologna.— 1881 or 82: Cerc.

conum Huber, 1896a, 578, for conus Crepl. (Dist.).

conus of Gurlt. 1831. 193.—1831: Dist. [1903: Opisthorchis felineus.]

combs Crep., 1825, 50, t. h. Felis catus dom., Canis vulpes.—1825; Dist. 1892; D. (Dicroccelium). 1896; Opisthorchis.

conviva Luehe 1901, 474, t. h. Conger conger; Coll. Berlin.—1901: Lecithochirium.
convolutum Brand. MS., in Braun, 1901i, 565, t. h. Platalea ajaja.—1901: Dist. [1902: Mesaulus grandis.]

copulans Cohn, 1902, 877, t. h. Cryptobranchus japonicus.—1902; Liolope (type).
 copulans Linst., 1904, 254, t. h. Árnoglossus laterna; ? Louvain.—1904; Synaptobothrium (type).
 1906; Lecithochirium.

cordatum Dies., 1850a, 308, t. h. Felis cat. fer.; M. C. V.—1850; Hemist.

cordiforme Braun, 1900f. 389, or 1900b, 225, t. h. Molossus sp.; Brazil.—1900: Lecithodendrium.

cordiformis Wolf, 1903, 602. t. h. Squalus sp.—1903: Braunina.

coregoni feræ Chauvannes. —. 62; see Dies.. 1858d. 283.—1858: Cercariæum.

cornifrons Leidy. 1878, 382, t. h. Donax fossor.—1878: Dist.

cornu Dies., 1839a. 235. t. h. Doras sp.=Cataphractus vacu; Forte do Rio Branco; Brazil.—1839: Amphist. [1860: A. nattereri.]

cornu Nitzsch. in Rud., 1819a, 89, t. h. Ardea cinerea, A. garzetta; Europe.—1819: Amphist, 1819 [p. 357]: Holost.

cornu Linst., 1878, 228, t. h. Abramis vimba; middle Europe.—1878; Dactylogyrus.
 cornu Zed., 1800a, xvii. t. h. Ardea cinerea; Europe.—1800; Dist. 1809; Amphist.
 1819; Monost. (Monost.)

cornucopia Mol., 1859, 287. t. h. Strix flammea ?; Batavia.—1859: Holost.

cornuta Bosc, 1802a. v. 3. 244.—1802: Cerc.

cornuta Osborn, 1903, 63, t. h. crawfish, black bass, and catfish: Lake Chautauqua, N. Y.—1903: Bunodera, 1904: Crepidost.

cornutum Rud.. 1898a. xxv: 1809a. 343: t. h. Charadrius pluvialis: Greifswald, Europe.—1808: Amphist. 1845: Holost.

cornutum Verrill, 1875, 40, t. h. Tetrapterus albidus; North America.—1875: Trist.

coronarium Cobbold, 1861e. 119. t. h. Alligator mississippiensis.—1861: Dist. 1899: Anoiktost. 1896: D. (Echinost.). 1899: Acanthost. 1901: Acanthochasmus.

coronata Fil., 1855b, 10, t. h. Lymnæus palustris. L. stagnalis: Moncalier.—1855: Cerc. 1858: C. (Gymnocephala).

coronatum Leidy, 1888, 127, t. h. terrapin; eastern U. S. A.—1888; Polyst.

coronatum Rud., 1819a, 686, t. h. Didelphis ?) virginiana; Brazil.—1819: Dist. (Echinost.). 1850: Rhopalophorus (type). 1892: Echinost. 1898: Rhopalias (type).

coronatum Rentsch. 1860, 38. t. h. Gasterosteus spinachia.—1860: Dist.

coronatum Wagener, 1852, 567, t. h. Corvina nigra: Nice.—1852: Dist. 1886: D. (Echinost.). 1898: Echinost. 1899. Anoiktost. (type).

corones Dies., 1858e, 322, t. h. Corvus corone; Ireland.—1858: Holost.

corpulentum Lint., 1905, 327, t. h. Lagodon rhomboides. Orthopristis chrysopterus; Beaufort, N. C.—1905: Dist.

corrigia Braun, 1901. 946, t. h. Tetrao tetrix: Coll. Vien.—1901: Lyperosomum.

corrugatum Duj., 1845a, 409. t. h. Sorex tetragonurus: Rennes.—1845: Dist. Brachylaimus).

corvinæ Stoss., 1886, 46, t. h. Corvina nigra; Triest.—1886; Dist. 1886; D. (Echinost.).
[1899; Anoiktost. (type).]

corvinum Stiles & Hass., 1894, 418, t. h. Corvus americanus, C. ossifragus; U. S. A.— 1894: Dist. longissimum. 1898: Opisthorchis.

coryphænæ Bosc. 1802, 271. t. h. "dorade".—1802: Fasc. 1809: Dist. [1850: D. clavatum: D. tornatum.]

coryphænæ hippuridis Tilesius in Litt. Rud.. 1809a. 436.—1809: Fasc. [1809: Dist. coryphænæ.]

coryphenæ Cobbold, 1879b, 460, for coryphænæ (Dist.).

coryphænæ Cobbold, 1879b, 461, for coryphænæ (Dist.).

[costæ D. Valle (Dist.), a tunicate.]

cotti Linst., 1889a, 80 based on Zschokke, 1884, 204, t. h. Cottus gobio; Lake Leman.—1884: Monost.

cotylophorum Fischder, 1901a, 370, t. h. Bos taurus at Togo, B. zebu in German East Africa.—1901: Paramphist.

cotylura Pag., 1862, 293, t. h. Trochus cinereus; Cette.—1862: Cerc.

craniaria Dies., 1858, 316, t. h. Cobitis fossilis.—1858: Tylodelphys. 1860: Diplost.

cranum Huber, 1896a, 575, misprint for crassum (Dist.).

crassa Rud., 1793a, 27, F. melis Gmelin renamed.—1793: Fasc.

crassa Sons., 1888, 124.—1888: Bilharzia. 1892: Gynæcophorus. 1899: Schistosoma.
 crassa Erc., 1881 or 1882, see Par., 1894, 164, t. h. Bythinia tentaculata; Bologna.—1881: Cerc.

crassicauda Erc., 1881 or 1882, see Par., 1894, 164, t. h. Bythinia tentaculata; Bologna.—1881: Cerc.

crassicaudatum Busch, 1851, 99, t. h. Sagitta.—1851: Dist.

crassicaudatum Leuck. in Kollar, 1836, 81, t. h. Acipenser stellatus.—1836: Diklibothrium. 1839: Diclibothrium. [1840: Hexacotyle elegans.] [1850: D. armatum.]

crassicolle Rud., 1809a, 378, for Fasc. salamandræ; t. h. Salamandra atra.—1809: Dist.
1845: Dist. (Brachycælium). [1896: Brachycæcum, type.] 1899: Brachycælium (type). 1899: Dist. (Dicrocælium). 1899: Lecithodendrium.

crassiusculum Rud., 1809a, 408, Planaria bilis Braun, renamed; t. h. Falco melanaëtus.—1809: Dist. 1892: D. (Brachylaimus). 1898: Opisthorchis. 1899: Metorchis.

crassiuscula var. janus Kowal., 1898h, 122, t. h. Anas boschas dom.—1898: Opisthorchis. 1898: Campula. [1899: Metorchis.]

crassiusculus Wedl, 1857, 258, t. h. Lucioperca sandra.—1857: Gyrodactylus. [1858: Tetraonchus unguiculatus.] 1890: Dactylogyrus.

crassum Cobbold, 1860a, 5, buskii renamed, t. h. Homo.—1860: Dist. 1899: Fasciolopsis.

crassum Olss., 1876, 25, t. h. Hirundo urbica.—1876: Dist.

crassum Sieb., 1836, 234, t. h. Hirundo urbica.—1836: Dist. 1892: D. (Dicroccelium). 1907: Eumegacetes.

crassus Looss, 19011, 568, t. h. Thalassochelys corticata; Egypt.—1901: Glyphicephalus. 1902: Epibathra (type).

crenata Rud., 1802, 76, t. h. Gasterosteus aculeatus, Pleuronectes maximus.—1802: Fasc. 1809: Dist. 1889: Apoblema. 1901: Hemiurus. 1905: Brachyphallus (type).

crenata Frœlich, 1802a, 60, t. h. Fulica chloropus; Europe.—1802: Fasc. [1814: Dist. uncinatum.]

crenata Mueller, 1774, 64.—1774: Fasc. 1776: Planaria.

crenatum Mol., 1859, 840, t. h. Centrolophus pompilius; Batavii.—1859: Dist. 1886: D. (Apoblema). 1889: Apoblema. 1899: Hemiurus. [1901: Lecithocladium excisum.] 1905: Brachyphallus.

crenulatum Rud., 1809a, 328, t. h. Motacilla phœnicurus; Greifswald.—1809: Monost. (Monost.).

crenulatum Cobbold, 1860a, 47, t. h. Anas nigra; Ireland.—1860: Holost.

cristallina Sons., 1893, 188, for crystallina Rud., 1819 (Tetracotyle).

[cristallinum Ren. (Dist.), tunicate.]

cristata La Valette, 1855, 23, t. h. Lymnæus stagnalis.—1855: Cerc. [1858: Lophocercaria fissicauda.]

cristatum Rud., 1819a, 117, t. h. Stromateus fiatola; Arimini.—1819: Dist. (Echinost.). 1860: Echinost.

croaticum Stoss., 1889, 183, t. h. Carbo graculus; Fiume, Croazia.—1889: Dist. 1892: Echinost.

crocodili Poir., 1886, 30, t. h. Crocodilus siamensis.—1886: Dist. 1895: Echinost.

crotali "Humboldt."—Dist., an arachnoid [now in Porocephalus].

crotali durissi Rud., 1809a, 433, for crotali.—1809: Dist. [1819: Pentast. proboscideum.]

cruciatus Wedl, 1857, 258, t. h. Cobitis fossilis.—1857: Gyrodactylus. 1858: Tetra-onchus. 1890: Dactylogyrus.

crucibulum Rud., 1819a, 83, t. h. Muræna conger, M. cassini (M. myroides); Naples.—1819: Monost. (Monost.). 1845: Dist. (Crossodera). 1850: Monost. 1859: Gasterost. 1905: Prosorhynchus.

crucifer Wagener, 1857, 55, t. h. Cyprinus erythrophthalmus.—1857: Dactylogyrus.

crumena Mueller, 1786, 129, in infuso Ulvæ linzæ marino.—1786: Cerc. 1815: Furcocerca. 1827: Crumena, type.

crumenifer Otto, 1896, 94, see crumeniferum.—1896: Gastrothylax.

crumeniferum Crep., 1847, 30, t. h. Bos taurus indicus.—1847: Amphist. 1883: Gastrothylax (type).

crumigerum Fischder., 1903h, 563.—1903: Amphist. [1903: Gastrothylax compressus.]

crux Levin., 1881, 80, t. h. Modiolaria discors; Egedisminde.—1881: Bucephalus.

cryptobothrium Ben., 1870, 1871a, 31, t. h. Trigla gurnardus; Belgium.—1870: Dist.
 crystallinum Rud., 1819a, 100, hosts Rana esculenta, R. temporaria, Bufo viridis,
 B. igneus, Vipera berus; Berlin.—1819: Dist. 1877: (ex parte) Tetracotyle.

cteniceps Leidy MS. in Stiles & Hass., 1894, 249, t. h. Fiber zibethicus.—1894: Dist. ctenolabri Staff., 1905, 682, t. h. Ctenolabrus adspersus.—1905: Dermocystis (type). cucullus Ziegler, 1883, 540, for cuculus (Bucephalus).

cuculus McCrady, 1874, Dec. 3, 176, t. h. Ostrea virginiana; Charleston, S. C.—1874: Bucephalus. [1874: Hydricuculus, type.]

cucumerina Erc., 1881, see Par., 1894, 364, t. h. Bythinia tentaculata; Bologna.—1881: Cerc.

cucumerina Rud., 1804, 166 in liver of Pleuronectes maximus.—1804: Fasc.

cucumerinum Rud., 1809a, 360, host Avis riparia, gen. sp. incert.—1809: Dist. 1889: Monost. 1902: Typhlocœlum.

cuneatum Rud., 1809a, 358, t. h. Otis tarda; Greifswald.—1809: Dist. 1901: Prosthogonimus.

cuspidatum Looss, 1896b, 96, t. h. Milvus parasiticus; Matarieh.—1896: Dist. 1899: Anoiktost. 1899: Centrocestus (type).

cuticola Nord., 1832, 49, t. h. Cyprinus, see p. 290.—1832; Holost. [1832; Cryptostomum.] 1850; Diplost. 1892; Tetracotyle. [1898; Hemist. denticulatum.] 1902; Tetracotyle.

cuticula Kroyer, 1852–53a, 1250, for cuticola (Holost.).

cycladis rivicolæ Dies., 1850a, 298, based on Sieb., 1837, 388, t. h. Cyclas rivicola.—
1850: Cerc. 1855: Cercariæum.

cyclidium Mueller, 1773, 68, in Aquis purioribus frequens.—1773: Cerc. 1827: Cyclidium, type.

cyclophora Braun, 1896, 3, t. h. Notothenia sp.; Navarin, Puerto Toro.—1896: Lophocotyle (type).

cyclopteri Rud., 1809a, 438, t. h. Cyclopterus lumpus.—1809: Dist. [1850: D. reflexum.]
 cygni oloris Dies., 1858e, 344, based on Bellingham, 1844, 427.—1858: Dist. [1858: D. echinatum.]

cygnoides Zed., 1800a, 163, t. h. Rana esculenta.—1800: Dist. 1845: D. (Dicroccelium.) 1889: D. (Polyorchis). 1898: Pleorchis. 1899: Phyllodist. 1899: Gorgodera (type).

cygnoides Schrank, 1803, 212, t. h. ——.—1803: Fasc.

cygnoidis Sons., 1893, 187, for cygnoides (Dist.).

cylindraceum Zed., 1800a, 163, t. h. Rana esculenta.—1800: Dist. 1802: Fasc. 1845: Dist. (Dicroccelium). 1847: Brachylæmus. 1899: Haplometra (type).

cylindraceum Zed., of Pachinger, 1888.—1888: Dist. [1894: D. variegatum.]

cylindrica Goeze, 1782a, 174, a "Klasse."—1782: Planaria. 1841: Dist.

cylindricum Dies., 1836d, 249, t. h. Cataphractus murica; Villa Maria, Brazil.—1836: Amphist.

cylindricum ranæ Mayer, 1841, 18.—1841: Dist.

cymbiforme Rud., 1819a, 96, t. h. Testudo mydas; Arimini.—1819: Dist. 1895: D. (Brachylaimus). 1899: Phyllodist. 1899: Spathidium. 1901: Plesiochorus (type).

cymbium Dies., 1850a, 320, t. h. Himantopus wilsonii; Caiçaræ, Brazil.—1850: Monost. [1819: M. flavum.] 1902: Hæmatotrephus.

cymbuliæ Græffe, 1860a, 47, t. h. Cymbulia peronii Les; Nizza.—1860: Cerc.

cymbuliæ delle Chiaje (1841a, 109).—Dist.

cyprinacea Schrank, 1790, 122, t. h. Cyprinus barbus.—1790: Festucaria.

cyprinaceum Zed., 1800a, 164, in intest. Cyprinorum.—1800: Dist. [1850: D. globiporum.]

cyprinæ Leach in Johnston, 1865, 35, t. h. Cyprina islandica; Plymouth, Eng.—1865: Monost. [1865: Malacobdella grossa.]

cyprini carassii Viborg, 1795, 242.—1795: Fasc.

cyprini idi (peritonei) Dies., 1858e, 367, t. h. Leuciscus idus; Rennes.—1858: Dist. [1858: ? Tetracotyle echinata.]

cyprini idi Moul., 1856, 233.—1856: Tetracotyle. [1858: T. typica.]

cysticola phalangii opilionis Dies., 1855, 64, see cystidicola 1846.—1855: Dist.

cysticum Crep., 1846a, 159, t. h. Planorbis sp.—1846: Dist.

cystidicola Crep., 1846, 156, t. h. Phalangium opilio.—1846: Dist.

cystophona Will.-Suhm, 1870, 5, for cystophora (Cerc.).

cystophora Wagener, 1866, 145, t. h. Planorbis marginatus.—1866: Cerc.

dactyliferum Braun, 1892a, 568, for dactylipherum (Dist.).

dactylipherum Poir., 1885, 10, t. h. Argonauta.—1885: Dist.

deflectens Rud., 1819a, 677, t. h. Sylvia sp., Motacilla sp.; Brazil.—1819: Dist. 1901: Dicroccelium.

delectans Braun, 1901g, 945, t. h. Myiothera ruficeps; Brazil.—1901: Dicroccelium.

delicatulum Dies., 1850a, 325, includes Dist. testudinis Rud., 1819a, 121, t. h. Emys europæa, Halichelys atra; Mus. Vien.—1850: Monost.

delicatulum Rud., 1809a, 373, t. h. Anas sponsa.—1809: Dist. 1828: Fasc.

deliciosum Olss., 1893, 10, t. h. Larus argentatus.—1893: Dist. 1900: Gymnophallus (type).

delitescens Looss, 1899b, 666, t. h. Chelonia mydas; apparently Egypt.—1899: Crico-cephalus (type). [See also albus.]

delphini Dies., 1850a, 330, t. h. Delphinus dalei; near Havre.—1850: Monost. [1860: M. blainvillei.] 1892: Monostomulum.

delphini Poir., 1886, 34, t. h. Delphinus delphis.—1886: Dist. 1892: Cladocœlium-1899: Brachycladium.

dendricum Ben., 1870, 36, for dendriticum (Dist.).

dendriticum Rud., 1819a, 93, t. h. Xiphias gladius.—1819: Dist. 1896: D. (Dicroccelium). 1899: Dicroccelium.

dendyi Haswell, 1893e, 96, t. h. Astacopsis bicarinatus; Australia.—1893: Temnocephala. dentatum Lint., 1900: 269, t. h. Paralichthys dentatus.—1900: Dist.

denticulata Rud., 1802, 91, t. h. Sterna hirundo; Greifswald, July.—1802: Fasc. 1809: Dist. (Echinost.). 1860: Echinost.

denticulata Rud., 1805, an arachnoid.—1808: Polyst. 1809: Polyst. (Pentast.), [now in Linguatula].

denticulatum Rud., 1819a, 90, t. h. Alcedo ispida; Mus. Vien., Europe.—1819: Amphist. 1845: Holost. 1850: Hemist.

denticulatum Olss., 1876, 10, t. h. Gadus virens.—1876: Octobothrium. 1895: Dactylocotyle.

depressum Polonio, 1859, teste Par., 1894, 149, t. h. Triton cristatus; Padua.—1859: Dist.

depressum Stoss., 1883, 118, t. h. Dentex vulgaris; Triest.—1883: Dist. 1886: D. (Brachylaimus).

detruncatum Braun, 1899g, 490, t. h. Mycteria americana, Ciconia americana; Brazil.—1899: Clinost. 1900: Dist.

diadema Mont., 1902, 137, t. h. Trygon violacea.—1902: Epibdella. 1903: Phylline. 1902: E. (Phylline).

diaphanum Odhn., 1902, 154, t. h. Polypterus bichir; White Nile.—1902: Callodist. (type).

diaphanum Staff., 1904, 494, t. h. Ambloplites rupestris; Canada.—1904: Protenteron (type).

diaphanum Cerf., 1894k, 936, t. h. Raja batis.—1894: Merizocotyle (type).

dichotoma Mueller, in LaValette, 1855, 38, free form.—1855: Cerc. 1858: C. (Schizocerca).

dicorynum Dies., 1850a, (359), 680, t. h. Lampris guttatus; new name for D. affine Dies.—1850: Dist. 1886: D. (Köllikeria?).

dicranocælum Fischder., 1901a, 369, t. h. Bos taurus indicus; Coll. Berl. Vet. School.— 1901: Paramphist.

dictyotus Mont., 1893, 156, reticulatum Looss, 1885, renamed.—1893: Dist. 1893: Mesogonimus (type).

didelphidis Par., 1896, 3, t. h. Didelphys azaræ; Paraguay.—1896: Dist. (Brachylaimus). 1904: Plagiorchis.

didelphydis Stoss., 1904, 2, for didelphidis Par.—1904: Dist. [D. (Brachylaimus).]

diesingi Braun, 1901, 561, for diesingii 1860.—1901: Dist. [1902: Microlistrum cochlear.]

diesingi Fil., 1837a, 334, t. h. Planorbis nitidus; Ticino.—1837: Diplodiscus. 1856: Cerc. [1850: Diplocotyle mutabile (type).]

diesingii Fil., 1854a, 6, for diesingi (Cerc.). 1854: Diplodiscus.

diesingir Cobbold, 1860a, 14 (D. cochlear Dies., renamed), t. h. Sterna cantiaca, S. minuta; Brazil.—1860; Dist. [Microlistrum cochlear.]

differens Sons., 1891, 261, t. h. Cantharus lineatus.—1891: Trochopus.

difformis Wagener, 1857, 63, t. h. Cyprinus erythrophthalmus.—1857: Dactylogyrus.

diffusocalciferum Gastaldi, 1854, 5, t. h. Rana esculenta.—1854: Dist.

digitata Mont., 1903a, 309, t. h. Palemonetes argentinus.—1902: Temnocephala.

digitatum Rathke, (1843), 242, t. h. Hippoglossus gigas; Norway.—1843: Octobothrium. [1850: Diclidophora palmata.] [1879: Octobothrium palmatum.]

digitatus Looss, 1899b. 641, t. h. Sphyræna vulgaris; Egypt.—1899: Hemiurus. 1901: Lecithochirium. 1907: Plerurus (type).

dilatatum Fischer de Waldheim, 1840a, 158, t. h. Gallus communis; Vilnæ.—1840: Dist. 1860: Echinost. [1892: E. echinatum.]

dilatatus Dady, 1905, 233, t. h. Colossoma brachypoma; Paraguay.—1905: Chiorchis. dilutatum Schneidemuehl. 1896, 303, misprint for dilatatum (Dist.).

dimidiatum Crep., 1829, 55, t. h. Acipenser sturio.—1859: Dist.

dimorphum Dies., 1850a, 353 [contains D. marginatum Rud., 1819a, 680, and Duj., 1845a, 446], t. h. Salmo carapus, etc., see page 184; Brazil.—1850: Dist. 1886: D. (Brachylaimus). 1899: Clinost.

dimorphum of Wagener, 1852, 555, see commutatum 1858, from chicken.—1852: Dist. 1890: Mesogonimus.

diodontis Oken, 1815, 182, t. h. Diodon sp.; Nolka to Cal.—1815: Phylline (type of P. Oken, teste Odhn.). [Trist. maculata.] [Capsala martinieri.]

diplachanthus Massa, 1903, 254, t. h. Trigla hirundo.—1903: Trochopus.

diplocotylea Pag., 1857, 25. t. h. Planorbis marginatus.—1857: Cerc. [1858: Diplocotyle mutabile.] [1885: Amphist. subclavatum.]

diplodiscoides Cohn, 1904, 240, t. h. Rana esculenta.—1902: Opisthodiscus (type).

diplorchis Odhn., 1905, 318, t. h. Lumprenus medius; Spitzbergen, off King's bay.— 1905: Monorcheides (type).

[dipsacum Lint., 1897a, 806, cestode (Octobothrium).]

discus Mueller, 1786, 138, pl. 20, fig. 3, in aqua palustri raro.—1786: Cerc. 1829: Cyclidium.

diserialis Ssinitzin (1896), in Rallus aquaticus.—1896: Notocotyle.

dispar Looss, 1902. 888, t. h. dogs and cats; Egypt.—1902: Heterophyes.

disticha Mueller, 1776, 224.—1776: Fasc. 1803: Dist.

distomatosa Linst.. 1889a, 118, based on Sons., 1884, 98, t. h. Cleopatra bulimoides; near Cairo, Egypt.—1889: Cerc.

distomi folii Looss, 1894a, 251.—1894: Cerc. [Dist. folium.]

distomi hepatici Looss, 1894a, 252.—1894: Cerc. [Fasc. hepatica.]

distomi homolostomi Linst., 1889a, 120, t. h. Limnæa stagnalis.—1889: Cerc. [Distholostomum.]

distomi militaris Braun, 1893a, 832.—1893. Cerc. [Dist. militare.]

distomi perlati Looss, 1894a, 32.—1894: Cerc. [Dist. perlatum.]

distomi retusi Linst., 1878a, 327, in Limnæa stagnalis.—1878: Cerc. [Dist. retusum.] divergens Rud., 1809a, 371, Fasc. blennii renamed, t. h. Blennius viviparus.—1809: Dist.

divergens Looss, 1902e, 640, t. h. Ranzania truncata; Triest.—1902: Orophocotyle. dolichocotyle Cohn, 1903, 37, t. h. Herpetodryas fuscus.—1903: Amphist. 1904: Ca-

tadiscus (type).

donavani Mont., 1888, 16, for donavini (Microcotyle).

donavini Ben. & Hesse, 1863; 1864, 114, t. h. Labrus donavini.—1863; 1864: Microcotyle (?type).

draconis Briot, 1904, 126, t. h. Trachinus draco; North Sea.—1904: Microcotyle.

dubia Gœze, 1782a, 177, see alata.—1782: Plan.

dubium Leidy, 1856, 45, t. h. Rusticola minor.—1856: Clinost. 1858: Dist.

dubium Cobbold, 1860, 45, t. h. Corvus corone.—1860: Holost. [1858: H. corones.]
dubium Cobbold, 1858b, 156 t. h. Gasterosteus spinachia.—1858: Monost. 1892: Monostomulum.

dubius Klein, 1905, 68, t. h. Coluber olivaceus.—1905: Halipegus.

dujardini MacCallum, 1904, 547, for dujardinii (Echinost.).

dujardinianus Dies., 1850a, 432, t. h. Cyprinus carpio, Leuciscus rutilus.—1850: Gyrodactylus. 1858. Dactylogyrus.

dujardinii Cobbold, 1860a, 37, Dist. histrix Duj., 1845a, 433, D. histrix Dies., 1850a, 393, renamed; t. h. Pleuronectes maximus, P. platessa—1860: Echinost.

dujardinii Cobbold, 1860a, 29, D. soleæ 1845, renamed, t. h. Solea vulgaris, see Pleuronectes solea.—1860: Dist.

dujardinii Dies., 1850a, 420, t. h. Cyprinus erythrophthalmus.—1850: Diporpa (type). [1879: Diplozoon paradoxum, type.]

dujonii Braun, 1893a, 917, see also dujonis (Monost.).

dujonis Leuck., (1874), 419, t. h. Halicore dujong.—1874: Monost.

duplicata Baer, 1826, 124, t. h. Anadonta ventricosa; Regiomontii.—1826: Dist. 1856: Cerc. [1850: Rhopalocerca tardigrada, type.]

duplicatum Rud., 1819a, 125, Polyst. thynni 1811, renamed, t. h. Scomber thynnus; Balearic Isles.—1819: Polyst. 1845: Polyst. (Hexacotyle). 1850: Plagiopeltis (type).

ecaudata Eichwald, 1829a, 248, t. h. Lymnæus stagnalis.—1829: Cerc.

echeneidis remoræ Rud. (1821-8), 163, t. h. Echeneis remora.—18—?: Dist.

echeneis Wagener, 1857, 99, t. h. Chrysophrys aurata.—1857: Dactylogyrus. 1889: Diplectanum.

echinata Dies., 1858e, 367, new name for Dist. cyprini idi (peritonei), see Duj., and T. acerinæ cernuæ, t. h. Leuciscus idus, Acerina cernua; Europe.—1858: Tetracotyle.

echinata Sieb., 1837, 187, t. h. Lymnæus stagnalis: Gedani.—1837: Cerc. 1855: Cerc. (Hormocerc.). 1858: Cerc. (Nephrocephala). [1858: Dist. echinatum.] 1861: Dist.

echinata Nord., 1840, 621.—1840: Fasc. [1840: F. trigonocephala.]

echinatoides Fil., 1854, 266, t. h. Paludina vivipara, P. achatina; Lake Varese, Lombardy.—1854: Cerc. 1855: Cerc. (Hormocerc.). 1858: C. (Nephrocephala). [1858: Dist. echiniferum.]

echinatoides Dies., 1858d, 263, for echinatoides anodontæ.—1858: Dist.

echinatoides anodontæ Pag., 1857, 32, t. h. Anadonta cygnea.—1857: Dist.

echinatum Linst., 1878, 223, t. h. Pandion haliaëtos.—1878: Monost.

echinatum Fil., 1837a, 338, t. h. Paludina impura; Italy.—1837: Heterost. (type). 1858: Cercariæum.

echinatum Zed., 1803a, 220, t. h. Anas domestica, A. querquedula; includes: Cucullanus conoideus Bloch, 1782; Planaria teres Gœze, 1782; Dist. anatis Zed., 1800; Festucaria' anatis Schrank; Fasc. anatis Gmelin.—1803: Dist. 1828: Echinost., type. 1809: D. (Echinost.).

cchinifera LaValette, 1855, 14, t. h. Paludina vivipara.—1855: Cerc. 1855: Dist. [1856: Cerc. echinatoides.] [1858: C. (Nephrocephala) echinatoides.] [1873: Dist.

militare.] 1892: Echinost.

echiniferum paludinæ Pag., 1857, 30, t. h. Paludina vivipara.—1857: Dist.

echinocephalum Rud., 1819a, 115, t. h. Falco milvus; includes D. milvi.—1819: Dist. (Echinost.). 1860: Echinost.

echinocerca Fil., 1855b, 17, t. h. Buccinum linnæi; Golf de Gênes.—1855: Cerc. 1858:

Histrionella. [1858: Dist. appendiculatum.]

echinostomum Dies., 1850a, 326, t. h. Cathartes aura and Sula fusca; Brazil; includes Dist. planicolle Rud., from Pelecanus sula in Brazil.—1850: Monost. [1902: Anoictost. planicolle.]

echiuri Greef, 1879a, 130, t. h. Echiurus pallasi.—1879: Dist.

efemera Ssinitzin, 1905, 158, for ephemera (Cerc.).

ehrenbergii Focke, 1836a, 191.—1836: Planaria.

elaphi Gmelin, 1790a, 3054, cervi renamed, t. h. Cervus elaphus.—1790: Fasc. Monost. [1850: Amphist. conicum.]

elegans Baer, 1826, 125, t. h. Stör=Acipenser sturio.—1826: Nitzschia (type). Trist.

elegans Ben., 1858a, 1861a, 60, t. h. Sciæna aquila; Belgium.—1858, 1861: Calceost. (type.)

elegans Dies., 1858e, 364, new name for Epibdella sciænæ, t. h. Sciæna aquila; near Ostend.—1858: Benedenia (type).

elegans Goto, 1894a, 188, t. h. Scombrops chilodipteroides; Japan.—1894: Microcotyle. elegans Looss, 1899b, 692, t. h. Phœnicopterus roseus; Gizeh.—1899: Echinost.

elegans Mont., 1890, 191, Raja clavata; Naples.—1890: Acanthocotyle.

elegans Mueller in LaValette, 1855, 13, free form.—1855: Cerc. 1858: Histrionella. elegans Olss., 1868, or 1869, 2, t. h. Chimæra monstrosa; Skagerrack.—1868: Macraspis (type). [1888] 1891: Aspidogaster.

elegans Rud., 1802, 65, t. h. Fringilla domestica; Greifswald.—1802: Fasc. 1809: Dist. 1845: D. (Brachylaimus). 1902: Plagiorchis.

elegans Nord., 1832, 106, t. h. Cyprinus brama.—1832: Gyrodactylus (type).

elegans Nord., 1840, 597, 600, includes crassicaudatum (Diklibothrium), t. h. Acipenser stellatus.—1840: Hexacotyle.

elephantis Dies., 1858, 354, based on Jackson, 1847, 317, t. h. Elephas indicus.—1858: Dist. 1892: Cladocælium.

elipticum Stoss., 1886, 64, for ellipticum Mol., 1858.—1886: D. (Echinost.).

ellicticum Mont., 1892, 715, for ellipticum (Monost.).

elliptica Dies., 1850a, 421, t. h. Labrax mucronatus.—1850: Plectanocotyle (type). 1858: Plectanophorus (type).

ellipticum Mol., 1858, 130, t. h. Acipenser nasus; Patavii.—1858: Dist. 1886: D. (Echinost.).

ellipticum Brand., 1888a, 59, t. h. Piaya cayana; Brazil.—1888: Hemist.

ellipticum Brand., 1888a, 67, t. h. Bubo magellanicus; Brazil.—1888: Holost.

ellipticum Rud., 1809a, 333, M. bombynæ 1800, renamed.—1809; Monost. (Monost.). [1853: Dist. variegatum.]

ellipticus Pratt, 1903, 25, t. h. Heterodon platvrhinus; North America.—1903: Renifer (type).

elongata Goto, 1894a, 210, t. h. Pagrus tumifrons; Mogi and Hakodate, Japan.— 1894: Diclidophora.

elongata Nitzsch, 1826, 150, t. h. Stör.—1826: Trist. 1840: Capsala. [1850: N. elegans.] 1865: Nitzschia.

elongatum Mehlis, 1831, 177, t. h. Larus argentatus, L. marinus, L. ridibundus.—1831:

elongatum J. Poir., 1883, 76, t. h. Palonia frontalis; Java.—1883: Gastrothylax. 1903: Amphist.

elongatus Pratt, 1903, 25, t. h. Heterodon platyrhinus; North America.—1903: Renifer. emarginata Olss., 1876, 11, t. h. Raja clavata.—1876: Onchocotyle. 1890: Octocotyle. emarginatum Rud., 1819a, 124.—1819: Pentast.

emarginatum Dies., 1839a, 237, t. h. Callithrix noctivaga; Matogrosso, Brazil.—1839: Amphist.

ematobia Sons., 1884, 20, for hæmatobia (Bilharzia).

émberizæ citrinellæ Dies., 1836d, 253.—1836: Amphist.

embryo Olfers, 1816, 110, t. h. Perca vulgaris, P. cernua.—1816: Dist.

emendatus Braun, 1901g, 895, new name for Dist. meropis of Par. [not of Rud.]—
Megacetes triangularis of Looss [not D. triangulare Dies.]; t. h. Merops apiaster.—1901: Eumegacetes (type).

[endemica Sons., 1884, 17, not as specific name.—1884: Dist. 1884: Bilharzia.]

endemicum Bælz, 1883, 235, t. h. Homo; Japan.—1883: Dist. hepatis. 1892: D. (Brachylaimus). [Opisthorchis sinensis.] 1907: Clonorchis.

endemicum hepatis St.-Remy, 1883, 528, cf. hepatis endemicum.—1883: Dist.

endobala Ssinitzin, 1906, 685, for endoloba (Opisthioglyphe).

endobola Ssinitzin, 1905, 121, for endoloba (Opisthioglyphe).

endoboluum Mont., 1891, 110, for endolobum (Dist.).

endolobium Gurlt, 1845a, 288, for endolobum (Dist.).

endolobum Duj., 1845a, 397, t. h. grenouilles vertes et rousses, et la Salamandre; Rennes.—1845: Dist. 1899: Opisthioglyphe (type).

engæi Haswell, 1893e, 97, t. h. Engæus fossor; Gippsland.—1893: Temnocephala.

engei Mont., 1899, 83, for engæi (Temnocephala).

enterarchos de Fil.—Dist. [1896: D. crassicolle.]

entzi Ratz, 1900, 534, t. h. Ardea purpurea.—1900: Opisthorchis.

epatica Brera, 1809a, 92, for hepatica (Fasc.).

epatica Rosa, 1794, 5, in Ardea purpurea.—1794: Fasc. [1850: Dist. heterostomum.] [1900: ? Clinost. heterostomum.]

epatico Galli-Valerio, 1893a, 178 (Dist.), see hepatica.

ephemera Nitzsch, 1807, 33, t. h. Planorbis corneus; Halle.—1807: Cerć. 1828: Histrionella H. & E., type. [1858: Glenocerc. flava.] [1858: Dist. trigonocephalum.]
epiclitum Fischder., 1904, 458, t. h. Bos taurus indicus, Buffelus indicus; Cochin China.—1904: Paramphist.

equi Gmelin, 1790a, 3054, t. h. Equus caballus.—1790: Fasc. hepatica.

equi Burke, 1882a, 322, t. h. Equus caballus.—1882: Hæmatobium.

ercolani Mont., 1893, 40, for ercolanii.—1893: Dist.

ercolanii Mont., 1893, 40.—1893: Dist. 1895: D. (Dicrocœlium). 1901: Telorchis. ericetorum Linst., 1898, 761, t. h. Xerophilus ericetorum; near Gættingen, Germany.—1898: Cerc.

erinacei E. Bl., 1847a, 300, t. h. Erinaceus europæus; Paris.—1847: Brachylæmus. [1850: Dist. linguæforme.] 1889: Dist. [1898: Mesogonimus.]

erinaceum Poir., 1886, 37, t. h. Delphinus delphis.—1886: Dist. 1892: D. (Dicroccelium). [1899: ?Astia.] 1904: Astiotrema.

eriocis Mueller, 1784, 92; [1788], 42, t. h. Salmo eriox.—[1788]: Fasc. 1803: Dist. [1850: D. hyalinum.]

erraticum Rud., 1819a, 120, t. h. Fringilla linaria, Motacilla alba, Parus cœruleus, P. major, P. palustris, P. pendulinus.—1819: Dist. 1899: Plagiorchis.

erraticum Rud., 1808a, 458; 1809a, 344, t. h. Larus septentrionalis; Greifswald, Europe.—1808: Amphist. 1845: Holost.

erythrini Ben. & Hesse, 1863, 115; 1864, 115, t. h. Pagellus erythrinus.—1863: Microcotyle.

erythrinis Braun, 1890a, 418, for erythrini (Microcotyle).

erythrops Dies., 1855a, 400, based on Cerc. paludinæ impuræ Baer, 1827b, 655.—1855: Cercariæum. 1858: Histrionellina.

esmarkii Scott, 1901, 147, t. h. Gadus esmarkii; Shetland.—1901: Octobothrium.

esocis lucii Rud., 1809a, 438, based on Rud., 1803, 29.—1809: Dist.

euculus McCrady, 1874, misprint for cuculus (Bucephalus).

euryporum Looss, 1896, 144, t. h. Milvus parasiticus; Cairo, Egypt.—1896: Echinost.
eurystomum Linst., 1877, 183, t. h. Anas clangula.—1877: Dist. 1892: D. (Dicrocelium).

eustemma Brand., 1888a, 65, Eustemma caryophyllum renamed, t. h. Accipiter pileatus; Brazil.—1888: Holost.

exacantha Mont., 1891, 104, for hexacantha (Placunella).

exachanthus Massa. 1903. 255. for hexacanthus Trochopus ...

exasperatum Rud., 1819a, 117, t. h. Sorex eremita: Mus. Vienna.—1819; Dist. 1819; D. (Echinost.). 1845; D. (Brachylaimus).

ercavata Rud., 1803a, 28. t. h. "Storch." Ardea ciconia: Greifswald.—1803: Fasc. 1809: Dist. 1819: Holost. 1819: Amphist. 1850: Hemist.

excavatum Nord., in Dies., 1850a, 428, renamed nordmanni 1850, in Brama mediterranea.—1850: Trist. [1850: Encotyllabe nordmanni.]

excisjorme Cohn. 1902k. 54. t. h. Scomber scomber.—1902: Lecithocladium.

excission Mont., 1891, 520, for excisum Apoblema .

excisum Rud., 1819a. 112. t. h. Scomber scomber at Arimini: S. colias at Naples.—1819: Dist. 1828: Fasc. 1845: Dist. Crossodera. 1886: D. Apoblema. 1889: Apoblema. 1899: Hemiurus. 1901: Lecithocladium type. 1902: D. Lecithocladium .

excisum Linst., 1906, 12. t. h. Egolius otus, Strix flammea.—1906: Holost.

exfoliata Moul., 1856a, 87, new name for Dist, of Leucochloridium paradoxum.—1856;

exigua Looss, 1896, 230, t. h. Cleopatra bulimoides: Egypt.—1896: Cerc.

exiguum Mueh.. 1898, 17. t. h. Circus rufus.—1898; Dist. 1898; Opisthorchis. 1899; Holometra type.

exiguum Mehlis in Crep., 1846, 145, t. h. Cygnus musicus.—1846; Holost.

exile Stoss., 1902, 17 t. h. Totanus ochropus.—1902: Cyclocoelum.

exilis Looss, 1899b, 628, t. h. Bagrus bayad; Cairo, Egypt.-1899; Leptalea type . 1900: Emoleptalea type .

exocuti Par. & Perugia, 1893. 1, includes Monost, filum G. Wagener [not Duj.], t. h. Exocœtus volitans: Genova.—1893: Didymozoon.

expansum Crep., 1842, 327, t. h. Aquila haliaetos.—1842; Monost, 1892; Dist. Tocotrema. 1903: Scaphanocephalus type .

explanatum Crep., 1847. 34. t. h. Bos taurus indicus; Berlin.-1847; Amphist. Paramphist.

caspinosum Hausmann, 1896, 391, t. h. Barbus fluviatilis.—1896; Dist. [D. perlatum.] 1899: Asymphylodora.

faba Bremser in Schmalz, 1831, 11, t. h. Parus major, Silvia sibilatrix, Motacilla bearula.—1831; Monost. 1860; Wedlia. 1904; Monostromum, misprint.

fabaceum Dies., 1838, 189, in Manatus australis: 1839a, 236, in Manatus exunguis; at Borba and Forte do Rio Branco, Brazil.—1839: Amphist. 1901: Chiorchis type). jabenii Mol., 1859, 289, t. h. Cantharus vulgaris; Batavii.—1859; Dist. 1886; D.

Brachylaimus .

jactum Mont., 1893. 32, misprint for fractum | Dist. |.

jalcatus Wedl. 1857. 258, t. h. Cyprinus sp.—1857: Gyrodactylus. 1858: Dactylogyrus.

falcolaca Leidy MS. in Stiles & Hass., 1894d, 250.—1894: Dist. [probably trapezium]. jalconis chrysaëti Rud., 1809a, 429. D. felleum falconis chrysaëti: Viborg.—1809: Dist. jalconis milei Rud., 1809a, 429, for Fasc, milvi Gmelin,—1809; Dist.

jalconis galumba Baird. 1853a. 47.—1853: Amphist. [1853: Holost. macrocephalum.]

falconis palumbarii Rud.. 1819a. 88. syn. of macrocephalum Amphist. ..

falconi: palumbi Viborg. 1795. 243.—1795: Strigea. 1809: Amphist.

jalconis peregrini Rud.. 1819a. 92. t. h. Falco peregrinus: Berlin, Germany.—1819: Amphist.

jalconis rufi Rud.. 1819a. 119, t. h. Falco rufus; Cat. Ent. Vien.—1819: Dist. [1850; D. lineola.

falconum Dies., 1858, 322, t. h. Falco nisus, F. rufus: Ireland.—1858: Holost.

fallar Rud., 1819a, 117. t. h. Uranoscopus scaber: Naples.—1819: Dist. Echinost... 1860: Echinost. 1899: Anoiktost. 1901: Anisogaster (type). 1902: Anisocladium type.

fallar Dies., 1850a, 297, Cerc. VII Baer, C. echinata Sieb., Dist. pacifica Steenstrup, renamed; t. h. Paludina vivipara; Regiomontii Baer,—1850; Cerc. 1855; C. Eucerc., [1858; Dist. militare.] 1858; C. Gymnocephala.

fallar Wagener, 1857, 55. t. h. Cyprinus erythrophthalmus, C. rutilus.—1857: Dactylogyrus.

farionis Mueller, 1784, 91, t. h. Salmo fario; [1788], 42.—[1788]: Fasc. [1814: Dist. laureatum.] 1891: Dist.

fasciata Haswell, 1887a, 284, t. h. Astacopsis serratus; N. S. Wales.—1887: Temnocephala.

fasciatum Rud., 1819a, 97, t. h. Labrus tinca, L. merops, Perca marina; Naples.—
1819: Dist. 1886: D. (Dicrocœlium). [1899: Creadiinæ.] 1901: Allocreadium.
1902: Helicometra. 1902: Loborchis.

fasciatus Stoss., 1902, 25, t. h. Numenius arquatus; locality?.—1902: Hæmatotrephus. fascicularis Villot, 1875, 480, t. h. Nassa reticulata; Roscoff.—1875: Cerc.

fasciolaris Mueller, 1788.—1788: Hirudo. [1790: Fasc. anatis.] [1850: Dist. ovatum.] fastosum Braun, 1901g, 896, t. h. Caprimulgus sp., Squatarola helvetica; Brazil.—1901: Stomylotrema.

felineum Rivolta, 1884, 20, t. h. Felis catus dom.; Italy.—1884: Dist. 1893: D. (Dicrocœlium). 1895: Opisthorchis (type). 1896: Dicrocœlium. [1896: Prosthometra, type.] 1898: Campula. ——: Dist. (Opisthorchis).

felineum of Ward, 1895, 152, see pseudofelineum.—1895: Dist.

felinum Ratz, 1896, 67, misprint for felineum (Dist.).

felleum falconis chrysaëti Viborg, 1795, 243.—1795: Dist. [1850: D. crassiusculum.]

fellis Olss., 1868, 44, t. h. Anarhichas lupus.—1868: Dist. 1886: D. (Brachylaimus).

ferocis Mont., 1888a, 14, apparently for ferox (Dist.).

ferox Rud., 1795a, 15, t. h. Ardea ciconia; Greifswald.—1795: Fasc. 1803: Dist.
 1809: D. (Echinost.). 1828: Echinost.

ferruginosum Linst., 1877, 184, t. h. Barbus fluviatilis.—1877; Dist. [1894; D. perlatum.]

ferrum-equinum Dies., 1836d, 238, t. h. Cataphractus murica, C. corome; Cuyaba and Matogrosso, Brazil.—1836: Amphist.

festæ Borelli, 1898, 6 pp.—1898: Plan.

filaria Bosc, 1802a, 261.—1802: Plan.

filarina Ben., 1858 or 1861, 108, t. h. Sciæna aquila; Ostend.—1858: Nematobothrium (type). 1859: Monost.

filicolle Rud., 1819a, 85, t. h. Brama raji; Naples.—1819: Monost. (Monost.). 1858: Dist. 1860: Köllikeria (type). 1886: D. (Köllikeria).

filicolle Mont., 1893, 150.—1893: Didymozoon.

filiferum Sars, 1885, 222, t. h. Nematoscelis megalops, Thysanoëssa gregaria; South Atlantic Ocean.—1885: Dist.

filiforme Rud., 1819a, 112, t. h. Cepola tænia, at Arimini; C. rubescens.—1819: Dist.

filigerum Rud., teste Risso, 1826, 262, de la castagnolle.—1826: Monost.

filum Duj., 1845a, 418, t. h. moineaux; Rennes.—1845: Dist. (Brachylaimus).

filum Duj., 1845a, 362, t. h. Scomber scombrus; Rennes.—1845: Monost. fimbriata Gœze, 1782a, 180.—1782: Fasc. [1810: Caryophyllæus mutabilis.]

fimbriatum Busch, 1851, 99, t. h. Sagitta.—1851: Dist.

fimbriatum Mol., 1859, 819, t. h. Anguilla vulgaris; Batavii.—1859: Gasterost.

fimbriatum Sieb., 1848, v. 1, 129, t. h. Perca fluviatilis, Lucioperca; Europe.—1848: Gasterost. (type).

fintæ Ben. & Hesse, 1863, 1864, 101, t. h. Alosa finta.—1863: Ophicotyle (type). 1879: Octobothrium.

fintæ Mont., 1888a, 13.—1888: Glossocotyle.

fissa Bory de St. Vincent, 1824a, 456.—1824: Histrionella. [1850: Malleolus furcatus.]

fissicanda Moul., 1856a, 109, for fissicauda (Cerc.).

fissicauda Dies., 1858d, 243, cristata Valette renamed.—1858: Lophocerc. (type).

fissicauda Dies., 1858d, 269, ocellata (Cerc.) renamed.—Histrionellina.

fissicauda La Valette, 1855, 21, t. h. Limnæus stagnalis.—1855: Cerc. 1858: C. (Schizocerca).

fissicaudata Mont., 1888, 196, for fissicauda (Cerc.).

flaccida Mueller, 1774, 57, free form.—1774: Fasc. 1776: Plan.

flagellatum Moniez, 1891, 27, t. h. Gymnotus electricus.—1891: Dist.

flava Stoss., 1903, 373, t. h. Centropristis hepatus; Triest.—1903: Helicometra,

flava La Valette, 1855, 24, C. ephemera Nitzsch renamed.—1855: Cerc. 1858: Glenocerc. 1858: Monost. [See flavum.]

flavescens Ben., 1870, 47, t. h. Gobius jozo, G. minutus.—1870: Dist. 1886: D. (Brachylaimus).

flavescens Pag., 1857, 34, t. h. Bulimus radiatus; Heidelberg.—1857: Dist. 1858: Cercariæum.

flavocinctum Linst., 1879, 183, t. h. Anguis fragilis.—1879: Dist. [1897: D. crassicolle.] flavopunctata Par., 1894, 703, for fulvopunctata (Cerc.).

flavum Mehlis, 1831, 172, t. h. Anas mollissima, A. fusca, A. marila, A. fuligula.— 1831: Monost. 1858: Glenocerc. 1902: Typhlocœlum (type). [See flava.]

flexum Lint., 1892, 98, t. h. Oedemia americana.—1892: Dist. 1899: Echinost.

flexuosum Rud., 1808a, 346; 1809a, 50, t. h. Talpa europæa.—1809: Dist. 1845: D. (Dicroccelium). 1899: Omphalometra (type).

fodicans Braun, 1901g, 896, t. h. Sterna nigra; Vien. Mus. no. 631.—1901: Cathæmasia. fæcundum Lint., 1900, 269, t. h. Lopholatilus chamæleonticeps.—1900: Dist.

fætorii Linst., 1876, 1, t. h. Fætorius putorius.—1876: Tetracotyle.

foliaceum Mol., 1859, 288, t. h. Gobius paganellus; Batavii.—1859: Dist.

foliaceum Rud., 1819a, 83, t. h. Accipenser sturio; Arimini.—1819: Monost. (Monost.). 1859: Amphilina.

foliaceum Goto, 1894a, 248, host unknown.—1894: Trist.

foliatum Lint., 1898, 532, t. h. Mola mola.—1898: Dist. 1902: Orophocotyle. 1904: Accaccelium.

folii Looss, 1894a, 256, Dist. duplicatum.—1894: Cerc.

foliiforme Crep., 1846, 149, t. h. Squalus griseus.—1846: Dist.

foliiforme Braun, 1899g, 490, t. h. Ardea purpurea; Italy.—1899: Clinost.

folium Olfers, 1816, 45, t. h. Esox lucius.—1816: Dist. [1894: Rhopalocerca tardigrada, type.] 1899: Spathidium (type). 1899: Phyllodist. (type). [1906: Gorgoderina.]

forceps Leuck. (1857), 26.—1857: Dactylogyrus.

forcipata Mueller, 1786, 134, in aqua palustri rarissime.—1786: Cerc. 1827: Dicranophorus.

formosum Sons., 1890, 134, t. h. Grus cinerea.—1890: Dist. 1892: Polyorchis.

formosum Staff., 1904, 486, t. h. Hippoglossus hippoglossus; Canada.—1904: Stegano-derma (type).

formosum Pratt, 1903, 34, t. h. a frog.—1903: Ostiolum (type).

fractum Rud., 1819a, 107, t. h. Sparus salpa; Naples.—1819: Dist. 1886: D. (Podocotyle). 1898: Podocotyle.

fragile Lint., 1900, 269, t. h. Mola mola.—1900: Dist. 1904: Stenocollum (type).

fragile Olss., 1869, 4, t. h. Raja batis; Norway.—1869: Microbothrium. 1890: Pseudocotyle. [1897: Micropharynx parasitica, type.]

fraterculus Odhn., 1905, 343, t. h. Phoca barbata, Odobenus rosmarus; Spitzbergen.—1905: Orthosplanchnus.

fraterna Odhn., 1902, 19, t. h. Harelda glacialis.—1902: Cyathocotyle.

fraternum Looss, 1894, 42, t. h. Pelecanus onocrotalus; Egypt.—1894: Dist. 1899: Cenogonimus. 1899: Cotylogonimus. 1902: Heterophyes.

frælichii Kowal., 1894, 3.—1894: Dist. 1894: Echinost. [1898: E. conoideum.]

fulgo punctata Braun, 1893a, 831 for fulvopunctata (Cerc.).

fuligulæ ferinæ Dies., 1858e, 355, based on Bellingham, 1844, v. 13, 430, t. h. Anas ferina; Ireland.—1855: Dist. [1892: Echinost. echinatum.]

fulva Bosc., 1802a, 257.—1802: Plan.

fulvopunctata Erc., 1881 or 1882, see Par., 1894, 161, t. h. Bythinia tentaculata; Bologna.—Cerc.

fulvum Rud., 1819a, 98, t. h. Gadus molva at Naples; G. mediterraneus.—1819: Dist. fulvum Duj., 1843, 340, t. h. Sorex.—1843: Brachylaima.

furcata Nitzsch, 1817, 49.—1817: Cerc. 1838: Malleolus (type).

furcatum Bremser, in Rud., 1819a, 107, t. h. Mullus surmuletus, M. rubescens at Arimini; Gadus molva at Naples.—1819: Dist. 1828: Fasc. 1845: D.(Podocotyle). 1898: Podocotyle. [furcatæ Eichwald, 1829a, 247.—Cerc.]

furcigerum Olss., 1868, 26, t. h. Pleuronectes limanda, P. limandoides.—1868: Dist. 1886: D.(Dicrocœlium). 1904: Leioderma (type). 1905: Steringophorus (type).

fusca Bosc., 1802, 271, in dorade.—1802: Fasc. 1885: Dist.

[fusca Sluiter, 1900, 7, (Dist.), a tunicate.]

fusca Pallas, 1774, 21, in aq. stag.; Europe.—1774: Fasc.

fusca Fabricius, 1798, 58, syn. Fasc. angulata.—1798: Plan.

[fusca, (Diplost.) mammal.]

fuscatum Rud., 1819a, 101, t. h. Tetrao coturnix; Ancona.—1819: Dist. ——: D. (Dieroccelium). 1902: Harmost.

fuscescens Rud., 1819a, 113, t. h. Sparus dentex; Arimini.—1819: Dist. 1886: D. (Dicrocœlium).

fuscescens Fabricius, 1798, 58.—1798: Plan.

fusiforme Luehe, 1901, 476, t. h. Conger conger; Coll. Berlin.—1901: Lecithochirium. fusiforme Zed., 1800a, 163, Fasc. upupæ Schrank, 1790, renamed.—1800: Dist. [1850:

D. involutum.]

fusiformis Goto, 1894a, 192, t. h. Centronotus rubulosus; Japan.—1894: Microcotyle. gadi Dies., 1855, 64, based on Bellingham, 1844, 428, t. h. Gadus æglefinus; Ireland.—1855: Dist.

gadi æglefini Dies., 1858e, 341, based on Bellingham, 1844, 428; renamed anonymum 1858e, 341.—1858: Dist.

gadorum Ben., 1870, 60, t. h. Merlangus carbonarius.—1870: Echinost.

gadorum Rathke, 1799, 68.—1799: Hydatula. 1851: Nematoideum. 1878: Gasterost. galactosomum Leidy, 1888i, 166, t. h. Labrax lineatus; U. S. A.—1888: Dist.

galeatum Rud., 1819a, 86, t. h. Centronotus glaucus; Naples. — 1819: Monost. (Monost.). 1898: Gasterost.

gammari Rentsch, 1860, 35, t. h. Gammarus ornatus.—1860: Dist.

gammari Linst., 1877, 186, t. h. Gammarus pulex.—1877: Dist.

gammari ornati Rentsch, 1860, pl. 12, fig. 2, see gammari (Dist.).

garzettæ MacCallum, 1904, 541, t. h. Garzetta nigripes Temm.—1904: Echinost.

gastrocolum Leidy, 1891a, 414, t. h. Trichiurus lepturus; U. S. A.—1891: Dist. gastroporus Luehe, 1901p, 166, t. h. Rana cyanophlyctis; India.—1901: Pleurogenes. gelatinosum Rud., 1819a, 102, t. h. Testudo mydas: Arimini.—1819: Dist. 1895: D.

(Dicrocœlium). 1901: Rhytidodes (type).

gelatinosum of Poir., see poirieri.

gemellatum Mont., 1892, 716, for gemellum (Monost.).

gemellum Steenstrup, 1860, 113, t. h. Sphyræna baracuda.—1860: Monost.

geminum Bremser in Schmalz, 1831, 13, for M. faba.—1831: Monost. [1853: M. faba.]
geminum Looss, 1896, 50, t. h. Milvus parasiticus: Cairo.—1896: Dist. 1898: Opisthorchis.

genata Looss, 1907, 488, t. h. Pelecanus onocrotalus; Egypt.—1907: Pygidiopsis (type).
geniculatum Dies., 1850a, 373, D. physophoræ renamed, t. h. Physophora tetrasticha;
Naples.—1850: Dist.

genu Rud., 1819a, 107, t. h. Labrus luscus; Naples.—1819: Dist. 1901: Allocreadium. gesserensis Bosc, 1802a, 262.—1802: Plan.

qiardi Stoss., 1898, 50, for giardii (Dist.).

giardii Stoss., 1889, 25, t. h. Naucrates ductor; Triest.—1889: Dist.

gibba Mueller, 1773, 120, in infusione jungermanniæ tamarisci.—1773: Cerc. 1827: Macrocercus.

gibba Fil., 1854b, 266, t. h. Lymnæus pereger.—1854: Cerc. 1855: C. (Xiphidiocerc.). 1858: C. (Acanthocephala).

gibbosa Rud., 1802b, 81, t. h. Esox belone.—1802: Fasc. 1809: Dist. 1845: D. (Podocotyle). 1901: Lecithaster.

gibbum Mehlis in Crep., 1846, 137, t. h. Fulica atra.—1846: Monost.

giganteum Dies., 1858e, 331, gigantica Cobbold, 1855, renamed.—1858: Dist. 1892: Cladocœlium.

giganteum Dies., 1836d, 238, pl. 22, figs. 5-6, t. h. Dicotyles albirostris at Nas Frechas and Caiçara, and D. torquatus at Matogrosso; Brazil.—1836: Amphist. 1901: Cladorchis (Stichorchis, type).

gigantica Cobbold, 1855a, 262, t. h. Giraffa camelopardalis.—1855: Fasc.

gigantocotyle Brand., in Otto, 1896, 103, t. h. Hippopotamus.—1896: Amphist.

gigas Nardo, 1827, 68, t. h. Proctostegus proctostegus.—1827: Dist.

glabrum Crep., 1846, 148, t. h. Salamandra maculosa.—1846: Dist.

glandulosum Looss, 1896, 64, t. h. Taphosus nudiventris; Ghizeh.—1896: Dist. 1899: Lecithodendrium.

glauca Mueller, 1774, 60.—1774: Fasc. 1787: Plan.

glauci Bergh, 1884, 18, t. h. Glaucus atlanticus, etc.—1884: Dist.

globicaudatum Crep., 1849a, 64, for globocaudatum (Dist.).

globifera Lamarck, 1816, 182, for globipora.—1816: Fasc. 1816: Dist.

globiparum Ehrenberg, 1837b, Feb., 199, for globiporum (Dist.).

globipora Rud., 1802b, 72, t. h. Cyprinus erythrophthalmus.—1802: Fasc. 1809: Dist. 1845: D. (Brachylaimus). 1886: D. (Dicrocœlium). 1898: Sphærost. (type).

globipora Erc., 1881 or 1882, see Par., 1894, 164, t. h. Bythinia tentaculata; Bologna.—Cerc.

globiporum tincæ Rud., see Dies., 1850a, 395.—1850: Dist. [1850: D. perlatum.]
globocaudatum Crep., 1825, 49. t. h. Corvus cornix.—1825: Dist. 1845: D. (Brachylaimus).

globosum Ben., 1858a, 1861a, 193, quotes Siebold.—1858: Dist. [See also D. orbiculare.]

globulus Rud., 1814a, 104, t. h. Anas fuligula; Greifswald.—1814: Dist. [1902: Psilost.]

glottoides Klein, 1905, 72, t. h. Rana hexadactyla.—1905: Ganeo (type).

gobii Rentsch, 1860, 43, t. h. Gobius minutus.—1860: Dist.

gobii Stoss., 1883, 116, t. h. Gobius jozo, in int.; Triest.—1883: Dist. 1886: D. (Dicroccelium). 1902: Loborchis. 1904: Helicometra.

gobii Stoss., 1898, 58, encysted on intest. of Gobius jozo; Triest, Austria.—1898: Agamodist.

gobii minuti Rentsch, 1860, for gobii Rentsch (Dist.).

goliath Ben., 1858b, 95, t. h. Balæna.—1858: Dist. [1902: Lecithodesmus (?type).] 1905: Lecithodesmus (type).

goliath of Lænnberg, 1891, 71.—Dist. [1902: Lecithodesmus (type).]

gonocephala Ackermann, 1905, 137.—1905: Plan.

gorgoderæ cygnoides Kowal., 1904, 24, in Cyclas cornea; Dublany.—1904: Cerc.

loossi (Cerc.) Ssintizin, 1905, see under loossi. pagenstecheri (Cerc.) Ssinitzin, 1905, see under pagenstecheri. varsoviensis (Cerc.) Ssinitzin, 1905, see under varsoviensis. vitelliloba (Cerc.) Ssinitzin, 1905, see under vitelliloba.

gorgon Lint., 1905, 327, t. h. Seriola lalandi; Beaufort, N. C.—1905: Gasterost.
gracile Fischder., 1901a, 368, t. h. Bos kerabau from Ceylon; Portax tragocamèlus.—
1901: Paramphist.

gracile Leidy, 1856, 45, t. h. Pomotis vulgaris, Esox.—1856: Clinost. (type). 1858: Dist.

gracile Rud., 1809a, 326, based on Acharius, 1790, 55, t. h. Salmo eperlanus.—1809: Monost. (Hypost.).

gracile Rud., 1819a, 89, t. h. Mergus merganser, M. albellus; Mus. Vienn.—1819: Amphist. 1845: Holost.

gracilescens Rud., 1819a. 111. t. h. Lophius piscatorius; Triest.—1819: Dist. 1852: Gasterost. 1858: Rhipidocotyle (?type). [Bucephalus haimeanus.]

gracilis Fil., 1837, 336, t. h. Planorbis nitidus; Italy.—1837: Redia (type). [1855: Diplocotyle mutabilis.]

gracilis Kath., 1894a, 129, t. h. see p. 278.— 1894: Gyrodactylus.

gracilis La Valette, 1855, 20, t. h. Planorbis corneus.—1855: Cerc. 1858: C. (Schizocerca).

gracilis Wedl, 1861, 480, t. h. Hydrocyon dentex.—1861: Dactylogyrus.

gracilis Looss, 1901e, 660, t. h. Uranoscopus scaber; Triest.—1901: Anisogaster.

grande Dies., 1839a, 237, t. h. Chelys, Phrynops, Peltocephalus, Podocnemis, Rhinemys; Brazil.—1839: Amphist.

grande Rud., 1819a, 676-677, t. h. Platalea ajaja; Brazil.—1819: Dist. 1902: Mesaulus (type).

grande Dies., 1850a, 307, t. h. Ardea leuce, A. agami; Brazil.—1850: Diplost. 1890: Hemist. [1890: H. macropterum.]

grandiporum Rud., 1819a, 110, t. h. Muræna helena; Naples.—1819: Dist. 1886: D. (Apoblema). 1889: Apoblema. 1899: Hemiurus. 1901: Lecithochirium.

granulosum Looss, 1907, 483, t. h. Vesperugo kuhli; Cairo, Egypt.—1907: Lecithodendrium.

granulum Rud., 1809a, 394, includes Fasc. scorpii Mueller; t. h. Cottus scorpius.—
1809: Dist.

grassum Biermer, 1863a, 395, for crassum (Dist.).

gregarius Looss, 1896b, 5, t. h. Egyptian buffalo; Egypt.—1896: Gastrothylax.

grisea Cerf., 1899a, 376, t. h. Hexacanchus griseus.—1899: Squalonchocotyle.

grisea Bosc, 1802, 257.—1802: Plan.

grænlandicus Levin., 1881a, 78, t. h. Cottus scorpius; Egedesminde.—1881: Gyrodactylus.

grossa Mueller, 1774, 67.—1774: Fasc. 1776: Plan.

grossa Mueller, 1788, 21.—1788: Hirudo. 1865: Phylline. 1828: Malacobdella.

grossa Goto, 1894a, 220, t. h. Thynnus sp.; Misaki, Japan.—1894: Hexacotyle.

gruis Gmelin, 1790a, 3055, t. h. Grus.—1790: Fasc. 1803: Dist. [1850: D. echinatum.]

guernei Moniez, 1891, 148, t. h. Thynnus alalonga; Europe.—1891: Nematobothrium. qulo Mueller, 1774, 56, free form.—1774: Fasc. 1776: Plan.

gulosum Lint., 1901, 415, t. h. Rhombus triacanthus.—1901: Dist.

gurltii Cobbold, 1860a, 42, t. h. Lacerta agilis.—1860: Monost.

gurnardi Ben. & Hesse, 1863, 1864, 103, t. h. Trigla gurnardus.—1863: Phyllocotyle (type).

gurnardi Ben. & Hesse, 1863, 1864, 96, t. h. Trigla gurnardus.—1863: Platycotyle (type).

[gutturalis (Bucephalus), reptile.]

gyrini Linst., 1884, 141, t. h. tadpole of Rana temporaria.—1884: Dist.

gyrinus Mueller, 1773, 64, in infusioni animali raro.—1773; Cerc. 1827: Macrocercus. hæmatobe Ben., 1858a, 1861a, 219, for hæmatobium (Dist.).

hæmatobia crassa, see crassa.—1895: Bilharzia.

hæmatobia magna Kowal., 1895, see magna.—1895: Bilharzia.

hæmatobium Bilharz, 1852a, 72, t. h. Homo; Egypt.—1852: Dist. 1858: Schistosoma (type). 1858: Gynæcophorus (type). 1859: Bilharzia (type). 1860: Thecosoma (type). 1886: D. (Bilharzia). [Hæmatobium.]

hæmatobium venæ portarum Hæk or Pag., 1859, 42, for hæmatobium (Dist.).

hæmatobium hominis Dies., 1855, 63, for hæmatobium.—1855: Dist. [1895: Bilharzia.] hæmatoma Braun, 1891d, 426, see hematoma (Dist.).

haimeanus Lacaze-Duthiers, 1854a, 294, t. h. Ostrea edulis, Cardium rusticum; Balearic Isles, Mahon, Cette.—1854: Bucephalus. 1855; B. (Bucephalopsis, type). 1856; Cerc. [Gasterost. grascilescens.]

haimejanus Erc., 1881e, 41, 87, for haimeanus (Bucephalus = Cerc.).

halecis Gmelin, 1790a, 3058, based on Leeuwenhoek, epist. 97, 47, t. h. halece.—1790: Fasc. 1803: Dist. [1809: Dist. ocreatum.]

halosauri Bell, 1887, 116, t. h. Halosaurus macrochis; Cape St. Vincent. 1887: Dist.
hamatum Rathke, 1843, 238, "for Phylline hippoglossi," on Pleuronectes hippoglossus;
Europe.—1843: Trist. [1850: Phylline hippoglossi.] [1858: Epibdella hippoglossi.]
[1879: Trist. hippoglossi.]

hameatobia Kowal., 1895g, 58, for hæmatobia (Bilharzia).

harengi Ben. & Hesse, 1863, 1864, 98, t. h. Clupea harengus.—1863: Octocotyle. 1879: Octobothrium. 1889: Octoplectanum.

hassalli Goto, 1895, 352, t. h. Kinosternon pennsylvanicum; Md., U.S. A.—1895: Polyst.

hasta Looss, 1902m, 686, t. h. Chelone mydas; Egypt.—1902: Octangium.

haukesi Sons., 1895, 187, for hawkesii, Amphist. (Pseudodiscus).

hawkesi Braun, 1893d, 466, for hawkesii (Amphist.).

hawkesii Cobbold, 1875n, 818, t. h. Elephas indicus; India.—1875: Amphist. 1895:
A. (Pseudodiscus).

helicis Meckel [? see helicis pomatiæ]. Cercariæum. [1899: Dist. leptosomum.]

helicis Leidy, 1847, 220, t. h. Helix alternata.—1847: Dist. [1855: Cercariæum helicis alternatæ.] [1858: Cercariæum vagans.]

helicis alternatæ Dies., 1855a, 389, Dist. helicis Leidy 1847 renamed.—1855: Cercariæum.

helicis asperx Dies., 1850a, 302, t. h. Helix aspera, based on Duj., 1845a, 472.—1850: Heterost. 1850: Dist. 1855: Cercariæum.

helicis aspersæ Dies., 1855a, 398, for helicis asperæ.—1855: Heterost. 1855: Cercariæum. 1856: Cerc.

helicis carthusianellæ Par., 1894, 164, t. h. Helix carthusianella; Bologna.—1894: Cerc.

helicis maculosæ Par., 1894, 164, t. h. Helix maculosa; Bologna.—1894: Cerc.

helicis pomatiæ Dies., 1850a, 303, based on Meckel, 1846, 5, t. h. Helix pomatia.—1850: Heterost. 1850: Dist. 1855: Cercariæum.

helicis viviparæ Dies., 1850a, 298, t. h. Paludina vivipara; Vilnæ.—1850: Cerc.

helluo Mueller, 1774, 64.—1774: Fasc. 1787: Planaria.

helostomatis MacCallum, 1905, 673, t. h. Helostoma temmincki; Palembang, Sumatra.—1905: Cladorchis.

heluans Braun, 1899g, 490, t. h. Ardea cœrulea, Nycticorax gardeni; Rio de Janeiro.—1899: Clinost.

hematoma Semprum, 1890, 596, in Homo; Cuba.—1890: Dist.

hemicyclum Mol., 1859, 829, t. h. Belone acus; Batavii.—1859: Dist. 1886: D. (Echinost.).

hendorstii Lint., 1889, 163, t. h. Coryphæna hippurus; Caleta buena, Chile.—1889: Phylline. 1894: Epibdella. 1902: E. (Phylline). 1903: E. (Benedenia).

hendorfi Mont., 1902, 144, for hendorffii (Epibdella).

hepatica Rœderer, 1762, 537, see Fasc. muris.

hepatica Linn., 1758a, 648, t. h. Ovis aries; Europe.—1758: Fasc. (type). ——: Dist. (type). 1836: Dyst. 1845: Dist. (Cladocœlium, type). 1845: Fasciolaria. 1863: Dist. (Fasc.). 1892: Cladocœlium (type).

hepatica angusta Rail., 1895, 338, t. h. cattle; St. Louis, Senegal, Africa.—1895: Fasc. 1898: Dist.

hepatica apri Gmelin, 1790a, 3054, based on Le Clerc, 1715a, 119; t. h. Susscrofa.—1790: Fasc.

hepatica boum Gmelin, 1790a, 3054, t. h. Bos taurus; Europe.—1790: Fasc.

hepatica caviæ Sons., 1896, 112, t. h. Cavia.—1896: Fasc.

hepatica cervi Gmelin, 1790a, 3054, t. h. Cervus.—1790: Fasc.

hepatica equi Gmelin, 1790a, 3054, t. h. Equus caballus.—1790: Fasc.

hepatica ovata plana Buchholzii Jærdens, 1802, 64.—1802: Fasc. [Dicrocælium lanceatum.]

hepatica porcorum Gmelin, 1790a, 3054, t. h. Sus scrofa dom.; Europe.—1790: Fasc.

hepaticum Betegh, in Gomy, 1898, 328, for "amphistome hépatique" = A. explanatum.—1898: Amphist.

hepaticum ægyptiaca Looss, 1896b, 33, t. h. buffles, bœufs, moutons; Egypt.—1896: Dist. 1898: Fasc.

hepaticum (perniciosum) Taylor, 1884, 52, see hepatis perniciosum.—1884: Dist.

hepaticum suis Willach, 1893, 40, t. h. Sus scrofa dom.—1893: Monost. [1894: Cysticercus tenuicollis.]

hepatis endemicum Bælz, 1883, 234, t. h. Homo; Japan.—1883: Dist. [1907: Clonorchis.]

hepatis innoccuum Caræs, 1888a, 41, for h. innocuum (Dist.).

hepatis innocuum Baelz, 1883, 236, t. h. Homo; Japan.—1883: Dist. [1907: Clonor-chis sinensis.]

hepatis perniciosum Baelz, 1883, 234, t. h. Homo; Japan.—1883: Dist. [1907: Clonorchis endemicum.]

hepatium Rivolta, 1884, 27, for hepaticum (Dist.).

herdmani Shipley & Hornell, 1904, 78, t. h. Margaritifera vulgaris; Ceylon.—1904: Musalia (type).

heteracanthus Massa, 1903, see heterachanthus (Trochopus).

heterachanthus Massa, 1903, 252, t. h. Trigla corax.—1903: Trochopus.

heterobranchi Wedl, 1861, 478, t. h. Heterobranchus anguillaris; Egypt.—1861: Monocerca, type.

heterocerca Goto, 1894a, 197, t. h. Seriola quinqueradiata; Hiroshima (Ujina Port), Mitsugahama, and Mitsaki, Japan.—1894: Axine.

heteroclitum Mol., 1859, 289, t. h. Perdix coturnix; Batavii.—1859: Dist. 1892: Mesogonimus. 1899: Clinost.

heterocotyle Ben., 1870, 67, t. h. Clupea sprattus.—1870: Octost. 1879: Octobothrium. 1885: Octoplectanum.

heterolecithodes Braun, 1899, 3, t. h. Porphyrio porphyrio; Madagascar, Africa.—1899: Dist. 1899: Athesmia (type).

heteromorphum Crep., 1837a, 317, t. h. Trigonocephalus sp.—1837: Dist.

heterophies Perroncito, 1879, 6, for heterophyes (Dist.).

heterophyes Sieb., 1853, 62, t. h. Homo; Egypt.—1853: Dist. 1866: [Heterophyes, type.] 1858: Dicrocœlium. 1860: Fasc. 1890: Mesogonimus. 1899: Cœnogonimus (type). 1899: Cotylogonimus (type). 1900: Heterophyes (type).

heterophyes hominis Dies., 1855, 64, for heterophyes.—1855: Dist.

heteroporum Duj., 1845a, 402, t. h. Vespertilio pipistrellus; Rennes.—1845: Dist. (Brachycœlium). 1899: Pycnoporus (type). 1899: Lecithodendrium.

heterostomum Rud., 1809a, 50, t. h. Ardea purpurea; t. l. apparently Europe.—1809: Dist. 1845: Dist. (Dicrocœlium). 1899: Clinost. 1899: Dicrocœlium.

heurteli Poir., 1885, 9, t. h. Thynnus vulgaris.—1885: Dist.

hexacantha Par. & Perugia, 1889, 740, t. h. Serranus gigas; Genova.—1889: Placunella. 1906: Trochopus.

hians Rud., 1809a, 359, t. h. Ardea nigra; Greifswald.—1809: Dist. 1845: Dist. (Dicrocœlium). 1899: Cathæmasia (type).

hiatulæ Goto, 1899a, 281, t. h. Hiatula onitis; Newport, R. I.—1899: Microcotyle.

hillii Johnston, 1904, 110, t. h. Larus novæ hollandiæ.—1904: Holost.

himantopodis Rud., 1819a, 87, t. h. Charadrius himantopus; Cat. Mus. Vien.—1819: Monost.

hippocrepis Dies.; 1850a, 324, t. h. Hydrochærus capybara; Brazil. 1850: Monost.
 hippoglossi Oken, see Tasch., 1878, 568.—1878: Trist. 1879: (Epibdella). 1899: Phyllonella.

hippoglossi pleuronectes Mont., 1889, 117 (Epibdella) see Trist. uncinatum.

hippoglossi Mueller, 1776a, 220 t. h. Hippoglossus; Denmark.—1776: Hirudo. 1815: Phylline (type?). 1828: Epibdella (type). 1878: Nitzschia. 1899: Phyllonella. 1905: Phyllonella (Epidella). [Entobdella.]

hippoglossii Ben., 1858a, 1861a, 21 for hippoglossi Mueller, 1776.—1858: Epibdella. 1858: Nitzschia. 1858: Nitchia. [1878: Trist.]

hippopodii Vogt, 1854, 97, t. h. Hippopodius leteus; Quoy et Gaimard, Mediterranean.— 1854: Dist.

hirndinis Brand., 1888a, 13, for hirudinis (Dist.).

hirsutum Looss, 1896b, 68, t. h. caméléon; Alexandria, Egypt.—1896: Dist. 1899: Lecithodendrium.

hirta Mueller, 1786, 128, in aqua marina bis tantum.—1786: Cerc. 1827: Coleps, type.
hirudinaceum Bartels, 1834, 61, t. h. Salmo lavaretus; St. Petersburg.—1834: Octobothrium. 1845: Octobothrium (Cyclocotyle). [1850: Discocotyle.] 1858: Placoplectanum (Discocotyle).

hirudinis Mont., 1888, 10, for hirundinis (Echinella).

hirudinis Henle, see Dies, 1850a, 418, t. h. Hirudo vulgaris.—1850: Dist. [1850: Hepast. hirudinum.]

hirundinum Zed., 1800a, xvii, 163, for hirundinis Frælich.—1800: Dist. [1850: D. maculosum.] [1902: Plagiorchis maculosus].

hirudinum Schomburgk, 1844, 136, includes hirudinis, t. h. Nephelis vulgaris, Clepsine complanatum.—1844: Heptost. (type). [Heptast.]

hirudo Dies., 1836d, 238, t. h. Palamedea cornuta; Engenho do Cap Gama, Brazil.— 1836: Amphist.

hirudo Johnston, 1846a, 437.—Planaria.

hirundinaceum Dies., 1850a, 424, for hirudinaceum (Discocotyle).

hirundinis Frælich, 1791, 75, t. h. Thurmschwalbe.—1791: Fasc. 1800: Dist.

hirundinis Ben. & Hesse, 1863, 1864, 94, t. h. Trigla hirundo.—1863: Echinella (type). 1878: Udonella.

hispida ventriculi accipenseris sturionis Viborg, 1795, 243; see hispidum.—1795: Dist.

hispidum Abildg. in Rud., 1819a, 118, 423, t. h. Accipenser sturio; Arimini, Berlin.— 1819: Dist. (Echinost.). 1858: Echinost. 1902: Deropristis (type).

histiophori Bell, 1891a, 534, t. h., Histiophorus brevirostris.—1891: Trist. [1899: T. læve.]

histrionella Ehrenberg.——Cerc.

histrix Dies., 1850a, 393, for hystrix (Dist.) Duj., 1845.—1850: Dist. 1904: Stephanochasmus.

histrix Mol., 1858, 128, t. h. Pelophylax esculentus; Patavii.—1858: Monost.

hamatobia Bourel-Roncière, 1878a, 116, for hamatobia (Bilharzia).

holostomoides Mehlis in Crep., 1846, 148, t. h. Colymbus cristatus.—1846: Monost. 1904: Taphrogonimus, type.

holostomum Rud., 1819a, 94, t. h. Rallus aquaticus; Vien. Mus.—1819: Dist. 1892: Cladocœlium. [1902: Urogonimus macrostomus.]

homoeostomum Dies., 1858, 343, based on Bellingham, 1844, 428, t. h. Trigla (Pini) cuculus; Ireland.—1858: Dist.

hominis Lewis & McConnell, 1876, 182, t. h. Homo; Calcutta, India.—1876: Amphist. 1895: A. (Gastrodiscus). 1896: Gastrodiscus.

hominis Dies., 1855, 63.—1855: Dist. hæmatobium.

hominis Dies., 1855, 64—1855: Dist. heterophyes.

hominis Taylor, 1884, 44, see sub Distomata.

homolostomum Linst., 1887, 104, t. h. Limnæa stagnalis.—1887: Dist.

horridum Leidy, 1850, 303, t. h. Boa constrictor.—1850: Dist. 1895: D. (Dicrocœ-lium). 1899: Plagiorchis.

horridus Dies., 1850a, 400, t. h. Didelphys myosurus, D. philander; Brazil.—1850: Rhopalophorus. 1898: Rhopalias.

hospes Looss, 1907, 478, t. h. cattle; Cairo, Egypt, from Soudan.—1907: Dicrocœlium.
 hospitale Staff., 1900, 403, t. h. Diemyctylus viridescens.—1900: Dist. 1902: D. (Brachycœlium). 1902: Brachycœlium.

humana Gmelin, 1790a, 3053, t. h. Homo.—1790: Fasc.

hyalinum Rud., 1809a, 389, eriocis renamed, t. h. Salmo eriox.—1809: Dist.

hyalinum Schlotthauber, 1860, 129, t. h. Machetes pugnax.—1860: Monost.

hyalocauda Haldeman (1840a).—Cerc.

hyans Moul., 1856a, 49, for hians (Dist.).

hyatinum Kroyer, 1843–45a, 624, for hyalinum (Dist.).

hylæ Rud., 1819a, 121, t. h. Hyla arborea.—1819: Dist. [1850: D. cygnoides.]

hymenocerca Villot, 1875, 479, t. h. Calyptræa sinensis; Roscoff.—1875: Cerc.

hystrix Duj., 1845a, 433, t. h. Pleuronectes maximus, P. platessa.—1845: Dist. (Echinost.). 1899: Anoiktost. 1899: Stephanost. 1904: Stephanochasmus.

hystrix Duj. of Olss.—1899: Stephanost.

hystrix Brand., 1892, 506, for histrix, t. h. Rana esculenta.—1892: Monost.

hyterophytes Cobbold, 1883, 401, for heterophyes (Dist.).

idi Rud., 1819a, 87, t. h. Cyprinus idus; Cat. Ent. Vienn.—1819: Monost. (?Hypost.). 1819: Monost. [1850: Aspidogaster limacoides.]

ignorata Raspail, 1902, 119.—1902: Planaria.

ignotum Nicoll, 1906, 514, t. h. Hæmatopus ostralegus.—1906: Monost.

ijima Goto, 1894a, 230, t. h. Trygon pastinaca; Japan.—1894: Monocotyle.

illatabile Braun, 1901g, 897, t. h. Falco nitidus; Brazil.—1901; Scaphiost. (type).

illense Ziegler, 1883, 543, t. h. Esox lucius, Leuciscus erythrophthalmus; Strassburg.—1883: Gasterost.

illiciens Braun, 1901g, 944, t. h. Rhamphastus sp., Pipra rupicola; Brazil.—1901: Dicroccelium.

[illotum Sluiter, 1898 (Dist.), a tunicate.]

imbricata Looss, 1893a, 20, t. h. Paludina vivipara La.; near Leipzig.—1893; Cerc. [1902: Notocotyle verrucosa.]

imbutiforme Mol., 1859, 844, t. h. Labrax lupus; Batavii.—1859: Dist. 1886: D. (Echinost.). 1898: Echinost. 1899: Anoiktost. 1901: Acanthochasmus.

imitans Muehl., 1898a, 17, t. h. Abramis brama; Kœnigsberg i. Pr.—1898: Dist. 1899: Asymphylodora.

imparispine Lint., 1905, 327, t. h. Rachycentron canadus; Beaufort, N. C.—1905: Dist.
 impleta Looss, 1899b, 590, t. h. Tetrodon fahaka; Cairo, Egypt.—1899: Astia.
 Astiotrema.

impudens Crep., 1846, 149, t. h. Squalus griseus.—1846: Monost.

incerta Cobbold, 1885g, 177, t. h. Coluber; Rio Plata.—1885; Dist.

incistidata Perroncito, 1880, July, 454, t. h. Rana esculenta.—1880: Cerc.

[incistidata Erc., 1881, 96.—1881: Dist.]

incisum Rud., 1809a, 361, anarrhichæ lupi renamed.—1809: Dist. 1904: Fellodist. (type).

incivile Leidy, 1856b, 44, t. h. Leiostomus obliquus.—1856: Dist.

inclusum Polonio, 1859, see Par., 1894, 149, t. h. Triton punctatus; Padova.—1859: Dist.

incommodum Leidy, 1856b, 43, t. h. Alligator mississippiensis; Florida.—1856: Monost. 1891: Dist.

incomptum Stoss., 1886, 51, misprint for incomtum 1819 (Dist.).

incomtum Rud., 1819a, 683, t. h. Chætodon sp.; Brazil.—1819: Dist.

inconstans Lint., 1905, 327, t. h. Chætoditperus faber; Beaufort, N. C.—1905: Dist.

incrassatum Dies., 1850a, 390, t. h. Lutra solitaria; Brazil.—1850: Dist. 1860: Echinost. 1892: D. (Echinost.).

increscens Olss., 1868, 36, t. h. Scomber, Merlucius, Hippoglossus.—1868: Dist. 1886: D. (Echinost.). [1905: Lepodora rachiæa, type.]

indicum Montgomery, 1906, 139, t. h. donkey; India.—1906: Schistosoma.

inerme Nitzsch, MS., in Rud., 1819a, 375, t. h. Anas boschas fera.—1819: Dist. [1850: D. oxycephalum.]

inerme Fil., 1857c, pl. 2, for D. inerme paludinæ impuræ.—1857: Dist.

inerme Linst., 1879, 183, t. h. Petromyzon fluviatilis.—1879: Dist.

inerme Fuhrmann, 1904, 63, t. h. Lutra.—1904: Echinost.

inerme Par. & Perugia, 1889, 747, t. h. Corvina nigra; Genova.—1889: Calceost.

inerme paludinæ impuræ Fil., 1857c, pl. 2.—1857: Dist.

inflatum Crep., 1849, 64, t. h. Alauda arvensis.—1849: Dist.

inflatum Mol., 1859, 826, t. h. Anguilla vulgaris; Batavii.—1859: Dist. 1886: D. (Echinost.). 1898: Echinost. 1899: Anoiktost. 1902: Deropristis.

inflexa Rud., 1802b, 82, t. h. Cyprinus jeses.—1802: Fasc. 1809: Dist.

ingens Moniez, 1886, 531, t. h. unknown.—1886: Dist. 1902: Hirudinella.

inhærens Dalyell, 1853a, 262, t. h. ling.—1853: Octodactylus (type). [1864: Pterocotyle palmata.] [1879: Octobothrium palmatum.]

innocuum Baelz of Taylor, 1884, 53, for D. hepatis innocuum.—1884: Dist. [1907: Clonorchis sinensis.]

innocuum hepatis La Clinica de Malaga, 1883, 309, for D. hepatis innocuum. 1883:

inocuum Laspeyres, 1904a, 6, for innocuum (Dist.).

inops Looss, 1902n, 887, t. h. Pelecanus onocrotalus, Milvus ægyptius; Egypt.—1902: Heterophyes.

inquieta Mueller, 1786, 121, in aqua marina.—1786: Cerc. 1850: Histrionella.

inquilina Graff, 1904, 457, t. h. snails.—1904: Planaria.

insigne Dies., 1850a, 347, D. scimna Risso, 1826, renamed, t. h. Echinorhinus spinosus.—1850: Dist. [1899: D. veliporum.]

insignis Looss, 1899b, 596, t. h. Fulica atra; Egypt.—1899: Urogonimus.

insignis Graff, 1904, 457, t. h. snails.—1904: Planaria.

insignis Leidy, 1858, 18, t. h. Anadonta fluviatilis, A. lacustris.—1858: Cotylaspis (type). 1893: Aspidogaster.

instabile Duj., 1845a, 412, t. h. Sorex fodiens; Rennes.—1845: Dist. (Brachylaimus). instar Looss, 1901, 562, t. h. Thalassochelys corticata.—1901: Enodiotrema.

integerrima Freelich, 1791a, 104, t. h. unknown.—1791: Linguatula. 1808: Polyst. (type). 1809: Polyst. (Hexast., type.) 1828: Hexathyridium. 1828: Hexathiridium.

integerrinum Ben. & Hesse, 1864, 84, for integerrinum (Polyst.).

integrum Dies., 1850a, 429, MS.—1850: Trist. [1850: Trist. coccineum.]

intermedium Mehlis in Creplin, 1846, 138, t. h. Colymbus cristatus, C. subcristatus.—1846: Dist.

intermedium Johnston, 1904, 109, t. h. Cygnus atratus.—1904: Hemist.

intermedius Uličný, 1878, 211, t. h. Anodonta cellensis.—1878: Bucephalus.

interruptum Mont., 1891, 101, t. h. Thynnus brachypterus.; Naples.—1891: Trist.

interruptus Braun, 1901g, 897, t. h. Alcedo viridirufa, Ardea virescens; Brazil.—1901: Opisthorchis.

intersectus Lænnec, 1812c, 9, t. h. Homo; Europe.—1812: Distomus (type).

intestinale Rud., 1819a, 119, see Dist. aluconis intestinale.

intestinale Tayler, see Carter, 1862a, xxxi, t. h. Homo; District of Dacca, India.—
1862: Dist.

intestinalis Mueller.— —: Planaria.

intestinalis Linnæus, 1758a, 649, in intestinis piscium.—1758: Fasc. [Ligula.]

intestinalis Gueldenstadt, teste Rud., 1810a, 24, t. h. Sorex moschatus.—Fasc. [1810: Ligula.] [1850: Cephalocotyleum.]

intestinalis ardex nigræ Viborg, 1795, 242.—1795: Dist.

intestinalis testudinis mydæ Viborg, see Rud., 1809a, 433.—Dist. [1809: Dist. testudinis mydæ.]

intestinalis vulpis Viborg, 1795, 242.—1795: Dist.

invaginatum Mayer, 1841, 17, D. appendiculatum, renamed.—1841: Dist.

inversus Looss, 1907, 486, t. h. Vesperugo kuhli; Cairo, Egypt.—1907: Pycnoporus.

involutum Rud., 1809a, 377, t. h. Upupa epops; includes Fasc. upupæ Schrank, 1790, and D. fusiforme from Upupa epops.—1809: Dist.

irroratum Rud., 1819a, 105, t. h. Testudo mydas; Arimini.—1819: Dist. [1899: Astia.] 1902: Pachypsolus (type).

isabellinum Ratzel, 1868, 153, t. h. Gadus æglefinus.—1868: Monost. [1878: Rhipidocotyle gracilescens.]

ischnum Leidy, 1890, 415, t. h. Saurus fœtens, at Beach Haven, N. J.—1890: Distishikawæ Goto, 1894a, 234, t. h. Lethrinus sp.; at Hagi, Japan.—1894: Epibdella. 1903: E. (Benedenia).

ishikawai St. Remy, 1898, 532, for ishikawæ (Epibdella).

isopori Looss, 1894a, 55, in Cyclas rivicola.—1894: Cerc. [Dist. isoporum.]

isoporum Looss, 1894, 49, t. h. Cyprinus carpio, Phoxinus lævis, Leuciscus rutilus,
 Abramis brama, Squalius cephalus, Tinca vulgaris, Esox lucius; Germany.—1894:
 Dist. 1899: Creadium (type). 1900: Allocreadium (type).

isoporum armatum MacCallum, 1895, 401, t. h. Aplodmotus grunniens; North America. 1895: Dist. [Creadium.]

isostomum Rud., 1819a, 105, t. h. Astacus fluviatilis.—1819: Dist.

isostomum Rud., 1814a, 100, Amphist. anatis tadornæ and S. candida renamed, t. h. Anas tadorna; Copenhagen.—1814: Amphist. 1845: Holost. [1850: H. erraticum.]

italicum Stoss., 1893, (6), t. h. Lichia amia; Triest.—1893: Dist.

jacksoni, see jacksonii (Fasc.).—1892: Dist. 1899: Fasciolopsis.

jacksonii Cobbold, 1869, 48, t. h. Elephas indicus.—1869: Fasc. 1892: Dist.

janus Kowal., 1898, 73 or 130, t. h. Anas boschas dom.—1898: O. crassiuscula var. 1898: Opisthorchis. 1898: Campula.

japonicum Katsurada, 1904, 147, t. h. Felis catus dom., Homo; Japan.—1904: Schistosomum. 1905: Schistosoma.

japonicum Bl., 1888a, 596, t. h. Homo; D. hepatis endemicum renamed, see endemicum.—1888: Dist. [1907: Clonorchis endemicus.]

japonicus Braun, 1901, 17, in hen's egg; Yedo, Japan.—1901: Prosthogonimus.

[jardinii (Bucephalus), reptile.]

jesis Gmelin, 1790a, 3058, t. h. Cyprinus jeses; Europe.—1790: Fasc.

jheringhii Mont., 1899, 79, for jheringii (Temnocephala).

jheringii Haswell, 1893e, 96, t. h. Ampullaria; Brazil.—1893: Temnocephala.

kampanulatum Schneidemuehl, 1896, 302, for campanulatum.—1896: Dist.

kantaniana, see cantaniana.

kölikerii Cobbold, 1860a, 30, pelagiæ Kölliker, renamed.—1860: Dist.

köllikerii Mont., 1893, 122, for kölikerii (Dist.).

kommutatum Schneidemuehl, 1896, 303, for commutatum Dies., q. v. (Dist.).

kongenitum Schneidemuehl, 1896, 302, for conjunctum (Dist.).

konica for conica.

konikum Schneidemuehl, 1896, 303, for conicum (Amphist.).

kowalewski Rail., 1899, 788, for kowalewskii (Schistosoma).—1902: Bilharzia.

kowalewskii Par. & Ariola (1896), 114, t. h. Larus melanocephalus.—1896: Bilharzia. 1899: Schistosoma. 1899: Bilharziella.

krassum Schneidemuehl, 1896, 302, for crassum (Dist.).

kræyeri Leuck., 1847, 147, t. h. Caligus sp. on Gadus sp.—1847: Amphibothrium (type).

kröyeri Wierezejski, 1877, 550, for kroyeri (Calicotyle).

kræyeri Hœk, 1856a, 507, for kroyeri (Calicotyle).

kræyerii Ben., 1870, 16, for kroyeri (Callicotyle).

krohnii Kœlliker, 1849d, 65, t. h. a cephalopod.—1849: Dyst. [Dist.]

kroyeri Dies., 1850a, 431, t. h. Raja radiata; Kattegat.—1850: Calicotyle (type). 1858: Callicotyle. 1898: Calycotyle.

kroyeri Dies., 1850a, 427, for kræyeri (Amphibothrium).

kuhni Cobbold, 1860a, 39, M. leporis, renamed, t. h. Lepus cuniculus—1860: Monost. [Cysticercus pisiformis.]

kuneatum Schneidemuehl, 1896, 303, for cuneatum (Dist.).

labiatum Rud., 1819a, 108, t. h. Syngnathus pelagicus; Naples.—1819: Dist.

labii Linst., 1889, 79, misprint for labri Stoss. (Dist.).

labracis Duj., 1845a, 398, t. h. Labrax lupus; Rennes.—1845: Dist. (Dicrocœlium). 1870: Echinost. 1870: D. (Echinost.). 1899: Dicrocœlium. 1901: Allocreadium.

labracis Cerf., 1895, 125, t. h. Labrax lupus; White Bank, N. Sea.—1895: Diclidophora. labracis Ben. & Hesse, 1863, 1864, 112, t. h. Labrax lupus.—1863: Microcotyle (?type). labri Rud., 1819a, 122, t. h. Labrus rupestris; C. E. V.—1819: Dist. [1850: D.

fasciatum.]

labri Ben., 1870, 45, n. sp., t. h. Labrus maculatus.—1870: Dist.

labri Stoss., 1886, 30, t. h. Labrus mixtus; locality not given.—1886: Dist. 1886: D. (Dicrocœlium). 1901: Allocreadium. 1902: Loborchis.

labri rupestris Olss., 1876, 20.—1876: Dist. 1886: D. (Apoblema). 1891: Apoblema.
lacertæ Dies., 1850a, 331, t. h. Lacerta agilis.—1850: Monost. [Cf. Piestocystis dithyridium.]

lacertæ Rud., 1819a, 121, t. h. Lacerta cærulescens; C. E. V.—1819: Dist. [1850: D. mentulatum.]

lachrymosus Braun, 1902b, 31,t. h. Larus maculipennis; Brazil.—1902: Philophthalmus.
laciniatum [Blainv., 1824] Duj. 1845a, 437, t. h. Simia maimon; Paris.—1845: Dist. [1824: "Alaire" type.] [1850: Alaria Blainv., type.]

laciniatum Mol., 1859, 821, apparently lapsus for fimbriatum.—1859: Gasterost.

lacrhyma Bory de St. Vincent, 1823a, 354, in infusions d'orge et d'avoine.—1823: Cerc. lactea Mueller, 1774, 61, free form.—1774: Fasc. 1787: Planaria.

lacteum Jægers., 1896, 167, t. h. Cottus scorpius.—1896: Monost. 1899: Galactosomum (type).

læve Lint. 1898, 517, t. h. Macrourus bairdii.—1898: Dist. 1899: Hemiurus.

læve Verrill, 1875, 40, t. h. Tetrapturus albidus; North America.—1875: Trist. [1906: T. ovale.]

laeve var. armata Goto, 1899a, 273, for læve Verrill and histiophori of Bell.—1899: Trist.

læve var. inermis Goto, 1899a, 273, ovale renamed.—1899: Trist. [1899: T. ovale.] lævis Ben. & Hesse, 1864a, 87, t. h. Mustelus lævis.—1864: Erpocotyle (type).

lagena Rud., 1809a, 366, as Distomate lagena, see lagena 1788.

lagena Braun, 1788, 237, t. h. Perca fluviatilis; Europe.—1788: Planaria. 1790: Fasc. 1809: Dist. [1809: Dist. nodulosum.]

lagena Mol., 1858, 127, t. h. Strix passerina; Patavii.—1858: Holost.

lagena Brand., 1888, 249, ascidia Ben., 1873, not Rud., 1819, renamed.—1888: Dist. 1899: Lecithodendrium (type).

lageniforme Lint., 1898, 524, t. h. Remora remora.—1898: Dist.

lambitans Braun, 1899g, 490, t. h. a "Reiher;" Semanabay, West Indies.—1899: Clinost.

lampetræ Gulliver, 1872, 103, t. h. Planer's lamprey; Canterbury.—1872: Neuronaia.
lampridis Lænnberg, 1891, 73, t. h. Lampris guttatus; Kristiania Mus.—1891: Didymozoon.

lancea Dies., 1850a, 334, t. h. Delphinus tacuschi; Barra do Rio Negro, Brazil.—1850: Dist. 1892: D. (Dicroccelium). 1901: Opisthorchis.

lanceatum Stiles & Hass., 1896, 158, lanceolatum Rud., 1803 [not Schrank, 1790] renamed.—1896: Dicrocœlium (type). 1899: Dist.

lanceolata Schrank, 1790, 123, t. h. Cyprinus brama.—1790: Fasc.

lanceolata Schulze in Zeringer (1829), see Crep., 1839a, 291.—? (1829): Cyclocotyla. (1839): Cyclocotyle. [1850: Discocotyle sagittata.] [1879: Octobothrium sagittatum.]

lanceolata Rud., 1803, 24, t. h. Homo; Europe.—1803: Fasc. 1825: Dist. 1845: D. (Dicroccelium, type). 1856: Dicroccelium.

lanceolatum F. S. Leuck., 1827a, 24, t. h. Clupea alosa; Germany.—1827: Octobothrium (type). 1850: Octocotyle (type). 1858: Octoplectanum (type). [Mazocraes alosæ.] [Octostoma alosæ.]

lanceolatum Wedl, 1858, 251, t. h. Himantopus rubropterus.—1858: Monost. 1902: Hæmatotrephus (type).

lanceolatus alosæ Mayer, 1841, 23, Octobothrium lanceolatum, renamed.—1841: Decacotylus.

lanceolum Gronkowski, 1902a, 519, for lanceolatum (Dist.).

lanceulatum Baldi, 1900a, 224, for lanceolatum (Dist.).

lapridis Sars, teste Nordmann, 1840, 600, in Lampris gullatus.—1840: Hexacotyle.

lari glauci Rud., 1819a, 92, t. h. Larus glaucus; Cat. Ent. Vien.—1819: Amphist. [1850: Hemist. spathaceum.]

lasium Leidy, 1891a, 415, t. h. Ilyanassa obsoleta; Beach Haven, N. J.—1891: Dist.

lata Lespés, 1857, 114, t. h. Venus decussata; Arcachon.—1857: Cerc. 1858: C. (Gymnocephala).

tateralis Looss, 1902h, 138, t. h. Mugil auratus, M. chelo.—1902: Haploporus.

laticolle Rud., 1819a, 117, t. h. Caranx trachurus; Naples and Arimini.—1819: Dist. (Echinost.) 1899: Tergestia. 1899: Echinost.

laticolle Mueh., 1896, 590 [not Rud., 1819], t. h. Anas glacialis; apparently Kænigsberg i. Pr. See platyurum.—1896: Dist.

latiuscula Geeze, 1782a, 169, includes Fasc. hepatica Linn.—1782: Planaria.

laureatum Zed., 1800a, 164, t. h. Salmo trutta; Europe.—1800: Dist. 1802: Fasc.
 1828: Lobostome, type. 1845: Dist. (Crossodera). 1860: Crossodera. 1900: Crepidost. ——: Bunodera.

[laysani Sluiter, 1900, 9 (Dist.) a tunicate.]

[leachii Stevens, 1827 (Distomus), beetle.]

lecithonotus Luehe, 1900, 555 [see philodryadum], t. h. Coluber sp., C. eririo, Philodryas schotti; Brazil.—1900: Opisthogonimus (type).

leidyi Cobbold, 1860a, 10, Clinost. dubium Leidy, renamed.—1860: Dist.

lemna Mueller, 1773, 67, in aquis paludosis.—1773: Cerc. (?type). 1850: Histrionella.

lenoiri Poir., (1885) 1886, 3, t. h. Tetrathyra vaillantii; Senegal.—1885: Cephalogonimus (type).

lenoiri Poir., 1886, 20 t. h. Tetrathyra vaillantii; Senegal.—1886: Aspidogaster. 1892: Platyaspis (type). 1902: Cotylaspis.

lenoirii Mont., 1888a, 16, for lenoiri (Aspidogaster).

lenticola Linst., 1878, 226, t. h. Abramis vimba.—1878: Diplost. 1892: Tetracotyle.
lentis Gescheidt, 1833, 421, t. h. Homo.—1833: Monost. 1860: Festuc. 1892: Monostomulum. 1894: Dist.

lepidotus Looss, 1907, 481, t. h. Vesperugo kuhli; Cairo, Egypt.—1907: Parabascus (type).

leporis Kuhn, 1829, 464, t. h. Lepus cuniculus.—1829: Monost. [Cysticercus pisiformis.]

leptogaster F. S. Leuck., 1830a, 612, t. h. Chimæra monstrosa; Europe.—1830: Octobothrium. 1845: Octobothrium (Cyclocotyle). 1850: Discocotyle. 1858: Placoplectanum (Discocotyle). 1890: Octobothrium).

leptosoma Villot, 1878, 32, cercarian stage of Dist. leptosomum, in Scrobicularia tenuis.—1878: Cerc.

leptosomum Crep., 1829, 57, t. h. Tringa variabilis.—1829: Dist. 1845: D. (Echinost.). 1860: Echinost.

leptostomum Olss., 1876, 18, t. h. Meles taxus.—1876: Dist. 1892: D. (Brachylaimus) 1899: Harmost. (type). 1899: Heterolope (type).

leucochloridii Leuck., 1858a, 114, see Leucochloridium paradoxum.—1858: Cerc. 1858: Dist.

lerinseni Odhn., 1905, 348, t. h. Gadus saida; East Greenland.—1905: Hemiurus.

levinsenii Setti, 1898, 311, for levinsenii (Trist.).

levinsenii Mont., 1891, 101 t. h. Thynnus sp.—1891: Trist.

lichiæ Ariola, 1899, 1, t. h. Lichia amia; Genova.—1899: Microcotyle.

lidiæ Par., 1902, 6, for lydiæ (Echinost.).

liqula Ben., 1870, 1871a, 17, t. h. Scymnodon ringens; Portugal.—1870: Dist.

liguloideum Dies., 1850a, 320, t. h. Vastres cuvieri; Borbæ, Brazil.—1850: Monost. 1892: Amphilina.

liliputanum Looss, 1896, 141, t. h. Pernis apivorus, at Alexandria, Egypt; Milvus parasiticus, at Matarijeh.—1896: Echinost.

lima Rud., 1809a, 408, t. h. Vespertilio auritus, V. murinus; Europe.—1809: Dist.
 (Echinost.). 1892: D. (Brachylaimus). 1899: Plagiorchis (type). [1899: Lepoderma.]

limacis Dies., 1850a, 302, based on Duj., 1845a, 472, in Limax agrestis, L. cinerea,
L. rufa; Rhedoni.—1850: Dist. 1850: Heterost. 1855: Cercariæum. 1856: Cerc.
[1858: C. (Acanthocephala) trigonocerca.]

limacoides Dies., 1834a, 1231; 1835c, 421, t. h. Cyprinus dobula, C. idus; Cat. Vien.— 1834, 1835; Aspidogaster. 1835; Monost.

limatulum Braun, 1900f, 389, t. h. Molossus sp.; from Brazil.—1900: Dist. [1907: ? Parabascus.]

limnææ ovatæ Linst., 1885, 251.—1885: Dist.

limnææ ovatæ Linst., 1884, 142.—1884: Cerc. [1889: Dist. endolobum Duj.]

limnææ truncatulæ Linst., 1892, 331, t.h. Limnæa truncatula.—1892: Cerc.

limnæi Dies., 1850a, 302, based on Duj., 1845a, 473, t. h. Limnæus palustris; Rhedoni.—1850: Dist. 1850: Heterost.

limnæi Pag., 1857, 32, t. h. Lymnæus stagnalis.—1857: Tetracotyle. [1858: T. typica.] limnophili Linst., 1879, 185, t. h. Limnophilus rhombicus; apparently Germany.—1879: Dist. [1889: D. endolobum.]

limuli Graff (1879), 202.—1879: Plan.

lineare Rud., 1819a, 83, t. h. Tringa vanellus; Mus. Vien.—1819: Monost. (Monost.).
[1850: Notocotyle triseriale.] [1896: N. verrucosum.]

linearis Looss, 1901, 618, t. h. Chelone mydas; Egypt.—1901: Pleurogonius.

linearis Rud., 1793a, 29, t. h. Phasianus gallus; Greifswald, October.—1793: Fasc. 1803: Dist. 1860: Crossodera. 1896: Bunodera.

linearis Mueller, 1774, 67, in foveis palustribus sylvarum.—1774: Fasc. 1787: Plan.

linearis Lespés, 1857, 117, t. h. Littorina littorea.—1857: Cerc. 1858: C. (Acanthocephala).

[linearis (Nitzschia) a diatome.]

linearis longa Linn. teste Pallas, 1781a, 95, in Cyprinus.—1781: Fasc.

lineata Mueller, 1774, 60, in littore maris Balthici.—1774: Fasc. 1776: Plan.

lineatus Scott, 1901, 143, t. h. Trigla lineata; Clyde.—1901: Trochopus.

lineola Dies., 1850a, 346, t. h. Falco rufus; C. E. V.—1850: Dist.

lingua Bosc, 1802a, 262.—1802: Plan.

lingua Crep., 1825, 47, t. h. Larus marinus.—1825: Dist. 1892: D. (Dicroccelium). Tocotrema (type). 1899: Cryptocotyle. 1899: Cotylogonimus (Cryptocotyle). 1903: Cryptocotyle.

linguæforme Dies., 1850a, 335, Brachylæmus erinacei Bl., 1847, renamed; t. h. Eri-

naceus europæus; Paris.—1850: Dist. 1898: Mesogonimus.

linguale Odhn., 1902, 66, t. h. Gymnarchus niloticus; Sudan.—1902: Phyllodist.

linguatula Rud., 1819a, 100, t. h. Rana; Brazil.—1819: Dist. ?: D. (Brachylaimus). linguatula Loos, 1899b, 668, t. h. Chelonia mydas; Egypt.—1899: Microscapha. 1902: Microscaphidium. 1902: Polyangium (type).

linstowi Mont., 1893, 102, for linstowii (Dist.).

linstowii Stoss., 1890, 42, Monost. aculeatum Linst., from Testudo græca, renamed.— 1890: Dist. 1895: D. (Dicrocœlium). 1899: Telorchis Luehe. 1899: Telorchis (type) Looss. 1900: T. (Cercorchis, type).

lintoni Pratt, in Lint., 1901b, 435, D. auriculatum Wedl of Lint., in Acipenser

rubicundus, renamed.—1901: Bunodera.

biorchis Fischder., 1901, 368, t. h. Cervus simplicicornis, C. campestris, C. mexicanus, C. rufus, C. dichotomus, C. namby; Brazil.—1901: Paramphist.

littoralis Herbst, 1787a, 35.—1787: Plan.

lobatum Rail., 1900, 241, t. h. Accipiter nisus.—1900: Dicroccelium. 1902: Lyperosomum.

lobatus Looss, 1901l, 619, t. h. Chelone mydas; Egypt.—1901: Glyphicephalus.

lobianchi Mont., 1888a, 7, t. h. Raia clavata.—1888: Acanthocotyle (type).

lobianchii Mont., 1891a, 104, for lobianchi (Acanthocotyle).

lobianchoi Goto, 1899, 285, for lobianchi (Acanthocotyle).

lobiancoi Mont., 1899, 75, for lobianchi (Acanthocotyle).

lobotes MacCallum, 1895, 406, t. h. Anguilla chrysopa, Perca flavescens, Stegostedion vitreum; North America.—1895: Dist. 1904: Centrovarium (type).

loliginis delle Chiaje, 1823, pl. 92, t. h. Loligo vulgaris; Naples.—1823: Polyst. [1850: Solenocotyle chiajei (type).]

loliginis Gmelin, 1790a, 3059, includes F. barbata.—1790: Fasc. 1803: Dist. [1850: Tetrabothriorhynchus migratorius.]

loliginis delle Chiaje, 1841a, 140.—1841: Amphyst. [Amphist.] 1894: Monost.

longe Braun, 1892a, 581, for longum (Diplost.).

longicauda Rud., 1809a, 372, t. h. Corvus cornix.—1809: Dist. [1819: D. macrourum.] 1899: Dicroccelium. [1899: Lyperosomum.] 1902: Lyperosomum (type).

longicauda Goto, 1899a, 282, t. h. Cynoscion regale; Newport, R. I.—1899: Microco-

longicaudata Piana, 1882, see Par., 1894, 161, t. h. Helix carthusiana.—1882: Cerc. [See also longocaudata.]

longicolle Crep., 1846, 154, t. h. Cottus gobio.—1846: Dist.

longicolle Crep., 1825, 57, t. h. Perca vulgaris, P. cernua; includes D. embryo Olfers.—1825: Dist. [1850: D. embryo.]

longicolle Rud., 1819a, 87, t. h. Ardea alba, A. stellaris, Larus ridibundus, L. atricilla; Mus. Vien.—1819: Amphist. 1845: Holost. 1905: Strigea.

longicollis Rud., 1809a, 380, based on Bloch, 1782a, 6, t. h. Esox lucius.—1782: Fasc. [1809: Dist. tereticolle.]

longicollis Abildg., [1788], 34, t. h. Coluber natrix.—[1788]: Fasc. 1860: Dist.

longicollis Frœlich, 1791, 73, t. h. Cyprinus carpio.—1791: Fasc. [1809: Dist. globi-porum.] 1881: Dist.

longicollis Dies., 1850a, 417, Octost. merlangi Kuhn, renamed, t. h. Merlangus vulgaris.—1850: Diclidophora. 1859: Octoplectanum. [1879: Octobothrium merlangi.]

longipenis Looss, 1899, 608, t. h. ape; Ghizeh Zool. Garden.—1899: Phaneropsolus.

longipes Dies., 1850a, 428, tubiporum 1835 renamed, t. h. Trigla hirundo.—1850: Trochopus (type). [1864: T. tubiporus.]

longiplexus Staff., 1902, 901, in lungs of American frogs and toads.—1902: Hæmatolæchus. 1905: Pneumonœces.

longispina Klein, 1905, 65, t. h. Rana hexadactyla; India.—1905: Halipegus.

longissimum Linst., 1883, 308, t. h. Ardea stellaris; Turkestan.—1883: Dist. 1892: D. (Dicrocœlium). 1896: Opisthorchis .

longissimum Poir., 1886, 29, t. h. Delphinus tursio.—1886: Dist. 1892; D. (Dicrocœlium). [1896: D. tursionis.]

longissimum corvinum Stiles & Hass., 1894, 418, t. h. Corvus americanus, C. ossifragus;
 U. S. A.—1894: Dist. (Dicrocœlium). 1899: Opisthorchis.

longissimum eorvinum Stiles & Hass., 1896, 155, misprint for corvinum (Dist. longissimum).

longiusculus Looss, 1902m, 582, misprint for longiusculus (Pleurogonius).

longiusculus Looss, 1901l, 568, t. h. Chelone mydas; Egypt.—1901: Pleurogonius (type).
 longocaudata Piana, 1882, teste Par., 1894, 622 (misprint?) in Helix carthusianella;
 Reggio Em.—1882 or 1894: Cerc.

longum Brand., 1888a, 25, t. h. Crocodilus; Brazil.—1888: Diplost.

longum Leidy, 1851, 206, t. h. Esox estor; at Cleveland, Ohio.—1851: Dist. 1904: Megadist. (type).

loossi Ssinitzin, 1905, 33, G. cygnoides of Looss renamed.—1906: Gordodera.

loossii Marshall & Gilbert, 1905, 483, t. h. Micropterus salmoides, Lucius lucius, Amia calva.—1905: Azygia.

lophocerca Fil., 1857c, 5, t. h. Paludina impura; Torino.—1857: Cerc. 1858: Glenocerc. lorenzii Mont., 1899, 1045, t. h. Trigla sp.; Rovigno.—1899: Plectanocotyle.

lorum Duj., 1845a, 407, t. h. Talpa europæa; Rennes.—1845: Dist. (Brachylaimus).
1893: Mesogonimus. 1899: Dolichosomum (type). [1899: Ityogonimus (type).]
1900: Dolichodemas (type).

loxiæ Rud., 1819a, 120, t. h. Loxia chloris, L. coccothraustes, L. pyrrhula; C. E. V.— 1819: Dist. [D. mesostomum.]

lubens Braun, 1901, 945, t. h. Pipra rupricola.—1901: Dicroccelium.

lucaneum Brand., 1892b, 511, for Monost. (Glenocerc.) lucanica in Planorbis parvus.— 1892: Monostomulum.

lucania Leidy, 1904a, 143, for lucanica (Monost.).

lucanica Leidy, 1877, 200, t. h. Planorbis parvus; U. S. A.—1877: Monost. (Glenocerc.). [1892: Monostomulum, see lucaneum.]

lucii Mueller, 1776, 224, t. h. Lucius, esoph., stomach.—1776: Fasc. 1782: Plan. 1800: Dist. [D. tereticolle.]

lucii Rud., 1819a, 122, for D. esocis lucii Rud., 1809a, 438, t. h. Esox lucius; Greifs-wald.—1819: Dist.

lucioperca Mueller, 1776, 223, t. h. Perca lucioperca.—1776: Fasc. 1803: Dist. [D. nodulosum.]

lucipetum Rud., 1819a, 94, t. h. Larus fuscus, L. glaucus; Mus. Vien.—1819: Dist.
 1828: Fasc. 1845: D. (Dicroccelium). 1899: Philophthalmus.

lühei Odhn., 1905, 351, t. h. Clupea harengus, C. sprattus.—1905: Hemiurus.

luna Mueller, 1786, 139, in aqua rarissime.—1786: Cerc. 1815: Furcocerca. 1827: Lecane.

lunatum Dies., 1836d, 238, t. h. Cervus dichotomus [?], Anas melanotus , Λ. ipecutiri, Himantopus wilsonii; Caiçara, Brazil.—1836: Amphist.

lunatus Looss, 19011, 558, t. h. Thalassochelys corticata; Triest.—1901: Pachypsolus (type). [See irroratus.]

8588-No. 37-08-4

lungocaudata Piana teste R. Bl., 1888a, 603 see also longicaudata), t. h. Helix carthu siana.—1888: Cerc.

lupi Ben. & Hesse, 1863, 92, t. h. Labrax lupus.—1863: Udonella.

lupus Mueller, 1773, 67, in aquosis, ubi Lemna vegetat, rara.—1773: Cerc. 1815: Furcocerca. 1827: Dicranophorus. [1828: Cyclogena (type).]

lusca Ben., 1868a, 7, for luscæ (Dactycotyle), in Morrhua lusca.

luscæ Ben. & Hesse. 1863; 1864, 111, t. h. Morrhua lusca.—1863, 1864; Dactycotyle. 1879; Dactylocotyle. 1879; Octobothrium. 1890; O. (Dactylocotyle).

lutea Ben. of Giard, 1897c, 954, t. h. Tapes decussatus and T. pullastra at Arcachon and Donax trunculus at Wimereux.—1897: Cerc. [1897: Brachycœlium luteum.] lutea Ben., 1870, 3. t. h. Scyllium canicula.—1870: Dist. 1897: Brachycœlium.

1897: Cerc. 1904: Diphterost. [D. betencourti.]

luteum Baer, 1826a. 125; 1827, 610. t. h. Paludina vivipara; Regiomonti,—1826: Dist. [1850: Heterost. ovatum.]

luzii Dies.. 1850a. 358, for lucii (Mueller) Zed., (Dist.).

lydiæ Stoss., 1896, 190, t. h. Orthagoriscus mola: Triest.—1896: Echinost. 1899: Dist.
 1899: Anoiktost. 1899: Stephanost. 1901: Dihemistephanus (type).

lymnæi auricularis Fil., 1854, 25, t. h. Lymnæus auricularis.—1854: Dist. 1855: Cercariæum. 1856: Cerc.

lymnæi obscuri Erc., 1881e, 33, t. h. Limnæus obscurus, L. stagnalis: Bologna.—1881:

lymnæi palustris Dies., 1855a, 399, for limnæi 1850a, 302.—1855: Cercariæum.

lymnæi peregri Dies., 1858d, 279, t. h. Lymnæus pereger; Turin.—1858: Cercariæum. lymphaticum Linst., 1903, 353, t. h. Mustelus vulgaris.—1903: Dist. [1906: D. megastomum.]

lyratum Schlotthauber, 1860, 129, t. h. Ardea cinerea.—1860: Holost.

macaci Cobbold, 1861, 119, t. h. Macacus radiatus.—1861: Cerc.

macconnelli Cobbold, 1876, 97, D. sinense renamed.—1876: Dist. [1907: Clonorchis sinensis.]

macdonaldi Mont., 1891, 120, t. h. Melo sp.: Shark Bay, West Australia.—1891: Aspidogaster.

macdonaldii Mont., 1891, 121, for macdonaldi (Aspidogaster).

macorcerca Looss, 1894a, 253, for macrocerca (Cerc.).

macrobothrium Ben., 1870, 70, t. h. Osmerus eperlanus.—1870: Dist.

macrocephalum Rud., 1803a. 21. t. h. Strix bubo: includes Strigea and Fasc. strigis.—
[1782: Planaria.] [1788: Festuc.] [1790: Fasc.] [1790: Strigea, type.] 1803: Amphist. (type). [1819: Holost. variabile, type.] 1828: Holost. (type). [1850: Hemist.]

macrocephalum [pars: falconis milvi] Rud., 1819a. 88.—1819: Amphist. [1840: A. striatum.] [1850: Hemist. spathula.]

macrocerca Fil., 1854a, 13, t. h. Cyclas cornea; Turin.—1854: Cerc. 1855: C. (Xiphidiocerc.) 1858: C. (Acanthocephala). [1858: Dist. cygnoides.] [1902: Gorgodera cygnoides.]

macrocotyle Dies., 1858, 342, t. h. Orthagoriscus mola, in Ireland; based on Bellingham, 1844, 429.—1858: Dist. [1860: D. macrocephalum.] 1886: D. (Cladocœ-lium). 1893: D. (Accacœlium). [1893: D. megnini.] 1898: Podocotyle. 1898: D. (Podocotyle). 1901: Accacelium.

macrolaimus Linst., 1894b. 334. t. h. Vesperugo pipistrellus.—1893: Dist. 1899: Lecithodendrium.

macrophallos Linst., 1875a, 190. t. h. Totanus hypoleucos.—1875: Dist. 1892: D. (Brachycœlium.) 1900: D. (Levinsenia). 1899: Levinsenia.

macrophallus Linst., 1887, 104, for macrophallos Dist.).

macropoculum Cobbold, 1860a. 25. macrocotyle 1858 renamed, t. h. Orthagoriscus mola.—1860: Dist.

macroporum Mont., 1893, 133, t. h. Lophius piscatorius.—1893: Dist.

macropterum Wien. MS. in Brandes, 1888a, 55. syn. of grande Dies.—1888: Hemist. [1888: Diplost. grande.]

macrorchis Brand., 1892, 508, t. h. marine turtles.—1892: Monost.

macrostoma Rud., 1803a, 26, t. h. Nachtigall [Motacilla luscinia]; Greifswald, June.—
1803: Fasc. 1809: Dist. 1893: Urogonimus (type). 1896: D. (Urogonimus).
[1835: Leucochloridium paradoxum, type.] 1907: Leucochloridium, type.

macrostomum Schlotthauber, 1860, 130, t. h. Petromyzon fluviatilis.—1860: Dist.

macrostomum Rud., 1809a, 337, t. h. Larus cinerarius; Greifswald.—1809: Monost. (Monost.).

macrostomum Jægers., 1900a, 33, t. h. Telmatias major; ?Upsala.—1900: Diplost.

macrourum Rud., 1819a, 98, t. h. Corvus cornix; D. longicauda Rudolphi, 1809a, 372 renamed.—1819: Dist. 1853: Dist. (Dicrocœlium). 1899: Dicrocœlium. [1902: Lyperosomum longicauda and Dicrocœlium albicolle.]

macrurum Schlotthauber, 1860, 129, t. h. Corvus glandarius.—1860: Monost.

macrurum Braun, 1892a, 746, for macrourum (Dist.).

maculata Leidy, 1847e, 252, free; Phila.—1847: Plan.

maculatum Looss, 1901d, 402, t. h. Labrus merula, Crenilabrus pavo, C. griseus; Triest.—1901: Dist.

maculatum Rud., 1819a, 123, t. h. Diodon; California.—[1811: Capsala martinieri, type.] 1819: Trist. 1840: Capsala. 1850: Trist.

maculosa Rud., 1802, 67, F. hirundinis renamed, t. h. Hirundo apus.—1802: Fasc. 1809: Dist. 1845: D. (Brachylaimus). 1892: D (Dicroccelium). 1901: Plagiorchis.

madagascariensis Vayssière (1892), 64.—1892: Temnocephala. 1899: Dactylocephala (type).

magna Cobbold, 1859d, 364, t. h. Cercopithecus fuliginosus.—1859: Bilharzia. 1892: Gynæcophorus. [1895: Bilharzia hæmatobia.] 1899: Schistosoma.

magna Pag., 1857, 22, t. h. Paludina vivipara.—1857: Cerc. 1858: C. (Gymnocephala).
magniovatum Stoss., 1898, 53, t. h. Puffinus kuhlii; Triest.—1898: Echinost.

magnum Bassi, 1875, 497, t. h. [Hirsch]; Mandria.—1875: Dist. 1889, 1894: Fasc.

maimonis Blainv., 1828, 586, t. h. Simia maimon.—1828: Fasc.

maior Tasch., 1879, 263, for major (Dactylogyrus).

major Goto, 1894a, 203, t. h. Scomber colias; Japan.—1894: Octocotyle.

major Nitzsch, 1817, 44.—1817: Cerc. [1850: Histrionella lemna.]

major Wagener, 1857, 99, "pl. 15, fig. 9," t. h. Gobius fluviatilis.—1857: Dactylogyrus. majus St.-Remy, 1898, 547, for major (Octocotyle).

malleus Linst., 1877, 182, t. h. Barbus fluviatilis.—1877: Dactylogyrus.

mancupatus Fischder., 1901, 371, t. h. African cattle.—1901: Gastrothylax.

mansoni Sambon, 1907, 365, t. h. Homo; Africa.—1907: Schistosomum.

marænulæ Rud., 1809a, 339, t. h. Salmo marænula; Europe.—1809: Monost. (Hypost.) 1892: Monostomulum.

marculentum Braun, 1901g, 948, t. h. Emberiza citronella; Vien. Mus.—1901: Dist.

margaritarum Dubois, 1901, 603, t. h. Mytilus edulis.—1901: Dist.

margaritiferæ Shipley & Hornell, 1904, 78, t. h. Margaritifera vulgaris; Ceylon.—1904: Muttua (type).

margaritiferæ Shipley & Hornell, 1904, 78, t. h. Margaritifera vulgaris; Cheval Paar, Ceylon.—1904: Aspidogaster.

marginatum Rud., 1819a, 680, t. h. Ardea sp.; Brazil.—1819: Dist. 1892: Mesogonimus. 1899: Clinost.

marginatum Mol., 1858, 128, t. h. Anas crecca; Batavii.—1858: Dist. 1892: D. (Brachylaimus).

marilæ Rud., 1819a, 87, t. h. Anas marila; Cat. Ent. Vien.—1819: Monost. [1850: Notocotyle triseriale.]

marillæ Dies., 1850a, 411, for marilæ (Monost.).

marina Garsin, 1730a, 387, t. h. Scomber pelamys.—1730: Hirudinella.

marionis St. Loup, 1885, 176, t. h. Mæna vulgaris.—1885: Choricotyle.

marmorosa Mueller, 1774, 71, in fossis aquaticis rara.—1774: Fasc. 1787: Plan.

marsupium Braun, 1901g, 941, t. h. Perdix rufina; Brazil.—1901: Harmost.

martinieri Bosc, 1811, 384, t. h. Diodon sp.—1811: Capsala (type).

martiranoi Stiles, 1903aa, 15, t. h. Anopheles claviger.—1903: Agamodist.

medians Olss., 1876, 25, t. h. Bufo vulgaris.—1876: Dist. 1899: Pleurogenes.

medians Olss. of Staff., 1900, Aug., 412.—1900: Dist. [1905: Loxogenes arcanum.]

medioplezus Staff., 1902, 901, t. h. American frogs and toads.—1902: Hæmatolæchus. 1905: Pneumonœces. [See Ostiolum formosum, type.]

medioximus Braun, 1901g, 895, t. h. Galbula grandis; Brazil.—1901: Eumegacetes.
medius Kath., 1894a, 129, t. h. Cobitis fossilis, Cyprinus carpio.—1894: Gyrodactylus.
megachondrus Looss, 1899b, 593, t. h. Testudo (græca?).—1899: Enodia (type).
1901: Enodiotrema (type).

megacotyla Dies., 1858d. 263, for Dist. echinatoides Pag., 1857, 32 [not Fil.], t. h. Anodonta cygnea; Heidelberg.—1858: Cerc. (Nephrocephala).

megacotyle Dies., 1850a, 379, D. velellæ Fil. renamed, t. h. Velella spirans; Naples.—1850: Dist.

megacotyle Dies., 1836d, 238, t. h. Silurus palmito; Matogrosso, Brazil.—1836: Amphist. megacotyle Linst., 1906, 176, t. h. Histiophorus sp.; Beruwala.—1906: Trist.

megacotylea Villot, 1878, 30, t. h. Mysis.—1878: Cerc.

megalocephalum Brand., 1888a, 67, t. h. Stomias sp.; Brazil.—1888: Holost.

megalocotyle Mont., 1893, 52.—1893: Dist.

megaloon Linst., 1879, 337, t. h. Lacerta agilis.—1879: Dist.

megastomum Wagener, 1857, 57, t. h. Cyprinus blicca, C. amarus.—1857: Dactylogyrus.
 megastomum Rud., 1819a, 102, t. h. Squalus galeus; Arimini.—1819: Dist. 1886: D. (Brachylaimus). 1900: Ptychogonimus (type).

megastomum Grobben, 1878a, 89, t. h. Portunus depurator.—1878: Dist.

megastomum leporis Kuhn (1829c).—1829: Dist.

megastomus Looss. 1902m, 533, t. h. Chelone mydas: Egyptian coast—1902: Cricocephalus.

megatocyle Linst., 1903, 354, misprint for megalocotyle Mont. (Dist.).

megnini J. Poir., 1885, 4, t. h. a fish.—1885: Dist. 1893: D. (Accacœlium). [1893: D. macrocotyle.]

megninii Mont., 1893, 102, for megnini 1885 (Dist.).

melanocystis Staff., 1904, 483, t. h. Lophius piscatorius; Canada.—1904: Xenodist (type).

melanoglena Dies., 1855a, 393, Melanoglena bipunctata renamed.—1855: Histrionella. 1858: Glenocerc.

melanoglena Pag., 1862, 298.—1862: Cerc.

melanops Dies., 1855a, 400, based on Cerc. paludinæ impuræ Baer, 1827b, 655.—1855: Cercariæum. 1858: Histrionellina.

melis Schrank, 1788, 17, t. h. Dachs, Meles; Europe: Planaria teres Goeze, pro parte, renamed.—1788: Fasc. 1800: Dist. 1809: Plan. [1850: D. trigonocephalum.]
melolonthæ Hammerschmidt, in Leuck., 1835, 88, nomen nudum, t. h. an insect;

Europe.—1835: Klepsitromis (type).

mentolatum Mueh., 1898, 19, for mentulatum.—1898: (Dist.).

mentulatum Rud., 1819a. 103, t. h. Coluber natrix Lacerta agilis. L. maculata; Europe— 1819: Dist. 1845: D. (Brachylaimus). 1899: Lepoderma. 1899: Plagiorchis.

mergi Rud., 1819a, 121, t. h. Mergus albellus; C. E. V.—1819: Dist. [1850: D. baculus.]

merlangi Kuhn, 1829b, t. h. Gadus merlangus and Cymothoa oestroides on Boops.— 1829: Octost. 1832: Octobothrium. 1832:? Polyst. 1838—40: Diclidophora. 1845: Octobothrium (Cyclocotyle). 1850: [Diclidophora longicollis]. 1888: Octocotyle. 1895: Dactylocotyle.

merlangi carbonarii Dies., 1858e, 341, renamed anonymum 1858e, 341: based on Bellingham, 1844, 428.—1858: Dist.

merlangi vulgaris Dies.. 1858e. 341, renamed anonymum 1858e, 341; based on Bellingham, 1844, 428.—1858: Dist.

merlangorum Dies., 1855, 64, based on Bellingham, 1844.—1855: Dist.

merluccii Ben. & Hesse, 1863, 1864, 105, t. h. Merlucius vulgaris.—1863 or 1864: Anthocotyle (type).

merlucii Ben. & Hesse, 1863, 1864, 93, t. h. Merlucius vulgaris.—1863: Udonella. merlucii Tasch., 1879k, 247, for merluccii (Anthocotyle).

meropis Rud., 1819a, 120, t. h. Merops apiaster; C. E. V.—1819: Dist. [1850: D. triangulare.] 1896: D. (Brachylaimus).

mesocolium Cohn, 1903, 35, t. h. Draco volans; Java.—1903: Hoploderma (type). [1907: Pintneria, type.]

mesostoma Rud., 1803a, 28, t. h. Krametsvogel [Turdus iliacus]; Greifswald, November.—1803: Fasc. 1809: Dist. 1892: D. (Brachylaimus). 1902: Harmost.

mesosternum Linst., 1873, 101, apparently for mesostomum (Dist.).

metœcus Braun, 1900f, 389, t. h. Vespertilio lasiopterus, V. noctua: Vien. Mus.—1900:

Dist. 1900: Crepidost. (type).

mexicana Vayssière, 1898, 227, t. h. Cambarus digneti; Mexico.—1898: Temnocephala. michælis Mont., 1892, Oct. 7, 168, t. h. Cantharus vulgaris: Triest.—1892: Cotylogaster (type).

micracantha Dies., 1858d, 259, syn. Cerc. armata Fil., 1855b, 3-5, t. h. Triton punctatus, Lymnæus palustris.—1858: Cerc. (Acanthocephala).

micracanthum Stoss., 1889, 29, t. h. Pagellus erythrinus; Triest.—1889: Dist. 1903:

D. (Dicrocœlium).

micracanthus Massa, 1903, see micrachanthus (Trochopus)

micrachanthus Massa, 1903, 225, t. h. Trigla hirundo.—1903: Trochopus. microbothorium Fischder., 1902a, 21, for microbothrium (Paramphist.).

microbothrium Fischder., 1901a, 369, t. h. Antelope dorcas; Coll. Vien. and Vet. School, Berlin.—1901: Paramphist.

microcanthus Massa, 1906, 66, for micracanthus (Trochopus).

microcephalum Baird, 1853, 58, t. h. Acanthias vulgaris; Falmouth Harbor.—1853: Dist. [D. veliporum.]

microcephalum Crep., 1837, 311, 1849, 64, in Corvus cornix.—1837: Dist. microcephalum Rud., 1819a, 88 lapsus for microstomum.—1819: Amphist.

micrococcum Rud., 1819a, 101, t. h. Glareola austriaca; Arimini.—1819: Dist. 1901: Phaneropsolus. 1892: D. (Brachylaimus).

microcotyla Fil., 1854a, 7; 1854b, 260; t. h. Paludina vivipara, P. achatina; Lake Varese and Lombardia.—1854: Cerc. 1855; C. (Xiphidiocerc.). 1858; C. (Acanthocephala) [1858: Dist. tetracystis.] [1905: Cystagora, type.]

microcotyle Dies., 1858e, 340, t. h. Rhombus maximus; Ireland; based on Bellingham, 1844, 428.—1858: Dist.

microcristata Erc., 1881 or 1882, see Par., 1894, 161, t. h. Bythinia cristata; Bologna.— 1881?: Cerc.

microdactyla Mont., 1903, 1, t. h. Dilocarcinus septemdentatus; Mattó Grosso.—1903: Temnocephala.

micropharyngeum Luehe, 1898g, 623, t. h. flamingo; Berberei.—1898: Dist. 1900: Gymnophallus.

microphylla Ben., 1870, 70, t. h. Osmerus eperlanus.—1870: Dist.

microporum Mont., 1889, 322, t. h. Plagyodus ferox: Madeira.—1889: Dist. 1891: Apoblema. 1899: Hemiurus.

micropteri Marshall & Gilbert, 1905, 481, t. h. Micropterus salmoides, M. dolomieu; near Madison, Wis.—1905: Leuceruthrus (type).

micropterygis Richardi, 1902, 4, t. h. Micropteryx dumerilii.—1902: Didymozoon. 1902: Monost. 1902: Didymost. [1902: D. bipartitum.]

microsoma Rud., 1809a, 109, t. h. Perca marina; Naples.—1819: Dist.

microsomum Dies., 1850a, 370, for microsoma (Dist.).

microstomum Rud., 1809a, 50, t. h. Pleuronectes solea; Paris.—1809: Dist.

microstomum Rud., 1809a, 342, t. h. Corvus caryocatactes: Greifswald, Europe.—1809: Amphist. 1845: Holost.

microstomum Crep., 1829, 49, t. h. Fulica atra; Greifswald.—1829: Monost. [1850: M. mutabile.

microtyla Moul., 1856a, 80, for microcotyle (Cerc.).

micrura Fil., 1857c, 5, t. h. Paludina impura.—1857: Cerc. 1858: C. Acanthocephala). [1894: Dist. globiporum.]

midæ Dies., 1850a, 325, for mydæ.—Planaria. [1850: Monost. trigonocephalum.]

midas Kuhl & van Hasselt, (1824 or 1822?).—Polyst.

miescheri Zschokke, 1890, 764, t. h. Trutta salar.—1890: Dist. (Cladocœlium).

miescherii Mont., 1893, 153, for miescheri (Dist.).

migrans Duj., 1845a, 407, Brachylaimus advena renamed, hence type of Brachylaima; hosts Sorex araneus, S. leucodon.—1845: Dist. (Brachylaimus, type).

migras Stoss., 1892, 19, misprint for migrans 1845 (Dist.).

miliaris Brown, 1881, 329, misprint for militare 1803 (Dist.).

militaris Rud., 1803a, 30, t. h. Scolopax arquata; Greifswald, January.—1803: Fasc. 1809: Dist. (Echinost.). 1860: Echinost.

milvi Gmelin, 1790a, 3054, t. h. Milvus.—1790: Fasc. 1803: Dist. [1819: D. echino-cephalum.]

minima Ben., 1870, 67, t. h. Clupea sprattus.—1870: Dist.

minimum Wagener, 1852, 558, t. h. Trigla microlepidota.—1852: Gasterost.—1858: Rhipidocotyle.

minimum Stoss., 1887. 96, t. h. Labrax lupus; Triest.—1887: Gasterost.

minor Haswell. 1887a, 284, t. h. Astacopsis bicarinatus; New South Wales.—1887: Temnocephala.

minor St.-Remy. 1898, 551, minus Olsson, 1876 [Octobothrium] renamed.—1898: Dactylocotyle.

minor Wagener, 1857, 60, t. h. Cyprinus alburnus.—1857: Dactylogyrus.

minor Goto, 1894a, 205. t. h. Scomber colias; Japan.—1894: Octocotyle.minor Mont., 1888, 16, t. h. Scyllium.—1888: Pseudocotyle. 1905: P. (Leptocotyle, type).

minor Looss, 1901d, 437, t. h. Labrus merula; Triest.—1901: Derogenes.

minus Cerf., 1898b, 330, t. h. Raja sp.; Roscoff.—1898: Merizocotyle.

minus Olss., 1876, 10, t. h. Gadus melanostomus.—1876: Octobothrium.

minuta Looss, 1899b, 585, t. h. dogs and cats at Cairo and Ardea cinerea at Damietta, Egypt.—1899: Ascocotyle.

minuta Erc., 1881 or 1882, see Par., 1894. 163, t. h. Bythinia tentaculata; Bologna.—1881?: Cerc.

minuta Nitzsch, 1817, 46, in various fresh-water mollusks; Halle.—1817: Cerc. 1855: C. (Eucercaria). 1858: C. (Gymnocephala).

minutissimum Stoss., 1896, 130. t. h. Anas boschas; Doberdò.—1896: Monost.

minutissimus Looss, 1901, 618, t. h. Chelone mydas; Egypt.—1901: Pleurogonius.

minutum Cobbold, 1859d, 364, t. h. Hæmatopus ostralegus.—1859: Dist.

minutus Looss, 1901e, 604, t. h. Uranoscopus scaber; apparently Triest.—1900: Stephanochasmus.

minutus Fischder., 1901, 372, t. h. Antilope sp., Tragelaphus scriptus: Kamerun.—1901: Gastrothylax.

miocerca Mont., 1888, 77, for myocerca (Cerc.).

mirabilis Braun, 1891c, 218. t. h. Limnæus palustris var. corvus.—1891: Cerc.

mirus Looss, 1901d, 439, t. h. Labrus merula; Triest.—1901: Zoogonus (type).

mustroides Mont., 1896. 144. constrictum Leared. renamed.—[1896: Dist.] [1896: Mesogonimus.] [1908: Hapalotrema (type).]

mitsukurii Goto, 1894a. 227, t. h. Rhina sp.: Mitsugahama. Japan.—1894: Calicotyle. 1898: Calycotyle.

[modestum Sluiter, 1898 (Dist.), a tunicate.]

molæ Bl., 1847a, 326, t. h. Orthagoriscus mola.—1847: Trist. [1850: T. rudolphianum.] molæ Maclaren, 1904, 573, t. h. Orthagoriscus mola.—1904: Nematobothrium.

molæ Rud., 1819a, 87, t. h. Orthagoriscus mola: Naples.—1819: Monost. [1850: Dist.

okenii.]
moleculum Linst., 1880, 51, t. h. Rallus pygmæus.—1880: Dist. 1892: D. (Brachy-

moleculum Linst., 1880, 51, t. h. Rallus pygmæus.—1880: Dist. 1892: D. (Brachy-laimus).

molinii Polonio, 1859, teste Par., 1894, 148, t. h. Rana esculenta; Padua.—1859: Dist. molinii Par., 1894, 148, for molini, 1859 (Dist.).

molle Leidy, 1856, 43. t. h. Sternotherus odoratus: U. S. A.—1856: Monost. 1894: Dist. (Polyorchis). 1896: Polyorchis. 1896: Pleorchis.

mollis Wedl, 1857, 272, t. h. Cyprinus carpio.—1857: Gyrodactylus. 1858: Dactylogyrus.

mollisimum Mont., 1893, 96, for mollissimum (Apoblema).

mollissimum Levin., 1881, 59, t. h. Cottus scorpius; Egedesminde.—1881: Dist. 1886: D. (Apoblema). 1889: Apoblema. 1889: Hemiurus. [1905: Lecithaster gibbosus. 1907: Lecithaster.

mollissimun Mont., 1891, 521, for mollissimum (Apoblema).

mollissinum Mont., 1891, 520, for mollissimum (Apoblema).

molvæ Ben. & Hesse, 1863, 1864, 94, t. h. Lota molva.—1863, 1864: Pteronella (type). 1878: Udonella.

molvæ Cerf., 1895h, 944.—1895: Dactylocotyle.

monas Rud., 1819a, 679, t. h. Amphisbæna sp.; Brazil.—1819: Dist. 1896: D. (Brachylaimus).

monenteron Wagener, 1857, 52, etc., t. h. Esox lucius.—1857: Dactylogyrus. 1858: Tetraonchus, probably type.

mongeotii Bory de St.-Vincent, 1823a, 354, in water.—1823: Cerc.

monorchis Stoss., 1890, 40, t. h. Cantharus orbicularis; Triest.—1890: Dist. 1893: D. (Monorchis, type). 1902: Monorchis, type.

monostomi Linst., 1896i, in 375, perhaps young of Monost. mutabile, t. h. Lymnæa ovata, L. peregra; Germany.—1896: Cerc.

monroi Goodsir, (1844), t. h. Gadus morrhua.—1844: Neuronaia, type.

monroii Goodsir, teste Gulliver, 1872, 103, see monroi (Neuronaia).

monstruosum Braun, 1901, 944, t. h. Corone venustissima; West Indies.—1901: Ochetosoma (type).

monticelli Mont., 1902, 143, for monticellii (Epibdella).

monticellii Lint., 1898, 518, t. h. Remora remora; Woods Hole.—1898: Dist. 1899: Hemiurus.

monticellii Stoss., 1904, 2, t. h. Tropidonotus viperinus, intestine; Naples.—1904: Astiotrema.

monticellii Scott, 1902, 300, t. h. Raia clavata; Scotland.—1902: Acanthocotyle.

monticellii Par. & Perugia (1895), 2.—1895: Phylline. 1896: Epibdella (Phylline). 1903: E. (Benedenia).

mordax Looss, 1899b, 688, t. h. Pelecanus onocrotalus; Egypt.—1899: Echinost.

mordens Braun, 1901g, 941, t. h. Rallus sp.; Brazil.—1901: Harmost.

mormiri Mont., 1888a, 34, for mormyri (Microcotyle).

mormyri Lorenz, 1878a, 21, t. h. Pagellus mormyrus; Triest.—1878: Microcotyle.

mormyri Stoss., 1885, 160, t. h. Pagellus mormyrus; Triest.—1885: Dist. 1886: D. (Brachylaimus). [1899: ?Creadium.]

mormyris Hausmann, 1897b, 28, for mormyri (Dist.).

morrhuæ Ben. & Hesse, 1863, 106, t. h. Gadus morrhua.—1863: Pterocotyle. 1879: Octobothrium. 1890: O. (Pterocotyle). 1898: Dactylocotyle.

motellæ Ben., 1870, 63, t. h. Motella mustela.—1870: Dist.

[mucosum von Dr. (Dist.), a tunicate.]

mühlingi Jægers., 1899a, 7, t. h. Larus ridibundus; Pillau.—1899: Dist. 1899: Tocotrema.

muchlingi Looss, 1899b, 586, for mühlingi (Tocotrema).

muellerii, see mülleri (Dist.).

mülleri Cobbold, 1860a, 50, t. h. Petromyzon fluviatilis.—1860: Diplost.

mülleri Levin., 1881a, 56, t. h. Cottus scorpius, Gadus ovak; Egedesminde.—1881: [1902: Genarches Dist. 1886: D. (Brachylaimus). 1899: Progonus (type). (type).] 1905: Genarches (type).

mugilis Vogt, 1878, 327, t. h. Mugil cephalus; Roscoff.—1878: Microcotyle.

mugylis Sons., 1891, 253, for mugilis (Microcotyle).

mullerii Mont., 1888, 34, 37, for mülleri (Dist.).

mulli Stoss., 1883, 114, t. h. Mullus barbatus; Triest.—1883: Dist. 1886: D. (Echinost.).

multilobum Cobbold, 1860a, 46, t. h. Charadrius pluvialis.—1860: Holost.

munroi Brown, 1899a, 490, for monroi (Neuronaia).

murænulæ Cobbold, 1860a, 43, for marænulæ (Monost.).—[1860: M. braunii.]

muris Erc., 1882c, pl. 3, figs. 16-21.—1882: Dist.

muris hepatica Rœderer, 1762, 537, t. h. Mus musculus; Germany.—1762: Fasc. [1905: Cysticercus fasciolaris.]

musculi Rud., 1819a, 119, t. h. Mus musculus; Cat. Vien.—1819: Dist. [See D. recurvum.]

musculicola Waldenburg, 1860, 12, t. h. Cyprinus, Perca fluviatilis.—1860: Holost. 1892: Diplost. 1894: Tetracotyle.

musculorum Braun, 1893, 871, musculorum percæ, 1860, renamed; in Perca fluviatilis.—1893: Dist.

musculorum percæ Waldenburg, 1860, 16, t. h. Perca fluviatilis.—1860: Dist.

musculorum suis Duncker, 1896, 279, t. h. Sus scrofa dom.; Germany.—1896: Dist. [1898: Agamodistomum suis.]

musculosum Johnston, 1904, 112, t. h. Sterna bergii.—1904: Holost.

musculum Brett, 1880a, 453, t. h. Ovis aries.—1880: Dist.

mutabile Mol., 1859, 833, t. h. Podarcis muralis; Padua, Italy.—1859: Dist. 1895: D. (Dicroccelium). 1901: Dicroccelium. 1902: Anchitrema.

mutabile Zed., 1800a, xvi, t. h. Fulica chloropus; Germany.—1800: Monost. 1809: Monost. (Monost.). 1835: Amphist. 1890 (1888?): Monost. (type). [1892: Cyclocœlum, type.] [1901: Cyclocœlum, type.] 1902: Cyclocœlum (type). 1904: (Cyclocœlium).

mutabile Dies., 1850a, 301, Diplodiscus diesingi renamed, t. h. Planorbis nitidus; Ticini.—1850: Diplocotyle (type). 1894: Diplodiscus (Diplocotyle). [1858: Diplodiscus subclavatus.]

mutabilis Schrank, 1803, 210.—1803: Fasc.

mutabilis Dies., 1850a, 317, urnigerum 1819 renamed, t. h. Pelophylax esculentus; Europe.—1850: Codonocephalus (type). 1896: Diplost. (Codonocephalus).

mutabilis Dies., 1839a, 234, t. h. Cataphractus n. sp. [Cichla temensis]; Rio Negro, Brazil.—1839: Aspidocotylus (type). 1850: Aspidocotyle (type).

mutabilis Stoss., 1902, 579, t. h. Anguillula vulgaris; Triest.—1902: Loborchis. 1903: Helicometra.

mutablilis Eichwald, ——.——: Planaria.

mydx Braun, in Rud., 1809a, 336, as syn. of Monost. trigonocephalum.—1809: Planaria. mydx, see midas (Polyst.).

mydas, see midas.

myliobatis Tasch., 1878, 574, t. h. Myliobatis aquila; Naples.—1878: Monocotyle (type).

myocerca Villot, 1878, 32, t. h. Scrobicularia tenuis.—1878: Cerc.

mystacidis Linst. 1896i, 379, t. h. Mystacides nigra.—1896: Dist.

myzura Pag., 1881, 18, t. h. Neritina fluviatilis.—1881: Cerc.

naia Duj., 1845a, 387, for naja.—1845: Dist. (Dicrocœlium).

naidis proboscidiæ Dies., 1858d, 283.—1858: Cercariæum.

naja Rud., 1819a, 99, t. h. Coluber natrix; Berlin.—1819: Dist. 1845: D. (Dicrocœ-lium). 1899: Macrodera (type). 1902: Saphedera (type).

nana Rud., 1802b, 68, t. h. Scolopax gallinula; Greifswald, July.—1802: Fasc. 1809: Dist. 1901: Plagiorchis.

nanodes Braun, 1901g, 942, t. h. Falco nitidus; Brazil.—1901: Urorygma (type).

nardoi Polonio, 1859, see Par., 1894, 147, t. h. Lacerta muralis; Padua.—1859: Dist. nassæ mutabilis Fil., 1855b, 22.—1855: Dist.

natator Claparède, 1863a, 13, free.—1863: Onchogaster (type).

nattereri Cobbold, 1860a, 52, cornu Dies., 1839a, renamed; t. h. Callichthys vacu; Brazil.—1860: Amphist.

negacotyle Dies., 1850a, 379, misprint for megacotyle (Dist.).

neglecta Fil., 1854b, 278, t. h. Lymnæus pereger; prope Augustam Taurinorum.—1854: Cerc. 1855: C. (Eucercaria). 1858: C. (Gymnocephala).

neglectum Linst., 1887d, 101, t. h. Rana temporaria.—1887: Dist. 1889: D. (Dicro-cœlium). [1899: Pleurogenes claviger.]

nematoides Cohn, 1904, 238, t. h. Aquila albicilla.—1904: Pronopharynx (type).

nematoides Crep., 1846, 129, t. h. Falco albicilla.—1846: Monost.

nematoides Mueh., 1898, 18, t. h. Tropidonotus natrix; East Prussia.—1898: Dist. 1899: Telorchis.

nephriticum Mehlis, in Crep., 1846, 138, t. h. Colymbus arcticus.—1846: Monost. 1904: Eucotyle (type).

nephrocephalum Dies., 1850a, 391, t. h. Turdus saxatilis; M. C. V.—1850: Dist. 1860: Echinost.

nephrocephalum Dies., 1858e, 327, in Sphargus coriacea; N. America; renicapite Leidy, 1856, renamed.—1858: Monost.

nephropis Cunningham, 1884a, 399, t. h. Nephrops norwegicus; Firth of Forth.—1884: Stichocotyle (type).

neuronaia monroii Maddox, 1867, 97, see monroii.—1867: Dist.

neuronaii monroii Cobbold, 1872b, 52, for neuronaia monroii (Dist.).

nigra Mueller, 1774, 54, in rivo.—1774: Fasc. 1787: Planaria.

nigrescens Olss., 1876, 19, t. h. Lophius piscatorius.—1876: Dist.

nigroflavum Rud., 1819a, 118, t. h. Orthagoriscus mola; Naples.—1819: Dist. (Echinost.). 1893: D. (Accacœlium). 1899: Accacœlium. 1903: Echinost.

nigropunctatum Linst., 1883, 310, t. h. a bird "Akatza;" Turkestan.—1883: Monost. nigrostavum Deslongchamps, 1829, 238, misprint for nigroflavum Rud. (Dist.).

nigrovenosum Bellingham, 1844, 429, t. h. Tropidonotus natrix; England.—1844: Dist. 1895: D. (Dicroccelium). 1899: Lecithodendrium. 1902: Brachyccelium.

nigrovenosum natricis torquatæ Dies., 1855, 64, based on Bellingham, 1844.—1855: Dist.

nigrum Linst., 1883, 307, t. h. Corvus cornix.—1883: Dist.

nipponicum Goto, 1891a, 151, t. h. Carassius vulgaris.—1891: Diplozoon.

nitens Lint., 1898c, 534, t. h. Tylosurus caribbæus; Woods Hole.—1898: Dist.

nitidum Leidy, 1856, 45, t. h. Rana pipiens.—1856: Holost.

[nitidum Sluiter, 1898, 17 (Dist.), a tunicate.]

noctulæ Cobbold, 1860a, 39, vespertilionis Rud., renamed, t. h. Vespertilio noctula.—1860: Monost.

noctulæ Rud., 1819a, 119, t. h. Vespertilio noctula; C. E. V.—1819: Dist.

nocturnus Looss, 1907, 479, t. h. Athene noctua; Egypt.—1907: Philophthalmus.

nodulosa Linst., 1873, 3, t. h. Bythinia tentaculata.—1873: Cerc. [1894: C. virgula Fil.]

nodulosa Freelich, 1791, 76, includes percæ cernuæ 1776, t. h. Perca cernua, P. fluviatilis.—1791: Fasc. 1800: Dist. 1845: Dist. (Crossodera, type). 1860: Crossodera. [1896: Bunodera, type.] 1899: Bunodera, type.

non coronata Crep., 1837a, 326.—1837: Dist. [As name of a group of distomes.]

nordmanni Dies., 1850a, 428, t. h. Brama mediterranea.—1850: Encotyllabe (type). 1878: Trist. 1890: Plectanocotyle.

normanni Braun, 1890a, 550, for nordmanni (Encotyllabe).

notidobiæ Linst., 1896i, 379, t. h. Notidobia ciliaris.—1876: Dist.

notulata Bosc, 1802a, 254.—1802: Planaria.

novæ-zelandiæ Haswell, 1887a, 284, t. h. Paranephrops setosus; New Zealand.—1887: Temnocephala.

noverca Braun, 1902, 836, "conjunctum Cobbold" of Lewis & Cunningham, 1872, renamed, t. h. Homo; India.—1902: Opisthorchis. 1907: Dist.

nozawæ Goto, 1894a, 249, t. h. Thynnus sibi; Japan.—1894: Trist.

obesa Ben., 1870, 33, t. h. Cottus scorpius.—1870: Dist.

obesum Looss, 1902h, 135, t. h. Mugil auratus, M. cephalus, M. chelo; Triest.—1902: Saccoccelium (type).

obesum Dies., 1850a, 361, t. h. Leporinus friderici, Salminus brevidens, Xiphostoma cuvieri; Brazil.—1850: Dist.

obesus Crep., 1851b, 292, t. h. Lacerta agilis.—1851: Tetrathyrus (type). 1866: Petrathyrus. [Monost. lacertæ.] [1860: Monost. gurltii.] [Dithyridium. Piestocystis. cestode.]

obliquus Looss, 1901, 30 Oct., 566, "trigonocephalus Rud." of Looss, 1899b, 666, 756, figs. 84-86.—1901: Pronocephalus (type).

oblonga Cobbold, 1858b, 168, t. h. Delphinus phocæna.—1858: Campula (type). 1891: Dist. 1892: D. (Brachylaimus). 1898: Opisthorchis.

oblongnum Mont., 1888a, 57, for oblongum (Polyst.).

oblongum Cobbold of Braun, see Looss, 1902m, 716 (Brachycladium).

oblongum R. Wright, 1879, 12, t. h. Aromochelys (Sternothærus) odoratus; Toronto, Canada.—1879: Polyst.

obovatum Mol., 1858, 288, t. h. Chrysophris aurata: Padua.—1859: Dist. 1886: D. (Brachylaimus). [1899: Creadiinæ.] 1901: Allocreadium.

obscura Mueller, 1774, 65, in piscinis.—1774: Fasc. 1776: Planaria.

obscura Sons., 1892, Oct. 7, 138, t. h. Limnæa natalensis; Cairo, Egypt.—1892: Cerc.

obscurum Staff., 1904, 484, t. h. Lophius piscatorius; Canada.—1904; Leptosoma (type).

obscurum Leidy, 1887, 24, t. h. Megalops thrissoides; Coll. Army Med. Mus., Wash.—1887; Monost.

obtusum Looss, 1896b. 78, t. h. caméléon, Alexandria, Egypt.—1896: Dist. 1899: Lecithodendrium.

occidentalis Nickerson, 1900, 250, t. h. sheepshead: Mississippi Valley.—1900: Cotylogaster.

occidualis Staff., 1905, 687, t. h. Rana clamata, R. catesbiana; Canada.—1905: Halipegus.

occultum Staff., 1905, 682, t. h. Diemyctylus viridescens Raf., Rana virescens; Canada.—1905; Manodist. (type).

ocellata LaValette, 1855, 22, t. h. Limnæus stagnalis.—1855: Cerc. [1858: Histrionellina fissicauda.]

ocellatum Rud., 1819a, 125, t. h. Testudo orbicularis; Arimini.—1819: Polyst. 1828: Hexacoctyla. 1828: Hexacotyle.

ochreatum Nitzsch, in Giebel, 1857, 265, t. h. Falco haliaëtos.—1857: Holost.

ocreata Rud., 1802b, 79, halecis 1790 renamed; t. h. Clupea harengus.—1802: Fasc. 1809: Dist. 1845: D. (Apoblema). 1889: Apoblema. 1899: Pronopyge (type).

ocreata Gœze, 1782a, 182, t. h. Maulwurf; Europe.—1782: Fasc. 1788: Cucullanus. 17—: Festucaria (Schrank). 1800: Monost. 1809: M. (Monost). 1899: Dist. [1899: Ityogonimus, type.] 1902: Ityogonimus. [Distomum lorum.]

ocreatum Mol., of Olss., 1868, 48.—1868: Dist. 1891: Apoblema. 1899: Hemiurus. [1905: Brachyphallus crenatus (Rud.).]

ocreatum Rud. of Mol., 1858, 209, in Clupea alosa; Padua.—1858: Dist.

octopodis Cuvier, 1829b, 147.—1829; Hectocotylus (type).

octopodis delle Chiaje, 1841, 139.—1841: Dist.

octopodis delle Chiaje, teste Par., 1894, 169, t. h. Octopus vulgaris; Naples.——: Monost.

ocular de Bonis, 1882, 180, for oculare.—1882: Dist. [Agamodist. ophthalmobium.] oculare Moquin-Tandon, 1860, 347, for D. oculi humani.—1860: Dist. 1860: Fasc. [Agamodist. ophthalmobium.]

oculatum Levin., 1881a, 64, t. h. Cottus scorpius; Egedesminde.—1881; Dist. 1886; D. (Echinost.). 1905; Acanthopsolus (type).

oculi humani Gescheidt, 1833, 434, t. h. Homo: Dresden, Saxony.—1833: Dist. 1858 Dicrocœlium. [Agamodist. ophthalmobium.]

oculis Moquin-Tandon, 1861, 375, for ocularis.—1861: Fasc. [Agamodist. ophthal-mobium.]

oculobium Cohn, 1902d, 712, t. h. Vanellus melanogast.; Greifswald coll.—1902: Monost.

odontocotyla Dies., 1858d, 264, t. h. Lymnæus stagnalis; Berlin.—1858: Cerc.

œsophagi ardex nigrx Viborg, 1795, 242.—1795: Dist. [1850: D. hians.]

oidemiæ nigræ Dies., 1858e, 322, based on Bellingham, 1844a, 340.—1858: Amphist. [1858: Holost. anatis nigræ.]

okeni Ariola, 1906, 186, for okenii (Dist., Köllikeria).

okenii Kœlliker, 1849c, 55, t. h. Brama raji.—1849: Dist. [1860: Köllikeria.]

okul' humani Schneidemuehl, 1896, 302, for oculi humani.—1896: Dist.

oligoon Linst., 1887d, 103, t. h. Gallinula chloropus.—1887: Dist. 1892: D. (Brachylaimus).

oligoterus Mont., 1899, 76, t. h. Raja clavata; Naples.—1899: Acanthocotyle.

oloris Dies., 1855, 64, based on Bellingham, 1844, 427.—1855: Dist.

olssoni Odhn., 1905, 326, t. h. Gadus melanostomus; Lumprenus maculatus from west coast of Sweden.—1905. Podocotyle.

olssoni Rail., 1900, 239, t. h. Apus apus.—1900: Dicrocœlium.

onchacanthus Massa, 1906, 44, t. h. unknown; Triest.—1906: Trochopus.

onchidiocotyle Setti, 1899, 121, t. h. "tonno;" Portoferraio.—1899: Trist.

onycephalum Galli-Valerio, 1898, 923, misprint for oxycephalum (Dist.).

opaca Bory de St. Vincent, 1823a, 354, in infusions de pois.—1823: Cerc.

opaca Staff., 1902, 416, t. h. Bufo lentiginosus; America.—1902: Gorgodera. 1905: Gorgoderina.

opacum Ward, 1894, 173, t. h. Amia calva, Cambarus propinquus, Ictalurus punctatus,
 Perca flavescens; Lake St. Clair.—1894: Dist. 1894: D. (Brachycœlium). 1899:
 Levinsenia. 1901: Microphallus (type).

operculata Herbst, 1787a, 36.—1787: Planaria.

ophthalmobium Dies., 1850a, 334, oculi humani renamed; Dresden.—1850: Dist. 1892: Agamodist.

ophthalmothium Luehe in Stiles, 1903u, 223, for ophthalmobium (Agamodist.).

opisthotrias Lutz, 1895, 181, t. h. Didelphys aurita.—1895: Dist. 1899: Harmost. 1899: Heterolope.

orbicolare Sons., 1891, 262, misprint for orbiculare (Monost.).

orbiculare Dies., 1850a, 349, t. h. Cebus trivirgatus; Brazil.—1850; Dist. 1901; Phaneropsolus.

orbiculare Rud., 1819a, 83, t. h. Sparus salpa; Naples.—1819: Monost. (Monost.).

1901: Mesometra (type).

orbis Mueller, 1786, 138, in aqua, ubi Lemna, rarissime.—1786: Cerc. 1815: Furcocerca. 1827: Lecane.

oricola Leidy, 1884a, 47, t. h. Alligator mississippiensis; Florida.—1884: Dist. [1895: D. pseudostomum.]

oricula Mont., 1892, 715, misprint for oricola (Dist.).

ornata LaValette, 1855, 18, t. h. Planorbis corneus.—1855: Cerc. 1858: C. (Acanthocephala). [1858: Dist. clavigerum, type of Pleurogenes 1899.]

ornata Will.-Suhm, 1870, 9, t. h. Alligator lucius; Charleston.—1870: Polycotyle (type).

ornata Odhn., 1902, 22, t. h. Nilkrokodil.—1902: Stephanoprora (type).

ornatum Leidy, 1856, 43, t. h. Rana pipiens; Phila.—1856: Monost.

ornatum Cobbold, 1882a, 240, t. h. Elephas indicus; India.—1882: Amphist. 1895: Pseudodiscus,

ornithorhynchi Johnston, 1901, 334, t. h. Ornithorhynchus anatinus Shaw.—1901: Dist. orphii Ben. & Hesse, 1863, 116; 1864, 116, t. h. Esox belone.—1863: Axine. [1879: A. belones.]

orthagorisci molæ Dies., 1855, 64, based on Bellingham, 1844, 429.—1855: Dist. [1850: D. macrocotyle.]

orthocœlium Fischder., 1901, 369, t. h. Bos kerabau; Ceylon.—1901: Paramphist. osculatum Looss, 1901, 654, t. h. Motella vulgaris.—1901: Dist.

osleri R. Wright, 1879, 66, t. h. Necturus lateralis;? Canada.—1879: Sphyranura (type). oslerii Mont., 1888a, 8, for osleri (Sphyranura).

oti Rud., 1819a, 354, for otidis, syn. of macrocephalum.—1819: Festuc.

otidis Frœlich, 1802, 53, t. h. Strix otus.—1802: Festuc. [Amphist. macrocephalum.] ovale Goto, 1894a, 241, t. h. Histiophorus orientalis; Misake, August.—1894: Trist. [1899: T. læve.]

ovalis Schrank, 1803, 86.—1803: Cerc.

ovata Villot, 1878, 20, t. h. Lygia oceanica.—1878: Cerc.

ovata Rud., 1803a, 25, t. h. Corvus frugilegus.—1803: Fasc. 1809: Dist. 1845: Dist. (Dicroccelium). 1892: Cephalogonimus. 1896: D. (Cephalogonimus). 1899: Prosthogonimus (type). 1899: Prymnoprion (type). 1901: Prosthogonimus (type). 1902: Prostogonimus.

ovata Linst., 1877, 192.—1877: Tetracotyle. [1894: Holost. variegatum.]

ovata Goto, 1894a, 234, t. h. Anthias schegeli; Misaki, Japan, August.—1894: Epibdella. 1902: Trist. 1903: E. (Benedenia).

ovatum Lint., 1900, 269, t. h. Lobotes surinamensis; Woods Hole.—1900: Gasterost.

ovatum Dies., 1850a, 302, for Dist. luteum Baer, 2, 610, pl. 29, figs. 20–22, in Paludina vivipara; Regiomontii.—1850: Heterost. 1858: Cercariæum. [1858. Dist. luteum Wagener.]

ovatum Mol., 1859, 822, t. h. Gallinula crex; Batavii.—1859: Monost.

ovicola Brand., 1894a, 303, t. h. Arius commersonii; Brazil.—1894: Fridericianella (type).

oviforme Poir., 1886, 26, t. h. Nycticebus javanicus.—1886: Dist. 1892: Dist. (Brachycœlium). 1899: Phaneropsolus. 1899: Lecithodendrium.

ovis Cobbold, 1885a, 499, t. h. Ovis.—1885: Bilharzia.

ovocaudatum Vulpian, 1859, 150, t. h. Rana viridis.—1859: Dist. 1899: Halipegus (type).

ovofarctum Odhn., 1902, 153, t. h. Synodontis sp.; Omdurman.—1902: Dist. ovopunctatum Stoss., 1902, 15, t. h. Numenius arquata.—1902: Cyclocælum.

ovum Crep., 1846, 159, t. h. Planorbis marginatus.—1846: Dist.

oxycephalikum Schneidemuehl, 1896, 303, for oxycephalum (Dist.).

oxycephalum Rud., 1819a, 98, t. h. Anas boschas, A. clypeata.—1819: Dist. 1896: Echinost.

oxycephalum Dies., 1836d, 238, t. h. Salmo auratus, S. pacu and Silurus megacephalus at Cuyaba, and Salmo pacupeba at Rio Panara and Rio Araguay; Brazil.—1836: Amphist. 1905: Chiorchis.

oxyurum Crep., 1825, 48, t. h. Anas marila; Germany.—1825: Dist. 1902: Psilost. pachisomum Par., 1894, 155, for pachysoma (Dist.).

pachycerca Dies., 1858d, 257, includes C. brachyura Lespés, t. h. Trochus cinereus; Francogalliæ.—1858: Cerc. (Acanthocephala).

pachycerca Claparède, 1863a, 12, free form; St. Vaast.—1863: Cerc.

pachyderma Braun, 1899a, 629, t. h. Chelone atra.—1899: Dist.

pachysoma Eysenhardt, 1829, 144, t. h. Mugil auratus.—1829: Dist. 1886: D. (Podocotyle.) 1893: D. (Monorchis). 1898: Podocotyle. 1902: Haplosplanchnus (type). pacifica Steenstrup, 1842, 74.—1842: Dist.

pacifica Ben., 1858a, 1861a, 86, as syn. of Dist. militare.—1858: Cerc.

pagelli Ben., 1870, 1871a, 43, t. h. Pagellus centrodontus; Belgium.—1870; Dist. 1886; D. (Cladocœlium).

pagelli Ben. & Hesse, 1863, 80, t. h. Pagellus centrodontus.—1863: Encotyllabe. 1878: Trist.

pagenstecheri Ssinitzin, 1905, 34, Dist. cygnoides Zed. of Pag.—1905: Gorgodera.

palæmonis Linst., 1877b, 186, t. h. Palæmon serratus.—1877: Dist.

[palea (Nitzschia), a diatom.]

paliatum Braun, 1892a, 608, misprint for palliatum (Dist.).

pallasii Poir., 1885, 13, t. h. Delphinus phocæna.—1885: Dist. 1892: D. (Dicrocœlium).

palleniscum Shipley & Hornell, 1905, 53, t. h. Balistes sp.; Ceylon.—1905: Dist.

pallens Rud., 1819a, 111, t. h. Sparus aurata; Naples.—1819: Dist. 1886: (Dicrocœlium).

palliatum Looss, 1885a, 390, t. h. Delphinus delphis.—1885: Dist. 1892: Cladocœlium. 1899: Brachycladium (type). 1901: Campula.

pallidum Staff., 1904, 487, t. h. Fundulus heteroclitus; Woods Hole.—1904: Homalometron (type).

pallidus Looss, 1902n, 889, t. h. Milvus ægyptiacus; Egypt.—1902: Heterophyes.

palmatum Rentsch, 1860, 38, t. h. Seestichling, Gasterosteus spinachia.—1860: Dist.

palmatum Leuck., 1830, 612, t. h. Gadus molva; Germany.—1830: Octobothrium.
1842: Octobothrium. 1845: Octobothrium (Cyclocotyle). 1850: Diclidophora.
1859: Octoplectanum. 1864: Pterocotyle (type). 1879: Octobothrium. 1890: Octobothrium (Pterocotyle).
1895: Dactylocotyle.

paloniæ J. Poir., 1883, 73, t. h. Palonia frontalis; Java.—1883: Homalogaster, type. palpebrarum Looss, 1899, 587, t. h. Corvus cornix, Milvus parasiticus: Egypt.—1899: Philophthalmus (type).

paludinæ impuræ Baer, see Dies., 1855a, 400.—1855: Cerc. 1855: Dist. 1855: Cercariæum.

paludinæ impuræ Fil., 1854, 279, t. h. Paludina impura.—1854: Dist. 1855: Cercariæum. [1858: C. p. i. inerme.]

paludinæ impuræ armatum Fil., 1857, 207, t. h. Paludina impura.—1857: Dist. 1858: Cercariæum.

paludinæ impuræ inerme Fil., 1857, 207.—1857: Dist. 1858: Cercariæum. [1858: Dist. perlatum.]

paludina impura (tentaculorum) Dies., 1855, 399, based on Baer, 1827b, 655, in Paludina impura; Regiomontii.—1855: Cercariæum.

paludinæ viviparæ Dies., 1855, 399=helicis viviparæ renamed, t. h. Paludina vivipara; Vilnæ.—1855: Cercariæum.

[pancerii D. Valle (Dist.) a tunicate.]

pancerii Sons., 1891, 303, t. h. Umbrina cirrhosa; Italy.—1891: Microcotyle.

pancreaticum Janson, 1889a, or Rail., 1890, 143, t. h. sheep; Japan.—1890: Dist.

1897: Dicrocœlium. 1907: Eurytrema, type.

pancreaticum Katsurada & Saita, 1906, 501, t. h. cattle; Japan.—1906: Dist. [Eurytrema.]

pancreatinum Ward, 1895, 335, for pancreaticum (Dist.).

pandum Braun, 1901a, 48, t. h. Thalassochelys caretta; Naples.—1901: Monost.

panduriforme Rail., 1900, 240, t. h. Pica pica.—1900: Dicrocœlium.

pangasii MacCallum, 1905, 668, t. h. Pangasius nasutus; Palembang, Sumatra.—1905: Cladorchis.

papillatum Rud., 1814a, 105, t. h. Anas boschas fera; Germany.—1814: Dist.

papillatum Ben., 1858a, 38.—1858: Trist.

papillatum Cobbold, 1882, 240, t. h. Elephas indicus; India.—1882: Amphist.

papilliferum Mol., 1859, 290, t. h. Belone acus; Batavii.—1859: Dist. 1886: D. (Crossodera).

papilliferum Braun, 1892a, 586, for papillatum Cobbold, 1882 (Amphist.).

papillorum Juel, 1889, 14, for papillosum (Trist.).

papillosa Erc., 1881 or 1882, see Par., 1894, 164, t. h. Bythinia tentaculata; Bologna.— 1881?: Cerc.

[papillosa Crep., 1837a, 326, name of a group of Dist.]

papillosa Lint., 1898, 508, t. h. Gadus callarias; Woods Hole.—1898: Nitzschia. 1904: Lintonia (type).

papillosum Dies., 1836, 313, t. h. Xiphias gladius.—1836: Trist. 1840: Capsala.

papillosum Dies., 1850a, 381, D. beroës Will, renamed; t. h. Beroe rufescens; Triest.— 1850: Dist. 1860: Crossodera.

paradoxum Nord., 1832a, 56, t. h. Cyprinus brama.—1832: Diplozoon (type).

paradoxum Carus, 1833a, 1835, 86, t. h. Succinea amphibia, Helix putris; Saxony.—
1835: Leucochloridium (type).

paradoxus Rud., 1810a, 257, t. h. Tetrodon mola; Europe.—1810: Schisturus (type).
paradoxus Crep., 1839a, 292, t. h. Perca lucioperca; Greifswald.—1839: Ancyrocephalus (type). 1878: Dactylogyrus. [1879: Tetraonchus unguiculatus.]

parallelum Looss, 19011, 622, t. h. Chelone mydas; Alexandria, from Egyptian coast.— 1901: Microscaphidium. 1902: Angiodictyum (type).

parasitica Jægers., 1897, 707, t. h. Raja clavata, R. lævis; Kattegat.—1897: Micro-pharynx (type) [ectoparasitic triclad].

paronæ Mont., 1893, 43, t. h. Seriola dumerilii.—1893: Dist.

parva Erc., 1881 or 1882, see Par., 1894, 164, t. h. Bythinia tentaculata; Bologna.— 1881?: Cerc.

[parva Sluiter, 1900, 6 (Dist.), a tunicate.]

parva Stoss., 1904, 10, t. h. Rana esculenta; Istria centrale.—1904: Brachymetra (type).

parvulum Staff., 1904, 494, t. h. Semotilus bullaris; Canada.—1904: Diplost.

parvulus Marshall & Gilbert, 1905, 478, t. h. Micropterus salmoides; near Madison, Wis.—1905: Cæcincola (type).

parvus Braun, 1901, 19, t. h. Testudo orbicularis; Vien. Mus.—1901: Telorchis. 1905: T. (Cercorchis).

parvus Looss, 1902i, 118, t. h. Sargus annularis, S. rondeletii; Triest.—1902: Monorchis.

pastinacæ Scott, 1904, 279, t. h. Trygon pastinaca; Scotland.—1904: Heterocotyle, type.

patagiatum Crep., 1846, 135, t. h. Ardea stellaris.—1846: Holost.

patellare Sturges, 1897, Sept., 57, t. h. Triturus (Molge) pyrrhogaster Boie.—1897;
 Dist. [1899: Phyllodist.] 1899: Spathidium. 1901: Phyllodist.

pectinata Huet, 1891a, 162, t. h. Donax anatinum.—1891: Cerc.

pectinatum Lint., 1905, 327, t. h. Trachinotus carolinus, Bairdiella chrysura; Beaufort. N. C.—1905: Dist.

pedata Schrank, (1796), 335, t. h. Anas querquedula.—1796: Festuc. [1905: Catatropis verrucosa.]

pedatum Looss, 1899, 592, t. h. Bagrus bayad, B. docmac; Cairo, Egypt.—1899: Glossidium (type).

pedatum Dies., 1850a, 309, t. h. Didelphis myosurus, D. cancrivorus; Brazil.—1850: Hemist.

pedatus Wagener, 1857, 99, t. h. Julis.—1857: Dactylogyrus. 1858: Diplectanum.

pedatus Dies., 1836c, 310, bellones 1794, renamed, t. h. Esox belone.—1836: Heteracanthus. [1794: Axine, type.]

pedicellatum Stoss., 1887, 184, t. h. Chrysophrys aurata; Triest.—1887: Dist. 1898: Podocotyle. [1899: Creadiinæ.]

pedocotyle Leidy, 1890, 282, t. h. Mola rotunda; New Jersey.—1890: Dist.

pegorchis Stoss., 1901, 94, t. h. Mæna smaris; Triest.—1901: Creadium. 1901: Allocreadium. 1904: Lepocreadium.

pelagiæ Kælliker, 1849, 53, t. h. Pelagia noctiluca; Naples.—1849: Dist. 1893: D. (Accacelium). 1900: Accacelium.

pelagica Mont., 1888, 45.—1888: Cerc.

pelagicum Staff., 1900, 399, at Passamaquoddy Bay, St. Andrews, New Brunswick.— 1900: Dist. 1902: Hemiurus (Apoblema).

pelamydis Tasch., 1878, 176, t. h. Pelamys sarda; Naples.—1878: Trist.

pelamydis Tasch., 1879, 612, t. h. Pelamys sarda; Naples.—1879: Didymozoon.

pellucidum Linst., 1873, 95, t. h. Gallus domesticus.—1873: Dist. 1890: Cephalogonimus. 1892: D. (Dicrocœlium). 1892: Mesogonimus. 1899: Prymnoprion. 1899: Prosthogonimus.

pellucidum Schlotthauber, 1860, 129, t. h. Petromyzon fluviatilis.—1860: Holost. pelophylacis esculenti Wedl, 1849, 197.—1849: Dist.

pendulum Looss, 1899b, 688, t. h. Recurvirostra avocetta; Egypt.—1899: Echinost. percæ Gmelin, 1790a, 3057, percæ cernuæ 1776 renamed; t. h. Perca cernua.—1790: Fasc. [1809: Dist. nodulosa.]

percæ Mont., 1888a, 7, 30.—1888: Tetracotyle.

percæ cernuæ Mueller, 1776, 224, t. h. Perca cernua.—1776: Fasc. [1809: D. nodulosum.] percæ fluviatilis Moul., 1856, 230.—1856: Tetracotyle. [1858: T. typica.]

percina Schrank, 1790, 123, t. h. Perca asper, P. vulgaris.—1790: Fasc. [1809: D. nodulosum.]

peregrinum Braun, 1900f, 389, t. h. Rhinolophus ferrum-equinum.—1900: Dist. 1900: Mesotretes (type).

pericardium Crep., 1849a, 78, D. helicis Leidy, renamed. 1849: Dist. [1858: Cercariæum vagans.]

perlatum Nord., 1832, 88, t. h. Cyprinus tinca.—1832: Dist. 1845: Dist. (Podocotyle). 1886: D. (Echinost.). 1899: Asymphylodora (type). 1903: Echinost.

perlatum var. exspinosum Hausmann, 1896, 390, t. h. Barbus fluviatilis; Switzerland.— 1896: Dist.

permixtus Braun, 1901, 943, t. h. Hirundo rustica; Coll. Vienna.—1901: Plagiorchis. 1904: Dist.

perniciosum Taylor, 1884, 53, see endemicum.—1884: Dist. [1907: Clonorchis endemicum.]

perpastum Braun, 1902b, 86, n. n. for singularis of Looss.—1902: Stomylotrema (type). perpusillus Looss, 1902, 134, t. h. Mugil chelo.—1902: Dicrogaster (type).

persicus Braun, 1901c, 334, t. h. Persian wolf.—1901: Cotylogonimus. 1902: Hetero-

personatum Poir., 1886, 11. host unknown; Gulf of Mexico.—1885: Dist.

perugiai Setti, 1898, 7 pp., t. h. Tetrapturus belone; Spezia.—1898: Trist. petalosum Lander, in Looss, 1902m, 454, t. h. Acipenser rubicundus.—1902: Dist. Bunodera]. 1904: Acrodactyla (type). See lintoni and cornuta.

petasatum Deslongchamps, 1824ee, 511, t. h. Hæmatopus ostralegus; Caen.—1824: Monost.

peteromyzi fluviatilis Dies., 1850a, 307, t. h. Peteromyzon fluviatilis.—1850: Diplost. 1892: Tylodelphys.

petiolatum Rail., 1900, 241, t. h. Garrulus glandarius.—1900: Dicrocælium.

petromyzonis fluviatilis Dies., 1858e, 316, for peteromyzi fluv. (Diplost.).—1858:

Tylodelphys.

petromyzontis Brown, 1899a, 489, t. h. Ammocœtes; Oxford, Eng.—1899: Tetracotyle.
phaneropsolus Stoss., 1902, 25, t. h. Totanus sp.; Yeddo, Japan.—1902: Hæmatotrephus.

phasiani galli Dies., 1855, 64.—1855: Dist.

philocholum Crep., 1845, 330, t. h. Delphinus phocæna, in liver.—1845: Dist.

philodryadum West, 1896, 322, t. h. Philodryas schottii; England.—1896: Dist. 1900: Opisthogonimus (type).

philomelæ Rud., 1819a, 120, t. h. Motacilla philomela; C. E. V.—1819: Dist. [1850:

D. macrostomum.]

phanicopteri Luehe, 1898g, 625, t. h. Flamingo; Berberei.—1898: Echinost.

phoxini Linst., 1896, 378, t. h. Phoxinus lævis.—1896: Dist.

phryganeæ Linst., 1877b, 185, t. h. Phryganea grandis.—1877: Dist.

phycidis Par. & Per., 1889, 743, t. h. Phycis blennoides; Genova.—1889: Dactylocotyle. 1890: Octobothrium. 1890: Dactycotyle. 1890: Octobothrium (Dactylocotyle).

physæ fontinalis Dies., 1855a, 400, t. h. Physa fontinalis; Regiomontii.—1855: Dist. 1855: Cercariæum.

physcon Luehe, 1901n, 478, t. h. Lophius piscatorius; Triest.—1901: Lecithochirium.
physophoræ Kælliker, 1849c, 53, t. h. Physophora tetrasticha; Naples.—1849: Dist.
[1850: D. geniculatum.]

pianæ Galli Valerio, 1898d, 145, t. h. Anas boschas; Busto Arsizio.—1898: Opistor-

chis. 1898: Opisthorchis.

picta Rud., 1802, 64, F. vespertilionis renamed; t. h. Fledermaus [Vespertilio murinus]; Greifswald.—1802: Fasc. [1809: Dist. lima.]

pictum Crep., 1837, 313, t. h. Ciconia alba.—1837: Dist. 1901: Stomylotrema.

pigmentata Sons., 1892, Oct. 7, 142, t. h. Physa alexandrina, P. micropleura; Cairo, Egypt.—1892: Cerc. [Amphist.]. [1902: A. conicum.]

pilchardei Mont., 1888a, 66, misprint for pilchardi (Octocotyle).

pilchardi Ben. & Hesse, 1863, 1864, 99, t. h. Clupea pilchardus.—1863: Octocotyle. 1879: Octobothrium. 1889: Octoplectanum.

pileata Rud., 1802, 65, t. h. Sterna hirundo; Greifswald.—1802: Festuc. 1803:
 Monost. 1809: M. (Monost.). 1819: Amphist. 1828: Holost. 1845: Holost.

pileatum Brand., 1888a, 59, t. h. Sterna caspica, Larus glaucus; Wien. Mus.—1888: Hemist.

pileatum Rud. of Bremser, 1824, pl. 8, figs. 28–29.—1824: Amphist. [1850: Hemist. commutatum.]

pingue Mehlis in Crep., 1846, 138, t. h. Colymbus cristatus.—1846: Monost. 1904: Renicola (type).

pinguicola Treutler, 1793, 6, t. h. Homo.—1793: Hexathyridium (type). 1800: Polyst. 1809: Polyst. (Hexast.). 1828: Hexathiridium. 1833: Hexacotyle. [Hexast.] [Linguatula.]

pinguiculum Joy, 1835a, 505, for pinguicola (Polyst.).

pini Ben. & Hesse, 1863, 1864, 72, t. h. Trigla pini.—1863: Placunella (type). 1878:
 Trist. 1903: Trochopus.

pınnarum Wagener, 1857, 26, t. h. Gasterosteus.—1857: Dist.

pinnatum Will.-Suhm, (1873), 341.—1873: Dist.

piriforme Crep., 1846, 142, in Anas fusca, A. glacialis; see pyriforme.—1846: Dist.

piriforme Sons., 1895, 184, for pyriforme (Amphist.).

piscicola Odhn., 1902, 152, t. h. Gymnarchus niloticus.—1902: Opisthorchis.

pittacium Braun, 1901, 947, t. h. Strepsilas interpres.—1901: Dist.

planci Stoss., 1899, 5, t. h. Ranzania truncata; Triest.—1899: Podocotyle. 1902: Orophocotyle (type).

planicolle Rud., 1819a, 682, t. h. Pelecanus sula; Brazil.—1819: Dist. (Echinost.). [1850: Monost. echinostomum.] [1901] 1902: Anoictost.

planorbis carinati Fil., 1857c, 13, t. h. Planorbis carinatus; Turin.—1857: Dist. 1858: Cerc. (Acanthocephala). 1858: Cercariæum.

planorbis cornei Dies., 1850a, 298, based on Henle, 1835, 597, t. h. Planorbis corneus.— 1850: Cerc. 1855: Cercariæum.

planorbis cornei Linst., 1877, 187.—1877: Dist.

planorbis cornei (hepatis) Dies., 1858d, 281, t. h. Planorbis corneus.—1858: Cercariæum.

planorbis cornei (ovariorum) Dies., 1858d, 281, for planorbis cornei 1850.—1858: Cercariæum.

platellæ Bosc, 1802a, v. 1, 273, see platessæ (Fasc.).

platessæ Mueller, 1784, 114, t. h. Pleuronectes platessa, int.—1784: Fasc. 1803: Dist. [1809: D. areolatum.]

platycephalum Crep., 1825a, 39, t. h. Colymbus rufogularis; Greifswald.—1825: Amphist. 1845: Holost.

platygaster Leuck., 1842a, 30, t. h. Gadus merlangus.—1842: Octobothrium. [1845: O. merlangi.]

platyura Crep., 1838b, 83, includes Axine belones, Heteracanthus pedatus, H. sagittatus.—1838: Axine.

platyura Leidy, 1891a, 416, free in pool with Lymnæus; Fort Bridger, Wyo.—1891: Cerc.

platyurum Much., 1896, 267, t. h. Harelda glacialis; East Prussia.—1896: Dist. 1899: Psilost. (type).

playfairii Forbes & Goodsir (1839).—1839: Tetrast.

plenus Staff., 1904, 484, t. h. Anarrhichas lupus; Canada.—1904: Derogenes. pleroticum Braun, 1899e, 631, host?; Brazil.—1899: Dist. 1901: Telorchis.

plesiostomum Linst., 1883, 305, t. h. Perdix græca; Turkestan.—1883: Dist. 1892: D. (Brachylaimus). 1899: Dicroccelium. [1899: Lyperosomum.]

pleurolophocerca Sons., 1892, Oct. 7, 138, t. h. Melania tubercolata, Cleopatra bulimoides; Cairo, Egypt.—1892: Cerc.

pleuronectes Mueller, 1773, 70, in aquis [etc.].—1773: Cerc. 1827: Phacus.

pleuronectis maximi Dies., 1855, 64, based on Bellingham, 1844, 428; renamed D. microcotyle. 1855: Dist.

plicatum Crep., 1829a, 878, t. h. Balæna rostrata; Rügen.—1827: Monost. 1891: Ogmogaster (type).

[plumbeum D. Valle, (Dist.), a tunicate.]

podomorphum Nitzsch, 1819, 399, t. h. Falco haliaëtos; Halle.—1819: Holost. 1850: Hemist.

podura Mueller, 1773, 66, in paludosis Lemna coopertis.—1773: Cerc. 1815: Furcocerca. 1827: Enchelys. 1828: Ichthydium, type.

poirieri Stoss., 1895, 227 (D. gelatinosum Poir., 1885).—1895: Dist. 1895: D. (Dicrocelium). 1899: Telorchis. 1902: Telorchis. 1905: T. (Cercorchis).

poirieri Giard & Billet, 1892, 615, t. h. cattle; Tonkin.—1892: Homalogaster.

pollachi Mont., 1888, 66, for pollachii (Dactycotyle).

pollachii Ben. & Hesse, 1863, 1864, 90, t. h. Merlangus pollachius.—1863: Udonella.

pollachii Ben. & Hesse, 1863; 1864, 110, t. h. Merlangus pollachius.—1863: Dactycotyle (type). 1879: Octobothrium. 1883: Dactylocotyle. 1890: O. (Dactylocotyle).

polonica Kowal., 1895g, 41, t. h. Anas boschas fera, A. crecca L.; Dublany.—1895: Bilharzia. 1898: Schistosoma. 1899: Bilharziella (type).

polonii Mol., 1859, 291, t. h. Caranx trachurus; Batavii.—1859: Dist. 1886: D. (Echinost.).

polychroa Tasch., 1879, 36.—1879: Planaria.

polyclinorum Pag., 1862, 306, t. h. Polyclinum; Golf von Spezia.—1862: Dist.

polymastos Leuck., 1880, or Lejtenyi, 1880, t. h. Equus caballus; Egypt.—1880: Gastrodiscus. [1896: G. ægyptiacus.]

polymorpha Rud., 1802b, 70, F. anguillæ Gmelin renamed; t. h. Aal.—1802: Fasc. 1809: Dist.

polymorphum Fil., 1837a, 337, t. h. Planorbis submarginata; see also brachyura Diesing, 1850.—1837: Dist.

polymorphus Baer, 1827b, 570, t. h. Unio, Anodonta.—1827: Bucephalus (type). 1855: Bucephalus (Eubucephalus, type). 1856: Cerc. ——: Cerc. (Bucephalus).

polyoon Linst., 1887, 103, t. h. Gallinula chloropus.—1887: Dist.

polyorchis Stoss., 1889, 24. t. h. Corvina nigra; Triest.—1889: Dist. 1894: D. (Polyorchis, type). 1896: Polyorchis. 1896: Pleorchis (type).

pomatiæ Vaney & Conté, 1899, 194, t. h. Helix pomatiæ; Lyon.—1899: Cerc.

pomatomi Goto, 1899a, 278, t. h. Pomatomus saltatrix; Newport, R. I.—1899: Microcotyle.

pontaliei Stoss., 1898, 178, for pontalliei (Dist.)

pontallici Cobbold, 1860a, 10, t. h. Ardea minuta; D. cladocalium Dies., renamed.— 1860: Dist.

porcorum Gmelin, 1790a, 3054.—1790: Fasc. hepatica.

poricola Schlotthauber, 1860, 129, t. h. Anas boschas.—1860: Ast. (type).

porrectum Braun, 1899b, 714, t. h. Saurophaga saurophaga; New Guinea.—1899: Dist. [1899: Lyperosomum.]

posticum Staff., 1905, Apr. 11, 692, t. h. Vespertilio subtilis Say; Canada; chilost. Mehlis of Staff., 1903, 827, renamed.—1905: Lecithodendrium.

poturzycensis Kowal., 1898g. 72, t. h. Anas boschas dom.—1898: Opisthorchis. 1898: O. simulans var. 1898: Campula. 1902: Metorchis.

præmorsum Nord., 1832, 55, t. h. Cyprinus brama; Europe.—1832: Monost. 1892: Monostomulum.

præteritus Looss, 1901e, 633, t. h. Labrax lupus; Triest.—1901: Acanthochasmus.
 prenanti St.-Remy, 1890, 41, t. h. Raja oxyrhynchus; Roscoff.—1890: Onchocotyle.
 1889: Rajonchocotyle.

pretiosus Ariola, 1902, 107, t. h. Thynnus vulgaris.—1902: Didymozoon.

prima Ssinitzin 1905, 147; 1906, 687, t. h. Aplexa hypnorum, Planorbis vortex compressa, Corethra, Ilybius; Warschau.—1905: Cerc.

primus Staff., 1905, 691, t. h. Eutenia sirtalis; Canada.—1905: Lechriorchis (type).
prismaticum Zed., 1800a, xvi. 150, t. h. Corvus frugilegus; Europe.—1800: Monost.
1809: M. (Monost.). [1892: Dist.]

pristiophori Johnston, 1902, 326, t. h. Pristiophorus cirratus Lath.—1902: Dist.

pris'is Deslongchamps, in Lamouroux, 1824, 563; 1821, 281, t. h. Merlangus communis;
 Normandy.—1824: Dist. 1845: D. (Echinost.). 1860: Echinost. 1899: Anoiktost. 1899: Stephanost. 1901: Stephanochasmus.

problematicum Stoss., 1902, 14, includes Looss, 1899b, 660, t. h. Totanus calidris, T. glottis.—[1899: Cyclocœlum.] 1902: Cyclocœlum.

proboscideum Rud., 1814a, 106, Echinorhynchus crotali, 1808, renamed; America.—
1814: Polyst. 1814: Polyst. (Pentast.). 1819: Pentast. [Porocephalus, type.]
producta Linst., 1903, 354, t. h. Solea vulgaris.—1903: Epibdella.

productus Staff., 1904, 485, t. h. Hemitripterus americanus; Canada.—1904: Sinis-

troporus. productus Odhn., 1902, 24, t. h. Nilkrokodil; Sudan.—1902: Acanthochasmus.

propinguum Braun, 1901, 942, t. h. Dendrocalaptes scandens; Brazil.—1901: Glaphyrost.

proteus Brand., 1891, 19, t. h. Chelonia viridis.—1891: Monost. 1892: Notocotyle. 1899: Baris (type). 1900: Deuterobaris (type).

proxima Lespés, 1857, 116, t. h. Littorina littorea.—1857: Cerc. 1858: C. (Gymnocephala).

prussica Mueh., 1896, 590, t. h. Anas glacialis.—1896: Cyathocotyle (type). pseudam phistomum Crep., 1846, 146, t. h. Chelonia mydas.—1846: Monost.

8588-No. 37-08-5

pseudoechinatum Olss., 1876, 21, t. h. Larus marinus; Scandinavia.—1876: Dist. 1892: Echinost.

pseudofelineus Ward, 1901, 180, "Dist. felineum" of Ward, 1895, not Rivolta, 1884, renamed, t. h. Felis catus dom.; Lincoln, Nebr.—1901: Opisthorchis.

pseudostoma Will.-Suhm, 1870, 11, t. h. Alligator lucius.—1870: Dist. 1886: Diplost. 1895: D. (Brachylaimus).

pudens Lint., 1900a, 269, t. h. Paralichthys dentatus; Woods Holl.—1900: Dist.

pugio Linst., 1887, 105, t. h. Lïmnæa ovata.—1887: Cerc.

pugnar LaValette, 1855, 19, t. h. Paludina vivipara.—1855: Cerc. [1855: C. microcotyla.]

pulchellum Rud., 1819a, 94, t. h. Labrus cynædus; Naples.—1819: Dist. 1902: Helicometra (type). 1903: D. (Dicrocœlium).

pulcherrima Weyenbergh, 1876, 167, t. h. Hypostomus plecostomus; Argentina.—1876: Amphist. 1878: Dist.

pulicis Linst., 1892, 333, t. h. Gammarus pulex.—1892: Dist.

pulmonale Linst., 1904, 678, t. h. Halicore australis; Australia.—1904: Opisthotremapulmonale Bælz, 1883, April, 236, t. h. Homo [see westermanii Kerbert, 1878].—1883: Dist. 1899: Dist. 1890: Mesogonimus. 1899: Paragonimus.

pulmonalis colubri natricis Viborg, 1795, 243.—1795: Dist. [D. naja].

pulmonar La Clínica de Málaga, 1883, 308, for pulmonale, see westermanii.—1883: Dist.

pulmonis Kiyona, Suga, and Yamagata, 1881, teste Ijima, 1889b, 148, see westermanii.—1881: Dist.

pulmonum Bælz, 1880, Sept., 721, t. h. Homo, see westermanii.—1880: Gregarina. 1884: Dist. [1899: Paragonimus.]

pulverulenta Braun, 1901g, 946, t. h. Anas querquedula; Dongola.—1901: Bilharziella. pulvinatum Braun, 1899, 630, t. h. Flussschildkræte; Brazil.—1899: Dist.

pumilio Looss, 1896, 154, t. h. Pelecanus onocrotalus; Egypt.—1896: Monost. 1899: Haplorchis (type).

punctata Mueller, 1774, 57, in pratis inundatis primovere.—1774: Fasc. 1787: Planaria.

punctatum Dies., 1850a, 329, misprint for punctum (Dist.).

punctum Zed., 1800a, 164, t. h. Cyprinus barbus; Europe.—1800: Dist. 1828: Fasc.
punctum Erc., 1881 or 1882, see Par., 1894, 164, t. h. Bythinia tentaculata: Bologna.—
1881: Cerc.

pungens Linst., 1894b, 333, t. h. Podiceps minor; Seeburger See.—1894: Dist. (Echinost.). 1899: Echinost.

pusilla Braun, D. M., 1790a, 62, t. h. Erinaceus europæus; Europe.—1790: Planaria. 1790: Fasc. 1803: Dist. 1892: Agamodist.

pusilla Looss, 1896b, 229, t. h. Vivipara unicolor Olivier; Damanhour.—1896: Cerc. pusilla Staff., 1904, 485, t. h. Anarrhichas lupus; Canada.—1904: Neophasis (type). pusillum Staff., 1904, 494, t. h. Stizostedion vitreum; Canada.—1904: Gasterost.

pusio Eichwald. ——: Planaria.

putorii Linst., 1877, 191, t. h. Fœtorius putorius.—1877: Diplost.

putorii Mol., 1858, 131, t. h. Mustela putorius; Patavia.—1858; Dist. 1892; Agamodist.
putorii Gmelin, 1790a, 3053, t. h. Mustela putorius.—1790; Fasc. 1790; Planaria.
1893; Dist. [1850; D. trigonocephalum.]

putrescens Looss, 1902, 135, t. h. Mugil auratus.—1902: Lecithobotrys (type).

pycnoporus Stoss., 1901, 92, t. h. Sargus salviani; Triest.—1901: Holorchis (type). pygmæa var. simile Jægers., 1900c, 737, t. h.—1900: Levinsenia. [1902: Spelotrema.]

pygmæa var. simile Jægers., 1900c, 737, t. h.—1900: Levinsenia. [1902: Spelotrema.] [1902: Dist.]

pygmæum Levin., 1881a, 73, t. h. Somateria mollisima; Egedesminde.—1881: Dist.
 1892: D. (Brachycœlium). 1899: Levinsenia. 1901: Spelotrema (type, Dec. 31).
 1903: Levinseniella.

pyramidum Looss, 1896b, 76, t. h. Rhinolopus hippocrepis; Ghizeh.—1896: Dist. 1899: Lecithodendrium.

pyriforme Dies., 1838a, 189, 1839a, 236, t. h. Tapirus americanus; Matogrosso or Cachoeira do Bananeira, Brazil.—1839: Amphist. 1901: Cladorchis.

pyriforme Crep., 1837a, 316, t. h. Eisente, Anas glacialis.—1837: Dist.

pyriforme Lint., 1900, 269, t. h. Palinurichthys perciformis; Woods Holl.—1900: Dist. pyridatum Bremser, in Rud., 1819a, 678, t. h. Crocodilus sclerops; Brazil.—1819: Dist.

pyrula Bory de St. Vincent, 1823a, 355, in infusion de chènevis.—1823: Cerc.

quadrangulata Bosc., 1802a, 257.—1802: Planaria.

quadrangularis Pallas, 1774, 20 in fossis; Hagæ Comitum.—1774: Fasc.

quadricornis Haswell, 1887a, 284, t. h. Astacopsis franklinii; Tasmania.—1887: Temnocephala.

quietum Staff., 1900, 403, t. h. "frog;" apparently Canada.—1900: Dist. 1905: Glypthelmins (type).

rachiæum Crep., 1839, 289, for rhachiæum (Diplost.).

rachidis Par., 1896, 2, for rhachidis.—1896: Diplost. (Tylodelphis).

rachieum Fraip., 1880c, 419, for rhachiæum (Diplost.).

rachineum Mueller, (1842).—Diplost.

rachion Cobbold, 1858b, 158, t. h. Morrhua æglefinus.—1858: Dist. 1886: D.(Echinost.). 1904: Lepidapedon (type). 1905: Lepodora (type).

rachiæa Odhn., 1905, 328, for rachion (Dist.).—1905: Lepodora (type).

radiata Mueller, 1774, 66, in aquis sylvestribus, rara.—1774: Fasc. 1776: Planaria. rachidis Hanover, 1864a, 3, see rhachidis 1850 (Tylodelphys).

radiatum Duj., 1845a, 427 t. h. Carbo cormoranus or Pelecanus carbo.—1845: Dist. (Echinost.).

radula Duj., 1845a, 433, t. h. Lymnæus palustris; Rennes.—1845: Dist. (Echinost.). ragazzi Linst., 1903t, 354, for ragazzii (Dist.).

ragazzii Setti, 1897, 8, t. h. Squalus sp.; Eritrea.—1897: Dist. (Polyorchis). 1899: Syncœlium (type).

rajæ Rud., 1809a, 435, for rajæ intestinale.—1809: Dist.

rajæ intestinale Viborg, 1795, 242.—1795: Dist.

ralli Rud., 1819a, 120, t. h. Rallus aquaticus.; M. C. V.—1819: Dist. [1850: D. holostomum.]

ramlianum Looss, 1896b, 36, t. h. caméléon; Ramleh.—1896: Dist. 1899: Lepoderma (type). 1899: Plagiorchis.

ramosum Sons., 1895, 123, t. h. Babulcus ibis: Nile Delta.—1895: Echinost.

ranæ Gmelin, 1790a, 3055, t. h. Rana, int.; Europe.—1790: Fasc. [1809: Amphist. subclavatum.]

ranæ Frælich, 1791, 69, t. h. Rana temporaria; Europe.—1791: Fasc. [1809: Amphist. subclavatum.] [1850: Dist. clavigerum.] [1894: D. endolobum.]

ranæ Zed., 1800a, xviii, 203, t. h. Rana temporaria; includes Planaria uncinulata 1790, from Rana esculenta; Europe.—1800: Polyst. (type). [1809: P. integerrimum.]

ranæ Valentin, 1843, 90 teste Dies., 1850a, 472.—Trematodum.

ranæ esculentæ Dies., 1850a, 388, based on Valentin, 1841, 54.—1850: Dist.

rarum Ben., 1858a, 1861a, 178, t. h. Cyprinus dobula.—1858: Dist.

rarus Braun, 1901, 17, t. h. Fulica atra, Coll. Berlin; Anas boschas.—1901: Prosthogonimus.

rastellus Olss., 1876, 16, t. h. Rana temporaria.—1876: Dist. [1887: D. endolobum.] rathonisi Šimon, 1896, 182, 192, misprint for rathouisi (Dist.).

rathonisii Huber, 1894, 2, misprint for rathouisi (Dist.).

rathouisi Poir., 1887, 203, t. h. Homo; Asia.—1887: Dist. 1892: D. (Dicrocœlium). 1903: Fasciolopsis.

rathouisii Braun, 1892a, 568, for rathouisi (Dist.).

raynerianum Nardo, 1827, 68, t. h. Luvarus imperialis; Venice.—1827: Dist. 1886: D. (Apoblema). 1901: Accacælium.

receptaculum Cobbold, 1860a, 29, t. h. Labrax lupus.—1860: Dist.

recurvatum Linst., 1873, 105, t. h. Anas marila.—1873: Dist. 1892: Echinost.

recurrum Duj., 1845a, 410, t. h. Mus sylvaticus; Rennes.—1845: Dist. (Brachylaimus).

redactum Nicoll, 1906, 515, t. h. Gasterosteus aculeatus; Scotland.—1906: Psilost. [1907: Podocotyle atomon.]

reductum Looss, 1901, 562, t. h. Thalassochelys corticata.—1901: Enodiotrema.

refertum Mueh., 1898, 18, t. h. Cypselus apus; East Prussia.—1898: Dist. 1899: Dicroccelium.

reficiens Braun, 1901, 945, t. h. Falco nitidus.—1901: Dicrocœlium.

[reflexum: Schistosoma, a term in teratology.]

reflexum Crep., 1825a, 54, t. h. Cyclopterus lumpus.—1825: Dist. 1886: D. (Dicrocelium). 1905: Podocotyle.

reinhardi Linst., 1903, 208, t. h. Astacus leptodactylus.—1903: Dist.

renale Chiaje, 1833, 13, t. h. Homo; Naples.—1833: Tetrast. (type).

renale Fil., 1855b, 19, t. h. Helix adspersa; near Turin.—1855; Dist. 1856; Cerc. 1858; C. (Gymnocephala).

renicapite Leidy, 1856, 43, t. h. Sphargis coriacea; America.—1856: Monost.

reniferum Looss, 1898, 461, unicum Looss, not Molin, renamed.—1898: Dist. 1899: Astia (type). 1904: Astiotrema (type).

reniforme Luehe, 1899, 531, for reniferum Looss (Dist).

reniformis Ariola, 1902, 101, t. h. Thynnus vulgaris; Naples.—1902: Didymocystis (type).

repandum Rud., 1819a, 681, t. h. Rana sp.; Brazil.—1819: Dist.

resectus Looss, 1902m, 537, t. h. Chelone mydas; Egyptian coast.—1902: Cricocephalus. reticulare Ben., 1859, 84, t. h. Chelonia midas.—1859: Monost. 1899: Microscapha (type). 1901: Microscaphidium (type).

reticulata Goto, 1894a, 189, t. h. Stromateus argenteus; Japan.—1894: Microcotyle. reticulatum Looss, 1885, 427, t. h. Wels; Costa Rica.—1885: Dist. 1899: Clinost. [1888: Mesogonimus (type).]

reticulatum Wright, 1879, 58, t. h. Ceryle alcyon.—1879: Dist. 1899: Fasc. 1892: D. (Brachylaimus.)

reticulatum Poir., 1886, 39, in Axinurus dugesii.—1886: Dist. [1899: Clinost. marginatum.]

retikulatum Looss, 1885b, 59, for reticulatum (Dist.).

retroconstrictum Srámek, 1901, 95, 108, fig. 62, t. h. Abramis brama.—1902: Dist.
retroflexum Mol., 1859, 290, t. h. Belone acus; Batavii.—1859: Dist. 1898: Podocotyle. 1886: D. (Podocotyle).

rctusum Duj., 1845a, 405, t. h. Rana temporaria; Rennes.—1845: Dist. (Brachycœ-lium). 1886: D. (Dicrocœlium).

revoluta Freelich, 1802, 58, t. h. Anas boschas; Europe.—1802: Fasc. 1899: Echinost. rhachiwum Henle, 1833a, 19, t. h. Rana.—1833: Diplost. 1892: Tylodelphys.

rhachidis Dies., 1850a, 305, rhachiæum 1833, renamed.—1850: Tylodelphys. 1896: Diplost. (Tylodelphis).

rhatonisii Simon, 1897, 223, misprint for rathouisii (Dist.).

rhizophisæ Mont., 1888a, 199, for rhizophysæ (Dist.).

rhizophysæ Studer, 1878, 12, t. h. Rhizophysa conifera.—1878: Dist.

[rhodopyge Sluiter, 1898 (Dist.), a tunicate.]

rhombi Ben., 1870, 72, t. h. Rhombus maximus.—1870: Dist.

rhombi Ben. & Hesse, 1863, 1864, 73, t. h. Rhombus maximus.—1863: Placunella. 1878: Trist. 1903: Trachopus.

rhombi lævis Dies., 1858e, 328, t. h. Rhombus lævis; Europe.—1855: Monost. 1892: Monostomulum.

rhopaloides Crep., 1839, 294, for ropaloides 1836, q. v.—1839: Amphist.

ricchiardii see richiardii (Dist.).—1902: Probolitrema (type).

richardii Brand., 1891b, 267, for richiardii (Dist.).

richiardi Mont., 1891, 500, for richiardii.—1891: Dist. (Polyorchis).

richiardii Lopez, 1888a, 137, t. h. Acanthias vulgaris.—1888: Dist. 1900: Anaporrhutum. 1902: Probolitrema (type).

richiardii Sons., 1890, 172, t. h. Pagrus orphus.—1890: Anoplodiscus (type).

rigens Linst., 1878a, 282, for ligula 1871 (Dist.).

rigonocerca Braun, 1892a, 809,? for trigonocerca.—1892: Cerc.

ringens Rud., 1819a, 101, t. h. Picus tridactylus; Vien. Mus.—1819: Dist. [1850: D. macrostomum.]

ringens Linst., 1878a, 360, for ligula 1871 (Dist.), t. h. Scymnodon ringens.

ringens Lint., 1905, 327, t. h. Micropogon undulatus, Trachinotus carolinus; Beaufort, N. C.—1905: Aspidogaster.

ringeri Cobbold, 1880, Aug., 139, t. h. Homo.—1880: Dist. 1890: Mesogonimus. 1897: Distomi. [See westermanii.]

ringerii for ringeri (Dist.).

ringers Rev. Sci., 1890, v. 46, 189, 190, for ringeri (Dist.).

robustum Lorenz, 1881, 583, t. h. Elephas africanus.—1881: Dist. 1892: D. (Brachylaimus).

robustum Stoss., 1902, 18, t. h. Fuligula cristata.—1902: Cyclocælum.

robustus Looss, 1901l, 621, t. h. Chelone mydas; Egyptian coast.—1901: Charaxicephalus (type).

rochebruni Poir., 1886, 36, t. h. Delphinus delphis.—1886: Dist. 1892: Cladocœlium. 1899: Brachycladium.

rochebrunni Braun, 1892a, 673, for rochebruni (Dist.).

rombi-lævis Mont., 1892, 717, for rhombi-lævis (Monost.).

ropaloides Leblond, 1836e, 290, t. h. Muræna conger; coast of Normandy.—1836: Amphist. [1850: Tetrabothriorhynchus migratorius.]

rosaceum Nord., 1832, 82, t. h. Gadus lota.—1832: Dist. 1856: D. [tereticolle].

rosarum Cobbold, 1860a, 21, misprint for rosaceum (Dist.).

rosea Mueller, 1774, 58, in sinu Dræbachiensi.—1774: Fasc. 1787: Plan. 1892: Amphiporus.

rosea Ben., 1870, 1871a, 90, t. h. Petromyzon omalii; Belgium.—1870: Dist.

rossittensis Mueh., 1898, 16, t. h. Turdus pilaris; East Prussia.—1898: Urogonimus 1899: Urotocus (type).

[rostellatum Dies., 1850a, 603.—1850: Octobothrium, cestode.]

rostrata Mueller, 1774, 65, in paludosis, primo vere annorum.—1774: Fasc. 1776: Plan.

rostrata Erc., 1881, see Par., 1894, 190, ?t. h. Bythinia tentaculata; Bologna.—1881: Cerc.

rostroaculeata Erc., 1881, see Par., 1894, 164, ?t. h. Bythinia tentaculata; Bologna.—1881: Cerc. [1894: C. nodulosa.]

rotundatum Linst., 1877, 187, t. h. Lanius collurio.—1877: Holost.

rotundum Goto, 1894a, 245, t. h. Xiphias gladius; Japan.—1894: Trist.

rubellum Olss., 1868, 40, t. h. Labrus maculatus.—1868: Dist. 1886: D. (Brachycelium). 1899: Lecithodendrium. 1902: Zoogonus.

rubens Duj., 1845a, 411, t. h. Sorex fodiens, S. tetragonurus; Rennes.—1845: Dist. (Brachylaimus).

ruber Luehe, 1900, 507, t. h. Trigla lineata.—1900: Derogenes (type).

ruber see also rubrum (Cricocephalus).

rubra Mueller, 1774, 59, Fucorum frequens.—1774: Fasc. 1776: Plan.

rubrum Kuhl & van Hasselt, 1822a, 1824a, 311, t. h. Chelonia midas; Iles des Cocotiers.—1824: Monost. 1902: Cricocephalus.

[rubrum Savigny (Dist.), a tunicate.]

rude Dies., 1850a, 360, t. h. Lutra brasiliensis; Brazil.—1850: Dist. [1899: Paragonimus.] 1900: Paragonimus.

rudectum Braun, 1901, 946, t. h. Ibis cœrulescens; Brazil.—1901: Lyperosomum.

rudolphianum Dies., 1850a, 429, t. h. Orthagoriscus mola; new name for T. coccineum of Rud., Phylline coccinea Schweigger, Capsala sanguinea Blainville, T. mola Bl.—1850: Trist. 1865: Capsala.

rufoviride Rud., 1819a, 110, t. h. Muraena conger; Naples.—1819: Dist. 1845: D. (Apoblema). 1899: Hemiurus. 1889: Apoblema. 1901: Lecithochirium (type).

rugosus Odhn., 1902, 32, t. h. Coluber pullatus.—1902: Cotylotretus (type).

rutilans Bosc, 1802a, 258.—1802: Plan.

saccata Goeze, 1782a, 221, based on Sack-Egel of Merrem, 1781, 169, pl. 1, figs. 3–7.—1781: Fasc. [1803: Cysticercus tæniæformis.]

sacidiornicola Stoss., 1902, 34, for sarcidiornicola (Monost.).

saginatum Ratz, 1898, 73, t. h. Ardea alba: Europe.—1898: Dist. 1903: Pegosomum

sagitata Lespés, 1857, 114, t. h. Nassa reticulata; Arcachon.—1857; Cerc. [C. (Gvm-

sagitta Looss, 1899b. 668, t. h. Chelonia mydas: Egypt.—1899: Microscapha. 1901: Microscaphidium. 1902: Octangium (type).

sagitta Pag., 1862, 297, for sagitata (Cerc.).

sagittalis Vaney & Conté. 1899, 196, C. sagittifera Sieb.. 1854, renamed. t. h. Helix pomatia.—1899: Cerc.

sagittata Dies., 1858d, 249, for sagitata 1857.—1858: Cerc. (Gymnocephala).

sagittatum Leuck., 1842, 49. t. h. Salmo fario.—1842: Octobothrium. cocotyle type. 1858: Placoplectanum type.

sagittatus Dies., 1836, 313, t. h. Esox belone: Europe. — 1836: Heteracanthus. [1850: Axine bellones.]

sagittifera Sieb., 1854. 18. t. h. Helix pomatia.—1854: Cerc. [1858: Cercariæum helicis pomatiæ.]

salamandræ Frœlich, 1789, 119, t. h. Salamandra atra.—1789; Fasc. 1803; Dist. [1809; D. crassicolle.]

salamandrinæ perspicillatæ Sons., 1896, 1: 1896, 116.—1896: Dist.

salebrosum Braun. 1901g. 946. t. h. Cypselus melba: Coll. Vien.—1901: Lypercsomum. salmonis Mueller, 1780 of Goze, 1782a, 173.—Fasc. [1901; ? Hemiurus crenatus Rud. Luehe.]

salmonis lavareti Linst.. 1878a, 266.—Trematodum.

salpæ Par. & Perugia. 1890. 207, t. h. Box salpa: Genova.—1890: Microcotyle.

sanguicola delle Chiaje, 1833, 14, venarum renamed, t. h. Homo.—1833: Polyst.

sanguinea Blainv.. see Dies.. 1850a, 429.—Capsala. [1850: Trist. rudolphianum.] sanguineum delle Chiaje, 1837b, 245, t. h. Homo.—1837: Polyst.

sanquineum Sons., 1894, 111. t. h. Camæleo vulgaris: Gabes in Tunisia.—1894: Dist. 1895: D. Brachylaimus . 1899: Anchitrema type .

sarcidiornicola Mégnin, 1890c, 87, t. h. Sarcidiornis melanota: Madagascar.—1890: Monost. 1902: Typhlocolum.

sargi and sargii Par. & Per., 1890, 4, t. h. Sargus rondeletii; Genova.—1890; Microcotyle.

sauromates Poir., 1886, 24, t. h. Elaphis sauromates.—1886; Dist. 1895; D. Dicrocœlium). 1904: Plagiorchis.

sauromatis Braun, 1893a, 876, for sauromates Dist.

savakinensis Looss, 1899b, 672, t. h. Larus sp.: Sawakin, Egypt.—1899; Stictodora type .

scaber Rud., 1819a, of Odhn., 1905, 353.—1905: Dist. [1905: Stephanochasmus.]

scabra Mueller, 1788, 31, t. h. Gadus barbatus.—1788: Fasc. 1893: Dist. 1889: Apoblema. 1899: D. Hemiurus scabrum.

scabridum Braun, 1900, 390, t. h. Noctilio macropus, Molossus, Phyllostoma; Brazil.— 1900: Urotrema (type).

scabrum of Rud., 1819a, 424.—1819; Dist. Echinost., 1860; Echinost, 1899; Anoiktost. [1899: Stephanost.] [1905: Stephanochasmus.]

schistocotyle Fischder.. 1901, 373, t. h. Dicotyles torquatus: Brazil.—1901: Cladorchis Taxorchis, type ..

schistolotyle Fischder. 1903f, 607, for schistocotyle (Cladorchis).

schlosseri Graff. 1904. 457, t. h. Botryllus schlosseri.—1904: Plan.

sciænæ Ben. & Hesse, 1863, 123. t. h. Sciæna aquila.—1863: Diplectanum.

sciana P. J. Ben., 1856, 502, t. h. Sciana aquila.—1856: Epibdella, 1891: Phylline, 1903: E. Benedenia). [1878: Trist.] [1858: Benedenia elegans, type.]

sciænæ Goto, 1894a. 194. t. h. Sciæna scina; Japan.—1894: Microcotyle.

sciænæ Ben. & Hesse, 1863, 1864, 93, t. h. Sciæna aquila.—1863: Udonella.

scianæ Tasch., 1878. 568, for sciænæ Ben., 1856.—1878: Trist.

scimna Risso, 1826, 262. t. h. Echinorhinus spinosus.—1826: Dist. [1850: D. insigne.]

sciana Ben. & Hesse, 1864, 123, for sciana (Diplectanum).

sciana Ben. & Hesse, 1864, 69, for sciana Epibdella).

scleroporum Crep., 1844a, 112, t. h. Chelonia mydas; Vratislaviæ.—1844: Amphist.

scoliocalium Fischder., 1904, 459, t.h. Buffelus indicus in Cochinchina and Annam; Bos taurus in Annam.—1904: Paramphist.

scombri Kuhn., 1829b, 361, t. h. Scomber scomber.—1829: Octost. 1832: Octobothrium. 1864: Octocotyle.

scombri Tasch., 1879, 612, t. h. Scomber colias; Naples.—1879: Didymozoon.

scombri of Grube, 1855a, 137, t. h. Scomber scombrus.—1855: Octobothrium. 1855: Tetracotyle. [1858: Grubea cochlear, type.] 1859: Pleurocotyle (type). [1859: Pleurocotylus, type.] 1864: Grubea.

scombri pelamidis Tilesius in Rud., 1809a, 437.—1809: Fasc. [1809: Dist. clavatum.] scombrinum Linst., 1889, 80, for sobrinum (Dist.).

scorpænæ Rud., 1819a, 122, t. h. Scorpæna scrofa: C. E. V.—1819: Dist. 1886: D. (Dicrocœlium). 1899: Dicrocœlium. [1899: Creadiinæ.]

scorpii Mueller, 1776, 223, t. h. Cottus scorpius.—1776: Fasc. 1803: Dist. [1850: D. granulum.]

scymna Villot, 1878, 3, for scimna (Dist.).

scymni Mont., 1893, 52, for scimna (Dist.).

scymni ainosi Dies., 1858e, 371, t. h. Scymnus ainosi.—[1857: Polyst. sp.] 1858: Polyst. 1858: Onchocotyle.

scyphocephalum Braun, 1899, 630, t. h. Testudo matemata: Brazil.—1899: Dist. 1901: Acanthochasmus.

sebastis Goto, 1894a, 187, t. h. Sebastes sp.; Japan.—1894: Microcotyle.

secunda Ssinitzin, 1905, 153; 1906, 687. t. h. Corethra, Ephemera; Warschau.—1905;

secundum Nicoll, 1906, 514, t. h. Hæmatopus ostralegus, Larus argentatus, L. ridibundus.—1906: Echinost.

secundus Looss, 1907, 134, t. h. mule; Assam.—1907: Gastrodiscus.

segmentatum Mueller, 1894, July, 113.—1894: Dist. (Echinost.).

semiarmatum Mol., 1858, 131, t. h. Acipenser naccari; Patavii.—1858: Dist. 1886: D. (Echinost.).

semiflavum Linst., 1880, 50, t. h. Petromyzon fluviatilis.—1880: Dist.

semifuscum Olss., 1876, 28, t. h. Sula bassana; Lund.—1876: Monost. semisquamosum Braun, 1900, 228, t. h. Vesperugo noctula.—1900: Dist. [1907: Parabascus.]

semperi Weber, 1889.—Temnocephala.

semperii Mont., 1891, 129, for semperi (Temnocephala).

sepiolæ delle Chiaje, —.—: Monost.

seriale Rud., 1808a, 351; 1809a, 368. t. h. Salmo alpinus: Greenland.—1808: Dist.

serialis Looss, 1901, 620, t. h. Thalassochelys corticata; Alexandria, from Egyptian coast.—1901: Adenogaster (type).

serotinus Staff., 1904, 493, t. h. Moxostoma macrolepidotum Le S.; Canada.—1904: Plagioporus (type).

serpens Nitzsch, in Rud., 1819a, 88, t. h. Falco haliaëtos; Halle.—1819: Amphist. 1819: Holost.

serpentatum Mol. 1859, 830, t. h. Sayris camperi; Batavii.—1859; Dist. 1896; D. (Brachylaimus).

serpentulum Carus, 1884, 131, for serpentatum (Dist.).

serrani Mont., 1889, 322, t. h. Serranus fimbriatus, at Madeira, and S. gigas at Naples.— 1889: Didymozoon.

serrata Freelich, 1789, 148 [an Arachnoid], t. h. hares.—1789: Linguatula (type). 1800: Polyst. 1809: Polyst. (Pentast.). 1819: Pentast.

scrrata Looss, 1899b, 592, t. h. Varanus niloticus; Zool. Inst. Leipzig.—1899: Styphlodora (type).

serratum Dies., 1850a, 385, t. h. Aranus scolopaceus: Brazil.—1850: Dist. 1860: Echinost.

serrulata Mueller, 1776.—Fasc. [1901: ? Hemiurus crenatus.]

sessilis Odhn., 1902, 27, t. h. Nilkrokodil.—1902: Nephrocephalus (type).

sessilis Goto, 1894a, 212, t. h. Chœrops japonicus: Mitsugahama (Prov. Iyo, Japan).— 1894: Diclidophora. 1895: Cyclobothrium (type).

setifera O. F. Mueller, 1786, 127, in aqua marina raro.—1786: Cerc. [1827: Trichoda.] setifera Moul., 1856a, 214 and Dies, 1858d, 250 based on J. Mueller, 1850, 497, free in water; Marseille or Triest.—1856: Cerc. 1858: C. Gymnocephala).

setifera Mueller of Villot, 1879 (Cerc.), in Scrobicularia tenuis. See villoti (Cerc.).

setigera Giard, 1897c, 955, for setifera, 1850.—1897: Cerc.

setosicauda Dadai. 1888f. 84.—1888: Histrionella.

setosicaudata Mont., 1888, 77, for setosicauda (Histrionella).

settenii Numan, 1640, 358, t. h. Equus caballus.—1840; Monost. 1850; Pentast. [1891; Oestrid larva.] 1892; Monostomulum.

sialidis Linst., 1892, 334, t. h. Sialis lutaria.—1892; Dist.

siamense Poir., 1886, 327, t. h. Crocodilus siamensis.—1886: Diplost.

sibiricum Winnogradow, 1892, 116, t. h. Homo; Siberia.—1892: Dist. [Opisthorchis

siemersii Buettel-Reepen, 1900, 589, t. h. Sphyræna barracuda; Atlantic.—1900: Dist.

[sigmoidea (Nitzschia), a diatom.]

sigmoideus Looss, 1899b, 608, t. h. Passer dom. at Alexandria, Caprimulgus europæus, at Cairo.—1899: Phaneropsolus type).

signatum Duj., 1845a, 415. t. h. Coluber natrix; Rennes.—1845: Dist. (Brachylaimus). signatum Duj., of Erc., 1881e, 73, in Tropidonotus natrix.—1881: Dist. 1899: Telor-chis. [1893: D. ercolanii.]

siluri Wagener, 1857, see Braun, 1890a, 544.—Dactylogyrus.

siluri glanidis Wagener, 1857, see Dies., 1858e, 379.—Dactylogyrus.

simile Sons., 1890, 105, t. h. Python molurus.—1890: Dist.

simile Jægers., 1900, 737, see pygmæa similis.—[1900: Levinsenia pygmæa var.] 1902: Spelotrema.

simile Looss, 1899b, 601, see similigenus.—1899; Dist. 1899; Hæmatolæchus. Pneumonœces.

similigenus Stiles & Hass., 1902d, 20, for simile (Dist.) Looss, (Hæmatolæchus).

similiplexus Staff., 1902, 901, t. h. American frogs and toads.—1902: Hæmatolæchus, 1905: Pneumonœces.

similis Stoss., 1902, 24, t. h. Himantopus atropterus: Egypt.—1902: Hæmatotrephus. simillimum Mueh., 1898, 18, t. h. Fuligula nyroca.—1898: Dist. 1899: Psilost. simplex Odhn., 1900, 62, t. h. Pleuronectes limanda: Kristineberg, Sweden.—1900:

Aporocotyle type.

simplex Rud., 1809a, 370 æglefini Mueller, 1776, renamed , t. h. Gadus æglefinus.-1809: Dist. 1886: D. (Dicroccelium). 1904: Sinistroporus type).

simplex Polonio, 1859, teste Par., 1894, 147, t. h. Lacerta muralis; Padua.—1859; Dist. simplex Looss, 1899b, 606,607, D. cygnoides var. B. of Bensley, 1892, renamed.—1899: Gorgodera. 1902: Gorgoderina type.

simplex Johnston, 1904, 112, t. h. Ardea novæ hollandiæ Lath.—1904: Holost.

simplissima Curtis. 1900a, 447.—1900: Planaria.

simulans Looss, 1896b, 52, t. h. Pernis apivorus; Alexandria, Egypt.—1896: Dist. 1898: Opisthorchis.

simulans poturzycensis Kowal.. 1898g, 71 or 1898h. 135, t. h. Anas boschas dom.—1898: Opisthorchis. 1898: Campula.

sinense Cobbold, 1875i, 423, t. h. Homo: China.—1875: Dist. 1895: Opisthorchis, 1896: Dicrocœlium. 1907: Clonorchis (type).

sinese Mosler & Peiper, 1894, 177 for sinense Dist.

sineuse MacConnell, 1876, 343, misprint for sinense Dist.).

singulare Mol., 1859. 288, t. h. Ibis falcinellus; Patavii.—1859: Dist. 1899: of Looss. Stomylus (type). 1892: D. (Dicrocœlium). [1900: of Looss. Stomylotrema, type.]

singulare Keferstein, 1862a, 131, t. h. Capitella rubicunda; St. Vaast.—1862: Loxosoma (type).

singularis Stoss., 1902. 29. t. h. Gallinula pusilla; loc. not given.—1902: Ophthalmophagus.

singularis Mol. of Looss, 1899b, renamed perpastum Braun.—1899: Stomylus (type). [1900: Stomylotrema, type.]

sinuatum Goto, 1894a, 239, t. h. Histiophorus sp.; Japan.—1894: Trist.

sinuatum Rud., 1819a, 97, t. h. Ophidium imberbe; Naples.—1819: Dist. 1901: Allocreadium. 1902: Helicometra.

sinuatuun Mont., 1899, 109, misprint for sinuatuum 1894 (Trist.).

siredonis Poir., 1886, 32, t. h. Siredon mexicanus, int.—1886: Dist. 1899: Opisthioglyphe.

sirenis Braun, 1893a, 870, for sirenis lacertinæ (Dist.).

sirenis lacertinæ Vaillant, 1863, 348, t. h. Siren lacertina.—1863: Dist.

sluiteri Brock, 1886, 543, t. h. Diacope metallicus; Java.—1886: Eurycœlum (type). 1892: Dist. 1892: Apoblema. 1899: Hemiurus.

smaris Ijima, in Goto, 1894a, 207, t. h. Smaris vulgaris (on caudal segment of a Cymothoa); Gulf of Naples.—1894: Octobothrium. 1894: Diclidophora.

sobrinum Levin., 1881a, 70, t. h. Cottus scorpius; Egedesminde.—1881: Dist. 1886: D. (Echinost.). 1899: Stephanost. 1899: Anoiktost. 1904: Stephanochasmus.

soccus Mol., 1858, 129, t. h. Mustelus plebejus; Patavii.—1858: Dist. 1886: D. (Brachylaimus). [1899: D. megastomum.]

sociale Luehe, 1901, 171, t. h. Bufo melanostictus Schneider; India.—1901: Dist.

solex Ben. & Hesse, 1863; 1864, 70, t. h. Solea vulgaris.—1863: Phyllonella (type). 1890: Epibdella. 1903: Phylline. 1879: Trist. (Phylonella). 1878: Trist.

solex Duj., 1845a, 417, t. h. Pleuronectes solea; Rennes.—1845: Dist. (Brachylaimus). solexforme Rud., 1809a, 384, t. h. Trigla gurnardus; for D. triglæ gurnardi.—1809 Dist. 1828: Fasc.

soleare Braun, 1899e, 629, t. h. Testudo midas.—1899: Dist. 1901: Cymatocarpus. solidus Looss, 1901l, 619, t. h. Chelone mydas; Egypt.—1901: Glyphicephalus (type). solitaria Looss, 1899b, 592, t. h. Thalassochelys corticata; Abukir.—1899: Styphlodora. 1903: Renifer.

solivagus Odhn., 1902, 29, t. h. Clemmys caspica.—1902: Telorchis.

solæiformis Blainv., 1828, 585, for soleæforme Rud., 1809a, 384.—1828: Fasc.

somateriæ Levin., 1881, 71, t. h. Somateria mollissima; Egedesminde.—1881: Dist.
 1892: D. (Brachyccelium). 1899: Lecithodendrium. 1900: Gymnophallus. 1902: Levcithodendrium. 1904: Leucithodendrium.

sonsinoi Giard, 1880a, lxviii.—Linst., 1889, 23, for sonsinoii 1877 (Gastrodiscus).—1889: Amphist.

sonsinoii Cobbold, 1877e, Apr., 233, t. h. Equus; Egypt.—1877: Gastrodiscus (type). 1877: Aspidocotylus. [1889: Amphist. sonsinoi.] [1896: Gastrodiscus ægyptiacus.]

sonsinoni Poir., 1883, 74, for sonsinoii (Gastrodiscus).

sonsinonis Cobbold, 1879b, 359, for sonsinoii (Gastrodiscus).

sophiæ Stoss., 1886, 44, t. h. Pagellus mormyrus; Triest.—1886: Dist. 1886: D. (Echinost.). [1899: Creadiinæ, ?Creadium.]

sorbens Braun, 1899g, 490, t. h. Tantalus loculator; Brazil.—1899: Clinost.

soricis Dies., 1858e, 354, t. h. Sorex araneus.—1858: Dist. [1855: D. (Dicrocœlium).] soricis Linst., 1877, 191, t. h. Sorex vulgaris.—1877: Tetracotyle.

soricis aranei Dies., 1855, 64; see soricis.—1855: Dist. (Dicroccelium).

spari Rud., 1819a, 122, t. h. Sparus erythrinus, S. smaris; C. E. V.—1819: Dist.

spataceum Stoss., 1896, 126, for spathaceum (Diplost.).

spathaceum Rud., 1819a, 109, t. h. Larus glaucus; Mus. Vien.—1819: Dist. 1845: Holost. 1850: Hemist. 1876: Diplost. 1898: Conchosomum.

spathula Brand., 1888a, 54, t. h. Falco palumbarius; Vienna Museum.—1888: Diplost.

1888: Hemist. spathula Crep., 1829, 50, t. h. Falco buteo, F. nisus, F. lagopus.—1829: Holost. 1850: Hemist. [1898: Conchosoma.]

spathulæforme Brand., 1888a, 44, t. h. Strix otus.—1888: Diplost.

spathulatum Leuck., 1876, 871, not Crep., 1849, t. h. Homo; for spatulatum Rud., 1819.—1876: Dist. [Opisthorchis sinensis.]

spathulatum Rud. of Crep., 1837, 310, for spatulatum.—1837: Dist. 1898: Echinost. 1902: D. (Echinost.). 1902: Sodalis (type).

spathulatum Dies., 1859c, 426, for spatulatum (Monost.).

spathulatum hepatis Simmonds, 1901, 110, see sinense (Dist.).

spatiosus Brand., 1898a, 197, t. h. Bos taurus; Dschidda, Arabia.—1898: Gastrothylax.
spatula Duj., 1845a, 394, t. h. Accentor modularis, int.; Rennes.—1845: Dist.
(Dicroccelium).

spatula Sieb., 1835, 57, for spathula Crep., 1829.—1836: Holost. 1898: Conchosomum.

spatula Odhn., 1902, 66, t. h. Bagrus docmac, B. bayad; Sudan.—1902: Phyllodist. 1902: Catoptroides (type).

spatulxforme Odhn., 1902, 67, t. h. Malapterurus electricus; Africa.—1902: Phyllodist. 1902: Catoptroides.

spatulatum Cobbold, 1879, 28, for spathulatum Leuck., 1876, 871; not spatulatum Rud., 1819.—1879: Dist. [Clonorchis sinensis.]

spatulatum Leidy, 1859, 111, t. h. a fish; America.—1859: Monost.

spatulatum Rud., 1819a, 109, t. h. Ardea minuta; Vien. Mus.—1819: Dist. 1845: Holost. 1902: Sodalis (type) spathulatus. 1898: Echinost. [spathulatum].

speciosus Stiles & Hass., 1896, 151, D. longissimum corvinum, 1894, renamed because of D. corvinæ, 1886; but corvinæ should not invalidate corvinum.—1896: Opisthorchis.

spenceri Haswell, 1893e, 97, t. h. Astacopsis bicarinatus; Australia.—1893: Craspedella (type).

spermatica Blumberg, see Chiaje, 1833, 34.— : Cerc.

sphæricus Klein, 1905, 68, t. h. Rana hexadactyla.—1905: Pleurogenes.

sphærocephalum Westrumb, 1823, 396, t. h. Coracias jugularis; Rio de Janeiro, Brazil.— 1823: Amphist. 1850: Holost.

sphærostomum Schlotthauber, 1860, 130, t. h. Corvus caryocatactes.—1860: Dist.

sphærula Looss, 1896b, 81, t. h. Rhinolophus hippocrepis Bonap.; Ghizeh.—1896: Dist. 1899: Lecithodendrium.

sphærula Rud., 1803a, 23, t. h. Corvus cornix; Greifswald.—1803: Amphist. 1845: Holost.

sphyra Braun, 1890a, 544, for sphyrna (Dactylogyrus).

sphyrænæ Tasch., 1879, 612, t. h. Sphyræna vulgaris; Naples.—1879: Didymozoon.

sphyrna Linst., 1878, 229, t. h. Abramis vimba; middle Europe.—1878: Dactylogyrus.
spiculator Duj., 1845a, 424, t. h. Mus decumanus; Rennes.—1845: Dist. (Echinost.).
1860: Echinost.

spiculigerum Muehling, 1898, 18, t. h. Fuligula nyroca.—1898: Dist. 1899: Psilost.
spinacis Goto, 1894a, 224, t. h. Spinax sp.; at Odawara, Japan.—1894: Onchocotyle.
1899: Squalonchocotyle.

spindalis Montgomery, 1906, 147, t. h. Bos indicus; India.—1906: Schistosoma.

spinetum Braun, 1901, 563, t. h. Rhynchops nigra L.; Brazil.—1901: Microlistrum.

spiniceps Looss, 1896b, 114, t. h. Bagrus bayad; Cairo, Egypt.—1896: Dist. 1899: Anoiktost. 1899: Acanthost. (type). 1901: Acanthochasmus (type).

spinifera La Valette, 1855, 17, t. h. Paludina vivipara, Planorbis corneus.—1855: Cerc. 1858: C. (Nephrocephala).

spiniferum Ratz, 1903, 422, t. h. Botaurus stellaris; Hungary.—1903: Pegasomum.

spinosissimum Stoss., 1883, 112, t. h. Box salpa; Triest.—1883: Monost. 1901: Centroderma (type).

spinosum Linst., 1880, 51, t. h. Sylvia rufa.—1880: Dist. 1892: D. (Dicroccelium). [1899: ?D. cirratum.]

spinulosum Rud., 1808a, 458; 1809a, 425, t. h. Larus nævius, L. cinerarius, Colymbus septentrionalis; Greifswald.—1809: Dist. (Echinost.). 1858: Echinost.

spinulosum Hofmann, 1899a, 184.—1899: Dist. 1899: Harmost.

spinulosum Hofmann, 1899a, 178, of pulmonates.—1899: Cercariæum. [1899: Dist. spinulosum.]

spinulosum Looss, 19011, 623, t. h. Chelone mydas; Alexandria, Egypt, from Egyptian coast.—1901: Amphist.

spirale Dies., 1850a, 325, t. h. Hypsilophus tuberculatus, Podocnemis tracaxa, Chelonsidis tuberculatus; Brazil.—1850: Monost. 1892: Dist.

spirale Fil., teste Par., 1896, 2, t. h. Dentex vulgaris.—Dist.

squali E. Bl., 1847, 327, t. h. Squalus; New Zealand.—1847: Trist. [1850: T. blanchardii.]

squali grisei Risso, ——, 38, see Dies., 1850a, 347.—Fasc. [1850: Dist. veliporum.] squamata Kerbert, 1881a, 556, for squamula (Dist.).

squamatum Linst., 1906, 174, t. h. Dissura episcopus; Palatupana.—1906: Lyperosomum.

squamatus Odhn., 1905, 297 (armatum Mol. of Olss.), t. h. Cottus scorpius; Belgium.—1905: Prosorhynchus (type).

squamosum Villot, 1878, 20, t. h. Strepsilas interpres.—1878: Holost. 1890: Dist.

squamula Rud., 1819a, 103, t. h. Mustela putorius: Mus. Vien.—1819: Dist. 1845: D. (Eurysoma [type]). 1850: Monost. 1892: D. (Dicroccelium). 1898: Eurysoma (type).

squamula Heath, 1902, 109, t. h. Paralichthys californicus.—1902: Epibdella. 1903: Phylline. 1905: E. (Phylline).

squatinæ Ben., 1865a, 11, t. h. Squatina angelus; Ostend, Belgium.—1865: Pseudocotvle (type).

squillarum; Triest.—1889; Mescocotyle (type). 1898; Dactylocotyle. 1895; Octobothrium. [1898; Dactylocotyle mer-

langi.]

stagnalis Mueller, 1774, 53, in stagnis.—1774: Fasc. 1776: Planaria.

stanleyi Cobbold, 1879b, 357, for stanleyii (Amphist.).—1879: Amphist. collinsii.

stanleyii Cobbold, 1875n, 819, t. h. Equus caballus; India.—1875: Amphist.

steenstrupi Odhn., 1902, 68, t. h. Anarrhichas minor s. pantherinus; Coll. Copenhagen.—1902: Lepidophyllum (type).

stenocotyle Cohn, 1902, 880, t. h. Herpetodryas fuscus; South America.—1902: Leptophyllum (type).

stentomi Goto, 1899a, 279, t. h. Stenotomus chrysops; Newport, R. I.—1899: Microcotyle.

sternæ cantiacæ Dies., 1858, 355, t. h. Sterna cantiaca; based on LaValette and Moul., 1856, 102.—1858: Dist.

stossichi Mont., 1893, 87, for stossichii (Apoblema).—1899: Hemiurus.

stossichi Luehe, 1901 of Odhn., for stossichii.—1905: Hemiurus. [1905: H. lühei.]

stossichi Braun, 1899, 80, t. h. Mustelus lævis.—1899: Calicotyle. stossichianum Mont., 1892, 12, t. h. Box salpa; Italy.—1892: Monost.

stossichii Mont., 1891, 512, t. h. Clupea aurita, C. pilchardus.—1891: Apoblema. 1893: Dist. 1899: Hemiurus. 1907: Aphanurus, type.

strephocelium Fischder., 1902a, 19, for streptocelium (Paramphist.).

streptocælium Fischder., 1901, 369, t. h. Bos kerabau; Ceylon.—1901: Paramphist.

striata Herbst, 1787a, 34.—1787: Planaria.

striata Par. & Perugia, 1890, 7, 21, t. h. Lichia amia; Triest.—1890: Vallisia (type). 1890: Octoboth; [Octobothrium arcuatum].

striatum Rud., 1809a, 343. t. h. Falco milvus; Europe.—1809; Amphist. [1819; A. macrocephalum.] [1850; Hemist. spathula.]

stridulæ Reich, 1801, 371, t. h. Strix stridula.—1801: Bist. 1801: Dist. [1809: D. apiculatum.]

strigata Mueller, 1774, 66, in aquis paludosis.—1774: Fasc. 1776: Planaria.

strigis Schrank, 1788a, 16, t. h. Weideneule.—1788: Festuc. 1790: Fasc. [Planaria.]
1790: Strigea, type. [1801: Amphist., type.] [1809: Amphist. macrocephalum, type.] [1850: Holost. variabile.]

strigis stridulæ Braun in Rud., 1809a, 347.—1809: Fasc.

strigosum Looss, 1899, 634, t. h. Merops apiaster; Marg.—1899: Dicrocœlium. 1899: Lyperosomum.

sturionis Rud., 1809a, 435, t. h. Acipenser sturio; Arimini.—1809: Dist. [——: D. hispidum.]

sturionis Abildg., 1794b, 55, t. h. Stören.—1794: Hirudo. ——: Trist. [1850: Nitzschia elegans.] 1852–53: Nitzschia. 「1878: T. elongatum.]

stylosa Linst., 1875, 193, t. h. Planorbis vortex.—1875: Cerc.

subclavata [Pallas, 1760, 29;] Gœze, 1782a, 178, t. h. Rana; Germany.—[1760: Fasc.]
 1782: Planaria. 1788: Fasc. 1800: Dist. 1802: Amphist. 1836: Diplodiscus (type). [1850: Diplocotyle.]

subclarata ore sessile Pallas, 1760, or 1761, 271, t. h. Rana.—1760: Fasc.

subelavatum Sons., 1893, 187, for subclavatum (Amphist.).

subflavum Sons., 1892, 91, t. h. Zamenis viridiflavus.—1892: Dist. 1895: D. (Brachylaimus).

[subtilis (Nitzschia), a diatom.]

subtriquetrum Rud., 1814a, 100, t. h. Castor fiber: Berlin, Germany.—1814: Amphist.
 1840: Dist. 1901: Cladorchis (Stichorchis). 1902: Stichorchis.

subtriquetrum giganteum Kuech., 1855, 192 (?for subtriquetrum, giganteum).—1855: Amphist.

subulata Herbst, 1787a, 36.—1787 Planaria.

subulo Pag., 1857, 19, t. h. Paludina vivipara—1857: Cerc. 1858: C. (Acanthocephala).
suis Stiles, 1898a, 22, musculorum suis Duncker, 1896, named binominally; t. h. Sus scrofa dom.; Germany.—1898: Agamodist.

sulcatum Linst., 1883, 309, t. h. Perdix græca; Turkestan.—1883: Dist. 1892: Clado-cœlium.

sulcatum Rud., 1809a, 337, t. h. Rana pipa, intestine.—1809: Monost. (Monost.).

superbum Staff., 1904, 492, t. h. Ameiurus nebulosus, Perca flavescens; Canada.—1904: Phyllodist.

suspensum Braun, 1901, 948, t. h. Corvus sp.—1901: Dist.

syamula, see squamula.

sygnoides Nord., 1840, 617, for cygnoides (Dist.).

sylviæ Rud., 1819a, 675, t. h. Sylvia cyanea; Brazil.—1819: Amphist.

synethes Fischder., 1901, 371, t. h. Bos kerabau; Ceylon.—1901: Gastrothylax.

syrinus Kuech., 1855, 471, misprint for gyrinus (Cerc.).

tabulatum Mueller, 1897, 21, in Numenius arquatus.—1897: Echinost.

tacapense Sons., 1894, 111: 1894, 2, t. h. Chamæleo vulgaris, Rana esculenta, Buío sp.; Gabes in Tunis.—1894: Dist. (Brachycœlium). 1899: Pleurogenes. [1899: D. medians Olss.]

tadornæ Rud., 1819a, 89, for anatis tadornæ, syn. of isostomum.—1819: Amphist.

tænioidea Nord., 1840, 595, for tænioides (Polyst.).

tænioides Rud., 1809a, 441, t. h. Canis familiaris; France.—1809: Polyst. (Pentast.).
 1819: Pentast. 1819: Prionoderma. [1824: prionoderme (type).] 1789: Linguatula, type.]

tænioides Mont., 1888a, 93, t. h. Orthagoriscus mola.—1888: Didymozoon. 1904: Nematobothrium (Didymozoon).

tagax Braun, 1901g, 896, t. h. Hirundo versicolor; Brazil.—1901: Stomylotrema.

talpæ Viborg, 1795, 242.—1795: Dist.

talpæ cæcæ Dies., 1850a, 472.—1850: Trematodum.

tanagræ Rud., 1819a, 674, t. h. Tanagra tatoa; Brazil.—1819: Amphist.

tarda Steenstrup, 1842, 75, Cerc. armata Siebold, renamed.—1842: Dist. [1858: Tetracotyle typica.]

tardigrada Dies., 1850a, 293, D. duplicatum, renamed.—1850: Rhopalocerca (type).

tartinii Stoss., 1899, 6, t. h. Oblata melanura; Triest.—1899: Dist.

taschenbergi Par., 1894, 704, for taschenbergii (Dactylocotyle).

taschenbergi Par. & Per., see St. Remy, 1898, 555.—Diclodophora.

taschenbergii Par. & Perugia, 1889, 743, t. h. Sargus rondeletii; Genova.—1889: Choricotyle. 1890: Octobothrium. 1894: Dactylocotyle (Choricotyle).

tasmanica Haswell, 1900, 430, t. h. Astacopsis tasmanicus.—1900: Temnocephala.

tectum Linst., 1873, 104, t. h. Osmerus eperlanus.—1873: Dist. [1905: Brachyphallus crenatus, type.]

tellinæ balticæ Dies., 1850a, 298, t. h. Tellina baltica; Gedani.—1850: Cerc. 1855: Cercariæum.

temperatus Staff., 1905, 689, t. h. Rana catesbiana, R. virescens; Canada.—1905: Diplodiscus.

tenax Mueller, 1773, 69, in infusione sordium dentium intra quatriduum.—1773: Cerc. 1827: Phacus.

tener Kowal., 1903, 517, t. h. Mergus merganser.—1903: Metorchis.

tenere Looss, 1898, 458, D. tacapense Sonsino of Looss renamed.—1898: Dist. 1898: Pleurogenes. 1899: Prosotocus. 1899: D. (Pleurogenes).

tenerum Looss, 1899b, 616, 622, see tenere (Dist.).

tensum Looss, 1902h, 141, t. h. Mugil chelo; Triest.—1902: Saccocælium.

tentaculata Mueller, 1774, 63, in aquis palustribus.—1774: Fasc. 1776: Planaria.

tenue Lint., 1898, 535, t. h. Roccus lineatus.—1898: Dist.

tenue tenuissime Lint., 1898, 536, t. h. Morone americana.—1898: Dist.

tenuicolle Rud., 1819a, 93, t. h. Phoca barbata.—1819: Dist. 1892: D. (Brachylaimus). 1896: Opisthorchis.

tenuicolle Rud., 1819a, 85, t. h. Lampris guttatus; Groningæ.—1819: Monost. (Monost.) [1850: Dist. affine Dies. nec Rud.].

tenuicolle Westrumb, 1823, 391, t. h. Falco rufus; [Mus. Cat. Vien.].—1823: Amphist. 1850: Holost.

tenuicollis-felineus Looss, 1899, 678, see tenuicollis.—1899: Opisthorchis.

[tenuis Muenster, 1842, 99.—Hirudinella. Hirudella, a leach.]

tenuis Wedl, 1857, 258, t. h. Perca fluviatilis.—1857: Gyrodactylus. 1858: Dactylogyrus.

tenuissime Lint., 1898, 536, t. h. Morone americana.—1901: Dist. [1898: D. tenue.] teres Gœze, 1782a, 173.—1782: Planaria.

teres duplici poro Gœze, 1782a, 173.—1782: Planaria.

teres poro simplici Gœze, 1782a, 173.—1782: Planaria.

tereticolle lucii Mayer, 1841, 18.—1841: Distoma.

tercticolle rosaceum (Nordmann, 1832) Dies., 1850a, 364.—1850: Dist.

tereticollis Rud., 1802, 74–75, lucii Mueller renamed.—1802: Fasc. 1808: Dist. 1845: Dist. (Brachylaimus). 1899: Azygia (type). 1904: Azigia.

teretiusculum Mont., 1893, 40, t. h. Solea klenii.—1893: Dist.

tergestinum Stoss., 1889, 28, t. h. Oblata melanura; Triest.—1889: Dist.

tergestinum Stoss., 1883, 119, t. h. Gobius niger, G. jozo; Triest.—1883: Gasterost. terrestris Mueller, 1774, 68, in asseribus muscisque humidis.—1774: Fasc. 1776:

Planaria. [Rhynchodesmus.]

terrestris Linst., 1889c, 241, t. h. Helix lens; Greece.—1889: Cerc.

terricola Linst. 1889c, 241, t. h. Helix ?vermiculata; Algiers.—1889: Cerc.

testudinis Rud., 1819a, 121, t. h. Testudo orbicularis; C. E. V.—1819: Dist. [1850: Monost. delicatulum.]

testudinis Braun, 1899, 630, MS. name in Vienna Coll.—1899: Monost. [1899: Dist. scyphocephalum.]

testudinis midæ Braun, 1899, 629, for t. mydæ (Dist.).

testudinis mydæ Rud., 1809a, 433, for D. intestinalis testudinis mydæ.—1809: Dist.

teihyæ Mont., 1892, 717.—1892: Monost.

tetracystis Gastaldi, 1854, 4, t. h. Rana esculenta.—1854: Dist. 1905: Cystagora (type). tetracystis ranæ esculentæ Dies., 1855, 64; see tetracystis.—1855: Dist.

tetragona Mueller, 1774, 69, in stagno ac foveis aquæ purioris nec vulgaris.—1774: Fasc. 1776: Planaria.

tetrodonis Goto, 1894a, 213, t. h. Tetrodon sp.; Hagi, Japan.—1894: Diclidophora.

1896: Heterobothrium (type).

tetrodontis St. Remy, 1898, 554, for tetrodonis (Diclidophora) (Heterobothrium).

texanicum Francis, 1891c, 135, t. h. Bos taurus; Texas.—1891: Dist. [Fasc.] [Fasc. magna.]

texicanum Leuck., 1892b, 797, for texanicum (Dist.).

thaumanthiadis Braun, 1889a, 357, for thaumantiatis (Cerc.).

thaumantiadis Pag., 1862, 298, for thaumantiatis (Cerc.).

thaumantiatis Græffe, 1860a, 49, t. h. Thaumantias.—1860: Cerc.

thethydis delle Chiaje, teste E. Bl., 1847a, 309.—Monost.

thetidicola Otto, 1823, 294, t. h. Thetis fimbria.—1823: Vertumnus (type). [1823: Phœnicurus varius.]

thetycola (delle Chiaje?), see Pag., 1862, 298.—Monost.

thompsonii Mueller, 1841, 147.—1841: Cyclocirra (? type).

thunninæ Par. & Per., 1889, 742, t. h. Thynnus thunnina; Genova.—1889: Octocotyle. 1890: Octobothrium. 1896: Hexacotyle.

thynni Delaroche, 1811a. 271, t. h. Scomber thynnus; Maroque.—1811: Polyst. [1815: Hexost., type.] 1828: Hexacotyla (type). 1840: Hexacotyle (type). [1850: Plagiopeltis duplicata, type]. 1890: Exacotyle. 1891: Plagiopeltis.

thynni Tasch., 1879, 612 (Monost. bipartitum, renamed), t. h. Thynnus vulgaris.— 1879: Didymozoon, type. [1902: Didymocystis reniformis, type].

thynni Par. & Per., 1891, 19, [lapsus for thunninæ?].—1891: Octocotyle.

tincæ Modeer, 1790, 127, t. h. Cyprinus tinca.—1790: Fasc. 1809: Dist. [1809: D. globiporum]. [1850: D. perlatum.]

tobiani Kræyer, 1846-53a, 592, t. h. Ammodytes tobianus.—1846-53: Dist.

todari delle Chiaje, 1841, 139.—1841: Dist.

todari delle Chiaje, see Par., 1894, 168, in Ommastrephes todarus; Naples.—Monost.

tornatum Rud., 1819a, 684, t. h. Coryphæna equiselis, C. hippuris.—1819: Dist. 1845: D. (Apoblema). 1889: Apoblema. 1899: Hemiurus. 1901: Lecithocladium. torosum Setti, 1897, 4, t. h. Squalus sp.; Massaua.—1897: Otiotrema (type). 1903: Dist.

torpedinis Chatin, 1874a, 11, t. h. Torpedo marmorata; Mediterranean.—1874: Amphibdella (type). 1890: Tetraonchus.

torulosum Rud., 1814a, 104, t. h. Silurus glanis; Greifswald.—1814: Dist.

torva Mueller, 1774, 62, in aquis.—1774: Fasc. 1776: Planaria.

totari E. Bl., 1847a, 309, for todari (Dist.).

totari E. Bl., 1847a, 309, for todari (Monost.).

trachea Montagu, 1811, 194. t. h. poultry, Gallus gallus.—1811: Fasc. 1819: Dist. [Syngamus trachealis, a nematode].

tracheicola Braun, 1901, 943, t. h. Anas fusca; Vienna, Austria.—1901; Orchipedum (type).

trachini Par. & Perugia, 1889, 744, t. h. Trachinus radiatus: Genova.—1889: Microcotyle.

trachuriBen. & Hesse. 1863, 1864, 118, t. h. Caranx trachurus.—1863: Gastrocotyle (type).

trachuri Par. & Perugia, 1890, 11, t. h. Caranx trachurus; Genoa.—1890: Pseudaxine (type).

translucida Staff., 1902, 413, t. h. Bufo lentiginosus, Rana virescens; America.—1902: Gorgodera. 1905: Gorgoderina.

transversalis Rud., 1802, 69, t. h. Cobitis fossilis; Berlin, Germany.—1802: Fasc. 1809: Dist. 1901: Allocreadium.

trapezium Leidy, 1891a, 414, t. h. Pandion carolinensis.—1890: Dist.

tremellaris Mueller, 1774, 72, in mare Balthico, Hafniam alluente.—1774: Fasc. 1776: Planaria.

tremoctopodis Kælliker, 1849e, 67.—1849: Hectocotylus. [1849: ♂ of Tremoctopus.] triangulare Johnston, 1904, 108, t. h. Dacelo gigas.—1904: Hemist.

triangulare Dies., 1850a, 351, t. h. Merops apiaster.—1850: Dist. 1899: Megacetes (type). 1901: Plagiorchis. 1900: Eumegacetes (type).

triangulare Dies. of Looss, see emendatus Braun.—1899: Megacetes (type). [1900: Eumegacetes, type].

triangulare Johnston, 1904, 108, t. h. Dacelo gigas.—1904: Hemist.

triangularis Goto, 1894a, 200, t. h. Acanthias schlegelii; Misaki, Japan.—1894: Axine. tricaudata Schrank, 1803, 86.—1803: Cerc.

tricolor Stiles & Hass., 1894, 729, t. h. Lepus sylvaticus, L. americanus; Maryland.—1894: Dist.

trifolium Braun, 1901, 947, t. h. Ardea coicoi; Brazil.—1901: Dist.

triganocephalum Kampmann, 1894b, 454, misprint for trigonocephalum (Dist.).

triglæ Ben. & Hesse, 1863; 1864, 92, t. h. Trigla.—1863: Udonella.

triglæ Ben. & Hesse, 1863, 117; 1864, 117, t. h. Trigla hirundo.—1863: Axine.

triglæ Ben., 1870, 30, t. h. Trigla hirundo; Belgium.—1870: Gasterost.

triglæ Rud., 1819a, 122, t. h. Trigla cuculus; C. E. V.—1819: Dist.

triglæ gurnardi Rathke, 1799, 68, t. h. Trigla gurnadus.—1799: Dist. [D. soleæforme.] triglæ pini Dies., 1855, 64, t. h. Trigla pini.—1855: Dist. [1858: D. homoeostomum.] trigonocephala Rud., 1802b, 87, F. melis Schrank, renamed, t. h. Schweinigel.—1802:

Fasc. 1809: Dist. (Echinost.). [1828: type of festucaire.] 1860: Echinost.

trigonocephalum Rud., 1809a, 336, t. h. Testudo mydas; Europe.—1809: Monost. (Monost.) 1901: Pleurogonimus.

trigonocephalum Rud. of Looss, 1899b, 666, renamed obliquus 1901.—1899: Pronocephalus (type, see obliquus).

trigonocerca Dies., 1858d, 259, C. limacis Moul., t. h. Limax cinereus, L. (Arion) rufus; Geneva.—1858: Cerc. (Acanthocephala).

trigonostoma Wagener, 1857, 90, t. h. Cyprinus rutilus.—1857: Dactylogyrus.

triloba Fil., 1857c, 3, t. h. Planorbis carinatus, Lymnæus stagnalis; Turin.—1857: Cerc. 1858: C. (Acanthocephala).

trilobum Rud., 1819a, 104, t. h. Pelecanus carbo; Mus. Vien.—1819: Dist. 1850: Hemist.

tringæ Brand., 1892, 507, t. h. Tringa variabilis; Sinai.—1892: Monost. [1902: Cyclocelum.] 1902: Hæmatotrephus.

tringæ helveticæ Rud., 1819a, 120, t. h. Tringa helvetica; C. E. V.—1819: Dist. [1850: D. cinctum.]

tripes Bosc, 1802a, v. 3, 227, see also tripos (Cerc.).

tripos Mueller, 1776, 206, in aqua marina.—1776: Cerc. 1827: Ceratium.

tripunctata Erc., 1881e, 23, t. h. Planorbis corneus; Italy.—1881: Cerc.

tripus Gmelin, 1790a, 3892, for tripos (Cerc.).

triserialis Dies., 1839a, 234, t. h. Anas anser dom. et ferus, etc.; Europe.—1839: Notocotylus (type). 1840: Nocotylus. 1850: Notocotyle (type). 1860: Monost.

troncatum Perroncito, 1882, 248, for truncatum (Dist). [Ercolani.]

truitta Bosc, 1802a, 274, for truttæ (Fasc.).

truncata Mueller, 1806, 35, t. h. Perca lucioperca.—1806: Fasc. 1814: Dist.

truncata Dies., 1850a, 422, Octostoma scombri renamed, t. h. Scomber scombrus; Rhedoni.—1850: Octocotyle. 1858: Octoplectanum. 1879: Octobothrium.

truncata Goto, 1894a, 191, t. h. Pristipoma japonicum; Japan.—1894: Microcotyle.

truncata Bosc, 1802a, 262.—1802: Planaria.

truncata Leidy, 1851b, in 224-227, free form; Newark, Del.—1857: Planaria.

truncatum Rud., 1819a, 91, t. h. Phoca vitulina; Europe, Berlin, Vratislaviæ.—1819: Amphist. 1886: Dist. 1896: Opisthorchis. 1899: Metorchis.

truncatum F. S. Leuck., 1842, 34, t. h. Sorex fodiens.—1842: Dist.

truncatum Erc., 1859a, 382, t. h. Canis familiaris; Italy.—1859: Dist. [D. truncatum (Rud.).]

trunkatum Schneidemuehl, 1896, 303, for truncatum (Amphist.).

truttæ Moul. of Dies., 1858, 356.—1858: Dist.

truttæ Frælich, 1789, 126, t. h. Salmo trutta; Europe.—1789: Fasc. [1809: Dist. laureatum.]

truttæ (intestinalis) Ræderer, 1762, 537, t. h. forelle.—1762: Fasc. [1810: Ligula nodosa.] [1809: ?Echinorhynchus fusiformis.] [1905: Echinorhynchus.]

tuba Linst., 1878, 228, t. h. Squalius leuciscus.—1878: Dactylogyrus.

tuba Braun (1805), 49.—1805: Hirudo. [1809: Amphist. subclavatum.]

tubarium Rud., 1819a, 111, t. h. Sciæna umbra; Spezia.—1819: Dist.

tuberculata Fil., 1857c, 8, t. h. Paludina impura.—1857: Cerc. 1858: C. (Gymnocephala).

tuberculatum Cobbold, 1875n, 819, t. h. ox; India.—1875: Amphist.

tubiporum Dies., 1836a, 14, t. h. Trigla hirundo.—1835: Trist. 1840: Capsala. [1850: Trochopus longipes, type.] 1864: Trochopus (type).

tubiporum Braun, 1900, 388, t. h. Vespertilio sp.—1900: Dist.

tubulatum Rud., 1819a, 675, t. h. Muræna sp.; Brazil.—1819: Dist.

tumbesiana Wacke, 1903, 1.—1903: Temnocephala.

tumidulum Rud., 1819a, 95, t. h. Syngnathus hippocampus; Vienna.—1819: Dist. 1901: Allocreadium.

turbo Mueller, 1786, 123, in aqua rivulari cum Lemna rarior.—1786: Cerc. [1827: Urocentrum, type.]

turdi Rud., 1819a, 120, t. h. Turdus saxatilis; C. E. V.—1819: Dist. [1850: D. mesostomum; D. nephrocephalum.]

turgidum Brand., 1888, 247, t. h. Rana esculenta.—1888: Dist. 1899: Brandesia (type).

tursionis Marchi, 1872, 304, t. h. Delphinus tursio, int.—1872: Dist. 1896: D. (Dicrocœlium). [D. (Dicrocœlium) longissimum Poir.]

tynni Crety, 1892c, 399, for thynni (Exacotyle).

typica Dies., 1858e, 366, t. h. see p. 369; Europe.—1858: Tetracotyle (type).

[typus, reptile, Bucephalus.]

umblæ Fabricius, 1780a, 329, t. h. Salmo umbla; Greenland.—1780: Fasc. 1803: Dist. [1809: D. seriale.]

umbonatum Odhn., 1902, 21, t. h. Krokodil; Nile.—1902: Echinost.

umbrinæ Stoss., 1885, 159, t. h. Umbrina cirrhosa; Triest.—1885: Dist. 1905: Allocreadium. 1886: D. (Brachylaimus).

unciforme Rud., 1819a, 674, t. h. Oriolus cristatus; Brazil.—1819: Amphist. 1850: Holost.

uncinata Dies., 1850a, 412, lapsus for uncinulata.—1850: Fasc. [1850: Polyst. integerrimum.]

uncinatum Macé (1880).—1880: Polyst.

uncinatum Zed., 1803a, 221, t. h. Fulica chloropus.—1803: Dist. 1809: D. (Echinost.). 1860: Echinost. [1892: E. cinctum.]

uncinatum Mont., 1889, 117, t. h. ?Pleuronectes; Coll. Leuck.—1889: Trist.

uncinatus Wagener, 1857, pl. 13, fig. 2, t. h. Perca fluviatilis.—1857: Dactylogyrus.

uncinulata Braun, 1790, 58, t. h. Rana esculenta: Germany.—1790: Planaria. 1790: Fasc. [1800: Polyst. ranæ, type.] [1850: Polyst. integerrimum.]

uncinulatum Macé in Braun, 1890a, 418, for uncinatum (Polyst.).

undulatus Looss, 1899, 594, t. h. Thalassochelys corticata; Abukir.—1899: Cymatocarpus (type).

unguicolatus Mont., 1888, 90, for unguiculatus Tetraonchus (Dactylogyrus).

unguiculatum Rud., 1819a, 91, t. h. Triton palustris; Berlin, Germany.—1819: Amphist. 1836: Diplodiscus.

unguiculatus Wagener, 1857, 61.—1857: Dactylogyrus. 1858: Tetraonchus.

unicum Mol., 1859, 835, t. h. Centrolophus pompilius; Padua.—1859: Dist. 1886: D. (Podocotyle).

unicum Odhn., 1902, 66, t. h. Serranus sp.; Sinai, Red Sea.—1902: Phyllodist.

unicum Looss, 1896b, 44, renamed reniferum; t. h. Trionyx nilotica; Egypt.—1896: Dist. 1899: Astia (type).

unionicola Graff, 1904, 457, t. h. Muscheln.—1904: Planaria.

upupæ Schrank, 1790, 123, t. h. Upupa epops, rectum.—1790: Fasc. [1809: Dist. involutum.]

urcatum Luehe, 1900, 489, misprint for furcatum (Dist.).

urna Looss, 1907, 485, t. h. Vesperugo kuhli; Cairo, Egypt.—1907: Lecithodendrium. urniceps Schlotthauber, 1860, 129, t. h. Falco pygargus.—1860: Holost.

urnigerum Rud., 1819a, 89. t. h. Rana esculenta; Mus. Vien.—1819: Amphist. 1845: Holost. [1850: Codonocephalus mutabilis, type.]

urocotyle Par., 1899, 6, t. h. Scorpæna scrofa; Portoferrajo.—1899: Pleorchis. 1905: Derogenes.

vagans Leidy, 1850, 304, t. h. Helix alternata, H. albolabris; D. helicis renamed.—1850: Dist. 1855: Cercariæum.

vaginatum Brand., 1888a, 64, t. h. Cathartes sp.; Brazil.—1888: Holost.

valdeinflatum Stoss., 1883, 114, t. h. Gobius jozo; Triest.—1883: Dist. 1886: D. (Echinost.) 1898: Agamodist. [1899: Stephanost. cesticillus.] valdemflatum Nicoll, 1907, 69, for valdeinflatum (Dist.).

validum Linst., 1886, 124, t. h. Delphinus sp.; South Atlantic.—1886: Dist. 1892: D. (Brachylaimus).

vallei Stoss., 1896, 129, t. h. Falco subbuteo.—1896: Dist.

vallei Stoss., 1899, 3, t. h. Thalassochelys caretta; Corfu.—1899: Aspidogaster. [1901: Lophotaspis adhærens.] 1902: Lophotaspis, type. 1902: Amphist., probably lapsus.

vallei Par. & Per., 1895, 3, t. h. Naucrates ductor; Genova.—1895: Placunella. 1903: Ancyrocotyle (type).

van benedeni St. Remy, 1898, 567, for van benedenii (Dactvlogyrus).

van benedenii Par. & Per., 1890, 96-97, t. h. Mugil auratus.—1890: Tetraonchus.

van benedenii Par. & Per., 1895, 2, t. h. Mugil auratus; Triest.—1895: Dactylogyrus. See also benedenii.

vanelli Rud., 1819a, 87, t. h. Tringa vanellus; Mus. Vien.—1819: Monost. [1850: M. mutabile.]

variabile Leidy, 1856, 44, t. h. Tropidonotus sipedon.—1856: Dist. 1903: Renifer.

variabile Nitzsch, 1819, 400, includes macrocephalum Rud., t. h. owls; Germany.— 1819: Holost. (type). 1860: Monost.

varica Mueller, 1784, 93, t. h. Salmo salar, stomach.—1784: Fasc. 1803: Dist. 1886: D. (Brachylaimus). 1899: Hemiurus. 1901: Derogenes.

varicans Abildg., 1794, 89, in aqua rivulari.—1794: Cerc. 1850: Cheilost. (type).

variegatum Crep., 1825a, 38, t. h. Larus marinus; Greifswald.—1825: Amphist. 1845: Holost.

variegatum Rud., 1819a. 99, t. h. Rana esculenta; Berlin.—1819: Dist. 1845: D. (Brachylaimus). 1847: Brachylæmus. 1850: Brachylemus. 1899: Hæmatolæchus (type). 1902: Pneumonœces (type).

varigatum Looss, 1892, 93, misprint for variegatum (Dist.).

variolaris Fuhrmann, 1904, 59, t. h. Rostrhamus sociabilis; South America.—1904: Bothriogaster (type).

[variolosus Gærtner (Dist.), a tunicate.]

varioplexus Staff., 1902, 901, t. h. Rana catesbiana; Canada.—1902: Hæmatolæchus. 1905: Pneumonœces.

varium Eysenhardt, 1829, 148, t. h. Gadus merluccius.—1829: Dist.

varsoviensis Ssinitzin, 1905, 36, t. h. frogs; Warschau.—1905: Gorgodera. 1906: Cerc. velellæ Fil., 1843, 66, see Dies., 1850a, 379, t. h. Velella spirans; Naples.—1843: Dist. [1850: D. megacotyle.]

velellæ Graff, 1904, 456.—1904: Plan.

reliporium Johnston, 1902, 329, misprint for veliporum (Dist.).

veliporum Crep., 1837a, 310, t. h. Squalus griseus.—1837: Dist. 1886: D. (Cladocœ-

lium). 1904: Otodist. (type).

venarum Treutler, 1793, iv, 23, t. h. Homo.—1793: Hexathyridium. 1803: Fasc. 1803: Polyst. 1828: Hexathiridium. 1836: Polyst. (Hexast.). 1850: Hexacotyle. [Hexast.] [1840: Linguatula.]

ventricolum Rud., 1809a, 334, misprint for ventricosum (Monost.).

ventricosa Rud., 1802, 20, t. h. Nachtigall; Greifswald.—1802: Festucaria. 1809: Monost. (Monost.).

ventricosa Bosc., 1802a, 257.—1802: Planaria.

ventricosa Pallas, 1774, 17, host not given; Amboyna.—1774: Fasc. 1853: Hirudinella. 1893: Dist.

ventricosum Rud., 1819a, 108, t. h. Clupea alosa; Arimini.—1819: Dist. 1886: D. (Apoblema). 1893: Apoblema. ventricosum Stoss., 1898, 29, t. h. Alausa finta; Triest.—1898: Dist. [1898: Apoblema

ocreatum Rud.]

ventricosum var. minor Shipley, 1900, 540, t. h. Pimelepterus sp.; New Britain.— 1900: Dist.

vereticolle lucii Rud., 1809a, 400, lapsus for tereticolle lucii (Dist.).

vermicularis Mueller, 1786, 133, in aqua ubi Lemna vegetat.—1786: Cerc. 1827: Dicranophorus.

verrilli Goto, 1899a, 283, t. h. a skate (Raja); Cape Cod.—1899: Acanthocotyle.

8588-No. 37-08-6

verrocosum Mont., 1892, 40, for verrucosum Notocotyle.

verrucosa Freelich. 1789, 112, t. h. domesticated geese.-1789: Fasc. 1800: Monost. 1809: Monost. Monost. . [1839: Notocotylus triserialis.] 1892: Notocotyle. 1905: Catatropis (type).

verrucosum Busch. 1851, 100, t. h. Ophidium barbatum.—1851: Dist. 1886: D.

verrucosum Mol., 1859, 842, t. h. Labrax lupus: Rennes, Padua,—1859; Dist, 1886; D. Dicroccelium.

verrucosum Poir., 1885, 10. t. h. Thynnus.-1885: Dist.

verrucosum Lint., 1892, 96, t. h. Larus californicus.—1892: Dist.

verucosa Odhn. LV. 1902. 63. for verrucosum Notocotyle.

resicata Uličný. 1878. 211. t. h. Cyclas rivicola: Mähren.—1878: Cerc. [1894: C. macrocerca.]

vesiculifera Dies., 1855a, 389. C. vesiculosa Fil., renamed.—1855; Cerc. 1855; C. Xiphidiocerc.). 1858; C. Acanthocephala).

resiculosa Dies., 1850a, 295. Cerc. I Baer, renamed; t. h. Paludina vivipara; Regiomontii.—1850; Cerc., 1855; C. Eucerc., 1858; C. Acanthocephala.

vesiculosa of Fil., 1854a, 12, t. h. Paludina vivipara,—1854: Cerc. [1855; C. Xiphidiocerc. vesiculiiera.]

vespertilionis Mueller. 1784. 95. t. h. Vespertilio auritus.—1784: Fasc. 1803: Dist. [--: Plan.] [1850: D. lima.] 1900: Plagiorchis.

respertilionis Rud., 1819a, 87, t. h. Vespertilio noctula: Cat. Ent. Vienn.—1819:

vetustum Staff., 1904, 487, t. h. Limanda ferruginea: Woods Hole.—1904: Stenakron

vexans Braun. 1901. 947. t. h. Turdus merula.—1901: Dist.

riber Lint., 1900, 269, t. h. Spheroides maculatus.—1900: Dist.

vicarium Braun, 1901g, 896, t. h. Ibis cœrulescens: Brazil.—1901: Stomylotrema.

vicinus Odhn., 1902, 24, t. h. Nilkrekodil: Sudan,—1902: Acanthochasmus.

villoti Mont., 1888, 194, C. setifera Mueller, of Villot, 1879, renamed, in Scrobicularia tenuis.—1888; Cerc.

vinal-edwardsii Lint., 1901, 416, t. h. Opsanus tau. Orthopristis chrysopterus: Beaufort, N. C.—1901: Monost.

vipera Linst., 1877, 180, t. h. Pelias berus.—1877; Dist. 1895; Agamodist.

vipera: Ben., 1870, 26, t. h. Trachinus vipera: Belgium.—1870: Gasterost.

virgula Fil., 1837, 338, t. h. Paludina impura, Valvata piscinalis: Ticini, Italy.—1837; Dist. 1850: Cerc. 1855: C. Eucerc. 1858: C. Acanthocephala . [1858: D.

viridata Bosc., 1802a, 258.—1802: Plan.

viridis Mueller. 1786. 126. in aquis fossarum stagnantibus primo vere.—1786: Cerc. 1815: Furcocerca. 1827: Enchelys.

viridis Bosc., 1802a, 256.—1802: Plan.

viridis Mueller, 1774. 59. in radicibus iucorum; Greenland.—1774: Fasc.

vitellatum Linst., 1875, 189, t. h. Totanus hypoleucus.—1875: Dist. 1901: Plagiorchis. 1892: D. Brachylaimus .

vitellilobum Olss., 1876, 14, t. h. Rana temporaria.—1876; Dist. 1889; D. Dicrocœlium. 1902: Gorgoderina. 1906: Gorgodera. 1906: Cerc.

vitellosum Lint., 1900, 269, t. h. Merluccius bilinearis; Woods Holl.—1900: Dist.

[vitreum Sars Dist., a tunicate.]

vitrina Linst., 1887, 105, t. h. Zebrina detrita: Frauenberg b. Sondershausen.—1887:

vitrinæ Targioni Tozzetti. 1873. 335. in vitrine.—1873: Dist.

vitta Duj., 1845a, 418. t. h. Mus musculus: Rennes.—1845: Dist. Brachylaimus.

vivæ Ben., 1870, 25. t. h. Trachinus draco: Belgium.—1870: Gasterost.

tirar Sons., 1892. Oct. 7, 137, t. h. Cleopatra bulimoides: Cairo, Egypt.—1892: Cerc.

viverrini Poir., 1886, 27. t. h. Felis viverrinus.—1886: Dist. 1892: D. (Dicrocœlium). 1896: Opisthorchis.

vivipara Ben., 1870, 28, t. h. Mugil chelo; Belgium.—1870: Dist.

viviparæ Linst., 1877, 185, t. h. Paludina vivipara.—1877: Monost. 1892: Monostomulum.

viriparæ fasciatæ Linst., 1877b, 186, t. h. Vivipara fasciata.—1877: Dist.

viviparum Olss., 1868, 28, t. h. Pleuronectes microcephalus.—1868: Dist. 1886: D. (Echinost.). 1901: Zoogonus. 1902: Zoogonoides (type).

vogtianum Baudon, 1881, 145, t. h. Succinea baudoni.—1881: Leucochloridium.

voluptarium Braun, 1901, 945, t. h. Falco sp.—1901: Dicrocœlium.

volvens Nord., 1832, 28, t. h. Gadus lota, Perca cernua, P. fluviatilis, P. lucioperca.—1832: Diplost. (type). 1833: Dist. [1898: Hemist. spathaceum.]

vulgaris Cerf., 1899a, 375, t. h. Mustela vulgaris.—1899: Squalonchocotyle.

vulpina Abildg., 1790, 63.—1790: Dist. [Hemist. alatum.]

vulpis Gmelin, 1790a, 3053, t. h. Canis vulpes; Europe.—1790: Fasc. [1809: D. alatum.] [1850: Hemist. alatum.]

vulpis Schrank, 1788, 52, t. h. Canis vulpes; Germany; Planaria alata vel dubia Gœze, 1782a, 176, pl. 14, figs. 11–13, renamed.—1788: Alaria (type).

wachniæ Rud., 1819a, 122, 427, t. h. Gadus wachnia.—1819: Dist. 1878: Trematodum.

wutsoni Conyngham, 1904, Aug. 13, 464; Sept. 17, 663. t. h. Homo; in Zola (Nigeria), came from Adamawa, German West Africa.—1904: Amphist. 1905: Cladorchis.

wedli Ariola, 1902, 105, t. h. Thynnus vulgaris; Naples.—1902; Didymocystis.

wedlii Cobbold, 1860a, 18, t. h. Rana esculenta.—1860: Dist.

wedlii Cobbold, 1860a, 43, t. h. Rhombus lævis, M. rhombi lævis Wedl, renamed.—
1860: Monost.

westermani Kerbert, 1881a, July, 529, for westermanii.—1881; Dist. 1891; Dist. 1899; Paragonimus (type). 1905; Paragonimus.

westermanii Kerbert, 1878a, 271, t. h. Kænigstiger.—1878: Dist. 1890: Mesogonimus. [1899: Paragonimus (type.)] [1899: Polysarcus, type.] 1900: Paragonimus.

westermanni Leuck., 1889, 404, 10r westermanii Kerbert, 1878.—1889: Dist. 1890: Mesogonimus. 1891: Dist. 1898: Dist. (Mesogonimus). [1899: Paragonimus (type).] 1899: Polysarcus (type).

westrumbii Cobbold, 1860a, 45, Amphist. sphærocephalum Westrumb, renamed; t. h. Coracina scutata; Brazil.—1860: Holost.

winogradoffi Jaksch, 1897a, 219 for sibiricum, t. h. Homo.—1889: Dist.

xanthostoma var. compascua Kowal., 1898, 71, t. h. Anas querquedula.—1898: Opisthorchis.

xanthosomum Crep., 1846a, 138, t. h. Colymbus septentrionalis; Germany.—1846: Dist. 1898: Opisthorchis. 1902: Metorchis.

zschokkei Volz, 1899, 231, t. h. Heterodon platyrhinus.—1899: Dist. 1903: Renifer. [For addenda, see p. 385.]

BIBLIOGRAPHY OF SUPERGENERIC, GENERIC, AND SPE-CIFIC NAMES USED IN TREMATODA AND OF DISEASES CAUSED BY TREMATODA.

The main portion of this index is made up of an alphabetical list of superspecific names used in connection with Trematoda. Under each generic or subgeneric name will be found a list of the species and subspecies which have been placed in the groups in question. The references following the technical names represent the places in literature which bear upon the technical names in question. Names of diseases caused by Trematoda are also inserted in this list.

In all cases possible, the type species a is mentioned for genera. In connection with the species, the hosts and locality, at least the type locality, are usually given.

(ACANTHOCEPHALA) Dies., 1858d, 253-255 (subg. of Cerc.) [not Acanthocephala Rud., 1808, worms, order: Acanthocephali Burm., 1837, worms, order; Acanthocephalus Kelreuter, 1771, worms, genus: Acanthocephalos Geze, 1782, for Acanthocephalus, 1771: Acanthocephala Laporte, 1832, hemipteron, genus; Acanthocephalus for Acanthocephala 1832], C. (Xiphidiocerc.), 1855, renamed, hence same type.a

armata (Sieb., 1837) Dies., 1858d, 251-252 (larva of Dist. endolobum Duj. [Opisthioglyphe 1899, type]) (in Planorbis corneus, Paludina impura, Lymnæus

stagnalis).

brachyura (Dies., 1850) Dies., 1858d, 257 (in Planorbis submarginatus; Ticini; P. nitidus, P. vortex).

buccini mutabilis (Fil., 1855) Dies., 1858d, 266 (in Buccinum (Nassa) mutabile; Genoa).

chlorotica (Dies., 1850) Dies., 1858d, 252-253 (syn. Cerc. (Eucerc.) chlorotica Dies.) (in Paludina vivipara).

gibba (Fil., 1854) Dies., 1858d, 257 (in Lymnæus pereger; Turin).

linearis (Lespés, 1857) Dies., 1858d, 258 (in Litorina litorea).

macrocerca (Fil., 1854) Dies., 1858d, 255 (larva of Dist. cygnoides Zed.) (in Cyclas cornea; Turin); 1858e, 334 (syn. of Dist. cygnoides; [type of Gorgodera, 1899]).

micracantha Dies., 1858d, 259-260 (syn. Cerc. armata Fil., 1855b) (in Lymnæus palustris, Triton punctatus).

microcotyla (Fil., 1854) Dies., 1858d. 253 (syn. C. pugnax La Valette) (in Paludina achatina. Lombardia: P. vivipara) (larva of Dist. tetracystis [type of Cystagora 1905]): 1858e, 348 (syn. of Dist. tetracystis).

micrura (Fil., 1857) Dies., 1858d, 258 (in Paludina vivipara; Turin). See globipo-

rum type of Sphærost.

ornata (La Valette, 1855) Dies., 1858d, 244, 255–256 (larva of Dist. clavigerum Rud. [type of Pleurogenes, 1899]) (in Planorbis corneus, Berlin: Hydrachna concharum, Heidelberg): 1858e, 338 (syn. of Dist. clavigerum).

pachycerca Dies., 1858d, 257-258 (syn. Cerc. brachyura Lespés) (in Trochus cinereus; Francogalliæ).

(ACANTHOCEPHALA)—Continued.

planorbis carinati (Fil., 1857) Dies.. 1858d, 266 (in Planorbis carinatus; Turin).

subulo (Pag., 1857) Dies., 1858d, 256-257 (in Paludina vivipara; Heidelberg).

trigonocerca Dies., 1858d, 259 (syn. Cerc. limacis Moul.) (in Limax cinereus, L. (Arion) rufus; Geneva).

triloba (Fil., 1857) Dies., 1858d, 252 (in Lymnæus stagnalis, Planorbis carinatus).
vesiculifera (Dies., 1855) Dies., 1858d, 254-255 (syns. Cerc. vesiculosa Fil., C. (Xiphidiocerc.) vesiculifera Dies., C. microcotyla Fil.) (in Paludina vivipara, P. achatinæ).

vesiculosa (Dies., 1850) Dies., 1858d, 254 (in Paludina vivipara; Regiomontii, Berlin, Heidelberg).

virgula (Fil., 1837) Dies., 1858d, 260 (larva of Dist. maculosum Rud.) (in Valvata piscinalis, Paludina impura).

ACANTHOCHASMUS Looss, 1900d, Dec. 3, 603, Acanthostomum Looss, 1899 [not Acanthostoma Kriechbaumer, 1895, insect] renamed, hence type spiniceps; χασμάσματ, den Rachen offen haben; 1901e, 595, 629, 634, 659; 1901f; 1902g; 1902m, 441, 780, 808, 838.—Braun, 1901b, 34, 35, 36; 1902b, 30.—Odhn., 1902, 23, 25, 37; 1902, 159.—Pratt, 1902a, 888, 894 (key).

absconditus Looss, 1901e, 631–632, fig. 8 (intest. of Bagrus bayad, B. docmac). coronarius (Cobbold, 1861) Braun, 1901b, 35, 36.—Odhn., 1902, 25 (in alligator).

imbutiformis (Mol., 1859, partim) Looss, 1901e, 632–633, 634, fig. 9 (intest, of Labrax lupus, Dentex vulgaris: Egyptian coast and Triest).—Odhn., 1902, 25.

præteritus Looss, 1901e, 633-634, fig. 10 (first half of intest. of Labrax lupus, Chrysophrys aurata, Corvina nigra; Triest and Egyptian coast).

productus Odhn., 1902, 24, 25 (in Nilkrokodil; Sudan).

scyphocephalus (Braun, 1899) Braun, 1901b, 35, fig. 14.—Odhn., 1902, 25.

spiniceps (Looss, 1896) Looss, 1901e. May 4, 629–631, fig. 7 (intest. of Bagrus bayad, B. docmac).—Braun, 1901a, 35, 36.—Odhn., 1902, 25.

vicinus Odhn., 1902, 24-25 (in Nilkrokodil; Sudan).

ACANTHOCOTILE Par. & Perugia, 1890, 11, 13, for Acanthocotyle.

ACANTHOCOTYLE Mont., 1888a, 7, 10, 11, 13, 16, 20, 30, 34, 35, 36, 37, 42, 43, 52, 55, 56, 57, 58, 59, 60, 66, 67 (Achantocotyle), 87 (m. lobianchi), 97 (in "subf." Tristomidæ, key); 1890, 189, 190, 208, figs. 1–5, pl. 8: 1891, 104, 106, 107, 111, 116; 1892, Oct. 7, 180, 213 (g. of Tristominæ): 1899a, 75–120, pls. 1–3: 1905, 71, 72, 73, 74, 75.—Braun, 1890a, 411, 412, 488, 523, 526, 529 (diagnosis); 1891d, 422; 1893a, 889.—Gamb., 1896a, 73.—Goto. 1899a, 283.—Massa, 1906, 45, 48.—Pratt, 1900a, 646, 649 (key), 655, fig. 10, 659.—Scott, 1902, 300, 301.

1888: Achantocotyle Mont., 1888, 67, misprint.

1900: Acanthocotile Par. & Perugia, 1890, 11, 13.

1902: Acanthetotyle Scott, 1902, 301, for Acanthocotyle.

concinna Scott, 1902, 301, no host given.

elegans Mont., 1890, 191, fig. 3 (on Raja clavata; Naples); 1899, 76, 77, 78, 79, 80, 81, 82, 85, 87, 88, 91, 93, 97, 99, 100, 101, 106, 107, 108, 110, 113-115, pl. 1, figs. 7, 8, pl. 2, figs. 13, 17, 19, 23, 25b, 28, 29, 32, pl. 3, figs. 44, 45; 1891, 107; 1899, 108; 1905, 73, 74, 75.—Braun, 1890a, 529, 547, 551.—Scott, 1902, 301.

lobianchi Mont., 1888a, 7, 13, 15, 16, 19, 22, 23, 26, 29, 31, 34, 56, 87 (on Raia clavata); 1890, 190; 1891, 102, 104 (lobianchii), 107, 116, 120, 130, pl. 5, figs. 9-10, pl. 6, figs. 36-39; 1893, 20: 1899, 108.—Braun, 1890a, 529, 547 (Naples), 551.

lobianchii Mont., 1891, 104, for lobianchi.

lobianchoi Goto, 1899a, 285, for lobianchi.

lobiancoi Mont., 1899, 75 (for lobianchi), 76, 77, 78, 80, 81, 83, 84, 85, 87, 89, 91, 94, 97, 100, 101, 106, 107, 108, 111-113, 116, 117, pl. 1, figs. 1, 3, 4, 5, 6, 10, pl. 2, figs. 11, 15, 16, 18, 21, 22, 25, 26, 31, 33, pl. 3, figs. 34-43, 46, 47, 49-58; 1905, 72, 73, 74.—Scott, 1902, 300.

monticelli Mont., 1905, 74, for monticellii.

monticellii Scott, 1902, 300–302, pl. 13, figs. 31–33 (in Raia clavata; Aberdeen, Scotland); 1905, 117.—Mont., 1905, 74, 75 (monticelli), figs. 4–5.

ACANTHOCOTYLE—Continued.

oligoterus Mont., 1899, 76, 77, 78, 80, 81, 84, 85, 87, 91, 100, 101, 105, 106, 107, 108, 111, 115–117, pl. 1, figs. 2, 9, pl. 2, figs. 12, 14, 20, 24, 25c, 27, 29, pl. 3, figs. 48 (on Raja clavata; Naples); 1905, 73, 74, 75 (in R. punctata).—Goto, 1899a, 283.—Scott, 1902, 301.

verrilli Goto, 1899a, 283–286, pl. 21, figs. 25–26 (on a skate, Raja; Cape Cod).—
Mont., 1905, 71–75.—Pratt, 1900a, 655, fig. 10, 657, 659 (key).—Scott, 1902, 302.—Staff., 1904, May 3, 482 (on Raja radiata Donovan; Canada).

ACANTHOCOTYLEA Dies., 1850a, 649, subtribe of Monocotylea.

ACANTHOCOTYLINÆ Mont., 1903, 335 (subf. of Tristomidæ).

ACANTHONCHOCOTYLE Cerf., 1899a, 347, 373, 402, 420, 436, 445, 446, 451, 455 [type by inclusion appendiculata].—Mont., 1903, 336 (syn. of Onchocotyle). appendiculata (Kuhn, 1829) Cerf., 1899a, 374, 379, 382, 461-462, pl. 19, figs. 6, 14a

(in Scyllium catulus Cuv.; Roscoff).—Type of Onchocotyle 1850.

canicula Cerf., 1899a, 374, 379, 382, 402, 462, pl. 19, figs. 5, 13, 14b.c. (syn. Onchocotyle appendiculata Kuhn of Stoss., 1887) (in Scyllium canicula).

ACANTHOPSOLUS Odhn., 1905, 328-331 (m. oculatus).

oculatus (Levin., 1881) Odhn., 1905, 328-331, pl. 2, fig. 11 (in Lycodes pallidus in East Greenland; Cottus scorpius, west coast of Sweden).

ACANTHOSTOMUM Looss, 1899b, 575, 577–579 (tod. spiniceps), 578, [not Acanthostoma Kriechbaumer, 1895, insect], ἡ ἄκαν βα, spine; 1900d, 603 (renamed Acanthochasmus); 1901e, 595.—Braun, 1901b, 34, 56; 1901i, 56.—Stiles, 1901, 189.

coronarium (Cobbold, 1861) Looss, 1899b, 578, 582.

spiniceps (Looss, 1896) Looss, 1899b, 578, 582.

ACANTHTCOTYLE Scott, 1902, 301, for Acanthocotyle.

ACCACELIUM Mont., 1893, 135 (contortum); subg. of Dist.—Raised to generic rank by Looss, 1899b, 631–632 (contortum as type); 1902, 839, 504.—Braun, 1901b, 27, 34, 37, 38.—Darr, 1902a, 698.—Luehe, 1900u, 487; 1901n, 481, 482–483, 485.—Odhn., 1905, 363.—Ofenheim, 1900, 183.—Pratt, 1902a, 889, 904 (key).

1893: Dist. (Accacelium) Mont., 1893, 135, q. v.

1894: Accacelum Zool. Record (1893) 1894, Vermes, 49.

calyptrocotyle (Mont., 1891) [Luehe, 1900u, 487].—To Orophocotyle in 1902.

contortum (Rud., 1819) Looss, 1899b, 632.—Darr, 1902a, 698.—Luehe, 1901n, 482, 483, 485.—Staff., 1904, 487 (Canada).

foliatum (Lint., 1898) Staff., 1904, 487 (Mola mola; Canada).

macrocotyle (Dies., 1858) Luehe, 1901n, 482.—Staff., 1904, 487 (Mola mola; Canada). nigroflavum (Rud., 1819) Luehe, 1901n, 482, 483.—Looss, 1902, 639.—Staff., 1904, 487 (Mola mola; Canada).

[pelagiæ (Kælliker, 1849), -

raynerianum (Nardo, 1827) Luehe, 1901n, 482, 483, 485.—Par., 1902, 6 (in Luyarus imperialis).

ACCACŒLUM Zool. Rec. (1893), 1894, Vermes, 49 (for Accacœlium).

ACERCÆ Mont., 1888, 94 (group of Cerc. without tail).

ACHANTOCOTYLE Mont., 1888, 67 (for Acanthocotyle).

ACOTYLEA Dies., 1850a, 286, 293 (subtribe of Dicranocœla), 287, 304 (tribe of Trematoda).—Brand., 1888a, 12.—Burm., 1856a, 250.—Goldb., 1855, 15, 16.—Mol., 1861, 191.—Mont., 1888, 83, 84; 1891, 111.—Tasch., 1879, 233.

ACOTYLOCEPHALA Dies., 1858e, 314, 374 (f. of Trematoda plectanophora).—Ben. & Hesse, 1864a, 119 (syn. of Gyrodactylidés).—Mont., 1888, 84.

ACRODACTYLA Staff., 1904, May 3, 491 (m. petalosa) [not Acrodactyla Hal., ante 1846, hymenopteron], ἄκρος, end; δάκτυλος, finger.

petalosa (Lander, 1902) Staff., 1904, May 3, 491 (m. Acipenser rubicundus Le S.; Canada); includes Dist. auriculatum Wedl. of Lint.

ACTINODACTYLELLA Haswell, 1893f, Sept., 153-158, pl. 16, corrigendum (m. blanchardi), Actinodactylus Haswell, 1893 [not Duchassaing, 1890], renamed; 1894a, 256; (18—), 589–590.—Pratt, 1900a, 646, 648 (key).—St.-Remy, 1898, 522, 531.

1893: Actinodactylus Haswell, 1893d, 477 [not Duchassaing, 1890] [type blanchardi].

1899: Actinodactynella Mont., 1899, 81, type haswelli=blanchardi.

ACTINODACTYLELLA—Continued.

blanchardi Haswell, 1893f, 153–158, pl. 16, corrigendum (Engæus fossor; Gippsland, Victoria, Australia).—Pratt, 1900a, 655, fig. 1, 657.—St.-Remy, 1898, 531–532, fig. 2 (on Eng. fos.; Australia) (syn. Actinodactylus sp.; Haswell).

ACTINODACTYLID. E Haswell.—St.-Remy, 1898, 522.

ACTINODACTYLUS Haswell, 1893d, 477 [not Duchassaing, 1890]; 1893f, 153–158, corrigendum, pl. 16, renamed Actinodactylella Haswell.—Mont., 1899, 116.

ACTINODACTYNELLA Mont., 1899, 81, 86, 116, 117, 118, 119, 122 (type haswelli Mont., 1898=blanchardi Haswell, 1893, see Actinodactylella.)

blanchardi Haswell, 1893, teste Mont., 1899, 122.

haswelli Mont. (1898), 1899, 122 (in Engeus fossor; Australia, Gippsland, Victoria). ACTINODACTYNELLINÆ Mont., 1899, 118, 121.—Pratt, 1900a, 646, 647 (key)

(apparently for Actinodactylellinæ), subf. of Temnocephalidæ.

ADENOGASTER Looss, 1901l, Nov. 7, 620–621 (m. serialis); 1902m, 570, 583, 591, 601, 603, 609, 612, 615.—Pratt, 1902a, 890, 909 (key).

serialis Looss, 19011, 620–621; 1902m, 545–548, pl. 26, figs. 71–74, 615, 875, pl. 26, figs. 71–74 (in Thalassochelys corticata; Alexandria, from Egyptian coast).

AGAMODISTOMA Rail., 1893a, 376 (for Agamodistomum, q. v.).

AGAMODISTOMUM Stoss., 1892, 4, 33–34 [collective group for immature forms, requires no type^a]; 1895, 33; 1895, 228–229; 1898, 58.—Mont., 1893, 154.—Stiles, 1898a, 28; 1904, 8, 11, 12, 13.—Stiles & Hass., 1898a, 82, 96; 1899, 117; 1900a, 559; 1902f, 360.

1892: Distomulum Brand., 1892b, Oct. 7, 510.

1893: Agamodistoma Rail., 1893a, 376.

anguis (Linst., 1885) Stoss., 1895, 229 (in Anguis fragilis).

capsulare (Dies., 1858) Stoss., 1892, 176 (in Ardea cinerea, A. purpurea, Crex pratensis, Nycticorax griseus, Podiceps auratus, P. nigricollis).

chimæræ Ariola, 1899a, 8–10, pl. 5, fig. 7 (in Chimæra monstrosa; Genova, Italy); (1899) v. 10 (in 129–138); (———), 299.

cælebs (Linst., 1875) Stoss., 1892, 176 (in Fringilla cælebs; Ratzeburg).

gobii Stoss., 1898, 58-59 (encysted in Gobius jozo; Triest).

martiranoi Stiles, 1903aa, 15 (in Anopheles claviger), based on Martirano, 1901d, 849–852, figs. 1–4 [see also Mart., 1901b, 1089–1091, figs. 1–4; 1901c, 274; 1902a, 73; 1902b, 358].

ophthalmobium (Dies., 1850) Stoss., 1892, 33 (in Homo).—Stiles, 1902s, 24, 29–34; 1903u, 223 (ophthalmothium); 1905cc, 54.—Ward, 1903, 866.

ophthalmothium Luehe in Stiles, 1903u, 223, for ophthalmobium.

pusillum (Braun, 1790) Stoss., 1892, 33 (in Erinaceus europæus; Greifswald; Rennes).

putorii (Mol., 1858) Stoss., 1892, 34 (in Putorius communis; Padova, Ratzeburg).
suis Stiles, 1898a, 22, 28, 29, 143, fig. 1 (in Sus scrofa domestica; Germany);
Dist. musculorum suis Duncker, 1896, 279, renamed.—Kastenbaum, 1899, (214), fig. 33, 4.—[See also Braun, 1893a, 870.—Duncker, 1881, 23-25; 1881, 141; 1881, 154; 1884, 39-42; 1896, 279-282,—Leuck., 1881, 46.—Moulé, 1885, 60-62.—Schndmhl., 1896, 302.—Stiles & Hass., 1900a, 559-560, fig. 23; 1902i, 360; 1904, 22.]

valdeinflatum (Stoss., 1883) Stoss., 1898, 59 (in Gobius jozo; Triest); "Rappresenta la larva dell' Echinostoma cesticillus Molin."—Barbagallo & Drago,

1903, 411 (in G. j.; Cantania).

viperæ (Linst., 1877) Stoss., 1895, 228 (in Pelias berus).

This name being French, has no status in nomenclature.

"ALAIRE" b Blainv., 1824a, 518,b see "Alaria Blainv." of Dies., 1850a.—Leblond, 1837a, 518, 59, pl. 14, fig. 15 (teste Dies., 1850a, 374).

"ALARIA Blainv." of Dies., 1850a, 331 (for "alaire" [q. v.] Blainv., 1824a, 518, see Duj., 1845a, 437). See also Alaria Schrank.

^a See p. 385.

b"Je propose de rétablir ce genre pour quelques espèces de fascioles cylindriques, avec une assez longue queue, et qui ont des espèces d'ailes de chaque côté du corps; j'en ai décrit une trouvée dans le pancréas du Simia maimon de Linné, à l'art. 'fasciole' du Dictionn. des sciences nat.''

ALARIA Schrank, 1788, 52 (m. vulpis) [not Alaria Dunc., 1841. lepidopteron; Morr. Lyc., 1850, mollusk].—Abildg., 1790, 63 (syn. of Dist. vulpina).—Brand., 1888a, 8.—Braun, 1893a, 884, 894, 902.—Dies., 1850a, 307 (syn. of Hemist.).—Encycl. méth., Par., v. 2, 20.—Knoch, 1862, 30.—Lamouroux, 1822a, 194. See also genera for which alata Gozze is type.

vulpis Schrank, 1788, 52 (Planaria alata vel dubia Gœze, 1782a, 176–177, pl. 14, figs. 11–13, from Canis vulpes. renamed).—Brand., 1888a, 60 (syn. of Hemist. alatum).—Dies., 1850a, 308 (syn. of Hemist. alatum; includes also alata Gœze as syn.).—Lamouroux, 1822a, 194.—Rud., 1809a, 402 (syn. of Dist.

alatum Zed.).

ALLEGREADIINE Engler, 1904, 188, for Allocreadiine.

ALLOCREADIIDÆ Stoss., 1904, 199.

ALLOCREADIINÆ Looss, 1902m, 841.—Odhn., 1905, 323, 326, 328.—Stoss., 1902, 578; 1903, 373.

ALLOCREADIUM Looss, 1900d, Dec. 3, 602 (Creadium Looss, 1899 [not Creadium and Creadion Vieill., 1816, bird], renamed, hence type isoporum; ἄλλος, another; 1901b, 199; 1902m, 760, 785, 839.—Braun, 1902b, 26, 147, 148.—Odhn., 1901, 483–520; 1905, 328.—Pratt, 1902a, 888, 897 (key).—Stoss., 1901, 95 (7); 1902, 578; 1903, 373; 1904, 199.

album (Stoss., 1890) Stoss., 1901, 95 (7); 1904, 199 (type of Lepocreadium).

angusticolle (Hausmann, 1896) Odhn., 1901, 517.—Stoss., 1901, 96 (S).

asymphyloporum Stoss., 1901, 96 (in Trutta trutta; Lake Plitvica, Croazia).

atomon (Rud., 1802) Odhn., 1901, 506–513, 516, pl. 33, figs. 9, 10; 1905, 320 (type of Podocotyle).—Stoss., 1902, 582.

commune (Olss., 1876) Odhn., 1901, 499–503, 508, 516, 517, pl. 33, fig. 6; 1905, 327.—Stoss., 1902, 582.

fasciatum (Rud., 1819) Odhn., 1901, 485–490, 492, 496, 516, pl. 33, fig. 1; 1902, 160.
genu (Rud., 1819) Odhn., 1901, 485, 496–499, 500, 501, 502, 517, pl. 33, figs. 3–5; 1905, 327.

isoporum (Looss, 1894) Looss, [1900d, 602:] 1902m, 785.—Kowal., 1902d, 27 (9).—Odhn., 1901, 505, 506, 516; 1905, 327.—Staff., 1904, May 3, 493 (in Semotilus bullaris Raf.; Canada).—Stoss., 1901, 93, 96 (in Gobius fluviatilis; Loitsch in Carniola).

labracis (Duj., 1845) Odhn., 1901, 514-516, pl. 33, fig. 11; 1905, 327.

labri (Stoss., 1886) Odhn., 1901, 490, 493–496, 516, pl. 33, fig. 2; 1902, 160.

obovatum (Mol., 1859) Stoss., 1901, 95–96 (7–8) (in Chrysophrys aurata; Triest). pegorchis Stoss., 1901, 94–95 (6–7), pl. 6, fig. 4 (Mæna smaris; Triest); 1904, 199. sinuatum (Rud., 1819) Odhn., 1901, 490–492, 516; 1902, 160.

transversale (Rud., 1802) Odhn., 1901, 505-506, 516; 1905, 327.

tumidulum (Rud., 1819) Odhn., 1901, 503–505, 516, pl. 33, figs. 7–8.

umbrinæ (Stoss., 1885) Odhn., 1905, 327.

AMFISTOMIDI Sons., 1895, see Amphistomidæ.

AMPHIBDELLA Chatin. 1874a, 11–16 (m. torpedinis); 1875, —.—Braun, 1890a, 412, 417, 444, 468, 484, 512, 519, 523, 542, 545 (diagnosis; m. torpedinis).— Maclaren, 1904, 583, 598, 599, 600.—Mont., 1889, 116; 1903, 336 (syn. of Tetraonchus).—Pratt, 1900a, 646, 654 (key), 657, fig. 47.—St.-Remy, 1898, 524.—Stoss., 1898, 17.

torpedinis Chatin, 1874a, 11–16. pl. 2 (IX), figs. 13–14 (in Torpedo marmorata; Mediterranean).—Brand., 1894a, 307.—Braun, 1890a, 418, 426, 444, 452, 545, 549, 552 (Genoa; Triest).—Maclaren, 1904, 587.—Mont., 1888, 93, 110; 1889, 116 (belongs to Gyrodactylidæ, Tetraonchus); 1890, 193–195 (on Torpedo narce, T. mar.); 1905. 79.—Par., 1894, 138, 578.—Par. & Perugia, 1890, 8; 1890, 363–367; 1890, Sept. 5, 335–336.—Pratt, 1900a, 657, fig. 47.—Stoss., 1898, 17–18 (on T. mar.; Triest).

AMPHIBDELLIDÆ Carus, 1885a, 121 (contains Amphibdella Chatin).—Mont., 1888, 93; 1889, 116 (4) (belongs to Gyrodactylidæ).

AMPHIBOTHRIUM Frey & Leuck., 1847, 147 (m. krœyeri)=Amphibothrium Leuck., 1847, see Braun, 1889a, 343).—Ben., 1858a, 1861a, 12.—Braun, 1890a, 518 (syn. of Udonella, 1855).—Dies., 1850a, 427 (syn. of Udonella Johnston).

kræyeri Leuck., 1847, 147-148, pl. 2, fig. 2 (on Caligus sp. parasitic on Gadus).

AMPHIBOTHRIUM—Continued.

kroyeri Dies., 1850a, 427 for krœyeri (syn. of Udonella caligorum Johnston) (in Caligorum corporis superficie, frequens, in Caligo hippoglossi vulgaris, April (Johnston); C. triglæ Gurnardi, June (Hyndman); C. curto (Kroyer).—Ben., 1858a, 1861a, 13 (syn. of Ud. cal. Johnst.).—Tasch., 1878, 573 (syn. of U. cal. F. & L.).

AMPHIOSTOMA Rud., 1819a, 88 for Amphistoma.

AMPHIOSTOMA Rud., 1819a, 88 for Amphistoma.

AMPHISTOMA ab Rud., 1801a, 50-51, 54 (Strigea Abildg., 1790, renamed, hence type strigis=macrocephalum); 1802b, 92-93; 1809a, 5, 21, 37, 340, pl. 5, figs. 4-7 [a, capite discreto, 6 species in birds; b, capite continuo, 3 species in birds, amphibia, and mammals]; 1819a, 87, 351-352, 589.—Baillet, 1866b, 99, 105-106.—Bellingham, 1844a, 338.—Ben. & Hesse, 1864, 61.—Blainv., 1828a, 582-583.—E. Bl., 1847a, 309-310.—R. Bl., 1886m, 841; 1888a, 543, 632, 636; 1895, 730.—Brand., 1888a, 9, 10, 13; 1891d, 12, 20.—Braun, 1883a, 69, 70; 1889i, 437; 1890a, 514, 515; 1892a, 568, 635, 650, 667, 681, 696, 698, 699, 707, 709, 710, 715, 720, 722, 738; 1892i, 49, 50, 51; 1893a, 851, 852, 872, 879, 880, 883, 884, 886, 890, 892, 893, 895, 902, 903, 904, 905, 906, 918; 1893i, 383; 1894a, 1147; 1895b, 134, 136, 137; 1898a, 1583; 1901b, 20, 55, 56 (sp. Bellingham).—Bremser, 1824, 132-133.—Burckhardt, 1891a, 63.—Burm., 1837a, 530; 1856a, 250.—Carus, 1863, 479.—Cobbold, 1875n, Nov., 817-821.—Cohn, 1903, in 35-42.—Crep., 1825a, 35-38; 1837a, 309, 313; 1839a, 286; 1847c, 30-35, pl. 2, figs. 1-5 (2 sp. in Zebu).—Cuv., 1817, 41.—Dav., 1877a, lxxx.—Deslongchamps, 1824c, 53-57.—Dies., 1836d, 238-246 (monograph); 1839a; 1850a, 288, 307 (sub Hemist.), 312 (sub Holost.), 317 (sub Codonocephalus), 318 (sub Diplodiscus), 319 (sub Monost.), 400-407, 573 (sub Tetrabothriorhynchus); 1858e, 312, 322 (of Bellingham, 1844a, 339) (syn. of Holost. falconum), 357-358.—Duj., 1845a, 327-331.—Eichwald, 1829a, 248.—Fil., 1837a, 334, 336.—Fischer, 1840a, 156.—Fischer., 1901a, 367-375 (revision with several new genera); 1902a, 6, 7, 10 (Strigea Abildg., renamed), 488, 489, 490 (type strigis=macrocephalum) (of mammals, revision).—Gamble, 1896a, 73.—Goldb, 1855a, 17.—Goubaux, 1863a, 882-exison).—Gamble, 1896a, 73.—Goldb, 1855 Abildg., renamed), 488, 489, 490 (type strigis=macrocephalum) (of mammals, revision).—Gamble, 1896a, 73.—Goldb., 1855a, 17.—Goubaux, 1863a, 882-884.—Gurlt, 1831a, 369.—Hahn & Lefevre, 1884, 806.—L'Herminier, 1826, 10.—Hoyle, 1890, 535, 539.—Jackson, 1888, 642, 644, 654.—Joy, 1835a, 504.— 10.—Hoyle, 1890, 535, 539.—Jackson, 1888, 642, 644, 654.—Joy, 1835a, 504.—Kath., 1894a, 130.—Kholodk., 1898a, 33; 1899a, 153.—Kitt, 1885a, 148.—Lamarck, 1816b, 187-188.—Lamouroux, 1822a, 297.—Leblond, 1836f, 4.—Lejtenyi, 1881a, 3, 9, 11.—Leuck., 1863, 451, 459, 462, 463, 524.—Looss, 1892a, 126; 1894a, 10, 20, 178, 249; 1896b, 173-177; 1901, 622; 1902m, 430, 438, 439, 638, 676, 746, 780 (Paramphist.).—Moniez, 1896, 87.—Mont., 1888a, 7, 12, 30, 46, 47, 52, 56, 63, 64, 71, 83 (Amphist.), 84, 90, 91, 103; 1892, 38; 1892, Oct. 7, 196, 197, 214; 1892, 700, 710; 1893, 27, 108, 208.—Moul., 1856a, 11, 12, 15.—Neumann, 1892, 345.—Nitzsch, 1819, 398-401.—Nord., 1840, 544, 614, 625-627, 628-629.—Olfers, 1816, 22, 47.—Otto, 1896, 78 pp., 30 figs.; 1896, 85-141, 275-296, figs. 1-30 (anat., histology).—Pag., 1857, 52.—Poir., 1883, 79.—Pratt, 1900a, 645.—Rafinesque, 1815, 151 (genus of Filaridia).—Rail., 1892, July 15, 633-634 (of domestic animals in Tonkin); 1893, April 15, 245-246; 1893a, 376 (Strigea).—Rousseau, 1833-34, 149-150.—Schndmhl. 245-246; 1893a, 376 (Strigea).—Rousseau, 1833-34, 149-150.—Schndmhl., 1896, 295, 303.—Schneider, 1866, 334.—Shipley, 1905, 7 (syn. of Strigea), 8 (type Holost. macrocephalum).—Shipley & Hornell, 1904, 80.—Sons., 1895, 181, 183, 184, 185, 186.—Stiles, 1898a, 24, 64.—Tasch., 1879, 232, 233, 256.—Vaullegeard, 1899, 53.—Vogt, 1878, 9, 10.—Wallenstedt, 1847, 7.—Ward, 1895, 256, 338 (sp. Cobb. in stomach of Equus caballus).—Westrumb, 1821, 46; 1823, 390–398, pl. 5, figs. 1–2. 1803: Amphystoma Rud., 1803a, 29 (misprint).

1819: Amphistomum Nitzsch, 1819, 397-401 (emendation).—Crep., 1825, 35; 1837, 309, 313.

1819: Amphiostoma Rud., 1819a, 88 (misprint).

1822: Amphistome Lamouroux, 1822a, 297 (French word or misprint).

a Rud. deliberately renamed a previously validly named genus, namely Strigea Abildgaard, 1790, referring clearly to this fact both in 1801a, 50–51, and 1802b, 92. He makes but one combination (Amphist. subclavatum), but since Amphist. is clearly a new name proposed for an older one (Strigea), which Rud. changed on the alleged ground that it was inappropriate, Amphist, should be suppressed in favor of Strigea and take the same species as type. b Since 1816 written almost indiscriminately Amphistoma or Amphistomum.

1888: Amphistomun Mont.. 1888, 83 (misprint).

anatis Rud., 1819a, 793, for anatis querquedulæ 1819a, 92.

anatis querquedulæ Rud., 1819a, 92 in Anas querquedula; C. E. V.).—Dies., 1836d, 253.—Westrumb, 1823, 398.

anatis tadornæ (Viborg. 1795) Rud., 1809a, 352; 1814a, 100 (=A. isostomum).—Dies., 1850a, 313 (syn. of Holost. erraticum Duj.).

asperum Dies., 1838a, 189 in Tapirus americanus; 1839a, 236, pl. 20, figs. 14–16 (in Tap. amer.; South America [Matogrosso and Cachoeira do Bananeira]); 1850a, 402.—Braun, 1892a, 580; 1893a, 894, 905; 1893d, 466.—Cobbold, 1875n, 819; 1879b, 402.—Duj., 1845a, 334.—Fischder., 1902a, 39 (syn. of Cladorchis asper).—Nord., 1840, 629.—Sons., 1895, 184.

attenuatum Dies., 1836d, 238, 252, pl. 24, figs. 9-12 (in Salmo paccu: Caiçara): 1850a, 406-407 (in Miletes bidens: Brazil).—Cobbold, 1860a, 54 (in M. bidens; Brazil).—Duj., 1845a, 341 (in Salmo pacu: Brazil).—MacCallum, 1905, 668,

677 (in Salmo).—Nord.. 1840, 629.

bothriophoron Braun, 1892f, 49, 50, 51 (in Bos indicus: Madagascar): 1893a, 738; 1893b, 185; 1893f, 383; 1901b, 20.—Fischder., 1903h, 538 (to Paramphist.).—Otto, R., 1896, 101–103, fig. 5.—Sons., 1895, 184.—Spengel, 1892.

bothriophorum Stiles. 1898a. 24, 67. fig. 56 (read bothriophoron).

brachycælium Cohn, 1903. 39 (host not given) ? lapsus

cervi Schrank, 1790) Stiles, 1898a, 24, 64, 65, 66, 67, 139, 140, 141, 142, figs, 49-55
 (Bos taurus).—Fischder., 1901a, 368 (type of Paramphist.).—Stiles & Hass., 1900a, 611, fig. 29; 1902f, 360; 1904c, 22.

cheloniæ imbricatæ Dies., 1858e. 358 (based on Bellingham. 1844a, 340. in Chelonia imbricata; Ireland).—Braun. M.. 1893a, 905.

chordale Burckhardt. 1891a. 21 Apr., 62-64 (Protopterus annectens); 1892a. 344-345 (a Tetracotyle, Holostomidæ, teste Brandes).

clavatum Steenstrup, 1842, 59 (in Rana temporaria); 1842, 109–110 [for subclavatum?].—Sieb., 1843, lx.

clavigerum Zed., 1803a, 199 [Fest. strigis Schrank. 1788, renamed] (in Strix).—Rud., 1809a, 341 (syn. of A. macrocephalum).

collinsi Sons.. 1895, 182 (for collinsii), 187 to (Pseudodiscus).—Fischder.. 1902a, 48 (in Equus caballus: India).

collinsii Cobbold, 18751, 741 (in Equus caballus); 1875n, 818, 819; 1879b, 357, 398; 1883x, 515.—Fischder., 1903h, 489 [to Pseudodiscus by Sons.).—Huber, 1896a, 580 (India).—Sons., 1895, 182, fig. 2.—Theobald, 1900, 51.

collinsii var. stanleyi Cobbold. 1879b. 357 for stanleyii 1875 (in Equus).—Fischder., 1902a, 48.—Piana & Stazzi, 1900, 519 [= A. hawkesi Cobbold].—Sons., 1895, 182.

conicum (Zed., 1803) Rud., 1809a, 349-352 (Bos taurus, Greifswald; Cervus elaphus), 356; 1819a, 91 (Ovis aries, Cervus dama), 360, 577, 589, 793.—Anacker, 1892c, 94.—Baillet, 1866b, 90, 106.—Bénion, 1874a, 628.—Bettendorf, 1897a, 7, 38; 1897, 311, 342.—E. Bl., 1847a, 310-316.—R. Bl., 1886, 295; 1888a, 584, 585.—Blumberg, 1871a, 40 pp., 1 pl. (anat.); ——, 496-499; 1872, v. 33, 190-191; ——, v. 8, 454.—Brand., 1891d, 7, 12, 13, 14, 15, 18, 20; 1898a, 222 (30).—Braun, 1892, 49, 50; 1892a, 576, 584, 587, 589, 591, 593, 595, 596, 597, 601, 603, 609, 613, 614, 615, 621, 622, 628, 632, 638, 640, 641, 642, 645, 646, 648, 654, 660, 661, 662, 664, 666, 667, 669, 671, 675, 676, 677, 681, 682, 683, 685, 688, 692, 693, 695, 703, 705, 711, 712, 713, 717, 718, 719, 724, 729, 730, 731, 733, 738, 739, 748, 762, 766; 1893a, 874, 879, 881, 905; 1893b, 186; 1893d, 466,—Cobb, 1891b, 614-615, 1 fig.—Cobbold, 1858b, 159; 1859, 51; 1875n, 818; 1879b, 331; 1884g, 976.—Conyngham, 1904, Sept. 17, 663.—Crep., 1837a, 311, 313, 315, 317, 321, 322, 323, 327; 1839a, 286; 1841, 80; 1847, 30.—Darr, 1902, 688.—Daubenton, ——, 250, pl. 16, fig. 3 (Bos taurus).—Dav., 1877a, lxxx, 234, 235.—Dies, 1835, 246; 1836d, 238, 239, 240, 243, 244, 245, 246-248, pl. 23, figs. 1-4 (Bos taurus dom., Ovis aries, Cervus elaphus, Cervus capreolus, Cervus dama, Cervus campestris, Cervus dichotomus, Cervus nambi, Cervus rufus, and Cervus simplicicornis; Brazil); 1850a, 401 (in Bos taurus indicus, Berlin; B. urus, Vilna; Capra hircus var.; Antilope dorcas, Berlin; Cervus capreolus; C. alces) (syns.; Fasc, hepatica Mueller; F. cervi Schrank; F. elaphi Gmelin; Festuc, cervi Zed., Monost, conicum Zed.); 1850a, 401.—Duj., 1845a, 331, 332-333.—Eichw., 1829a, 248.—Falk, ——, 6-7.—Fischer,

1840, 157; 1883a, 20.—Fischder, 1901, 368, 375; 1902a, 11 (syn. of Paramphist. cervi). 13 (syn. of P. liorchis), 41, 50 (syn. of Balanorchis anastrophus); 1903h, 485 (in Bos taurus; Kænigsberg i. Pr.), 504–505 (syn. of P. cervi). 506, 508 (of Dies., 1835, 247, in Cervus dichotomus in Brazil as syn. of Bal. anast.), 508 (of Dies., 1835, 247, in Cervus simplicicornis, C. campestris, C. mexicanus, C. rufus, C. dichotomus, C. namby in Brazil as syn. of P. liorchis), 509 (of Gurlt, 1831, 369 in Bos taurus indicus as syn. of P. dicranocœlium), 515 (of Gurlf, 1831, 369 in Bos taurus indicus as syn. of P. dicranocceium), 519 (or Dies., 1835, 247, in South American Cervus simplicicornis, campestris, mexicanus, rufus, dichotomus, namby, as syn. of P. liorchis), 528 (in B. tau. ind., as syn. of P. dicranoccelium), 535 (in Antilope dorcas as syn. of P. microbothrium), 584 (syn. of Gastrothylax mancupatus); 1904, 459.—Giard & Billet, 1892a, 615.—Giebel. 1857, 266.—Gronkowski, 1902a, 511, 514, 515, 517–518, 519, 520, 523, 529–531, 532, 533, fig. A (4, 7, 8, 10–11, 12, 13, 16, 22–24, 25, 26, pl. 13, figs. 2, 6, 7).—Gurlt, 1831, 156, 369–370, pl. 8, figs. 25–28.—Jancon 1892e, 261, 1897a, 103.—Kastenbaum, 1899 (244), fig. 33, 6.—Kath. son, 1893c, 261; 1897a, 103.—Kastenbaum, 1899 (244), fig. 33, 6.—Kath., 1894a, 131, 143.—Kerbert, 1881a, 548, 551.—Kitt, 1885a, 148.—Lamarck, 1816b, 189.—Lamouroux, 1822a, 297.—Laurer, J. F., 1830a, 20 pp., 1 pl.; 18160, 189.—Lamouroux, 1822a, 297.—Laurer, J. F., 1830a, 20 pp., 1 pl.; 1830, 3, 4, figs. 1–14.—Lejtenyi, 1881a, 3, 5, 6, 7, 8, 11, 16, 18, 20.—R. Leuck., 1863, 479, fig. 157; 1876, 869.—Linst., 1883, 310.—Looss, 1885b, 6, 17; 1894a, 124, 142, 146, 206, 237; 1894d, 17, 22, 24; 1896b, 5, 23, 31, 32–33, 170, 173, 176, 178, 185–191, pl. 12, figs. 125–134 (syn. Fest. cervi) (in Physa alexandrina; P. micropleura; buffles; Egypt); 1898a, 459; 1902m, 438, 444 (Cerc. pigmentata Sons.), 638.—Macé, 1882, 9, 30, 80, 81, 85.—Mégnin, 1882, 456.—Mingazzini, 1899a, 10 pp., figs. 1–5.—Miram, 1840, 157.—Moul., 1856a, 18.—Mueh., 1898, 20.—Neumann, 1888, 336, fig. 124; 1892, 353, fig. 140; 1892, 363, fig. 140.—Nitzsch, 1819, 398 (syn. Monost. conicum Zed.).—Nord., 1832a, 38, 92, 93, 97; 1840, 547; 1840, 627 (syns. Fasc. elaphi, Gmelin; Monost. 363, fig. 140.—Nitzsch, 1819, 398 (syn. Monost. conicum Zed.).—Nord., 1832a, 38, 92, 93, 97; 1840, 547; 1840, 627 (syns. Fasc. elaphi Gmelin; Monost. conicum Zed.).—Olfers, 1816, 47.—Ostertag, 1899, 412.—Otto, 1896, 97–101. fig. 4, figs. 10, 13.—Poir., 1885, 1, 3, 101, 102.—Rail., 1886, 301, fig. 195; 1890, 143; 1892, 633; 1893a, 376–377 (synonymy), fig. 249.—Rail. & Gomy, 1899, 348, 349.—Rubridge, 1892, 61.—Schmalz, 1831, 24.—Schneider, 1866, pl. 28. fig. 4.—Siebold, 1835, 56, 57, 58, 59, 62, 65.—Slawikowski, 1819, 52.—Sons., 1895, 181, 184, 185; 1896, 314; 1897, 252.—Staff., 1905, April 11, 693 (syn. of Paramphist. cervi Zed.).—Stiles, 1898a, 64.—Tasch., 1878, 176.—Trollip, 1893, 6 (22), 2. Nov., 424–425 (in ox).—Verrill, 1870, 176, 177, 220.—Veterinarius, v. 21, 328.—Villot, 1878, 16.—Volz, 1899, 232.—Ward, 1895, 256 (syns. Fest. cervi Zed.; Fasc. elaphi Gmelin: Monost. conicum Zed.); 1895, 332 (in Bos taurus). 335 (in Ovis aries), 338 (in Equus caballus).—Westrumb, 1823, 396, 397.—Zeit. f. Fleisch-u. Milchhyg., July. 1894, 200.—Ziegler, 1883, 546.—Zuern, 1882, 220, 221, pl. 4; fig. 10.—Also reported for Capreolus caprea, Cariacus paludosus. caprea, Cariacus paludosus.

cornu (Zed., 1800) Rud., 1809a, 346–347 (Ardea cinerea; Europe); 1819a, 85, 345, 793 (to Monost.).—Brand., 1888a, 10.—Dies., 1850a, 327 (to Monost.).
 Duj., 1845a, 349 (to Monost.).—Lamarck, 1816b, 189.—Nord., 1840, 626–627.—Olfers, 1816, 48.

cornu (Nitzsch, 1819) Rud., 1819a, 89–90, 357, 793 (Ardea cinerea, A. garzetta;
Europe) [nec (Zed., 1800) Rud., 1809].—Baird, 1853a, 47.—Bellingham, 1844,
339.—Dies., 1850a, 315 (to Holost.); 1858e, 321.—Stoss., 1898, 22 (to Holost.).—
Westrumb, 1823, 394–395.

cornu Dies., 1839a, 235, pl. 20, figs. 12–13 (Dorcas n. sp.=Cataphractus vacu Natt. Cat.; Forte do Rio Branco, Brazil) [nec (Zed., 1800) Rud., 1809a; nec (Nitzsch 1819) Rud., 1819]; 1850a, 402.—Braun, 1893a, 905.—Cobbold, 1860a, 53 (renamed nattereri).—Duj., 1845a, 340.—MacCallum, 1905, 668.

cornutum Rud., 1808a, xxv. pl. 4, figs. 4–7; 1809a, 343–344 (in Charadrius pluvialis, intestine; Greifswald); 1819a, 90, 589, 793.—Dies., 1850a, 317 (to Holost.).—Duj., 1845a, 372 (to Holost.).—Lamarck, 1816, 188.—Lamouroux, 1822a, 297.—Nord., 1840, 626.—Olfers, 1816, 47.—Risso, 1826, 262.—Westrumb, 1823, 395.

Tumeniferum Crep., 1847, 30–34, pl. 2, figs. 1–5 (in Bos taurus indicus, rumen).—

**Baillet, 1866b, 106.—Brand., 1891d, 17; 1898a, 196, 216 (4, 24).—Braun, 1892a, 576, 738, 739; 1892, 49, 51.—Cobbold, 1875m, 819; 1879b, 332.—Dav., 1877a., lxxx.—Dies., 1850a, 402–403.—Fischder., 1902a, 27 (to Gastrothylax); 1903h, 488, 557 (type of Gastrothylax), 558, 559, 575; 1904, 459.—Giard & Billet, 1892a, 615.—Neumann, 1892, 363.—Poir., 1883, 76 (to Gastrothylax).—Sons., 1895, 185; 1896, 298.—Stiles, 1898a, 67.—Verrill, 1870, 177, 220.—Ward, 1895, 332 (in Bos taurus).

crumigerum Fischder., 1903h, 563 (syn. of Gastrothylax compressus Brand.).

cylindricum Dies., 1836d, 249, pl. 23, figs. 13–15 (in Cataphractus murica; at Villa Maria, Brazil); 1850a, 405 (in Doras murica; Brazil).—Braun, 1893a, 905.—Duj., 1845a, 340.—MacCallum, 1905, 668, 672, 673 (in Callichthys).—Nord., 1840, 628.

denticulatum Rud., 1819a, 90, 358 (in Alcedo ispida; Mus. Vien.), 793.—Dies., 1850a, 311 (to Hemist.).—Duj., 1845a, 372 (to Holost.).—Lamouroux, 1822a, 297.—Villot, 1898, 538.—Westrumb, 1823, 395–396.

dolichocotyle Cohn, 1903, May 30, 37–39, fig. 3 (in Herpetodryas fuscus); 1904, 242.— Type of Catadiscus, 1904.

elongatum (Poir., 1883) Fischder., 1903h, 577 (to Gastrothylax) (in Anoa depressicornis; Berlin Zool. Garden).

emarginatum Dies., 1839a, 237 (in Callithrix noctivaga: at Matogrosso, Brazil); 1850a, 407 (in Cebus trivirgatus), ex Dies., 1839.—Braun, 1893a, 905; 1901e, 311.—Cobbold, 1879b, 289.—Duj., 1845a, 331 (ex Dies., 1839).—Fischder., 1902a, 49 (in Nictipithecus trivirgatus: Brazil).—Nord., 1840, 629.

emberizæ citrinellæ Dies., 1836d, 253.

erraticum Rud., 1808a, 458 (nomen nudum); 1809a, 344–345 (in Larus septentrionalis; Greifswald, Europe); 1819a, 89 (in Scolopax gallinago, Greifswald; Colymbus arcticus, Scolopax rusticola), 356, 793.—Baird, 1853a, 47 (to Holost.).—Crep., 1839, 288.—Dies., 1850a, 313 (to Holost.).—Duj., 1845a, 373 (to Holost.).—Erc., 1881e, 86 (eggs); 1882a, 322.—Lamarck, 1816b, 189.—Lamouroux, 1822a, 297.—Nord., 1840, 626.—Olfers, 1816, 47.—Westrumb, 1823, 393–394.

excavatum (Rud., 1803) Nitzsch, 1819, 399 to (Holost.).

explanatum Crep., 1847, 34–35 (in Bos taurus indicus, liver and gall bladder; Berlin).—Baillet, 1866b, 106.—Braun, 1892a, 738; 1893a, 875, 905; 1893d, 467 (in Bos taurus indicus).—Cobbold, 1875n, 819; 1879b, 332.—Dav., 1877a, lxxx.—Dies., 1850a, 404–405 (Berlin).—Fischder., 1902a, 34 (in Bos taurus ind.; Calcutta); 1904, 454, 459.—Giard, & Billet 1892a, 615.—Gomy, 1897a, 377.—Rail. & Gomy, 1897, 2 July, 610–613 (in cattle; Cochin China); 1897, 1 Aug., 474–475; 1899, 347, 349.—Sons., 1895, 184.—Stiles, 1898a, 24, 67, 140.—Verrill, 1870, 177.—Veterinarius, v. 21, 328.—Ward, 1895, 332 (in Bos taurus).

fabaceum Dies., 1838, 189 (in Manatus australis); 1839a, 236, pl. 20, figs. 19–23 (in Manatus exunguis; Borba and Forte do Rio Branco, Brazil); 1850a, 403–404 (in Man. exung.; Brazil).—Braun, 1893a, 905.—Cobbold, 1879b, 429.—Duj., 1845a, 334.—Fischder., 1901, 374 (type of Chiorchis n. g.); 1902a, 44, (to Chiorchis); 1903h, 621, 622 (in Manatus latirostris; North America).—Leidy, 1891a, 413–414 (in Manatus latirostris).—Nord., 1840, 629.—Stedman, 1889, v. 11, 85–101, pls. 1–2.

falconis palumbæ Baird, 1853a, 47 (syn. of Holost. macrocephalum).

falconis palumbarii Rud., 1819a, 88 (syn. of A. macrocephalum), 793.—Dies., 1850a, 309 (syn. of Hemist. spathula).

falconis palumbi (Viborg, 1795) Rud., 1809a, 352, sp. inq.

falconis peregrini Rud., 1819a, 92, 362, sp. dub. (Falco peregrinus; Berlin), 793.—Dies., 1836d, 253.—Westrumb, 1823, 397.

ferrum-equinum Dies., 1836d, 238, 250, pl. 23, figs. 16–18 (Cataphractus murica at Cuyaba and C. corome at Cuyaba and Matogrosso); 1850a, 405 (Doras muricas, D. costatus).—Braun, 1893a, 905.—Duj., 1845a, 340–341.—MacCallum, 1905, 668.—Nord., 1840, 629.

giganteum Dies., 1836d, 238, 239, 240, 243, 244, 245, 248, pl. 22, figs. 5–6 (Dicotyles albirostris at Nas Frechas and Caiçara, and D. torquatus at Matogrosso); 1850a, 403.—Brand., 1891d, 17.—Braun, 1892a, 578, 601; 1893a, 874, 905; 1893d, 466 (in Dic. labiatus; D. torq.).—Cobbold, 1875n, 819; 1879b, 404.—Crep., 1841, 80 (in Dic. lab., D. torq.).—Duj., 1845a, 333–334.—Fischder., 1901, 373; 1902a, 40 (e. p. syn. of Cladorchis (Taxorchis) schistocotyle).—Moul., 1856a, 19.—Nord., 1840, 547, 628.—Sons., 1895, 184.

gigantocotyle Brand., in Otto, 1896, 16 Apr., 103–105, figs. 6–7 (in Hippopotamus).—Fischder., 1902a, 9, 34 (in Hip. amphibius; Africa); 1903h, 495, 496.

gracile Rud., 1819a, 89, 355 (Mergus merganser and M. albellus), 793.—Bellingham, 1844, 339.—Dies., 1850a, 315 (to Holost.); 1858e, 321.—Duj., 1845a, 578 (to Holost.).—Lamouroux, 1822a, 297.—Westrumb, 1823, 393.

 grande Dies., 1839a, 237, pl. 20, figs. 25–27 (in Chelys, Phrynops, Peltocephalus,
 Podocnemis, Rhinemys; South America); 1850a, 406 (in Chelys fimbriata,
 Phrynops geoffroanus, P. miliusii, P. gibbus, Peltocephalus dumerilianus, Podocnemis erythrocephala, P. expansa, P. tracaxa, Rhinemys nasuta; Brazil).—Brand., 1891d, 17.—Braun, 1892a, 586; 1893a, 879, 905; 1899b, 715, 719; 1901b, 55.—Crep., 1844a, 114.—Duj., 1845a, 336.—Leidy, 1888, 127.—Looss, 1902m, 430, 437, 440.—Mont., 1892, 715; 1896, 165.—Nord., 1840, 629.

haukesi Sons., 1895, 182; 1895, 187, fig. (to Pseudodiscus); 1896, 310.

hawkesi Braun, 1893d, 466, for hawkesii Cobbold.

hawkesii Cobbold, 1875n, 818, 819 (in Elephas indicus; India); 1877, 234; 1879b, 393, 396, 399; 1882, 238–240, fig. 8; 1883x, 515.—Braun, 1893a, 874, 905; 1893d, 466 (hawkesi).—Fischder., 1902a, 48 (hawkesi) (in E. indicus, India); 1903h, 489 (to Pseudodiscus by Sons., 1895).—Galli-Valerio, 1901c, 364 (elephant).—Huber, 1896a, 579–580 (elephant).—Looss, 1902m, 439 (hawkesi).—Mégnin, 1882, 455.—Piana & Stazzi, 1900, 511, 519-525, 529, figs. 12-14 (hawkesi); 1901, 416.

hepaticum Betegh in Gomy, 1898, 328–329 [Veterinarius, v. 21 (11), June 1]; [Gomy, 1897, Aug. 21, 401].—Rail. & Gomy, 1899, 348.—Refers to "amphistome hépatique" = A. explanatum.

hirudo Dies., 1836d, 238, 249, pl. 23, figs. 10-12 (Palamedea cornuta; Engenho do Cap Gama, Brazil); 1850a, 407.—Braun, 1893a, 874, 905.—Duj., 1845a, 355.— Nord., 1840, 628.

hominis Lewis & McConnell, 1876a, 182-186, 1 text fig., pl. 3, figs. 1-3 (Homo; Calcutta, India):—Anders, 1903, 1245.—B1., 1888a, 632–636, figs. 327–329; 1895, 744; 1900, 488.—Braun, 1883. 69; 1892a, 663; 1893a, 874, 905; 1895, 137–138, 744; 1900, 488.—Braun, 1883, 69; 1892a, 663; 1893a, 874, 905; 1895, 137–138, fig. 52; 1903, 146 (to Gastrodiscus).—Cobbold, 1879, 36–38, fig. 6; 1883x, 515.—Dav., 1877a, cxxxii.—Dunglison, 1893, 50, 533.—Fischder., 1901, 374; 1902a, 9, 46 (to Gastrodiscus); 1903h, 496.—Gamb., 1896, 63.—Giles, 1890, 125.—Hackley, 1886, 519.—Huber, 1896, 579–580.—Ijima, 1889, 157.—Jamison, 1897a, 74.—Kholod., 1898, 33, pl. 11, figs. 26–27; 1899a, 153.—Looss, 1902, 746; 1905, 110 (to Gastrodiscus).—Manson, 1901, 541, 543; 1903, 3. ed., 663, fig. 106.—Mon., 1896, 86, 87–89, figs. 17–18.—Mosler & Peiper, 1894, 185–186.—Schneidemuehl, 1896, 303.—Simon, 1897, 209; 1897, 223–224.—Sons., 1895, 181, 183; 1895, 187, fig. 6 to (Gastrodiscus); 1896, 310.—Stiles, 1904, 46 (to Gastrodiscus).—Veterinarian, Lond., 1877, v. 50, 82.—Vogt, 1878, 10.—Ward, 1895, 327 (in Homo); 1903, 865 (to Gastrodiscus).

isostomum Rud., 1814a, 100 (Strigea candida and Amphist. anatis tadornæ, renamed; in Anas tadorna; Copenhagen); 1819a, 89, 355, 793.—Bellingham, 1844a, 339.—Duj., 1845a, 377 (to Holost.).—Dies., 1850a, 313 (syn. of Holost. erraticum); 1858e, 320.—Lamouroux, 1822a, 297.—Olfers, 1816, 48.—Westrumb, 1823, 393.

konikum Schneidemuehl, 1896, 303, for conicum.

lari glauci Rud., 1819a, 92 (in Larus glaucus; Cat. Ent. Vien.), 793.—Dies., 1836d, 253; 1850a, 310 (syn. of Hemist. spathaceum).

loliginis delle Chiaje, 1841a, 140 (sub Amphystoma).—Par., 1894.

longicolle Rud., 1819a, 87–88, 92, 352–353 (Ardea alba, A. stellaris, Larus ridibundus, and L. atricilla; Mus. Vienn.), 793.—Baird, 1853a, 48 (to Holost.).—Bellingham, 1844, 338.—de Blainv., 1828, 583.—Bremser, 1824, 132; 1824c, pl. 8, figs. 15–16.—Dies., 1850a, 316 (to Holost.); 1858e, 321.—Fischer, 1840, 157.—Lamouroux, 1822a, 297.—Stoss., 1898, 21.—Westrumb, 1823, 390–391.

lunatum Dies., 1836d, 238, 240, 250–251, pl. 23, figs. 21–22 (Cervus dichotomus, Anas melanotus, Anas ipecutiri, Himantopus wilsonii; Caiçara); 1850a, 405–406 ("Cervus dichotomus, nisi lapsus calami;" other hosts as in 1836d).—Braun, 1892a, 570; 1893a, 874, 905.—Crep., 1839, 286.—Duj., 1845a, 335–336.—Fischder., 1902a, 53–54 in Cervus dichotomus?, Anas ipecuteri, A. himantopus, A. melanotus, A. moschata fer.; Brazil).—Nord., 1840, 629.—Also reported for Cariacus paludosus.

macrocephalum Rud., 1803a, 21–23 (Strix bubo) (includes: Fasc. strigis Gmelin, 1790a, and Strigea Abildg., 1790 [Copenhagen]) [type of Amphist., by inclusion]; 1809a, 50, 340–342 (in Strix ulula, S. otus, S. bubo, S. flammea, Scolopax gallinago); 1814a, 99–100; 1819a, 88 (additional hosts: Falco albicilla, F. aluco, F. apivorus, F. buteo, F. chrysaëtos, F. cineraceus, F. cyaneus, F. gallicus, F. haliaëtos, F. lagopus, F. lanarius, F. leucosoma, F. lithofalco, F. milvus,

F. nævius, F. nisus, F. palumbarius, F. pennatus, F. peregrinus, F. rufipes, F. subbuteo, F. tinnunculus, Strix brachyotus, S. dasypus, S. passerina), 354–355 (includes Holost. variabile Nitzsch, and Fest. oti), 793.—Baird, 1853a. 47, 48 to Holost.).—Bellingham, 1844a, 338.—Brand., 1888a, 10.—Bremser, 1824, pl. 8, figs. 18, 19, 21, 22.—Cobbold, 1858b, 164 (syn. of Hemist. spathula Dies., see next entry).—Crep., 1839, 288.—Dies., 1850a, 312–313 (A. mac. of Rud., 1809a, 340; 1819a, 88. exc. syn.; Westrumb, 1823, 391; Bremser, 1824, pl. 8, figs. 18, 19, 21, 22, all as syn. of Holost. variabile); 1858e, 320 (syn. of Holost. variabile).—Fischer., 1840, 157.—Fischder, 1901, 367; 1902a, 7 (e-Plan. strigis): 1903h, 490 (=strigis; type of Strigea, Amphist., Holost.).—Lamarck, 1816, 188.—Lamouroux, 1822a, 297.—Nitzsch, 1819, 400 (syn. of Holost. variabile).—Nord., 1840, 625 (syns. Fasc. strigis, Holost. variabile).—Olfers, 1816, 47.—Stoss., 1898, 21 (syn. of Holost. variabile).—Westrumb, 1823, 392–393.

macrocephalum Rud., 1819a, 88 (pars: falconis milvi).—Baird, 1853a, 48.—Bremser, 1824. pl. 8, figs. 17, 20.—Dies., 1850a, 309 (syn. of Hemist. spathula).—

Linst., 1905, 191.—Nord., 1840, 626 syn. of A. striatum).

megacotyle Dies., 1836d, 238, 250, pl. 23, figs. 19-20 (Silurus palmito; Matogrosso); 1850a, 405 (Trachicirrhus nattereri; Brazil).—Braun, 1893a, 905.—Duj., 1845a, 340 (in Sil. palm.; Brazil).—MacCallum, 1905, 668 (in Silurus).—Nord., 1840, 629.

microcephalum Rud., 1819a, 88, lapsus for A. microstomum Rud.

microstomum Rud., 1809a. 342–343 (Corvus carvocatactes: Europe); 1819a. 88, 793.—Dies., 1850a. 314 (to Holost.): Duj.. 1845a, 371 (Greiswald), Ito Holost.).—Lamouroux, 1822a, 297.—Olfers, 1816, 47.—Westrumb, 1823, 393.

mutabile (Zed., 1800) Sieb., 1835, 70, lapsus for Monost. mut.

nattereri Cobbold, 1860a, 52; cornu Dies., 1839a [not (Zed., 1800) Rud., 1809], renamed; t. h. Callichthys vacu; Brazil.

oidemiæ nigræ Dies., 1858e, 322 (syn. of Holost. anatis nigræ), based on Bellingham, 1844a, 340, Ireland.

ornatum Cobbold, 1882a, March. 240, fig. 9 (Elephas indicus).—Braun, 1893a, 882, 905.— ischder., 1902a, 49 (in Elephas indicus; India).—Looss, 1902m, 439.—Mégnin, 1882v, 454–456 (in elephant).—Sons., 1895, 183.

oxycephalum Dies., 1836d, 238. 240, 251, pl. 24, figs. 1-8 (Salmo auratus, S. pacu, Silurus megacephalus at Cuyaba, Salmo pacupeba at Rio Panara and Rio Araguay); 1850a, 407.—Braun, 1892a, 586; 1893a, 905.—Duj., 1845a, 339-340.—MacCallum, 1905, 668, 672 (in Salmo and Silurus).—Nord., 1840, 629.

papillatum Cobbold, 1882, 240–242, fig. 10, pl. 24, fig. 11 (Elephas indicus).—
 Braun, 1892a, 580, 663; 1893a, 874, 905; 1893d, 466.—Fischder., 1902a, 49
 (in Elephas indicus; India).—Sons., 1895, 184, 187, figs. 4–5.

papilliferum Braun, 1892a, 586, for A. papillatum Cobbold.

pileatum (Rud., 1802) Rud., 1819a, 90, 358, 793.—Blainv., 1828, 584 (to Holost.).—Dies., 1850a, 314 (to Holost.).—Lamouroux, 1822a, 297.—Westrumb, 1823, 395.

pileatum Rud., of Bremser, 1824, pl. 8, figs. 28-29.—Dies., 1850a, 311 (syn. of Hemist. commutatum).

piriforme Sons., 1895, 184 (for pyriforme).

platycephatum Crep., 1825a, 39-41, 81-82 (Colymbus rufogularis; Greifswald); 1846, 138, 139, 140, 141.—Dies., 1850a, 313 (to Holost.).—Duj., 1845a, 376 (to Holost.).

pulcherrima Weyenbergh, 1876, 167–169, pl. 9, figs. 75–78 (in Hypostomus plecostomus Val.; Argentina); 1878, 354 (to Dist.).—Looss, 1885b, 55.

pyriforme Dies., 1838a, 189 (in Tapirus americanus); 1839a, 236. pl. 20, figs. 17–18
(Tap. amer.; Matogrosso or Cachoeira do Bananeira); 1850a, 403 (Brazil).—
Braun, 1893a. 874, 905; 1893d, 466 (in Tap. amer.).—Cobbold. 1875n. 819; 1879b, 402.—Duj., 1845a, 334.—Fischder., 1902a, 36 (to Cladorchis).—Nord., 1840, 629.—Sons., 1895, 184 (piriforme).—Weyenbergh, 1878, 167–169.

rhopalsides Crep., 1839, 294; 1846, 168 ('ialsch geschrieben ropaloides'' Lebland).—Baird, 1853a. 116 syn. of Tetrabothriorhynchus barbatus).—Dies., 1850a. 574 syn. of Tetrab. migratorius in Conger vulgaris) (includes Lebland, 1836e. 290, pl. 16. figs. 1-3; 251.—Deslongchamps,—, 294.—Duj., 1845a, 341.—Sieb. 1837, 265; 1838, 306).—Lacaze-Duthiers, 1854a, 294.

ropaloides Leblond, 1836e, 290, pl. 16, figs. 1–3 (Muræna conger; coast of Normandie); 1836f, 4, pl. 16, figs. 1–3 (in Muræna conger).—Crep., 1839, 294.—Deslongchamps, ——, 249.—Duj., 1845a, 341 (coast of Normandie).—Sieb., 1837, 265; 1838, 306.—Vaullegeard, 1899, 82.

scleroporum Rud., of Brand., 1891d, 19 (for Creplin?).

scleroporum Crep., 1844a, 112–115, pl. 3, fig. A (Chelonia mydas); 1846, 146.— Braun, 1893a, 905; 1899b, 715 (in Chelone viridis), 725 (C. mydas); 1901a, 39, 40, 55, 56, 1 fig.—Dies., 1850a, 406 (Halichelys atra; Vratislaviæ).—Looss, 1901, 623, 624; 1902m, 430, 437, 438, 440, 562.—Mont., 1896, 165.—Walter, 1892, 248.

serpens (Nitzsch, 1819) Rud., 1819a, 88, 353-354 (in Falco haliaëtus; September),
 793.—Dies., 1850a, 316 (to Holost.).—Duj., 1845a, 371, at Halle (to Holost.).—
 Lamouroux, 1822a, 297.—Nord., 1840, 628 (to Holost.).—Westrumb, 1823, 392.

sonsinoi Linst., 1889, 23 (for sonsinoii) (in Equus caballus).—Ward, 1895, 338, (syn. of Gastrodiscus ægyptiacus) (in Equus caballus).

species Bellingham, 1844, 340.—Braun, 1901a, 55, 56.—Looss, 1902m, 430.

species Cobbold, 1879, in Equus caballus.—Ward, 1895, 338.

species Fischder., 1903, v. 17, 594; 1904, 468, syn.? of Paramphist. scolioccelium. species in Felis catus.

species in Phoca grænlandica.

sphærocephalum Westrumb, 1823, 396 (Coracias jugularis; at Rio Janeiro).— Brandes, 1888a, 65 (to Holost.); 1890a, 592.—Dies., 1850a, 314 (to Holost.; in Coracina scutata; Brazil).

sphærula Rud., 1803a, 23–24 (Corvus cornix; Greifswald); 1809a, 345–346; 1819a, 90, 358.—Bellingham, 1843a, 343; 1844, 339.—Dies., 1850a, 314 (to Holost.).—Duj., 1845a, 371 (to Holost.).—Lamouroux, 1822a, 297.—Olfers, 1816, 47.—Westrumb, 1823, 395.

spinulosum Looss, 19011, 7 Nov., 623–624 (in Chelone mydas; Alexandria, from Egyptian coast); 1902m, 415, 430–444, pl. 21, figs. 10, 11; pl. 22, figs. 12–16, 430, 434, 436, 437, 438, 439, 442, 649, 676, 869 (colon of Ch. mydas), 871.—Shipley, 1905, v. 2, 846.

stanleyi (Cobbold, 1879) Ward, 1895, 338 (in Equus caballus) (for stanleyii Cobbold).

stanleyii Cobbold, 1875n, 819.—Fischder., 1902a, 48 (stanleyi) (in Equus caballus; India).

striatum Rud., 1809a, 343 (Falco milvus; Europe); 1819a, 88 (syn. of A. macrocephalum, 793.—Baird, 1853a, 47 (syn. of Holost. macroceph.).—Dies., 1850a. 309 (syn. of Hemist. spathula).—Lamarck, 1816b, 188.—Nord., 1840, 626 (syn. A. macroceph. Rud.).—Olfers, 1816, 47.

subclavatum (Geze, 1782) Rud., 1802b, 92–93; 1809a, 348–349 (in Rana temporaria, R. bufo, R. arborea, R. esculenta); 1819a, 90–91 (in Bufo igneus, B. cinereus), 358–359, 589, 793.—Baillet, 1866b, 96 (of Nitzsch).—Ben., 1858a, 1861a, 81–84, 171, 203, 215 (syns. Plan. subclavata, Dist. subclavatum, Diplodiscus subclavatus, Dip. diesingii, Diplocotyle mutabile).—Bettend., 1897a, 8, 37; 1897, 312.—Blainv., 1828a, 53.—E. Bl., 1847a, 316–317, pl. 14, fig. 1.—Brand., 1891d, 17.—Braun, 1892a, 592, 798: 1893a, 845, 848, 851, 852, 854, 858, 864, 869, 874, 879; 1893b, 177, 179, 180, 182; 1895b, 11.—Bremser, 1824c, pl. 8, figs, 30–31; 1824, 133.—Cobbold, 1872b, 91 (to Diplodiscus); 1879b, 49, 452, 454.—Crep., 1839a, 286.—Creutzburg, 1890a, 21.—Darr, 1902a, 678, 688.—Dies., 1836d, 237, 238, 240, 253 (to Diplodiscus); 1850a, 318 (in Dendrohyas viridis, Rana temporaria, Pelophylax esculentus, Phryne vulgaris, Bufo viridis, Bombinator igneus, Leptodactylus sibilatrix); 1858e, 360, 1859c, 435.—Duj., 1845a, 336, 339 (in Rana esculenta; Rennes).—Erc., 1881e, 57 (embryo), pl. 2, fig. 9; 1882a, 293.—Fil., 1837a, 338 (syn. Diplodiscus subcl.); 1854a, 6; 1855b, 13–17, pl. 2, figs. 14–16 (in Planorbis vortex; Moncalier); 1857c, 32.—Fischder., 1903h, 487 (to Diplodiscus by Dies.).—Fraip., 1880c, 419.—Gerv. & Ben., 1859b, 212.—Gronkowski, 1902a, 515 (8).—Hoyle, 1890, 539.—Ijima, 1884c, 638.—Kitt, 1885a, 148.—Knoch, 1894a, 10.—Lamarck, 1816b, 189.—Lamouroux, 1822a, 297 (Amphistome).—Lang, 1892a, 81–89, 1 fig. (syn. Diplodiscus subc. Dies.) (Cerc. of); 1893a, 479.—Lejtenyi, 1881a, 5, 8, 10, 18, 20.—Leuck., 1863, 82, 488, 491; 1879, 107; 1886d, 80.—Levin., 1881a, 63, 69.—Looss, 1892b, 147–167, 1 fig., pls. 19–20 in text (devel-

opment); 1894a, 8, 85, 117, 133, 136, 137, 142, 146, 150, 169, 178, 206, 227, 234, 237, 248; 1894d, 17, 21, 22; 1896b, 17, 31, 138, 171, 172, 178, 179, 180, 181, 182, 183, 188, 189, 190; 1901, 441; 1902m, 438, 444 (Cerc. diplecotylea).—Macé, 1882, 12.—Mont., 1888, 39, 49, 50, 73, 78; 1891, 117; 1893, 68, 160, 205.—Moul., 1856a, 18, 20 (eye of Cerc.), 47 (embryo, 125µ), 50, 125 (eye of Cerc.), 208-211, pl. 6, fig. 10 (Cerc.=Diplodiscus diesingii) (in Planorbis vortex), 212.—Mueller,———Nitzsch, 1819, 398-399 (syn. Plan. subcl. Geze).—Nord., 1840, 627 (syns. Plan. subcl., Fasciolaria ranæ Gmelin, Diplodiscus subcl. Dies.).—Olfers, 1816, 45, 48.—Pag., 1857, 25, 26, 49-50, 52, pl. 6, figs. 5-9 (in braunen Frosche).—Poir., 1885, 102.—Risso, 1826, 262.—Roosbach, 1906, 368, 369, 375, 392, 394, 402, 405, 408, 413, 414, 423, 429, 431.—Sieb., 1835, 56, 57, 67, 70; —, v. 1, 160.—Slawikowski, 1819, 49.—Sons., 1884, 59, 60, 61; 1893, 187 (subelavatum) 188 (in Rana temporaria), 189 (in Triton cristatus, Rana esculenta L.).—Ssinitzin, 1904, 768, figs. a, b; 1906, 684.—Stephens, 1906, 10.—Tennent, 1906, 650.—Villot, 1878, 6.—Wagener, 1857, 26, 27.—Walter, 1858, v. 1, in 268-297, pls. 11-13; 1866, 64-65.—Westrumb, 1823, 396.—Zed., 1803a, 198, pl. 3, fig. 3. claratum Rud. of Staff., 1900, 405, 412 ("rectum of frogs, chiefly of the small

subclaratum Rud. of Staff., 1900, 405, 412 ("rectum of frogs, chiefly of the small grass frogs;" Canada); 1905, 689-690 (Rana virescens Kalm, R. catesbiana Shaw; Canada), renamed Diplodiscus temperatus.—Looss, 1885b, 24.

subelavatum Sons., 1893, 187, for subclavatum.

subtriquetrum Rud., 1814a, 100 (Castor fiber; Berlin, May); 1819a, 91–92, 360–361, 577, 589, 793.—Blainv., 1828, 583.—Bojanus, 1821, 164, pl. 2, figs. 5–12.—Braun, 1892a, 578, 601, 608, 680, 681, 694, 746; 1893a, 905.—Bremser, 1824c, pl. 8, figs. 32–33.—Civinini, 1842, ——.—Cobbold, 1879, 317.—Crep., 1837, 313, 323; 1841, 80.—Dies., 1836d, 238, 244, 245, 246, 248–249, pl. 23, figs. 7–9 (in Castor fiber) (syn. Dist. amphistomoides); 1850a, 402 (syn. Dist. amphistomoides).—Duj., 1845a, 331–332.—Eichwald, 1829a, 249.—Fischer, 1840, 157.—Fischder., 1901, 373; 1902a, 41, 42 to Cladorchis (Stichorchis); 1903h, 506, 507.—Kuech., 1855, 192.—Lamouroux, 1822a, 297.—Leidy, 1888, 126.—Leuck., 1863, 460.—Macé, 1882, 9.—Mol., 1859, 849 (in Castor fiber).—Nitzsch., 1819, 398, 399.—Nord., 1840, 547.—Olfers, 1816, 47.—Otto, 1896, 105–108, fig. 8.—Schmalz, 1831, 24, 27.—Sieb., 1835, 64, 65.—Slawikowski, 1819, 52.—Sons., 1895, 184, 187, fig. 7.—Westrumb, 1823, 397.—Also reported for Arvicola campestris, Fiber zibethieus.

subtriquetrum giganteum Kuech., 1855, 192.

sylvix Rud., 1819a, 675 (Sylvia cyanea; Brazil), 793.—Dies., 1836, 253.—Westrumb, 1823, 398.

tadornæ Rud., 1819a, 89 (for Λ. anatis tadornæ and syn. of Λ. isostomum), 793.
 tanagræ Rud., 1819a, 674-675 (Tanagra tatoa; Brazil), 793.—Dies., 1836, 253.—Duj., 1845a, 335.—Westrumb, 1823, 397.

tenuicolle Westrumb, 1823, 391–392, pl. 5, fig. 2 (Falco rufus).—Dies., 1850a, 316

(to Holost.); Mus. Cat. Vien.

truncatum Rud., 1819a, 91, 359–360 (in Phoca vitulina; Berlin and Vratislaviæ), 793.—Anacker, 1888b, 314; 1892c, 94.—Baillet, 1866b, 105 (syn. of Dist. conus).—Braun, 1892a, 578, 586; 1893a, 875, 905; 1893, 348, 349, 350; 1893d, 467 (in Phoca sp.); 1893f, 382, 383, 384, 385, 386, 389, 424, fig. 1; 1893g, 802 (syn. of Dist. conus Crep.).—Cobbold, 1875n, 819; 1879b, 307, 313.—Crep., 1839, 286.—Dav., 1877a, lxxx.—Dies., 1836, 238, 252, pl. 24, figs. 13–15; 1850a, 404; 1858e, 350 (syn. of Dist. conus), 358 (in Phoca grænlandica).—Duj., 1845a, 331.—de Jong, 1896a, 3, 4, 5.—Lamouroux, 1822a, 297.—Linst., 1878, 44.—Mol., 1859, 849 (in Felis catus).—Mont., 1889, 321.—Verrill, 1870, 175, 220.—Ward, 1895, 341 (in Canis familiaris).—Westrumb, 1823, 397.—Zuern, 1882, 223.

truncatum of Dies., 1850a, 404 (in Felis catus); 1858e, 332 (syn. of Dist. lanceolatum).—Kitt, 1885a, 148 (in cat).—Sons., 1889, 281 (syn. of Dist. conus Crep.).—Stoss., 1892, 25 (syn. of Dist. conus Crep.).

trunkatum Schneidemuehl, 1896, 303, for truncatum.

tuberculatum Cobbold, 1875n, 819 (in ox: India); 1879b, 332.—Fischder., 1902a,
 49 (in Bos taurus; India).—Stiles, 1898a, 67, 140.—Ward, 1895, 332 (in Bos taurus).

unciforme Rud., 1819a, 674 (Oriolus cristatus: Brazil), 793.—Braun, 1892a, 578;
1893a, 905.—Dies., 1836d, 238, 252-253, pl. 24, figs. 16~18; 1850a, 404.—Duj.,
1845a, 335.—Westrumb, 1823, 397.

unguiculatum Rud., 1819a, 91, 360 (Triton palustris; Berlin), 793.—Baird, 1853a, 44 (syn. of A. subclavatum).—Crep., 1839a, 286.—Dies., 1836d, 237, 254; 1850a, 319 (to Diplodiscus).—Fischder., 1903h, 487 (to Diplodiscus by Dies.).—Lamouroux, 1822a, 297.—Nitzsch, 1819, 399 (in Wassersalamander).— Westrumb, 1823, 397.

westerm Rud., 1819a, 89, 356–357 (in Rana esculenta; Mus. Vien.), 793.—
Baird, 1853a, 49 (to Holost.).—Blainv., 1828a, 583.—Braun, 1892a, 796.—
Bremser, 1824c, pl. 8, figs. 24–27.—Crep., 1825a, 41–45, 83.—Dies., 1850a, 318 (syn. of Codonocephalus mutabilis; type of Codonoc.).—Duj., 1845a, 378, 379 (to Holost.) (in Rana esculenta; Paris).—Lamouroux, 1822a, 297.—
Moul., 1856a, 16 (type of Codonocephalus Dies., 1850).—Nitzsch, 1819, 400.—Sans 1893, 190 (syn. of Cod. mut. Dies.) —Westrumb, 1823, 304 400.—Sons., 1893, 190 (syn. of Cod. mut. Dies.).—Westrumb, 1823, 394.

vallei (Stoss., 1899) Looss, 1902m, [418], 430 (to Lophotaspis), 890; the reference on p. 890 (index) is apparently a lapsus for A. spinulosum.

variegatum Crep., 1825a, 38–39, figs. 4–6 (in Larus marinus; Greifswald); 1846, 139, 140, 146.—Dies., 1850a, 315 (to Holst.).—Duj., 1845a, 376.

watsoni Conyngham, 1904, Aug. 13, 464, Aug. 15, 252, Aug. 27, 355; 1904, Sept. 17, 663, figs. 1–2 (in Homo; Africa); 1904, Sept. 29, 1480; 1904, Oct. 8, 710; 1904, Sept. 8, 1092.—Shipley, 1905, 8 (syn. of Cladorchis watsoni); original patient in Zola (Nigeria) came from Adamawa, German West Africa.

AMPHISTOMÆ Mont., 1888a, 18.

AMPHISTOMATA Bojanus, 1817b, 275, 276.

AMPHISTOMATIDÆ Gamb., 1896a, 73.

AMPHISTOME Lamouroux, 1822a, 297, for Amphistoma.

AMPHISTOMEÆ Zuern, 1882, 113.—Brand., 1888, 49.—Braun, 1893a, 886.—Mont., 1888a, "mihi," 7, 11, 12, 14, 15, 16, 18, 34, 36, 38, 41, 51, 52, 56, 57, 60, 90, 91, 102, 103; 1892, Oct. 7, 195.

AMPHISTOMIDÆ E. Bl., 1847a, 309.—R. Bl., 1895, 730.—Brand., 1890a, 576.—Braun, 1893a, 886, 890, 895, 900, 904; 1895b, 136.—Cobbold, 1877, 235; 1879b, 4.—Cohn, 1904, 242.—Fischder., 1901, 367; 1902a, 8 (renamed Paramphistomidæ; 1903h, 489, 490 (syn. of Paramphistomidæ).—Kholodk., 1898, 25, 33.—Looss, 1899, 541, 542, 543.—MacCallum, 1905, 667.—Mont., 1888a, "mihi," 21, 90, 91, 103, 108; 1892, Oct. 7, 214 (f. of Malaccoctylea); 1893, 82.—Much., 1898, 20.—Nickerson, 1902, 612.—Poir., 1883, 74; 1885, 147.—Schneidemuchl, 1896, 295.—Sons., 1895, 184, 185, 186.—Stiles, 1898a, 22, 24, 27, 64.—Stiles & Hass., 1898, 87.—Zuern, 1882, 113. 64.—Stiles & Hass., 1898, 87.—Zuern, 1882, 113. 1895: Amfistomidi Sons., 1895.

[AMPHISTOMINA Lankester, E. R., 1890, 846 (f. of Gromiidea, Protozoa).]

AMPHISTOMINÆ Mont., 1892, Oct. 7, 214 (subf. of Amphistomidæ).—Braun, 1893a, 890.—Looss, 1899, 541, 543.—Not Amphistominæ, subf. of Gromidæ (Protozoa), see Calkins, 1901b, 106.

AMPHISTOMULUM Brand., 1892b, Oct. 7, 510 (proposed as an artificial collective group to contain immature amphistomes).—Stiles, 1902, 28, 48.

AMPHISTOMUM Nitzsch, 1819, 397.—Crep., 1825a, 35 (for Amphistoma, q. v.).

[AMPHITYPIE, a term, not a zoological name, see Looss, 1902m, 789-792.]

AMPHYSTOMA Rud., 1803a, 29 (for Amphistoma, q. v.)

ANADASMUS Looss, 1899b, 568–569 (m. amphiorchis) [not Anadasmus Walsingham, 1897, insect]; 1900d, 601, 602 (renamed Orchidasma); 1902m, 839.—Braun, 1901b, 20.—Luehe, 1901, 488.—Stiles, 1901, 185, 189, 191.

amphiorchis (Braun, 1899) Looss, [1899b, 568, 569]; 1902m, 463 (to Orchidasma).

ANAPORRHUTINÆ Looss, 1901, 205 (subf. of Fasciolidæ); 1901, 558; 1902, 479, 485, 844, 862-863 (diagnosis).—Odhn., 1902, 65, 68.—Pratt, 1902a, 888, 901 (key) (includes Anaporrhutum, Plesiorchorus; related genus Callodistomum).

ANAPORRHUTUM Ofenheim, 1900, 145–186 (type albidum Brand., designated by Looss, 1900b, 204).—Braun, 1902b, 23.—Darr, 1902, 652.—Looss, 1901b, 204, 205, 209; 1901l, 557, 558; 1902m, 479, 480, 481, 482, 484, 485, 526, 621, 813, 814, 844, 848, 851, 852, 853, 854, 855, 857 (Anaporrhutinæ), 858, 859, 860, fig. B. V, 863 (diagnosis).—Odhn., 1902, 65, 67.—Pratt, 1902a, 888, 900 (key), 901 (key).

ANAPORRHUTUM—Continued.

albidum Brand. in Ofenheim, 1900, 145–186, figs. 1–4 in text, pl. 3, figs. 1–8 (in body cavity and pericardium of Aëtobatis narinari, in Pacific).—Hollack, 1902a, 868.—Looss. 1901b, 204 (type of genus), 206, 209; 1901l, 557; 1902m, 479, 482, 483, 484, 620, 622, 623. 624, 625, 628, 791 (Amphitypie), 844, 848, 851, 852, 855, 857, 863.—Odhn., 1902, 154.

ricchiardii (Lopez, 1888) Looss, 1901b, 204, 206, 209; 1901l, 557, 558; 1902, 479, 482, 483, 484, 621, 622, 791 (Amphitypie), 844, 848, 852, 853, 854, 855 (to Proboli-

trema as type).

ricchiardii (Lopez of Ofenheim) Looss, 1902m, 479, 482, 483, 484, 621, 622, 852, 854, 855 (renamed Probolitrema capense).

richiardii of Ofenheim, 1900, 169–180, 178, 179, 180, 185, 186, pl. 3, figs. 9–12 (in Scyllium sp., Atlantic, near Kapstadt).—Odhn., 1901, 61, 1902, 61.

ANCHITREMA Looss, 1899b. 637 (m. sanguineum); ~\(\tilde{x}\gamma\gamma\text{z}\), near related; 1901b, 200; 1902m, 839.—Klein, 1905, 78; 1905, 20.—Luehe, 1901, 487.—Odhn., 1902, 41.—Ofenheim, 1900, 183.—Pratt, 1902a, 889, 905 (key).

mutabile (Mol., 1859) Rizzo, 1902, 27-28, fig. 1 (Ascalobotes mauritanicus, Lacerta

agilis).

sanguineum (Sons., 1894) Looss, 1899b, 637; 1902m, 492, 818, 823.—Heymann, 1905, 97.—Luehe, 1900aa, 565, 566.

ANCYROCEPHALUS Crep., 1839a, 292 (m. paradoxus),—Ceri, 1895h, 918; 1896, 514.—Dies., 1850a, 416; 1858e, 314, 368 (f. Polycotylea, subf. Aplacocotylea).—Mont., 1903, 336 (syn. of Tetraonchus).—Stiles & Hass., 1905b, 85.

paradoxus Crep., 1839a, 292 (in Perca lucioperca); 1846, 155.—Dies., 1850a, 416 (Greifswald, Vienna, Gedani); 1858e, 368, pl. 1 (in Lucioperca sandra).—Duj., 1845a. 654 (in P. luc.).—Kroyer, 1838—40a, 579 (in Lucioperca sandra).—Mont., 1888, 90; 1889, v. 3(2), 113—116 (syn. of Tetraonchus unguiculatus Wagener; in Lucioperca sandra, Perca fluviatilis); 1889, 26 Oct., 516—517; 1891, 109; 1893, 113—116.—Sieb.,—, 298.—Tasch., 1879, 238, 264 (syn. of of Tetraonchus unguiculatus Wagener).

ANCYROCOTILE Massa, 1903, 252, for Ancyrocotyle.

ANCYROCOTYLE Par. & Mont., 1903, v. 7 (1), 117–123, pl. 3, figs. 1–6 (m. vallei); 1904, May 17, 280.—Massa, 1903, 252 (Ancyrocotile); 1906, 44, 48.—Mont., 1903, 335 (subf. Ancyrocotylinæ; f. Tristomidæ).

vallei (Par. & Perugia, 1895) Par. & Mont., 1903, 117-121, pl. 3, figs. 1-6 (Naucrates

ductor).

ANCYROCOTYLINE Mont., 1903, 335 (subf. of Tristomidæ).

ANGIODICTGUM Looss, 1902m, 648 (misprint for Angiodictyum).

ANGIODICTYIDÆ Looss, 1902m. 484. 617-684 (anat. histol.) 683 (diagnosis, key to genera). 699, 843 (contains Deuterobaris, Octangium, Microscaphidium, Angiodictyum, Polyangium).

ANGIODICTYUM Looss, 1902m. 433, 632, 634, 639, 641, 642, 645, 647, [648], 652, 657, 658, 659, 665, 666, 668, 675, 679, 681, 683, 684, 688–689 (diagnosis, m. parallelum [Looss]), 691, 698, 699.

1902: Angiodictgum Looss, 1902m, 648.

parallelum (Looss, 1901) Looss, 1902m, 632, 689-690, 696, 698, pl. 29, figs. 134-136, pl. 30, figs. 137-142.

ANISOCLADIUM Looss, 1902e, 637, new name for Anisogaster Looss, 1901, 658 (not Anisogaster Deyr, 1863, coleopteron), hence type fallax; 1902m, 789.

fallax (Rud., 1819) Looss, 1902m, 789.

ANISOCŒLIINÆ Looss, 1901e, 658.

ANISOCŒLINÆ Pratt, 1902a, 888, 894 (key), 899 (for Anisocœliinæ Looss) contains Anisocœlium and Anisogaster.

ANISOCŒLIUM Luehe, 1900. 504–507 (m. capitellatum).—Looss, 1901. 656, 658, 659; 1901, 206.—Odhn., 1905, 314.—Pratt, 1902a, 888, 895 (key), 899.

capitellatum (Rud., 1819) Luehe, 1900, 504-507, 508; 1901, 399.—Hollack, 1902a, 868.—Looss, 1901, 656-658, 659, 660, fig. 12; 1902, 789.—Stoss., 1901, 97 (in Uranoscopus scaber) (9).

ANISOCOTYLINE Mont., 1903, 336 (f. Monocotylidæ); 1904, 118; 1905, 70, 71; 1905, 117.

- ANISOGASTER Looss, 1901e, 658 (tod. fallax Rud.) [not Anisogaster Deyr, 1863, coleopteron]; renamed Anisocladium Looss, 1902e, 637; 1902e, 789.—Pratt, 1902a, 888, 894 (key), 899.
 - fallax (Rud., 1819) Looss, 1901e, 658–660, 661, fig. 13.
 - gracilis Looss, 1901e, 660–661, fig. 14 (in Uranoscopus scaber; Triest).
- "ANOICTOSTOMA Stoss." of Braun, 1899, 789, for Anoiktost. 1901b, 34; 1901f, 568; 1902b, 28, 30, 31.
 - planicolle (Rud., 1819) Braun, [1901f, 567;] 1902b, 28–31, fig. 20 (syns.: Dist. planicolle Rud., 1819; Duj., 1845; D. (Echinost.) planicolle Brand., 1892; Monost. echinostomum Dies., Mont.).
- ANOIKTASTOMA Pratt, 1902a, 888, 894 (key), for Anoiktost.; related to Centrocestinæ.
- ANOIKTOSTOMA Stoss., 1899, 11, 15–16 (coronatum=corvinæ=aloysiæ).—Looss, 1899b, 580, 581, 582, 583 (tld. coronatum=corvinæ=aloysiæ), 625.
 - 1899: Anoictostoma Braun, 1899, 789.
 - 1901: Anoitostoma Vaullegeard, 1901, 143.
 - 1902: Anoiktastoma Pratt, 1902a, 888, 894 (key).
 - aloysiæ (Stoss., 1885) Stiles & Hass., 1908, 99.
 - cesticillus (Mol., 1858) Stoss., 1899, 15 (includes Dist. bicoronatum Stoss., 1883, 113, pl. 1, figs. 1–3) (in Corvina nigra, Umbrina cirrhosa, Lophius piscatorius, Zeus faber).—Looss, 1899b, 576 (type of Stephanost.).
 - coleostomum (Looss, 1896) Stoss., 1899, 15.—Looss, 1899b, 585 (type of Ascocotyle).
 - coronarium (Cobbold, 1861) Stoss., 1899, 16 (int. of Alligator missisipiensis).— Looss, 1899b, 578 (to Acanthost.); 1901 (to Acanthochasmus).
 - coronatum (Wagener, 1852) Stoss., 1899, 15 (including Dist. corvinæ and Dist. aloysiæ Stoss.) (int. of Corvina nigra); [name not available, see corvinæ] (type by designation of Looss, 1899b, 583).
 - corvinæ (Stoss., 1886) (type by designation, Looss, 1899b [581–583], [see aloysiæ 1885].
 - cuspidatum (Looss, 1896) Stoss., 1899, 15 (int. of Milvus parasiticus; Egypt).— Looss, 1899b, 584 (to Centrocestus as type).
 - fallax (Rud., 1819) Stoss., 1899, 16 (intest. of Uranoscopus scaber.—Type of Anisogaster, 1901; type of Anisocladium, 1902.
 - hystrix (Duj., 1845) Stoss., 1899, 15 (stomach of Lepidoleprus trachyrhynchus, Lophius piscatorius, Merlangus carbonarius, etc.).—Looss, 1899b, 576 (to Stephanost.).
 - imbutiforme (Mol., 1859) Stoss., 1899, 16 (intest. of Labrax lupus).
 - inflatum (Mol., 1859) Stoss., 1899, 16 (as doubtful) (intest. of Anguilla).
 - lydiæ (Stoss., 1896) Stoss., 1899, 15 (intest. of Orthagoriscus mola).—Looss, 1899b, 576 (to Stephanost.).
 - planicolle (Rud., 1819) Braun, [1901e, 567 (sub Anoictost.);] 1902b, 28, fig. 20 (includes Dist. planicolle Rud., 1819a, 687; Duj., 1845a, 430; Monost. echinostomum Dies., 1850a, 326; Mont., 1892, 30; Dist. (Echinost.) planicolle of Brand., 1892, 506).
 - pristis (Deslongchamps, 1824) Stoss., 1899, 15 (intest. of Gadus euxinus).—Looss, 1899b, 576 (to Stephanost.).
 - scabrum (Mueller, 1788) Stoss., 1899, 16 (intest. of Gadus morrhua, Merlangus pollachius, Lota molya).—Looss, 1899b, 582 to D. (Hemiurus).
 - sobrinum (Levin., 1881) Stoss., 1899, 15 (intest. of Cottus scorpius.)—Looss, 1899b, 576 (to Stephanost.).
 - spiniceps (Looss, 1896) Stoss., 1899, 15 (in intest. of Bagrus bayad).—Looss, 1899b, 578, 582 (to Acanthosomum as type).
- ANOITOSTOMA Vaullegeard, 1901, 143 (misprint for Anoiktost.).
- ANOPLODISCUS Sons., 1890, 172 (m. richiardii); 1891, v. 15, 147–148.—Braun, 1891d, 422; 1893a, 890.—Gamb., 1896a, 73.—St.-Remy, 1898, 522, 524.—Mont., 1892, Oct. 7, 213 (g. of Calceostominæ); 1903, 336 (subf. Anisocotylinæ); 1905, 65, 66, 68, 70.—Pratt, 1900a, 646 (key), 654.
 - richiardii Sons., 1890, v. 7, 16. Nov., 172 (t. h. Pagrus orphus); 1891, v. 15, 147–148.— Braun, 1891d, 422.—Mont., 1891, 108; 1905, 65–68, figs. 1–3 (host P. orphus).— Par., 1894, 138, 672, at Pisa.

ANTHOCOTILE Par. & Perugia, 1890, 11, 13, for Anthocotyle.

ANTHOCOTYLE Ben. & Hesse, 1863; 1864, 96, 104 (m. merluccii).—Braun, 1890a, 413, 415, 516, 517, 523, 534, 536, 546; 1893a, 890.—Cerf., 1895d, 476–478; 1895e, 510–527, 1 pl., figs. 1–9; 1895g, 536; 1895h, 913, 918, 920; 1896b, 422–423; 1896d, 497-510, 514, 515; 1899a, 345, 391.—Gamb., 1896a, 73.—Hoyle, 1890, 539 ("one species (A. merlucii) found in the hake").—Mont., 1888a, 11, 66, 86, 89, 99; 1892, Oct. 7, 213 (subf. of Octocotylinæ); 1903, 336, (subf. Diaphorocotylinæ).—Pratt, 1900a, 646, 653 (key), 656, fig. 37.—St.-Remy, 1898, 556.— Scott, 1901. 148.—Sons., 1895, v. 6 (6), 118-121; 1896, June 4, 790.—Tasch., 1879, 69; 1879, 238, 247.

merluccii Ben. & Hesse, 1863; 1864, 105, pl. 10, figs. 8-12 (in Merluccius vulgaris).— Cerf., 1895e, 510, 511–525, figs. 1–8; 1896, 497, 500, 510; 1896, 498–509.—Par. & Perugia, 1890, 7.—Pratt, 1900a, 656, fig. 37, 657.—Scott, 1901, 148, pl. 8, fig. 4 (in Merlucius merlucius; Aberdeen); 1905, 117.—Staff., 1904, 482 (gills of Merluccius bilinearis Mit.; Canada).

merlucii Tasch., 1879, 247 (for merluccii)—Braun, 1900a, 419, 453, 510, 536, 548, 551.—Hoyle, 1890, 539.—Mont., 1888, 7, 8.—Sons., 1895, 118, 119, 120.

(APOBLEMA) Duj., 1845a, 383, 389, 420, as subg. of Dist.; raised to generic rank by E. Bl., 1847, 302. See Apoblema and Hemiurus. Type appendiculatum.

affine (Rud., 1819) Stoss., 1886, 44.—See Derogenes.

apertum (Rud., 1819) Duj., 1845a, 422.

appendiculatum (Rud., 1802) Duj., 1845a, 420-421.—Braun, 1891d, 423. See Apoblema.

caudiporum (Rud., 1819) Duj., 1845a, 422. See Lecithochirium.

crenatum (Mol., 1859) Stoss., 1886, 58. See Apoblema.

excisum (Rud., 1819a) Stoss., 1886, 58. See Apoblema.

grandiporum (Rud., 1819a) Stoss., 1886, 63. See Apoblema.

labri rupestris (Olss., 1876) Stoss., 1886, 51.

mollissimum (Levin., 1881) Stoss., 1886, 58. See Apoblema.

ocreatum (Rud., 1802) Duj., 1845a, 422-423.—Type of Pronopyge, 1899. See Apoblema.

raynerianum (Nardo, 1827) Stoss., 1886, 58.

rufoviride (Rud., 1819a) Duj., 1845a, 421. See Apoblema.

tornatum (Rud., 1819a) Duj., 1845a, 421–422. See Apoblema.

ventricosum (Rud., 1819) Stoss., 1886, 59. See Apoblema.

APOBLEMA (Duj., 1845) E. Bl., 1847, 302–303 (appendiculatum.)—Bettend., 1897a, 17; 1897. 321.—R. Bl., 1891, 609.—Brand., 1891b, 267; 1891d, 19, 20.—Braun, 1891d, 423; 1892a, 570, 590, 603, 608, o41, 682, 710, 713, 715, 720, 721, 744; 1892h, 728, 729; 1893a, 838, 879, 885, 886, 890, 893, 894, 909, 911; 1895, 744; 1892h, 728, 729; 1893a, 838, 879, 885, 886, 890, 893, 894, 909, 911; 1895, 128, 138; 1899, 3; 1900h, 3.—Dies., 1850a, 331 (syn. of Dist.).—Hausmann, 1897b, 38.—Juel, 1889a, 46 pp., figs. 1–18 (anatomy); 1889b, 46 pp., figs. 1–18; 1890a, pp. 54–55.—Kath., 1894a, 130.—Looss, 1894a, 122, 129, 142, 170, 200, 201, 204, 224; 1894d, 42; 1896b, 122, 123, 125, 127, 134; 1899, 527, 535, 538, 542, 582, 637, 638, 729; 1902, 756, 839.—Luehe, 1900, 509; 1901, 394, 484.—Mont., 1888a, 92, 105; 1891d, 496, 522, 1 pl.; 1891e, 32 pp., 17 figs.; 1892, Oct. 7, 214 (gen. of Distominæ); 1891h, 742; 1893, 6, 27, 33, 34, 43, 53, 61, 84, 85, 86, 88, 90, 107, 123, 149, 151, 152, 153, 154, 182.—Nickerson, 1902, 609.—Ofenheim, 1900, 156, 179, 181.—Pratt. 1898, 352, 368, 375.—Stiles, 1901, 177, 178, 197.—Stiles & Hass., 1898a, 83, 90, 91 (syn. of Hemiurus).—Stoss., 1892, 4; 1898, 27. See Hemiurus.

appendiculatum (Rud., 1802) E. Bl., 1847, 302-303, pl. 12, fig. 3.—Barbagallo & pradiculatum (Rud., 1802) E. Bl., 1847, 302–303, pl. 12, fig. 3.—Barbagallo & Drago, 1903, 409 (in Lichia amia, Lophius piscatorius, Cantania; Scomber colias, S. scomber).—Braun, 1892a, 714.—Dies., 1850a, 371 (to Dist.).—Juel, 1889, 4, 5, 7, 8, 13, 19, 22, 23, 24, 25, 26, 27, 28, 30, 31, 32, 33, 34, 35, 36, 39, 41, fig. 17.—Lander, 1904a, 4, to (Hemiurus), 7, 14.—Looss, 1894a, 224; 1896, 123, 124, 125, 131–140, pl. 9, figs. 88–90 (in Alosa finta; Cairo).—Luche, 1901, 396, 397, 399, 400.—Mont., 1891, 501, 502–508, 513, 516, 517, 518, 520, 521 (syns. D. app. Dies., 1858, D. ventricosum Wagener, 1860, D. ocreatum Olsson, 1868); 1891, 9; 1891, 502; 1891, 11, 12; 1893, 27, 61, 84, 95, 191, pl. 1, fig. 3.—Much., 1898, 21.—Odhn., 1905, 350, 351.—Par., 1902, 5 (in Clupea alosa, Scomber colias, S. scomber).—Pratt, 1898, 351–388, 3 pls. (anat., life history); 1900, 371.—Stoss., 1898, 28–29.—Also reported for Acipenser sturio, Trutta salar, Pseudocalanus elongatus. APOBLEMA-Continued.

crenatum (Rud., 1802) Juel, 1889, 5, 7, 34.—Mont., 1893, 95.—Stoss., 1898, 27–28.
crenatum of Mol.—Juel, 1889, 5, 7, 34.—Mont., 1891, 520, 521, 522; 1893, 95.—Stoss., 1898, 27–28 (in Merlucius esculentus; Triest).

excissum Mont., 1891, 520 (for excisum).

excisum (Rud., 1819) Juel, 1889, 5, 7, 8, 9, 10, 12, 13, 14, 15, 16, 17, 18, 19, 20, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 38, 39, 40, 41, figs. 1–16.—Braun, 1892, 48; 1892a, 607, 609, 711, 713, 744; 1892h, 728, 729.—Lander, 1904a, 7, 21, 24.—Looss, 1894a, 173, 210, 216, 224; 1896b, 133.—Luehe, 1898, 624.—Mont., 1891, 503, 517, 520 (excissum), 521, 522: 1893, 6, 7, 95.—Pratt, 1898, 356.—Par., 1902, 5 (in Scomber colias, S. scomber), 375.—Stoss., 1898, 28.

grandiporum Mol. See Mont., 1891, 521.

grandiporum (Rud., 1819a) Juel, 1889, 6, 7, 23, 29, 34.—Looss, 1896b, 123; 1899,
 641 (to Hemiurus).—Mont., 1891, 520.

labri rupestris (Olss., 1876) Mont., 1891, 518, 522.

microporum (Mont., 1889) Mont., 1891, 516, 521.

mollisimum Mont., 1893, 96 (for mollissimum).

mollissimum (Levin., 1881) Juel, 1889, 6–7, 29, 34.—Jacoby, 1900, 3.—Lander, 1904a, 14, 20 (to Hemiurus).—Looss, 1896b, 121–130, 136, pl. 9, figs. 85–87 (in Alosa finta); 1899b, 641 (to Hemiurus), 728 (H. bothryophorus) (in Alosa finta; Egypt).—Mont., 1891, 520 (mollissinum), 521 (mollissimun): 1893, 95, 102, 105.—Odhn., 1905, 359 (of Looss, 1896, as syn. of Lecithaster confusus).—Ofenheim, 1900, 179.—Staff., 1904. May 3, 484 (of Levin., as syn. of Lecithaster bothryophorus).—Stoss., 1898, 29–30.

mollissimun Mont., 1891, 521 (for mollissimum).

mollissinum Mont., 1891, 520 (for mollissimum).

ocreatum (Olss., 1868 nec Rud.) Mont., 1891, 520.

ocreatum (Rud., 1802a) Juel, 1889, 4-5, 7, 31, 33, 34, 40.—Braun, 1891d, 423; 1892a, 643.—Lander, 1904a, 1 (of Juel, syn. of Hemiurus crenatus (Rud.)).—Looss, 1899b, 639 (=Dist. carolinæ Stoss., type of Pronopyge, 1899).—Luehe, 1901, 399.—Mont., 1891, 15; 1891, 501, 508-512, 514, 516, 521, pl. 4, figs. 1, 5, 7, 10, 11, 14, 15c (syns. D. ventricosum Ben., D. carolinæ Stoss., 1889; 1893, 508; 1893, 61, 84, 95, 98, 99, 102, 191, pl. 1, fig. 16.—Pratt, 1898, 352.—Stoss., 1898, 29.

rufoviride (Rud., 1819) Juel, 1889, 6, 7, 8, 13, 19, 22, 23, 24, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 39, 41, fig. 18.—Barbagallo & Drago, 1903, 409 (in Anguilla vulgaris, Conger vulgaris; Catania).—Braun, 1892a, 711.—Jacoby, 1900, 3.—Lander, 1904a, 7.—Looss, 1894a, 224.—Mont., 1891, 520, 521; 1893, 53, 95, 191, pl. 1, fig. 5.—Par., 1902, 5 (in Anguilla vulgaris. Conger vulgaris, Nettastoma melanura, Rhombus lævis, Saurus griseus, Trigla cuculus).—Stoss., 1898, 29.

scabrum (Mueller, 1788) Juel, 1889, 5.—Mont., 1891, 520.

sluiteri (Brock, 1886) Braun, 1892, 48; 1892h, 729; 1893b, 179.—Looss, 1894a, 234 (to Eurycœlum, type).

stossichi Mont., 1893, 87 (for stossichii).

stossichii Mont., 1891, 501, 502, 512–516, 520, 521, pl. 4, figs. 2, 3, 4, 8, 9, 13, 15a, 16, 17 (syn. Dist. ocreatum Mont., v. 1, p. 87 (in Clupca aurita Cuv., C. pilchardus Wall.); 1891, 19 (D. ocreatum p. p.); 1893, 53, 61, 95, 96.—Barbagallo & Drago, 1903, 410 (stossichi) (in Alosa sardina; Catania).—Jacoby, 1900, 3.—Luehe, 1901, 399.—Ohdn., 1905, 351.—Stoss., 1898, 30.

tornatum (Rud., 1819) Juel, 1889, 6, 29, 31, 34, 40.—Looss, 1896, 125.—Mont., 1891, 516, 520, 521; 1893, 95.

ventricosum (Rud., 1819) Mont., 1893, 191.

[APOLEMA Hulst, see Zool. Rec., 1896, v. 33, Ins. 250.]

APOPLEMA Linst., 1904, 252 (for Apoblema).

APOROCOTYLE Odhn., 1900, 62-66 (m. simplex).

simplex Odhn., 1900, 62-66, 1 fig. (on gills of Pleuronectes flesus and especially Pl. limanda, the latter given as type host; Kristineberg, Sweden).—Mont., 1903, 335.

ASCOCŒLI Burm., 1837, 531.—Braun, 1890a, 515 (includes Gyrodactylus, Hirudinei).—Mont., 1888, 83.—Tasch., 1879, 233.

- ASCOCOTYLE Looss, 1899b, Dec., 584–585, 586, 611 (tod. coleostoma); ὁ ἀσκός, Schlauch; ἡ κοτύλη, Např; 1902m, 441, 824, 832, 833.—Braun, 1902b, 30.— Jægers., 1903a, 14.—Pratt, 1902a, 888, 894 (key).
 - coleostoma (Looss, 1896) Looss, 1899b, 582, 585, 699.
 - minuta Looss, 1899b, 585, 698-699 (in middle portion of small intest. of dogs and cats, in Cairo, Egypt, and in intest. of Ardea cinerea at Damietta; probably contracted from eating fish), 700, 701, fig. 23; 1901, 205.
- ASPIDOBOTHRIA Mont., 1888, 91 (f. for Aspidogaster); 1892, Oct. 7, 195.—Braun, 1893b, 188.
- ASPIDOBOTHRIDÆ Mont., 1888, 51 (Aspidobotridæ), 63, 67 (Aspidobothridæ), 71, 72, 90, 91 (Aspidobotridæ), 103, 107, 108; 1892, Oct. 7, 168 (revision), 195, 196, 197, 198 (diagnosis), 199 (key), 198-209, 213 (f. of Aspidocotylea); 1893, 36, 82, 114.—Braun, 1893a, 886, 888, 890, 891, 894, 895; 1893b, 188.—Gamb., 1896a, 73.—Kofoid, 1899, 179.—Nickerson, 1901, Mar. 8, 378; 1902, 597, 599, 606, 608, 612, 613, 614, 616.—Poche, 1907, 125.—Pratt, 1902a, 887, 891 (key; contains: Macraspis, Stichocotyle, Aspidocotylus, Platyaspis, Cotylogaster, Aspidogaster, Lophotaspis).—Stoss., 1898, 19 (Aspydobothridæ), 1899, 3, 4.
- ASPIDOBOTHRII Burm., 1856a, 252.—Braun, 1890a, 516; 1893b, 188.—Mont., 1888, 84, 107.—Poche, 1907, 125.—Tasch., 1879, 233, 258.
- ASPIDOBOTRIDÆ Mont., 1888, 52, 63, 91 (for Aspidobothridæ); 1892, Oct. 7, 213 (f. of Aspidocotylea).
- ASPIDOCOTYLEA Mont., 1892, Oct. 7, 213 (suborder of Trematoda, for Aspidobothridæ).—Braun, 1893a, 890, 891, 894, 895, 917; 1893b, 188; 1895, 136.—Gamb., 1896, 73.—Kofoid, 1899, 179.—Looss, 1899, 543.—MacCallum, 1902, 636.—Odhn., 1902, 43, 44, 45.—Pratt, 1900a, 645, 646 (included in Digenea of Ben.), 647 (key); 1902a, 887, 891 (key).—Schneidemuehl, 1896, 295.—Ward, 1903, 864.
 - 1896: Aspidokotylea Schneidemuehl, 1896, 295.
- ASPIDOCOTYLE Dies.. 1850a, 288, 413 (m. mutabile) for Aspidocotylus; 1858e, 314, 372.—Braun, 1890a, 546; 1893a, 887, 888, 890, 891, 892, 895, 904 (g. of Amphistomidæ), 907, 918; 1893b, 188.—Burm.. 1856a, 251.—Hoyle, 1890, 539.—Kofoid, 1899, 181.—Looss, 1902, 428.—Mont.. 1888a, 89, 91; 1892, 207; 1892, Oct. 7, 196, 197, 198, 213 (g. of Aspidobothridæ).—Nickerson, 1902, 612.—Pratt, 1900a, 647.—Tasch., 1879, 256, 259–260.
 - 1892: Aspitocotyle Mont., 1892, Oct. 7, 197, misprint.
 - cochleariforme (Dies., 1838) Mont., 1892. Oct. 7, 207–208, fig. 6 (in Cichla temensis; Rio Negro, Brazil) (syn. mutabile).
 - mutabile (Dies., 1839) Dies., 1850a, 413 (int. of Cataphractus n. sp.; Rio Negro, Brazil) (includes A. cochleariformis as syn.).—Braun, 1893a, 907 (in Cichla temensis; Brazil).
 - mutabilis (Dies., 1839) Dies., 1858e, 372 (in Cichla temensis Humboldt).—Tasch., 1879, 256, 260 (syns. Aspidocotylus cochleariformis Dies., A. mutabilis Dies.) (in Cataphractus sp.; Rio Negro).
- [ASPIDOCOTYLEEN Looss, 1902m, 418. German name.]
- ASPIDOCOTYLUS Dies., 1838a, 189; 1839a, 234 (m. mutabilis); 1850a, 413, renamed Aspidocotyle.—Braun, 1892a, 568; 1893a, 879, 884.—Cobbold, 1877f, 326; 1877, 235, 237, 238; 1879b, 360.—Crep., 1841, 82.—Duj., 1845a, 479.—Lejtenyi, 1881a, 2.—Nord., 1840, 601.—Pratt, 1902a, 887, 891 (key).
 - 1850: Aspidocotyle Dies., 1850a, 288, 413 (m. mutabile).
 - cochleariformis Dies., 1838a. 189 (in Cataphractus N. 150); 1839a, 234; 1850a, 413 (syn. of Aspidocotyle mutabile).—Cobbold, 1877. 237.—Mont., 1892, Oct. 7, 207 (to Aspidocotyle).—Tasch., 1879, 260 (syn. of Aspidocotyle mut.).
 - mutabilis Dies., 1839a, 234, pl. 15, figs. 20–22 (in Cataphractus; Rio Negro, Brazil)
 A. cochleariformis, renamed; 1850a, 413 (to Aspidocotyle); 1858e, 372.—
 Braun, 1892a, 580.—Cobbold, 1877, 237, 238; 1879b, 360.—Crep., 1841, 82.—
 Duj., 1845a, 479.—Mont., 1892, Oct. 7, 207 (syn. of Aspidocotyle cochleariforme).—Nord., 1840, 601 (in Cataphractus sp.; Amériq, méridion.).—Tasch., 1879, 260 (to Aspidocotyle).
 - sonsinoii (Cobbold, 1877) Cobbold, 1877e, 238, to Gastrodiscus.
- ASPIDODASTER Mont., 1892, Oct. 7, 213, misprint for Aspidogaster.

ASPIDOGASTER Baer, 1826a, 124; 1827b, 525–557 (m. conchicola); 1828e, 671–678.—Ben., 1858a, 1861a, 11, 172, 345; 1882b, 14, 15, 16.—Bettend., 1897, 339.—Blainv., 1828, 584–585.—Blochmann & Bettend., 1895a, 216, 217.—Braun, 1890a, 480, 515, 516; 1890b, 127; 1892a, 568, 576, 586, 603, 614, 660, 671, 672, 673, 675, 688, 689, 704, 709, 711, 715, 716, 730, 731, 757, 758, 774, 775, 777, 779, 792, 843, 879, 884, 886, 887, 888, 889, 890, 891, 894, 896, 897, 917; 1893b, 177, 178, 179, 187, 188; 1894a, 1149; 1895, 26, 125.—Burm., 1837, 530; 1856a, 240, 252.—Carus, 1863, 478.—Cobbold, 1877, 238.—Crep., 1839, 285–286.—Dies., 1835c, 427; 1839a, 234; 1850a, 288, 414; 1858e, 314, 372–373; 1859c, 438.—Duj., 1845a, 324.—Eichwald, 1829a, 248.—Fraip., 1880c, 418, 443, 445.—Gamb., 1896, 63, 73.—Goldb., 1855a, 18.—Hoyle, 1890, 539.—Jackson, 1888, 642, 643, 646, 654.—Kofoid, 1899, 180, 181, 182, 183, 184.—Lander, 1904a, 5, 10.—Leuck., 1863, 489; 1879, 93, 119, 150, 151; 1886d, 70, 90, 117.—Looss, 1893b, 814; 1894a, 212; 1901, 625; 1902m, 428, 429, 430.—Mont., 1888, 12, 37, 38, 40, 43, 52, 56, 57, 63, 68, 71, 72 (Asptdogaster), 83, 84, 89, 91, 95, 103, 107; 1891, 121; 1892, Oct. 7, 170, 172, 175, 176, 180, 184, 185, 186, 187, 188, 189, 194, 195, 196, 197, 198, 199–205, 199 (syns.: Monost. Rud., 1819a, 87, Aspidonotus Keber, 1851; Cotylaspis Leidy, 1857, 18), 199 (diagnosis), 200 (key), 213 (g. of Aspidobothridæ); 1893, 36, 111.—Moul., 1856a, 235.—Nickerson, 1902, 599, 603, 604, 605, 609, 610, 612, 614, 617, 619.—Nord., 1840, 603 (1. conchicola; 2. limacoides).—Pag., 1857, 6.—Par., 1894, 140.—Poche, 1907, 125.—Poir., 1885, 102; 1886, 20, 21.—Pratt, 1902a, 887, 892 (key).—Raspail, 1829, v. 22, 556–564.—Shipley, 1904, in 77–106.—Staff., 1900, 405, 412; 1903, 824.—Stoss., 1899, v. 19, in 1–6, 1. pl.—Tasch., 1879, 233, 255, 258 (syns. Monost. Rud., Aspidonotus Keber).—Volz, 1899, 232.—Wagener, 1857, 25, 29.—Wallenstedt, 1847, 7.

1851: Aspidonotus Keber, 1851a, 19 (m. conchicola).

1888: Asptdogaster Mont., 1888, 72, misprint.

1892: Aspidodaster Mont., 1892, Oct. 7, 213, misprint.

ascidiæ Dies., 1858e, 373 (in Mentula marina, by Redi), sp. inq., based on Baer, 1827b, 549 (Redi's worm in Ascidia mentula).

cochleariformis (Dies., 1838) Cobbold, 1879b, 360 (syn. of Cotylegaster cochleariform.), lapsus, see Gastrodiscus sonsinoii.

conchicola Baer, 1826a, 124 (in Anodonta, Unio; Prussia); 1827b, 527–557, pl. 28; 1828, 671–678.—Aubert, 1855, 349–376, pls. 14–15 (anat., development).—Ben., 1858a, 1861a, 206, 210.—Bettend., 1897a, 25.—Blainv., 1828a, 584.—Blochmann & Bettend., 1895a, 216.—Braun, 1883a, 41; 1892a, 579, 592, 597, 598, 602, 631, 632, 637, 639, 640, 641, 642, 645, 655, 677, 678, 679, 687, 693, 699, 707, 712, 713, 715, 725, 731, 747, 749, 776, 780, 783, 790; 1893a, 841, 869, 870, 880, 883, 891, 897; 1893b, 183 (in Vivipara unicolor; Cleopatra bulimoides); 1893d, 468.—Burm., 1837, 530.—Crep., 1839, 286.—Dies., 1834a, 1231; 1835c, 423, 427; 1858e, 373 (in Unio purpureus, U. nasutus, U. radiatus, U. cariosus, U. pictorum, Anodonta marginata, A. anatina); 1859c, 438.—Duj., 1845a, 324–327.—Eichwald, 1829a, 248.—Fil., 1855b, 20; 1857c, 22, 27.—Fraip., 1880c, 418.—Gamb., 1896a, 63.—Gronkowski, 1902a, 515 (8) (conchicole).—Hoyle, 1890, 539, fig. 3f, 540 (description).—Jackson, 1886, 650.—Jægers., 1899, 203.—Kath., 1894a, 131.—Keber, 1851a, 19, 90.—Kofoid, 1899, 179, 180, 181, 182, 183, 184, 185.—Kuech., 1855, 184, 185; 1856c, 277.—Leidy, 1851b, 224; 1856b, 45; 1857a, 18; 1858a, 110 (Anodonta fluviatilis and A. lacustris; Phila.); 1877f, 260; 1904a, 53, 88, 108, 110, 111, 148.—Leuck., 1879, 149, figs. 48, 74; 1886d, 115, figs. 48, 74.—Looss, 1894a, 203; 1901, 624; 1902m, 420, 428.—Maclaren, 1904, 609.—Mont., 1888, 40, 43 (conchiola); 1891, 100, 103, 109, 121; 1892, Oct. 7, 172, 186, 188, 191, 195, 197, 198, 200–202, 203, fig. 2 (extensive bibliography); 1893, 111, 114.—Moul., 1856a, 13 (conchycola), 25, 144.—Mueh., 1898, 11.—Nickerson, 1902, 606, 614, 620 (in Unionidæ; Europe and N. America).—Nord., 1832a, 38; 1840, 603.—Odhn., 1902, 44, 45.—Pag., 1857, 35–36, pl. 4, figs. 1–5 (in Anodonta anatina).—Poir., 1886, 21.—Sieb., 1836, 217; 1836, 233; 1837, 263; 1838, 143, 144, 156.—Shipley & Hornell. 1904, 95, 98.—Staff., 1895, 92 July, 282–284; 1895. Oct., 536; 1896, 6 July, 477–542, pls. 36–39 (anat.); 1898, 698.—Steenstrup, 1842, 55.—Stoss.,

conchicole Gronkowski, 1902a, 515, for conchicola.

conchilega Braun, 1883a, 41, for conchicola. conchiola Mont., 1888, 40, 43, for conchicola.

ASPIDOGASTER-Continued.

conchycola Dies., 1850a, 414 (for conchicola).

elegans (Olss., 1868) Mont., 1891, 122 (see 1888, 91); 1892, Oct. 7, 196.

insignis (Leidy, 1858) Braun, 1893a, 897.—Kofoid, 1899, 184.—Shipley & Hornell, 1904, 98.

lenoiri Poir., 1886, 20–22, pl. 1, figs. 1–3 (in Tetrathyra vaillantii; Sénégal).—
Braun, 1892a, 580, 716; 1893a, 888, 891; 1893b, 188.—Kofoid, 1899, 181, 182, 184.—Mont., 1888, 42; 1891, 121, 122; 1892, Oct. 7, 198; 1893, 111.—Nickerson, 1902, 613.—Sons., 1896, 294.—Stoss., 1899, 4.

lenoirii Mont., 1888, 16, 38, 53, 54, 60, 71, 107 (for lenoiri).

limacoides Dies., 1834a, 1231; 1835c, 420–430, 1 pl., figs. 1–19 (in Cyprinus dobula, C. idus); 1850a, 414–415 (intest. of Leuciscus idus, L. dobula); 1858e, 373.—
Braun, 1892a, 580; 1893a, 897.—Crep., 1839a, 286.—Duj., 1845a, 327.—Jackson, 1888, 650.—Kofoid, 1899, 184.—Kroyer, 1846–53a, 462 [671] (in Leuciscus idus).—Looss, 1902m, 420.—Mont., 1888, 71; 1891, 109, 121, 122; 1892,Oct. 7, 172, 195, 200, 202–203, fig. 3; 1893, 114.—Nickerson, 1902, 620 (in Leuciscus sp.; Europe).—Nord., 1840, 603.—Shipley & Hornell, 1904, 98.—Stoss., 1899, 3.—Tasch., 1879, 259 (in Leuciscus idus, L. dobula).—Voeltzkow, 1888, v. 8, 290–292.

macdonaldi Mont., 1891, 120; 1892, Oct. 7, 200, 203–204, fig. 4 [in Melo; Shark Bay, Australia].—Braun, 1893a, 870, 897; 1893d, 468.—Looss, 1901, 625; 1902m, 429, 430 (probably belongs to Lophotaspis).—Nickerson, 1902, 604, 620 (in Melo; Australia).—Shipley & Hornell, 1904, 98.—Stoss., 1899, 3.

macdonaldii Mont., 1891, 121, 122, for macdonaldi.

margaritiferæ Shipley & Hornell, 1904, 78, 90, 95–98, pl. 4, figs. 60–62, 66, 68, 69 (Margaritifera vulgaris; at Cheval Paar, Ceylon); 1905, 55.

ringens Lint., 1905, 327, 333, 367, 397, figs. 243-249 (in Micropogon undulatus, Trachinotus carolinus; Beaufort, N. C.).

sp. Braun, 1892a, 595.

vallei Stoss., 1899, 3-4, figs. 6-8 (in Thalassochelys caretta, at Corfù).—Looss, 1902m, 415, 418 (to Lophotaspis as type) (stomach of Thalassochelys corticata; Egyptian coast), 746.

ASPIDOGASTRIDÆ Poche, 1907, 125.

ASPIDOKOTYLEA Schneidemuehl, 1896, 295 (for Aspidocotylea).

ASPIDONOTUS Keber, 1851a, 19–20, 65, 66, 69, 90 (new name for Aspidogaster, hence type conchicola).—Braun, 1893a, 896.—Mont., 1892, Oct. 7, 199 (syn. of Aspidogaster).—Tasch., 1879, 258 (syn. of Aspidogaster Baer).

conchicola (Baer, 1826) Keber, 1851a, 19, 65, 66, 69, 90; 1854, 45.—Dies., 1858e,
373 (to Aspidogaster).—Mont., 1892, Oct. 7, 200, 201 (to Aspidogaster).—
Tasch., 1879, 259 (to Aspidogaster).

ASPITOCOTYLE Mont., 1892, Oct. 7, 197 (for Aspidocotyle).

ASPTDOGASTER Mont., 1888, 72, misprint for Aspidogaster.

ASPYLOGASTER Mont., 1892, Oct. 7, 187 (for Cotylogaster).

ASTIA Looss, 1899b, 590-591, 592 (tod. renifera) [not Astia Koch, 1879, Arach.; Astea, Asteia, Asteja], ἀστεῖος, feingebildet; 1900, 602 (renamed Astiotrema).—Braun, 1901b, 37.—Pratt, 1903, 34.

impleta Looss, 1899b. 590, 703-705, fig. 26 (intestine of Tetrodon fahaka; in Nile, at Cairo, Egypt).—Braun, 1901b, 37.—Luehe, 1900, 561.

renifera (Looss, 1898) Looss, 1899b, 590.

ASTIOTREMA Looss, 1900, Dec. 3, 602, 607, Astia Looss, 1899 [not Koch, 1879, Arach.], renamed, hence type renifera; 1901, 560; 1902, 487, 505, 821.—Pratt, 1902a, 888, 899 (key).

erinacea (Poir., 1886) Stoss., 1904, 2 (in Delphinus delphis).

impleta (Looss, 1899) Stoss., 1904, 2 (in Tetrodon fahaka).

monticellii Stoss., 1904, 2-3, fig. 3 (in Tropidonotus viperinus; Naples).

renifera (Looss, 1898) Stoss., 1904, 2 (in Trionyx nilotica; Egypt).—Staff., 1905, Apr. 11, 693 (in Trionyx nilotica).

ASTOMUM Schlotthauber, 1860, 129 (m. poricola).

poricola Schlotthauber, 1860, 129 (in Anas boschas).

ASYMPHYLODERA Kowal., 1902d, 26 [8], misprint for Asymphylodora. perlata (Nord., 1832) Kowal., 1902d, 26 (8).

ASYMPHYLODORA Looss, 1899b, Dec., 598-599 (tod. perlata); άσυμφυλος, unlike; ή δορά, skin; 1902, 760.—Luehe, 1900, 487; 1901, 488.—Ofenheim, 1900, 183.—Pratt, 1902a, 888, 902 (key) (related to Haplometrinæ).

1902: Asymphylodera Kowa¹. 1902d, 26, [8].

exspinosa (Hausmann, 1896) Looss, 1899b, 598, 599.

imitans (Mueh., 1898) Looss, 1899b, 598.

perlata (Nord., 1832) Looss, 1899b, 598, 599.—Odhn., 1905, 322.

ATHESMIA Looss, 1899b, Dec., 635-637 (m. heterolecithodes) [not Athetmia 1816, lepidopteron]; άθεσμιος, gesetzlos.—Braun, 1901b, 34; 1901h, 702.—Pratt, 1902a, 889, 904 (key).—Stiles, 1901, 189.

heterolecithodes (Braun, 1899) Looss, 1899b, 635; 1902, 789.—Braun, 1901h, 702.— Engler, 1904, 186.—Hollack, 1902a, 867, 868.

AURIDISTOMUM Staff., 1905, Apr. 11, 690-691 (m. chelydræ); auris, ear.

chelydræ (Staff., 1900) Staff., 1905, 690 (in Chelydra serpentina; Canada).

AXIME Moul., 1856a, 10, misprint.—Cosmovici, 1887a, 127 (for Axine).

AXIME Moul., 1856a, 10, misprint.—Cosmovici, 1887a, 127 (for Axine).

AXINE Abildg., 1794, 59-60 (m. belones) [not Oken, 1835, crustacean].—Ben., 1856e, in 643-654, 1 pl., figs. 1-21; 1858a, 1861a, 11, 52, 345.—Ben. & Hesse, 1864, 6, 96.—Braun, 1890a, 407, 414, 416, 420, 426, 437, 445, 448, 451, 453, 454, 458, 463, 468, 469, 483, 484, 485, 490, 492, 494, 498, 511, 515, 516, 517, 518, 523, 540, 541, 546; 1890b, 125; 1891d, 422.—Burm., 1837, 530; 1856a, 251.—Carus, 1863, 478.—Cerf., 1895h, 91; 1896, 514.—Cosmovici, 1887a, 127 (Axime).—Crep., 1839, 291.—Dies., 1850a, 290, 424, 425; 1858e, 315, 385-386.—Duj., 1845a, 317.—Gamb., 1896, 56, 73.—Goldb., 1855a, 19.—Goto, 1891a, 161, 169, 178, 183, 184; 1891c, 103; 1893a, 798.—Haswell, 1892a, 459; 1892b, 149; 1893e, 114.—Hoyle, 1890, 537, 539.—Ijima, 1884c, 638, 639.—Jackson, 1888, 642, 644, 645, 646, 647, 648, 654.—Juel, 1889, 33.—Kath., 1894a, 143, 152, 155.—Kerbert, 1881a, 573.—Kroyer, 1846–53a, 273; 1852–53a, 1221 (in Belone rostrata Fab.).—Looss, 1885b, 5, 10, 15, 17, 18; 1892, 72.—Lorenz, 1878a, in 405–436, pls. 31–33.—Mont., 1888, 11, 15, 34, 40, 52, 53, 59, 60, 61, 66, 83, 84, Rostata Fab.).—Looss, 18890, 5, 10, 10, 17, 18, 1892, 72.—Lorenz, 1878a, 11 405–436, pls. 31–33.—Mont., 1888, 11, 15, 34, 40, 52, 53, 59, 60, 61, 66, 83, 84, 86, 89, 101; 1893, 111; 1903, 336 (subf. Axininæ).—Moul., 1856a, 10 (Axime).—Par. & Perugia, 1890, 13.—Pratt, 1900a, 646, 653 (key), 657, fig. 40.—St.-Remy 1898, 562–563.—Stoss., 1898, 14.—Tasch., 1879, 40, 45, 56, 58, 61, 66, 69; 1879, 233, 255–256 (syn. Heteracanthus Dies.).—Wallenstedt, 1847, 7.— Ziegler, 1883, 557.

1856: Axime Moul., 1856a, 10.

aberrans Goto, 1894a, 198-199, pl. 7, figs. 5-6 (in Belone schismatorhynchus; Hagi,

Japan).—St.-Remy, 1898, 563.

belones Abildg., 1794d, 59-60, pl. 6, fig. 3, a-b (in Belone acus).—Ben., 1856a; 1856e; 1857i; 1858a, 1861a, 52–54, 168, 169, 196 (in Esox belone); 1870, 80.—Blainy., 1828a, 568.—Braun, 1889k, 621; 1890a, 418, 424, 428, 436, 437, 443, Blainv., 1828a, 568.—Braun, 1889k, 621; 1890a, 418, 424, 428, 436, 437, 443, 472, 477, 488, 494, 514, 541, 549, 550; 1891a, 52.—Crep., 1838b, 83-96 (syns. Heteracanthus pedatus, H. sagittatus); 1839a, 291; 1839, 163.—Dies., 1850a, 425 (syns. Heteracanthus pedatus, H. sagittatus); 1858e, 385-386; 1859c, 444.—Duj., 1845a, 317.—Goto, 1894a, 196.—Hoyle, 1890, 539, 540.—Juel, 1889, 36.—Kath., 1894a, 155.—Kroyer, 1846-53a, 273 (in Belone rostrata).—Lorenz, 1878, —.—Mont., 1888, 8, 15; 1890, 422; 1893, 111.—MoquinTandon, 1846, 13, 395.—Nord., 1840, 598 (syn. of Heteracanthus pedatus Dies.) (in Esox belone).—Oken, —, 357, pl. 10.—Par., 1902, 3 (in Belone acus, Exocœtus volitans; Portoferrajo).—Par. & Perugia, 1890, 8.—Sieb., 1839, 163.—Stoss., 1890, 44; 1890, 52; 1898, 14.—Tasch., 1879, 35; 1879, 256 (syns. Axine orphii Ben. & Hesse, Heteracanthus pedatus, H. sagittatus Dies.) (in Belone acus).—Ziegler, 1883, 545.—Also reported for Belone vulgaris. (in Belone acus).—Ziegler, 1883, 545.—Also reported for Belone vulgaris.

heterocerca Goto, 1894a, 197–198, pl. 7, figs. 1–4, pl. 8 (on Seriola quinqueradiata; Japan).—Pratt, 1900a, 657, fig. 40.—St.-Remy, 1898, 563.

orphii Ben. & Hesse, 1863, 116; 1864, 116-117, pl. 12, figs. 19-27 (in Esox belone).-Braun, 1890a, 418.—Tasch., 1879, 256 (syn. of A. belones).—Also reported for Belone vulgaris.

platyura Crep., 1838b, 83 (syns. A. belones, Heteracanthus pedatus, H. sagittatus).— Sieb., 1839, 163.

triangularis Goto, 1894a, 200–201, pl. 7, figs. 7–8 (on Acanthias schlegelii; Misaki, Japan).—St.-Remy, 1898, 563.

triglæ Ben. & Hesse, 1863, 117; 1864, 117 (Trigla hirundo).—Braun, 1890a, 418, 541, 549 (Brest), 552.—Tasch., 1879, 256 (in T. hirundo).

AXININ.E Mont., 1903, 336 (subf.; f. Microcotylidæ).

AZIGIA Staff., 1904, May 3, 488, misprint for Azygia.

tereticolle (Rud., 1802) Staff., 1904, 488 (Esox lucius, Lota maculosa, Ameiurus nigricans; Canada).

AZYGIA Looss, 1899b, Dec., 569–570, 579, 580 (m. tereticollis); ἄζυξ, ἄζυγος, unverbunden; 1902, 839.—Braun, 1901b, 33.—Darr, 1902, 667.—Luehe, 1900, 489, 490.—Marshall & Gilbert, 1905, 477, 484.—Odhn., 1905, 328.—Pratt, 1902a, 888, 897 (key).—Staff., 1904, May 3, 483, related to Otodistomum, 489.—Stoss., 1904, 199.

1904: Azigia Staff., 1904, May 3, 488, misprint.

loossii Marshall & Gilbert, 1905, 483–485, pl. 15, figs. 5–7 (Micropterus salmoides, Lucius lucius, Amia calva).

tereticollis (Rud., 1802) Looss, 1899b, 570; 1901, 119; 1902, 456, 457.—Cohn, 1902k, 47.—Darr, 1902, 688.—Heymann, 1905, 85.—Kowal., 1902d, 26 (8) (in Perca fluviatilis); 1904, 24 (9) (in Esox lucius; Galicia).—Luehe, 1900, 491.—Marshall & Gilbert, 1905, 484, 485.—Odhn., 1905, 363.—Staff., 1904, 488.—Ward, 1903, 861.—Reported for Esox lucius L. Lota maculosa Le S., Ameiurus nigricans Le S.

BALANORCHIS Fischder., 1901, 374–375 (m. anastrophus); 1902a, 49–50; 1903h, 490.—Pratt, 1902a, 887, 893 (key).—Shipley, 1905, 8.

anastrophus Fischder., 1901, 375 (Cervus dichotomus); 1902a, 50–52, fig. 4 (syn. Amphist. conicum Dies., 1835 e. p.) (in C. dichotomus; Brazil).

BARIS Looss, 1899b, Dec., 669–670 (m. proteus) [not Baris Germ., 1817, coleopteron]; η $\beta \alpha \rho \iota \tau$, ägyptischer Nachen; 1900d, 602 (renamed Deuterobaris).—Stiles, 1901, 189.

proteus (Brand., 1891), Looss, 1899b, 669, 770–772, fig. 82.—Braun. 1901b, 38, 54. BATHYCOTYLE Darr, 1902a, 644–662 (m. branchialis). 678, 687, 691, 696; 1902b.

branchialis Darr, 1902a, 644-662, pl. 23, figs. 1-15 (in mackerel; Pemba, German East Africa); 1902b, 735-736.

BENEDENIA Dies., 1858e, 313, 363–364 (m. elegans=sciænæ; in Sciæna aquila; Ostend) [not of Gray, 1864, mammal; not of Schneider, 1875, protozoon]: 1859c, 437.—Braun, 1890a, 518.—Mont., 1888, 87; 1902, 143.—Odhn., 1905, 371.—Tasch., 1878, 566 (syn. of Trist. Cuv.).

elegans Dies., 1858e, 364 (new name for Epibdella sciænæ in Sciæna aquila; prope Ostendam); 1859c, 437.—Sons., 1891, 263.—Tasch., 1878, 565.

hendorffi (Linst., 1889) Linst., 1903, 355 (in Coryphæna). Epibdella (Benedenia). ishikawa (Goto, 1894) Linst., 1903, 356, E. (Benedenia), from Lethrinus.

monticellii (Par. & Perugia, 1895) Linst., 1903, 356, E. (Benedenia), from Mugil. orata (Goto, 1894) Linst., 1903, 356, E. (Benedenia), from Anthiæ.

sciænæ (Ben., 1856) Linst., 1903, 355, E. (Benedenia), from Sciæna.

BILARZIA Mazzei, 1905a, 657, for Bilharzia.

BILHARCIA de Bonis, 1882, 115, for Bilharzia.

BILHARTZIA Sons., 1877, 652, for Bilharzia.

BILHARXIA Semprum, 1890, 596, for Bilharzia.

(BILHARZIA) written as subg. of Dist. by some authors, as Bomford, 1887a, but probably not intended as a subg. in all cases.

BILHARZIA Cobbold, 1859d, 364 (m. hæmatobia); 1876, 488, 756; 1879, 17, 39, 42, 44, 45, 47, 48, 49, 52; 1883i, 106–107, figs. 1–2; 1885a, 498–500, figs. 211–213.— Alexander, 1906, Mar. 1, 69.—Beach, 1899, 103.—R. Bl., 1888a, 541, 543; 1888u, 832, 833, 1 fig.; 1904, 153.—Brand., 1892, 507.—Braun, 1891d, 426; 1892a, 573, 574, 603, 698, 707, 715, 735, 760, 764, 765, 788, 790; 1893a, 878, 879, 882, 885, 886, 890, 893, 894, 895, 908, 912, 918; 1895b, 125, 127, 129, 136, 151; 1900h, 3; 1901, 562; 1902b, 140, 142; 1903, 3 ed., 168.—Brock, 1893b, 622–625.— Catto, J., 1904, Nov. 19, 1411 (new species of).—Chatin, 1887, 1003–1006; 1887, 595.—Coe, 1896a, 562, 564, 566, 567.—Darr, 1902a, 649.—Fischer, 1883a, 41.—Fritsch, 1888a, 192–223, 1 fig., pls. 11–12, figs. 1–14 (anat.): 1888b, 588.—Gamb., 1896a, 73.—Grassi & Rovelli, 1888c, 799.—Hahn & Lefèvre, 1884a, 515; 1884, 805, 806.—Harrison, 1889, 163.—Higgins, 1906, 887.—Hoyle, 1890, 539.—Huber, 1894, 297.—Jackson, 1888, 644, 646, 648, 649, 654.—Juel, 1889, 37.—Kartulis, 1885e, 364, 1 fig.—Kholodk, 1898, 23, 25, 32.—Kowal., 1895g, 2, 3, 5, 15, 23, 24, 25, 42, 43, 45, 55, 63, 64, 65; 1896g, 70; 1896i, 8 [352].—

BILHARZIA—Continued.

Leuck., 1863, 617.—Looss, 1892, 81, 82; 1894a, 179; 1896b, 2, 167; 1899b, 536, 538, 542; 1901, 196.—Moniez, 1896, 83, 154.—Montgomery, 1906, Feb. 12, 18.—Mont., 1888a, 7, 11, 15, 34, 37, 51, 61, 62, 84, 92, 105; 1892, Oct. 7, 214; 1893, 43, 82, 87, 94, 107, 154; 1896, 162, 163.—Sambon, 1899, v. 2, 117–120, 2 figs.—Schneidemuehl, 1896, 295.—Shipley, 1905, v. 6 (1), 4.—da Silva Lima, 1877, Nov., 489.—Sons., 1874, 71–83; 1882, June 10, 620–621; 1890, 134.—Stiles, 1898, 60; 1903, 77.—Stiles & Hass., 1898a, 83, 93, 94.—Villeneuve, 1892, 153–157.—Vogt, 1878, 9, 14.—See also Schistosoma. For additional medical papers on infections with species of Bilharzia, see also Bilharziosis.

1877: Bilhartzia, Sons., 1877, 652. 1882: Bilharcia de Bonis, 1882, 115. 1890: Bilharxia Semprum, 1890, 596. 1905: Bilarzia Mazzei, 1905a, 657.

bovis Sons., 1876, 84–87, fig. (in Bos taurus; Egypt); 1876, 280; 1876, 400; 1876, 594–595; 1876, Oct. 18, 631–632; see Gynæcophorus crassus; 1877, 651.—Barbagallo, 1899c, 279.—R. Bl., 1888a, 650.—Bomford, 1887a, 54; 1887b, 345–346.—Braun, 1892a, 763; 1893a, 912; 1895, 154.—Cobbold, 1876, 489; 1876m, 781; 1879b, 332.—Dav., 1877, 943.—Grassi & Rovelli, 1888.—Huber, 1894, 302.—Kowal., 1895g, [3] 43 (syn. of B. crassa).—Laveran & R. Bl., 1895, 101–104.—Stiles, 1898a, 60.—Stoss., 1892, 6 (syn. of Gynæcophorus crassus).—Ward, 1895, 332 (in Bos taurus), 335 (in Ovis aries).

capensis Harley, 1864a, 63 (in Homo; Cape of Good Hope).—R. Bl., 1888a, 644.— Bourel-Roncière, 1888a, 130.—Cobbold, 1872b, 89 (syn. of B. hæmatobium).— Dolley, 1894a, 982.—Dav., 1877, 941.—Gues, 1879a, 169.—Stiles, 1898a, 58.—Ward, 1903, 872.

crassa Sons., 1888, 124, 125; 1895, 124; 1896, 79; 1896, 309, 318.—Barbagallo, 1899a, 10 pp. (in Sicilia); 1899b, 15–16; 1899c, 277–285.—R. Bl., 1888a, 650.—Braun, 1893a, 876, 882; 1893d, 467; 1902b, 143.—Cobbold, 1885a, 498, 499, 500, fig. 213; 1885b, 985.—Colloridi, 1891a, 856.—Gamb., 1896, ——Gomy, 1897a, 377.—Kowal., 1895g, [3, 4], 43, 44 (syn. B. bovis); 1896g, 64; 1896i, [8], 352; 1897b, 2, 3; 1898h, 150, [47].—Looss, 1892a, 81; 1895, 3, 12, 13, 32, 38, 39, 48, 98; 1896b, 159.—Moniez, 1896, 159, 168.—Sanfelice & Loi, 1896, Sept. 12, 305–307.—Stiles, 1898a, 60.—Ward, 1895, 332 (in Bos taurus), 335 (in Ovis aries).

ematobia Sons., 1884, 20, 21.

endemica Sons., 1884, 17–21 [not used as specific name, but in the sense of Bilharzia, endemic in, etc.].—Huber, 1894, 303.

hæmatobia (Bilharz, 1852) Cobbold, 1859d, 364; 1864g, 157; 1866, 6; 1872b, 89–92, figs. a-w; 1872c, 636–646, figs. a-u; 1876m, 780; 1876h, 211, 212; 1876n, 488; 1879b, 38–56, 181, 185, 289, figs. 7–16, 19; 1880e, 59; 1882, Jan., 2; 1882c, 84; 1882f, 272; 1883i, 107, figs. 2–3; 1883, 401; 1885a, 498, 499, 500, figs. 211–212 (syns. B. magna, Dist. hæmatobium); 1885b, 985.—Agnew, 1881, 538.—Aitken, 1866, 804, 839–841; 1872, 146.—Albany M. Ann., v. 9, 202.—Albarran & Bernard, 1897a, 645–647; 1897b, 1096–1123, pl. 11.—Allen, 1882, July 15, 51–53; 1883, 15, 660–661; 1888, 310.—Almeida Couto, 1872, 4, 5, 6, 7, 10, 13, 14, 16, 17, 18, 24, 30, 42.—Auden, 1904, Apr. 30, 1235.—Barbagallo, 1899c, 277, 278, 279, 282.—Batho, 1872a.—Bays, 1901a, 825.—Belleli, 1885b, 54–56, figs. 19–20.—Ben., 1878a, 276.—Birch-Hirschfield, 1889, 302.—Birt, 1904 (10), 421–422.—Bizzozero, 1883, 220, fig. 80.—R. Bl., 1887a, 500–507; 1888a, 636–652, figs. 330–335 (syns. Dist. hæm., Gynækophorus hæm., Schist. hæm., Thecosoma hæm., Dist. capense); 1888e, 193; 1888r, 51; 1890, 54; 1891p, 604–615.—de Bonis, 1882, 115.—Booth, 1882a, 81–84.—Bowlby, 1889a, 891; 1889b, 786; 1891a, 136; 1891b, 194–195.—Brault.—Braun, 1891d, 426; 1892a, 571, 572, 602, 606, 640, 673, 674, 740, 764, 765, 766, 778, 783, 785, 876, 882, 911; 1893a, 912, pl. 25, fig. 10; 1893b, 187; 1893d, 467; 1895b, 152–154, figs. 69–71.—Brock, G. S., 1893a, 52–74, pls. 8–10, figs. 1–17.—Brocks, 1897a, 492–493.—Bull. med., 1888, v. 2, 918; 1890, v. 4, 281.—Buttel-Reepen, 1900a, 590; 1902, 188, 191, 193.—Cadet, 1876.—Calvert, 1902c, 1523 (hematobia).—Chaker, 1890a.—Chatin, 1881, 11; 1887a, 595–597; 1887, 129; 1887, 1003.—Childe, 1899a, 644; 1899, 318.—Chute, 1888a, 85–87, 89–91, figs. 1–8.—Coles, 1902a, 1137–1138.—Colloridi, 1891a, 854–864.—Cortet, 1893.—da Costa, 1884, 935.—Crevaux, 1874a, 177.—Crocker, 1883a, 25–28.—Cureton & Webb, 1899a, 156 (from S. Africa).—Curtis, 1896a, 56–58.—Damaschino, 1882, 949.—Dav., 1877, 909, 952.—Davies, 1884, 181.—Deblenne, 1885.—Dight, 1887a, 420–424

BILHARZIA—Continued.

(hæmatobium).—Dunglison, 1893, 142, 502, 518, 820.—Ebstein, 1884, 45, etc.—Ensor, 1904, Nov., 575–576.—Eve, 1888a, 184.—Fenwick, 1888a, 344—346.—Fenwick & Harris, 1888a, 183–184.—Fouquet, 1885a, 677–680, 693–696.—Fritsch, 1885, 30.—Gamb., 1896, 4, 63, 68–70, fig. 34.—Gautrelet, 1885a, 577–579; 1885, 138.—Gozalez, 1904, 193–194 (hematobium).—Grassi & Rovelli, 1888, 799; 1888d, 284.—Gues, 1879a, 169–175.—Guillemard, 1882; 1883a; 1884, 4; 1894a.—Gutch, 1900a, 1222.—Harley, 1864a, 62, 63 (to Dist.).—Harrison, 1889a, 163.—Harrley, 1887a, 214.—Hatch, 1875a, 874–875, figs. 1–2 Harrison, 1889a. 163.—Hartley, 1887a. 214.—Hatch, 1878a, 874–875, figs. 1–2; 1887a, 875; 1887b, 760–761; 1903a, 772 (in India); 1903b, 225.—Handford, 1894, 48.—Herff, 1894, 415 (syn. Dist. bilharzii).—Hertwig, 1892.—Hirsch, 1887a, 875; 1887b, 760–761; 1903a, 772 (in India); 1903b, 225.—Handford, 1894, 48.—Herff, 1894, 415 (syn. Dist. bilharzii).—Hertwig, 1892.—Hirsch, 1883, 206–209.—Hoyle, 1890, 538–540, fig. 4a.—Huber, 1896a, 580–582.—Ijima, 1889b, 155.—Jackson, 1888, 653.—Jacoby, 1900, 3.—Jaksch, 1889, 42; 1892, 122.—Jamieson, 1897b, 147.—J. Trop. M., 1905, 335.—Kartulis, 1885e, 364, 1 fig.—Keating, —, 768–773, fig. 7.—Kholodk, 1898, 32–33, pl. 11, figs. 22–25.—Kowal, 1895g, [3, 4, 6, 7, 8, 9, 10, 11, 12, 13, 15, 16, 17, 18, 19, 20, 23, 24, 25, 26], 43, 44, 46, 47, 48, 49, 50, 51, 52, 53, 55, 56, 57, 58, 59, 60, 63, 64, 65, 66; 1896g, 64, 66, 67, 68, 70, 71; 1896i, [4, 7, 8, 9, 10], 348, 351, 352, 353, 354; 1896l, 146, 147; 1898g, 75; 1898h, 150, 151, 152, 155, 158, [47, 48, 49, 52, 55].—Kuech., 1879, 340–354.—Kuech. & Zuern, 1881, 340, pl. 8, fig. 13.—Lawson, 1904a, 263–271, figs. 1–6.—Leuck., 1876, 873; 1893, 464–534, fig. 231.—Lewis, 1900a, 1057–1058.—Lockwood, 1901, 2. ed., 821.—Looss, 1892a, 82; 1896b, 2, 64, 158–166, 167, pl. 11, figs. 107–114; 1905, 100.—Lortet & Vialleton, 1894, 8.—Lyle, 1883, 113.—Mackie, 1882, Oct., 661.—Maddlen, 1899.—Manson, 1901, 542 (syn. Dist. hæm.); 1903, 3. ed., 605–616, 639, figs. 86–89.—Mitchell, 1870a, 3.—Moniez, 1896, 170.—Montgomery, 1906, Feb., 18.—Mont., 1888, 9, 15, 17, 18, 67, 68, 69; 1893, 27, 86.—Monvenoux, 1884.—Moore, 1885, 89.—Moty, 1893, 51.—Nachtigal, 1879, 151.—Napier, 1887, 460; 1890, 88–91; 1892.—Neumann, 1888, 547–548, fig. 272; 1892, 2. ed., 610–611, fig. 321; 1892, 632–633, fig. 321.—N. Y. M. J., 1896, v. 64, 586.—Nitze, 1891, Feb. 16.—Num, 1888, 407–410.—Packard, 18—. 523.—Pariera, 1877.—Parker, 1889, 71; 1891, 80, 124, 683.—Perroncito, 1882, 279–283, figs. 122–123—Petrie, 1903, July, 133–134.—Rail., 1892, 161; 1893a, 371–375, figs. 245–246.—Rathelot, 1892.—Rathery, 1907, 48.—Ratz, 1899, 617.—Rawitz, 1893, 858–859.—Rendall, 1904, Mar., 755–756.—Roberts, 1891, 468.—Ross, 1904, 687.—Rossbach, 1906, 414.—Ruault, 1885, 56.—Ruettimeyer, 1892, 144.—Seligmann, 1897, Dec. 834—Sewe Rogers, 1901, 95.—Rosenstein, 1892; 1894, 654.—Ross, 1904, 687.—Rossbach, 1906, 414.—Ruault, 1885, 56.—Ruetimeyer, 1892, 144.—Seligmann, 1897, Dec., 834.—Sewell, 1904, Nov., 575.—Shattock, 1891, 196.—Shaw, 1901, 19.—Silva Araujo, 1877, Nov., 499.—Silva Lima, 1877, Sept., 388, 389.—Simon, 1897, 99.—Simpson, 1871, 212-215; 1872, Mar., 123.—Sons., —, 743; 1874, 502; 1874, 71; 1874, 305-321; 1875, 738; 1876, 3-17, 33-46, 1 pl., figs. 1-9; 1876, 233-235; 1876, 376; 1876, 283; 1877, 297-307; 1877, 651; 1878, 611; 1879, 9; 1884, 365; 1885, 1197; 1887; 1888, 119-127; 1894, 754; 1896, 79; 1896, 287, 295, 298-299, 307, 309, 311, 314, 315, 320-329; 1897, 259.—Stevenson, 1892, 274.—Stoss., 1892, 5 (to Gynæcophorus).—Symmers, 1905, v. 168, 1138.—Thurn, 1882.—Tyson, 1903, 3rd, 1181.—Ultzmann, 1878, 34.—Unione M. egiz., 1884-5, 1. nos. 20-21.—Verrill, 1870, 170.—Villeneuve, 1891, 321; 1891, 556; 1891, 398; 1892, 153, etc.—Virchow, 1888, 368; 1891.—Vegt, 1878, 10, fig. 1, 36.—Walker, 1900, 651; 1900, 390-392; 1905, 207-209.—Ward, 1895. 10, fig. 1, 36.—Walker, 1900, 651; 1900, 390–392; 1905, 207–209.—Ward, 1895, 253, 327; 1903, 872 (to Schist.).—Wortabet, 1880, 603; 1882.—Worth, 1904, Feb. 27, 611.—Zancarol, 4882, 410.—Zuelzer, 1894, 205.—Zuern, 1882, 220.

anatomy and histology of: Brock, 1893a; 1893b; 1894a.—Chatin, 1887a, 595—597; 1887b; 1887c; 1887d (excr., genital organs).—Fritsch, 1885a, 407, 411; 1888a, 192–223, pls. 11–12; 1888b, 588.—Looss, 1895c, 1–108, figs. 1–30; 1896a, 329–332.

——, eggs of: Belleli, 1885a, 1–3 (in lungs).—Brock, 1893b, 622–625, figs. 1–6, 1894a, 774.—Cahier, 1892a, 570–576, figs. 1–4 (egg and embryo).—Chatin, 1884d, 364–365; —, 740.—Le Dantec, 1904, Dec., 399–400.—Jones, 1893a, 145 (in hematuria).—Kartulis, 1885a, 139–145, figs. 1–4 (in pelvic organs); 1885b, 188–189.—Looss, 1893, 521–528; 1905, 101–102.—Low, 1903b, Feb., 67–68; 1903l, May, 232.—Manson, 1901, 542; 1903, 3. ed., 606–607, 613, fig. 87.—Mazzei, 1905, 659 (in urine and feces); 1905, Aug., 550; 1905, July, 1083.—Milton, 1902.—Seligmann, 1898.—Smith, 1905, Oct., 656–659.

BILHARZIA—Continued.

, life history of: Cobbold, 1872b, July, 89–92.—Lazarus-Barlow & Douglas, 193a.—Looss, 1894c, 286–292, 340–346; 1905, 903.—Manson, 1903, 3. ed., 608.—Sons., 1884, 380–394, 1 pl., figs. 1–6; 1888, 100–101; 1893, Sept., 621–622; 1893, Oct., 466–467; 1893, Nov., 707–708; 1893, 997–1000; [1894, 9–10;] [1894, Jan., 10–14;] 1894, Nov., 754–755.

hæmatobia crassa (Sons., 1877) Kowal.. 1894; 1895g, 18, 19, 27 [58, 59, 67], pl. 2, fig. 2 (in Bos taurus, Ovis arics; Africa, Sicily).—Stiles, 1898a, 60.

hæmatobia hominis (Dies., 1855) Kowal.. 1895g, 18, 19, 26–27 [58, 59, 66–67], pl. 2, fig. 1 (in Homo; Africa).—Stiles, 1898a, 58.

hæmatobia magna (Cobbold, 1859) Kowal., 1895g, 27 [67] (in Cercopithecus fuliginosus; Africa?).—Stiles, 1898a, 58.

hamcatobia crassa Kowal., 1895g, 18 [58], for hæmatobia.

hæmatobia Bourel-Roncière, 1878a, 116, 117, for hæmatobia.

kowalewski (Rail., 1899) Par., 1902, 6, for kowalewskii.

kowalcwskii Par. & Ariola (1896), 114-116 (1-3), 1 fig. (in Larus melanocephalus); 1896, Oct. 24,620.—Kowal., 1896i, 9, 353.—Looss, 1899b, 658 (to Bilharziella).— Par., 1899, 7, 1 fig.; 1902, 6 (kowalewski) (in Larus melanoc.).—Sons., 1896, 319.

magna Cobbold, 1859d, 364, pl. 63, fig. 8-9 (in Cercopithecus fuliginosus); 1861, 118; 1872b, 89 (syn. of B. hæmatobia); 1885a, 498 (syn. of B. hæmatobia).— Braun, 1893a, 876, 911; 1893d, 467.—Colloridi, 1891a, 854 (syn. of B. hæm.).—Dav., 1877, 942.—Leuck., 1863, 619.—Kowal., 1895g, 3 [43, 64].—Moniez, 1896, 155.—Sons., 1888, 124.—Stiles, 1898a, 58, 59, 138.—Stoss., 1892, 6 (to Gynæcophorus).

magnum of Montgomery, 1906, Feb., 12, 45, for magna.

ovis, see Cobbold, 1885a, 499 (in Ovis).

polonica Kowal., 1895g [1-27], 41-70, pl. 2, 20 figs. (in Anas boschas fera, A. creeca L.; Dublany); 1896d, 4 (254); 1896g, 63-72; 1896h, 265-266; 1896i; 345-356, pl. 12, figs. 1-9; 1896k, 345-356, pl. 12, figs. 1-9; 1896l, 145-148, 1896m, 145-148; 1897a, 41-70, pl. 2, figs. 1020; 1897b, 198-200 (1-3); 1898g, 75-77, figs. 30-36 (in Anas acuta, A. boschas dom., A. cinerca, A. querquedula, D. Caraco, 1898, Dublany, Poturzyca, Gênes); 1898h, 135, 148-158 (32, 45-51, 52, 54, 55), figs. 30-36.—Barbagallo, 1899c, 281.—Looss, 1899, 567, 658 (type of Bilharziella).—Sons., 1896, 318; 1896, Mar. 1, 79-80; 1897, Jan. 17, 198-200.—Sons. & Kowal., 1897, 3 pp.—Also reported for Ardea cinerca, Mergus albellus.

BILHARZIELLA Looss, 1899b, 543, 657-658 (tod. polonica); 1902, 746.—Braun, 1901, 562; 1902b, 149, 142; 1903, 169 (in birds).—Darr, 1902, 660.—Montgomery, 1906, Feb. 12, 18.—Pratt, 1902a, 889, 907 (key).

?canaliculata (Rud., 1819) Braun, 1902b, 142-146, figs. 58-88 (includes Dist. canaliculatum Rud., 1819a, 676; Duj., 1845a, 449; and Dist. canaliculatum (Rud.) Dies., 1850a, 346; Stoss., 1892, 36; Braun, 1901, 562).

kowalewskii (Par. & Ariola, 1896) Looss, 1899b, 658.—Braun, 1902b, 140.

polonica (Kowal., 1895) Looss, 1899b, 543, 658.—Braun, 1902b, 140, 141, 145.— Kowal., 1902d, 28 (10) (in Anas querquedula, A. acuta, A. crecca, A. boschas, A. boschas dom., Mergus albellus); 1904, 8 (23) (in Fuligula leucophthalmos; Dublany).—Montgomery, 1906, Feb. 12, 18.

pulverulenta Braun, 1901g, 946-947 (in Anas querquedula; Dongola); 1902b, 140,

141 (in A. quer.).

BILHARZIOSIS (name of disease): Allen, 1882, July 15, 51-53; 1888, Apr., 310-320 (parasitic hæmaturia).—Ashford, King & Igaravidez, 1904, Dec., 91 (in Porto Rico).—R. Bl., 1904, Feb., 153;—Chute, 1888a.—Douglas & Hardy, 1903, Harley, 1864a, 55–12; 1869a, 379–387 (endemic hiematuria).—Harch, 1903a, 772 (in India); 1903b, 225.—Kanellis, 1904b, 45–51 (bilious hemoglobinuric fever).—Katsurada, 1904e, Aug. (endemic disease).—Kautsky, 1903a, 649–652.—Lawson, 1904a, Sept.. 263–271.—Leão, 1897a, 337–366, figs. 1–2, pl. 2, figs. 1–3; 1898, 504.—Letulle, 1905b, 37.—Looss, 1903a. 32; 1905, 93.—Lortet, 1893a, 618–620.—Lortet & Vialleton, 1894a, 1–111, pls.. figs.; 1895a, 265–269.—Lyle, 1883, Jan., 9 (endemic hæmaturia).—Madden, 1902, June, 165; 1903, 1; 1903, 866; 1904, 73–80, figs, 1–6.—Manson, 1901, 542 (endemic hæmaturia);

RZIOS1S—Continued.

1903, 3. ed., 605–616, figs, 86–88.—Martinez, 1904, Dec., 193–194; Milton, 1902c, 165–170, 191–192, 200–203, 213–219 (3 lectures); 1904a, 107–122, 1 pl.—McL., K., 1903, June, 255.—Montgomery, 1906a, 15–46, 2 pls. (in animals in India); 1902b, 32 pp., 2 pls.; 1906c, 138–174, pls. 1–2; 1906d, 37 pp., 2 pls.; 1906e, 164; 1906f, 531–532; 1906g, 834; 1906h, 251–252, 259–260; 1906i, 564–565.—Posnett, 1901, 318.—Rankin, 1904, July, 354; 1904, July, 210.—Rochard, 1871, 298.—Reutimeyer, 1894.—Roux, 1906. 105–107.—Sandwith, 1904, Oct., 460–477; 1904, Nov. 15, 360; 1904, Dec. 6, 2200.—Scheiss-Bey, —, 303.—Scott, 1904, 725 (in Persia).—Semaine méd., 1903, 3.—Sewell, 1904; 1904, Nov., 360 (in India).—Sons., 1876, 650 (Egypt).—Villeneuve, 1891, 398, 421 (in Tunisie); 1891, 556.—Virchow, 1888, 361–385.—Voorthius, 1905, 1665–1672.—Wortabet, 1880, Sept., 578.

—, cases of: Anders & Callahan, 1905, 509–519, figs. 1–2.—Benham, 1906, 922.—
R. Bl., 1904, Dec., 148–151; 1906, Jan. 18, 662–663.—Booth, 1882, 81–84.—
Bowlby, 1889, Apr. 20, 891.—Brault, 1891a; 1891c; 1891, 409; 1892a.—
Brooks, 1897a, 492–493; 1897, 617.—Cahier, 1894, Apr., 363–365.—Childe, 1899a, 644; 1899b, 318; 1899c, 598.—Cobbold, 1878h, 358.—Cortez. 1905, 97–103.—Crocker, 1883a, 25–28.—Cureton & Webb, 1899a, 156.—Davies, 1884, 181.—Le Dantec, 1904, Nov., 399–400.—Douglas & Hardy, 1903, Oct., 1009–1012; 1903, Oct., 476; 1903, Oct., 259; 1903, Oct., 835–866; 1903, Nov., 1032; 1903, Nov., 735; 1903, Dec., 263; 1904, Jan., 34; 1904, Jan., 60; 1904, Oct., 476.—Duncan, 1902–03, 187–189.—Fenwick, 1888, 344–346.—Freeman, 1905, 145–148.—Frothingham, 1904, 453–457, 1 pl.—Gautrelet, 1885a, 577–579.—Gunn, 1906, Apr., 1031–1032.—Gutch, 1900a, 1222.—Handford, 1887b, 240–245, figs. 1–3; 1889a, 424–425; 1894a, 48–49.—Hillmantel, 1893a, 230–233, figs. 1–3; 1893b, 4 pp., 3 figs.—Jones, 1893a, 145.—Lelean, 1904f, 425–428.—Letulle, 1904a, 48; 1905a, 420–421.—Lewis, 1904a, 345–346.—Lillie, 1901a, 212.—Madden, 1899, June, 1716; 1899, July, 114; 1901. May., 143–144.—Manson, 1902, Dec., 384–385; 1902, Dec., 1894–1895; 1903, Jan., 15; 1903, Jan., 121; 1903, Feb., 15; 1903, June, 293.—Moore, 1885, 89–92.—Nitz, 1891, Jan. 28.—Pool, 1903, Apr., 632–633; 1903, June, 1146.—Rafferty & Rafferty, 1904, June, 918–919. 6 figs.; 1904, June, 1148–1149; 1904. June, 1661; 1904, Nov., 523.—Sewell, 1904, Mar., 346–347; 1904, Nov., 360.—Simpson, 1872, 320–321.—Smith, 1905, Oct., 656–659, figs. 1–4; 1905, Oct., 1359; 1905–06, 515–518.—Sondern, 1897, May, 554–557, figs. 1–6 (see case of Brook, 1897, 492).—Symmers, 1906, Jan. 18, 662; 1906, Jan. 7, 22.—Tottenham Posuett, 1901, Oct., 318–319.—Villeneuve, 1891, 321–324.—Walker, 1900, Feb., 390–392, 8 figs.; 1900, 651.—Wallace, 1901, Feb., 121–124; 1901, Feb., 304–305; 1902, Mar., 203.—Wardrop, 1906, Sept., 282–283.—Webb, 1899, Jan., 156.—Wortabet, 1800.

complications of: Ali Labib, 1902 (urinary fistulæ).—Bouchut, 1879a, 874–877 (chyluria).—Letulle, 1905c, Apr., 607–609 (phlebitis): 1905d, Apr., 231–232.— Looss, 1905, 94, 95 (fistulæ).—Madden, 1902, Dec., 241–257 (calculus, vesical); 1902. July, 317–329.—Manson, 1903, 3. ed., 610 (fistulæ).—Milton, 1902, 166–167 (carcinoma), 167–168 (stricture), 200–201 (fistulæ), 215.—Mouvenoux, 1884, 1144 pp. (chyluria).—N. Y. M. J., 1896, v. 64, 586 (urethral fistulæ).—Panagiotatou, 1900, Mar., 649–650 (chylothorax).—Silva Lima, 1877, —; 1878, Feb., 171.—Trekaki, 1896, Oct., 586 (urethral fistulæ).—Trekaki & Eichstorff, 1896, Sept., 769 (urethral fistulæ).

——, diagnosis of: Jaksch, 1892, —.—Looss, 1905, 105.—Manson, 1903, 3. ed., 613-

614

——, geographic distribution of: Anders & Callahan, —, 509–519 (U. S.).—Ashford, King & Igaravidez, 1904, 91 (Porto Rico).—Batho, 1872, Dec., 502–504 (Cape of Good Hope).—R. Bl., 1891, 611–612 (Cuba).—Brault, 1891a, 382–385, 3 figs. (Lyon); 1891b, Aug., 409 (Tunisie); 1891c, 449–453; 1892a, 51–56.—Braun, 1903, 3. ed., 169–172, figs. 118–122 (Egypt, Capland, Abessinien, Sudan, Mozambique, Natal, Gold Coast, Tunis, Algiers, Arabia (Mekka).—Cahier, 1893a, 101–116 (Tunisie).—Chevreau & de Chazal, 1890a, June, 44 pp. (à l'île Maurice).—Cobbold, 1876, 487–489 (Australia, Egypt); 1878h, 357 (Natal).—Le Dantec, 1904, Nov., 399–400 [62–63] (Natal).—Davies, 1884,

181 (Cape of Good Hope).—Deblenne, 1883, 271 pp. (Nosi-Bé).—Eyles, 1887a, 659–660 (West Africa).—Fagge & Pye Smith, 1902, 4. ed., 475 (Egypt & Natal).—Faichnie, 1905, Nov., 638–639 (England).—Felkin, 1889.—Freeman, 1905, 145–148 (England).—Gonzales Martinez, 1904 (Porto Rico).—Grassi & Rovelli, 1888c, June, 799 (Sicilia).—Gunn, 1905, May 23 (Porto Ricans at San Francisco); 1905, 1953–1954; 1906, Apr., —; 1906, July, 224.—Harley, 1864a, 55–72 (Cape of Good Hope); 1869a, 379–387 (Cape of Good Hope and Natal); 1871a, 47-62, pl. 1 (southeastern coast of Africa).—Hatch, 1903a, 772 Natal); 1871a, 47-62, pl. 1 (southeastern coast of Africa).—Haten, 1903a, 72 (India); 1903b, 225 (India).—Hillmantel, 1893a, 230-233, figs. 1-3 (?U. S. A.); 1893b, 4 pp., 3 figs. (?U. S. A.).—Katsurada, 1904e, Aug., — (Japan: Yamanashi, Hiroshima, Saga, etc.).—Lahille, 1906, 262-265, 2 figs. (Antilles).—Legrain, 1898, 148-169, 8 figs. (Algeria).—Letulle, 1904a, 46-48 (Martinique); 1905a, 420-421 (Martinique).—Lillie, 1901a, 212 (Orange River Colony).—Lyle, 1882a, 9-10 (southeast coast of Africa); 1883a, 113-132.—Manson, 1901, 542 (Africa; Persia; Egypt); 1902, Dec., 384-385 (West Indies); 1902, Dec., 1894-1895 (West Indies); 1903, Jan., 18 (West Indies); 1903, Jan., 121 (West Indies); 1903, Feb., 15 (West Indies); 1903, June, 293 (West Indies); 1903, 3. ed., 605 (Egypt, Natal, Mauritius, Gold Coast, eastern coast of Africa to Port Elizabeth; Uganda; Mesopotamia; Cyprus; possibly Sicily; West Indies).—Martinez, 1904, 32 pp. (Puerto Rico); 1904, Dec., 193-194 (Puerto Rico).—McL., K., 1903, June, 225 (India).—Milton, 1897, 93-106 (Egypt); 1903, Mar. (Egypt); 1903, May, 779-780 (Egypt).—Mohammed Bey Talaat, 1904a, statistics (Eygpt).—Montgomery, 1906, in animals (India); 1906, 1-32, pl. 1, figs. 1-4, pl. 2, figs. 1-3; (2), pp. 1-37, pl. 1, figs. 1-4, pl. 2, figs. 1-3 (India); 1906, Jan., 15-46 (India); 1906, Feb. (India); 1906, Apr., 138-174, pls. 1-2 (India); 1906, May, 164 (India); 1906, July, 531-532 (India); 1906, Aug., 251-252 (India); 1906, Aug., 259-260 (India).—Peyrot, 1905, 105-111 (Tombouctou).—Posnett, 1901, Oct., 318-319; 1901, Oct., 672; 1901, —Powell, 1903, Feb., 490 (India).—Rafferty, 1904, June, 918-919 (U. S. A.); 1904, June, 1148-1149 (U. S. A.); 1904, June, 1661 (U. S. A.); 1904, Nov., 523 (U. S. A.).—Renoult, ——, 366-370 (Egypt).—Sandwith & Harding, 1906, 217 (South Africa).—Scott, 1904, Mar., 726 (Persia).—Sewell, 1904, Nov. (India); 1904, Nov., 360 (India).—Smith, 1905, Oct., 656-659 (U. S. A.).—Sons., 1874–71-83 (Egypt): 1874–Aug. 502-510. Sept., 521-539 figs. 1-7 (Egypt). (India); 1903b, 225 (India).—Hillmantel, 1893a, 230-233, figs. 1-3 (?U. S. A.); (South Africa).—Scott, 1904, Mar., 726 (Persia).—Sewell, 1904, Nov. (India); 1904, Nov., 360 (India).—Smith, 1905, Oct., 656-659 (U. S. A.).—Sons., 1874, 71-83 (Egypt); 1874, Aug., 502-510, Sept., 521-539, figs. 1-7 (Egypt); 1876, Sept., 521-539, figs. 1-7 (Egypt); 1876, Dec., 738-748 (Egypt); 1876, 3 (Egypt); 1876, June, 650 (Egypt); 1876, Jan., 376 (Egypt); 1876, 652-673, figs. 1-9 (Egypt); 1887 (Egypt); 1887, 12 pp. (Massana).—Sturrock, 1899, Dec., 1543 (Mesopotamia); 1899, Dec., 1212-1213 (Mesopotamia).—Tyson, 1903, 3. ed., 1181 (Egypt & Africa).—Villeneuve, 1891, 398 (Tunisie); 1891, 421 (Tunisie); 1892, 153-157 (Tunisie); 1892, 556 (Tunisie); 1893, 49-53 (Tunisie).—Williamson, 1902, Aug., 475 (Cyprus); 1902, Aug., 313 (Cyprus); 1902, Sept., 956 (Cyprus); 1902, Dec., 625 (Cyprus); 1905, 956 (Cyprus).

Abdominal organs: Kartulis, 1885a, 139-145, figs. 1-4; 1885b, 188-189.

Liver: Looss, 1905, 96, 100.—Manson, 1903, 3. ed., 610, 613 (ova in gallstone).—Phillips, 1904, Sept., 657.—Ruault, 1885, 56-59; 1885, 145-147.— Symmers, 1903, Dec., 237-239, pl. 21, figs. 1-2.

Mesentery: Ruault, 1885, 56-57; 1885, 145-147.

Omentum: Madden, 1901c, 143-144.

Pancreas: Looss, 1905, 96.

Peritoneum: Madden, 1901c, 143–144 (and omentum); 1902, July, 787–788.— Manson, 1903, 3. ed., 613.

Spleen: Looss, 1905, 96.

Heart: Manson, 1903, 3. ed., 613.

Blood, hematology, eosinophilia: Balfour, 1903, Dec., 1649; 1904, Jan., 35; 1904, Jan., 39; 1904, Aug., 374; 1905, Feb., 125.—Coles, 1902a, 1137-1138; 1902,

Nov., 402; 1902b, 914; 1902c, 955; 1902d, 787; 1902e, 993.—Douglas, 1903, Oct., 10; 1904, Oct., 476.—Le Dantec, 1904, Nov. 18, 400.—Fagge & Pye Smith, 1902, 4. ed., 475.—Kautsky Bey, 1904a-e; 1905a; 1905, Oct., 665-666; 1905, Oct., 477.—Lockwood, 1901a, 2. ed., 821.—Looss, 1905, 97-100.—Manson, 1903, 3. ed., 606.—Rep. Wellcome Research Lab., Khartoun, 1904, 58-61.—Russell, 1902, Dec. 6, 1540; 1902, Dec. 18, 326; 1902, Dec. 20, 988; 1902, Dec. 22, 1234-1235; 1902, Dec. 27, 1134.—Symmers, 1905, Jan. 18, 22; 1906, Jan. 7, 662.—Tyson, 1903, 3. ed., 1181.

Intestines: Damaschino, 1882a, 949 (and urinary); 1883a.—Firket, 1897, June.—; 1899, v. 3 (1), 71.—Lahille, 1906, v. 9 (2), 262–265, 2 figs.—Letulle, 1904a, 46–48; 1905a, 420–421; 1905e, 329–439, figs. 1–16, pls. 1–2.—Mackie, 1882, Oct. 7, 661.—Symmers, 1905, Jan. 7, 22.—Zancarol, 1882, 410–412, pl. 26 (and bladder).

Appendix: Burfield & Shaw, 1906, Feb. 10, 368–370, figs. 1–2; 1906, Mar. 3, 359; 1906, Mar. 10, 757; 1906, May 29, 1076; 1906, July 16, 532.—Crimp, 1906, Mar. 10, 672.

Rectum: Belleli, 1885b, 54–56, figs. 19–20.—Bowlby, 1891a, 136.—Letulle, 1904, Jan. 13, —; 1905b, 37; 1905f, 659–714, figs. 1–19, pls. 1–2; 1905g, 672; 1906, Jan. 31, 64–65.—Looss, 1905, 95, 99, 105.—Madden, 1899, May, 566–568; 1899, July, 159; 1902. Feb., —; 1902. May 10, 1285.—Manson, 1903, 3. ed., 610, 612–613.—Milton, 1902, 169.—Ruault, 1885, 56–57; 1885, 145–147.

Female genitalia: Goebel, 1905; 1379–1382; 1905, Dec. 23, 1344; 1905, Nov. 28, 2333; 1906, Jan. 18.—Horwood, 1906, Mar. 10, ——; 1906, Aug. 15, 633.—Looss, 1905, 95.—Madden, 1899, June 24, 1716; 1899, July, 114.—Manson, 1903, 3. ed., 610, 612, 613.—Milton, 1902, 169, 202.

Urinary system: Bowlby, 1891b, 194–195.—Damaschino, 1882a; 1883a, 150–156.—Eve, 1888a, 184.—Huber, 1896, 581.—Manson, 1903, 3. ed., 612.—Milton, 1902, 181–182.—Trekaki, 1903, Jan., 309–325; 1903, Mar., 329–349; 1903, Apr., 396–408; 1903, June 30, 800.

Calculi: Ebstein, 1884a.—Looss, 1905, 94, 106.—Madden, 1902, Dec., 241–257.—Milton, 1902, 200.—Reyer, 1856, ——.—Zancarol, 1882, 76.

Kidney: Eve, 1888a, 184.—Manson, 1903, 3. ed., 613.—Ruault, 1885, 56–57; 1885, 145–147.

Ureter: Milton, 1902, 167.

Bladder: Eve, 1888a, 184.—Fenwick, 1888a, 344–346 (epicystitis); Fenwick & Harris, 1888a, 183–184.—Goebel, 1903a; 1903b; 1903g; 1904, Oct., 1810; 1905, v. 3 (3): 1905, 369–513 (carcinoma): 1905, Dec., 957–958: 1906, Mar., 45–46; 1906, July, 315.—Harrison, 1889a, 163.—Huber, 1896, 581.—Kutner, 1905, v. 16 (12), 649–657, 1 fig.: 1906, Jan., 78.—Lewis, 1900a, 1057–1058.—Loss, 1905, 97, 98, 100.—Manson, 1903, 3. ed., 610–612.—Milton, 1902a, 181–182; 1902, 166, 191, 192, 214.—Peyrot, 1905, 105–111.—Ruault, 1885, 56–57; 1885, 145–147.—Shattock, 1891, 196–197.—Zancarol, 1881–1882, 410–412, pl. 26.

Prostate: Ruault, 1885, 56-57; 1885, 145-147.

Urine: Fagge & Pye Smith, 1902, 4. ed., 475.—Gibson, 1904, 219.—Looss, 1905, 105.

Hematuria (bilharzian, endemic, Egyptian): Allen, 1888, Apr., 310–320.—Batho, 1872a, 502–504.—Bilharz, 1856a, 49–52, 65–68.—Cantani, 1886, 9.—Chaker, 1890a, 72 pp., figs. —; 1890b, 594.—Chute, 1888a, 85–87, 89–91.—Cobbold, 1885a, 985.—Crevaux, 1872a-b; 1874a, 165: 1875a.—Le Dantec, 1904, Nov., 399–400.—Davies, 1884a, 181–187, figs. 1–7.—Douglas & Hardy, 1903, Oct. 10, 1009–1012; 1903, Oct. 22, 259: 1903, Oct. 31, 865–866; 1903, Nov. 28, 1032; 1903, Nov. 30, 735; 1903, Dec., 263: 1904, Jan. 30, 60; 1904, Jan. 2, 34.—Fagge & Pye Smith, 1902, 4. ed., 475, 679.—Fayrer, 1879b, 189 (not clear whether this refers to the disease or to the parasite).—Frothingham, 1904, 453–457, 1 pl.—Griesinger, 1866, 96.—Guès, 1879a, 161–190.—Guillemard, 1882a, 61 pp.—Gutch, 1900a, 1222; 1900b.—Handford, 1887b, 240–245, figs. 1–3; 1889a, 424–425; 1894a, 48–49.—Harley, 1864a, 55–72, figs. 1–16; 1864b, 173–175; 1864c, 156–157; 1864d, 161–163; 1864e, 515–517; 1865a, 161–173, figs. a–b; 1869a, 379–387; 1869b, 394; 1871a, 47–62, pl. 1.—Jourdan, 1877a, — Looss, 1905, 33, 93, 94.—Lyle, 1882a, 9–10; 1883a, 113–132.—Renoult, 1808, 366–370.—Roberts, 1879, 461–466.—Simpson, 1872, Sept. 21, 320–321.—Sons., 1874, 71–83; 1874, 502–521; 1874, 305–321; 1876, 233–235; 1884, 17–21.—Thurn, 1882, —.—Ultzmann, 1878, 34, fig. 12.

Lungs: Belleli, 1885a, 1–3.—Bowlby, 1891b.—Looss, 1905, 96.—Manson, 1903, 3. ed., 610, 613.

Tumors: Albarran & Bernand, 1897a, 645–647 (epithelial); 1897b, 1096–1023, pl. 11.—Belleli, 1885, 54–56.—Looss, 1905, 95, 96.—Madden, 1903, Jan., 1–2; 1903, Jan., 14.

——, symptomatology of: Belleli, 1886a, 4–5, 12–14, 18–21, 28–29, 35–37.—Le Dantec, 1904, Nov., 399–400.—Goebel, 1903a-i.—Lelean, 1902a-b; 1904f.—Lockwood, 1901, 2. ed., 821.—Looss, 1905, 94, 95, 105.—Manson, 1901, 543; 1903, 608–609, 610.—Milton, 1902, 191, 201–202.—Moty, 1893, Feb., 51–56 (urine), figs. 1–4.—Tyson, 1903, 3. ed., 1181.

——, prognosis: Lockwood, 1901, 2. ed., 821.—Looss, 1905, 93, 96, 106.—Manson, 1901, 543; 1903, 3. ed., 609, 610, 614, 615.—Milton, 1902, 191, 200 (death due

to exhaustion).

, treatment of: Allen, 1883, 660.—Belleli, 1886a,.—Cadet, 1876a, —; 1876b, —.—Le Dantec, 1904, Nov., 400.—Ensor, 1904, Nov., 575–576 (chrysoidine); 1904, Nov., 360.—Fouquet, 1885a, 677–680, 693–696; 1885b, 88–95.—Goebel, 1903a-b.—Harley, 1871a.—Looss, 1905, 106, 107.—Manson, 1901, 544; 1903, 3. ed., 615, 616.—Milton, 1897a, 93–106; 1902, 214, 218; 1902, 93–106 (surgery); 1903a, 866–869; 1903b, 714; 1903c, 659; 1903d, 779–780.—Sons., 1885, June, 1197–1198.—Wortabet, 1882, Dec., 979–980 (turpentine).

BISTOMA Reich, 1801, 371, lapsus for Dist. = Fasc. —Stiles & Hass., 1898a, 83 (to Dist.).

stridulæ Reich, 1801, 371, in Strix stridula.

BOTHRIOGASTER Fuhrmann, 1904, Sept. 23, 59-61 (m. variolaris), subf. Syncœliinæ.

variolaris Fuhrmann, 1904, 59-61, figs. 1-2 (in Rostrhamus sociabilis; South America).

BRACHYCÆCUM Rail., 1896, 15 Mar., 160, Brachycœlium Duj., 1845a [not Brachycœlus Chaudoir, coleopteron] renamed, hence type crassicolle.—Stiles & Hass., 1898, 83 (for Brachycœlium).—Stoss., 1898, 31.

brusinai (Stoss., 1889 [Looss, 1901]) Barbagallo & Drago, 1903, 410 (in Oblata melanura; Catania), Br. as subg. of Dist.

BRACHYCELIUM Mont., 1888, 38 for Brachycelium.

BRACHYCLADIINÆ Odhn., 1905, 346, 347, f. Fasciolidæ.

BRACHYCLADIUM Looss, 1899b, Dec., 558–560 (tod. palliatum); 1901, 208; 1902m, 707, 708, 709, 711, 715, 716, 717, 718, 755–778 (cf. Campula; Opisthorchis), 811.—Braun, 1900g, 251, 254; 1901b, 38; 1905, 339, 344, 345, 346, 347–348.—Odhn., 1905, 347a–348.—Stiles 1901, 203, 204, 205.

delphini (Poir., 1886) Looss, 1899b, 558; 1902m, 707, 708.—Odhn., 1905, 348.

oblongum (Cobbold of Braun) Looss, 1902m, 716.—Odhn., 1905, 343, 347.

palliatum (Looss, 1885) Looss, 1899b, 558, 560; 1902, 707, 708, 711, 715.—Daar, 1902, 653.—Odhn., 1905, 341, 342, 347, 348.

rochebruni (Poir., 1886) Looss, 1899b, 558; 1902m, 707, 811.—Odhn., 1905, 344, 346, 347.

BRACHYCŒLIINÆ Looss, 1899, Dec., 607, 611, 612, 614; 1902, 839, 841.—Luche, 1900, 561; 1901, 173.—Odhn., 1902, 40, 42.—Pratt, 1902a, 889, 902 (key); contains Phaneropsolus, Lecithodendrium, Pycnoporus, Brachycœlium; related genera Cymatocarpus, Brandesia.

BRACHYCŒLINÆ Ward, 1901, 185.—Jægers., 1903a, 14.

(BRACHYCCLIUM) Duj., 1845a, 383, 388, 402 (subg. of Dist.), type crassicolle after Stiles & Hass., 1898a, 83, but arrectum after Luehe, 1899k, 536. Raised to generic rank by Stiles & Hass., 1898a, 83.

arrectum Duj., 1845a, 387, 403 (in Lacerta viridis; Rennes).—Stoss., 1895, 225 (to Dicroccelium).

brachysomum (Crep., 1846) Stoss., 1892, 148, 189, 190, 191; 1899, 10 (to Levinsenia). claviforme (Brand., 1888) Stoss., 1892, 148–149, 191. See Brachycelium.

clavigerum (Rud., 1819) Duj., 1845a, 387, 404 (misdetermined).—Stoss., 1889, 576 (to Dicroccelium).See confusus Looss, 1894.

crassicolle (Rud., 1809) Duj., 1845a, 386, 404–405.—Cohn, 1903, 42.—Stiles & Hass., 1898a, 83 (designated type of Brachycœlium). See Brachycœlium.

(BRACHYCELIUM)—Continued.

heteroporum Duj., 1845a. 387, 402–403 (in Vespertilio pipistrellus; Rennes).—Looss, 1899b, 611 (type of Pycnoporus).

macrophallos (Linst., 1875) Stoss., 1892, 147, 190.

nigrovenosum (Bellingham, 1844) Looss, 1902m, 822.

oviforme (Poir., 1886) Stoss., 1892, 12-13, 40.

pygmæum (Levin., 1881) Stoss., 1892, 147 (in Somateria mollissima; Greenland), 189; 1899, 10 (to Levinsenia).

retusum Duj., 1845a, 386, 405-406 (in Rana temporaria at Rennes).

rubellum (Olss., 1868) Stoss., 1886, 60.

somateriæ (Levin., 1881) Stoss., 1892, 146, 189.

tacapense (Sons., 1894) Stoss., 1895, 215, 235.

BRACHYCŒLIUM (Duj., 1845a) Stiles & Hass., 1898a, 83 (type crassicolle after Stiles & Hass., arrectum after Luehe).—Baird, 1853a, 52.—R. Bl., 1891, 609.— Braun, 1892a, 663, 672; 1893a, 885, 909, 910; 1900h, 6.—Giard, 1897c, 955, 956, sp. in Pélécypodes; 1897d, 956–957.—Looss, 1899b, 535, 536, 611–614, 617, 618, 620, 622, 623, 625, 647; 1902m, 705, 768–775, 814, 815, 816, 822, 823.— Luche, 1899k, 536 (type arrectum); 1900aa. 562, 563.—Mont., 1888, 38 (Brachycelium), 92. 105, 1892, 687; 1893, 43. 150.—Pratt. 1902a. 889, 903 (key).—Rail., 1896, 160 (renamed Brachycæcum).—Staff.. 1903, 827, 828; 1905, Apr. 11, 684.—Stiles, 1901, 179, 196, 197. 198, 199, 200, 201, 202, 203.—Stiles & Hass.. 1898a, 83, 97, 98 (type Dist. crassicolle Duj., 1845) (syns. Dist. (Brachycæclium), Brachycæcum Rail., 1896).—Stoss., 1892, 12; 1899, 7.—Ward, 1901, 175 Ward, 1901, 175.

1845: Distoma (Brachycœlium) Duj., 1845a, 383, 388, 402 (tld. crassicolle).

1888: Brachycelium Mont., 1888, 38, misprint.

1896: Brachycæcum Rail., 1896, 160, Brachycælium [not Brachycælus] renamed.

crassicolle (Rud., 1809) Looss, 1899b, 611, 612; 1902, 768, 769, 772, 774. 775. 813, 814, 815, 816, 818, 821, 822, 823, 832. 835.—Cohn, 1903, 42,—Luehe, 1900, 562.—Staff., 1903, 827, 828; 1905, Apr. 11, 693.

hospitale (Staff., 1900) Looss, 1902, 822.—Staff., 1903, 824-828, figs 4-5; 1905, 682 (int. of Diemyctylus viridescens and Plethodon erythronotus; Canada).

luteum (Ben., 1870) Giard, 1897c, 955; 1897d, 956, 957.—Caullery & Chappellier, 1906, 325.—Reported for Donax, Scyllium canicula, S. stellare.

nigrovenosum (Bellingham, 1844) Looss, 1902m, 822, as doubtful, species Giard, 1897d, 956–957 (in Donax trunculus, Tellina fabula, T. tenuis, T. solidula, T. baltica).—Leger, 1879h, 957–958.

BRACHYLEMUS E. Bl., 1847a, 295–302 (for Brachylaimus).—Dies., 1850a, 331.— Stiles & Hass., 1898a, 83, 84 (syn. of Brachylaima Duj., 1843).

cylindraceus (Zed., 1800) E. Bl., 1847a, 295–298, pl. 12, fig. 2.—Baird, 1853a, 51.— Dies., 1850a. 368 (to Dist.).—Looss, 1894a, 64 (to Dist.); 1899b, 600 (type of Haplometra).

erinacei E. Bl., 1847a, 300-302, pl. 9. fig. 2 (Erinaceus europæus; Paris).—Dies., 1850a, 335 (renamed Dist. linguæforme).

variegatus (Rud., 1819) E. Bl., 1847a, 298-300, pl. 13, fig. 1.—Looss, 1894a, 71 (to Dist.).—Type of Hæmatolæchus 1899 and Pneumonæces 1902.

BRACHYLAIMA Duj., 1843a, 338-341 (tld. advena).—Luehe, 1899k, 536.—Stiles & Hass., 1898a, 83-84, 96 (type advena = Dist. migrans 1845) (syns. Dist. (Brachylaimus) Duj., 1845. Brachylæmus Duj. of Bl.).

advena Duj., 1843a, 338-341 (in Sorex).—Stiles & Hass., 1898a, 83, 84, 96 (designated type) (syn. Dist. migrans Duj., 1845).

fulvum Duj., 1843a, 340-341 (in musaraigne).—Stiles & Hass., 1898a, 83 (in Sorex araneus).

(BRACHYLAIMUS) Duj., 1845a, 382, 388, 407-420 (Brachylaima, 1843, renamed, hence type advena), used as subg. of Dist. See also Brachylaimus.

æquale Duj., 1845a, 410.

aloysix (Stoss., 1885) Stoss., 1886, 58.

andersoni (Cobbold, 1876) Stoss., 1892, 19, 37.

annuligerum (Nord., 1832) Stoss., 1886, 44.

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(BRACHYLAIMUS)—Continued.
   arcuatum Duj., 1845a, 410-411.
   aristotelis Stoss., 1892, 14-15, 40.
   ascidioides (Ben., 1873) Stoss., 1892, 16, 40.
   baraldii (Sons., 1892) Stoss., 1895, 218-219, 235.
    bergense (Olss., 1868) Stoss., 1886, 63.
   bothryophoron (Olss., 1868) Stoss., 1886, 57.
   carnosum (Rud., 1819) Stoss., 1886, 59.
   caudatum (Linst., 1873) Stoss., 1892, 39.
   characis (Stoss., 1886) Stoss, 1886, 59.
   cirratum (Rud., 1808) Duj., 1845a, 413.
   conjunctum (Cobbold, 1860) Rail., 1893a, 365.
   conostomum (Olss., 1876) Stoss., 1886, 62.
   corrugatum Duj., 1845a, 409-410.
   crassiusculum (Rud., 1809) Stoss., 1892, 151-152, 194.
   cymbiforme (Rud., 1819) Stoss., 1895, 215-216, 236.
   depressum Stoss., 1883; 1886, 59.
   didel phidis Par., 1896, 3-5, fig. 1a (in Didelphis marsupialis azaræ; Paraguay).
   dimorphum (Dies., 1850) Stoss., 1886, 60.
   elegans (Rud., 1802) Duj., 1845a, 414-415.
   endemicum (Bælz, 1883) Stoss., 1892, 39, 40.
   exasperatum (Rud., 1819) Duj., 1845a, 411.
   fabenii (Mol., 1859) Stoss., 1886, 59.
   fellis (Olss., 1868) Stoss., 1886, 57.
   filum Duj., 1845a, 418.
   flavescens (Ben., 1870) Stoss., 1886, 58 (in Gobius minutus).
   globiporum (Rud., 1802) Duj., 1845a, 417.
   globocaudatum (Crep., 1825) Duj., 1845a, 413-414.
    instabile Duj., 1845a, 412.
    leptostomum (Olss., 1876) Stoss., 1892, 17, 39.
    lima (Rud., 1809) Stoss., 1892, 14, 39, 40.
    linguatula (Rud., 1819) ——?—
    lorum Duj., 1845a, 407.
    maculosum (Rud., 1802) Duj., 1845a, 412-413.
    marginatum (Mol., 1858) Stoss., 1892, 149, 189.
    megastomum (Rud., 1819) Stoss., 1886, 64.
    mentulatum (Rud., 1819) Duj., 1845a, 415.
    mesostomum (Rud., 1803) Stoss., 1892, 149-150, 193, 194.
    micrococcum (Rud., 1819) Stoss., 1892, 150, 190.
    migrans Duj., 1845a, 407-409=Brachylaima advena Duj., 1843, renamed.
    moleculum (Linst., 1880) Stoss., 1892, 150, 192.
    monas (Rud., 1819) Parona, 1896, 12-13 (in Siphonops annulatus; Brazil).
    mormyri (Stoss., 1885) Stoss., 1886, 59.
    mülleri (Levin., 1881) Stoss., 1886, 61.
    oblongum (Cobbold, 1858) Stoss., 1892, 16-17, 37.
    obovatum (Mol., 1859) Stoss., 1886, 59.
    oligoon (Linst., 1887) Stoss., 1892, 149–192.
    plesiostomum (Linst., 1883) Stoss., 1892, 152, 192.
    pseudostomum (Will.-Suhm., 1870) Stoss., 1895, 219–236.
    recurvum Duj., 1845a, 410.
    reticulatum (Wright, 1879) Stoss., 1892, 154-155, 192.
    robustum (Lorenz, 1881) Stoss., 1892, 18, 38.
    rubens Duj., 1845a, 411.
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(BRACHYLAIMUS)—Continued.

sanguineum (Sons., 1894) Stoss., 1895, 217, 235.

serpentatum (Mol., 1859) Par., 1896, 18–19 (in Sayris camperii=Scomberesox rondeletii; Padova).

signatum Duj., 1845a, 415-416.

soccus (Mol., 1858) Stoss., 1886, 64.

solea Duj., 1845a, 417-418.

subflavum (Sons., 1892) Stoss., 1895, 216, 235.

tenuicolle (Rud., 1819) Stoss., 1892, 18-19, 39.

tereticolle (Rud., 1802) Duj., 1845a, 419-420.

umbrinæ (Stoss., 1885) Stoss., 1886, 58.

validum (Linst., 1886) Stoss., 1892, 15-16.

varicum (Mueller, 1784) Stoss., 1886, 58.

variegatum (Rud., 1819) Duj., 1845a. 416-417.

vitellatum (Linst., 1875) Stoss., 1892, 154, 190.

vitta Duj.. 1845a, 418.

- BRACHYLAIMUS (Duj., 1845a) E. Bl., 1847a, 298 (Brachylaima renamed, hence type advena).—R. Bl., 1891, 609, 610.—Braun, 1892a, 643; 1893a, 885, 894, 909, 911; 1901b, 11; 1901e, 341, 342.—Giard & Billet, 1892, 614.—Johnston, 1902, 329.—Looss, 1899b, 535, 609.—Luehe, 1899, 530.—Mont., 1888, 92, 105; 1893, 43, 150.—Rail. & Marotel, 1898, 31, 38.—Sons., 1890, 141.—Stiles & Hass., 1898a, 84.—Stoss., 1892, 13; 1898, 32.—Volz, 1899, 231.—West, 1896, 323.—See also (Brachylaimus).
- BRACHYLEMUS Dies., 1850a, 331, for Brachylaimus, q. v. variegatus (Rud., 1819) Dies., 1850a, 355 (to Dist.).—Molin, 1859, 828 (to Dist.).
- BRACHYMETRA Stoss., 1904, 10 (m. parva) [not Mayr. 1865, insect]. parva Stoss., 1904, 10–12, fig. 1 (in Rana esculenta: Istria centrale).
- BRACHYPHALLUS Odhn., 1905, 350, 356, 359 (tod. crenatus).—Nicoll, 1907, 84.

 crenatus (Rud., 1802) Odhn., 1905, 349, 352-355, pl. 4, figs. 3-5 (includes Fasc. crenata Rud., 1802, 76, pl. 2, fig. 5: Dist. crenatum (Rud.) Rud., 1809a, 404, pl. 5, fig. 1; Mol., 1868, 48, pl. 5, figs. 96-98; Dist. appendiculatum p. p. Rud., 1819a, 404; Dist. tectum Linst., 1873, 104, pl. 5, fig. 4; Dist. ocreatum Mol. p. p. Lint., 1899, 288, pl. 35, figs. 16-24; Hemiurus crenatu (Rud.) Luehe, 1901, 24; Dist. crenatum Rud., 1810a, 376; Mol., 1859, 25, pl. 1, fig. 3); (hosts reported by Odhn. for Scandinavia: Cottus scorpius, Pleuronectes limanda, Gasterosteus aculeatus, Ammodytes tobianus, Salmo salar, S. trutta, Osmerus eperlanus); 1906, 62.—Nicoll, 1907, 72, 84, 88-89 (in Ammodytes tobianus).
- BRANDESIA Stoss., 1899, 7, 10 (m. turgida).—Braun, 1900h, 6.—Looss, 1899b, 617, 623, 624–625.—Pratt, 1902a, 889, 903 (key), related to Brachycœllinæ—Staff., 1905, Apr. 11, 683, 684, 685, 686.—Stiles, 1901, 189, 190.
 - turgida (Brand , 1888) Stoss., 1899. 10.—Cohn, 1904, 235.—Kowal., 1902d, 27 [9].—Looss, 1899, 624, 775-777, figs. A. B.—Staff., 1905, Apr. 11, 684, 685.
- BRAUNINA Heider, 1900a, 19–22, figs. a–d (Hemistomidæ): 1900b, 19–22, figs. 2–d.—Pratt, 1902a, 908 (key).—Wolf, 1903, 603–626, 1 pl., 11 figs.

cordiformis Wolf, 1903. 623, 1 pl., 11 figs. (in Squalus sp; Rio Janeiro).

BRAUNININE Wolf, 1903, 622, 623, n. subf.

BUCEPHALIDÆ Poche, 1907, 125.

(BUCEPHALOPSIS) Dies., 1855a, 396 (m. haimeanus) subg. of Bucephalus Baer.—Ziegler, 1883, 540.

aculeatus Dies., 1858, 275, based on Wagener's Furcocerce in Planorbis marginatus.—Ziegler, 1883, 540.

haimeanus (Lacaze-Duthiers, 1854) Dies., 1855a, 396; 1858d, 275–276 (syns. Bucephalus (Bucephalopsis) haim. Lacaze-Duthiers; Cerc. haim. Moul.) (in Ostrea edulis, Cardium rusticum; Balearic Isles).—Tennent, 1906, 642.—Ziegler, 1883, 540.—See also sub Bucephalus.

BUCEPHALUS Baer, 1826a, 124–125 [not Smith, 1829, reptile] [m. polymorphus]; 1827b, 570–589 (m. polymorphus).—Biehringer, 1884, 20; 1888a, 231, 232, 235.—Brand., 1891d, 8.—Braun, 1892a, 589, 592, 599, 636, 640, 768, 769, 770, 771; 1893a, 823, 825, 834, 884, 892.—Burm., 1856a, 244, 245, 250.—Carus, 1835a, 94.—Cobbold, 1876, 304; 1879b, 453.—Dies., 1850a, 286, 294; 1855a, 395; 1858d, 271, 274.—Duj., 1845a, 478.—Erc., 1881e, 60; 1882a, 296.—Fil., 1854a, 6; 1857c, 22.—Garner, 1875a, 102 (in freshwater mussel).—Goldb., 1855a, 15.—Gronkowski, 1902a, 522 (15).—Hoyle, 1890, 539 (larva of Gasterost.), 540, fig. 4-F.—Jackson, 1888, 643, 651 (= Gasterost.; in Anodonta and oyster), 643, 654.—Kath., 1894a, 138.—Keber, 1852,—.—Lacaze-Duthiers, 1854a, 294 ff.—Leuck., 1863; 454, 495, 503, 512.—Looss, 1885b, 6; 1892a, 125; 1894a, 55, 252.—Mont., 1888, 9, 77, 79, 80, 92, 94; 1893, 6, 9, 10.—Moul., 1856a, 65, 121, 124.—Odhn., 1905, 293, 304.—Pag., 1857, 7, 10, 29, 52, 53.—Poche, 1907, 125.—Spengel, 1905, 258.—Uličný, 1878, 214.—Woods, 1875, v. 4 (29), Aug., 58-66, pls. 4–5.—Ziegler, 1883, v. 39 (4), Dec. 31, in 537-571, pls. 32-33, figs. 1-28; 1883, v. 6 (148), 10 Sept., 487-492; 1905, 36.

1855: Eubucephalus Dies., 1855a, 395 (m. polymorphus).

aculeatus Dies., 1858, 275 (in Planorbis marginatus) to (Bucephalopsis).

[bellii, see Smith, reptile.]

crux Levin., 1881a, 80–81, pl. 3, fig. 7a-j (in Modiolaria discors; Egedesminde).—
 Braun, 1893a, 834.—Odhn., 1905, 304.—Tennent, 1906, 641, 643 (in Modiolaria discors).

cucullus Ziegler, 1883, 540 (for cuculus).

cuculus McCrady, 1874, Dec. 3, 176–192, figs. 2, 1–8 (in Ostrea virginiana; Charleston, S. C.)—Braun, 1893a, 834.—Tennent, 1906, 641, 644–646, 682 (syn. of B. haimeanus Lacaze-Duthiers) (in oyster; Charleston, S. C.).—Ziegler, 1883, 540 (cucullus).

euculus McCrady, 1874, descr. of fig., misprint.

[gutturalis, see Smith, reptile.]

haimeanus Lacaze-Duthiers, 1854a, 294–302, pl. 6, figs. 1–10 (in Ostrea edulis, Cardium rusticum; Balearic Isles, Mahon, Cette).—Babcock, 1875a, 144, 145, 146, pl. 98, figs. 6–7.—Braun, 1893a, 834.—Caullery & Chappellier, 1906, 325.—Claparède, 1863, 10, 11 (to Cerc.).—Dies., 1855a, 379, 396, type of B. (Bucephalopsis); 1858d, 276 to (Bucephalopsis).—Erc., 1881e, 41, 87 (haimejanus); 1882a, 277, 323 (haimejanus).—Fil., 1855b, 21.—Giard, 1874e, 485–487 (encystation) (in Cardium rusticum, Ostrea edulis; Mediterranean); 1874i, 375–377; 1874g, 276–278; 1875a, 466–467; 1897, 955.—Huet, 1889a, 145–149; 1893a, 40–41.—McCrady, 1874, 176, 178, 179, 180.—Moore, 1875a, in 50–57, pl. 3.—Moul., 1856a, 86, 87, pl. 5, fig. 7, 178 (in Ostrea edulis, Cardium rusticum; Balearic Isles, Mahon, Cette).—Pag., 1857, 10; 1862, 295, 298 to (Bucephalopsis), 300, 301.—Tennent, 1906, Feb., v. 49 (4), 635–690, pls. 39, figs. 1–17, pl. 40, figs. 18–39, pl. 41, figs. 41–43, 51 (larva of Gasterost gracilescens in oysters, Ostrea edulis, Cardium rusticum).—Uličný, 1878, 214, 215, 216.—Vaullegeard, 1894, v. 8, 8–14 (in Tapes decussatus, T. pullastra); 1894, 343.—Villot, 1875, 478.—Ziegler, 1883, 539, 540, 567.—Also reported for Belone vulgaris, Cardium edule, Mactra solida.

haimejanus Erc., 1881e, 87; 1882a, 323 (for haimeanus).

intermedius Uličný, 1878, 211–217, pl. 6, fig. 6 (in Anodonta cellensis).—Braun, 1893a, 834.—Tennent, 1906, 641, 643 (in Anodonta cellensis).—Ziegler, 1883, 541.

[jardinii, see Smith, reptile.]

polymorphus Baer, [1826a specific name not given]; 1827b, 570–589, pl. 30, figs. 1–27 (in Unio and Anodonta).—Badcock, 1875a, 141–146, pl. 98, figs. 1–5; 1875b, 149–150.—Ben., 1858a, 1861a, 220.—Bettend., 1897a, 13; 1897, 317.—Biehringer, 1884, 3; 1888a, 230.—Braun, 1883a, 54; 1893a, 834, 835, 838, 847.—Claparède, 1863, 10, 11.—Crep., 1839a, 293.—Desmonceaux, 1868, 22.—Dies., 1850a, 294; 1855a, 379, 395 (in Unio pictorum, An. anatina, A. cellensis); 1858d, 274–275 (syn. Cerc. polym.) (in U. pict., An. cell., A. anat.).—Duj., 1845a, 478.—Erc., 1881e, 37, 38, 41, 44, 45, 60, 87; 1882a, 273, 274, 277, 280, 281, 296, 323.—Fil., 1855b, 21, 22; 1857c, 27, 31.—Gamb., 1896a, 72.—Garner, 1838, 830.—Giard, 1874e, 485, 486.—Hahn & Lefevre, 1884, 806.—Hessling, 1852, 315.—Hoyle, 1890, 540, fig. 4–e.—Jacobson, 301, pl. 8, figs. 1–3.—Juel, 1889, 11.—Keber, 1851a, 99, 100; 1852a, 72.—Kuech., 1856c, 280.—

BUCEPHALUS-Continued.

Lacaze—Duthiers, 1854a, 297, 299, 300, 301.—Levin., 1881a, 80, 81.—Looss, 1885b, 19; 1892a, 122.—McCrady, 1874, 176, 178, 179.—Mont., 1888, 33, 77.—Moul., 1856a, 86, 145, 174, pl. 5 bis, fig. 6 (in An. anat., A. cell., U. pict.), see Cerc. polym.—Mueh., 1898, 11.—Nord., 1840, 548; 1840, 617, 630, 631.—Pag., 1857, 6.8, 10, 27-28, pl. 3, figs. 2-8 in An. anat.; 1862, 300, 302.—Poche, 1907, 124, 125.—Sieb., 1839, 165; 1854, 14.—Stewart, 1875, July, 1-2, pl. 107.—Tennent, 1906, 638, 641, 643, 646, 655, 660, 662, 663, 678, 682 (in An. mutabilis var. anatina and cellensis, U. pict.).—Uličný, 1878, 214, 215, 216.—Wagener, 1857, 22, 45; 1866, 145.—Ziegler, 1883, 488, 491; 1883, 537, 539, 540, 541-567, pl. 32, figs. 1-12, pl. 33, fig. 17, 23, 25 (in An. mutabilis var. cellensis).

[typus, see Smith, reptile.]

BUNODERA Rail., 1896, Mar. 15, 160, Crossodera Duj., 1845 [not Gould, 1837, bird] renamed. hence type nodulosa.—Braun. 1900, 232.—Hass., 1896a, 7.—Heymann, 1905, 83, 89.—Looss, 1899, 580, 594, 595, 597, 598, 599, 604, 671; 1902, 453, 454, 760.—Odhn., 1905, 296.—Pratt. 1902a, 888, 898 [key.,—Staff., 1904, May 3, 491.—Stiles & Hass., 1898, 84–86, 96 [syns., Dist., (Crossodera) Duj., Crossodera Duj., 1860 [by Cobbold] [not Crossodera Gould, 1837, Aves] [type Dist., nodulosum Zed., 1800—Fasc. luciopercæ Mueller, 1776).

auriculata Wedl, 1857 Osborn, 1903, 63, 64, 67, 70.

cornuta Osborn, 1903, 63–73, figs. 1–7 / crawfish, black bass, rock bass, and catfish in Lake Chautauqua, N. Y. (; 1905, Jan. 31, 22,

linearis Rud., 1793 Rail., 1896, 160.

lintoni Pratt in Lint., 1901b. 435 (syn. Dist. auriculatum of Lint. in Acipenser rubicundus).—Cf. petalosum Lander.

nodulosa Freelich, 1791 [Rail., 1896, 160] Looss, 1899b, 598; 1901l, 564; 1902m, 452, 453.—Engler, 1904, 186.—Heymann, 1905, 83, 85.—Kowal., 1902d, 26 (8); 1904, 24 (9) in Perca fluviatilis; Galicia).—Osborn, 1903, 63, 64, 67, 68, 70, 71, 72.—Staff., 1904, May 3, 489—490 in P. flavescens Mit.; Canada).

BUNODERINÆ Looss, 1902, 453.—Pratt, 1902a, 888, 898 (key), contains Bunodera, Tergestia.

CECINCOLA Marshall & Gilbert, 1905, 477, 478-481 m. parvulus .

parvulus Marshall & Gilbert, 1905, 478–481, pl. 15, figs, 1–4 (in Micropterus salmoides: near Madison, Wis.).

CALCEOSTOMA Ben., 1858a, 1861a, 11, 57, 59-60, 194, 196 (m. elegans).—Ben. & Hesse, 1864, 121.—Brand., 1894a, 305, 306.—Braun, 1890a, 411, 412, 416, 443, 451, 468, 472, 478, 483, 484, 492, 512, 514, 516, 517, 523, 542; 1893a, 890.—Dies., 1858e, 379 (syn. of Dactylogyrus calceostoma Wagener); 1859c, 441 (1 sp. elegans=calceostoma .—Gamb., 1896a, 73.—Hoyle, 1890, 539 on Sciæna aquila).—Jackson, 1888, 642, 646, 648, 654.—Maclaren, 1904, 598, 599, 600.—Mont., 1888, 14, 52, 84, 86, 101, 107; 1891, 108; 1892, Oct. 7, 213 (g. of Calceostomine): 1903, 336 (subf. Calceostomine): 1905, 65, 66, 67, 68.—Par. & Perugia, 1890, 12; 1890, 8, sp.—Pratt, 1900a, 646, 654 (key), 657, fg. 49.—St.-Remy, 1898, 524, 564, sp.—Sons., 1890, 172; 1890, 174–176.—Tasch., 1879, 69; 1879, 265 (syn. Dactylogyrus Wag.).

1859: Calceostomum Dies.. 1859c, 441. for Calceostoma.

elegans Ben., 1858a, 1861a, 60-63, 169, 170, 189, 190, 196, pl. 7, figs. 1-8 (in Sciæna aquila; Belgium, —Ben. & Hesse, 1864, 125-126.—Braun, 1890a, 417, 452, 543, 549, 552.—Dies., 1859c, 441.—Jackson, 1888, 646.—Mont., 1888, 8, 10.—Pratt, 1900a, 657, fig. 49.—St.-Remy, 1898, 564.—Sons., 1890, 174, 175-176 (in Sc. umbra; 1891, 263 (in Sc. um.).—Tasch., 1879, 265 (syn. Dactylogyrus calceostoma Ben.) (in Sci. aq.).

inerme Par. & Perugia, 1889, 747 in Corvina nigra : 1890, 747 (in Corv. nig.); 1890,
 8.—Parona, 1894, 139, Genova.—Braun, 1890a, 418, 543, 549, 550; 1891d,
 422.—Sons., 1890, 174, 175-176 (in Corv. nigra, Umbrina cirrosa); 1891, 263

(in Um. cirrhosa).

CALCEOSTOMIDÆ Par. & Perugia. 1890. 19 [as subf.].—Mont.. 1903, 336 [raised from subf. to f. rank] (subf. 1 Calceostominæ (g. Calceostoma. Fridericianella).

CALCEOSTOMINÆ Mont., 1892, Oct. 7, 213 (subf. of Gyrodactylidæ); 1903, 336 (f. Calceostomidæ).—Braun, 1893a, 890.—Gamb., 1896, 73.

CALICOTYLE Dies.. 1850a, 290, 431, 651 (m. kroyeri).—Braun, 1890a, 412, 415, 426, 435, 442, 447, 452, 454, 455, 468, 471, 472, 478, 483, 484, 491, 492, 511, 515, 516, 517, 523, 530, 531; 1893a, 890; 1896b, 7; 1899c, 80–82, 1 fig.—Cerf., 1894, 947; 1898b, 347, 352, 356, 362 (Calycotyle).—Goldb., 1855a, 20.—Goto, 1894a, 226–227; 1893a, 798.—Haswell, 1893e, 112.—Hoyle, 1890, 539.—Ijima, 1884c, 638.—Jackson, 1888, 646, 647, 650, 653.—Juel, 1889, 33.—Kerbert, 1881a, 572 (Callicotyle).—Looss, 1892a, 72.—Mont., 1888, 10, 11, 13, 37, 52, 57, 59, 65, 66, 84, 86, 88, 97, 98 (Callicotyle); 1888, 56, 57, 58, 60 (Callycotyle); 1891, 109, 127; 1903, 336 (subf. Calycotyline); 1905, 70.—St.-Remy, 1898, 523, 540.—Stoss., 1898, 9.—Tasch., 1878, 176 (Callicotyle); 1878, 573; 1879, 49, 50, 54, 57, 58, 60, 62, 64, 65, 66, 68; 1879, 236.

1850: Calycotyle Dies., 1850a, 650.

1858: Callicotyle Dies., 1858e, 313, 362.

1888: Callycotyle Mont., 1888, 56, 57, 58, 60.

1902: Calliocotyle Scott, 1902, 299-300.

1905: Callocotyle Scott, 1905, 117.

kroyeri Dies., 1850a, 431 (on Raja radiata; Kattegat).—Braun, 1890a, 410, 418, 420, 428, 434, 438, 449, 456, 461, 487, 512, 514, 531, 547, 551; 1899, 80, 81.—Cerf., 1894, 948.—Cunningham, 1887a, 278.—Goto, 1891a, 159, 184.—Ijima, 1884c, 638.—Juel, 1889, 14, 37.—Kath., 1894a, 152.—Kroyer, 1852–53a, 961 (kroyeri) (in Raja radiata Don.).—Looss, 1885b, 5, 18.—Mont., 1888, 7, 15, 53 (Callicotyle); 1891, 108, 111, 116, pl. 6, figs. 33–35.—Par. & Perugia, 1890, 6.—Pratt, 1900a, 656, fig. 20, 657.—Stoss., 1898, 9–10.—Tasch., 1878, 176 (Callicotyle); 1878, 573 (Callicotyle); 1879, 48, 49, 52, 55, 56, 61.—Wiergejski, 1877, 18 Oct., 550–561, pl. 31.—Ziegler, 1883, 545.

kröyeri Wierzejski, 1877, 550, pl. 31.—Hoyle, 1890, 539.

kræyeri Hæk, 1856a, 507-512; 1857a, 157-160, 1 fig.

mitsukurii Goto, 1894a, 227–229, pl. 19, t. h. Rhina sp.; Mitsugahama.— Braun, 1899, 80.—Cerf., 1898b, 341.

stossichi Braun, 1899, 80-82, 1 fig. (in Mustelus lævis; Berlin Aquarium).

CALICOTYLEA Dies., 1850a, 290, 431, subtribe of Monocotylea.—Goldb., 1855, 20.

CALICOTYLINÆ Mont., 1903; 1905, 70.

CALLICOTYLE Dies., 1858e, 313, 362 (see Calicotyle).—Ben. & Hesse, 1864, 66, 79.—Carus, 1863, 477.—Mont., 1893, 118.—Scott, 1902, 299–300.—Stoss., 1885, 162.

kroyeri (Dies., 1850) Dies., 1858e, 362 (on Raja radiata, R. batis).—Ben. & Hesse, 1864, 79.—Mont., 1888, 7, 15, 53.—Tasch., 1878, 176; 1878, 573.

kröyeri Wierzejski, 1877, 550-551.—Scott, 1902, 299-300, pl. 13, fig. 30 (on Raja clavata).

kræyerii Ben., 1870, 16.

CALLIOCOTYLE Scott, 1902, 299-300, for Calicotyle, q. v.

kröyeri (Wierzejski, 1877) Scott, 1902, 299–300, pl. 13, fig. 30 (in Raia radiata, R. clavata).

CALLOCOTYLE Scott, 1905. p. 117. for Calicotyle, q. v.

kröyeri (Wierzejski, 1877) Scott, 1905, 117.

CALLODISTOMUM Odhn., 1902, 154 (m. diaphanum).—Pratt, 1902a, 888, 900 (key), related to Anaporrhutinæ.

diaphanum Odhn., 1902, 154 (in Polypterus bichir; White Nile River).

CALYCODES Looss, 1901l, 565 (m. anthos); 1902m, 462 (καλυκώδηε, knospenartig), 463, 839.—Pratt, 1902a. 888, 896 (key).

anthos (Braun, 1899) Looss, 1901l, 565–566; 1902m, 458–463, 869. pl. 21, figs. 17, 18 (includes Dist. anthos Braun, 1899, 720; 1901, 27, pl. 2. figs. 20–22, 24, 31).

CALYCOTYLE Dies., 1850a, 650, for Calicotyle, q. v.

kroyeri (Dies., 1850) St.-Remy, 1898, 540, 541 (in Rhombus maximus). mitsukurii (Goto, 1894) St.-Remy, 1898, 540–541.—Ccrf., 1898b, 341.

CALYCOTYLINÆ Mont., 1903, 336 (subf. of Monocotylidæ).

CAMPULA Cobbold, 1858b, 168; 1859d, 363 (m. oblonga).—Braun, 1893a, 885, 894; 1900g, 249-250, 254; 1901b, 34, 37, 38; 1902b, 5 pp..—Jackson, 1888, 648.—Looss, 1899, 558, 559, 560; 1901, 658; 1901, 208; 1902, 504, 708, 709, 715, 717, 718, 719, 730, 775-778.—Mont., 1888, 92; 1893, 153.—Odhn., 1905, 339, 344.—Pratt, 1902a, 887, 893 (key).—Stiles, 1901, 203, 204, 205.—Stiles & Hass., 1898, 85, 88, 97 ([error] syn. Opisthorchis R. Bl., 1895).

bilis (Braun, 1790) Rail., 1898, 412.

crassiuscula var. janus (Kowal., 1898) Rail., 1898, 412.

felinea (Rivolta, 1884) Kholodk., 1898, 354-355.—Type of Opisthorchis.

janus (Kowal., 1898) Rail., 1898, 412.

oblonga Cobbold, 1858b, 168, pl. 33, figs. 84–85 (in Delphinus phocæna) 1860a, 4; 1879, 419.—Braun, 1900g, 249–254, figs. 1–3; 1902b, 5.—Looss, 1899b, 558, 559, 560; 1902m, 708, 709, 710, 711, 712, 713, 714, 715, 716, 717, 718, 775, 776, 777, 778.—Mont., 1893, 44.—Stiles, 1901, 203, 204, 205.—Stiles & Hass., 1898, 85, 88, 97 (type of Campula).—Stoss., 1892, 16 (to Dist.).

palliata (Looss, 1885) Looss, 1901, 208.—Type of Brachycladium.

poturzycensis (Kowal., 1898) Rail., 1898, 412.

simulans poturzycensis (Kowal., 1898) Rail., 1898, 412.

CAPSALA Bosc, 1811, 384–385 (m. martinieri).—Ben., 1858a, 1861a, 11, 38.—Blainv., 1828, 568–569.—Braun, 1890a, 518.—Dies., 1850a, 428 (syn. of Trist. Cuv. and pp. syn. of Trochopus Dies.).—Johnston, 1865, 30, 33.—Encycl. méthodique, 1824, 164.—Massa, 1906, 51 (of Nord., syn. of Trochopus.)—Nitzsch, 1826, 150–151.—Nord., 1840, 602.—Tasch., 1878, 563, 566 (syn. of Trist.).

coccinea (Cuv., 1817) Blainv., 1828a, 569.—Baird, 1853a, 42.—Dies., 1850a, 429 (to Trist.).—Moquin-Tandon, 1846, 396.—Tasch., 1878, 567 (to Trist.).

elongata (Nitzsch, 1826) Nord., in Lam., 1840, 602 (syn. Nitzschia elegans Baer).— Baird, 1853a, 42 (includes: Hirudo sturionis, Phylline hippoglossi, Trist. elongatum, Nitzschia elegans).—Dies., 1850a, 426 (syn. of Nitzschia elegans).—Johnston, 1865, 33.

maculata (Rud., 1819) Nord. in Lam., 1840, 602.—Dies., 1850a, 430 (to Trist.).—

Tasch., 1878, 567 (to Trist.).

martinieri Bosc, 1811, 384–385 (in Diodon sp. incerta).—Blainv., 1828a, 569.—Dies., 1850a, 430 (syn. of Trist. maculatum).—Moquin-Tandon, 1846, 396–397.—Nitzsch, 1826, 150.—Tasch., 1878, 567 (to Trist.).

papillosa (Dies., 1836) Nord. in Lamarck, 1840, 602 (in Xiphias gladius).—Dies., 1850a, 431 (to Trist.)—Stoss., 1898, 6.—Tasch., 1878, 567 (to Trist.).

rudolphiana (Dies., 1850a) Johnston, 1865, 33.

sanguinea Blainv., see Dies., 1850a, 429 (syn. of Trist. rudolphianum).—Nord., 1840, 602 (syn. Trist. coccineum Rud.) (in Xiphias).—Stoss., 1898, 5.—Tasch., 1878, 567 (syn. of Trist. molæ Blanch.).

tubipora (Dies., 1835)
Nord. in Lamarck, 1840, 602.—Dies., 1850a, 428 (type of Trochopus; renamed T. longipes).—Massa, 1906, 53 (to Trochopus).—Mont., 1891, 123.—Stoss., 1898.
7.—Tasch., 1878, 568 (to Trist.).

CAPSALIDÆ Baird, 1853a, 41.—Johnston, 1865, 32, 299.

CATADISCUS Cohn, 1904, 243 (m. dolichocotyle).

dolichocotyle (Cohn, 1903) Cohn, 1904, 242–243.

CATATROPIS Odhn., 1905, 366, 367 (tod. verrucosa).

verrucosa (Frœlich, 1789) Odhn., 1905, 366–370, fig. 4, pl. 4, fig. 10 (includes Fasc. verrucosa Frœlich, 1789, 112, pl. 4, figs. 5–7; F. anseris Gmelin, 1790a, 3055; Monost. verrucosum Zed. of Levin., 1881, 78; ? e. p. Notocotyle verrucosum Frœlich of Mont., 1892, 40); in Somateria mollissima, S. spectabilis, Mergus serrator; west coast of Sweden and Arctic region; also in Anser domesticus, A. cinereus, A. leucopsis, Cygnus musicus, Bucephala clangula, from Pommern.

species from Spatula clypeata; Egypt.—Odhn., 1905, 369, see Looss, 1899b, 664.

CATHÆMASIA Looss, 1899b, Dec., 562–563 (tod. hians); κατά nach unten; ἡ αίμασιά die Dornen.—Braun, 1901b, 33; 1901g, 896; 1902b, 4.—Pratt, 1902a, 887, 896 (key).—Stoss., 1901, 93 (5).

fodicans Braun, 1901g, 896-897 (in Sterna nigra; Vien. Mus.), 631; 1902b, 4, 5 (in S. nig.).

hians (Rud., 1809) Looss, 1899b, 563 (of Mueh.).—Braun, 1902b, 5.—Stoss., 1901, 93 (5).

CATOPTROIDES Odhn., in Looss, 1902m, 857 (Gorgoderinæ), 861, 862 (tod. spatula).

**spatula* (Odhn., 1902) Odhn. in Looss, 1902m, 857, 862.

spatulæforme (Odhn., 1902) Odhn. in Looss, 1902m, 857.

- CENTROCESTINE Looss, 1899b, 586.—Jægers., 1903a, 14.—Pratt, 1902a, 888, 894 (key), contains Centrocestus, Ascocotyle; related genera, Acanthocasmus, Anoiktastoma.
- CENTROCESTUS Looss, 1899b, Dec., 584, 586 (m. cuspidatus); τὸ κέντρον, spine; ὁ κεστός, Gürtel; 1902m, 832, 833.—Braun, 1902b, 30.—Jægers., 1903a, 14.—Pratt, 1902a, 888, 894 (key).

cuspidatus (Looss, 1896) Looss, 1899b, 582, 584.

- CENTRODERMA Luehe, 1901, 59 (tod. spinossimum). spinosissimum (Stoss., 1883) [Luehe, 1901d, 59].
- CENTROVARIUM Staff., 1904, May 3, 493 (m. lobotes).
 - lobotes (MacCallum, 1895) Staff., 1904, May 3, 493 (in Esox lucius, Stizostedion vitreum).
- CEPHALOBDELLIDEA Dies., 1850a, 291 (subtribe of Monocotylea to contain Astacobdella, Peltogaster, Pachybdella, Trachelobdella, Podobdella, Pontobdella, Ichthiobdella, Branchiobdella), 433, 650.—Goldb., 1855, 20.
- CEPHALOGONIMIN.E Looss, 1899b, Dec., 628; 1900, 561.—Luehe, 1901, 487.488.—Pratt, 1902a, 889, 901 (key), includes Cephalogonimus, Emoleptalea, Prosthogonimus; related genus Stromylotrema.
- CEPHALOGONIMUS Poir., 1886, 22 (m. lenoiri) κεφαλή, γόνιμος.—Braun, 1892a, 642, 645, 696, 704, 705, 713, 715, 733; 1893a, 885, 886, 890, 893, 909, 911; 1895, 138; 1900h, 3; 1902b, 68.—Looss, 1894a, 173, 174; 1899b, 536, 538, 541, 625, 626-627, 628, 721.—Luehe, 1899, 539; 1900, 555.—Moniez, 1896, 89.— Mont., 1888a, 15, 34, 92, 104; 1892, Oct. 7, 214 (gen. of Distominæ); 1893, 82, 95, 154, 157; 1896, 167.—Neumann, 1892, 345.—Pratt. 1902a, 889, 901 (key).— Stiles & Hass., 1898a, 85-86, 96.—Stoss., 1892, 4; 1898, 23.
 - americanus Staff., 1902, 5 Nov., 719-725, 1 pl., figs. 1-4 (in Rana virescens; Ashbridge's Bay, Toronto; May; also? R. clamata); 1902, 30 Dec., 844; 1905, July, 52; 1905, Apr. 11, 687 (int. R. vir.; R. clamata).
 - lenoiri Poir. [1885, 3, pl. 2, figs. 1–2;], 1886, 22–24, pl. 2, figs. 1–3 (in Tetrathyra vaillantii Roch; Sénégal).—Braun, 1892a, 734; 1899b, 715; 1902b, 68.—Looss, 1899b, 626, 627; 1902m, 418 (in Trionyx nilotica), 783.—Luehe, 1899, 539.—Mont., 1893, 83, 102, 105, 106, 107, 157; 1896, 167 (lenori).—Stiles & Hass., 1898a, 85, 96.—Staff., 1902, 719, 725.—Stoss., 1895, 213.

lenori Mont., 1896, 167, for lenoiri.

- ovatus (Rud., 1803) Stoss., 1892, 144; 1896, 126; 1898, 23; 1902, 13 (of 1896, 126, syn. of Cyclocœlum mutabile Zed.).—Hass., 1896a, 2 (syns. Fasc. ov. Rud., Dist. ov. (Rud.); D. bursicola Crep.) (in Gallus dom.).—Looss, 1899b, 629 (type of Prymnoprion), 720, 721.—Luche, 1899, 539 (type of Prosthogonimus).—Mont., 1893, 157.—Rail., 1893a, 368.—Sons., 1890, 134.—Stiles & Hass., 1898a, 85.—Reported also for Accipiter nisus, Anser cinereus dom., Ardea cinerea. Bucephala clangula, Buteo vulgaris, Corvus cornix, Gallinago scolopacina, Harelda glacialis.
- pellucidus (Linst., 1873) Rail., 1890, 138; 1893a, 369.—Braun, 1902b, 68.—Hass.,
 1896a, 2 (in Gallus dom.).—Looss, 1894a, 174.—Mont., 1893, 157.—Stiles & Hass., 1898a, 85.
- CERATIUM Schrank, 1793.—Nitzsch, 1827, 69, contains Cerc. tripos [cf. Ceratium for Keratella Bory, 1824].
- CERCARIA Mueller, 1773, 64–70 (contains gyrinus, catellus, podura, lupus, lemna, cyclidium, tenax, pleuronectes) [apparently lemna is type; cf. Braun, 1889a, 312, 315]; 1786.—Abildg., 1793, 88.—Aitken, 1866, 801, 806, 837; 1872, 144, 148, 204.—Assenova, 1899, 64–65.—Ben., 1882b, 14.—R. Bl., 1888a, 551, 557, 558, 559, 560, 561, 562, 604, 605, 610, 620, 627, 628, 647; 1890h, 2–3.—Bojanus, 1818a, 729–730 [688–689].—Bory de St. Vincent, 1823a, 354–355; 1823b, 355–356 (cercariées); 1824d, 189–191.—Bosc, 1802a, 223–224.—Braun, 1889a, 312 (only malleus and lemna of Mueller's original species of 1773 are true Cercaria); 315 (only inquieta and lemna of Mueller's, 1786, Cercaria are true Cercaria); 1890a, 515; 1892a, 767, 775; 1893a, 884; 1893b, 183 (of Dist recurvatum Linst., in Physa alexandrina).—Bruguière, 1792, 456, 458.—Burm., 1837, 529; 1856a, 246, 250.—Cobbold, 1858a, 9 (structure); 1859d, 365 (in Cervus axis).—Dadai, 1888c, 107–109; 1888e, 105–106, pl. 3, figs. 16; 1888f, 84–86, pl. 3, figs.

11, 13 (Gulf of Naples); 1888g, 107–109, pl. 3, figs. 11, 13.—Dies., 1850a, 286, 293 (of Abildg., syn. of Cheilost.), 294 (of Nitzsch, syn. of Malleolus), 295–298, 299 (of Nitzsch, syn. of Histrionella); 1855a, 377–400 (revision); 1858d, 239–290, 245–246.—Duj., 1845a, 475–478; 1845b, 57–58.—Eichwald, 1829a, 247.—Fewkes, 1822a, 134–145 (with caudal setæ); 1882b, 192.—Gmelin, 1790a, 3891.—Goldb., 1855a, 16.—Herbst, 1787a, 19; 1789a, 129.—Hogg, 1870a, 232–235, pl. 50, figs. 1–3.—Jackson, 1888, 643, 651, 652, 653.—Johnston, 1865, 18.—Lereboullet, 1847a, 300 (migrations); 1847b, 266 (in insects).—Lespès, 1857b, 113–117, figs. 11–16 (of marine mollusks).—Leuck., 1863, 34, 73, 74, 493; 1879a, 41, 96; 1886d, 31, 72, 376.—Lint., 1905, 333, 401 (in Monacanthus hispidus).—Looss, 1892a, 124.—Mont., 1888, 4, 14, 24, 35, 37, 44, 45, 46, 74, 76, 79, 83, 94; 1892, 38.—Mueller, 1786, 119; 1850, 496.—Moul., 1856, 123.—Nitzsch, 1827, 66–69.—Nord., 1840, 617, 630–632.—Pag., 1857, 3, 4.—Parkes, 1891, 267.—Perroncito, 1879, 7–9 (in Rana esculenta); 1880, 454; 1884, 154; 1885, 208–209.—Piana, 1882, 12 pp. (of mollusks); 1882, v. 5.—Quintart, 1905, 724–725 of Barleeia rubra (Adams).—Schrank, 1803, 20.—Sieb., 1839, 153; 1850, 668.—Sons., 1884, 15 Dec., 57–61 (cellules à bâtonnets).—Steenstrup, 1842, 28.—Tasch., 1879, 233.—Vaney, 1901, 29 Apr., 1062–1064; 1901, 105 (in land mollusks).—Villot, 1875, 479 (in Conus mediterraneus).—Wagener, 1834, v. 27 (2), 131–132, pl. 1, fig. 4; 1857, 21.—Ward, 1903, 863, 864.—Wagner, 1883, 120.

1888: Cercavia Mont., 1888, 80, misprint.

I. Baer, 1827b, 621, pl. 31, figs. 1-4.—Dies., 1850a, 295 (syn. of Cerc. vesiculosa).

II. Baer, 1827b, 622, pl. 31.—Dies., 1850a, 296 (syn. of Cerc. chlorotica).

III. Baer, 1827b, 623, pl. 31, fig. 3.—Dies., 1850a, 296–297 (syn. of Cerc. brunnea); 1855a, 387.

V. Baer, 1827b, 625, pl. 31, figs. 5a, a.—Dies., 1850a, 299 (syn. of Histrionella ephemera).

VI. Baer, 1826a; 1827b, 627, pl. 31, figs. 6a, 6b.—Dies., 1850a, 295 (syn. of Malleolus furcatus).

VII. Baer, 1827b, 629, pl. 31, figs. 7a-b.—Dies., 1850a, 297 (syn. of Cerc. fallax); 1855a, 388.

acerca Biehringer, 1884a, 3 (in Onchidium carpenteri Stearns).—Braun, 1892a, 807 (in O. carp.).

aculeata Erc., 1881. See Par., 1894, 163 (in Lymnæa auricularia; Bologna).

affinis Eichwald, 1829a, 247–248, pl. 1, fig. 15a-c (in Lymnæus stagnalis).

agili3 Fil., 1857c, 4–5, pl. 1, fig. 2 (in Lymnæus stagnalis; Turin).—Ben., 1858a, 1861a, 215, 216.—Dies., 1858d, 248–249 to (Gymnocephala).—Par., 1894, 162.
agilis Leidy, 1858a, 110 (free in Delaware River); 1904a, 111.

alata (Hemp. & Ehrenberg, 1828) Moul., 1856a, 213.

alba Erc., 1881e, 12 (for C. brunnea var. Dies.); 1882a, 248.

amphistomi subclavati Ben., 1858a, 82 (in Cyclas cornea, etc.).—Dies., 1859c, 435 (syn. of Diplodiscus subclavatus).—Erc., 1881e, 24, 35; 1882a, 260, 271.—Gerv. & Ben., 1859, 212.—Moul., 1856a, 20, 95, 106–107, 125, 208–211, pl. 6, fig. 10 (syns. Redia gracilis Fil., 1837, Diplodiscus diesingii Fil., Diplocotyle mutabile Dies.) (in Planorbis nitidus; Ticini; P. vortex; Moncalier).

arcuata Steenstrup.—Erc., ——.—See Par., 1894, 164 (in Lymnæa obscura, L.

stagnalis; Bologna).

armata Sieb., 1837, 187 (based on Wagener, 1834, 131); [1835, 336;] 1850, 646, 669; 1854, 18, 21, 26, 27, 28.—Baillet, 1866b, 9, 92, 93, 94, 96.—Ben., 1858a, 1861a, 91, 92, 96, 168, 180, 220.—Biehringer, 1884, 2, 11, 14, 17, 24, 25.—Braun, 1892a, 598, 634, 636, 642, 701, 769, 771, 797, 806, 807, 809, 810, 814; 1893a, 820, 822, 829, 839, 848, 859, 963; 1900, 225.—Cobbold, 1879b, 454.—Desmonceaux, 1868, 21.—Dies., 1850a, 298 (includes Dist. tarda); 1855a, 377, 381, 388 to (Xiphidiocercaria) (in Lymnæus stagnalis, Planorbis corneus); 1858d, 251—252 to (Acanthocephala), larva of Dist. endolobum Duj. (in Plan. corn., Lymn. stag., Paludina impura), 259 (of Fil., 1855b, 305, pl. 1, figs. 2–4, syn. of C. (Acanthocephala) micracantha Dies.); 1859c, 434.—Dowker, 1882a, 9, 11.—Erc., 1881e, 13, 15, 18, 21–23, 48, pl. 1, figs. 32–37; 1882a, 249, 251, 254, 257–259, 284; 1881e, 19–21; 1882a, 255–257 (of Steenstrup).—Fil., 1854a, 6, 7, 8, 9, 10, 12, 22, 24; 1855b, 3–5, 7, 20, 21, 23, pl. 1, figs. 1–4 (in Lymn. pal.; Mon-

calier); 1856a, 257; 1857c, 3, 16, 32.—Florance, 1866a, 4, 11.—Fraipont, 1880a, 397; 1880c, 417, 419, 441; 1881b, 4; 1883a, 35.—Hahn & Lerèvre, 1884a, 516.—Harz, 1881c, 4.—Hoyle, 1890, 538 (cf. Dist. signatum; D. muris).—Juel, 1889, 15.—Kath., 1894a, 136, 139.—Kerbert, 1881a, 556.—Knoch, 1862, 100, 103.—Leuck., 1863, 73, 504, 506, 508, 510, 511, 515, 522, figs. 146, 163, 175, 176, 177; 1879a, 96.—Linst., 1873, 1 (larva of Dist. endolobum Duj.); 1887, 98, 99, 100; 1887, 102 (syn. of Dist. ascidia Ben.).—Looss, 1885b, 19; 1892a, 128; 1894a, 90 (armate), 237, 252.—Macé, 1882, 61.—Mont., 1888, 26, 41, 45, 77, 78, 80; 1888, 196, 197.—Moul., 1856a, 77 (Lymn. stag., Plan. corn.), 78-79, pl. 5, figs. 2-4, 80, 102, 103, 116, 118, 146–152, pl. 5, bis fig. 10, 153, 154, 155, 156, 162, 163, 186, 217, 226, 227, 234 (in Pal. imp. Valette; Berlin; Lymn. stag., Plan. corn., Lymn. palustris; Moncalier).—Nord., 1840, 631.—Pag., 1857, 9, 18-19, 39 (in Lymn. stag.).—Par., 1894, 162.—Roewer, 1966, 217.—Rossbach, 1906, 363.—Schwarze, 1886, 63.—Sons., 1897, 253.—Steenstrup, 1842, 42-47, 51, 57, 78-94, pl. 3, figs. 1-6.—Stoss., 1892, 21.—Tennent, 1906, 661.—Valette St. George, 1855, 18, pl. 1, figs. P-Q.—Villot, 1882, 507.—Vogt, 1878, —, fig. 33.—Wagener, 1834, 131, pl. 1, fig. 4; 1857, 22. calier); 1856a, 257; 1857c, 3, 16, 32.—Florance, 1866a, 4, 11.—Fraipont,

armata minor (Distomi retusi) Ben., 1858, 98, pl. 11, figs. 9-27 (in Lymnæus stagnalis; Belgium).—Dies., 1859c, 434 (syn. of Dist. retusum).

armatæ Looss, 1894a, 90, not as specific name, but as descriptive of several forms and used in the plural.

bilineata Haldemann, 1840a, 3 (in Limnea catascopium; Camden on Delaware).— Dies., 1850a, 300 (to Histrionella).

bipartita Sons., 1897, 253 (in Limnæa palustris; Pisa); 1897, 4.

brachysoma Villot, 1878, 27-28, pl. 9, figs. 1-3 (in Anthura gracilis Leach).—Mont., 1888, 94 (larva of Dist. brachysoma).

brachywra Fil., 1837a, 337, figs. 8-14 (in Planorbis submarginatus: Pavia), teste Par., 1894, 161 (in Plan. submarg.; Pavia), see next entry.

brachyura Dies., 1850a, 296 (Dist. polymorphum Fil., 1837a, 337. figs. 8–14, renamed) (in Planorbis submarginatus; Ticini); 1855a, 386 to (Eucercaria); 1858d, 257. to (Acanthocephala) (syns. C. (Eucerc.) br.; (in Plan. submarg.; Ticini; P. nitidus, P. vortex).—Moul., 1856a, 213 (syn. Dist. polymorphum Fil., 1837).

brachyura Lespés, 1857b, 117, pl. 1, fig. 15 (in Trochus cynereus).—Braun, 1893a,
831.—Dies., 1858d, 257 syn. of C. (Acanthocephala) pachycerca.—Pag., 1862,
297 (in Trochus cinereus).—Villot, 1875, 479 (in Trochus cinereus).

brevicaudata Piana, 1882 (in Helix carthusiana; Reggio Em.) teste Par., 1894, 161, 622.

brunnea Dies., 1850a, 296-297 (Cerc. III. Baer, 1827b, 623, pl. 31, renamed) (in Limnæus stagnalis; Regiomontii); 1855a, 387 to (Eucercaria); 1858d, 247, to (Gymnocephala), syn.C. (Eucerc.) br.; (in Lymn. stag.).—Baillet, 1866b, 96.—Ben., 1858a, 1861a, 89, 180 (syn. of Dist. echinata).—Erc., 1881e, 8–9, 11, 12, pl. 1, figs. 3–9; 1882a, 244–245, 247, 248 (in Paludina vivipara).—Linst., 1887, 104.—Moul., 1856a, 80–81, 118, 157–158 (in Lymn. stag.).—Par., 1894, 162 (in Paludina vivipara). Pal. vivip.; Bologna, Erc.).

buccini mutabilis (Fil., 1855) Dies., 1858d, 266, to (Acanthocephala) (syn. Dist. bucc. mutab. Fil., 1855b) (in Buccinum (Nassa) mutabile; Genoa).—Mont., 1888, 75.—Moul., 1856a, 85, 104, 168.

bucephalus Erc., 1881e, 40, 41–44, 60, 87, pl. 1, figs. 38–42; 1882a, 276, 277–280,
296, 323 (from Unio pictorum).—Braun. 1893a, 834, 839.—Tennent, 1906,
641, 643 (in Unio, Anodonta).—Ziegler, 1883, 541.

capriciosa Cuénot, 1892, in 1-23 (in Synapta inhærens) teste Braun, 1893a, 921 (syn.? Cerc. megacotyle Vill.); 1893b, 183.

capsularia Sons., 1892, 7. Oct., 144-146, pl. 18, figs. 6-7 (in Cleopatra bulimoides; Cairo, Egypt).—Braun, 1893b, 183.—Looss, 1896b, 223, 226, 227, pl. 16, figs. 183–190 (in Cl. bul. Jick.; near Alexandria).

caryophyllata Bory St. Vincent, 1823a, 354 (in Infusions de chènevis).

catellina Mueller, 1786, 130-131, pl. 20, figs. 12-13 (in Aqua fossarum, ubi Lemna).— Bosc, 1802a, 226-227.—Bruguière, 1792a, 456, 462-463.—Nitzsch, 1827, 68 to Dicranophorus.

catellus Mueller, 1773, 65–66 ("in aqua anno ferre in vasculo cum Hydrachnis servata ac in aqua flores per septimanam nutriente"); 1776, 206; 1786, 129–130, pl. 20, figs. 10–11.—Bosc, 1802a, v. 3, 226.—Bruguière, 1792a, 456, 462.—Gmelin, 1790a, 3892.—Herbst, 1789a, 129.—Nitzsch, 1827, 68 to Dicranophorus.

cellulosa sp. inq. Looss, 1896b. 227-229, 232. pl. 14, figs. 159-161 (in Melania

tuberculata Bourg.; near Alexandria, Egypt).

cercopitheci Cobbold, 1861e, 119 (in Cercopithecus fuliginosus).

cervi Cobbold, 1861e, 119 (in Cervus axis).

chlorotica Dies., 1850a. 296 (Cerc. II. Baer, 1827b, renamed) (in Paludina vivipara; Regiomontii); 1855a, 386 to (Eucercaria); 1858d, 252–253 to (Manthocephala) (syn. C. (Eucerc.) chlor. Dies.) (in Pal. vivip.).—Baer, 1827b, 622, pl. 31.—R. Bl., 1888a. 554.—Erc., 1881e. 7–8. 9. 11, 12. pl. 1, figs. 1–2 (in Pal. vivip., P. achatina); 1882a, 243–244, 245, 247, 248.—Fil., 1854a, 7.— Mont., 1888, 75.—Moul., 1856a, 80. pl. 5, fig. 7, 103 (syn. of C. microcotyla Fil.), 157, pl. 5, bis fig. 12 (syn. Cerc. II of Baer; in Pal. vivip.).—Par., 1894, 163.

clausi Mont., 1888, 79 (for clausii).

clausii Mont., 1888, 77, 79 (clausi); 1891, 110.—Eraun, 1893a, 837, 854.—Giard, 1897c, 954, 955 (clausi).—Pintner, 1891, 285-294, 1 pl.; 1892, 619.

columbellæ Pag.. 1862, 306, pl. 29, figs. 1-3 (in Columbella rustica).—Braun, 1889a, 362 (in Col. rust.); 1893a. 831.—Par.. 1894. 163 (in Col. rust.; Spezia).

cometa Bory St. Vincent, 1823a, 354 (in Intusions d'orge).

coni mediterranci Fil., 1857c, 14.—Mont., 1893, 2.—Par., 1894, 163 (in Conus mediterraneus: Mediterranean).

conum Erc., see Par., 1894, 164 (in Bythinia tentaculata: Bologna).

cornuta Bosc, 1802a, v. 3, 224, pl. 32, fig. 2.

coronata Fil., 1855b, 10–13, 15, 23, pl. 1, figs. 11–13 (in Lymnæus palustris. L. stagnalis; Moncalier); 1857b, 426–429, pl. 1, figs. 11–13; 1857c, 4.—Biehringer, 1884, 5, 22.—Dies., 1858d, 249, 250, to (Gymnocephala) (in Lymn. pal., L. stag.).—Moul., 1856a, 100, 111–114 (redia), 117, 118, 202–203 (in Lymn. stag.; Moncalier; L. pal.).—Par., 1894, 162.

cotylura Pag., 1862, 293-305. pls. 28-29, figs. 9-10 (in genital glands of Trochus cinereus; Cette).—Biehringer, 1884. 15.—Braun, 1892a, 809; 1893a, 831,

839.—Levin., 1881a. 81.—Ziegler, 1883, 540.

crassa Erc., 1881 or 1882, see Par., 1894, 164 (in Bythinia tentaculata: Bologna).— Looss, 1894a, 32.

crassicauda Erc., 1881 or 1882, see Par., 1894, 164 (in Bythinia tentaculata; Bologna).

cristata LaValette, 1855, 23. pl. 2. fig. K (in Limnæus stagnalis).—Braun, 1893a, 821. 832, 834, 839; 1893b. 183 (in Cleopatra bulimoides; Cairo. Egypt).—Dies., 1858d. 243 (syn. of Lophocercaria fissicauda Dies.).—Erc., 1881e, 37-41, 42. 43. 44. 87. pl. 1, figs. 23-26 (in Limn. stag.); 1882a. 273-277, 278. 279, 280, 323.—Fil.. 1856b. 86; 1857c. 7-8. pl. 1, fig. 11 (in Valvata piscinalis, Paludina impura. Planorbis submarginatus. Lymn. stag., L. palustris).—Looss, 1896b. 210.—Moul., 1856a, 110, 118, 141, 173 (in Lymn. stag.; Berlin).—Par., 1894, 162.—Ziegler, 1883, 564.

crumena Mueller, 1786, 129, pl. 20, figs. 4–6 (in infusio Ulvæ linzæ marino).— Bosc. 1802a, v. 3, 226.—Bruguière, 1792a, 456, 462.—Nitzsch. 1827, 68–69

(type of Crumena).

cucumerina Erc., 1881, see Par., 1894, 164 (in Bythinia tentaculata; Bologna).— Braun, 1893a, 831.

cycladis rivicolæ Dies., 1850a, 298 (based on Sieb., 1837, 388) (in Cyclas rivicola, liver); 1855a, 400 to Cercariæum.—Moul., 1856a, 85, 168 (in Cyc. rivic.).—Uliený, 1878, 212.

cyclidium Mueller. 1773. 68-69 (in "aquis purioribus frequens"); 1776, 206; 1786, 137-138, pl. 20, fig. 2.—Bosc, 1802a, v. 3, 228.—Bruguière, 1792a, 457, 465.—Gmelin, 1790a, 3891.—Herbst. 1789a, 130.—Nitzsch, 1827, 69 (type of Cyclidium).—Schrank, 1803, 84.

cymbuliæ Græffe, 1860a, 47–49, pl. 10, figs. 4–9 (in Cymbulia peronii Les.; Nizza).— Braun, 1892a, 672; 1893a. 833.—Mont., 1888, 75, 77, 81; 1893, 209.—Pag., 1862, 298.—Par., 1894, 165.

cystophona Will.-Suhm, 1870, 5 (for cystophora).

cystophora Wagener, 1866, 145, 146 (in Planorbis marginatus).—Biehringer, 1884, 227, pl. 16.—Rossbach, 1906, 369.—Sons., 1897, 253.—Ssinitzin, 1906, 686 (larva of Halipegus ovocaudatus).—Stoss., 1892, 23.—Will.-Suhm, 1870, 4, 5.—Ziegler, 1883, 540.—Zuern, 1882, 209.

dichotoma Mueller, in La Valette. 1855, 38 [see Mueller, 1850, 485-500].—Dies., 1858d, 265 (free; Nice), to (Schizocerca).—Mont.. 1888. 77; 1896. 164.—Moul., 1856a, 213-214.—Pag., 1862. 298.—Villot, 1875. 477, 479; 1878, 37.

diesingii (Fil., 1837) Moul., 1856a, 95–96, pl. 5, bis fig. 5 (syns. Diplodiscus diesingii; Redia gracilis) (in Planorbis nitidus, P. vortex).—Dies., 1858d, 272 (syn. Dip-

locotyle mutabilis).

diplocotylea Pag., 1857, 25–27, 49, 52, pl. 3, figs. 9–12 (in Planorbis marginatus; includes Redia gracilis Filippi); 1862, 301.—Braun. 1893a, 839.—Cobbold, 1879b, 452, 454.—Dies., 1858d, 241, 242, 273 (syn. of Diplocotyle mutabilis).—Fil., 1857c, 32.—Kitt, 1885a, 148 (larva of Amphist, subclavatum).—Looss, 1902m, 444 (syn. of Amphist, subcl.).—Villot, 1878, 36 (syn. Diplodiscus diesingii Filippi).

discus Mueller, 1786, 138, pl. 20, fig. 3 (in aqua palustri raro).—Bosc, 1802a, v. 3, 228.—Bruguière, 1792, 457, 465–466.—Nitzsch, 1827, 69 (to Cyclidium).

distomatosa Linst., 1889a, 118, based on Sons., 1884, 98-102 (in Cleopatra bulimoides).—Braun, 1893b, 183 (in Cl. bul.).—Looss, 1896b, 197-204, pl. 14, figs. 152-158 (in Cl. bul.; Fgypt).—Sons. [1884. 61 (Cercaire distomateuse)], 1892, Oct. 7, 144 (in Cl. bul.; Cairo, Egypt).

distomi folii Looss, 1894a, 251 (syn. Dist. duplicatum).

d[istomi] hepatici Looss, 1894a, 252.

distomi homolostomi Linst., 1889a, 120, based on Linst., 1887d, 104-105, pl. 2, figs. 5-6, 17a (in Limnæa stagnalis).

distomi militaris Braun, 1893a, 832, based on Ben., 1858a.

distomi perlati Looss, 1894a, 32.

distomi retusi Linst., 1878a, 327, based on Ben., 1858a, 98, pl. 11, figs. 7-27 (in Limnæa stagnalis).

duplicata (Baer, 1826) Moul., 1856, 77-78, 100, 116, 119, 143-146, 173, 177. pl. 5, fig. 1, pl. 5, bis fig. 9 (to Dist.) (in Anodonta ventricosa, A. anatina. A. cygnea).—Braun, 1893a, 832.—Dies., 1858d, 271 (syn. of Rhopalocerca tardigrada Dies.).—Looss, 1894a, 23, 256, 264.

ccaudata Eichwald, 1829a, 248, pl. 1, figs. 16a-c (in Lymnæus stagnalis).

echinata Sieb., 1837, 187 (in Lymnæus stagnalis; Gedani); 1845, 228; 1850, 671.-Baillet, 1866b, 93 (of Duj.).—Ben., 1858a, 1861a, 86, 180 (syn. of Dist. militare), 89 (to Dist.), 204, 215, 221.—Biehringer, 1884, 5, 7.—Braun, 1883a, 53, fig. 13; 1892a, 770, 772, 797; 1893a, 863; 1906, 144, 79.—Chatin, 1894b, 1356–1358 (excretory canals); 1895a, 20.—Dav., 1877, lxxi. fig. 33.—Dies., 1850a, 297 (syn. of C. fallax); 1855a, 377, 378 (tail), 380, 388, 390 to (Hormocercaria), 391; 1858d, 246, 248, 260-262 to (Nephrocephala) (syn. C. (Hormocerc.) ech. Siebold) (in Paludina vivipara, Planorbis corneus, Lymn. stag.) cerc.) ech. Stebold) (in Paludina vivipara, Planorbis corneus, Lynn. stag.) (larva of Dist. ech.), 261 to (Hormocercaria); 1858e.344 (C. Nephrocephala) ech. syn. of Dist. ech.)—Erc. 1881e, [14], 15, 22, 25–33, pl. 1, figs. 50–58, pl. 2, figs. 1, 2–5, 46, 47, 48, 52, 57, 70, 73, 88, 90; 1882a, [250], 251, 258, 261–269, 282, 283, 284, 288, 293, 307, 309, 324, 326.—Fil. 1854a, 6, 9, 14, 16, 17, 18, 21, 22, 24; 1855b, 10, 11, 19, 20, 21, 22, 25; 1856a, 267; 1857c, 4, 21.—Hahn & Lefevre, 1884a, 516.—Henle, ——, 6.—Keber, 1851a, 90.—Leuck., 1863, 494, 498, 516, 518, 519, figs. 169, 174.—Linst., 1873, 1 (larva of Dist. ech. Zed.); 1873, 106

51–78. pl. 2.—Tennent, 1906, 661.—La Valette, 1855, 16, pl. 1, figs. C-M.—Wagener, 1857, 39, pl. 36A, fig. 12.—Waldenberg, 1860, 21.

echinatoides Fil., 1854a, 14–24, 25, 26, 28, pl. 2, figs. 19–25; 1854b, 266–278, pl. 2, figs. 19–25 (in Paludina vivipara, P. achatina; Lake Varese; Lombardy); 1855b, 11, 12, 13, 16, 18, 19, 23, 25 (syn. C. echinifera La Valette; 1857c, 4, 16, 21.—Baillet. 1866b. 93. 94. 96, 98.—Braun. 1892a. 797.—Dies., 1855a. 377, 380, 382, 387, 391–392 to (Hormocercaria); 1858d. 261, 262–263 to (Nephrocephala (syns. C. (Hormocerc.) echinatoides Fil.: Cerc. echinifera La Cephala (syns. C. (Hormocerc.) echinatoides Fil.: Cerc. echinifera La Valette; Dist. echiniferum Pag.) (adult of Dist. echiniferum La Valette) in Pal. vivip.. P. achatina: 1858e, 346 (C. (Nephrocephala) echinatoides as syn. of Dist. echiniferum).—Erc.. 1881e, 25, 26, 28, 29, 31: 1882a, 261, 262, 264, 265, 267.—Harz, 1881c, 3.—Linst., 1873, 106 larva of Dist. trigonocephalum Rud.) (in Pal. vivip., P. achatina:—Moul., 1856a, [72], 74, 93-94, pl. 5, bis fig. 1, 97, 105-106, 116, 149, 190, 191-201, pl. 6, fig. 6, 202, 203, 226, 227, 228, 229, 234 (syn. C. echinifera La Val.) (in Pal. vivip.: Berlin).

echinifera La Valette. 1855. 14–16. pl. 1. figs. A–F (in Paludina vivipara).—R. Bl., 1888a, 605.—Braun. 1892a, 772.—Dies., 1858d, 261, 262 (syn. of C. (Nephrocephala) echinatoides Fil. —Dolley, 1894a, 984 (= echinatoides .— Erc., 1881e, 25 (= echinatoides), 26, 28, 29, 31; 1882a, 261, 262, 264, 265, 267.—Fil. 1855b, 25 (syn. of C. echinatoides Fil.); 1857c, 21.—Hahn & Lefèvre, 1884a, 516.—Linst., 1873, 106 (larva of Dist. militare Rud.) (in Pal. vivip.).—Moul., 1856a, 105, 109, 110, 191, 192, 200 (syn. of C. echinatoides Fil.).—Pag., 1857, 53.—Will.-Suhm, 1870, 4.

echinocerca Fil., 1855b, 17-19, 22, 23, 25, pl. 2, figs. 19-20 (in Buccinum linnæi; Gulf of Gênes); 1857, 433-435, pl. 2, figs. 19-20.—Dies., 1858d, 268 (Histrionella echinocerca Dies.).—Mont., 1888, 10, 77; 1888, 195, 198; 1888, 194; 1893, 2, 124.—Moul., 1856a, 114-115, 116, 211-212 (in Bucc, linn.); Gulf of Gênes).—Villot, 1875, 479 (in Bucc, linn.).

efemera Ssinitzin. 1905, 158 for ephemera.

elegans Mueller, in La Valette. 1855, 13, 38, pl. 2, fig. 2, based on Mueller 1850, 496, free form.—Braun, 1893a, 831, 832.—Dies., 1858d, 269 (to Histrionella).— Mont., 1888, 194, to (Hist.).—Moul., 1856a, 214.—Pag., 1862, 298.—Villot, 1875, 479: 1878. 35.

ephemera Nitzsch (1807). 33-36; 1817, 29. pl. 1. figs. 1-13 (in Planorbis corneus; Halle); 1827. 67-68 | Pl. corn.).—Baillet, 1866b, 95, 96, 98.—Biehringer, 1884. 5.—Braun, 1892a, 770, 772; 1893a, 848, 849, 851, 858.—Brett, 1881b, 141.—Dies., 1850a, 299 to Histrionella ; 1855a, 392 (to Hist.); 1858d, 244 (of Sieb., syn. of Glenocerc. flava). 267 (of Wagener to Hist.).—Duj., 1845a, 478.—Eichwald, 1829a, 247, 248.—Erc., 1881e, 23; 1882a, 259 (of Nitzsch as doubtful syn. of Cerc. tripunctata Erc. .-Fil., 1855b, 20, 22, 25 (syn. C. doubtful syn, of Cerc. tripunctata Erc.).—F11., 1855b, 20, 22, 25 (syn. C. flava LaValette); 1857c, 31.—Florance, 1866a, 12.—Hemp. & Ehrenb., 1828a (to Hist.).—Leuck., 1863, 494, 515, 522, 523.—Looss, 1896b, 194, 197.—Moul., 1856a, 77, 94-95, pl. 5, bis fig. 3, 116, 204-208, pl. 6, figs. 4, 5 (syns. Hist. ephemera Hemp. & Ehrenb.; C. flava LaVal.) (in Plan. corn. by Sieb. and Val.; Paludina impura by Baer; P. vivipara by Nitzsch and Baer).—Nord., 1840, 631 (eyes; cf. Hist. Ehrenb.).—Pag., 1857, 24-25, pl. 2, figs. 7-12 (in Plan. corn.).—Sieb., 1835, 70; 1837, 187, 189; 1850, 670; 1854, 14, 18, 19, 25, figs. 1, 4-9, 13, 14.—Sons., 1897, 252.—Ssinitzin, 1905, 158 (efemera).—Steensstrap, 1842, 37, 51-55, 56, 65, 94, —Wagener, 1857, 21 strup, 1842, 37, 51-55, 56, 65, 94.—Wagener, 1857, 21.

ericetorum Linst., 1898, 761-762, fig. 8 in Xerophilus ericetorum; near Gœttingen, Germany).

- erfoliata Moul., 1856a, 87, 116, 180-183, pl. 6, figs. 13, 14 [new name for Dist. of Leucochloridium paradoxum].—Cobbold, 1879b, 435.—Dies., 1858d, 277 (syn. of Leuc. par. Carus).—Erc., 1881e, 55 (Cerc. of Leuc. par.); 1882a, 291.—Zeller, 1874.
- exiqua Looss, 1896b, 230-232, pl. 16, figs. 181-182 (in Cleopatra bulimoides; Egypt).

fallax Dies., 1850a, 297 (includes "Cerc. echinatum Siebold?" Dist. pacifica; Cerc. VII. Baer) (in Paludina vivipara, Limnæus stagnalis); 1855a, 380, 387 to (Eucerc.) 388, 391 (Dies., 1850a, partim as syn. of C. (Hormocerc.) echinata); 1858d, 247–248, 262, to (Gymnocephala) (syn. C. (Eucerc.) fallax Dies.) (in Lym. stag.; Pal. vivip.).—Baer, 1827, 629, pl. 31, figs. 7a, 7b.—Ben., 1858a, 1861a, 86 (syn. of Dist. militare).—Bojanus, 1818, 729, pl. 9, figs. A-F.—Fil., 1857c, 32.—Henle, ——, 6.—Moul., 1856a, 184 (syn. of C. echinata Sieb.).—Mueller, 1843.—Pag., 1857, 3, 4, 6, 21, 23–24, pl. 3, fig. 1 (in Lim. stag.).—Schwammerdam, ——, 75.—Sieb., 1845, 228; ——, 187.—Steenstrup, ——, 51, 78, pl. 2.

fascicularis Villot, 1875, 480, pl. 14, fig. 4 (in Nassa reticulata; Roscoff).

fissicanda Moul., 1856a, 109 (for fissicauda).

fissicauda La Valette, 1855, 21, pl. 2, figs. 6, H (in Limnæus stagnalis).—Braun, 1893a, 821; 1893b, 183 (in Physa alexandrina).—Dies., 1858d, 265 to (Schizocerca) (in Lym. stag.; Berlin).—Fil., 1857c, 23.—Mont., 1888, 196, 197 (fissicaudata); 1888, 9, 76, 77 (fissicaudata).—Moul., 1856a, 109 (fissicanda), 118, 171 (in Lym. stag.; Berlin).—Sons., 1897, 253.—Villot, 1878, 32, 36–37, pl. 10, figs. 9–11 (in Scrobicularia tenuis).

fissicaudata Mont., 1888, 196; 1888, 9, 76, 77 (for fissicauda).

flava La Valette, 1855, 24–25, pl. 2, figs. 8, a. g. (=C. ephemera Nitzsch, renamed.)—Dies., 1858d, 244 (to Glenocerc.).—Fil., 1855b, 25 (syn. of C. ephemera).—Moul., 1856a, 204 (syn. of C. ephemera).

flavopunctata Par., 1894, 703 (for C. fulvopunctata).

folii Looss, 1894a, 256 (syn. Dist. duplicatum).

forcipata Mueller 1786, 134–135, pl. 20, figs. 21–23 (in aqua palustri rarissime).— Bosc, 1802a, v. 3, 227.—Bruguière, 1792, 457, 464.—Nitzsch, 1827, 68 to Dicranophorus.

fulgopunctata Braun, 1893a, 831 (for fulvopunctata).

fulvopunctata Erc., 1881 or 1882, see Par., 1894, 161 (in Bythinia tentaculata; Bologna).—Spengel, 1905, 258.

furcata Nitzsch, [1817], 49, pl. 2, figs. 12–18; 1827, 68 (syn. Vibrio malleus Mueller).—Baer, 1827, 626–629.—Braun, 1893a, 821, 846.—Dies., 1850a, 295 (to Malleolus, only species, ?type by tautonymy malleus).—Fil., 1855b, 21, 22, 23; 1857c, 19, 23, pl. 2, fig. 31.—Moul., 1856a, 77, 84–85, pl. 5, fig. 13, 115, 116, 118, 120, 168, 169, 170, 171 (= Cerc. VI. of Baer) (in Lymnæus stagnalis; Paludina vivipara by Baer).—Nitzsch, 1827, 68 (syn. Vibrio malleus).—Nord., 1840, 631.—Pag., 1857, 4.—La Valette, 1855, 22, pl. 2, fig. J.

[furcatæ Eichwald, 1829a, 247.]

gibba Mueller, 1773, 120; 1786, 120–121, pl. 18, fig. 2 (in infusione jungermanniæ tamarisci).—Bory St. Vincent, 1823a, 355 (in infusions de Jungermannes).—Bruguière, 1792, 456, 458.—Nitzsch, 1827, 68, to Macrocercus.—Schrank, 1803, 85.

gibba Fil., 1854a, 13, 26, pl. 1, fig. 18 (in Lymnæus pereger); 1854, 266, pl. 10, fig.
15: 1855b, 23.—Bosc, 1802a, v. 3, 225.—Dies., 1855a, 389 to (Xiphidiocerc.);
1858d, 257, to (Acanthocephala) (syn. C. (Xiphidiocerc.) g. Fil.) (in Lym. per.; Turin).—Erc., 1881e, 17-18, pl. 1, figs. 29-31; 1882a, 253-254.—Moul., 1856, 82, 159-160, pl. 5, bis fig. 15 (in Lym. per.).—Par., 1894, 162.—Ssinitzin, 1906, 686 (in Limnæa stagnalis, L. palustris; Warschau), larva of Opisthioglyphe endoloba.

globipora Erc., 1881 or 1882, see Par., 1894. 164 (in Bythinia tentaculata; Bologna).—Looss, 1894a, 47, 48.

gorgoderæ cygnoides Kowal., 1904. 24 (9) (in Cyclas cornea; Dublany).

gorgoderæ loossi Ssinitzin, 1905, 44–46, pl. 1, figs. 3, 14 (syn. Cerc. macrocerca Wagener); 1906, 683 (in Epithecea; Warschau).

gorgoderæ pagenstecheri Ssinitzin, 1905, 46-47, pl. 1, figs. 4, 7, 13, pl. 2, figs. 15, 19, 20 (syn. C. macrocerca Thiry); 1906, 683.

gorgoderæ varsoviensis Ssinitzin, 1905, 47–49, pl. 1, fig. 5, pl. 2, figs. 16, 18; 1906, 683.

gorgoderæ vitelliloba Ssinitzin, 1905, 49-51 (syn. Cerc. macrocerca Fil., 26, pl. 10, fig. 7); 1906, 683.

gracilis La Valette, 1855, 20-21, pl. 1, fig. 13 (in Planorbis corneus).—Ben., 1858a. 1861a, 220.—R. Bl., 1888a, 555.—Dies., 1858d, 264—265 (in Plan. corn.; Berlin (to (Schizocerca), 270.—Fil., 1857c, 23.—Moul., 1856a, 108–109, 116, 170– 171 (in Plan. corn.: Berlin).—Wagener, 1866, 146.

nyrinus Mueller, 1773, 64-65 in infusioni animali raro : 1776, 206; 1786, 119-120 (syn. Macrocercus corpore globoso), pl. 18, fig. 1.—Bory St. Vincent. 1823a, 354–355 (in infusions animales).—Bosc, 1802a, v. 3, 224–225.—Bruguière, 1792, 456, 458.—Gmelin. 1790a. 3892–3893.—Herbst, 1789a, 129, pl. 80.— Nitzsch, 1827. 68 to Macrocercus.

haimeana (Lacaze-Duthiers, 1854) Moul., 1856, [87] 178-179, pl. 6, fig. 12 (in Ostrea edulis, Cardium rusticum: Mahon, Cette).—Badcock, 1875a, 145.—Claparède, 1863, 10-12, pl. 4, figs. 8-9 (syn. Bucephalus haimeanus Lacaze-Duthiers.—Dies., 1858d, 276 (to Bucephalopsis).—Giard, 1874e, 486.—McCrady, 1874. 180.

haimejana Erc., 1881e, 41 (Bucephalus haimeanus, renamed); 1882a, 277.

helicis aspersæ (Dies., 1855) Moul., 1856c, 83-84, 166-167 (based on Duj., 1845a, 472 in Helix aspersa .—Dies., 1858d, 277 (to Cercariæum).—Par., 1894, 164.

helicis carthusianellæ [Erc., 1881e, 95, pl. 1. figs. 45–47; 1882a, 331 (Cerc. dell' Helix carthusianella)].—Par., 1894, 164 (Bologna).

helicis maculosæ [Erc., 1881e, 95, pl. 1, figs. 48, 49; 1882a, 331].—Par., 1894, 164 (Bologna).

helicis viviparæ Dies., 1850a, 298 for Dist. Bojanus, 1818, 730; Vilnæ ; 1855a, 399. hirta Mueller, 1786, 128. pl. 19, figs. 17, 18 in aqua marina bis tantum).—Bosc, 1802a, 226.—Bruguière, 1792, 456, 461.—Nitzsch, 1827, 69 (type of Coleps).

histrionella Ehrenb.—Wagener, 1834, 131-132.

hyalocauda Haldemann 1840a .- Evarts, 1880a, 230-232, figs. 34-36 (in Physa heterostropha Say

hymenocerca Villot, 1875, 479-480, pl. 14, figs. 5-7 in Calyptræa sinensis; Roscoff). imbricata Looss, 1893a, 20 (in Bythinia tentaculata Gray=Paludina vivipara Lam.; near Leipzig; 1896b, 192-197 (sub Monost, verrucosum; in Melania tuberculata Bourg.); 1902m. 444 (sub Notocotyle verrucosa).—Sons., 1897, 252.

incistidata Perroncito [1879, 7-9.]. 1880, 454-457, 1 fig. in Rana esculenta).—Erc., 1881e. 13. 14. 15, 30, 32, 57, 63; 1882a, 249, 250, 251, 266, 268, 293, 299.

inermes Looss. 1892a, 128, not as specific name, but meaning unarmed cercariæ.

inquieta Mueller, 1786, 121-122. pl. 18, figs. 3-7 (in aqua marina).—Baer, 1826a, 125.—Ben., 1858a, 1861a, 312.—Bory St. Vincent, 1825b, 253.—Bosc, 1802a, v. 3, 225.—Bruguière, 1792, 456, 458-459.—Dies., 1850a, 300 (to Histrionella).—Mueller, J., 1850, 496.—Moul., 1856a, 213 (of Nitzsch, 1817).—Nitzsch, 1817, 47; 1827, 67, 68.—Villot, 1875, 479.

isopori Looss, 1894. 55 (in Cyclas rivicola).

lacrhyma Bory St. Vincent, 1823a, 354 in infusions d'orge et d'avoine .

lota Lespès. 1857. 114-116. pl. 1. fig. 13 (in Venus decussata; Arcachon).—Braun, 1893a. 832.—Dies., 1858d, 251. to (Gymnocephala) (in V. dec.).—Florance, 1866a, 7.—Pag., 1862, 297, 298, 299 (in V. dec.).—Villot, 1875, 479 (in V. dec.).

 lemna Mueller. 1773. 67-68 (in aquis paludosis): 1776, 206; 1786, 122-123. pl. 18, figs. 8-12; ——. 2485.—Abildg., 1793. 89.—Baer, 1826a, 125.—Bory St. Vincent. 1825b, 253.—Bosc, 1802a, v. 3, 225.—Bruguière, 1792a, 456, 459.—Dies., 1850a, 299 (to Histrionella).—Gmelin, 1790a, 3892.—Herbst, 1789a, 130.—Hermann, —, 160. pl. 3. figs. 43a-d.—Moul., 1856a, 213.—Nitzsch, 1827, 67, 68.—Schrank, 1803, 79–80.—Wagner, 1832, 394. pl. 4, figs. 1–6.

leptosoma Villot, 1878, 32-33, pl. 9, figs. 4-5 (=Cerc. stage of Dist. leptosomum Crep.) (in Scrobicularia tenuis).—Nicoll, 1906, 517, 518 [larva of Echinost.

leptosomum] (in Scrob. ten.).

leucochloridii Leuck., 1858a, 114. see Leucochloridium paradoxum.

 limacis (Dies., 1850; Moul., 1856, 83, pl. 5, fig. 11, pl. 8, fig. 12, pl. 9, figs. 8–9; 103, 163–164 pl. 8, figs. 19, 20, pl. 19, figs. 11–12; 165 in Limax cinerea; Genève).—
 R. Bl., 1888a, 554.—Braun, 1893a, 831.—Dies., 1858d, 259 (syn. of C. (Acanthocephala) trigonocerca Dies.).—Erc., 1881e, 59; 1882a, 295.

limnææ ovatæ Linst., 1884, 142; 1887d, 98, 99, 100 (in Limnophilus rhombicus), 105.—Stoss., 1889, 66 (syn. of Dist. endolobum).

limnææ truncatulæ Linst., 1892, 331–332, pl. 15, fig. 17 (in Limnæa truncatula).

linearis Lespès, 1857, 117, pl. 1, fig. 16 (in Littorina littorea).—Braun, 1893a. 381.—Dies., 1858d, 258 (in Litorina litorea) to (Acanthocephala).—Pag., 1862, 297 (in Litt. litt.).—Villot, 1875, 479.

longicaudata Piana, 1882, see Par., 1894, 161 (in Helix nemoralis, H. carthusiana; Reggio Em.) (see also longocaudata).—Dolley, 1894a.

longocaudata Piana, 1882, teste Par., 1894, 622 (misprint?) (in Helix carthusiana), as larva of Dist. lanceolatum.

lophocerca Fil., 1857c, 5, pl. 1, figs. 3-4 (in Paludina impura).—Braun, 1893a, 832.—Dies., 1858d, 245 (to Glenocerc.).—Erc., 1881e, 24, 25, 35; 1882a, 260, 261, 271.—Looss, 1894a, 252.—Par., 1894, 163 (in Bythinia tentaculata, Pal. vivipara, P. achatina; Torino by Fil., 1858).

luna Mueller, 1786, 139, pl. 20, figs. 8, 9 (in eadem aqua, ac proxime precedens, rarissime [C. orbis]; [Zool. Dan. prodr. addend., 1776, 280].—Bosc, 1802a, v. 3, 228.—Bruguière, 1792a, 457, 466.—Nitzsch, 1827, 69 to Lecane.

lungo-caudata Piana, see R. Bl., 1888a, 603 (in Helix carthusiana), see also longicaudata.

lupus Mueller, 1773, 67 (in aquosis, ubi Lemna vegetat, rara); 1776, 206; 1786, 131–133, pl. 20, figs. 14–17.—Bosc, 1802a, v. 3, 227.—Bruguière, 1792a, 456, 463.—Gmelin, 1790a, 3892.—Hemp. & Ehrenb., 1828a (type of Cycloglena).—Herbst, 1789a, 130.—Nitzsch, 1827, 68, to Dicranophorus.—Schrank, 1803, 83-84.

lutea (Ben., 1870) Giard, 1897c, 954-956 (to Brachyccelium luteum Ben.) (in Tapes decussatus, T. pullastra at Arcachon; Donax trunculus; Wimereux); 1897d, 957; 1903h, 27–28.—Caullery & Chappellier, 1906, 325.—Pelseneer, 1895, 357.—Also reported for Pholas candida.

lymnæi auricularis (Fil., 1854) Moul., 1856, 84, pl. 5, fig. 12; 167 (in Lymnæus auricularis by Fil.).—Dies., 1858d, 279 (to Cercariæum).—Looss, 1894a, 32,

to (Dist.).

lymnæi obscuri Erc., 1881e, 33–35, pl. 1, figs. 16–17, 46; 1882a, 269–271, 282 (in (Limnæus obscurus, L. stagnalis).—Par., 1894, 162 (in Lymn. obs.; Bologna

macaci Cobbold, 1861, 119 (in Macacus radiatus).

macorcerca Looss, 1894a, 253 (misprint for macrocerca).

rocerca Fil., 1854a, 13, 26, pl. 1, figs. 15–18 (in Cyclas cornea; Turin); 1855, 341, pl. 1, figs. 15–17; 1855b, 22, 24.—Baillet, 1866b, 93.—Biehringer, 1884, 2–3, 7, 9, 16, 17, 18, 22, 26.—Braun, 1883, 56; 1890e, 595; 1891c, 218; 1891f, 369; 1892a, 634, 808, 809, 810; 1892b, 188; 1893a, 831, 832, 854.—Claparède, 1863a, 12.—Cobbold, 1879b, 470.—Dies., 1855a, 389 to (Xiphidiocerc.); 1858d, 255, to (Acanthocephala) (syn. C. (Xiphidiocerc.) macr. Fil.) (larva of Dist. cygnoides Zed.) (in Cyclas cornea; Turin); 1858e, 334 to (Acanthocephala) (as syn. of Dist. cyg.)—Fraipont, 1880a, 397; 1880c, 419, 441; 1883a, 35.—Gamb., 1896a, 72.—Hahn & Lefèvre, 1884a, 516.—Jackson, 1888, 651.—Juel, 1889, 15.—Kerbert, 1881a, 556.—Kowal., 1902d, 27 (9) [larva of Gorgodera cygnoides, in Cyclas cornea].—Leuck., 1863, 511, 512.—Linst.. 1873, 1 (larva of Dist. cyg. Zed.).—Looss, 1885b, 19; 1892a, 125; 1894a, 23, 63, 129, 236, 244, 251, 253 (macorcerca), 256, 264, pl. 6, fig. 129; 1896b, 227; 1902m, 444 (sub Gorgodera cygnoides).—Mace, 1882, 61.—Mont., 1888, 41.—Moul., 1856, 82, 119, 160–161, pl. 6, figs. 1, 2; 173 (in Cyc. corn.; Turin).—Pag., 1862, 299.—Par., 1894, 165.—Pratt, 1898, 361.—Ssinitzin, 1905, 49 (of Fil., syn. of vitelliloba), 46 (of Thiry, syn. of pagenstecheri), 44 (of Wagener, syn. of macrocerca Fil., 1854a, 13, 26, pl. 1, figs. 15-18 (in Cyclas cornea; Turin); 1855, 341, vitelliloba), 46 (of Thiry, syn. of pagenstecheri), 44 (of Wagener, syn. of loossi); 1906, 682.—Thiry (1859), v. 10 (2), 271–277, pls. 20–21, figs. 1–13; (1860), v. 10, 271–277, 2 pls.—Uličný, 1878, 212, 214.—Villot, 1882, 507.—Wagener, 1857, pl. 29.—Ziegler, 1883, 540, 547 (in Limnæa sp.).

magna Pag., 1857, 3, 4, 22–23, pl. 2, figs. 13–14 (in Paludina vivipara).—Dies., 1858d, 247, to (Gymnocephala) (in Pal. vivip.; Heidelberg).

major Nitzsch, 1817, 44, pl. 2, figs. 1–8; 1827, 68 (syn.? Brachionus proteus).— Dies., 1850a, 299 (syn. of Histrionella lemna).—Moul., 1856a, 213.—Mueller, 1850, 497.—Pag., 1857, 23.

megacotyla Dies., 1858d, 263, to (Nephrocephala) (syn. Dist. echinatoides Pag., nec Fil.) (in Anodonta cygnea; Heidelberg).

megacotylea Villot, 1878, 30, pl. 9, fig. 7 (in Mysis).—Braun, 1893b, 183 (?syn. of

C. capriciosa | .—Cuénot, 1892.

melanoglena Pag., 1862, 298 [?Melanoglena bipunctata renamed].

micracantha Dies., 1858d, 259–260 syn. C. armata Fil., 1855b, 3–5) to (Acanthocephala) in Triton punctatus. Lymnæus palustris).—Linst., 1887, 100; 1878a, 205 in Tr. tæniatus).

205 (in Tr. tæniatus).

microcotyla Fil., 1854a, 7-12, 13, 26, pl. 1, figs. 5-10 (in Paludina vivipara, P. achatina: Lake Varese and Lombardy): 1854b, 260-265, pl. 10, figs. 5-10; 1855b, 6, 22, 23, 25 (syn. C. pugnax La Valette): 1856b, 85, 86; 1857c, 7, 15, 31.—Baillet, 1866b, 93.—Biehringer, 1884, 15.—Braun, 1892a, 809; 1893b, 183 (in Melania tuberculata; Cleopatra bulimoides.—Dies., 1855a, 378, 390 to (Xiphidiocerc.): 1858d, 253 to (Acanthocephala) (syn. C. (Xiphidiocerc.) micr. Fil.: C. pugnax La Valette) (in Pal. achat.: Lombardia: P. vivip.) (larva of Dist. tetracystis Gast., 254 (syn. of C. Acanthocephala) vesiculifera Dies.): 1858e, 348 (sub Acanthocephala as syn. of Dist. tetracystis.—Erc., 1881e, 7, 8, 9, 10, 11-15, pl. 1, figs. 10-15 (in Pal. vivip., P. achat.; liver and genital organs: Limnæus obscurus. Planorbis corneus, 71: 1882a, 243, 244, 245, 246, 247-251, 307.—Linst., 1873, 1 (larva of Dist. tetracystis Gast.).—Moul., 1856a, 80 (microtyla), pl. 5, figs. 5-6, 103, 104, 153-157, pl. 5, bis fig. 11:160, 161 (syns. C. pugnax La Valette, C. chlorotica Dies., Cerc. II. Baer (in Pal. vivip., P. achatina: Lac de Varèze and Berlin).—Sons., 1884, 58: ——, 136-137; 1897, 253.

microcristata Erc., 1881 or 1882. see Par., 1894, 161 (in Bythinia tentaculata; Bologna).

microtyla Moul., 1856a, 80 for microcotyle).

micrura Fil., 1857c, 5–6. pl. 1, figs. 5, 6 (in Paludina impura); 1859. — .—Biehringer, 1884, 3, 26.—Braun, 1893a, 831.—Dies., 1858d, 258. to (Acanthocephala) (in Pal. imp.: Turin .—Looss, 1894a, 47 (= C, globipora=Dist, globiporum Rud.) (in Bythinia tentaculata : 1896b, 208.—Mont, 1888, 76.—Par, 1894, 163.—Ssinitzin, 1906, 685 (in Bythinia tentaculata : 686 (in Nephelis vulgaris but not Limnea . larva of Sphærost globiporum.—Villot, 1878, 36.

minuta Erc. 1881 or 1882, see Par. 1894, 163 (in Bythinia tentaculata; Bologna), minuta Nitzsch. 1817, 46, pl. 2, figs. 9-11 (in various mollusks at Halle); 1827, 68.—Dies., 1850a, 295; 1855a, 385 to Eucerc.; 1858d, 246 to (Gymnocephala) (in various fresh-water mollusks at Halle).—Moul., 1856a, 213.—Spengel, 1905, 258.

miocerca Mont., 1888, 77 for myocerca ..

mirabilis Braun. 1891c. 218 (in Limnæus palustris var. corvus); 1891f. 369; 1892b. 187-188 (in Limn. pal. corv.: Kurischer Lowlands); 1893a, 821, 832, 834; 1893b. 183.

mongeotii Bory St. Vincent. 1823a. 354 in water .

monostomi Linst. 1896i. in 376-377 perhaps young of Monost. mutabile) (in Lymnæa ovata. L. peregra: Weende. Germany): 1897a, 109.

myocerca Villot. 1878, 35–36, pl. 10, figs. 12–13 in Scrobicularia tenuis .—Braun, 1893a, 832.—Mont., 1888, 9, 77 miocerca .

myzura Pag., 1881, 18 in Neritina fluviatilis).—Braun, 1893a, 831.

neglecta Fil., 1854a, 24, 26, pl. 2, figs. 26, 27 (in Lymnæus pereger); 1854, 278, pl. 11, figs. 26–27; 1855b, 23.—Dies., 1855a, 386 to (Eucerc.), 387; 1858d, 246 to (Gymnocephala).—Moul., 1856, 94, pl. 5, bis. fig. 2; 116, 204, pl. 6, fig. 9 (in Lymn, per.).—Par. 1894, 165.

nodulosa Linst., 1873, 3, pl. 1, fig. 5 in Bythinia tentaculata): 1873, 142, figs. 30-32: 1878a, 323 = Dist. nodulosum .—Looss, 1894a, 40 in By. ten. by Linst. (=C, virgula Fil., and C, rostroaculeata) (not Dist. nodulosum).

obscura Sons., 1892. Oct. 7, 138 (in Limnæa natalensis; Cairo, Egypt): 1896, 321.—Braun, 1893b, 183 (in Limn, nat.).

ocellata La Valette. 1855, 22–23, pl. 2, fig. 5 (in Limnæus stagnalis).—Braun, 1893a, 821, 834.—Dies., 1858d, 270 (syn. of Histrionellina fissicauda).—Erc., 1881e, 35–36 (in Planorbis corneus, liver J. 49, pl. 1, figs. 43, 44, pl. 2, fig. 18; 1882a, 271–272, 285.—Moul., 1856a, 109–110, 116, 118, 172–173 (in Lymn, stag.; Berlin).—Par. 1894, 161.—Sons., 1897, 253.

odontocotyla Dies., 1858d, 264 (in Lymnæus stagnalis; Berlin).

opaca Bory St. Vincent, 1823a, 354 (in infusions de pois).

orbis Mueller, 1786, 138–139, pl. 20, fig. 7 (in aqua, ubi Lemna, rarissime); [Zool. dan. prod., 1776, 280].—Bosc, 1802a, v. 3, 228.—Bruguière, 1792a, 457, 466.— Nitzsch, 1827, 69 to Lecane.

ornata La Valette, 1855, 18-19, pl. 1, figs. N-O (in Planorbis corneus).—Braun, 1892a, 806; 1893a, 832.—Cobbold, 1876. 211; 1879, 454.—Desmonceaux, 1868, 22.—Dies., 1858d, 241, 244 to (Acanthocephala), 255-256 to (Acanth.) (larva of Dist. clavigerum Rud.) (in Plan. corn., Berlin; Hydrachna concharum; Heidelberg); 1858e, 388 to (Acanth.); 1859c, 434.—Fil., 1857c, 31, 32.—Florance, 1866a, 11.—Harz, 1881c, 4.—Leuck., 1863, 495, fig. 171.—Linst., 1873, 1 (larva of Dist. clavigerum Rud.).—Looss, 1894a, 90, 99, 237, 250, 252, 257.—Moul., 1856, 108, 120, 152-153 (in Plan. corn.; Berlin).—Pag., 1857, 13-18, 52, pl. 1, figs. 1-13 (in Plan. corn.); 1862, 301.—Rossbach, 1906, 389. 1906, 389.

ovalis Schrank, 1803, 86.

orata Villot, 1878, 29–30, pl. 9, fig. 6 (in Lygia oceanica). pachycerca Dies., 1858d, 257–258. to (Acanthocephala) (syn. C. brachyura Lespès) (in Trochus cinereus; Francogalliæ).

pachycerca Claperède, 1863a, 12-13, pl. 18, fig. 1 (free form; St. Vaast).—Braun, 1893a, 831.—Par., 1894, 165 (in Cydippe sp.; Nizza; see Leuck., in Carus, 4884, Prod., 133).—Villot, 1878, 35 (of Dies. or Clap.?).

pacifica Ben., 1858a, 1861a, 86 (syn. of Dist. militare).

paludinæ impuræ Baer, 1827. 655, see Dies., 1855a, 400 (syn. of Cercariæum erythrops, and of C. melanops).—Moul., 1856, 96, pl. 5, bis fig. 4, 212–213, pl. 6, fig. 8; 216 (syn. Dist. paludinæ impuræ Fil.) (in Paludina impura; Lake de Varèse).

papillosa Erc., 1881 or 1882, see Par., 1894, 164 (in Bythinia tentaculata; Bologna). parva Erc., 1881 or 1882.—Par., 1894, 164 (in Bythinia tentaculata; Bologna).

pectinata Huet. 1891a, 12, Apr., 162–165, figs. 1–4 (in Donax anatinum). pelagica Mont., 1888, 45, 46.

pigmentata Sons., 1892, Oct. 7, 142–144 (in Physa alexandrina, P. micropleura; Cairo, Egypt) of an Amphist. (sp.).—Braun. 1893b, 183 (in Ph. micr.).—Looss, 1902m, 444 (sub Amphist. conicum).—Stiles, 1898a, 64, 65, 143.

planorbis carinati Dies., 1858d, 266, to (Acanthocephala) (in Planorbis carinatus: Turin).

planorbis cornei Dies., 1850a, 298 (in Planorbis corneus) (based on Henle, 1835. 597); 1855a, 400 (to Cercariæum).—Moul., 1856a, 85 (in Plan. corn.) (Henle,

platyura Leidy, 1891a, 416 (free in pool with Lymnæus; Fort Bridger, Wyo.).

pleurolophocerca Sons., 1892, Oct. 7, 138-139 (in Melania tubercolata. Cleopatra bulimoides; Cairo, Egypt).—Braun, 1893b, 183 (in Mel. tub., Cl. bul.).— Looss, 1896b, 204–210, pl. 13, figs. 140–145 (in Mel. tub.; Egypt).

pleuronectes Mueller, 1773, 70 ("In aqua ultra sex septimanas in vasculo servata); 1776, 206; 1786, 135, pl. 19, figs. 19–21.—Bosc, 1802a, v. 3, 227.—Bruguière, 1792a, 457, 464.—Gmelin, 1790a, 3891.—Herbst, 1789a, 130.—Nitzsch, 1827, 69 to Phacus.—Schrank, 1803, 85-86.

 podura Mueller, 1773, 66 (''Ín paludosis Lemna coopertis, Novembri et Decembri''); 1776, 206; 1786, 124–125. pl. 19, figs. 1–5.—Bory. St. Vincent, 1825a, 84 (to Furcocerca).—Bosc, 1802a, v. 3. 225.—Bruguière, 1792a, 456, 460.—Gmelin, 1790a, 3892.—Hemprich & Ehrenberg, 1828a, type of Ichthydium.—Herbst, 1789a, 130.—Nitzsch, 1827, 68 to Enchelys.

polymorpha (Baer, 1827) Moul., 1856, 86, 116, 174–177, pl. 6, fig. 11; 179.—Dies., 1858d, 275 (to Bucephalus).—Erc., 1881e, 41, 42, 43, 44–45, 60, 87; 1882a, 277, 278, 279, 280–281, 296, 323.—Par., 1894, 164 to (Bucephalus).—Ziegler,, 1883, 541.

pomatiæ Vaney & Conté, 1899, 194-196, figs. 1-2 (in Helix pomatia; Lyon).

prima Ssinitzin, 1905, 147-153; 1906, 687 (in Aplexa hypnorum, Planorbis vortex compressa; Warschau) (encysts in Corethra, Ilvbius; Warschau).

proxima Lespès, 1857, 116, pl. 1, fig. 14 (in Littorina littorea).—Dies., 1858d, 249-250, to (Gymnocephala).—Pag., 1862, 297 (in Litt. litt.).—Villot, 1875, 479.

pugio Linst., 1887, 105, pl. 2, figs. 7, 17c (in Limnæa ovata).

pugnax La Valette, 1855, 19, pl. 1, fig. R (in Paludina vivipara).—Dies., 1858d, 247.—Erc., 1881e, 11 (= C. microcotyla Fil.), 12; 1882a, 247, 248.—Fil., 1855b, 25 (syn. of C. microcotyla Fil.); 1857c, ——.—Moul., 1856, 103, 153, 157 (syn. of C. microcotyla).—Pag., 1857, 20-21, 22, pl. 1, figs. 17-24 (in Pal. vivip.).

punctum Erc., 1881 or 1882, see Par., 1894, 164 (in Bythinia tentaculata; Bologna).
pusilla Looss, 1896b, 229–230, 231, 232, pl. 16, figs. 178–180 (in Vivipara unicolor Olivier; Damanhour).

pyrula Bory St. Vincent, 1823a, 355 (in infusion de chènevis).

renale (Fil., 1855) Moul., 1856, 164–165 (in Helix aspera; Turin).—Dies., 1858d, 265–266 (renalis) to (Gymnocephala) (in H. adspersa; Turin).

rigonocerca Braun, 1892a, 809 (for trigonocerca).

rostrata Erc., 1881 or 1882, see Par., 1894, 164 (in Bythinia tentaculata; Bologna).—Looss, 1894a, 40.

rostroaculeata Erc., 1881, see Par., 1894, 164 (in Bythinia tentaculata; Bologna).—Looss, 1894a, 40 (see nodulosa).

sagitata Lespès, 1857, 114, pl. 1, fig. 12 (in Nassa reticulata; Arcachon).—Dies., 1858d, 249.

sagitta Pag., 1862, 297 (in Nassa reticulata), for sagitata.

sagittalis Vaney & Conte, 1899, 196 (C. sagittifera Sieb., 1854, renamed) (in Helix pomatia).

sagittata Dies., 1858d, 249, to (Gymnocephala) for sagitata (in Buccinum (Nassa) reticulatum), 250.—Villot, 1875, 479.

sagittifera Sieb., 1854, 18, fig. 3 (in Helix pomatia).—Dies., 1855a, 398 (syn. of Cercariæum helicis pomatiæ); 1858d, 278 (syn. of Cercariæum h. p.).—Moul., 1856, 82, pl. 5, fig. 10; 83, 163 (in H. pom.).

secunda Ssinitzin, 1905, 153; 1906, 687 (in Corethra, Ephemera larvæ; Warschau).

setifera O. F. Mueller, 1786, 127–128, pl. 19, figs. 14–16 (in aqua marina raro).— Bosc, 1802a, v. 3, 226.—Bruguière. 1792a, 456, 461.—Mont., 1888c, 193–199.— Nitzsch, 1827, 69 to Trichoda.

setifera Moul., 1856a, 214, and Dies., 1858d, 250-251 (Triest), to (Gymnocephala), based on J. Müeller, 1850, 497 (free in sea water; Marseilles).—Braun, 1893a, 832, 853.—Claparède, 1863a, 12, 13.—Dadai, 1888f, 85; 1888g, 108.—Giard, 1897c, 954, 955 (setigera).—Mont., 1888, 9, 10, 23, 77, 78, 79, 80 (Cercavia); 1888, v. 2, 193-199; 1891, 519; 1892, 717; 1893, 1, 2, 13, 47, 52, 55, 61, 70, 122, 124, 125, 209.—Moul., 1856a, 214.—Pag., 1862, 298.—Par., 1894, 165 (syn. Macrurochæta acalepharum).—Villot, 1875, 477, 479; 1878, 32, 33-35, pl. 10, figs. 1-8 (in Scrobicularia tenuis).

setigera Giard, 1897c, 955 (for setifera, 1856).

species Wagener, 1857, in liver of Limnæus stagnalis.—Braun, 1893a, 821 (is an Echinost.).

spermatica Blumenbach, see Chiaje, 1833, 34.

spinifcra La Valette, 1855, 17, pl. 1, figs. 10–11 (in Paludina vivipara, Planorbis corneus).—Braun, 1893a, 832.—Dies., 1858d, 263–264, to (Nephrocephala) (in Plan. corn.; Berlin).—Erc., 1881e, 25, 26; 1882a, 261, 262.—Looss, 1894a, 252.—Moul., 1856, 109, 110, 202, 203 (in Plan. corn.; Berlin).

stylosa Linst., 1875, 193-195, pl. 3, figs. 16-17 (in Planorbis vortex); 1878a, 329 (in Plan. vor.).

subulo Pag., 1857, 19-20, pl. 1, figs. 14-16 (in Paludina vivipara).—Dies., 1858d, 256-257, to (Acanthocephala) (in Pal. vivip.; Heidelberg).

syrinus Kûech., 1855, 471, misprint for gyrinus.

tellinæ balticæ Dies., 1850a, 298 (in Tellina baltica; Gedani), based on Sieb., 1837, 388; 1855a, 400 (to Cercariæum).—Moul., 1856, 85, 168 (in Tell. balt.).—Villot, 1875, 479 (in Tell. balt.).

tenax Mueller, 1773, 69–70 (in infusione sordium dentium intra quatriduum); 1776, 206; 1786, 136–137, pl. 20, fig. 1.—Bosc, 1802a, v. 3, 228.—Bruguière, 1792a, 457, 465.—Chiaje, 1833, 37–38.—Gmelin, 1790a, 3891.—Herbst, 1789a, 130.—Nitzsch, 1827, 69, to Phacus.—Olfers, 1816, 24.—Schrank, 1803, 84–85.

terrestris Linst., 1889c, 241, pl. 16, fig. 14 (in Helix lens; Greece); 1889d.

terricola Linst., 1889c, 241, pl. 16, fig. 13 (in Helix vermiculata; Algiers); 1889d. thaumanthiadis Braun, 1889a, 357 (in Eucope); 1893a, 832, for thaumantiatis.

thaumantiadis Pag., 1862, 298, for thaumantiatis.—Linst., 1887a, 334 (in Eucope

sp.).

thaumantiatis Græffe, 1860a, 49-51, pl. 10, figs. 10-12 (in Thaumantias Græffe, ccelenterate, not Bonaparte, 1854, bird, compare Thaumantias Eschsch., 1829, Acal.).—Braun, 1889a, 357 (thaumanthiadis in Eucope); 1893a, 832 (thaumanthiadis).—Mont., 1888, 193, 194, 195; 1888, 77.

tricaudata Schrank, 1803, 86.

trigonocerca Dies., 1858d, 259, to (Acanthocephala) (syn. C. limacis Moul.) (in Limax cinereus, L. (Arion) rufus; Geneva).—Biehringer, 1884, 15.

triloba Fil., 1857c, 3. pl. 1, fig. 1 (in Planorbis carinatus, Lymnæus stagnalis;
Turin).—Dies., 1858d, 252, to (Acanthocephala) (in Lym. stag., Plan. car.).—
Erc., 1881e, 15-17, pl. 1, figs. 20-22; 1882a, 251-253 (in Lim. stag., Plan. car.;
Po, in Piemonte).—Par., 1894, 161 (in Plan. corn., P. carinatus, Lym. stag.; Torino).

tripes Bosc, 1802a, v. 3, 227.

tripos Mûeller, 1776, 206 ("in aqua marina"); 1786, 136, pl. 19, fig. 22.—Bruguière, 1792a, 457, 464–465.—Nitzsch, 1827, 69.

tripunctata Erc., 1881e, 23–25, pl. 1, figs. 27–28; 1882a, 259–261 (syn. C. ephemera Nitzsch) (in Planorbis corneus; liver and genital organs; Italy).—Mont., 1888, 10.—Par., 1894, 161 (in Plan. corn.; Bologna).

tripus Gmelin, 1790a, 3892, for tripos Mueller.

tuberculata Fil., 1857c, 8-9, pl. 2, figs. 19, 20 (in Paludina impura).—Braun, 1893a, 827.—Dies., 1858d, 248, to (Gymnocephala) (in Pal. imp.; Turin).—Harz, 1881c, 4.—Looss, 1894a, 138.—Par., 1894, 163.

turbo Müeller, 1786, 123–124, pl. 18, figs. 13–16 (in aqua rivulari cum Lemna rarior).—Abildg., 1793, 79.—Bosc, 1802a, v. 3, 225.—Bruguière, 1792a, 456, 459–460.—Nitzsch, 1827, 68, type of Urocentrum.

varicans Abildg., 1794, 89, pl. 3a, figs. 1–4 ("Hab. primitivum ignotum, in aqua rivulari").—Dies., 1850a, 293 (to Cheilostomum as type).—Moul., 1856a, 213 (syn. Ch. varicans Dies.).—Pag., 1857, 6.

varsoviensis Ssinitzin, 1906, 683, to (Gorgodera).

vermicularis Mueller, 1786, 133-134, pl. 20, figs. 18-20 (in aqua ubi Lemna vegetat).—Bosc, 1802a, v. 3, 227.—Bruguière, 1792a, 457, 463–464.—Nitzsch, 1827, 68, to Dicranophorus.

vesicata Uličný, 1878, 211–217, pl. 6, figs. 1–5 (in Cylcas rivicola; Mähren).— Braun. 1893a, 831, 832; 1893b, 183; 1898a, 1581.—Looss, 1894a, 227 (= C. macrocerca Fil.).

vesiculifera Dies., 1855a, 378, 389, to (Xiphidiocerc.) (C. vesiculosa Fil., 1854, renamed) (in Paludina vivipara); 1858d, 254-255, to (Acanthocephala) (syns. C. vesiculosa Fil.; C. (Xiphidiocerc.) vesiculifera Dies.; C. microcotyla Fil.) (in Pal. vivip., P. achatina).

vesiculosa Dies., 1850a, 295–296 (in Paludina vivipara; based on Cerc. I. Baer, 1827b; Regiomontii, Berlin, Heidelberg); 1855a, 385 to (Eucerc.); 1858d, 254 (of Fil.), syn. of C. (Acanthocephala) vesiculifera, 254 (syn. C. (Eucerc.) vesiculosa Dies.) to (Acanthocephala).—Braun, 1892a, 797, 806.—Erc., 1881e, 9–10, pl. 1, figs. 18–19, 12, 52; 1882a, 245–246, 248, 288 (in Pal. achatina, P. vivip.; Italy).—Fil., 1854a, 12–13, 22, pl. 1, figs. 12–14; 1855b, 23; 1857c, 14–15, 16, 17, pl. 2, figs. 22–23, 24, 25 (in Pal. vivip., P. ach.; Lake Majeur).— Moul., 1856, 81–82, 103–104, 158–159, pl. 5, bis figs. 13. 14; 260 (in Pal. vivip.; Berlin).—Pag., 1857, 21–22, pl. 2, figs. 1–4 (in Pal. vivip.).—Par., 1894, 163.—La Valette, 1855, 19–20, pl.1, fig. S.

vesiculosa of Fil., 1854a, 12, 13, 22, 30, pl. 1, figs. 12–14 (in Paludina vivipara).— Dies., 1855a, 389 (renamed C. (Xiphidiocerc.) vesiculifera); 1858d, 254 (syn.

of C. (Acanthocephala) vesiculifera Dies.).

villoti Mont., 1888, 194 (C. setifera Mûeller of Villot, 1879, in Scrobicularia tenuis, renamed), 195, 197; 1888, 77.—Giard, 1897c, 954.

rirgula (Fil., 1837) Dies., 1850a, 296: 1855a, 386. to (Eucerc.); 1858d, 260, to (Acanthocephala) (syn. C. (Eucerc.) virg. Dies.) (larva of Dist. maculosum Rud.) (in Valvata piscinalis, Paludina impura).—Biehringer. 1884, 15, 18.—Braun, 1892a, 749, 809; 1893a, 859.—Fil., 1855b, 5, 10, 19, 22, 23, pl. 1, figs. 5–10 (= Dist. virg.) (in Pal. imp.; Pavia Moncalier); 1856b, 85; 1857c, 6-7, pl. 1, figs. 9, 10 (in Pal. imp.).—Florance, 1866a, 8.—Harz. 1881c, 4, 7, 8.—Leuck., 1863, 504, 505, 506, fig. 173.—Linst., 1901, 197.—Looss, 1894a, 40.—Moul., 1856a, 51, 82, 99, 104, 161-163 (in Pal. imp.; Pavia Moncalier).—Par., 1894, 163.

viridis Mueller, 1786, 126–127, pl. 19, figs. 6–13 (in aquis fossarum stagnantibus primo vere).—Bory, 1825a, 84.—Bosc 1802a, v. 3, 226.—Bruguière, 1792a, 456., 460–461.—Nitzsch, 1827, 68, to Enchelys.—Schrank, 1803, 80–83.

vitrina Linst., 1887, 105–106, pl. 2, figs. 8, 17c (in Zebrina detrita; Frauenberg b. Sondershausen).—Braun, 1893a, 831.

vittelliloba (Olss., 1876) Ssinitzin, 1906, 683, to (Gorgodera).

vivax Sons., 1892, Oct. 7, 137–138, pl. 18, fig. 3 (in Cleopatra bulimoides; Cairo, Egypt).—Braun, 1893b, 183 (in Cl. bul.).—Looss, 1896b, 210–223, pl. 15, figs. 162-177 (in Cl. bul.; Egypt, Tunis; Melanopsis præmorsa).—Rossbach, 1906, 370.

CERCARIÆ, plural of Cercaria.

gorgoderæ cygnoidis Zeder, 1800, of Kowal., 1904 (9), 24 (in Cyclas cornea; Dub-

CERCARIÆA Nitzsch, see Dies., 1850a, 286, 292; 1855a, 383-400; 1858d, 241-243.— Goldb., 1855a, 15.—Mont., 1888, 83.

Goldb., 1855a, 15.—Mont., 1888, 83.

CERCARLEUM Dies., 1855a, 397-400 (collective group and not as true generic name, for little known and doubtful forms of Cercariæa Nitzsch).—Bettend., 1897a, 4, 5, 7, 9, 12, 23, 30, 32, 33, 37, 39, 41, 42, 43, pl. 1, figs. 1, 2, 3, 5, 7, pl. 2, figs. 11, 15, 16, 17, pl. 3, figs. 18, 19, 20, 21, 22, 23, pl. 4, figs. 24, 25, 30, pl. 5, figs. 31, 32; 1897, 309, 311, 313, 316, 327, 334, 336, 337, 343, 345, 346, pl. 28, figs. 1-3, 5, 7, pl. 29, figs. 11, 15-17, pl. 30, figs. 18-23, pl. 31, figs. 24, 25, 30, pl. 32, figs. 31, 32 (in Helix hortensis).—Blochmann, 1892b, 649-652 (in Helix hortensis).—Blochmann & Bettend., 1895a, 218, 219, figs. 4, 5 (in Helix hortensis).—Brand., 1898a, 214 (22).—Braun, 1893a, 836, 857.—Darr, 1902, 694.—Kowal., 1898h, 158 (55).—Looss, 1894a, 236.—Sons., 1897, 252 (Cercaryæum).

ancyli lacustris Dies., 1855a, 400, based on Baer, 1827b, 656, in Ancylus lacustris; 1858d, 282.

coni mediterranei (Fil., 1857c) Dies., 1858d, 282-283.

coregoni feræ Chavannes, —, 62, see Dies., 1858d, 283.—Linst., 1879a, 266 (in Coregonus fera Jur.).

cycladis rivicola (Dies., 1850) Dies., 1855a, 400 (in Cyclas rivicola); 1858d, 282.— Braun, 1893a, 830.

echinatum (Fil., 1837a) Dies., 1858d, 280 (syn. Heterost. ech. Dies.) (in Paludina impura; Ticinum).

erythrops Dies., 1855a, 400 (based on Cerc. paludinæ impuræ Baer, 1827b) (Regiomontii); 1858d, 270 (to Histrionellina).

flavescens (Pag., 1857) Dies., 1858d, 278 (in Bulimus radiatus; Heidelberg).

helicis Meckel [? see helicis pemetia].—Praun, 1891d, 424 (in Helix nemoralis; H. strigella); 1893a, 829, 856.—Hofmann, 1899a, 174, 175, 177, 178–185, 188, 193, 201, pl. 11, figs. 1, 3, 4, pl. 12, figs. 3, 4 (becomes mature in Erinaceus europæus to Dist. leptosomum, syn. D. caudatum).—Roewer, 1906, v. 41, 185–228, figs. 1, 2, 3, pl. 14, figs. 1–7, pl. 15, figs. 8–22; 1906, 5 Oct., 616–617; 1906, 9 Oct., 596–599; 1906, 15 Nov., 340–341.—Ziegler, 1905, 37.

helicis alternatæ Dies., 1855a, 398 (=Dist. helicis Leidy, renamed) (in Helix alternata; Philadelphia); 1858d, 278 (syn. of C. vagans).—Leidy, 1857, 44.

helicis aspersæ Dies., 1855a, 398 (for helicis asperæ Dies., 1850a, 302, based on Duj., 1845a, 472); 1858d, 277 (syn. Cerc. h. a. Moulinie) (in Helix aspersa; Rennes). helicis pomatiæ (Dies., 1850) Dies., 1855a, 398; 1858d, 278 (syn. Cercaria sagittifera

Siebold) (in Helix pomatia).

limacis (Dies., 1850) Dies., 1855a, 397; 1858d, 277 (in Limax agrestis, L. cinereus, L. rufus; Rhedoni).—Braun, 1893a, 831.—Jackson, 1888, 651.

CERCARIÆUM-Continued.

lymnæi auricularis (Fil., 1854) Dies., 1855a, 398; 1858d, 278-279 (in Lymnæus

lymnæi palustris Dies., 1855a, 399 (for Heterost. limnæi Dies., 1850a, 302, based on Duj., 1845a, 473) (=Dist. lymnæi Dies., 1850, renamed); 1858d, 279 (in Lymnæus palustris; Rennes).—Sons., 1897, 252.

lymnæi peregri Dies., 1858d, 279 (in Lymnæus pereger; Turin).

melanops Dies., 1855a, 400 (based on Cercaria paludinæ impuræ Baer, 1827b, 655); 1858d, 270 (to Histrionellina).

naidis proboscidex Dies., 1858d, 283.

ovatum (Dies., 1850) Dies., 1858d, 279-280 (syns. Heterost. ov. Dies., Dist. luteum La Valette) (adult=Dist.luteum Wagener, in Esox lucius) (in Paludina vivipara) (Berlin, Heidelberg, Regiomontii).—Mont., 1893, 190.

paludinæ impuræ (Fil., 1854) Dies., 1855a, 399 (in Paludina impura); 1858d, 280

(syn. of C. pal. imp. inerme).

paludina impura armatum (Fil., 1854a) Dies., 1858d, 281 (in Paludina impura).— Par., 1894, 161 (in Bythinia tentaculata).

paludinæ impuræ inerme (Fil., 1854a) Dies., 1858d, 280 (syns. C. pal. imp. Dies., Cerc. pal. inerme Morl., Dist. pal. imp. inerme Fil., adult=Dist. perlatum Nord.) (in Paludina impura).—Linst., 1873. 1 (larva of Dist. perlatum Nord.).—Par., 1894, 162 (in Bythinia tentaculata; Pisa).

paludinæ impuræ (tentaculorum) Dies., 1855a, 399 (based on Dist. pal. imp. Baer, 1827b, 655) (in Paludina impura; Regiomontii); 1858d, 281.

paludina vivipara Dies., 1855a, 399 (=Cercaria helicis vivipara Dies., 1850a, 298, renamed); 1858d, 279 (in Paludina vivipara; Vilnae).

physæ fontinalis (Baer, 1827) Dies., 1855a, 400; 1858d, 282.

planorbis carinati (Fil., 1857) Dies., 1858d, 281–282.

planorbis cornei (Dies., 1850) Dies., 1855a, 400; 1858d, 281 (syn. of C. plan. cornei (ovariorum)).

planorbis cornei (hepatis) Dies., 1858d, 281 (in Planorbis corneus).

planorbis cornei (ovariorum) Dies., 1858d, 281 (syn. C. planorbis cornei Dies.) (in Planorbis corneus).

spinulosum Hofmann, 1899a, 178, 185, 193, 201, pl. 11, fig. 2, of pulmonates, see Dist. spinulosum of Erinaceus europæus.

tellinæ balticæ (Dies., 1850) Dies., 1855a, 400; 1858d, 283.—Braun, 1893a, 830.— Pag., 1862, 298.

vagans (Leidy, 1850) Dies., 1855a, 398; 1858d, 278 (syns. Dist. helicis Leidy, D. pericardium Crep., D. vagans Leidy, C. helicis alternatæ Dies.) (in Helix albolabris, H. alternata; Philadelphia).

CERCARYÆUM Sons., 1897, 252, for Cercariæum.

CERCORCHIS Luehe, 1900aa, 566 (tod. linstowi), subg. of Telorchis; κέρκος, tail.— Braun, 1901a, 14; 1901b, 58; 1901i, 58.—Looss, 1902m, 831, 832, 834, 835. 1899: Telorchis Looss, 1899b, 566 (tod. linstowi).

linstowi[i] (Stoss., 1890) Luehe, 1900aa, 566.

CHARAXICEPHALUS Looss, 19011, 7 Nov., 621–622 (m. robustus); 1902, 541, 542, 562, 579, 582, 589, 590, 591, 596, 597, 598, 599, 600, 601, 602, 609, 611, 612–613 (diagnosis) (Pronocephalidæ).—Pratt, 1902a, 890, 910 (key).

robustus Looss, 19011, 621–622 (in Chelone mydas; Egyptian coast); 1902m, 416,

541-545, pl. 25, figs. 65-69, pl. 26, fig. 89, 548, 597, 599, 612.

CHEILOSTOMUM Dies., 1850a, 286, 293 (m. varicans); 1855a, 379, 381, 394; 1858d, 273.—Burm., 1856a, 250.—Fil., 1857c, 8.—Goldb., 1855a, 15.—Moul., 1856a, 121, 123.

varicans (Abildg., 1794) Dies., 1850a, 293; 1855a, 394 (Hab. primitivum ignotum); 1858d, 273 (free; Dania).—Moul., 1856a, 213 (to Cerc.).—Pag., 1857, 6.

CHIORCHIS Fischder., 1901, 374 (m. fabaceus); 1902a, 43-44.—Looss, 1902m, 442.— Pratt, 1902a, 887, 893 (key).—Shipley, 1905, v. 6, 8.

dilatatus Daday, 1905, 233 (in Colossoma brachypoma; Paraguay).

fabaceus (Dies., 1839) Fischder., 1901, 374; 1902a, 44-46, 53 (in Manatus exunguis, M. latirostris; Brazil and North America); 1903h, 528.

oxycephalus (Dies., 1836) Daday, 1905, 233 (in Colossoma brachypoma).

CHONCHOSOMUM Galli-Valerio, 1901c, 364, for Conchosomum. alatum (Gœze, 1782) Galli-Valerio, 1901c, 364 (in dog).

CHORICOTILE Parona & Perugia, 1889, 743, (for Choricotyle).

CHORICOTYLE Ben. & Hesse, 1863; 1864, 96, 109 (m. chrysophryi).—Braun, 1890a, 477, 498, 516, 517, 522, 546.—Cerf., 1895h, 918, 920; 1896, 514, 515.—Mont., 1888, 8, 11, 86, 89, 99; 1903, 336 (syn. of Diclidophora).—Par. & Per., 1890, 10, 13.—Tasch., 1879, 240.

1889: Choricotile Par. & Perugia, 1889, 743, for Choricotyle.

chryophrii Cerf., 1898a, 303 (for chrysophryi).

chrysophris Mont., 1888, 11, 16 (for chrysophryi).

chrysophryi Ben. & Hesse, 1863; 1864, 109–110, pl. 11, figs. 16–22 (in Chrysophrys aurata).—Cerf., 1898a, 303 (chryophrii).—Mont., 1888, 11, 16 (chrysophris).—Tasch., 1879, 247 (to Octobothrium).

marionis St. Loup, 1885, 176 (in Mæna vulgaris).

taschenbergii Par. & Perugia, 1889, 743 (Choricotile); 1890, 743 (Choricotile) (in Sargus rondeletii; Genova); 1890, 6.—Braun, 1890a, 418, 535, to Octobothrium.—Cerf., 1898a, 303.

CLACOCŒLIUM Stiles & Hass., 1898a, 89, misprint for Cladocœlium.

CLADOCALIUM Pontallié, 1853, 104, 105 (misprint for Cladoccelium).—Stiles, 1904i, 21.—Stiles & Hass., 1898, 86, 89.

CLADOCŒLIUM Duj., 1845a, 382, 388, 389 (m. hepaticum), as subg. of Dist.—R. Bl., 1891, 609, 610.—Braun, 1893a, 885, 886, 909, 910; 1895, 138.—Looss, 1899b, 535.—Mont., 1888, 38, 92, 105; 1893, 150, 154.—Stiles, 1904i, 21.—Stiles & Hass., 1898a, 86, 89 (Clacocalium).—Stoss., 1892, 4, 7, raised to generic rank.

1853: Cladocalium Pontallié, 1853, 104, 105, misprint.

1898: Clacocœlium Stiles & Hass., 1898a, 89, misprint.

delphini (Poir., 1886) Stoss., 1892, 10 (in Delphinus delphis).

elephantis (Dies., 1858) Stoss., 1892, 9 (in Elephas indicus; Rangoon).

giganteum (Dies., 1858) Stoss., 1892, 9–10 (syns. Dist. gig., Fasc. gig., Dist. magnum) (in Camelopardalis giraffa, Cervus aristotelis, C. canadensis, C. elaphus, Dama vulgaris, Portax picta).—Stiles, 1898a, 49, 51; 1904i.—Ward, 1895, 253 (syn. of Fasc. magna); 1903, 866 (in part syn. of Fasc. magna).

hepaticum (Linn., 1758) Stoss., 1892, 7–9 (syns. Fasc. hepat., Dist. hepat., Dist. caviæ) (in Antilope dorcas; Bos bubalis, Egitto; B. taurus; Padova, America; Camelus bactrianus, Cavia cobaja, Capra hircus, Cervus capreolus, C. elaphus; C. virginianus, North America; Castor fiber, Dama vulgaris, Elaphus indicus, Equus asinus, E. caballus, Padova, North America; Felis domestica, Homo, Dalmazia, Spagna, Zurigo, Liverpool: Lepus cuniculus, L. timidus, Macropus giganteus; Ovis aries, Padova, Normandia, Mecklenburg, Offenbach, Buenos Aires; Ovis argali; Portax picta, India; Sus scrofa).—Braun, 1903, 147 (to Fasc.): 1906, 150, fig. 83 (to Fasc.).—Duj., 1845a, 389.—Rail., 1893a, 342.—Stiles, 1898a, 29; 1904i, 22.—Ward, 1895, 327 (in Homo); 332 (in Bos taurus), 335 (in Ovis aries), 338 (in Equus caballus); 1903, 865 (to Fasc.).

holostomum (Rud., 1819) Stoss., 1892, 145.—Braun, 1902b, 136, syn. of Urogonimus macrostomus.

palliatum (Looss, 1885) Stoss., 1892, 10–11 (in Delphinus delphis). rochebruni (Poir., 1886) Stoss., 1892, 11 (in Delphinus delphis). sulcatum (Linst., 1883) Stoss., 1892, 145.

(CLADOCŒLIUM) Duj., 1845a, 382, 388, 389 (m. hepaticum), subg. of Dist.

hepaticum (Linn., 1758) Duj., 1845a, 389–390, see Fasc.

macrocotyle (Dies., 1858) Stoss., 1886, 63.

miescheri Zschokke, 1890, 764 (in Trutta salar).

pagelli (Ben., 1870) Stoss., 1886, 59.

veliporum (Crep., 1837) Stoss., 1886, 64.

CLADORCHINÆ Fischder., 1901a, 372; 1902a, 35; 1903h, 490, subf. of Paramphistomidæ.—Looss, 1902m, 439.—Pratt, 1902a, 887, 892 (key), includes Gastrodiscus, Homalogaster, Diplodiscus, Cladorchis, Chiorchis; related genus Balanorchis.—MacCallum, 1905, 668.—Shipley, 1905, v. 6, 4, 8.—Stiles, 1904i, 45.

- CLADORCHIS Fischder., 1901a, 372 (tod. pyriformis); 1902a, 35–36, 43, 45.—Looss, 1902m, 836.—MacCallum, 1905, 668.—Pratt, 1902a, 887, 893 (key).—Shipley, 1905, v. 6, 8.
 - asper (Dies., 1839) Fischder., 1901, 373; 1902a, 39 (in Tapirus americanus); 1903f, 602-606, pl. 29, figs. 82-85.
 - giganteus (Dies., 1836d) Fischder., 1901, 373 to (Stichorchis); 1902a, 41–42, 43 (in Dicotyles albirostris, D. torquatus); 1903f, 611–614, pl. 30, figs. 89–92.
 - helostomatis MacCallum, 1905, 673–678, fig. B (in Helostoma temmincki; Palembang, Sumatra).
 - pangasii MacCallum, 1905, 668-673, fig. A (in Pangasius nasutus; Palembang, Sumatra).
 - pyriformis (Dies., 1839) Fischder., 1901, 373; 1902a, 36–38, 39, fig. 3 (in Tapirus americanus); 1903h, 596–602, figs. 78–81.
 - schistocotyle Fischder., 1901, 373, to (Taxorchis) (in Dicotyles torquatus); 1902a, 40–41 (syn. Amphist. giganteum Dies., 1835 e. p.; Brazil), to (Taxorchis) (in Dic. torq.); 1903f, 607–610, pl. 29, figs. 86–87, pl. 30, fig. 88 (schistolotyle).
 - subtriquetrus (Rud., 1814) Fischder., 1901, 373–374, to (Stichorchis); 1902a, 35, 42–43, to (Stichorchis) (syns. Amphist. subtriq. Rud., Dist. amphistomoides Boj.) (in Castor fiber); 1903h, 567, to (Stichorchis); 1903f, 615–620, pl. 30, figs. 93–96.
 - watsoni (Conyngham, 1904) Shipley, 1905, 129–135, pl. 4, figs. 1–10 (in Homo; Africa); 1905, 1–9, pl. 4, figs. 1–10; 1905, April, 205; 1905, 8 April, 950; 1905, 2 Nov., 1298; 1905, 9 pp., 10 figs., 190.
- CLINOSTOMIDÆ Luehe, 1901, 488.—Looss, 1902m, 839 (includes Clinost., Harmost., Ityogonimus, Urotocus, Hapalotrema.—Odhn., 1902, 42.
- CLINOSTOMINÆ Pratt, 1902a, 889, 907 (key), includes Clinost., Nephrocephalus; additional genera Sperost., Microlistrum, Mesotretes, Hapalometra, and Distoma oricola).—Odhn., 1902, 42.
- CLINOSTOMUM Leidy, 1856, 45 (tld. gracile) [not Clinostomus Giard, 1856, 211, fish].—Braun, 1893a, 886, 887, 894; 1899, 1, 3; 1899, 465; 1899g, 484-488, 489-493; 1899h, 1-3; 1900h, 1-48, pls. 1-2, figs. 1-20; 1900c, 24-32; 1900, 27, 31; 1901b, 9, 34; 1901, 561; 1904, v. 2 (3), 24-25.—Darr, 1902, 655.—Looss, 1899b, 649-651 (see æquale, heteroclitum, commutatum); 1902m, 839.—Luehe, 1899, 538; 1900, 557; 1901, 488.—MacCallum, 1899, 707.—Mont., 1893, 156.—Odhn., 1902, 29, 42.—Stiles & Hass., 1898a, 86 (gracile, designated type), 96.—Odhn., 1902, 29.—Pratt, 1902a, 889, 907 (key).—Stiles, 1901, 184.
 - africanum Stess., 1906, in Galli-Valerio, 1906, 49–50, fig. 2, in int. of fish; French Kongo.
 - complanatum (Rud., 1814) Braun, 1899g, 490, 491 (in Ardea cinerea; Berlin); 1900, 27–28, 31; 1900h, 23–25, 26, 41, 42, pl. 1, figs 6, 7.—Stoss., 1901, 97 (9) (in Ardea cinerea; Albona).
 - detruncatum Braun, 1899g, 490 (Dist. dimorphum Dies., p. p.) (in Mycteria americana, Ciconia americana; Brazil); 1900, 29–30, 31; 1900h, 28, 32–34, 35, 37, 39, 40, pl. 2, fig. 11.
 - dimorphum (Dies., 1850) Braun, 1899g, 490 (in Ardea cocoi; Brazil); 1900h, 12, 36–38, pl. 1, figs. 3, 9, pl. 2, figs. 12, 18; 1900, 29, 31.—Locss, 1899b, 650.
 - dubium Leidy, 1856, 45 (in Rusticola minor); 1904a, 88.—Braun, 1899g, 484;
 1900h, 2.—Cobbold, 1859, 10.—Dies., 1858e, 336.—Stoss., 1892, 181.—Stiles & Hass., 1898a, 86.
 - foliiforme Braun, 1899g, 490 (in Ardea purpurea; Italy); 1899h, 2; 1900h, 20, 30–32, 42, pl. 1, fig. 5.
 - gracile Leidy, 1856, 45 (in Pomotis vulgaris, encysted; Esox, intest.); 1904a, 88.—Brand., 1898a, 203 (11).—Braun, 1892a, 651; 1899g, 484, 486, 491 (doubts exist as to identity of gracile and heterostomum); 1900h, 2, 4, 5, 6, 13, 42; 1900, 26, 31.—Dies., 1858e, 336 to Dist.—Looss, 1894a, 137, 138, 171; 1899b, 585, 649, 650, 651 (gracile not identical with heterostomum).—MacCallum, 1899a, 704, 705, 707, 708.—Mont., 1888a, 92; 1893, 155.—Staff., 1904, May 3, 493 (gills of Perca flavescens Mit.; Canada).—Stiles, 1901, 176.—Stiles & Hass., 1894, 414; 1898a, 86, 96 (type of C. and syn. of Dist. heterostomum Rud.).

^a Looss considers that gracile is not the young form of heterostomum, but he accepts the latter as type; this is doubtless a lapsus.

CLINOSTOMUM—Continued.

heluans Braun, 1899g, 490 (in Ardea cœrulea and Nycticorax gardeni; Rio de Janeiro); 1900, 30, 31; 1900h, 8, 11, 39–41, pl. 2, fig. 10.

heterostomum (Rud., 1809) Braun, 1899g, 487, 489–490 [t. h. Ardea purpurea; t. l. apparently Europe]; 1900c, 30–31; 1900h, 11, 19–22 (includes Dist. het. Rud., 1809a, 381; 1819a, 102, 388; Duj., 1845a, 400; Dies., 1850a, 353; Stoss., 1892, 64; Par., 1896, no. 258; D. hians p. p. Par., 1887, 331 from Nycticorax griseus), figs. 1–2, (29), 42, (44, 45); 1902b, 124.—Looss, 1899, 650.

heterostomum MacCallum, 1899a, 697-710, figs. 1-7 (in Ardea herodias; Dunville, Ontario, Canada).—Braun, 1900e, 140–141 (identical with Dist. marginatum Rud. and? D. complanatum Rud. and? D. heterostomum Wright); 1900h, 29, 45 (syn. of Clinost. marginatum); 1904, 23.

lambitans Braun, 1899g, 490 ("in Schlunde eines Reihers; Semanabay, West-indien"); 1900c, 30, 31; 1900h, 41-42, pl. 2, figs. 16, 17.

marginatum (Rud., 1819) Braun, 1899g, 490 (in Ardea sp., Brazil; Ardea cocoi and Mycteria americana in Brazil), 491; 1900c, 28, 31; 1900h, 25–30, 32, 37, 41, 45, 46, pl. 1, figs. 4, 8, pl. 2, figs. 19, 20.

reticulatum (Looss, 1885) Braun, 1899g, 491.—Looss, 1899, 651.

sorbens Braun, 1899g, 490 (in Tantalus loculator; Brazil); 1900c, 30, 31; 1900h, 34, 37, 45, pl. 2, figs. 13–15; 1902b, 129.

CLONORCHIS Looss, 1907, Feb. 1, 147–152 (tod. sinensis).

endemicus (Baelz, 1883) Looss, 1907, Feb. 1, 150-152.

sinensis (Cobbold, 1875) Looss, 1907, Feb. 1, 148-150 (syn. Dist. innocuum Baelz, 1883).

CODONOCEPHALUS Dies., 1850a, 287, 317–318 (m. mutabilis); 1858e, 312, 323, 393.—Brand., 1888a, 12, 13, 51; 1890a, 578.—Braun, 1894l, 166; 1895b, 132.—Goldb., 1855a, 17.—Mont., 1888, 84, 92.—Moul., 1856a, 16.—Villot, 1898, 542.

mutabilis Dies., 1850a, 317–318, Amphist urnigerum Rud., 1819, renamed (in Pelophylax esculentus); 1858e, 323 (in Pel. esc.).—Baird, 1853a, 49 (—Holst. urn. Duj.).—Brand., 1888a, 13.—Braun, 1892a, 796; 1893a, 870; 1894, 167.—Gastaldi, 1854a, 14 pp., 2 pls.—Giebel, 1857, 266 (Amphist. urn. Rud).—Hannover, 1864a, 3, 4.—Mol., 1858, 128; 1859, 818–819, pl. 1, fig. 2 (in Pel. esc.; Padua); 1861, 197.—Mueh., 1898, 16.—Sons., 1893, 188, 190 (syns. Amphist. urn. Rud.; Holost. urn. Duj.) (in Rana esculenta L.).

CŒNOGONÍMINÆ Looss, 1899b, Dec., 573, 583–584, 586, 619; 1902, 839.—Jægers., 1903a, 14, 15.—Luehe, 1901, 488.—Odhn., 1900, 13, 21; 1905, 314.

CŒNOGONIMUS Looss, 1899b, Dec., 585, 586, 619 (tod. heterophyes); κοινός, gemeinsam; γονιμος, Erzeugende; 1900d, 608; 1902m, 515, 833, 835.—Braun, 1901b, 56; 1901e, 334 (syn. of Cotylogonimus Luehe); 1901i, 56.—Jægers., 1900c, 736; 1903a, 10, 11, 13, 15.—Luehe, 1900aa, 557.—Odhn., 1900, 21, 22.—Ofenheim, 1900, 183.—Stiles, 1904i, 43.—Stiles & Hass., 1900a, 563.

fraternum (Looss, 1894) Jægers., 1903a, 3, for fraternus.

fraternus (Looss, 1894) Looss, 1899b, 585, 700-701; 1901b, 205.

heterophyes (Sieb., 1853) Looss, 1899b, 585, 678, 699-700, 701.—Braun, 1903, 164 (to Cotylogonimus).—Jægers., 1903a, 3.—Ward, 1903, 870.

COLEPS Nitzsch, 1827, 69 (m. Cerc. hirta).

CONCHOSOMA Stoss., 1898, 20, for Conchosomum, q. v.

CONCHOSOMINÆ Rail., 1896, 160.

CONCHOSOMUM Rail., 1896, Mar. 15, 160, Hemist. Dies., 1850 [not Swainson] renamed, tod. alatum.—Stoss., 1898, 20.—See also Alaria 1788.

1898: Conchosoma Stoss., 1898, 20.

1901: Chonchosomum Calli Valoria 1001.

1901: Chonchosomum Galli-Valerio, 1901c, 364.

 $alatum~({\rm Geeze,\,1782})$ Rail., 1896, 160 (in intest. of Canidæ).—Piana, 1898, 3. $spathaceum~({\rm Rud.,\,1819})$ Stoss., 1898, 20–21 (in Larus canus, L. argentatus michahelesii; Triest).

spatula (Dies., 1836) Stoss., 1898, 20 (in Buteo vulgaris at Triest, Iamiano; Falco peregrinus at Ospo; Circus aeruginosus at Verteneglio, Monfalcone, Doberdò, Iamiano).—See spathula.

COSMOCOTYLEA Dies., 1858e, 313, 360, 365 (subf. of Monocotylea, contains Gasterost., Rhipidocotyle, Callicotyle).—Tasch., 1878, 565, 566.

COTYLAGASTER Mont., 1904, 65, misprint for Cotylogaster.

COTYLASPIS Leidy, 1857a, 18 (m. insignis); 1858a, 110; 1904a, 110.—Braun, 1893a, 886, 887, 888, 891, 894, 896.—Dies., 1859c, 438-439.—Hoyle, 1890, 539 (one species; in Anodonta).—Jackson, 1888, 650 (endoparasitic, American Anodonta), 654.—Kofoid, 1899, 180, 181, 182, 183, 184.—Mont., 1888, 89, 91, 1892, Oct. 7, 196, 197, 198, 199, (syn. of Aspidogaster) 202.—Nickerson, 1902; 599, 603, 604, 605, 612, 613, 615, 617.—Pratt, 1902a, 887, 891 (key).—Shipley & Hornell, 1904, 98.—Tasch., 1879, 259.

insignis Leidy, 1857a, 18 (in Anodonta fluviatilis, A. lacustris); 1858a, 110.—Dies., gnis Leidy, 1857a, 18 (Ili Andodolta Ildvathis, A. factistris); 1858a, 110.—Dies., 1859c, 439.—Kofoid, 1899a, 179–186 (identity with Platyaspis anodontæ); 1899b, 179–186.—Mont., 1892, Oct. 7, 197, 201 (?syn. of Aspidogaster conchicola), 202.—Nickerson, 1902, 612, 613, 614, 619 (in Unionidæ; North America).—Osborn, 1905a, 201–242 (habits and structure), figs. 1 (a-f), 1–81; 1905, 201–242, 3 pls., 1 fig.; 1905, 317–318.—Tasch., 1879, 259 (in Anod. fluv., A. lac.; North America).—Also reported for Anodonta corpulenta, Unio alatus, U. anadontoides, U. confragosus, U. edentulus, U. elegans, U. gracilis, U. katharinæ, U. licementians, U. reported for Anodonta corpusations, U. katharinæ, U. ligamentinus, U. rectus, U. tuberculatus).

lenoiri (Poir., 1886) Nickerson, 1902, 613, 614, 619 (in Tetrathyra; Senegal).

COTYLEA Dies., 1850a, 286, 293 [not of Mont., 1891, 111], subtribe of Dicranocola to contain Rhopalocerca, Bucephalus, Malleolus, Cerc., Histrionella, Diplocotyle.—Goldb., 1855a, 15.

COTYLEGASTER Sieb., 1877 MS. in Cobbold, 1877e, 235, 237 (lapsus for Aspido-

cotylus) [not Cotylogaster Mont., 1892].

cochleariforme (Dies., 1838) Sieb., 1877 MS. in Cobbold, 1877e, 235, 237 (lapsus), see Gastrodiscus sonsinoii; 1879b, 360 (syn. Aspidogaster cochleariformis) "not to be confused with Gastrodiscus."—Ward, 1895, 338 ("error—Gastrodiscus ægyptiacus," in Equus caballus).

cochleariformis (Dies., 1838) Zuern, 1882, 222 (see cochleariforme).—Lejtenyi, 1881a, 1.—Sons., 1895, 179, 180; 1896, 297.

COTYLOCEPHALA Dies., 1858e, 315 (f. of Trematoda plectanophora to contain subf. Aplacoplectana, Placoplectana), 382.—Ben. & Hesse, 1864, 95.—Mont., 1888a, 84.

COTYLOGASTER Mont., 1892a, Oct. 7, 168, 169, 170, 173, 174, 175, 176, 177, 179, 180, 183, 184, 185, 186, 187 (Aspylogaster), 188, 189, 190, 191, 194, 195, 196, 197, 198, 206^a (m. michælis), 213 (g. of Aspidobothridæ); 1893, 37, 111, 114, 202, 205, 206, 209.—Braun, 1893a, 888, 890, 891, 894, 896, 897, 917, 918; 1893b, 177, 178, 179, 182, 188.—Gamb., 1896, 73.—Kofoid, 1899, 182.—Lejtenyi, 1881a, 2.—Looss, 1894a, 11, 134; 1902, 428, 429.—Nickerson, 1902, 602, 604, 606, 610, 611, 612, 617, 1902, 1803, 100 606, 610, 611, 612, 617.—Pratt, 1902a, 887, 891 (key).—Stoss., 1898, 19.

1892: Aspylogaster Mont., 1892, Oct. 7, 187, for Cotylogaster.

michælis Mont., 1892, Oct. 7, 168-173, 198, 206-207, pls. 21-22 (in Cantharus vulgaris; Triest); 1893, 52, 82, 145, 202; 1902, 139.—Braun. 1893a, 887, 898; 1893b, 184.—Jægers., 1899, 203, 207.—Looss, 1902, 420, 421, 428.—Nickerson, 1902, 597, 598, 599, 600, 601, 602, 603, 605, 606, 610, 611, 618 (in Can. vulg.; Triest).—Stoss., 1898, 19; 1899, 3.

occidentalis Nickerson, 1900, Feb. 16, 250 (in Sheepshead; Mississippi Valley); 1901, Mar. 8, 378; 1902, 597–624, pl. 32, figs. 1–7, pl. 33, figs. 8–21 (in Aplo-

dinotus grunniens; Minnesota).

COTYLOGONIMINÆ Pratt, 1902a, 888, 897 (key), includes Cotylogonimus and Cryptocotyle.

COTYLOGONIMUS Luehe, 1899k, Dec., 538, 539 (tod. heterophyes); 1900, 555, 557.—Braun, 1900h, 6; 1901e, 334, 338; 1901i, 56; 1903, 164; 1906, 171-172.— Pratt, 1902a, 888, 898 (key).—Looss, 1900, 607; 1902, 813, 833.—Stiles, 1904i, 43.—See Heterophyes.

[concavum (Crep., 1825), 1899 to (Cryptocotyle).]

fraternus (Looss, 1894) Braun, 1901e, 337.—Fischder., 1903h, 548.

heterophyes (Sieb., 1853) [Luehe, 1899k, 539].—Braun, 1901e, 335, 337, 338; 1903, 3 ed., 164–166, fig. 114; 1906, 136, fig. 71, 172–173, fig. 105.—Fischder., 1903h, 548.—Ward, 1903, 870 (to Heterophyes).—Type of Heterophyes, q. v.

persicus Braun, 1901e, 334-338, pl. 20. fig. 13 (in Persian wolf).

COTYLOPHORA Mont., 1888a, 84, 93, for Trematoda cotylophora Dies., 1858e, 312, a subtribe containing fams. Monocotylea, Tricotylea, Polycotylea [not Cotylophori Mayer, 1841a, 24, 25].

COTYLOTRETUS Odhn., 1902, 32 (m. rugosus).—Pratt, 1902a, 888, 897 (key). rugosus Odhn., 1902, 32–45, fig. 3 (in Coluber pullatus = Spilotes pullatus).

CRANIOCEPHALA Mont., 1905, 21-24 (m. birói), a temnocephala; κράνειον, cornelian-tree.

birói Mont., 1905, 21–24 (named after L. Biró), figs. 1–2 (in Sesarma gracilipes A. Edw.; Sattelberg, New Guinea).

CRASPEDELLA Haswell, 1893e, 96, 98, 99, 112, 114, 132, 141, 143, 145 (m. spenceri); 1893f, 154.—Mont., 1899, 84, 115, 116, 117, 120, 121.—Pratt, 1900a, 646, 647 (key).—St.-Remy, 1898, 522, 529.

spenceri Haswell, 1893e, 97, 98, 113, 142, pl. 13, fig. 21, pl. 15, fig. 3 (in Astacopsis bicarinatus); 1900, 433.—Mont., 1899, 84, 121 type (in Ast. bic.; Austr.).—
Pratt, 1900a, 655, 657, fig. 2.—St.-Remy, 1898, 529-530, fig. 1 (in Ast. bic.).

CREADIINÆ Looss, 1899b, Dec., 571.—Luehe, 1900, 487.—Odhn., 1901, 483.

CREADIUM Looss, 1899b, Dec., 570–571, 574, 580, 595 [not Creadium, Creadion Vieill., 1816, bird] tod. isoporum; 1900d, 3 Dec., 602 (renamed Allocreadium).—Braun, 1900, 232; 1901b, 33.—Luche, 1900, 487, 489; 1901, 397.—Odhn., 1901, 483.—Stiles, 1901, 189.—[See also Dist. commune, D. scorpænæ, D. fasciatum, D. sophiæ, D. bacillare, D. pedicellatum, D. album, D. mormyri, D. obovatum.—Looss, 1899b, 571.

angusticolle (Hausmann, 1896) Looss, 1899b, 571 [possibly identical with D. com-

mune Olss., 1868, 31].

isoporum (Looss, 1894) Looss, 1899b, 570, 571, 595.

pegorchis Stoss., 1901, descr. of pl. 6, fig. 4, see Allocreadium.

CREPIDASTOMUM Pratt, 1902a, 888, 896, misprint for Crepidostomum.

CREPIDOSTOMUM Braun, 1900b, 232 (tod. metœcus); 1901b, 31, 33.—Heymann, 1905, 83, 89 (Fasciolidæ, Bunoderinæ).—Looss, 1902, 452, 453, 454.—Odhn., 1905, 296.—Pratt, 1902a, 888 (Crepidast., Fasciolidæ, Psilostominæ), 896 (key).—Staff., 1904, 491.

1902: Crepidastomum Pratt, 1902a, 888, 896, misprint.

cornutum (Osborn, 1903) Staff., 1904, May 8, 490 (int. of Ambloplites rupestris Raf.; Canada), 491.

laureatum (Zed., 1800) Braun, 1900b, 232 (in fish).—Heymann, 1905, 87.—Looss, 1901, 564; 1902, 452, 453.—Odhn., 1905, 296.—Staff., 1904, May 3, 490 (int. Salvelinus fontinalis Mit.; Canada), 1. var. in Perca flavescens and Stizostedion vitreum, 2. var. in Necturus maculatus; 1905, Apr. 11, 682 (int. Necturus maculatus; Canada).

metacus (Braun, 1900) Braun, 1900b, 230–232, pl. 10, fig. 13.—Heymann, 1905, 87.—Looss, 1901, 564; 1902, 451 (in Vespertilio lasiopterus), 452, 453.—Odhn.,

1905, 296.

CRICOCEPHALUS Looss, 1899b, Dec., 551, 666-667 (tod. delitescens), δ κρίκος, ring; 1901b, 209; 1901l, 620; 1902m, 531, 532, 534, 536, 537, 540, 542, 547, 562, 570, 576, 579, 580, 581, 582, 583, 585, 586, 587, 589, 591, 593, 599, 600, 601, 603, 604, 605, 609, 612, 614α (delitescens=albus).—Braun, 1901a, 50.—Pratt, 1902a, 890, 910 (key).

albus (Kuhl & Hasselt, 1822) Looss, 1899b, 667; 1901l, 621; 1902m, 511, 532 (syndelitescens), pl. 25, fig. 64; pl. 32, figs. 170–174; 533, 535, 536, 537, 539, 540,

541, 600, 605, 614.—Braun, 1901a, 45, figs. 10, 13, 15, 16.

delitescens Looss, 1899b, 666, 667, 759-762 (in Chelonia mydas; apparently Egypt), 773, figs. 76-80; 1902m, 531, 532 (syn. of Cr. albus), 549, 605.—Braun, 1901b, 38, 44, 45, 48.

megastomus Looss, 1902, 533–536, 540, 541, 605, pl. 25, figs. 57–60, pl. 27, fig. 90 (in Chelone mydas; Egyptian coast).

resectus Looss, 1902m, 537–541, 813, pl. 25, figs. 61–63, 813 (in Chelone mydas; Egyptian coast).

ruber (Kuhl & Hasselt, 1822) Looss, 1899, 762.

CROBYLOPHORUS Kroyer, 1852–53a, 813 (m. chimæræ).—Dies., 1859c, 447 (syn. of Gyrocotyle).

chimæræ Kroyer, 1852–1853a, 813, 1226–1227 (in Chimæra monstrosa L.).—Dies., 1859c, 447 (syn. of Gyrocotyle amphiptyches).

CROSSEDERA Mont., 1888, 92 (for Crossodera).

(CROSSODERA) Duj., 1845a, 382, 389, 434–437 [not Gould, 1837, aves], subg. (tld. nodulosa) of Dist.; raised to generic rank by Cobbold, 1860a, 31. See Crossodera.—Mont., 1888a, 12.—Stiles & Hass., 1898a, 84 (syn. of Bunodera).

auriculatum (Wedl, 1857) Stoss., 1886, 63.

campanula Duj., 1845a, 435, as doubtful (in Esox lucius; Rennes, France).

crucibulum (Rud., 1819) Duj., 1845a, 363, 435.

excisum (Rud., 1819) Duj., 1845a, 436-437.

laureatum (Zed., 1800) Duj., 1845a, 435-436. See Crossodera.

nodulosum (Freelich, 1791) Duj., 1845a, 434–435.—Stiles & Hass., 1898a, 85 (designated type of subg). See Crossodera.

papilliferum (Mol., 1861) Stoss., 1886, 60.

verrucosum (Busch, 1851) Stoss., 1886, 61.

CROSSODERA (Duj., 1845a, tld. nodulosa [nec Crossodera Gould, 1837, aves]) Cobbold, 1860a, 31–32.—R. Bl., 1891, 609.—Braun, 1893a, 885, 890, 909, 911; 1900b, 232; 1900h, 3.—Fil., 1857c, 14.—Hass., 1896a, 7 (syn. of Bunodera Rail.).—Looss, 1899b, 535, 538, 542, 580, 594, 598.—Mont., 1888, 92 (Crossedera), 105; 1892, Oct. 7, 214 (g. of Distominæ); 1893, 150, 152, 153, 155.—Rail., 1896, 160 (renamed Bunodera).—Stiles, 1901, 197.—Stiles & Hass., 1898a, 84, 85 (nodulosa type by designation), 87.—Stoss., 1892, 4, 5; 1899, 11.—Wagener, 1860, 165.

1888: Crossedera Mont., 1888, 92, misprint.

campanula (Duj., 1845) Cobbold, 1860a, 32 (in Esox lucius).

laureata (Zed., 1800) Cobbold, 1860a, 32 (in Salmo trutta, S. fario, S. umbla, Thymallus vexillifer).

linearis (Rud., 1793) Cobbold, 1860a, 32 (in Phasianus gallus).—Hass., 1896a, 2 (in Gallus dom.).—Rail., 1893a, 368.—Stoss., 1892, 146.

nodulosa (Frœlich, 1791) Cobbold, 1860a, 32 (in Perca fluviatilis, Acerina vulgaris, Aspro vulgaris, A. zingel, Lucioperca sandra, Esox lucius, Barbus communis).

papillosa (Dies., 1850) Cobbold, 1860a 32 (in Beroë rufescens) (syn. Dist. beroës Will).

CRUMENA Nitzsch, 1827, 68-69 (m. Cerc. crumena).

(CRYPTOCOTYLE) Luehe, 1899, 539 (tod. concavum), subg. of Cotylogonimus; 1900, 557.—Braun, 1900h, 6; 1901b, 56; 1901i, 56.—Jægers., 1901, 981.—Looss, 1900, 607 (type concavum); 1902, 813, 833.—Pratt, 1902a, 888, 897 (key).

concavum (Crep., 1825) Fischder, 1903h, 548.

lingua (Crep., 1825) Fischder, 1903h, 548.

CRYPTOGONIMUS Osborn, 1903, 315–318 (m. chili); 1903, 533–534.—Marshall & Gilbert, 1905, 479.

chili Osborn, 1903, 315-318, fig. 2 (in Micropterus dolomieu; Chautauqua); 1903, 533-534; 1904, 308.—Marshall & Gilbert, 1905, 478.—Staff., 1905, 682 (in Ambloplites rupestris Raf.).

CRYPTOSTOMUM Nitzsch, 1819, sub Amphist.—See Nord., 1832a, 30, 50 (syn. Holost. cuticola); 1840, 561, 628.—Moul., 1856a, 15 (syn. of Hemist.).—[Not Cryptostoma Blainv., 1818, mollusk: Cryptostoma Latr., 1825, coleopteron; Cryptostomus Blainv., 1818, mollusk.]

CUCULANUS Mueller, 1777.—See Nematoda.

conoideus Bloch, 1782a, 35, pl. 10, figs. 5-7 (Cuculanus) (t. h. Anas boschas dom.; Europe).—Dies., 1850a, 383 (syn. of Dist. echinatum).—Herbst, 1787a, 42.—Looss, 1899b, 680.—Mueller, 1788b, 18.—Rail., 1886, 297 (to Dist.).—Rud., 1809a, 418, 420, 432.

CYATHOCOTYLE Much., 1896, v. 20, 590 (m. prussica) (Holostomidæ); 1897, v. 1, in 243–279, pl. 4; 1897, in 127–128; 1897, in 478–479.—Odhn., 1902, 19.—Pratt, 1902a, 889, 908 (key).

fraterna Odhn., 1902, 19-21 (in Harelda glacialis).

prussica Mueh., 1896, 590; 1896, 270-278, figs. 7, 15-17 (in Anas glacialis); 1898, 4, 16, 80.—Braun, 1901, 259; 1902b, 154.—Odhn., 1902, 19, 20.

CYATHOCOTYLINÆ Mueh., 1898b, 20, subf. of Holostomidæ, contains Cyathocotyle.—Pratt, 1902a, 889 (includes Cyathocotyle).

CYCLATELLA Ben. & Hesse, 1863; 1864a, 65, 66, 81–83 (m. annelidicola) (Tristomidæ).—Ben., 1869a, 640, is a bryozoon, see Loxosoma.—Braun, 1890a, 518 (=Loxosoma).—Mont., 1888a, 86.—Tasch., 1879, 56.

anellidicola Mont., 1888a, 88, for annelidicola.

annelidicola Ben. & Hesse, 1863; 1864a, 82–83, pl. 7, figs. 12–13 (on ?Clymene).— Mont., 1888a, 88 (anellidicola).—Prouho, 1891, 91–116 (to Loxosoma).— Sons., 1895, 119.

CYCLIDIUM Mueller, 1786.—Nitzsch, 1827, 69, contains Cerc. cyclidium [type by absolute tautonymy], C. discus.

CYCLOBOTHRIUM Cerf., 1895m, 141, 142, 144–145 (m. sessilis); 1896d, 548, 550–551.— Mont., 1903, 336 (subf. Diclidophorinæ).—St.-Remy, 1898, 552.—Zool. Anz., 1895, 327.

sessilis (Goto, 1894) Cerf., 1895m, 141, 142, 145 (in Chœrops japonicus; Japan); 1896, 548, 551.—St.-Remy, 1898, 554 (=Diclidophora sessilis).

sessile (Goto, 1894), Zool. Anz., 1895, 327.

CYCLOCIRRA Mueller, 1841, 147 (apparently m. thompsonii).

thompsonii Mueller, 1841, 147, based on Thompson, epizoon on Comatula.

CYCLOCŒLEUM Fuhrmann, 1904, 61, misprint for Cyclocœlum.

CYCLOCŒLINA Cohn, 1904, 232, misprint for Cyclocælinæ.

CYCLOCELINÆ Cohn, 1904, 231, 232 (contains Typhlæcælum=Typhlocælum).— Stoss., 1902, 2, 3, 4, 7, 8, 9, 11, 12, 31, 34, 35.

CYCLOCŒLIUM Fuhrmann, 1904, 59, for Cyclocælum.

mutabile (Zed., 1800) Fuhrmann, 1904, 59, 61, to (Cyclocælium).

CYCLOCŒLUM Brand., 1892b, 507 (for Monost. mutabile, M. flavum, M. arcuatum, M. tringæ, M. ellipticum) [tld. mutabile].—Braun, 1901e, 346.—Cohn, 1904, 229, 231, 232 (in syn. of Monost. flavum).—Looss, 1899b, 659–661, 662; 1901b, 192 (type, Monost. mutabile); 1902m, 701, 702, 719, 720, 721, 729, 730, 780.—Luehe, 1901, 174 (type, Monost. mutabile); 1901, 481.—Pratt, 1902, 890, 909 (key).—Stoss., 1902, 5, 6, 7, 8, 12.

1902: Cgclocœlum Stoss., 1902, 40, misprint.

1904: Cyclocœleum Fuhrmann, 1904, 61, misprint.

1904: Cyclocælium Fuhrmann, 1904, 59, misprint.

adolphi Stoss., 1902, 19–20, pl. 4, figs. 13, 14 (in Grus cinerea, Ardea cinerea; Siberia).

arcuatum (Brand., 1892) [Brand., 1892b, 507].—Stoss., 1902, 20-22. pl. 4, figs. 15, 16 (syn. Monost. mutabile Sieb.) (Anser dom., Bucephala clangula; Berlin).

brasilianum Stoss., 1902, 16–17, pl. 2, figs. 7, 8 (in Scolopax flaviceps; Brazil).

exile Stoss., 1902, 17-18, pl. 3, figs. 9, 10 (in Totanus ochropus).

mutabile (Zed., 1800) [Brand., 1892b, 507].—Stoss., 1902, 12, 13–14, pl. 1. figs. 1, 2
 (syns. Monost. mutabile Zed., 1800; M. microstomum Crep., Cephalogonimus ovatus Stoss.) (type of genus) (in Gallinula chloropus, Fulica atra).—[Braun, 1901e, 346].—Cohn, 1904, 231, 232.—[Luehe, 1901 p, 174.]

ovopunctatum Stoss., 1902, 15-16, pl. 2, figs. 5, 6 (syn. Monost. mutabile Ben.) (in

Numenius arquata).

problematicum Stoss., 1902, 14–15, pl. 1, figs. 3, 4. pl. 7, fig. 25, pl. 8, fig. 29 (syn. Cyclocelum sp. Looss, 1899b, 660, in Totanus calidris; Egypt) (in Totanus calidris, T. glottis).

robustum Stoss., 1902, 18, pl. 3, figs. 11, 12 (syn. Monost. fiavum Par., v. 11 (258) (in Fuligula cristata).

CYCLOCOTYLA Otto, 1823, 300–302 (m. bellones).—Baer, 1826a, 126.—Ben. & Hesse, 1864, 47.—Blainv., 1828a, 570.—Braun, 1889a, 326.—Cobbold, 1877, 238.—Dies., 1840, 234; 1850a, 419 (syn. of Cyclocotyle).—Zæringer, 1829.

1823: Octostoma Otto, 1823, 302 (preoccupied), same as Cyclocotyla.

1823: Cyclostoma Otto, 1823, 302 (preoccupied), same as Cyclocotyla.

bellones Otto, 1823, 300, pl. 41, fig. 2, a-c (on Hornhecht=Belone acus; Naples).—
Baer, —, 687, pl. 32, fig. 9.—Blainv., 1828a, 570.—Bl., 1847, 336.—Braun,
1889a, 326.—Dies., 1850a, 419, to Cyclocotyle.—Nord., 1832a, 78, 80; 1840,
600 (to Octobothrium).

- CYCLOCOTYLA—Continued.
 - belones Crep., 1839, 291, for bellones.—Kroyer, 1852-53a, 1221 (in Belone rostrata Fab.).
 - lanceolata Zæringer, (1829), 21.—Braun, 1890a, 553 (on Salmo fario) (to Octobothrium).—Crep., 1839, 291.
- CYCLOCOTYLE for Cyclocotyla.—Burm., 1837, 530.—Braun, 1890a, 515-518 (belongs to Octobothrium Leuck.).—Crep., 1838, 84.—Dies., 1850a. 289. 419 (of Otto, syn. of Octobothrium Nord.), 423 (of Schultze, syn. of Discocotyle); 1858e, 314, 372 (of Otto).—Gold., 1855a, 19.—Mont., 1888a, 83.—Tasch., 1879, 233, 239.—See Cyclocotyla.
 - bellones (Otto, 1823) Dies., 1850a, 419 (on Belone acus) (syns. Cyclocotyla bellones, Octobothrium bellones); 1858e, 372.—Tasch., 1879, 245 (to Octobothrium).
 - lanceolata Schulze in Zæringer (1829).—Crep., 1839a, 291, note 25.—Dies., 1850a, 424 (syn. of Discocotyle sagittata).—Tasch., 1879, 244 (syn. of Octobothrium sagittatum Leuck.).
- CYCLOGENA Hemp. & Ehrenb., 1828a (m. Cerc. lupus; Berlin), fam. Hydatinorum.
- CYCLOSTOMA Otto, 1823, 302 (as syn. of Cyclocotyla) [not Cyclostoma Lam., 1801, mollusk; Cyclostoma Nilss., 1832, fish; Cyclostoma for Ciclostoma Scacchi, 1836, mollusk].
- CYMATOCARPUS Looss, 1899b, Dec., 550, 551, 593-594 (tod. undulatus), 607, 611, 633; $\tau \acute{o} \kappa \tilde{v} \mu \alpha =$ wave, $\acute{o} \kappa \alpha \rho \pi \acute{o}_5 =$ fruit.—Braun, 1901b, 22.—Odhn., 1902, 38, 41, 42.—Pratt, 1902a, 889, 903 (key).—Staff., 1903, 828.
 - solearis (Braun, 1899) Braun, 1901b, 22, 23, fig. 8, or Looss. 1901l, Oct. 30, 563; 1902m, 508.
 - undulatus Looss, 1899b, 594, 711-712, figs. 32-34 (in Thalassochelys undulatus; Abukir, July); 1901l, 563; 1902m, 414, 508.—Braun, 1901a, 22, 23.—Heymann, 1905, 84.—Luehe, 1900, 561.
- CYSTAGORA Staff., 1905, Apr. 11, 683 (m. tetracystis); κύστις, bladder; άγορά, assembly.
 - tetracystis (Gastaldi, 1854) Staff., 1905. 683 (in Rana catesbiana. R. virescens; Canada).
- DACTYCOTYLE Ben. & Hesse, 1863; 1864a, 96, 110 (pollachii [type by present designation], luscæ); δάκτυλος, finger.—E. Ben., 1868a, 4–7; 1868, 22–36, 1 pl.—Braun, 1890a, 446.—Mont., 1888a, 8, 11, 16, 52, 59, 66, 86, 89, 99; 1903, Dec., 336 (syn. Pterocotyle) (in subf. Octocotylinæ; f. Octocotylidæ).—Par., 1894, 135 (syn. of Dactylocotyle Par. & Perugia).—Par. & Perugia, 1890, 4, 7; 1890, 10.—Tasch., 1879, 59, 73; 1879, 240.

- 1889: Dactyocotyle Braun, 1889a, 364, misprint.
- lusca Ben., 1868a, 7 (in Morrhua lusca) (for luscæ).
- luscæ Ben. & Hesse, 1864, 111-112 (in Morrhua lusca).—Ben., 1868, 36.—Braun, 1889a, 364 (on M. l.); 1890a, 535, to Octobothrium (Dactylocotyle).—Tasch... 1879, 247, to Octobothrium.
- phycidis (Par. & Perugia, 1889), Par. & Perugia, 1890, 4; 1890, 7.—Braun, 1890a, 535, to Octobothrium (Dactylocotyle).
- pollachi Mont., 1888, 66 (for pollachii).
- pollachii Ben. & Hesse, 1863; 1864, 110-111, pl. 11, figs. 23-30 (in Merlangus pollachius).—Ben., 1868a. 7; 1868, 36, 1 pl.—Braun, 1889a, 364 (on Mer. pol.); 1890a. 535, to Octobothrium (Dactylocotyle).—Mont., 1888, 66 (pollachi).—Par. & Perugia, 1890, 4.—Scott, 1901, —, pl. 8; 1905, 117.—Tasch., 1879, 242, 246 (to Octobothrium).—Can best be taken as type of genus.
- DACTYLOCEPHALA Mont., 1899, 115, 116, 117, 120, 121 (type Temnocephala madagascariensis).—Pratt, 1900, 646, 647 (Temnocephalidæ, Temnocepha linæ).
 - madagascariensis (Vayssière, 1892) Mont., 1899, 121 (in Astacoides madagascariensis; Madagascar).

DACTYLOCOTYLE Marschall, 1873, 430, 795, and 796, apparently for Dactycotyle.

Ben. & Hesse, 1863.—Braun, 1890a, 469, 471, 477, 484, 490, 498, 516, 517, 522, 535 (syn. or subg. of Octobothrium), 546.—Cerf., 1895h, 913-946, pls. 1-2, figs. 1-20; 1895i, 831-834; 1895k, 634; 1895m, 125, 126, 137, 138, 139; 1896c, 232-233; 1896, 510-535, 536, 545, 546, 547; 1898a, 301, 302, 305-315, 322-323; 1899a, 345, 370, 383, 411.—Lint., 1905, 333, 352, fig. 151 (sp. on Brevoortia tyrannus).—Par., 1894, 135 (syn. Dactycotyle Ben. & Hesse).—Pratt. 1900a, 646, Polystomide. Octoortyling, 652 (on gills of fish, esp. Gadide), 656, 666 646 (Polystomidæ, Octocotylinæ) 652 (on gills of fish, esp. Gadidæ), 656, 660, fig. 35.—St.-Remy, 1898, 523, 545, 548.—Scott, 1901, 150.—Scudder, 1884 (1882), part 2, 91 [gives date as "1883 [1862]" and quotes Marschall, which is 1873].—Stoss., 1898, 12.

carbonarii Cerf., 1895h, 929, 931, pl. 1, figs. 1, 5, 9, 10, 11, pl. 2, figs. 12-20.

denticulatum (Olss., 1876). Ceri., 1895h, 922, 923-938, 941, 943, type of genusa in Gadus carbonarius; 1896, 517, 518-530, 532-533, 534, pl. 23, figs. 1, 5, 9, 10, 11, pl. 24, figs. 12-20; 1898a, 301, 305 a, 307, 312, 313, 317, 318, 323, 325, pl. 12, figs. 6-9 in Gadus carbonarius (Linn.).—Lint., 1901, 408, 414 in Pollachius virens, 474.—Par., 1899, 4.—Pratt, 1900a, 656, fig. 35, 657, 660 (key).—St.-Remy, 1898, 548-549, 550 in Gadus virens, 6, carbonarius.)—Staff, 1904, May 3, 482 (gills of Pollachius vireus [virens] L.; Canada).

*Usex** (Ben. & Hesse, 1863)
 Tasch., 1879, 247 to Octobothrium).—Braun. 1890a, 535 as Octobothrium (Dactylocotyle) lusex, on Morrhua lusea.—Cerf., 1895h, 922; 1896, 517; 1898a, 302, 507, 312, 315-322, 326, pl. 12, figs. 1-5 in Morrhua lusea; 1899a, 372.—Par., 1899, 4 to Octobothrium) (on Gadus minutus; Portoferraio).—St.-Remy, 1898, 551.

merlangi (Kuhn, 1830) Cerf., 1895h, 922, 939, 942–943. pl. 1, figs. 4, 8 (in Gadus merlangus); 1896, 517, 531, 534, pl. 23, figs. 4, 8; 1898a, 301, 304, 310, 312, 324–325, pl. 12, figs. 12, 13 (in Gadus merlangus L.); 1899a, 370.—St.-Remy, 1898, 550, 556.

minor St.-Remy. 1898. 551=Octobothrium minus Olss., 1876.

molvæ Ceri., 1895h, [944], pl. 1, figs. 3, 7; 1896, pl. 23, figs. 3, 7.

morrhuæ Ben. & Hesse, 1863 St.-Remy, 1898, 551 (=Pterocotyle morrhuæ).

palmatum (Leuck., 1830) Cerf., 1895h, 922, 939-940, 943-944 (in Gadus molva); 1896, 517, 531, 534-535; 1898a, 302, 312, 325-326 in G. mol. L.; 1899a, 370, St.-Remy, 1898, 550-551 syn. Octodactylus inhærens Dalyell).—Scott. 1897, Apr., 127 (in the Moray Firth).

phycidis Par. & Perugia, 1889, 743-744, fig. 5 (in Phycis blennoides: Genova); 1890. 743-744. fig. 5.—Braun. 1890a. 535 to (Octobothrium (Dactylocotyle) on Phycis blenuoides [blennoides]).—Cerr., 1895h. 922; 1896. 517; 1898a. 302.—Par., 1894, 135-136 in Phycis blennoides: Genova).—St.-Remy, 1898, 551.—Staff., 1904, May 3, 482 gills of Phycis chuss Walb.; Canada

pollachii Ben. & Hesse. 1863 Braun, 1883a, 50: 1890a, 472, 476, 492, 498, 535, to Octobothrium Dactylocotyle, on Merlangus pollachius.—Cert. 1895h, 914–915, 921, 922, 938, 941–942, pl. 1, figs. 2, 6 in Gadus pollachius): 1896, 211, 516, 517, 530, 533–534, pl. 23, figs. 2, 6; 1898a, 301, 311, 312, 317, 318, 319, 321, 324 in G. p. 1—8t. Remy, 1898, 549–550, 551.—Scott, 1901, 150, pl. 8, figs. 28, 29; 1901. 343 in G. p.; Irish Sea).

squillarum Par. & Perugia. 1889 Stoss., 1898, 12-13 on Bopyrus squillarum; Triest).

taschenbergi Par., 1894, 704 for taschenbergii.

taschenbergii (Par. & Perugia, 1889 Par., 1894, 136, to (Choricotyle) in Sargus rondeletii; Genova ...—Cerf., 1895h. 922. 923: 1896. 517.

DACTYLODA Mont., 1905, 25. Mai, 403 order for Temnocephalidæ).

DACTYLODISCUS Olss., 1893, 7–8 (m. borealis).—Mont., 1903, 336 (syn. of Tetra-onchus); 1905, 79–80.—Pratt, 1900a, 646, 554 (key), 657, fig. 48.—St.-Remy, 1898, 524, 564, 567.

borealis Olss., 1893, 7-8, pl. 1, figs. 7-10 on Thymallus vulgaris, in lacubus Jemtlandiæ Sällsjön, Nälden, Ockesjön et in flumine ad Mörsil: Coregonus lavaretus, in lacu Ockesjön).—Mont., 1905, 79.—St.-Remy, 1898, 568, fig. 6 (in Coregonus lavaretus).—Pratt, 1900a, 657, fig. 48.

a "Dans la description, le Dactylocotyle denticulatum avait été pris comme type, parce que c'est dans cette forme que les organes de fixation : aractéristiques du genre, présentaient la plus grande complication."—Cerf., 1898, p. 705. It is impossible to take this species as type. See International Code, Art. 30.

- DACTYLOGYRUS Dies., 1850a, 290, 433, 650, 651–652 (n.. auriculatus); 1858e, 374, 375; 1859c, 440.—Ben., 1858a, 1861a, 63.—Ben. & Hesse, 1864, 121.—Brand., 1894a, 309.—Braun, 1890a, 412, 413, 417, 426, 434, 438, 444, 445, 452, 458, 465, 468, 478, 479, 486, 497, 499, 503, 512, 515, 517, 523, 542, 543; 1893a, 890.—Carus, 1863, 478.—Cobbold, 1879, 41, 42.—Fraip., 1880c, 418.—Gamb., 1896a, 73.—Goldb., 1855a, 20.—Goto, 1891a, 161, 166, 183, 185; 1893a, 798, 801, fig. 3.—Haswell, 1892b, 150; 1893e, 114.—Hoyle, 1890, 539 (about 20 species, on fishes).—Jackson, 1888, 654.—Kath., 1894a, 128, 133, 144, 155, 156, 157.—Leuck., 1886d, 45, 48.—Maclaren, 1904b, 574, 598, 599, 600.—Mont., 1888a, 10, 14, 40, 84, 86, 101, 107; 1889, 114; 1891, 109, 111; 1892, Oct. 7, 186, 213 (gen. of Gyrodactylinæ); 1903, 336 (of subf. Gyrodactylinæ).—Olss., 1893, 7.—Pratt, 1900a, 646, 654 (key), 657, fig. 44.—St.-Remy, 1898, 524, 566.—Stoss., 1898, 18.—Tasch., 1879, 69; 1879, 260 (syn. of Gryodactylus Nord.), 261, 263 (of Wagener, syn. of Tetraonchus Dies.), 265 (of Wagener, syn. of Calceost. Ben.).—Wagener, 1857, 28, 29, 50, 54–55, 57–76.
 - **aquans* Wagener, 1857, 99, pl. 15, fig. 14 (in Labrax lupus); —, 84.—Ben. & Hesse, 1864, 121, 122 (to Diplectanum).—Dies., 1858e, 381 (to Diplectanum).—Tasch., 1879, 264 (to Diplectanum).
 - alatus Linst., 1878, 227, fig. 10 (on Blicca bjærkna; Europe); 1889a, 91 (in Bl. bj., Alburnus lucidus).—Braun, 1890a, 544, 549, 550.
 - amphibothrium Wagener, 1857, 57, 58, 60, 66, 70, pl. 11, figs. 3, 4, pl. 12, figs. 1-4, pl. 15, fig. 11 (in Acerrina cernua).—Braun, 1890a, 544, 549, 550.—Dies., 1858e, 377.—Linst., 1873, 99; 1878, 229–230, fig. 15.—Tasch., 1879, 262 (in Acerina cernua).
 - anchoratus (Duj., 1845) Wagener, 1857, 49, 52, 55, 62, 99, pl. 15, fig. 8.—Dies.,
 1858e, 376.—Kath., 1894a, 156.—Reported for Carassius auratus, Cyprinus carpio.
 - auriculatus (Nord., 1832) Dies., 1850a, 433, 651-652; 1858e, 375-376; 1859c, 440.—
 Ben., 1858a, 1861a, 66 (to Gyrodactylus).—Braun, 1890a, 419, 544, 549, 550, 551.—Kath., 1894a, 156.—Kroyer, 1852-53a, 1223, 1225, 1226 (in Abramis brama L.; Phoxinus aphya L.).—Par. & Perugia, 1890, 9.—Pratt, 1900a, 657, fig. 44.—Tasch., 1879, 261 (syns. Gyrodactylus auric. Nord.; G. anchoratus Duj.) (in Abramis brama, Cyprinus carpio, Phoxinus kevis).—Wagener, 1857, 76.
 - benedeni St.-Remy, 1898, 566, 567 for Dactylogyrus vanbenedeni Par. & Perugia. calceostoma Wagener, 1857, 99 (in Sciæna aquila).—Dies., 1858e, 379 (in Sc. aq.); 1859c, 441 (syn. of Calceost. elegans Ben., type of Calceostomum).—Tasch., 1879, 265 (syn. of Calceost. elegans Ben.) (of Ben.).
 - cochlea (Wedl, 1857) Braun, 1890a, 408, 417.
 - cornu Linst., 1878, 228–229, fig. 13 (on Abramis vimba).—Braun, 1890a, 544, 549, 550 (on Ab. v.; Middle Europe).
 - crassiusculus (Wedl, 1857) Braun, 1890a, 417, 435.
 - cruciatus (Wedl, 1857) Braun, 1890a, 417.
 - crucifer Wagener, 1857, 55, 60, 63, 73, pl. 14, fig. 3 (in Cyprinus erythrophthalmus).—Braun, 1890a, 544, 549, 551.—Dies., 1858e, 377–378.—Tasch., 1879, 262 (in Leuciscus erythrophthalmus).
 - difformis Wagener, 1857, 63, pl. 15, fig. 6 (in Cyprinus erythrophthalmus).— Braun, 1890a, 544, 549, 551.—Dies., 1858e, 378 (in Leuciscus erythrophthalmus).—Tasch., 1879, 262 (in L. ery.).—Also reported for Scardinius erythrophthalmus).
 - dujardinianus (Dies., 1850) Dies., 1858e, 376 (in Cyprinus carpio, Leuciscus rutilus); 1859c, 440 (syn. Gyrodactylus auriculatus Ben.?).—Braun, 1890a, 544,549,550,551.—Kath., 1894a, 156.—Linst., 1877, 182–183.—Olss., 1893, 7.—St.-Remy, 1898, 566–567.—Tasch., 1879, 261 (syn. Gyrodac. auric. Nord. of Duj., 1845a) (in Cypr. carp., Leuc. rut., Abramis brama).—Also reported for Cypr. gibelio, Leuc. idus, L. prasinus.
 - echeneis Wagener, 1857, 99, pl. 15, fig. 13 (in Chrysophrys aurata).—Braun, 1890a,
 544, 549 (Genua), 550.—Dies., 1858e, 379 (in Chrys. aur.).—Par. & Perugia,
 1889, 746 (in Chrys. aur. Wagener); 1890, 746.—Tasch., 1879, 263 (in Chrys. aur.).
 - falcatus (Wedl, 1857) Dies., 1858e, 377 (in Cyprinus sp.).—Braun, 1890a, 417, 544, 549.—Tasch., 1879. 261–262 (syn. Gyrodactylus falcatus Wedl).

DACTYLOGYRUS-Continued.

fallax Wagener, 1857, 55, 70, pl. 11, figs. 1, 2, pl. 15, fig. 7 (in Cyprinus erythrophthalmus, C. rutilus).—Braun, 1890a, 544, 549, 551.—Dies., 1858e, 376–377.—Tasch., 1879, 261 (in Leuciscus rutilus, L. erythrophthalmus).—Also reported for Leuc. idus, L. prasinus, Scardinius erythrophthalmus.

forceps Leuck. (1857), 26 (in Chondrostoma nasus); 1858a. 118 (in Cyprinus nasus, C. dobula).—Linst.. 1878a, 254 (in Squalius cephalus), 257 (in Chond. nasus).

gracilis Wedl. 1861, 480, pl. 3, figs. 41–42 (on Hydrocyon dentex).—Braun, 1889a, 362.

maior Tasch., 1879, 263 (in Gobio fluviatilis) (for major).

major Wagener, 1857, 99, "pl. 15, fig. 9" (in Gobius fluviatilis).—Braun, 1890a,
 544, 549, 550.—Dies., 1858e, 379 (in Gobio flaviatilis).—Tasch., 1879, 263 (maior) (in Gobio fluviatilis).

malleus Linst., 1877, 182, pl. 12. figs. 12–13 (in Barbus fluviatilis).—Braun, 1890a, 544, 549, 550.—Sramek, 1901, 95, 110, fig. 64 (in Barb. fluv.).

megastoma [also megastomum] Wagener, 1857, 57, 62, 67, 99, pl. 14, fig. 5, pl. 36A, fig. 2 (in Cyprinus blicca. C. amarus).—Braun, 1890a, 544, 549, 551.—Dies., 1858e, 378 (in Rhodeus (Cyprinus) amarus).—Tasch., 1879, 262 (in Rhod. am.).

megastomum, see megastoma.

minor Wagener. 1857, 60, 63, pl. 14, fig. 4 (in Cyprinus alburnus).—Braun. 1890a, 544, 549, 550.—Dies., 1858e, 378 (in Aspius alburnus).—Linst., 1878. 227–228, fig. 11.—Tasch., 1879, 262 (in Aspius alburnus).—Also reported for Alburnus lucidus, Blicca bjærkna).

mollis (Wedl, 1857) Dies., 1858e, 378–379 (in Cyprinus carpio).—Braun, 1890a, 544, 549, 550.—Linst., 1885, 252. pl. 15. fig. 29.—Mont. 1888, 90.—Tasch., 1879, 262 (syn. Gyrodactylus mollis Wedl) (in Cyprinus carpio).

monenteron Wagener, 1857, 52, 55, 56, 58, 61, 63, 64, 65, 66, 67, 69, 70, 71, 72, 73, 76, pl. 13, fig. 1, pl. 36A, fig. 3f (in Esox lucius).—Braun, 1890a, 424, 452, 486.—Dies., 1858e, 380 (to Tetraonchus).—Olss., 1893, 7.

paradoxus (Crep., 1839a) Linst., 1878a, 210 (in Lucioperca sandra).

pedatus Wagener, 1857, 99, pl. 15, fig. 12, pl. 36A, fig. 3 (in Julis).—Ben. & Hesse, 1864, 121.—Dies., 1858e, 382 (to Diplectanum).—Tasch., 1879, 264 (to Diplectanum).—siluri Wagener, 1857, see Braun, 1890a, 544, 549, 552 (in Silurus glanis; middle

Europe).

siluri glanidis Wagener, 1857, see Dies., 1858e, 379 (in Silurus glanis).

species Olsson (in Coregonus lavaretus).

species Sramek, 1901. 95 (in Blicca björkna L.).

species Sramek, 1901, 95 (in Carassius vulgaris L.).

species Sramek, 1901, 95 (in Idus melanotus L.).

species Sramek, 1901, 96 (in Scardinius erythrophthalmus Bonap.).

sphyra Braun, 1890a, 544, 549, 550, for sphyrna.

sphyrna Linst., 1878, 229. fig. 14 (in Abramis vimba).—Braun, 1890a, 544 (sphyra), 549 (middle Europe), 550.

tenuis (Wedl, 1857) Dies., 1858e, 379 (in Perca fluviatilis).—Braun, 1890a, 417, 544, 549, 551.—Tasch., 1879, 262–263 (syn. Gyrodactylus tenuis Wedl) (in Perca fluviatilis).

trigonostoma Wagener. 1857, 99, "pl. 15, fig. 10" (in Cyprinus rutilus).—Braun, 1890a, 544, 549, 551.—Dies., 1858e, 379 (in Leuciscus rutilus).—Tasch., 1879, 263 (in Silurus glanis).—Also reported for Leuc. idus, L. prasinus.

tuba Linst., 1878, 228, fig. 12 (in Squalius leuciscus).

uncinatus Wagener, 1857, pl. 13, fig. 2 (in Perca fluviatilis).

unguiculatus Wagener, 1857, 61, 62, 64, 65.—Dies., 1858e, 381 (to Tetraonchus).—Tasch., 1879, 263 (to Tetraonchus).

van benedenii St.-Remy, 1898, 567 (syn. of Tetraonchus benedenii), for van benedenii. van benedenii Par. & Perugia, 1895, 2 (in Mugil auratus; Triest. Dec.), see also benedenii

DECACOTYLUS Mayer, 1841a, 24, D. lanceolatus alosæ.

lanceolatus alosa Mayer, 1841a, 24, Octobothrium lanceolatum renamed.

DENDROCCELI Burm.—Braun. 1890a, 515.—Mont., 1888a. 83.—Tasch., 1879. 233. contained f. Malacobothrii for Diplost., Cerc., Dist., Amphist., Holost.,

DENDROCELI-Continued.

Polyst., Monost., Caryophyllæus, and f. Plectobothrii for Aspidogaster, Trist., Octobothrium, Diplozoon, Hectocotyle, Cyclocotyle, Nitzschia, Axine).

DERMOCYSTIS Staff., 1905, Apr. 11, 682 (m. ctenolabri); δέρμα, skin; κύστις bladder.

ctenolabri Staff., 1905, Apr. 11, 682 (gills and skin of Ctenolabrus adspersus Walb.) includes Linton, 1899, 281, 296, pl. 40, figs. 76–81.

DEROGENES Luehe, 1900w, 507–509 (m. ruber); 1901n, 395, 396, 476, 478–479, 480, 481, 484 (Hemiuridæ).—Looss, 1901, 206; 1901, 438; 1901, 658; 1902, 839.—Odhn., 1905, 363, 364, 365, 366.—Pratt, 1902a, 889, 905 (key).—Stoss., 1902, 582.

affinis (Rud., 1819) Luehe, 1901n, 479 (in Scorpæna cirrosa).—Odhn., 1905, 364. cacozelus Nicoll, 1907, 72, 90–91, pl. 3, fig. 10 (in Pleuronectes limanda, Hippoglossus vulgaris).

minor Looss, 1901d, 437–439, fig. 5 (in Labrus merula; Triest).—Johnstone, 1907, 191–192.—Luehe, 1901n, 479.—Odhn., 1905, 361, 364.

plenus Staff., 1904, May 3, 484 (in Anarrhichas lupus; Canada); 1905, Apr. 11, 682.
 ruber Luehe, 1900, 507-509 (in gall bladder of Trigla lineata); 1901, 396, 479, 483.—Looss, 1901, 437, 438, 439.—Odhn., 1905, 362, 363, 364.

urocotyle (Par., 1899) Odhn., 1905, 364.

varicus (Mueller, 1784) Looss, 1901d, 438, 439.—Johnstone, 1907, 188–192, fig. 18 (in G. merl.; Cumberland).—Luche, 1901, 479.—Nicoll, 1907, 68, 70, 71, 72, 90, 91 (in Cottus bubalis, C. scorpius, Gadus merlangus, Hippoglossus vulgaris, Pleuronectes limanda, Rhombus lævis, R. maximus).—Odhn., 1905, 360–364, 365, pl. 4, figs. 6-7 (syns. Fasc. varica Mueller, Dist. dimidiatum.—Staff., 1904, May 3, 483 (in Salmo salar, Gadus callarias, Melanogrammus æglefinus, Pollachius virens, Clupea harengus, Osmerus mordax, Sebastes marinus, Anguilla anguilla, Cryptacanthodes maculatus, Acanthocottus scorpius, Hemitripterus americanus, Lophius piscatorius, Hippoglossus hippoglossus, Limanda ferruginea, Platysomatichthys hippoglossoides. Hippoglossoides platessoides; Canada), 484; 1905, Apr. 11, 682 (abundant in copepods, Acartia, Aug.).

DEROPRISTIS Odhn., 1902, 154-156 (tod. hispida).—Pratt, 1902a, 888, 896 (key).

hispida (Abildg., 1819) Odhn., 1902, 155, 156, 159.—Staff., 1904, May 3, 492 (hispidus, int. Acipenser rubicundus), 493.

inflata (Mol., 1859) Odhn., 1902, 157–160, figs. 1–2 (in Anguilla vulgaris; Triest).— Staf., 1904, May 3, 485 (int. A. anguilla; Canada).

DEUTEROBARIDINE Looss, 1902m, 699.

DEUTEROBARIS Looss, 1900, Dec. 3, 602—Baris Looss, 1899 [not Germ., 1817] renamed, hence type proteus; 1902, 546, 634, 635, 639, 646, 647, 648, 659, 660, 664, 669, 674, 675, 682, 684, 693–694, 695, 698, 699.—Pratt, 1902, 890, 909 (key).

proteus (Brandes, 1891) Looss, 1902, 416, 633, 694, pl. 31, figs. 151–168 (in first, widened, portion of large intestine of Chelone mydas).

DIAPHOROCOTYLINÆ Mont., 1903, 336 (f. Octocotylidæ).

DICLIBOTHRIUM Leuck., 1836a, 764, see Diclybothrium.

DICLIDOPHORA Dies., 1850a, 289, 417–418 (longicollis and palmata), 425; 1858e, 315, 383–384.—Braun, 1890a, 518.—Burm., 1856a, 251.—Cerf., 1895h, 920; 1895m, 130, 134, 137, 139, 140, 141, 142–143; 1896, 516, 540, 542, 545, 546, 547, 548, 549–550.—Gamb., 1896a, 73.—Goldb., 1855a, 19.—Johnston, 1865a, 31.—Mont., 1903, 336 (syn. Choricotyle) (subf. Diclidophorinæ).—Pratt, 1900a, 646, 652 (key), 656, 660, fig. 36.—St.-Remy, 1898, 523, 545, 551–552.—Tasch., 1879, 239.

affinis (Lint., 1898) Lint., 1901, 408, 414, 482 (in Paralichthys dentatus).—Pratt, 1900a, 656, fig. 36, 657, 660 (key).

chrysophri St.-Remy, 1898, 555, for chrysophryi.

chrysophryi (Ben. & Hesse, 1863) ————, 1895.

elongata Goto, 1894a, 210–212 (in Pagrus tumifrons, sometimes on Cymothoa;
 Mogi and Hakodate, Japan).—Ceri., 1895m, 126, 142, 144 (in Pag. tum.;
 Japan); 1896, 536, 547, 548, 550; 1898a, 303.—St.-Remy, 1898, 553.

DICLIDOPHORA—Continued.

labracis Cerf., 1895m, 126–142, 143, pl. 3, figs. 1–15 (in Labrax lupus; White Bank, N. Sea); 1896, 535–537, 539, 540, 548, 550, pl. 25; 1896, 230–232; 1898b, 338; 1899a, 371, 412.—Ben., 1895, 20–21.—Braun, 1896a, 1346.—St.-Remy, 1898, 554–555.

longicollis Dies., 1850a, 417 (Octost. merlangi Kuhn, 1830, renamed) (in Merlangus communis); 1858e, 384; 1859c, 443 (to Octoplectanum).—Ben., 1858a, 1861a, 44, 49 (syn. of Octobothrium merlangi.—Cerf., 1895h, 915 (syn. of Octob. merl.); 1895m, 139; 1896, 512, 547.—Tasch., 1879, 245 (syn. of Octob. merl. Kuhn).

merlangi (Kuhn, 1829) Kroyer, 1838–40a, 606 (in Merlangus vulgaris L.).

palmata (Leuck., 1830) Dies., 1850a, 417—418 (includes Octobothrium digitatum Rathke); 1858e, 384; 1859c, 443 (to Octoplectanum).—Cerf., 1895f, 20–21; 1895h, 917 (to Octobothrium); 1895m, 139; 1896, 513, 547.—Johnston, 1865a, 31.—Kroyer, 1838—40a, 608, 611 (in Lota molva L., Hippoglossus maximus Mind).—Tasch., 1879, 246 (to Octobothrium).

sessilis Goto, 1894a, 212–213 (in Chœrops japonicus; Mitsugahama, Japan).—Cerf., 1895m, 126, 140 (in Ch. jap.); 1896, 536, 547.—St.-Remy, 1898, 553–554.

smaris (Ijima, 1894) Goto, 1894a, 207–210, fig. 1 (syn. Octobothrium smaris); on Smaris vulgaris, on caudal segment of a Cymothoa; Gulf of Naples).—Cerf., 1895m, 126, 140, 142, 143–144 (in Cymothoa sp. in buccal cavity of Smaris vulgaris; Gulf of Naples); 1896, 536, 547, 548, 550; 1898a, 303 (in Cymothoa on Sm. vul.).—St.-Remy, 1898, 552–553, 556.

sp. Lint., 1905, 333, 380 (in Orthopristis chrysopterus; Beaufort, N. C.).

taschenbergi Par. & Perugia, 1892, 95, pl. 2, fig. 4, pl. 3, figs. 13–14, teste St.-Remy, 1898, 555.

tetrodonis Goto, 1894a, 213–215, pl. 10, figs. 1–4 (in Tetrodon sp.; Hagi, Japan).— Cerf., 1895m, 126, 140 (in Tetrodon sp.); 1896, 536, 547.—St.-Remy, 1898, 554.

DICLIDOPHORIN.E Cerf., 1895f, 20–21; 1895m, 132, 142; 1896, 535–552; 1898a, 303; 1899a, 345, 370.—Mont., 1903, 336 (f. Octocotylidæ).—St.-Remy, 1898, 552.

DICLYBOTHRIUM F. S. Leuck., 1835a, 88 (m. armatum); 1836a, 764 (Diclibothrium).—Ben. & Hesse, 1864, 84.—Braun, 1890a, 518.—Burm., 1856a, 251.—Crep., 1839, 292.—Dies., 1850a, 289, 421, 425; 1858e, 315, 383 (armatum); 1859c, 443.—Goldb., 1855a, 19.—Tasch., 1879, 254, 255 (syn. of Diplobothrium Leuck.).

1836: Diclibothrium Leuck., 1836a, 764.

1836: Diklibothrium Leuck., in Kollar, 1836, 81.

1842: Diplobothrium Leuck., 1842a, 18 (armatum).

armatum Leuck., 1835a, 88 (in Acipenser rostratus); 1836a, 764; 1836, 219.—Braun, 1889a, 332.—Crep., 1839, 292.—Dies., 1850a., 421 (includes D. crassicaudatum Leuck., Hexacotyle elegans Nord., Diplobothrium armatum Leuck., Polyst. armatum Duj.); 1858e, 383.—Staff., 1904, May 3, 488 (to Diplobothrium; on gills of Acipenser rubicundus).—Tasch., 1879, 254 (to Diplobothrium).

crassicaudatum Leuck., in Kollar, 1836, 81 (Diklibothrium) (in Acipenser stellatus).—Crep., 1839a, 292.—Dies., 1850a, 421 (syn. of Diclib. armatum).—

Tasch., 1879, 254 (syn. of Diplobothrium armatum Leuck.).

DICOTYLE—Stiles, 1898a, 59 (quoted from Sonsino; a larval trematode).

DICOTYLID. E Mont., 1903, 336 (subf. Dicotylinæ, g. Sphyranura).

DICOTYLINE Mont., 1903, 336 (g. Sphyranura).

DICRANOCŒLA Dies., 1850a, 286, 288, 293 (as tribe: "Tractus intestinalis bifurcatus"), 408.—Braun, 1893a, 892.—Goldb., 1855a, 15, 17.—Mont. 1888a, 83; 1892, Oct. 7, 196.

DICRANOCŒLI Burm., 1837, 528.—Braun, 1890a, 515.—Mont., 1888a, 83.—Tasch., 1879, 233.

DICRANOCŒLIA Dies., 1850a, 408 (subtribe I of Polycotylea, tribus III).

DICRANOPHORUS Nitzsch, 1827, 68 (contains Cerc. catellina, C. lupus, C. vermicularis, C. forcipata,? C. catellus).

(DICROCELIUM) Stiles & Hass., 1894e, 413, misprint for (Dicrocœlium).

DICROCCELIIN.E Looss, 1899b, Dec., 635; 1902, 839.—Hollack, Johanne, 1902a, 867–869 (amphitypie); 1903a, 536; 1905, July, 53–54.—Odhn., 1902, 40.—Pratt, 1902a, 889, 904 (key), includes Dicroccelium, Lyperosomum, Athesmia, related genera Eumegacetes, Anchitrema.—Ward, 1903, 863.

(DICROCŒLIUM) Duj., 1845a, 391–401 (tld. lanceolatum = lanceatum), subg. of Dist. See Dicrocœlium.

albicolle (Rud., 1819) Duj., 1845a, 387, 393 (as variety of D. (D.) attenuatum; in Falco pennatus). See Dicrocœlium.

albidum Braun, 1893e, 353.

arrectum (Duj., 1845) Stoss., 1895, 225-226, 236.

ascidia (Ben., 1873) Stoss., 1892, 21, 39, 40.

assula Duj., 1845a, 387, 398 (in Coluber natrix; Toulouse).

atomon (Rud., 1802) Stoss., 1886, 57.

attenuatum Duj., 1845a, 387, 392–393 (in Turdus merula; Rennes) [not attenuatum Rud., 1814]. See Dicrocœlium.

bacillare (Mol., 1859) Stoss., 1886, 58.—Looss, 1899b, 571 (as described by Stoss., would belong to Creadiine, possibly to Creadium).

boscii (Cobbold, 1859) Stoss., 1895, 226.

buski (Lankester, 1857) Rail., 1893a, 363-364, fig. 243. See Dicrocœlium.

calceolus (Mol., 1858) Stoss., 1886, 63.

chilostomum (Mehlis, 1831) Stoss., 1892, 20-21, 39, 40.

choledochum (Linst., 1883) Stoss., 1892, 161.

clathratum (Deslongchamps, 1824) Duj., 1845a, 393–394 (as var. of D. (D.) attenuatum). See Dicrocœlium.

clavigerum (Rud., 1819) Stoss., 1889, 64-65.

cælomaticum (Giard & Billet, 1892) Rail., 1893a, 360. See Dicrocælium.

commune (Olss., 1867) Stoss., 1886, 58.

complanatum (Rud., 1814) Duj., 1845a, 399–400 (believes it identical with D. (D.) hians.

complexum Stiles & Hass., 1894e, June, 425–426, pl. 1, figs. 1–3 (in Felis catus dom.; U. S. A.), 1894.

concavum (Crep., 1825) Stoss., 1892, 158-159, 188, 189.

conus (Crep., 1825) Stoss., 1892, 24–25, 39, 40.—Stiles & Hass., 1896a, 156 (to Opisthorchis).

crassicolle (Rud., 1809) Stoss., 1889, 63-64.

crassum (Sieb., 1836) Stoss., 1892, 156-157, 192.

 $\textit{cygnoides} \; (\textbf{Zed.}, \, 1800) \; \textbf{Duj.}, \, 1845a, \, 396-397. \\ \textbf{—Stoss.}, \, 1898, \, 31 \; (\text{to Pleorchis}).$

 $cylindraceum~({\bf Zed.,~1800})$ Duj., 1845a, 386, 395–396.—E. Bl., 1847, 295 (to Brachylæmus).—Looss, 1899b, 600 (type of Haplometra).

dendriticum (Rud., 1819) -----, 1896. See Dicrocœlium.

endolobum Duj., 1845a, 386, 397 (in "Grenouilles vertes et rousses, et de la Salamandre;" Rennes).—Looss, 1899b, 589 (type of Opisthioglyphe).

ercolanii (Mont., 1893) Stoss., 1895, 223–224, 234.—Looss, 1899b, 567 (probably identical with Telorchis linstowii).

erinaceum (Poir., 1886) Stoss., 1892, 22, 37.—Looss, 1899b, 570 (probably an Astia). eurystomum (Linst., 1877) Stoss., 1892, 159–160, 189.

felineum (Rivolta, 1884) Rail., 1893a, 361. See Dicrocœlium.

fasciatum (Rud., 1819) Stoss., 1886. 59.—Barbagallo & Drago, 1903, 410 (in Crenilabrus cœruleus, Serranus scriba; Catania).—Looss, 1899b, 571 (as described by Stoss., this belongs to Creadiinæ, possibly to Creadium).

flexuosum (Rud., 1809) Duj., 1845a, 386, 398–399.—Type of Omphalometra 1899.

furcigerum (Olss., 1868) Stoss., 1886, 58.

fuscatum (Rud., 1819) -? -, date?

fuscescens (Rud., 1819) Stoss., 1886, 59.

gelatinosum (Rud., 1819) Stoss., 1895, 226–227, 236, 237.—Looss, 1899b, 567–568. globiporum (Rud., 1802) Stoss., 1886, 59.

gobii.(Stoss., 1883) Stoss., 1886, 58; 1892.

heterostomum (Rud., 1809) Duj., 1845a, 400 (thinks it identical with hians).— Braun, 1899, 1 (to Dicroccelium); 1899, 487 (to Clinost.). See Dicroccelium. hians (Rud., 1809) Duj., 1845a, 399.—Looss, 1899b, 563 (type of Cathæmasia).

horridum (Leidy, 1850) Stoss., 1895, 220, 234. See Plagiorchis.

(DICROCŒLIUM)—Continued.

labracis Duj., 1845a, 386, 398 (in Labrax lupus; Rennes).—Ben., 1870, 45 (to Echinost.). See Dicrocœlium.

labri (Stoss., 1886) Stoss., 1886, 60.

lancea (Dies., 1850) Stoss., 1892, 26-27, 37.

lanceolatum (Rud., 1803) Duj., 1845a, 386, 391-392 [not Schrank, 1790]. See Dicroccelium lanceatum.

lingua (Crep., 1825) Stoss., 1892. 158. 190.—Looss. 1899b, 586 (type of Tocotrema).
 linstowi (Stoss., 1890) Stoss., 1895, 224–225. 237.—Looss. 1899b, 566 (to Telorchis).
 longissimum (Linst., 1883) Stoss., 1892. 161. 191.—Stiles & Hass., 1896c, 155 (to Opisthorchis).

longissimum (Poir., 1886) Stoss., 1892, 25-26, 37.

lucipetum (Rud., 1819) Duj., 1845a, 386. 400-401.—Looss, 1899b, 587 (to Philophthalmus).

macrourum (Rud., 1819) Baird, 1853a, 50.—Par., 1899 (to Dicroccelium); 1902. 4 (in Sturnus vulgaris, Turdus musicus; Elba). See Dicroccelium.

mutabile (Mol., 1895) Stoss., 1895, 224, 236. See Dicrocœlium.

naia (Rud., 1819) Duj., 1845a, 387, for naja.

naja (Rud., 1819) Duj., 1845a, 395.—Looss, 1899b, 604 (type of Macrodera).

neglectum (Linst., 1887) Stoss., 1889, 65.—Looss, 1899b, 617 (= Pleurogenes claviger).

nigrovenosum (Bellingham, 1844) Stoss., 1895, 222–223, 234.—Luehe, 1899, 535 (thinks D. nigrovenosum belongs to Lecithodendrium).

ovatum (Rud., 1803) Duj.. 1845a. 388, 394–395.—Stoss., 1892, 144 (to Cephalogonimus).—Type of Prosthogonimus 1899 and Prymnoprion 1899.

pallasii (Poir., 1885) Stoss., 1892, 27, 37.

pallens (Rud., 1819) Stoss., 1886, 59.

pellucidum (Linst., 1873) Stoss., 1892, 157, 192.—Looss, 1899b. 629 (to Prymnoprion). poirieri Stoss., 1895, 227, 237.—Looss, 1899b (to Telorchis).

pulchellum (Rud., 1819) Barbagallo & Drago. 1903, 410 (in Gobius jozo; Catania). rathouisi (Poir., 1887) Stoss., 1892, 27–28, 40.

reflexum (Crep., 1825) Stoss., 1886, 57.

retusum (Duj., 1845) Stoss., 1886, 63.

sauromates (Poir., 1886) Stoss., 1895, 220-221, 235.

scorpænæ (Rud., 1819) Stoss., 1886, 59.—Par., 1899, 5 (to Dicroccelium). See Dicroccelium.

simplex (Rud., 1809) Stoss., 1886, 58.

singulare (Mol., 1861) Stoss., 1892, 162, 191.—Looss, 1899b, 596 (type of Stomylus). spatula Duj., 1845a, 386, 394 (in Accentor modularis; Rennes).

spinosum (Linst., 1880) Stoss., 1892, 162, 193.—Luehe, 1899, 531 (perhaps syn. with Dist. cirratum).

squamula (Rud., 1819) Stoss., 1892, 20, 39.—Type of Eurysoma.

verrucosum (Mol., 1859) Stoss., 1900, 59. Homonym.

vitellilobum (Olss., 1876) Stoss., 1889. 67.—Looss, 1899b, 606-607.

viverrini (Poir., 1886) Stoss., 1892, 24, 39.—Stiles & Hass., 1896, 155 (to Opisthorchis).

DICROCŒLIUM (Duj., 1845) E. Bl., 1847, 291 (tld. lanceatum).—R. Bl., 1891, 609, 610; 1895, 730.—Braun, 1892a, 643; 1893a, 885, 909, 910; 1893, 353, 354; 1893f, 388; 1991a, 34; 1901, 562, 563, 565; 1901h, Nov. (n. sp. in Zibeth-katze), 700, 701, 702; 1902, 17 June, 356; 1902b, 97, 100, 102; 1903, 166; 1906, 173–174.—Cohn, 1903, 37.—Darr, 1902, 698.—Gomy, 1897a, 374.—Hausmann, 1897b, 29.—Hollack, 1902a, 867–869.—Hoyle, 1890, 540.—Klein, 1905, 20; 1905, 78.—Leuck,, 1863a, 586.—Looss, 1899b, 535, 610, 632–635, 648, 650, 721—Luehe, 1899, 530, 533; 1900, 490; 1901, 173.—Mont., 1888a, 92, 105; 1893, 150; 1896, 168.—Pratt, 1902a, 889, 904 (key).—Rail., 1900, 212; 1900, 239–242 (of birds); 1900, 514.—Rail, & Marotel, 1898, 33, 37, 38.—Sons., 1889, 276.—Stiles, 1898a, 22, 55; 1901, 193, 197.—Stiles & Hass., 1898a, 87 (lanceatum designated type), 97.—Stoss., 1892, 20; 1898, 41.—Ward, 1903, 871.

DICROCŒLIUM—Continued.

albicolle (Rud., 1819) Braun, 1901, 562; 1902b, 99-101, fig. 58 (includes macrourum Stoss., 1892, longicauda p. p. Muehling, 1890), in Aquila pennata; Vienna.

attenuatum (Duj., 1845) Rail., 1900, 241 (in Turdus merula).

buski R.Bl., 1888a, 622, for buskii.—Stiles, 1904i, 41.—Ward, 1895, 328 (in Homo). buskii (Lankester, 1857) Weinland, 1858, 87; 1859, 281,—Cobbold, 1866, 6; 1876. 303; 1879, 20.—Stiles, 1904i.

clathratum (Deslongchamps, 1824) Looss, 1899b, 634.—Braun, 1902b, 100, 101.— Rail., 1900, 239.

cwlomaticum (Giard & Billet, 1892) Rail., 1896, 160.—Gomy, 1897a, 374, 375.

concinnum Braun, 1901, 700-702, 1 fig. (in Viverra zibetha).—Hollack, 1902a, 869.

deflectens (Rud., 1819), 1901h, 702; Braun, [1901, 563]; 1902b, 101-102, fig. 59 (in Thryothorus hypoxanthus; Brazil).

delectans Braun, 1901g, 945-946 (in Myiothera ruficeps; Brazil); 1901h, 702; 1902b, 102–103, fig. 60.

dendriticum (Rud., 1819) Looss, 1899b, 634.—Braun, 1901h, 702.

felineum (Rivolta, 1884) Moniez, 1896, 136.—Type of Opisthorchis.

heterophyes (Sieb., 1852) Weinl., 1858, 86; 1859, 281.—R. Bl., 1888a, 625.—Cobbold, 1866, 6.—Ward, 1895, 328.—Type of Heterophyes.

heterostomum (Rud., 1809) Braun, 1899, 1; 1900h, 19, 30.

hospes Looss, 1907, Mar. 5, 478–479, fig. 1 (in cattle from Soudan, in Cairo, Egypt). illiciens Braun, 1901g, 944–945 (in Rhamphastus sp., Pipra rupricola: Brazil); 1901h, 702; 1902b, 102, 103, 104, 105–106, fig. 6.

labracis (Duj., 1845) Par., 1899, 5; 1902, 4 (syn. D. verrucosum Mol.) (in Labrax lupus; Elba).

lanceatum Stiles & Hass., 1896c, 158 (lanceolata Rud., 1803, not Schrank, 1790, renamed); 1898a, 87, 97 (type).—R. Bl., 1900, 488; 1901b, 204, 207, 209, 210; 1901c, 581 (in sheep), 584, 586, 587.—Bossuat, 1902, 188, 189.—Braun, 1901h, 702; 1903, 166–168, figs. 115–117; 1906, 174–176, figs. 106–108 (in Limax, Arion, 102, 1905, 190-108, ligs. 119-117; 1906, 174-176, ligs. 106-108 (in Limax, Arion, sheep, ox, goat, ass. horse, deer. hare rabbit, pig. man; Egypt, Siberia, Turkestan, North and South America).—Gomy, 1897a, 374.—Hollack, 1902a, 868.—Kamensky, 1902, 57-62 (in dogs).—Looss, 1899b. 633, 634, 635; 1905, 88 (rare in man); 1907, Feb. 1, 125, 126, 132, 134; 1907, Mar. 5, 479.—Rail., 1899, 345.—Rail. & Marotel, 1898, 33, 38.—Stiles, 1898a, 23, 55, 56, 137, 138, 139, 140, 141, 142, 143, figs. 36, 38; 1901, 193; 1902, 25, 28, 29, 33, 34; 1902, 204; 1903, 8, 84; 1904, Aug., 9, 12, 13, 29, 30, figs. 39-41.—Stoss., 1901, 97 (in Lepus timidus; Triest).—Ward, 1903, 407; 1903, 864, 866, 869, 871 (description) (description).

lanceolatum (Rud., 1803) Weinl., 1858, 86; 1859, 280.—R. Bl., 1888a, 602, 626; 1895, 734.—Braun, 1903, 166.—Engler, 1904, 186.—Galli-Valerio, 1898c, 7, 8; 1901c, 364 (mouton, cheval).—Kamensky, 1902a, 57–62 (in dog), 63–64.— Luehe, 1901, 172, 173, 1901, 487.—Stiles, 1898a, 55.

lobatum Rail., 1900, 241–242 (in Accipiter nisus).—Braun, 1901, 565; 1902b, 109, 110, 111 (to Lyperosomum).

longicauda (Rud., 1809) Looss, 1899, 634.—Braun, 1902b, 106, 107 (to Lyperosomum).—Rail., 1900, 240, 241.

lubens Braun, 1901g, 945 (in Pipra rupricola); 1901h, 702; 1902b, 102, 103, fig. 63 (Pipra rupricola).

macrourum (Rud., 1819) Par., 1899, 4.

mutabile (Mol., 1859) Braun, 1901, 702.

oculi humani (Gescheidt, 1833) Weinland, 1858, 86; 1859, 281.—R. Bl., 1888a, 630.—Stiles, 1902s, 29, 34.—Ward, 1895, 328 (in Homo).

olssoni Rail., 1900, 239 (Apus apus) "Dist. clathratum [Deslongchamps] of Olss. et Mueh.," renamed.—Braun, 1902b, 109.

 pancreaticum (Rail., 1890) Rail., 1897, 371–377, 1 fig.—Braun, 1901h, 702.—Gomy, 1897a.—Looss, 1899b, 634; 1907, Feb. 1, 128, 132 (of Rail. & Marotel, 1898, syn. of Eurytrema cœlomaticum).—Stiles, 1898a, 23, 55, 56, 57, 140, fig. 40.

panduriforme Rail., 1900, 240-241 (in Pica pica).—Braun, 1902b, 109.

DICROCELIUM-Continued.

petiolatum Rail., 1900, 241 (in Garrulus glandarius).—Braun, 1901, 946; 1901h, 702; 1902b, 98, 99, 109, fig. 57.

plesiostomum (Linst., 1883) Looss, 1899b, 634.

refertum (Mueh., 1898) Looss, 1899b, 634.

reficiens Braun. 1901g. 945 (in Falco nitidus); 1901h, 702; 1902b, 98, 103, fig. 62. scorpænæ (Rud., 1819) Par., 1899, 5; 1902. 5 (in Scorpæna scroia; Elba).

sinense (Cobbold, 1875) Moniez, 1896, 125.

strigosumLooss, 1899b, Dec., 634, 635, 727–728, fig. 47 (in Merops apiaster L. & Gm.; Marg).—Braun. 1902b, 109.

voluptarium Braun, 1901g, 945 (in Falco sp.); 1901h, 702; 1902b, 102, 103.

DICROGASTER Looss, 1902h, 134 (tod. perpusillus) (subf. Haploporinæ). contractus Looss, 1902h, 136, figs. 3–4 (in Mugil chelo).

perpusillus Looss, 1902h, 134, 135–136, figs. 1–2 (in Mugil chelo).

DIDYMOCISTIS Ariola, 1902, 105, for Didymocystis.

DIDYMOCYSTIS Ariola, 1902, 101-103 (tod. reniformis) fam. Didymozoonidæ.

reniformis Ariola, 1902, 101–103, 104, 107, figs. 1–3 (in Thynnus vulgaris; Naples) (includes Monost, bipartitum 2d form Wagener, 254, pl. 9, fig. 2, and Didymozoon thynni Tasch., 1879, 612).

wedli Ariola, 1902. 105–107, figs. 7–10 (in Thynnus vulgaris; Naples; Th. vul. and Th. tunnina from Gulf of Genoa) (includes Monost. bipartitum 3d form Wagener. pl. 9, figs. 5–8, and Didymozoon thynni partim Braun, Vermes IV, pl. 26, fig. 6d.).

DIDYMOSTOMA Ariola, 1902, 103–105 (evidently m. bipartitum), Didymozoonidæ.—

This genus falls as syn, of Wedlia 1860.

bipartitum (Wedl, 1855) Ariola. 1902, 103–105, figs. 4–6 (includes Monost. bipartitum Wedl partim, Didymozoon thynni partim, Monost. micropterygis Richiardi) (in Thynnus vulgaris; Naples).

micropterygis (Richiardi, 1901) Ariola, 1902, 105. See also under syn. of bipartitum.

DIDYMOZOON Tasch., 1878, 716 (no sp. mentioned); 1879, 72; 1879, 605–617 (Wedlia renamed, hence type thynni = bipartitum), pl. 6, figs. 1–5; διδυμον, ζωον, — Ariola, 1902, 25 July, 99–108, figs. 1–11; 1903, 14 Aug., 533; 1905, July, 61.— Braun, 1883, 42, 57; 1891d, 424; 1892a, 569, 574, 577, 660, 696, 698, 699, 700, 704, 707, 711, 727, 734; 1893a, 878, 879, 886, 890, 894, 895, 913, 918; 1895, 126, 136.—Hoyle, 1890, 539.—Jackson, 1888, 654.—Looss, 1892a, 81; 1899b, 542.— Maclaren, 1904b, 602, 612.—Moniez, 1891, 186, 187.—Mont., 1888a, 9, 11, 15, 18, 34, 52, 53, 57, 92, 93, 106; 1892, Oct. 7, 214 (g. of Didymozoonidæ); 1892, 689, 690, 693, 713, 716; 1893, 24, 27, 137, 149, 150.—Par. & Perugia, 1893, 2, 3.—Poche, 1907, 125.—Pratt. 1902a, 890, 908 (key).—Richiardi, 1902–3, 4–5 (n. sp.).—Schneidemuehl, 1896, 295.—Stoss., 1898, 62.

auxis Tasch., 1879, 613, pl. 6, figs. 1, 3 (in Auxis rochei; Naples).—Braun, 1883,
 41, 57; 1892a, 574; 1893a, 913.—Par., 1894, 169.—Par. & Perugia, 1890, 10;

1893, 2 (in A. roch.).

benedenii Mont., 1893, 137 (in Orthagoriscus).—Maclaren, 1904, 602.

exocæti Par. & Perugia, 1893, 1–4 (in Exocœtus volitans; Genova) (syn. Monost. filum Wagener not Duj.).—Par., 1902, 7 (in E. vol.).

filicolle Mont., 1893, 150.

lampridis Leennb., 1891, 73–75, pl. 2, fig. 9 (in Lampris guttatus; Kristiania Museum).—Braun, 1891d, 424; 1893a, 914.

micropterygis Richiardi, 1902, 4–5 (in Micropteryx dumerilii Cuv.).

pelamydis Tasch., 1879. 612, 614, 615, 616, pl. 6, figs. ii, v (in Pelamys sarda: Naples).—Braun, 1883a, 41: 1893a, 913.—Moniez, 1891, 186.—Par., 1894, 169; 1902, 7 (in P. s.).—Par. & Perugia, 1890, 10; 1893, 2 (in Pelamys sarda).

pretiosus Ariola, 1902, 107-108, fig. 11 (in Thynnus vulgaris).

scombri Tasch., 1879, 612, 615, 616, pl. 6, fig. iv (in Scomber colias; Naples).— Braun, 1892a, 734; 1893a, 913.—Mont., 1890, 427.—Par. & Perugia, 1890, 10; 1893, 2 (in Scomber scomber).—Stoss., 1890, 50; 1898, 63.

serrani Mont., 1889, 322–323, pl. 33, fig. 6 (in Serranus fimbriatus at Madeira and S. gigas at Naples).—Braun, 1893a, 914.—Par. & Perugia, 1893, 2 (in Serranus

gigas).

species Par. & Perugia, 1890, 10.

DIDYMOZOON—Continued.

sphyrænæ Tasch., 1879, 612, 614, 615 (in Sphyræna vulgaris; Naples).—Barbagallo & Drago, 1903, 411 (in Sphyræna vulgaris; Catania).—Brand., 1891d, 18.—Braun, 1893a, 913.—Maclaren, 1904b, 608.—Mont., 1888a, 7, 17; 1892, 716.—Par., 1894, 169; 1899, 7; 1902, 7 (in S. vul.).—Par. & Perugia, 1890, 10; 1893, 2 (in Sphyræna vulgaris).

tænioides Mont., 1888a, 93 (in Orthagoriscus mola); 1893, 137.—Maclaren, 1904b, 602.—Stoss., 1900, 8, figs. 10–11 (in Or. mola; Triest).

thynni Tasch., 1879, 612, 613, 614, 615, 616 (syn. Monost. bipartitum Wedl [type of Wedlia 1860]) (in Thynnus vulgaris).—Ariola, 1902, 100, 101 (of Tasch. syn. of Didymocystis reniformis Ariola), 103 (of Braun's Vermes, figs. 6–7, syn. of Didymostoma bipartitum), 105 (of Braun's Vermes, pl. 26, fig. 6d, syn. of Didymocystis wedli Ariola.—Barbagallo & Drago, 1903, 411 (in Thynnus vulgaris; Tonnara di Sa. Panagia).—Braun, 1883, 57; 1892a, 660, 734; 1893a, 914.—Gamb., 1896a, 71.—Moniez, 1891, 186.—Par., 1894, 169.—Par. & Perugia, 1889, 746–747 (syn. Monost. bipartitum Wedl.) (in Thynnus thunnina); 1890, 746–747; 1890, 9–10; 1893, 2.—Stoss., 1898, 62–63.

DIDYMOZOONIDÆ Mont., 1888a, 17, 18, 23, 52, 93, 106; 1892, 7 Oct., 214 (f. of suborder Malacocotylea); 1892, 689; 1893, 12.—Ariola, 1902, 101.—Brand., 1892, 505.—Braun, 1893a, 887, 890, 895, 900, 913; 1895, 136.—Looss, 1899b, 541, 543, 659.—Luehe, 1901, 488.—Maclaren, 1904b, 603, 612.—Par. & Perugia, 1890, 9.—Poche, 1907, 125.—Pratt, 1902a, 890, 908 (key), includes Didymozoon, Nematobothrium.—Schneidemuehl, 1896, 295.—Stoss., 1898, 62.

DIDYMOZOONTIDÆ Gamb., 1896a, 73.

DIGENA Olsson, 1893, for Digenea.

DIGENEA Ben., 1858, see Braun, 1883, 38, 58; 1890a, 473, 486; 1892a, 567, 574, 601, 775; 1893a, 888, 891, 895, 898; 1895, 136.—Carus, 1863, 478.—Gamb., 1896a, 73.—Hahn & Lefèvre, 1884, 804.—Hoyle, 1890. 539 (includes Monostomidæ, Distomidæ, Gasterostomidæ, Holostomidæ).—Kholodk., 1899a,149.—Knoch, 1894a, 11.—Kofoid, 1899, 183.—Looss, 1899, 543.—Odhn., 1902, 42, 43; 1905, 293–295.—Pratt, 1900a, 645, 646.—Stiles, 1898a, 27.—Ward, 1903, 865.

1893: Digena Olss., 1893, for Digenea.

DIGENEA s. strict. Leuck., see Looss, 1899b, 543.—Luehe, 1901, 488.

DIHEMISTEPHANUS Looss, 1901, 605-606, 628-629 (m. lydiæ).—Pratt, 1902a, 888, 894 (key).

lydiæ (Stoss., 1896) Looss, 1901, 605-606, 628-629.

DIKLIBOTHRIUM Leuck. in Kollar, 1836, 81.—Nord., 1840, 600, for Diclybothrium. crassicaudatum Leuck. in Kollar, 1836, 81.—Nord., 1840, 600 (syn. of Hexacotyle elegans).

DIONCHUS Goto, 1899a, 286 (m. agassizi), 291; 1901, 351–352.—Mont., 1903, 336 (Anisocotylinæ, subf.); 1905, 70.—Pratt, 1900a, 646, 650 (key), 655, 659,

agassizi Goto, 1899a, 286-291, pl. 21, figs. 19-24 (in Remora brachyptera; Newport, R. I.).—Pratt, 1900a, 655, 657. fig. 18, 659 (key).

agassizii Mont., 1905, 69, for agassizi.

DIOSTOMUM Duffek, 1903, 843 (misprint for Distomum).

DIPHTEROSTOMINÆ Stoss., 1904, 198.

DIPHTEROSTOMUM Stoss., 1904, 197–198 (tod. brusinai=brusinæ).

1905: Diphtherostomum Staff., 1905, Apr. 11, 684.

brusinæ (Stoss., 1889) Staff., 1905, Apr. 11, 684.

brusinai (Looss, 1901) Stoss., 1904, 197-198.

luteum (Ben., 1870) Stoss., 1904, 197 [name not available].—Staff., 1905, Apr. 11 (syn. D. betencourti Mont.).

DIPHTHEROSTOMUM Staff., 1905, Apr. 11, 684, for Diphterostomum.

DIPLECTANINE Mont., 1903, 336 (f. Gyrodactylidæ); 1905, 80.

DIPLECTANUM Dies., 1858e, 315, 381–382 (æquans [probably type], pedatum).—
Ben. & Hesse, 1864, 121, 122.—Braun, 1883, 48; 1890a, 412, 416, 452, 465, 469, 478, 479, 512, 517, 523, 542, 546; 1893a, 890.—Gamb., 1896a, 73.—Hoyle, 1890, 539.—Maclaren, 1904b, 574, 582, 583, 584, 589, 590, 595, 596, 597, 598, 599, 600, 601.—Mont., 1888a, 10, 14, 16, 66, 84, 86, 102; 1891, 111; 1892, Oct. 7, 213 (g. of Gyrodactylinæ); 1903, 336 (subf. Diplectaninæ); 1905,

DIPLECTANUM—Continued.

80.—Pratt, 1900a, 646, 654 (key). 657. fig. 45.—St.-Remy, 1898, 568.—Stoss., 1898, 18.—Tasch., 1879, 69; 1879, 264 (syn. Dactylogyrus Wagener).

acquans Sons., 1891, 263 (for æquans).

aculeatum Par. & Perugia, 1889, 745 (in Corvina nigra: Genova); 1890, 745; 1890, 4, 9.—Braun, 1890a, 417, 546, 549, 550.—Maclaren, 1904b, 576-577, 593, 601.—Par., 1894.

aquans (Wagener, 1857) Dies., 1858e, 381 (in Labrax lupus).—Ben. & Hesse, 1864, 121, 122–123, pl. 13, figs. 9–22 (in Lab. lup.).—Braun, 1890a, 408, 409, 417, 468, 479, 489, 514, 546, 549, 551: 1891d. 422.—Kerbert, 1881a, 572.—Maclaren. 1904b, 573, 574–602, pl. 20, figs. 1–12, pl. 21, figs. 13–16, 18, 20, 22, text figs. A. C. 1–5: 1904, 9 June, 443–444; 1905, 31 Jan., 20–21; 1905, June, 317; 1905, Dec., 703.—Mont., 1888a, 7, 8, 10, 13, 14.—Par. & Perugia, 1890, 4, 9, 11.—St.-Remy, 1898, 568.—Scott, 1905, 117, pl. 6, fig. 24 (in Lab. lup.).—Sons., 1890, 173–174 (in Umbrina cirrosa): 1891, 263 (acquans), 264 (in Umb. cirr-rhosa).—Stoss., 1885, 162: 1898, 18.—Tasch., 1879, 59; 1879, 264 (in Lab. lup.).

echeneis (Wagener, 1857) Par. & Perugia, 1889, 746 (in Sargus rondeletii; Genova); 1890, 746; 1890, 4, 9; 1895, 1.—Braun, 1890a, 417, 546, 549, 552.—Maclaren, 1904b, 576, 577–578, 601.—Par., 1894, 594.—Stoss., 1898, 18.—Also reported

for Chrysophrys aurata.

pedatum (Wagener, 1857) Dies., 1858e, 382 (in Julis sp. ad branchias).—Braun, 1890a, 546, 549, 551.—Maclaren, 1904b, 574, 575, 601.—Par. & Perugia, 1890,

9.—Tasch., 1879, 264 (in Julis sp.).

sciænæ Ben. & Hesse, 1863; 1864, 123–124, pl. 13. figs. 23–31 (in Sciæna aquila).— Braun, 1890a, 417, 546, 549, 552.—Maclaren, 1904b. 575, 601.—Mont., 1888a, 7, 4.—Pratt, 1900a, 657, fig. 45.—Sons., 1891, 263–264 (in Sci. umbra).— Tasch., 1879, 264 (in Sci. aq.).

sciænæ Ben. & Hesse, 1864, 123, misprint.

DIPLOBOTHRINÆ Mont., 1903, 336 (f. Hexacotylidæ); 1905, 78.

DIPLOBOTHRIUM Leuck.. 1842. 13 Diclibothrium renamed, hence (type by inclusion armatum) [not Diplobothrium Beneden. 1889, cestode].—Braun, 1889a, 338 (=Diclibothrium): 1890a, 413, 415, 511, 516, 517, 518, 523, 538, 539, 546; 1893a, 890.—Dies. 1850a, 421 (syn. of Diclibothrium).—Hoyle, 1890, 539 ("one species from the gills of a sturgeon").—Mont., 1888a, 8, 11, 13, 89, 100; 1892, Oct. 7, 213 (g. of Polystominæ): 1903, 336 (subf. Diplobothrinæ): 1905, 77, 78.—Pratt. 1900a, 646, 651 (key), 656, fig. 25.—Tasch., 1879, 69; 1879, 254 (syns. Diclibothrium Leuck., Hexacotyle Nord., Polyst. Duj.).

[affine Lænnberg, 1892. cestode.]

armatum (Leuck., 1835) Leuck., 1842, 13–18, pl. 1, figs. 6a–f.—Braun, 1890a, 414, 419, 539, 548, 550.—Dies., 1850a, 421 (to Diclibothrium).—Pratt, 1900a, 656, 657, fig. 25.—Staff., 1904, 3 May, 488 (on gills of Accipenser rubicundus; Canada).—Tasch., 1879, 254–255 (syns. Diclibothrium crassicaudatum Leuck., D. armatum Leuck., Hexacotyle elegans Nord., Polyst. armatum Duj.).—Reported also for Accipenser stellatus.

[simile Ben., 1889, cestode.]

DIPLOCOTYLE Dies., 1850a, 286, 301 (m. mutabile); syn. Diplodiscus [not Diplocotyle Krabbe, 1874, cestode]; 1855a, 378, 381, 393, 394; 1858d, 272.—Ariola, 1901, 412.—Burm., 1856a, 250.—Fil., 1854a, 6.—Goldb., 1855, 16.—Moul., 1856a, 121, 123.

[cohærens Linst., 1903, cestode.]

mutabile Dies., 1850a, 301 (in Planorbis nitidus: Ticini) (Diplodiscus diesingi Fil., renamed); 1855a, 394; 1858d, 241, 242, 272–273 (mutabilis) (syns. Diplodiscus diesingii, Cerc. diplocotylea Pag.) (larva of Diplodiscus subclavatus) (in Planorbis marginatus, P. nitidus, P. vortex; Heidelberg, Ticini, Moncalier).—Ben., 1858a, 1861a, 82 (syn. of Amphist. subclavatus).—Linst., 1873, 1 (larva of Diplodiscus subclavatus Dies.) (mutabilis).—Moul., 1856a, 107 (=Diplodiscus diesingii), 208 (syn. of Cerc. amphistomi subclavati).

mutabilis Dies., 1855a, 394 (for mutabile).

[olrikii Krabbe, 1874b, 22, cestode.]

[rudolphii (Mont., 1890) Mont., 1890, cestode.]

[serrata Linst., 1901, cestode.]

DIPLODISCINÆ Cohn, 1904, 242.

DIPLODISCUS Dies., 1836d, 237, 238, 253–255 [type subclavatus]; 1850a, 287, 301 (of Fil., syn. of Diplocotyle), 318–319 (syns. Planaria, Fasc., Dist., Hirudo, Amphist.); 1855a, 380, 384, 393 (cf. Diplocotyle); 1858e, 312, 359; 1859e, 435–436 (mentions only subclavatus, but reference is not necessarily designation of type).—Brand., 1888a, 12.—Braun, 1892a, 613, 650, 663, 664, 665, 698, 712, 715, 814; 1893a, 817, 820, 823, 827, 872, 884, 886, 890, 894, 895, 904, 906, 918; 1895, 136.—Cobbold, 1872b, 91 (subg. of Amphist.).—Cohn, 1904, 242.—Crep., 1839a, 286.—Fil., 1837a, 334 (mentions only diesingi); 1855b, 24.—Fischder., 1902a, 6; 1903h, 487, 489.—Gamb., 1896a, 73.—Goldb., 1855a, 17.—Looss, 1902m, 440, 442.—Mont., 1888a, 7, 12, 35, 84, 91, 103; 1892, Oct. 7, 214 (g. of Amphistominæ).—Moul., 1856a, 16, 121.—Nord., 1840, 629.—Piana & Stazzi, 1900, 523.—Poir., 1883, 79.—Pratt, 1902a, 887, 893 (key).—Schneidemuehl, 1896, 295 (Diplodiskus).—Sieb., 1854, 19, 20.—Sons., 1895, 185, 186.—Wagener, 1857, 27, 39, 45, pl. 16, fig. 4, pl. 17, fig. 2, pl. 18, figs. 1-4, pl. 19, figs. 1-4.

1850: Diplocotyle Dies., 1850a, 286, 301 (m. mutabile=subclavatum) [not

Krabbe, 1874, cestode].

1896: Diplodiskus Schneidemuehl, 1896, 295 (for Diplodiscus).

conicum Polonio, 1859; 1860, see Par., 1894, 140 (in Natrix torquata; Padova). [See also Dist. conicum Polonio.]

diesingi Fil., 1837a, 334–336, figs. 1–5, 337 (in Planorbis nitidus; Ticino).—Dies., 1850a, 301 (syn. of Diplocotyle mutabile; type of Diplocotyle).

diesingii Fil., 1854a, 6, pl. 1, figs. 1–4 [type of Diplocotyle] (in Planorbis nitidus) for diesingi; 1855b, 14, 20, 22, 23.—Ben., 1858a, 1861a, 81, 82 (syn. of Amphist. subclavatum).—Dies., 1855a, 394 (syn. of Diplocotyle mutabilis); 1858d, 272 (syn. of Diplocotyle mut.).—Levin., 1881a, 63.—Moul., 1856a, 95 (to Cerc.) (syn. Redia gracilis Fil.), 106, 107, 208, 209 (syn. of Cerc. amphistomi subclavati).—Villot, 1878, 36 (syn. of Cerc. diplocotylea Dies.).

mutabilis (Dies., 1850) Par., 1894, 140 to (Diplocotyle) (in Planorbis nitidus;

Pavia).

subclavatus (Pallas, 1760) Dies., 1836d, 238, 240, 253–254, pl. 24, figs. 19–24 (in Hyla arborea, Rana temporaria, R. esculenta, Bufo cinereus, B. igneus, Leptodactylus sibilatrix); 1850a, 318–319 (includes Hirudo tuba Braun); 1858e, 359–360 (in Dendrohyas viridis, Pelophylax esculentus, Rana temporaria, R. pipiens, Phryne vulgaris, Bufo viridis, Bombinator igneus, Leptodactylus sibilatrix, Lissotriton punctatus); 1859c, 435–436 (includes Amphist. subclavatum Ben., 1858, 81, Cerc. amphistomi subclavati Ben., 1858, 82, in Cyclas cornea, etc.).—Ben., 1858a, 1861a, 82 (to Amphist.).—Bettend., 1897a, 4, 37 (of Geze); 1897, 308.—Blochmann & Bettend., 1895a, 217, fig. 3.—Braun, 1892a, 598, 613, 620, 641, 650, 693, 698, 711, 759, 760, 766, 768, 776, 779, 783, 785, 798, 804, 806, 807, 809, 812, 813, 814; 1893a, 817, 818, 823, 824, 828, 830, 831, 906.—Cobbold, 1872b, 92; 1879, 51.—Cohn, 1903, 39; 1904, 240, 241, 242.—Crep., 1839a, 286, 287.—Darr, 1902, 679.—Fil., 1837a, 338 (to Amphist.); 1854; 1855.—Fischder., 1903h, 488.—Gamb., 1896a, 71.—Giebel, 1857, 266.—Kowal., 1894, 2.—Lang, 1892a, 81 (to Amphist.).—Linst., 1873, 1 (larva=Diplocotyle mutabilis Dies.); 1877, 13–14.—MacCallum, 1905, 669.—Mol., 1859, 27.—Much., 1898, 20.—Nord., 1840, 627 (to Amphist.), 629.—Par., 1894, 140 to (Amphist.) (in Rana esculenta; Padova).—Polonio, 1859.—Sieb., 1837, 263.—Sons., 1895, 180, 185.—Ssinitzin, 1905, 157–158; 1906, 685.—Stoss., 1890, 51.—Wagener, 1857, 26, 100, pl. 16, fig. 4, pl. 17, fig. 1.

subclavatus Dies., of Leidy, 1856b, 45 (in Rana pipiens); 1904a, 88.—Staff., 1905,

689–690 (syn. of Diplodiscus temperatus n. sp.).

temperatus Staff., 1905, 689 (in Rana catesbiana, R. virescens; Canada), includes subclavatus Dies. of Leidy, 1856.

unguiculatus (Rud., 1819) Dies., 1836d, 238, 254–255, pl. 24, figs. 25–27 (in Salamandra palustris); 1850a, 319.—Baird, 1853a, 44 (= Amphist. subclavatum Rud).—Braun, 1892a, 613.—Crep., 1839, 287.—Fischder., 1903h, 488.—Nord., 1840, 629 (in Triton lacustris).

DIPLODISKUS Schneidemuehl, 1896, 295, for Diplodiscus.

DIPLOOZON Mont., 1888a, 64, for Diplozoon.

DIPLOOZOON Mont., 1888a, 64, for Diplozoon.

DIPLOSTOMA Cobbold, 1860a, 49, for Diplostomum.

DIPLOSTOMATUM Olss., 1893, 8, see Diplostoma.

DIPLOSTOMEA Olss., 1893, 8, f. name for Diplostomeæ.

- DIPLOSTOME E Mont., 1888a, 6, 7, 11, 12, 14, 15, 18, 26, 28, 33, 34, 41, 46, 47, 51, 52, 53, 54, 56, 57, 60, 64, 67, 90, 91, 92, 94, 102, 103, 108; 1891, 110.—Brand., 1888a, 49; 1890a, 576, 580.—Braun, 1893a, 886, 887.
- DIPLOSTOMIDÆ Poir., 1886, v. 4 (3), 327–346, pls. 18–20; (1887) v. 1 (14), 425–426,— Brand., 1888a, 17, 49, 53; 1890a, 575.—Braun, 1893a, 886, 887.—Mont., 1888a, 10, 15, 22, 53, 91, 103.
- DIPLOSTOMINÆ Brand.—Braun. 1893a. 890. 895. 901.—Mont., 1892. Oct. 7, 214 (subf. of Holostomidæ).—Pratt, 1902a. 890, 908 (key) (includes Diplost., Polycotyle).
- DIPLOSTOMULUM Brand., 1892b, 511, as collective group, not as genus, hence no type.

constrictum (Dies., 1850) Brand., 1892b, 511.

- DIPLOSTOMUM Nord.. 1832a, 27–28, 34, 39, 47, 69 (type of first group Dip. volvens: type of second group Dip. clavatum; type by elimination and page precedence volvens: [not Diplostoma Rafinesque 1817, mammall: 1840, 561, 602, 617, 629–630.—Brand.. 1888a, 11, 12, 17, 18, 49, 50 (proposed as new genus; not of Nord., 52, 54–58; 1890a, 575, 576, 578, 580.—Braun, 1883, 40, 56; 1890a, 515; 1892a, 600, 658, 715, 774, 792, 793, 794; 1893a, 872, 879, 881, 884, 886, 887, 890, 892, 895, 900, 901, 917; 1893b, 187; 1894, 166; 1895, 132, 136.—Burm., 1837a, 529; 1856a, 250.—Cobbold, 1860a, 49 (Diplostoma).—Crep., 1839a, 289.—Dies., 1850a, 287, 304 (syn. of Tylodelphys, 305–307; 1855, 60; 1858e, 312, 317–318 (volvens, cuticola, grande; 1859c, 424.—Duj., 1845a, 473–474.—Fischer, 1840a, 156.—Gamb., 1896a, 64, 73.—Goldb., 1855a, 17.—Heider, 1900, 21.—Henle, 1835a, 587.—Hoyle, 1890, 539.—Jackson, 1888, 652, 653, 654.—Leuck., 1863a, 475, 525.—Mol., 1858, 287; 1861, 191.—Mont., 1888a, 15, 54, 57, 61, 71 (Displost.), 83, 84, 91 (Diplostostomum, 104; 1892, Oct., 7, 214 (g. of Diplostomime).—Moul., 1856, 12, 16 (probably young holostomes).—Pavesi, 1881, 615.—Poir., 1886, 327–346.—Pratt, 1902a, 890, 908 (key).—Schneidemuehl, 1896, 295.—Tasch., 1879, 233.—Villot, 1898, 541, 542.—Wyman, 1869, 42.—Wolf, 1903, 605.
 - 1860: Diplostoma Cobbold. 1860a, 49-51, for Diplostomum.
 - 1888: Diplostostomum Mont., 1888a, 91, misprint.
 - 1888: Displostomum Mont., 1888a, 71. misprint.
 - abbreviatum Brand., 1888a, 55 (in Crocodilus sp.; Brazil); 1890a, 581–582, pl. 39, figs. 15–17 (in Croc. sp.; Brazil).—Braun, 1893a, 901.
 - ægyptiaca Cobbold, 1876t. 757.—Fischder., 1903h. 488.—Sons., 1895, 179.
 - *agyptiacum** Cobbold. 1876t. 757 (in Equus: Egypt): 1877. 233, 235, 238.—Fischder., 1902a, 46 (syn. of Gastrodiscus polymastos Leuck.).—Lejtenyi, 1881a. 1.—Looss, 1896b. 13 (to Gastrodiscus).—Sons., 1895, 179.—Ward, 1895, 338 (in Equus caballus).—Zuern, 1882, 222.
 - alatum (Gœze, 1782) Par., 1894, 140 as Dip. Hemist.) alatum Dies. (in Vulpes vulgaris, Padova; Canis familiaris, Torino).
 - auriflavum Mol., 1859, 287 (in Ardea nycticorax: Batavii); 1861, 191–192, pl. 1, figs. 1–2.—Braun, 1893a, 901.—Dies., 1859c, 424.
 - bifurcatum (Wedl. 1861) Brand., 1888a, 57–58 in Crocodilus vulgaris; Egypt); 1890a, 584.—Braun, 1893a, 901.—Odhn., 1902, 19.
 - brévicaudatum (Nord., 1832) Dies., 1850a., 306.—Braun, 1892a., 796.—Kroyer, 1852-53a, 1222 (in Barbus fluviatilis Ag.).—Villot, 1898, 542.
 - clavatum Nord.. 1832a, 42–49, pl. 3, figs. 5–10. pl. 4, fig. 5 (in Perca cernua. P. fluviatilis, P. lucioperca): 1833, 286–295, pl. 18, fig. 4, pl. 19, fig. 2: 1840, 630.—Braun, 1892a, 654, 794.—Claparède, ——, 198, pl. 8, figs. 1–3.—Crep.. 1839a, 289.—Dies., 1850a, 305 (to Tylodelphys): 1858e, 316 (to Tylodelphys).—Duj., 1845a, 475.—Gescheidt, ——, 428.—Henle, 1833a, 22.—Kroyer, 1838–40a, 21; 1846–53a, 253 (in Perca fluviatilis, Esox lucius).—Moul.. 1856a, 220–221 (in eyes of perch by Nord.: Esox lucius Duj.; P. fluviatilis Moul.: Lake Leman).—Olss., 1893, 8.—Steenstrup, 1842, 58.
 - cobitidis Linst., 1890d, 179 (in Cobitis barbatula .—Braun, 1891d, 424 in Cob. barb.).
 - craniarium (Dies., 1858) Cobbold, 1860, 49-50 (includes Trematodum sp. Leydig) in Cobitis fossilis .
 - cuticola (Nord., 1832) Dies., 1850a, 306; 1858e, 317-318 to Holost. (in Pomotis vulgaris: Phila.: Leuciscus dobula, L. rutilus: Anglia: Gobio vulgaris. Phoxinus lævis).—Braun, 1892a, 796.—Koch, 1904, 795.—Kroyer, 1838-40a,

DIPLOSTOMUM—Continued.

578 (in Perca fluviatilis L.).—Lint., 1898, 513, pl. 41, figs. 6-10, pl. 42, figs. 1-5.—Staff., 1904, May 3, 494 (encysted in liver of Ambloplites rupestris: Canada).—Villot, 1897; 1898, 541, 542 (larva of Hemist. denticulatum) (in Phoxinus lævis).—Wellner, 1899, 51-53, 2 figs. (in Weissfisch).—Also reported for Abramis brama, A. vimba, Blicca bjærkna, Gobitis tænia, Chænobrythus gulosus, Chondrostoma nasus, Cyprinus carpio, Eupomotis auratus, E. pallidus, Gobio fluviatilis, Idus melanotus. Lepomis auritus, Leuciscus erythrophthalmus, L. idus, L. vulgaris, Scardinius erythrophthalmus.

[fusca=Geomys bursarius, mammal].—Stiles, 1896, 183.

grande Dies., 1850a, 305, 307 (in Ardea leuce, A. agami; Matogrosso, Brazil); 1855, 60, pl. 1, figs. 1–12; 1858e, 318; 1859c, 424 (in Strix nivea).—Brand., 1888a, 12, 18, 26, 50, 54–55 (syn. Hemist. macropterum) (in Ardea leuce; Brazil); 1890a, 554, 576, 581, pl. 39, fig. 14 (in Ardea leuce: Brazil).—Braun. 1892a, 582; 1893a, 901.—Leidy, 1859, 110.—Mont., 1888a, 12.—Poir., 1886, 328.

lenticola Linst., 1878, 226–227, fig. 9 (in Abramis vimba).—Braun, 1892a, 796.— Poir., 1886, 336 (in Ab. vim.).

longe Braun, 1892a, 581, see longum.

longum Brand., 1888a, 25, 55, 57, 61 (in Crocodilus: Brazil); 1890a, 553, 582, 584, pl. 39, figs. 1-9 (in Croc. by Natterer; Brazil).—Braun, 1892a, 569, 582, 586, 672; 1893a, 901.

macrostomum Jægers., 1900a, Jan. 6, 33-37, figs. 1-5 (in Telmatias major; ?Upsala). mülleri Cobbold, 1860a, 50 (includes D. petromyzi fluvialis Mueller, D. rachiæum Mueller?, Tylodelphys Dies. Revis., p. 12) (in Petromyzon fluviatilis).

musculicola (Waldenburg, 1860) Braun, 1892a, 796.

mutabile (Dies., 1850) Par., 1896, 2, to (Codonocephalus) (in Rana esculenta; Italy).

parvulum Staff., 1904, 494 (in Semotilus bullaris; Canada).

peteromyzi fluviatilis Dies., 1850a, 307, based on Mueller, 1840, 30 (in Peteromyzon fluviatilis); 1858e, 318 (to Tylodelphys).

pseudostomum (Will.-Suhm, 1870)
Poir., 1886, 334–339, 345, pl. 18, figs. 3–4, 5, 6, pl. 19, figs. 2, 4.—Brand., 1888a, 34, 50, 55–56 (in Crocodilus; Brazil); 1890a, 563, 576, 582–583, pl. 39, figs. 10, 11 (in Croc. sp.).—Braun, 1892a, 593; 1893a, 901.

putorii Linst., 1877, 191, pl. 14, fig. 21 (in Fœtorius putorius).—Brand., 1888a, 42; 1890a, 572.—Braun, 1894a, 796. rachiæum Crep., 1839, 289.—Mueller, 1839, 198.

rachidis Par., 1896, 2, for rhachiæum to (Tylodelphis).

rachieum Fraip., 1880c, 419, for rhachiæum.

rachineum Mueller, (1842).

rhachiæum Henle, 1833a, 19-22, pl. 1, figs. 18-22 (in Rana).—Brand., 1888a, 15.— Braun, 1892a, 654 (rachiæum), 795.—Claparède, ——, 198, pl. 8 (rachiæum).— Crep., 1839, 289 (rachiæum).—Dies., 1850a, 305 (syn. of Tylodelphys rhachidis); 1858e, 316 (rachiæum), (syn. of Tylodelphys rachidis).—Duj., 1845a, Hofman, 1899, 184 (rachiæum).—Jackson, 1888, 645 (excretory system).— Kuech., 1855, 187 (rachiæum).—Leidy, ——, 383.—Leuck., 1863a, 526 (rachiæum).—Mont., 1888a, 41 (rachieum).—Moul., 1856a, 221 (rachiæum).— Mueller, 1839, 198 (rachiæum); 1842, — (rachineum) (in Petromyzon fluviatilis).—Par., 1896.

siamense Poir., 1886, 327-333, 336, 340, 345, pl. 18, figs. 1, 2, pl. 19, fig. 7, pl. 20, figs. 1–4 (in Crocodilus siamensis).—Brand., 1888a, 26, 34, 50, 56–57 (in Croc. si.); 1890a, 554, 563, 576, 583.—Braun, 1892a, 717; 1893a, 901.—Mont., 1888a, 12.

spataceum Stoss., 1896, 126 (for spathaceum).

spathaceum (Rud., 1819) Olss., 1876, 29-30.—Braun, 1893a, 901.—Stoss., 1896, 126 (spataceum); 1898, 20 (to Conchosomum).

spathula Brand., 1888a, 54 (syn. Hemist. spathula) (in Falco palumbarius; Wien. Museum); 1890a, 580, pl. 39, fig. 13.—Braun, 1892a, 582; 1893a, 901.—Also reported for Astur palumbarius.

DIPLOSTOMUM—Continued.

spathulæforme Brand., 1888a, 44, 54 (in Strix otus); 1890a, 574, 580–581, pl. 39, fig. 12 (in Otus vulgaris).—Braun, 1892a, 582; 1893a, 844, 845, 901; 1894, 166.—Wolffhuegel, 1900, 9, 11, 12, 14.—Also reported for Accipiter nisus, Ægolius otus, Buteo vulgaris, Sarcorhamphus gryphus.

sp. Kroyer, 1852–53a, 1051 (in Petromyzon fluviatilis L.).

sp. Lint., 1901, 415, 442, 471.

volvens Nord., 1832a, 28–41, 43, pl. 1, fig. 3, pl. 2, figs. I–10, pl. 3, figs. 1–4, 9, pl. 4, fig. 6 (in Cyprinus erythrophthalmus, Gadus lota, Perca cernua, P. fluviatilis, P. lucioperca); 1833, 270–286, pls. 18, 19, figs. 1–3 (pl. 19, fig. 1, gives Dist. volvens, probably lapsus); 1840, 630.—Aubert, 1853, 90.—Brand., 1888a, 12; 1890a, 569.—Braun, 1892a, 636, 637, 648, 653, 654, 794–796; 1894, 166, 167; 1894k, 681 (larva of Hemist. spathaceum).—Brown, Λ. W., 1899d, 490, 493, 494.—Claparède, —, 100, pls. 4, 5.—Cobbold, 1858b, 159.—Crep., 1839a, 289; 1846, 150, 153.—Dies., 1850a, 306; 1858e, 317.—Duj., 1845a, 474–475.—Fraip., 1880a, 400; 1880c, 419, 420, 429–433, 441, 442, 443, pl. 18, figs. 18–21 (in Leuciscus rutilus, Chondrostoma nasus); 1881b, 1, 7; 1883a, 36.—Gamb., 1896a, 64, fig. 31 (as Dip. (Tetracotyle) volvens).—Gescheidt, 1833, v. 3, 426–428.—Henle, 1883a, 22.—Jackson, 1888, 645 (excretory system).—Kroyer, 1838–40a, 21, 579, 609; 1852–53a, 1224 (in Leuciscus rutilus L. L. erythrophthalmus, Lota vulgaris, Perca fluviatilis L., Acerina cernua L.).—Looss, 1885b, 20.—Mont., 1888a, 41, 42, 43, 45, 46.—Moul., 1856a, 220 (in eye of Perca fluviatilis, P. cernua, P. lucioperca, Gadus lota, Cyprinus erythrophthalmus).—Mueh., 1898, 16.—Olss., 1893, 9 (in Leuciscus phoxinus).—Steenstrup, 1842, 58.—Villot, 1898, 542 (larva of Hemist. spathaceum).—Also reported for Acerina cernua Λ. vulgaris, Lota communis, Lucioperca sandra.

DIPLOSTOMUM Brand., 1888a, 50, 54-58, proposed as new genus [no type given], [not Nord., 1832], for abbreviatum, bifurcatum, grande, longum, pseudostomum, siamense, spathula, spathulæforme see under foregoing entry; 1890a.—

The generic name Diplostomum Brand. falls as a homonym.

DIPLOSTOSTOMUM Mont., 1888a, 91 for Diplostomum.

DIPLOOZON Mont., 1888a, 64 (for Diplozoon).

DIPLOOZOON Mont., 1888a, 64 (for Diplozoon).

DIPLOZOON Nord., 1832a. 56–76 (m. paradoxum); 1840, 599.—Beard. 1905, v. 93, 383, 2 figs.—Ben., 1858a. 1861a, 11, 38, 168, 172, [200], 345; 1868, 28.—Ben. & Hesse, 1864, 61, 64, 96.—Brand., 1891b. 266; 1894a, 308.—Braun, 1889k, 620, 621; 1890b, 125, 127, 128; 1891d. 422; 1890a, 413, 414, 442, 451, 461, 468, 472, 475, 477, 480, 481, 482, 485, 491, 492, 493, 494, 495, 498, 500, 506, 507, 511, 514, 515, 516, 517, 518, 520, 522, 523, 533, 534, 535, 546; 1893a, 890; 1895, 125; 1899, 3.—Burm., 1837a, 530; 1856a, 251.—Carus, 1833, 477.—Cerf., 1895h, 918, 920; 1896, 514, 515; 1899a, 451.—Chatin, 1880f, 591–592; 1881a, 310–312; 1887d, 1005.—Crep., 1838, 84; 1839a, 292.—Dies., 1850a, 90, 422–423, 425; 1858e, 269–272, 315 (in subf. Placoplectana), 387.—Duj., 1845a, 315.—Gamb., 1896a, 55, 57, 61, 73.—Goldb., 1855a, 19.—Goto, 1891a, 167, 168, 169, 170, 176, 177, 178, 181, 183, 186, 187; 1891c, 103; 1893a, 800.—Haswell, 1892b, 149; 1893c, 114.—Hoyle, 1890, 539.—Ijima, 1884, 637, 638.—Jackson, 1886, 649 (=Diporpa), 653.—Juel, 1889, 33.—Kath., 1894a, 148, 155.—Leuck., 1863a, 48; 1879, 58, 62.—Leuck., 1886d, 45, 48.—Looss, 1892a, 72–73; 1893b, 810.—Mayer, 1841a, 34.—Mont., 1888a, 11, 15, 37, 49, 51, 52, 53, 55, 57, 58, 59, 60, 61, 63, 64 (Diploozon, Diploozoon), 65, 66, 70, 83, 84, 86, 89, 98, 99; 1892, Oct. 7, 186, 213 (in Octocotylinæ); 1893, 111; 1903, 336 (subf. Octobohtrinæ).—Moul., 1856a, 10.—Pintner, 1891, 728.—Pratt, 1900a, 646, 651 (key), 656, fig. 31.—Tasch., 1879, 35, 69; 1879, 232, 233, 238, 249 (syn. Diporpa Duj.).—Wallenstedt, 1847, 7.

1835: Diplozoum Burm., 1835b, 187, for Diplozoon.

1888: Diploozoon Mont., 1888a, 64, for Diplozoon.

1888: Diploozon Mont., 1888a, 64, misprint.

1900: Diplozoön Pratt, 1900a, 646, 651.

nipponicum Goto, 1891a, 151–192, pls. 21–23 (in Carassius vulgaris); 1891b, 472–473.—Braun, 1891d, 422.—Mont., 1893, 8, 111.

paradoxum Nord., 1832a, 56-57, pl. 5, figs. 1-5, pl. 6, figs. 1-2 (in Cyprinus brama); 1833, 373-396, pl. 20, figs. 1-4; 1840, 547, 597, 599.—Ben., 1858a, 1861a, 39-44, 53, 66, 99, 169, 176, 186, pl. 4, figs. 1-12.—Braun, 1883, 45, 51, 57, 71; 1890a, 409, 415, 420, 428, 438, 439, 445, 451, 452, 455, 456, 457, 465, 472, 479, 483, 489,

DIPLOZOON—Continued.

492, 499, 506, 512, 535, 548, 550, 551; 1891a, 52, 53.—Burm., 1837a, 530.—Cobbold, 1872b, 90.—Crep., 1838, 89 (in Cyprinus balerus, C. brama, C. jeses, C. rutilus, C. vimba; on gills), 164; 1839a, 292.—Dies., 1850a, 423; 1858e, 282, 387 (in Carassius gibelio; Rhodeus amarus); 1859c, 444.—Duj., 1845a, 316.—Ehrenb., 1835, 128.—Fraip., 1880a, 400 (paraodoxum); 1880c, 416, 418, 433; 1883a, 36.—Gamb., 1896a, 59–61, figs. 27, 28.—Gosse, 1857a, 125.—Goto, 1891a, 151, 175, 185.—Hausmann, 1897b, 4, 7, 20, 22, 36–38 (in Abramis brama, Cottus gobio, Gobio fluviatilis, Phoxinus lævis).—Heller, 1857a, 109–110, 5 figs.—Hoyle, 1890, 539, fig. 3, A.—Ijima, 1884c, 637.—Jackson, 1888, 650.—Jacoby, 1900, 3.—Kerbert, 1881a, 556.—Kholodk., 1899a, 148, fig. 176.—Kollar, 1835, 81; —, 82.—Kroyer, 1846–53a, 388, 410, 419, 434, 446, 462; 1852–53a, 1223, 1224 (in Abramis balerus, A. blicca, A. brama, A. vimba; Leuciscus erythrophthalmus, L. idus, L. rutilus).—Lamarck, —, 599.—Leuck., 1863a, 450.—Looss, 1885b, 18; 1892a, 81.—Mayer, 1841a, 23.—Mont., 1888a, 9, 38, 40, 52, 58, 70; 1890, 421; 1892, Oct. 7, 186; 1893, 110, 111.—Mueh., 1898, 12, 18.—Olss., 1893, 6.—Par., 1896, 1.—Par. & Perugia, 1890, 7.—Paulson, 1862, 16 pp., 1 pl.; 1867, 1–24.—Pratt, 1900a, 656, 657, fig. 31.—Sieb., 1835, 58, 59; 1836, 105; 1836, 238; 1839, 163, 164; 1842, 459; 1850, 36–37; 1851, 10 Mar., 62–68 (conjugation); 1854, 52–58, 201–206.—Sons., 1897, 259.—Sramek, 1901, 95, 109, fig. 63 (in Abramis brama Cuv.).—Tasch., 1879, 249 (syn. Diporpa dujardini Dies.); 1879, 613.—Vogt, 1841, 33–36, pl. 2, figs. 10–12.—Wagener, 1857, 96, pls. 9–10.—Zeller, 1872, 3 May, 168–180, pl. 12, figs. 1–13 (development); 1872, Sept., 99–112; 1888, 23 Mar., 233–239, pl. 19 (genital organs); 1888, 427–428.—Also reported for Blicca bjerkna, Carassius vulgaris, Chondrostoma nasus, Cyprinus blicca, C. cephalus, C. gobio, C. leuciscus, Gasterosteus aculeatus, Idus melanotus, Leuciscus cavedanus, L. prasinus, Lota vulgaris, Scardinius erythrophthalmus.

DIPLOZOUM Burm., 1835b, 187, for Diplozoon.

DIPORPA Duj., 1845a, 316–317, pl. 3, fg. C (m. dujardinii Dies., 1850).—Ben., 1858a, 1861a, 38, 199.—Ben. & Hesse, 1864, 61, 64.—E. Bl., 1847, 337–338.—Braun, 1883, 57; 1890a, 424, 442, 452, 506, 507, 518, 535.—Chatin, 1880f, 591–592; 1881a, 310–312.—Cunningham, 1887a, 279.—Dies., 1850a, 289, 420, 425; 1858c, 269–272; 1858e, 315, 386.—Gamb., 1896a, 61.—Goldb., 1855a, 19.—Heller, 1857a, 110.—Jackson, 1888, 649.—Looss, 1892, 81.—Kroyer, 1846–53a, 434 (in Leuciscus erythrophthalmus).—Mont., 1888a, 70, 83, 84, 89.—Tasch., 1879, 232, 249 (syn. Diplozoon Nord.).

dujardinii Dies., 1850a, 420 (in Cyprinus erythrophthalmus) (based on Duj., 1845a, 316-317, pl. 3, fig. c); 1858c, 271; 1858e, 386 (in Phoxinus laevis).—Ehrenb., 1852, 28.—Olss., 1893, 6.—Sieb., 1850, 36-37; ——, 62.—Tasch., 1879, 249 (syn. of Diplozoon paradoxum Nord.).—Also reported for Abramis

alburnus.

DISCOCOTYLE Dies., 1850a, 290, 423–424 (only positive species, hence type sagittata), 425.—Goldb., 1855a, 19; Tasch., 1879, 239.

1858: Placoplectanum Dies., 1858e, 315, 384, Discocotyle renamed, hence type sagittatum.

1890: Discotyle Braun, 1890a, 518, misprint.

hirundinaceum (Bartel, 1834) Dies., 1850a, 424 (sp. inq., nomen nudum except for habitat, Coregonus wartmanni).

leptogaster (Leuck., 1830) Dies., 1850a, 424 (in Chimera monstrosa; Christiania).— Kroyer, 1852–53a, 812 (in Ch. mon. L.).—Tasch., 1879, 245 (to Octobothrium).

sagittata (Leuck., 1842) Dies., 1850a, 423–424 (in Salmo fario).—Kroyer, 1838–40a, 616 (in Salmo fario).—Tasch., 1879, 244 (to Octobothrium).

DISCOCOTYLEA Dies., 1850a, 419, 422 (supergeneric).

DISCOCOTYLEÆ Dies., 1850a, 288, 289, 290 (supergeneric).—Goldb., 1855a, 17, 19.

DISCOTYLE Braun, 1890a, 518, misprint for Discocotyle.

DISCOTYLEA Cerf., 1899a, 351.

DISPLOSTOMUM Mont., 1888a, 71 (for Diplost.).

DISSICHYA Cosmovici, 1887a, 129, 131 (includes Amphist., Bilharzia, Dist. Gasterost., Hemist., Holost.).—Mont., 1888a, 84.

DISTOAM Cobbold, 1875n, misprint for Distoma.

DISTOMA Retzius (1782); 1790, 32 (Fasciola Linn., 1758, renamed, hence type hepatica) [not Distoma Savigny, 1816, mollusk; Distomus Gærtner 1774,

Ascidiæ compositæ; Distomus Steph., 1827, coleopteron; not Distoma Gærtner of Herdmann, 1890].—Abildg., 1790, 36 (syns: Fasc. hep. Linn., Fasc. Mueller, Planaria Gæze, Fasc. Bloch).—Adam, 1879, 159.—Aitken, 1872, 203, fig. 36a.—Albarran, 1897b, 1096.—Anacker, 1885e, 438; 1885d, 380, 381; 1887b, 350.—Andral, 1829d, 617.—Ariola, 1899a; 1899 (in 129-138); —, 299.—Ashley, 1856, 7. 8.—Audouin, 1824a, 563-564 (polyp).—Baillet, 1866b, 90.—Bellingham, 1844a, 22, 423.—Rep., 1870a, 142.—Rep., 442. —, 299.—Ashley, 1856, 7. 8.—Audouin, 1824a, 563–564 (polyp).—Baillet, 1866b, 99.—Bellingham, 1844a, 422–423.—Ben., 1870c, 142.—Ben. & Hesse, 1864a, 61.—E. Bl., 1847a, 291–292.—R. Bl., 1888a, 543, 567, 570, 575, 577, 584, 585, 590, 593, 594, 595, 597, 598, 599, 601, 602, 603, 605, 614, 615, 622, 624, 625, 627, 628. 630, 631, 636, 641, 643, 644, 646, 647; 1891p, 609; [1892b], 1026–1030, figs. 1–7.—Bojanus, 1817, pl. 9.—Brand., 1888a, 9; 1892b, 506.—Braun, 1883a, 59, 70; 1890a, 473, 479, 514, 515; 1891d, 423; 1892a, 570, 599, 635, 696, 715, 720, 735, 767, 769, 770; 1893a, 825, 837, 853, 857, 872, 879, 880, 884, 885, 886, 887, 890, 893, 894, 895, 902, 908, 913, 918; 1893f, 382, 383, 388, 389; 1893g, 802; 1894i, 604; 1895b, 17, 134, 136, 138; 1900b, in 217–236, 1 pl.; 1900. 3; 1900g, 254; 1900h, 3, 6; 1901e, 323, 332, 333, 338, 339, 340, 342; 1902b, 41, 51, 52, 53, 79, 117.—Burm., 1837a, 529; 1856a, 250.—Bremser, 1824, 133–134.—Buttel-Reepen, 1900a, 585–598, figs. 1–9 (2 species); 1902a, 282–283.—Carus, 1835a, 90, 91, 93; 1863, 479.—Chiaje, 1833, 11.—Cobbold, 1879b, 15; 1883p, 401, fig. 16; 1883x, 514.—Cohn, 1902h, 880.—Cosmovici, 1887a, 128.—Crep., 1837a, 309–329; 1839a, 288.—Daldorf, 1793, 159 (syn. of Fasc. hep.).—Davaine, 1877a, 1xxiy.—Delafield & Prudden, 1897a, 130.—Dies., 1850a, 288, Davaine, 1877a, lxxiv.—Delafield & Prudden, 1897a, 130.—Dies., 1850a, 288, 293 (of Baer syn. of Rhopalocerca), 295 (of Fil. and Steenstrup syn. of Cerc.), 301 (of Baer syn. of Heterost. Fil.), 307 (of Rud. syn. of Hemist. Dies.), 318 (of Zed. syn. of ? Diplodiscus Dies.), 319 (of Zed. syn. of Monost. Zed.), 329 (of Ammon, syn. of Monost, lentis Nord.), 331-339 (syns. Fasc, Linn., Planaria Auct., Schisturus Rud., Alaria Blainv., Brachylemus E. Bl., Apoblema E. Bl.), 400 (of Bojanus syn. of Amphist, Rud. ex parte; of Rud. syn. of Rhopalophorus Dies.), 418 (of Henle syn. of Heptast. Otto Schomburgk), 573 (of Zed. syn. of Tetrabothriorhynchus Dies.), 609 (of Humboldt syn. of Pentast. Rud.); of Tetrabotinfornynchus Dies.), 609 (of Humboldt syn. of Pentast. Rud.); 1855a, 377, 380, 383 (of Fil., Sieb., & Steenstrup, syn. of Cercariæa), 384, 385, 396; 1858e, 312, 329 (syns. Fasc., Plan., Dist., Schisturus, Alaria, Brachylemus, Apoblema, Clinost.), 331.—Doyère, 1838a, 131–132; 1838b, 398–399.—Duj., 1845a, 381–388.—Dunglison, 1893a, 338, 710, 1174.—Eichwald, 1829a, 248.—Eiss, 1838a, 21.—Encyl. méthodique, Par., 1824, v. 2, 256–285.—Fabricius, 1794, 26; [1799a], 149.—Fil., 1837a, 337.—Fischer, 1840, 157.—Froriep, 1833, 428–432.—Gamb., 1896a, 73.—Goldb., [1855a], 17, 27.—Goto, 1891a, 158; 1893a, fig. 2.—Gunther, 1858, 205.—Gurlt, 1831, 370.—Hackley, (1886a), 518–519, figs. 880–885.—Hahn & Lefèvre, 1884a, 515–549; 1884, 806.—Hausman, 1897b, 16.—Henle, 1835a, 597.—Herdmann, 1890, 617 (Distoma of Hausmann, 1897b, 16.—Henle, 1835a, 597.—Herdmann, 1890, 617 (Distoma of Gærtner, 1890, 617, f. Distomidæ of Ascidiæ compositæ).—Hoyle, 1890, 535, 539, 540.—Huber, 1896a, 574 (from διστομος).—Jacoby, 1899c, 1–30, pls. 1–2, figs. 1–16; 1899d, 30 pp., 2 pls., figs. 1–16.—Jægers., 1901b, 979.—Joy, 1835a, 504.—Kajama & Nanba, 1892a, 32–36, 42–46 (Japan).—Kholodk., 1898a, 25, 26.—Knoch, 1862d, 19.—Kolenati, 1857, 11.—Kowal., 1895g, 1, 24 1898a, 25, 26.—Knoch, 1862d, 19.—Kolenati, 1857, 11.—Kowat., 1859g, 1, 24 (41, 64); 1896g, 70, 71.—Kuech., 1855a, 182.—Lamouroux, 1824a, 559–563.—Leblond, 1836e; 1836f, 4.—Leidy, 1850a, 301–310, pl. 43, figs. 1–16 (2 species); 1884a, 47-48.—Leuck., 1863a, 451, 461, 524, 528–530; 1879, 136; 1886d, 7, 34, 35, 62, 67, 105, 117, 204, 715.—Linst., 1879a, 165–188.—Lint., 1901b, 408.—Looss, 1885a (2 species); 1885b, 56; 1892a, 126; 1893b, 816, 817, fig. 1: 1896b, in 1–252, 16 pls.; 1899b (divided into various genera), 533, 535, 536, 537, 538, 539, 540, 542, 543, 545, 546, 556, 658; 1901b, 191, 192, 193, 195, 196, 197, 200, 210, 1904d, 602, 1902m, 701, 719 (in ref. to Manust, prismaticum), 720, 732 210: 1900d, 603; 1902m, 701, 719 (in ref. to Monost. prismaticum), 720, 732, 746, 750, 751, 752, 753, 756, 775, 795, 796, 812, 813 (as collective genus), 842, 865.—Meltzer, 1894a, 406–407, figs. 1–3; 1895a, 137.—Milner, 1858, 17.—de Miranda, (1903a), 137–139; 1904a, 280 — Miyake, 1894, 1–6, 1 pl.— Mlinarich, Miranda, (1903a), 137–139; 1904a, 280.—Miyake, 1894, 1–6, 1 pl.—Mlinarich, 1832, 13.—Moniez, 1896, 86, 89–90.—Mont., 1888 (thesis), 4, 7, 8, 9, 12, 22, 24, 31, 33, 38, 43, 50, 53, 54, 57, 64, 71, 83, 84, 89, 92, 104; 1889k, 132–134; 1889l, 612–613; 1892, 29; 1892, 687; 1892, Oct. 7, 189; 1893, 229, pl. 1; 1893, 427; 1894, Feb. 1, 16–21.—Moul., 1856a, 12, 15, 121.—Mueller, 1839, in 171–251, pls. 1–4.—Muller, 1850, 496.—Neumann, 1892, 345.—Nord., 1840, 544, 613, 614 (of Zed. syn. of Fasc.).—Odhn., 1902, 43.—Osborn, 1898, 301–310, figs. 1–4 (in Anodonta plana).—Perrier, 1903, 682.—Pontallié, 1853, 103–105 (2 species).—Pratt, 1900a, 645; 1902a, 889; 1902 (4 species).—Raf., 1815, 151 (of Zed. renamed Distomopsis).—Rail. & Marotel, 1898, 37.—Rathke, 1799, 69.—Rawitz, 1893, v. 1, 1694–1697.—Rapitsch, 1890, 14.—Rud., 1809, 6, pl. 5, figs. Rawitz. 1893, v. 1, 1694-1697.—Rentsch. 1869, 14.—Rud., 1809, 6, pl. 5, figs. 1-3, pl. 6, figs. 7-8, 21-22, 37-38, 352; 1819, 92, 362-363.—Schneidemuehl, 1896,

MA—Continued.
295, 296.—Schneider, 1866, 334.—Seely, 1906, Apr., 249–254, figs. 1–3 (2 species).—Setti, 1897, in 50 pp., 2 pls.; 1897, in 198–247, 2 pls.; ——, 49.—Shaw, 1901, 263, 809.—Sieb., 1836, 232–240 (as genus); 1854, 20, 29.—Simon, 1897, 259 (Distomi).—Sluiter, 1898, 1–64, pls. 1–7.—Sommer, 1880, 195–202, figs. 3–10; 1895, 116–123, figs. 15–22.—Sons., 1889, 276.—Staff., 1900, in 399–414 (5 species).—Steenstrup [1859, 167–170].—Stiles, 1901, 163, 164, 165, 172, 174, 194, 196; 1905z, 14.—Stiles & Hass., 1898a, 87, 88, 89.—Stoss., 1892, 42 pp.; 1892, 4, 5, 7, 12; 1898, 31; 1899, in 1–6, 1 pl.; 1903, 193–201; 1905, Jan. 31, 23.—Stowell, 1879, Jan., 85–93, pl. 9.—Tasch., 1879, 232, 233.—Valentin, 1811, 6.—Verrill, 1870, 172, 219, 220.—Veterinarian, Lond., 1897, v. 70, Oct., 522 (in the heart).—Villot, 1870, 9–13 (adults).—Vogt, 1878, 9.—Wagner, 1883, 120–122.—Ward, 1896, in 257–272, figs. 1–10.—Weltner, 1896, 199–200, 3 figs. (in Wasseriungfern).—Wolf, 1903, 610.—Zed., 1800a, 161–164.—Ztschr. f. (in Wasserjungfern).—Wolf, 1903, 610.—Zed., 1800a, 161–164.—Ztschr. f. Fleisch. u. Milchhyg., 1906, Nov., 66.

, anatomy of: Baer, 1828f, 197-198 (anus).—Chatin, 1882d, 200-202 (muscles).— Crety, 1892d, 21-26, figs. 1-2 (suckers, tactile organs); 1893a, 380-384 (suckers, tactile organs).—Jackson, 1888, 643, 644.—Linst., 1873e, 95–108, pl. 5, figs. 1–6 (new species and female organs); 1873f, 231–232 (sexual organs).—Macé, 1882, 91 pp., 3 pls. (grande douve du foie).—Nardo, 1827, v. 1, 68–69 (anus). biology of: Braun, 1890d, 568.—Brown, 1882b, 624–630 (liver-fluke).—Harms, 1891a, 249–250 (Aufnahme).—Pontallié, 1851, 217–219 (encysted adults).—Schauinsland, 1882, Nov. 25, 494–498, pls. 19–20 (embryology).—Wymann,

1851, v. 6, 65.

classification of: R. Bl., 1885a, 541; 1891p, 609–611.—Duj., 1845a.—Looss, 1899b, 521–784, pls. 24–32; 1900a, 390–401; 1900b, 458–466; 1901b; 1901c; 1901, Feb.—Luehe, 1899k, 524-539; 1900p, 305-306.—Mont., 1892, Oct. 7, 214.— Stiles & Hass., 1898a.

, geographic distribution of: Billet, 1898a, 279–282, figs. 22–23 (Haut-Tonkin). Cobbold, 1876h, 209–212 (Asia).—Johnston, 1901a, 334–338, pl. 22, figs. 1-4 (Australia); 1901b, 598 (Aust.); 1902a, 326-330, pl. 13, figs. 1-7 (Aust.); 1902b, (Aust.); 1903a, 899 (Aust.).—Lint., 1900a, 267-304 (in fish at Woods Hole); 1901a, 267-304 (Woods Hole); 1901b, 405-492 (Woods Hole); 1902a, 449-450 (Woods Hole).—Looss, 1900a, 390–401 (Egypt); 1900b, 458–466 (Egypt).—Mochizuki & Tsutsumi, 1899, Nov. 20, 13–24 (Shigaken).

-, in collections: Stoss, 1904, June 23, 1-14, pl. 2, figs. 1-3 (Naples); 1905, Jan. 31, 23-24 (Naples).

-, in various animals: Bavay, 1902a, 199-200, 1 fig. (sp. in Rhizostoma cuvieri); 1905, July, 63.—Heymann, 1905, 97-98, fig. B (Kachuga tectum).—Weltner, 1896, 199-200, 3 figs. (Libellulidæ).

-, in amphibia: Groenouw, 1898a, 60-62, 1 fig., 85-92 (eye of Rana esculenta); in amphibia: Groenouw, 1898a, 60–62, 1 fig., 85–92 (eye of Rana esculenta); 1903a, 65 (frog).—Guenther, 1853a, 95–99, pl. 1, figs. 1–6 (in Rana temporaria).—Klein, 1905, 59–80, pl. 5, figs. 1–8 (Rana hexadactyla); 1905, 24 pp. (R. hex.); 1906, Jan. 30, 41–42 (R. hex.).—Looss, 1894a, 296 pp., 192 figs. (and fish); 1894b.—Luehe, 1901p, 166–177 (Indian Anura); 1901q, 658 (Indian Anura).—Schellenberg, 1895, 170–171, figs. 1–2 (frog: muscle); 1895, Aug. 24, 183.—Staff., 1903, Oct. 21, 822–830, 1 pl., figs. 1–5 (Canadian Urodela); 1904, May 17, 281 (Canadian Urodela).—Stoss, 1889, v. 11, 60–74; 1889, Oct., 521; 1889, Nov. 8, 581–582.—Vallada (1882), v. 29, 35–39 (crustacea).—Vulpian, 1859, 150–152 (frogs); 1860, pl. 11, fig. 4 1859, 150-152 (frogs); 1860, pl. 11, fig. 4.

-, in aves: Braun, 1899a, 1–4 (in Porphyrio).—Linst., 1906, 174 (in Plotus melanogaster).—de Miranda Ribeiro, 1903a, 137–139; 1904a, 280.—Nicoll, 1906, 515 (Colymbus septentrionalis).—Rail., 1890, 131 (pigeon).—Spencer, 1889, 109–110 (egg of fowl).—Stoss., 1892, v. 13 (2), 143–196; 1892, 54 pp.

-, in fishes: Ariola, 1899, 129–138 (marine).—Catois, 1897, xxxiii (meninges of in fishes: Ariola, 1899, 129–138 (marine).—Catois, 1897, xxxiii (meninges of Gadus).—Chavannes, 1851a, 210 (Corregonus fera).—Johnston, 1902a, 326–330 (Pristiophorus cirratus); 1905, July, 57 (Pr. cir.).—Kroyer, 1838–1840a, 612 (Rhombus vulgaris); 1843–1845a. 517 (Cyclopterus lumpus): 1852–53a, 745 (Orthagoriscus mola).—Lint., 1900a, 267–303; 1901a, 267–304; 1901b, 405–492; 1902a, 449–450; 1905, 349, 350, 353, 359, 360, 361, 364, 366, 372, 373, 374, 382, 385, 389, 393, 397, 402, 403, 404, 410, 413, 414, 415, 416, figs. 167, 168, 169, 171, 172, 173, 179, 198, 199, 205, 208, 209, 210, 213, 214, 215.—Looss, 1894a, 296 pp., 192 figs. (and frogs): 1894b, 706–711 (and frogs); 1901d, 398–405, 437–442, figs. 5–6 (Labriden; Triest).—Luehe, 1900w, 504–509 (gall-bladder;



298 (of Bojanus syn. of Cerc. helicis viviparæ sp. inq.: in Paludina vivipara).—
Dubois, 1903, 178–179 (in Mytilus gallo-provincialis).—Huet, —, 20 pp.,
1 pl. (Cardium edule).—Leidy, 1847c, 220–221 (Helix alternata).—Osborn,
1898, 301–310, figs. 1–4 (Anodonta plana; Chautauqua, N. Y.).

——, in insects: Ruge, 1904, June 30, 520; 1904, 174–176; 1905, Apr. 15, 420 (Anopheles maculipennis).—Schoo, 1902, Feb. 8, 283–286, figs. 1–2 (in Anophelaviger); 1902, June 17, 358–359; 1905, July 1, 63.

- in mammalia: Braun. 1893e, 347–355 (cat); 1893f, 381–392, 422–428; 1894, 691–696; 1900f, 387–391 (Chiroptera).—Cobb, 1897a, 453–481, 1 pl., 26 figs. (sheep).—Duncker, 1881a, 23–25, 55, 154, 159–160; 1884a, 39–42 (muscle, hogs).—Erc., 1875a, 391–441, 1 pl., figs. 1–9 (dogs); 1875b, 88–91 (dogs); 1875c, 413–416 (dogs); 1875e, 33–40 (dogs); 1875f, 274–279 (dogs); 1875g, 254–255 (dogs); 1876a, 379 (dogs).—Galli-Valerio, 1893a, 173–182, pl. 2, figs. 1–6 (embolism, horse); 1895c, 266 (horse).—[Hoppen, 1881a, 39 (muscle, hog).]—Johnston, 1901a, 334–338, figs. 1–4 (Platypus); 1901b, 598.—Hutcheon, 1900i, 497 (Duiker antelope).—Katsurada & Saito, —, v. 39 (3), 506; 1906, Nov. 13, 2264 (pancreas. cattle).—Leuck., 1881b, 46 (muscle, hog).—Marchi, 1873, 304, pl. 5, fig. B (Delphinus tursio).—Meltzer, 1894a; 1895a (in lungs of cattle).—Morot, 1889, Jan. 30, 37 (calf).—von Ratz, 1893, 249 (horse).—Rec. d. méd. vét., Par., 1885, Jan. 15, 60–62 (pork).—Rivolta, 1884, v. 16, 20–28, pl. 1 (cat and dog).—Sons., 1895, 1157–1160 (Carnivora); 1896, v. 20, 709.—Stoss., 1892, 1–42.—Tright, 1885, Apr., 84–85 (dog).—Villot, 1875, 467 (Delphinus delphis).—Welsch, 1881, June 15, 97 (pork).—Willach, 1892, Apr., 131 (lungs, horse); 1892, 118–124 (horse); 1892, 239–241 (bull); 1893, Sept. 1, 289 (horse); 1893, v. 8 (6–7), 80 (cattle).
- ----, in man (Homo): Askanazy. 1904, May 28, 897; 1904, July. 210; 1904, July 1, 74-75.—R. Bl., 1888a, 631.—Braun, 1894i. 602-606.—Lankester, 1857b, 433-437.—Mueller, 1842, v. 2, 559; 1842. 85 (fœtus: spinal marrow).—Poir. 1888, v. 8.1, 49.—Sons., 1895, 1157-1160; 1896, v. 20, 709.—Tommasi (1881, fasc. II).—Valentin, 1840, 317-319 (fœtus: spinal canal).
- in reptiles: Heymann, 1905, v. 22 (1-2), 81-100, figs. 1-5 (Chelonien); 1905, 25 pp.; 1905, 96-97. fig. a (sp. in Dermatemys mavii); 1906, Jan. 30, 41.—
 Luehe, 1900aa, 555-566 (snakes and lizards); 1900bb, 928-930.—Sons., 1892 (Zamenis viridiflavus).—Stoss., 1895, v. 16, 213-239.—Voltz, 1899, Oct., 231-240, pl. 20 (snakes).
- ----, larvæ of: Calkins, 1901b, 8, 12.
- ——, reproduction of: Ben., 1858d, 858 (fertilization); 1858e, 159–160, 223–224; 1858h, 312–314.—Fielde, 1888a, 115.—Linst., 1904p, 252–254, figs. 1–4; 1906, Jan. 30, 42.—Ward, 1903, 864.
- species of: Braun, 1892a, 761; 1893a, 826.—Lint., 1898, 540 (larva), pl. 53, figs. 12-13, pl. 54, fig. 1; 1898, 537-538, pl. 53, figs. 1-2; 1900, 269, 295, 296, pl. 39, figs. 71, 72-81.—Linst., 1906, 174.—Looss, 1899, 521.—Ssinitzin, 1904, 768, fig. c (n. g.).—Staff. 1902, 481.—Wedl, 1857, 247-248, pl. 1, fig. 8.—Weltner, 1896, 199-200.—Wolffluegel, 1900, 36.

acanthocephalum Stoss., 1887, 94. pl. 10, fig. 40 (in Belone acus; Triest).—Braun, 1892a, 583, 584, 736; 1893a, 874, 911.—Looss, 1899b, 578, 580 (to Tergestia).—Mont., 1893, 102.

acanthoides Rud., 1819a, 114, 415–416 (in Phoca vitulina; Berlin).—Braun, 1901e, 315–316.—Cobbold, 1860a, 33 to Echinost.; 1879, 313.—Dies., 1850a, 382.—Duj., 1845a, 424.—Stoss., 1892, 29 (to Echinost.).

acervocalcaphorum Erc., 1881e, 21 (for acervocalciferum); 1882a, 257.

acervocalciferum Gastaldi, 1854, 6–7, pl. 1, figs. 6–9 (in Rana esculenta).—Braun, 1893a, 870.—Cobbold, 1860a, 18.—Dies., 1858e, 340–341 (in Pelophylax esculentus).—Linst., 1875, 193.—Rizzo, 1902, 29–30, fig. 2 (in Tropidonotus natrix).—Stoss., 1889, 68.

acervocalciferum ranæ esculentæ Gastaldi, see Dies., 1855, 64, footnote 11.

acervocalcoforum Linst., 1875a, 193 (for acervocalciferum).

acervocalcophorum Erc., 1881e, 20; 1882a, 256 (for acervocalciferum).

actxonis Pag., 1862, 306, pl. 29, fig. 5a (in Acteon viridis; Cette).

aculeatum Nitzsch, in Giebel, 1857, 266 (in Strix bubo).—Braun, 1901a, 14.— Stoss., 1892, 177 (in Bubo maximus).

acutum Leuck., 1842, 33–34, pl. 1, figs. 7a-b (in Mustela putorius).—Braun, 1893a, 877, 881; 1893d, 467; 1901e, 324.—Cobbold, 1860a, 8 (November).— Dies., 1850a, 364.—Duj., 1845a, 439.—Moniez, 1890e, Mar. 1, 242; 1890f, 1; 1890g, Apr. 29, 542–543.—Stoss., 1892, 34 (in Putorius communis).

[adriaticum von Dr., a tunicate.]

aduncum Lint., 1905, 327, 333, 409, figs. 195-197 (in Opeanus tau; Beaufort, S. C.).

advena (Duj., 1843) Braun, 1892a, 772 [type of Brachylaima 1843]; 1893a, 831, 864,
 880, 894.—Gamb., 1896a, 71.—Reported for Sorex araneus, Limax.

xglefini Ben., 1870, 57.—Braun, 1892a, 728, 734, 759.—Linst., 1873e, 99.—Mont., 1893, 193.—Reported for Gadus euxinus, G. æglefinus, G. morrhua, Merlangus vulgaris.—[See also next entry.]

æglefini (Mueller, 1776) Zed., 1803a, 211 (in Gadus æglefinus; int.).—Dies., 1850a, 343 [syn. of D. simplex Rud.].—Nicoll, 1907, 73.—Rud., 1809a, 370 (renamed D. simplex).—[Type of Sinistroporus, 1904.]

xgyptiacum Looss, 1896b, 33-36, 192, 196, pl. 3, fig. 16, pl. 11, figs. 117-118 (to Fasc.).

æquale Duj., 1845a, 410 (in Strix flammea) to (Brachylaimus).—Braun, 1892a, 767; 1901e, 341.—Cobbold, 1859d, 365 (in Strix perlata); 1860a, 14 (in S. perlata); 1861, 118; 1879, 447.—Dies., 1850a, 363.—Looss, 1899, 650 (perhaps a Clinost.).—Mont., 1893, 155.—Stoss., 1892, 174 (to Mesogonimus).

affine Dies., 1850a, 359 (in Lampris guttatus; Groningæ) [nec Rud., 1819a], 680 (renamed dicorynum) (Monost. tenuicolle Rud., 1819, renamed [not Dist.

tenuicolle Rud., 1819a]); 1859c, 430.

affine Rud., 1819a, 110, 406 (in Perca cirrosa; Arimini) [nec Dies., 1850a] (to Derogenes).—Baird, 1853a, 54 (D. appendiculatum Rud.?).—Carus, 1884, 131.—Cobbold, 1860a, 27 (in Scorpæna cirrhosa).—Dies., 1850a, 371 [nec Dies., 1850a, 371]. 359].—Luehe, 1901, 479.—Odhn., 1905, 364.—Stoss., 1886, 44.—Wagener, 1860, 190 (in Perca cirrhosa).

agamos Linst., 1872, 1-5, pl. 1, figs. a-c (in Gammarus pulex); 1877, 185; 1878a, 315 (in Gam. pulex L.).—Jackson, 1888, 648 (in G. p.), 652.—Looss, 1885b, 36;

1894, 175.

alacre Looss, 1901, 401–402, 403, fig. 2 (in Labrus maculatus, L. merula, Creni-

labrus pavo, C. quinquemaculatus, C. griseus).

alatum (Geeze, 1782) Zed., 1800a, 177-180; 1803a, 213.—Anacker, 1892c, 94.— Blainv., 1824a, 519 (a holostome).—Brand., 1888a, 9, 60 (to Hemist.).—Crep., 1829, 66–67; 1837, 310; 1839, 287.—Dies., 1850a, 308 (to Hemist.).—Fischer, 1840, 158.—Gurlt, 1831, 375–376, pl. 8, figs. 39–40.—Lamouroux, 1822a, 194; 1824a, 563 (Distome ailé).—Mehlis, 1831, col. 184.—Nitzsch, 1819, 399 (to Holost).—Nord., 1840, 628 (Holost. alatum).—Olfers, 1816, 46.—Rud., 1809a, 400, 403, 404 (included Physical Court of Taylor alatic properties of the court of 400, 402–404 (includes Plan. alata Gœze, 1782; Alaria vulpis Schrank, 1788 [type of Alaria]; Fasc. vulpis Gmelin, 1790; Dist. vulpina Abild., 1790; Festucaria alata Schrank, 1790; Fasc. alata Rud., 1793); 1819a, 112, 412–413.—Also reported for Canis lupus and C. vulpes.

albicolle Rud., 1819a, 98-99, 376-377 (in Falco pennatus; Mus. Vien.).—Braun, 1893a, 875; 1901, 561, 562; 1902b, 99 (to Dicrocœlium).—Bremser, 1824, pl. 9, figs. 3-4.—Cobbold, 1860a, 12.—Crep., 1837, 310.—Dies., 1850a, 348.—Duj., 1845a, 393.—Rail., 1900, 239, 240.—Stoss., 1892, 156 (D. macrourum Rud.).—

Also reported for Aquila pennata.

albidum Braun, 1893e, 347-355 (to Dicrocælium) (in Felis domestica); 1893f, 390, 391, 392, 424, 427, fig. 2; 1894g, 129; 1894i, 606; 1895b, 151.—Askanazy, 1900b, 498, 501; 1901, 72; 1906, 128 (in cats fed on fish; intermediate host). de Jong, 1896a, 3, 4.—Looss, 1899b, 564-565 (type of Metorchis), 677.—

Moniez, 1896, 139, 140.—Mueh., 1898, 15, 23 (in Halichærus grypus), 87, 88, 89 (cf. D. crassiusculum).—Ratz, 1900, 141.—Stiles & Hass., 1894e, 424–425, pl. 1, figs. 1–2.—Ward, 1895, 341.

albocæruleum Stoss., 1889, 28–29 (in Sargus salviani; Triest); 1898, 37.

album Stoss., 1890, 42, pl. 16, fig. 73 (in Cantharus orbicularis; Triest); 1898, 49; 1901, 94.—Braun, 1891d, 424 (in C. orb.).—Looss, 1899, 571 (may belong to Creadiinæ, possibly to Creadium).—Mont., 1893, 85, 86, 94, 102.—Odhn., 1905, 328.—Also reported for Cantharus vulgaris.

allostomum Dies., 1850a, 367 (in Tropidonotus natrix var.) (includes D. colubri murorum Rud., 1819a).—Braun, 1893a, 863.—Cobbold, 1860a, 20.—Erc., 1881e, 67, 68, 69, 70, 73; 1882a, 303, 304, 305, 306, 309 (larva in Helix carthusianella).—Mont., 1893, 187.—Par., 1894, 147.—Stoss., 1895, 229.—Also reported for Natrix torquata, Tropidonotus viperinus.

aloysix Stoss., 1885, 161, pl. 6, fig. 28 (in Corvina nigra; Triest); 1886; 24; 1898, 56; 1899, 15.—Braun, 1892a, 567, 663, 737, 1902b, 31.—Looss, 1899, 581.

altemon Ben., 1870, 48, for atomon.

aluconis intestinale Rud., 1819a, 119 (in Strix aluco).—Dies., 1850a, 396.—Duj. 1845a, 442.—Stoss., 1892, 173, syn. of Echinost. apiculatum (Rud.) Cobbold.

aluconis thoracicum Rud., 1819a, 119 (in Strix aluco).—Dies., 1850a, 396.— Duj., 1845a, 442.—Stoss., 1892, 177.

americanum (Hass., 1891) Stiles, 1892e, 148; 1892f; 1892g, 732–733; 1898a, 51 (syn. of Fasc. magna).

amphileucum Looss, 1896b, 55–60, pl. 4, figs. 31–35 (in Naja haje; Alexandria, Egypt); 1899b, 565 (to Metorchis).

amphiorchis Braun, 1899b, 719 (in Thalassochelys corticata at Triest; Chelone mydas; Podocnemis expansa); 1899e, 629; 1901b, 20, 36.—Looss, 1899b, 568, 569 (type of Anadasmus); 1902, 463 (to Orchidasma).

amphistoma Nord., 1840, 616 [probably lapsus].

amphistomoïdes Bojanus, 1817b, 270–277, pl. 9, figs. 1–6 (in Castor fiber) (Dystoma).—Baird, 1853a, 43 (=Amphist. subtriquetrum Rud.).—Dies., 1836, 248; 1850a, 402 (syn. of Amphist. subtr.).—Fischder., 1902a, 42 (syn. of Cladorchis (Stichorchis) subtriquetrus).

ampullaceum Buttel-Reepen, 1900a, 586-596, 597, 598, figs. 1-7 (in Cetacean;
Indian Ocean); 1902, Dec. 8, in 165-236; 1902, pl. 6, fig. 25; pls. 7-10, figs.
27-53; text figs. b-g; 1904, Jan. 26, 24-25; 1905, July, in 52-53.—Darr, 1902, 698.

anarhichæ Jacoby, 1900, 13, see anarrhichæ.

anarrhichæ Rud., 1819a, 121–122 (D. anarrhichæ lupi Rathke, from intestine, renamed).—Dies., 1850a, 398 (possibly syn. of D. appendiculatum).

anarrhichæ lupi Rathke, 1799, 70, 146, pl. 2, figs, 3, a, b (in Anarrhichæ lupus).— Dies., 1850a. 339, from stomach (syn. of D. incisum Rud.).—Jacoby, 1900, 12.—Rud., 1809a, 361, from stomach (syn. of D. incisum), 435—436 (see anarrhichæ).

anatis (Schrank, 1788) Zed., 1800a, xvii, 164, 196–198.—Dies., 1850a, 383 (syn. of D. echinatum Zed.).—Looss, 1899b, 680.—Rud., 1809a, 418 (syn. of D. echinatum Zed.).

anatis domesticæ Rud., 1809a, 431–432 (Hirudo fasciolaris Mueller, Fasc. anatis Bruguière, 1791, renamed); 1819a, 121.—Dies., 1850a, 335 (syn. of D. ovatum Rud.).

anatis fuscæ Viborg, 1795, 243 (at Copenhagen).—Dies., 1850a, 397–398.—Dij., 1845a, 450.—Knoch, 1862e, 104.—Rud., 1809a, 431; 1819a, 120–121.—Stoss., 1892, 167, syn. of Echinost. echinatum (Zed.) Cobbold.

anceolatum Braun, 1892a, 677 (for lanceolatum).

anceps Mol., 1859, 845–846 (in Fulica atra; Padua).—Stoss., 1892, 168 (to Echinost.).
ancyli lacustris Dies., 1855a, 400 (to Cercariæum), based on Baer, 1827b, 656 (in Ancylus lacustris).

andersoni Cobbold, 1876, 46, pl. 10, fig. 3 (in Platanista gangetica); 1879, 420.— Linst., 1886, 125.—Stoss., 1892, 19 to (Brachylaimus).

anguillæ (Gmelin, 1790) Zed., 1803a, 222, in Anguilla.—Dies., 1850a, 340 (=? D. polymorphum Rud.).—Nord., 1840, 618 (to Fasc.).—Rud., 1809a, 363 (syn. of

D. polymorphum); 1814a, 101; 1819a, 369 (of Abildgaard, as ? syn. of D polymorphum).—Thompson, 1844, 439 (in Anguilla conger).

anguillulæ Dies., 1850a, 340 (syn. of D. polymorphum), lapsus for anguillæ.

anguis Linst., 1885, 250–251, pl. 15, fig. 27 (in Anguis fragilis).—Braun, 1892a, 642, 663, 693.

angulatum Duj., 1845a, 401–402 to (Podocotyle [type]) (in anguille) [not Fasc. angulata Mueller, 1776].—Braun, 1893a, 910.—Cobbold, 1860a, 29 (in Anguilla vulgaris).—Dies., 1850a, 379.—Kroyer, 1846–1853a, 641 (in Ang. migratoria).—Looss, 1902m, 757, 770, 771 (possibly only one testicle), 772, 827 (type of Podocotyle).—Luehe, 1900, 487, 491, 492.—Odhn., 1905, 320, 321 (syn. of Pod. atomon).—Stiles, 1901r, 193.—Stiles & Hass., 1898a, 92, 93, 97 (type of Pod.).—Stoss., 1886, 17; 1902, 582.

angusticolle Hausmann, 1896a, 391–392 (in Cottus gobio; Basel); 1897b, 4, 6, 20,
22, 24–29, 31, pl. 1, figs. 1–3.—Looss, 1899b, 571 (to Creadium: possibly identical with D. commune Olss., 1868, 31).—Odhn., 1901, 483, 502.

angustum Staff., 1900, 407-408, fig. 6 (in Chrysemys picta).

aniarum Leidy, 1891a, 414 (in Tropidonotus sipedon).

annulatum Dies., 1850a, 386–387 (in Gymnotus electricus; Brazil); 1855, 67–68, pl. 3, figs. 18–21; 1858e, 347.—Braun, 1892a, 571, 584.—Cobbold, 1860a, 36 (to Echinost.).—Looss, 1899b, 595.—Mont., 1888a, 14.—Stoss., 1886, 36.

annuligerum Nord., 1832a, 43, 53–54, 55, 102, pl. 1, figs. 4–10 (in Flussbarsch).— Braun, 1893a, 871.—Cobbold, 1860a, 28; 1879, 458.—Crep., 1837, 310, 313, 326.—Dies., 1850a, 377.—Duj., 1845a, 455.—Gescheidt, 1833a, 431.—Kroyer, 1838–1840a, 21 (in Perca fluviatilis).—Leuck., 1863a, 526.—Moul., 1856, 218–219 (in eye of perch).—Steenstrup, 1842, 59.—Stoss., 1886, 44 to (Brachylaimus).

anonymum Dies., 1858e, 341 (in Gadus æglefinus, G. euxinus, Merlangus vulgaris, M. carbonarius; Ireland) (includes D. Gadi æglefini, Merlangi vulgaris, et M. carbonarii based on Bellingham, 1844a, 428).—Cobbold, 1860a, 25.—Stoss., 1886, 45.

anthos Braun, 1899b, 720 (in Cheloniæ; Yedo); 1901b, 27–29, 33, figs. 20–22, 24,
31.—Looss, 1899b, 575 (appears to be closely related to Echinost.); 1901l, 565,
566; 1902m, 458, 462 (type of Calycodes).

apertum Rud., 1819a, 108, 400–401, 779 (in Apogon ruber =Mullus imberbis; Naples).—Cobbold, 1860a, 27.—Dies., 1850a, 370.—Duj., 1845a, 422 to (Apoblema).—Mont., 1891, 522.—Stoss., 1886, 45.—Wagener, 1860, 189 (in Mullus imberbe).

apiculatum (Rud., 1803) Rud., 1809a, 423, to (Echinost.) includes D. stridulæ, 1801 (in Strix stridula, S. flammea); 1819a, 116, 119.—Braun, 1893a, 874.—Cobbold, 1860a, 35 (to Echinost.) (in Strix aluco, S. flammea).—Crep., 1837, 311.—Dies., 1850a, 386 (in Strix aluco, S. flammea; Gryphiæ).—Duj., 1845a, 425-426.—Giebel, 1857, 266.—Looss, 1899, 703.—Olfers, 1816, 46.—Stoss., 1892, 173 (to Echinost.).

apodis Packard, 1882, 142, fig. 1 (in egg sacks of Apus).

apolaimum Heymann, 1905, Mar. 25, 91–94, pl. 6, figs. 4–5 (in Kachuga tectum (Gray), small intestine); λαιμός, Schlund.

appendiculata Leidy, 1877, 202 (in Helix arborea) [nec Rud., 1802]; 1891a, 416 (syn. of D. centrappendiculatum).

appendiculatum (Rud., 1802) Rud., 1808a, xxiv [descr. of pls.], pl. 5, fig. 2 [nec Frælich, 1802]; 1809a, 387, 400-401, 405, 407, 438, pl. 5, fig. 2; 1819a, 110, 122, 404-406.—Baird, 1853a, 54.—Bellingham, 1844a, 425.—Ben., 1858a, 1861a, 178, 189; 1870, 16, 30, 66.—Braun, 1891d, 423 to (Apoblema); 1892a, 705; 1893a, 853, 864, 879, 911; 1893d, 468; 1902b, 124.—Cobbold, 1858b, 158; 1860a, 20.—Crep., 1837, 310, 313, 326; 1839a, 288.—Dies., 1850a, 342, 370-371, 398 (includes Apoblema ap. Bl., Fasc. alosæ Hermann, F. clupeæ Schrank, F. ap. Rud., F. crenata Rud., Dist. clupeæ Zed., D. clupeæ rhenanæ Rud., D. crenatum Rud., D. varium Eysenhardt); 1858d, 268 (larva Histrionella echinocerca Dies.); 1858e, 342 (in Acipenser sturio, Ammodytes lancea); 1859c, 431-432.—Duj., 1845a, 420-421 to (Apoblema).—Fil., 1855b, 19.—Fraip., 1880b, 106; 1881b, 4.—Gamb., 1896a, 71.—Hausmann, 1897b, 4, 6, 20, 22, 38-39, pl. 1, figs. 11-12 (in Trutta salar).—Johnstone, 1907, 180-182, fig. 14 (in Gadus merlangus; Shoals).—Juel, 1889, 4, 7.—Kroyer, 1838-

1840a, 187, 578, 582, 583, 595, 605, 609, 611; 1843–1845a, 444, 488, 581; 1846–1853a, 218, 253, 641; 1852–1853a, 778, 1219 (in Gasterosteus aculeatus L., Perca fluviatilis Linn., Trigla hirundo Linn., Cottus scorpius Linn., Scomber scombrus Linn., Gadus morthua Linn., Lota yulgaris Cuv., Platessa flesus Linn.. Rhombus maximus Linn., Solea vulgaris Čuv., Salmo salar Linn., Alosa finta Cuv., Esox lucius Linn., Anguilla migratoria Kroyer, Acipenser sturio Linn., Clupea harengus Linn.)—Lander, 1904a, 14, to (Hemiurus).—Leuck., 1863a. 453, 454, 481.—Levin., 1881a, 9, 58-59, 59 (Rud. of Olsson, 46, syn. of D. a. Rud. Mol.), 61, 64 (includes D. ventricosum Wagener, D. ap. Rud. of Olsson) (in Cottus scorpius, Gadus ovak).—Linst., 1903, 354.-Lint., 1900, 269, 283, 289, pl. 36, figs. 25–26; 1901, 408, 415, 418, 437, 439, 440, 445, 449, 459, 460, 467, 471, 475, 478, 482, 486, 487, figs. 312–314; 1905, 328, 333, 352, 365, 374, 378, 382, 393, 404, 405, 415, figs. 152, 153, 160 (in Brevoortia tyrannus, Caranx hippos, Coryphæna equisetis, Lagodon rhomboides, ta tyrannus, Caranx hippos, Coryphæna equisetis, Lagodon rhomboides, Leiostomus xanthurus, Lophopsetta maculata, Orthopristis chrysopterus, Prionotus scitulus, P. tribulus).—Looss, 1894a, 204; 1899b, 640 (type of Hemiurus).—Luehe, 1901n, 394, 396, 397, 398, 399, 401.—Mayer, 1841a, 17–18, pl. 3, fig. 12.—Mol., 1858, 289 (in Alausa vulgaris: Patavii); 1859, 825–826 (in Anguilla vulgaris; Padua); 1861, 204–205, pl. 2, fig. 3.—Mont., 1888, 17, 38, 43; 1888a, 198; 1891, 9 (to Apoblema); 1891, 496, 497, 498, 499, 500, 501, 502, 522 (to Apoblema); 1893, 123.—Nicoll, 1906, 525.—Olfers, 1816, 46.—Olss., 1868, 46; 1876, 20; 1893, 11.—Sieb., 1842, 365.—Staff., 1902, 895.—Stiles & Hass., 1898a, 90.—Stoss., 1883, 115; 1885, 159; 1886, 47; 1886, 13; 1887, 90; 1887, 184; 1890, 40; 1902, 582.—Wagener, 1860, v. 1, 165–190, pls. 8–9.—Will.-Suhm., 1871 (3), 25 July, 380–396.—Reported also for Accip pls. 8-9.—Will.-Suhm., 1871 (3), 25 July, 380–396.—Reported also for Accippenser sturio, Achirus fasciatus, Alausa finta, A. vulgaris, Alosa vulgaris, Anguilla vulgaris, Brosmius brosme, Capros aper, Centropages hamatus, Citharus linguatula, Clupanodon pseudohispanicus, Clupea alosa, C. sprattus. Coregonus oxyrrhynchus, Cottus bubalis. Cynoscion regalis, Decapterus macarellus, Gadus æglefinus, G. callarias, G. euxinus, G. melanostomus, G. minutus, G. morrhua, G. ovak, G. polachius, G. virens, Gobius jozo, Hippoglossus maximus, Labrax lupus, Lichia amia, Lophius piscatorius, Lota molva' Lucullus acuspes, Merlangus pollachius, M. vulgaris, Microgadus tomcod, Molva vulgaris, Myxocephalus æneus, Paralichthys dentatus, Pomolobus mediocris, P. pseudoharengus, Prionotus carolinus, Pseudopleuronectes americanus, Saurus griseus, S. lacerta, S. saurus, Stenotomus chrysops, Stotenophorus brownii. Torpedo marmorata, Trachurops crumenophthalmus, Urophycis chuss.

appendiculatum Rud., 1819a, 404, e. p.—Odhn., 1905, 352 (syn. of Brachyphallus crenatus).

appendiculatum Rud. of Olss., 1868.—Levin., 1881, 59 (p. 46 Olss., syn. of D. a., Rud. of Mol.).—Odhn., 1905, 348 (syn. of Hemiurus levinseni; includes D. a. "(e. p. ?) Levin., 1881, 58"), 351 (Ölss., 1868, pl. 5, fig. 95, syn. of Hemiurus communis Odhn.; includes also D. a. Juel, 1889), 352 (Olss., 1868, e. p., syn. of Hemiurus lühei Odhn.).

aquilæ Leidy, 1887b, 24 (in Haliaëtos leucocephalus); 1904a, 197.—Braun, 1893a,
 876; 1900h, 16, 42.—MacCallum, 1899, 706, 707.—Stiles & Hass., 1894, 414.

arcanum Nickerson, 1900, Oct., 811–815, fig. 1 (in frogs).—Staff., 1902, 482, to
 (Pleurogenes) (includes D. medians Olss., of Staff., 1900); 1902, 724; 1905,
 Apr. 11, 683 (type of Loxogenes).

arcuatum Duj., 1845a, 410–411 (in Corvus glandarius; Rennes) to (Brachylaimus).— Braun, 1901e, 341.—Cobbold, 1860a, 15.—Dies., 1850a, 389.—Looss, 1899b, 703.—Stoss., 1892, 151.—Wolffhuegel, 1900, 9, 24, 37, 38, 39, 40, 41, 42.—Reported also for Corvus corone, Garrulus glandarius.

ardeæ (Gmelin, 1790) Zed., 1803a, 222.—Dies., 1850a, 388 (syn. of D. ferox Zed.).— Rud., 1809a, 432.

ardex minutx Pontallié (1853), to (Cladocalium).—Dies., 1855, 64, foot-note 5, to (Cladocalium).

ardex nigræ Viborg, 1795, see Rud., 1809a, index (D. hians Rud.).

ardex stellaris Rud., 1809a, 432–433 (Fasc. ardex Gmelin, 1790, renamed); 1819a,
 120.—Dies., 1850a, 388 (syn. of D. ferox Zed.).—Duj., 1845a, 447.—Stoss.,
 1892, 165, syn. of Echinost. ferox (Rud.).

arenula Crep., 1825a, 53-54 (in Fulica atra); 1837, 317.—Braun, 1902b, 155, 156, 157, fig. 99.—Cobbold, 1860a, 15.—Dies., 1850a, 364-365.—Duj., 1845a, 447.—Linst., 1887, 104.—Stoss., 1892, 177.

sura, Micropogon undulatus, Orthopristis chrysopterus, Sciænops ocellatus).-Mont., 1893, 193.—Olfers, 1816, 46.—Stoss., 1886, 43, 60 to (Echinost.).— Wagener, 1860, 185.

aristotelis Stoss., 1892, 14-15 (D. chilostomum Mehlis of Ben., 1873b) (in Rhinolophus hippocrepis, Vespertilio murinus, V. desycneme, V. daubentonii, V. emarginatus, V. mystacinus, Nannugo pipistrellus).—[Ben., 1873, 27.]—Braun, 1900, 221, 223, 388.—Stoss., 1892, 14, to (Brachylaimus).

armatissimum Linst., 1903, 280, fig. 15 (in Iguana sp.).

armatum (Rud., 1793) Zed., 1803a, 220.—Caruccio, 1886, 293.—Dies., 1850a, 382 (syn. of D. trigonocephalum Rud.).—Rud., 1809a, 416.—Stoss., 1892, 167

(Echinost. echinatum (Zed.) Cobbold).

armatum Mol., 1858, 130 [nec (Rud., 1793) Zed., 1803] (in Phasianus gallus; Patavii); 1861, 217.—Anacker, 1887c, 513 (in Gallus dom.).—Braun, 1893a, 874 (in Gallus dom.).—Dies., 1858e, 347.—Hass., 1896a, 2, 3 (syn. of Echinost. echinatum (Zed.).—Landois, 1882, 23.—Schneidemuehl, 1896, 303 (in chickens).—Sons., 1889, 11 (in Gallina).—Stoss., 1898, 52.

armatum MacCallum, 1895, 401 (as variety of D. isoporum).

armatum paludinæ impuræ Fil., 1857c, pl. 2, figs. 14, 15 [doubtless equals D. palu-

dinæ impuræ armatum].

arrectum Duj., 1845a, 403 (in Lézard vert), to (Brachycœlium).—Cobbold, 1860a, 20.—Dies., 1850a, 389–390 (in Lacerta viridis; Rhedoni).—Linst., 1873, 101; 1879, 185.—Looss, 1899b, 567, 568, 611, 614; 1902m, 816.—Luehe, 1899k, 536.—Mol., 1859, 831–833 (in Lacerta viridis, Rhedoni; Podarcis muralis, Padua).—Par., 1894, 147.—Stiles, 1901r, 197, 199, 201.—Stoss., 1895, 225— 226.—Also reported for Lacerta muralis.

arrectum of Mol. [1859, 831-833], and of Stoss. [1895, 225-226], are, according to Looss, 1899b, 567-568, probably misdeterminations. Molin's form appar-

ently belongs in Telorchis.—Braun, 1901a, 13.—Luehe, 1899, 530.

ascidia Rud., 1819a, 108, 399–400 [nec Ben., 1873] (in Sparus boops, S. pagrus; Arimini and Naples).—Barbagallo & Drago, 1903, 410, to (Brachylaimus) (in Box boops, Pagrus vulgaris; Catania).—Carus, 1884, 130.—Dies., 1850a, 369 (in Box vulgaris, Pagrus vulg.; Armini and Naples).—Duj., 1845a, 458.—Looss, 1899b, 609.—Stoss., 1886, 45; 1898, 41.

Loos, 1899b, 609.—Stoss., 1886, 45; 1898, 41.

ascidia Ben., 1873c, 328–329 (see D. lagena Brand.) [nec Rud., 1819a].—Brand., 1889b, 249, 250.—Braun, 1892a, 577, 618, 642, 663; 1893a, 864; 1900, 388; 1900b, 221, 223, 224, 225.—Cobbold, 1879b, 294.—Gamb., 1896a, 71.—Kowal., 1894, 2.—Linst., 1884, 140, fig. 25; 1885, 248–249; 1887, 102–103, pl. 2, figs. 4, 17f (in Vesperugo nathusii) (includes Cerc. armata Sieb.).—Looss, 1894a, 2, 125, 127, 167, 168, 181, 184, 185, 186, 212, 270, 275, pl. 3, fig. 52, pl. 4, figs. 72–73; 1896b, 86; 1898, 453, 454, 455, 457, 458, fig. 1, ii; 1899, 547, 556, 609 (type of Lecithodendrium), 610, 613, 618, 718.—Luehe, 1899, 536.—Macé, 1881, 421, in Vespertilio murinus; 1882, 62.—Stiles, 1901r, 200.—Stoss., 1892, 21 (in Plecotus auritus, Rhinolophus hippocrepis, Vespertilio dasycneme, V. daubentonii, V. murinus, V. mystaginus, Vesperuso, nathusi, Vesperus V. daubentonii, V. murinus, V. mystacinus, Vesperugo nathusi, Vesperus serotinus, Nannugo pipistrellus).—Also reported for Rhinolophus ferrum equinum, R. hipposideros, Limnæa stagnalis, Planorbis corneus, Ephemera, Perla, Chironomus plumosus.

ascidioides Ben., 1873, 332 (in Vesperugo noctula).—R. Bl., 1891, 467.—Braun, 1892a, 579; 1893b, 185 (in Vespertilio murinus); 1900, 221, 222; 1900, 388.—Cobbold, 1879b, 294.—Linst., 1885, 249–250; 1887, 103 (in Rhinolophus hipposideros).—Loos, 1894a, 2, 167, 168, 181, 182, 184, 185, 210, 211, 212, 275, pl. 3, fig. 51; 1896b, 86; 1898a, 453, 454, 455, 457, 458, fig. 1, i; 1899b, 547, 609, 610 (to Lecithodendrium).—Mont., 1892, 712.—Staff., 1902, 483 (in Vespertilio subulatus) to (Lecith.); 1905, Apr. 11, 692 (syn. of Lecith. chilostomum Mehlis).—Stiles, 1901, 200.—Stoss., 1892, 16 (in Rhinolophus hippocrepis, in

Belgio; Vesperugo noctula; Vespertilio murinus; in Clarte-Dieu, Francia; Vesperus serotinus).

- ascoidea Leidy, 1877e, 201 (in Planorbis parvus) to (Gymnocephala); 1904a, 144.— Linst., 1889a, 122 (ascoideum).
- asperum Wright, 1879, 57–58, pl. 1, figs. 3–5 (in Ardea minor).—Braun, 1892a, 583.—Mont., 1893, 94.—Stoss., 1892, 164 (to Echinost.).
- aspidophori Ben., 1870, 34, pl. 4, fig. 16 (in Aspidophorus europæus).—Braun, 1892a, 643.—Stoss., 1886, 45.
- assula Duj., 1845a, 398 (in Coluber natrix; Toulouse) to (Dicroccelium).—Cobbold, 1860a, 20.—Dies., 1850a, 390.—Erc., 1881e, 78; 1882a, 314 (from Tropidonoton).—Kampmann, 1894b, 454.—Linst., 1879, 184.—Mont., 1893, 187, 188, 189.—Par., ——, 147.—Stoss., 1895, 222.—Volz, 1899, 235, 237.
- atomon (Rud., 1802) Rud., 1809a, 362–363 (in Pleuronectes flesus); 1810a, 328; 1819a, 95.—Ben., 1870, 48 (D. altemon).—Braun, 1892a, 763, 764, 766; 1893a, 873.—Carus, 1884, 132.—Cobbold, 1858b, 160–161, pl. 32, figs. 30–32 (in Hippoglossus vulgaris); 1860a, 30.—Dies., 1850a, 340 (in Platessa flesus; Gryphiae); 1859c, 427–428.—Duj., 1845a, 466–467.—Kroyer, 1843–1845a, 297 (in Platessa flesus).—Linst., 1878, 225, fig. 7.—Mol., 1858, 288 (in Platessa passer; Patavii); 1859. 828; (in Platessa passer; Padua); 1861, 199.—Mont., 1893, 193.—Nicoll, 1907, 73 (to Podocotyle).—Odhn., 1901, 484, 485, 499, 502. 503, 506, 507, 508, 509, 511, 512, 513, 514; 1905, 320, 321 (to Podocotyle).—Olfers, 1816, 45.—Olss., 1868, 30.—Stiles & Hass., 1898a, 95.—Stoss., 1886, 33 (in Anarrhichas lupus); 1887, 185, 186; 1898, 47–48.—Wagener, 1860, 183, 188.
- atriventre Weinland, 1856, 24 (in Physa heterostropha).—Braun, 1893a, 865.—Gamb., 1896a, 71.
- attenuatum Rud., 1814a, 103, F. longicollis Abildg., renamed [later renamed naja] [nec Duj., 1845a].—Olfers, 1816, 46.
- attenuatum Duj., 1845a, 392–393 [nec Rud., 1814] (in Turdus merula; Rennes) (includes ?D. longicauda Rud., ?D. macrourum Rud., ?D. albicolle Rud., ?D. clathratum Deslongchamps).—Braun, 1902b, 109.—Dies., 1850a, 345 (syn. of D. macrourum Rud.).—Rail., 1900, 240.—Schlotthauber, 1860, 129.—Stoss., 1892, 156.
- attenuatum Bremser, MS., in Rhynchops nigra; Brazil.—Braun, 1901f, 563 (renamed Microlistrum spinetum); 1902b, 60.
- auriculatum Wedl, 1857, 242–243, pl. 1, fig. 2 (in Acipenser ruthenus).—Braun, 1892a, 586; 1900, 231.—Carus, 1884, 128.—Cobbold, 1860a, 25.—Dies., 1858e, 343.
- auriculatum Wedl of Lint., 1898, 521, pl. 45, figs. 1–7 (U. S. Nat. Mus. 4845); 1901, 415, 435.—Looss, 1902, 454 (of Lint., 1898) (probably a Bunodera and identical with D. petalosum).—Staff., 1904, May 3, 491 (of Lint., as syn. of Acrodactyla petalosa Lander).—Stoss., 1886, 18, 63 to (Crossodera).—See also lintoni.
- baccigerum Rud., 1819a, 108, 398–399 (in Atherina hepsetus; Naples).—Braun, 1892a, 700, 720, 721, 734.—Carus, 1884, 129.—Cobbold, 1860a, 27.—Dies., 1850a, 369.—Duj., 1845a, 461.—Mont., 1893, 95.—Stoss., 1886, 45; 1889, 27; 1898, 44–45.
- bacillare Mol., 1859, 834–835 (in Centrolophus pompilius; Padua).—Barbagallo & Drago, 1903, 410 (to Dicrocœlium) (in Scomber scombrus; Catania).—Braun, 1892a, 720, 737.—Carus, 1884, 130.—Looss, 1899b, 571 (this form, as described by Stoss., belongs to the Creadiine, possibly to Creadium).—Mont., 1893, 86, 94, 102.—Odhn., 1905, 328, 338.—Stoss., 1886, 31 to (Dicrocœlium); 1887, 92–93; 1898, 50.
- baculus Dies., 1850a, 391 (in Mergus albellus) (D. mergi Rud., 1819a, 121, renamed).—Braun, 1892a, 584; 1893a, 911.—Cobbold, 1860a, 36, 163 (to Echinost.).—Linst., 1877, 183, pl. 13, fig. 15.—Stoss., 1892, 163.
- bælzi Cobbold, 1884g, 976 (syn. of D. pulmonale Bælz).
- bagri incapsulatum Wedl, 1861, 479–480, pl. 3, fig. 40 (in Bagrus sp.).
- baraldii Sons., 1892, 91, 92, 93, 94 (in Zamenis viridiflavus Lacep.); 1893, 499; 1893, 184; 1896, 116.—Braun, 1893b, 185 (in Z. vir.).—Rizzo, 1902, 28 (in Coluber viridiflavus; Catania).—Stoss., 1895, 218-219, to (Brachylaimus).—Volz, 1899, 235, 237.—West, 1896, 323.
- barbatum (Linn., 1761) Zed. of Rud., 1809a, 441.—[See Dies., 1850a, 573] (syn. of Tetrabothriorhynchus migratorius).

batryophorum Ben., 1870, 51, see botryophorum.

beleocephalum Linst., 1873, 104–105, 106, pl. 5, fig. 2 (in Ardea cinerea).—Stoss., 1892, 169 (to Echinost.).

belones Braun, 1893a, 871 (in Belone vulgaris), based on Wedl, 1855, 382-383.

belones vulgaris Dies., 1855, 64, based on Wedl, 1855, 382–383 (gives no specific name); 1858e, 355, 356 (in Belone vulgaris).—Carus, 1884, 128.—Stoss., 1886, 46.

benedeni Stoss., 1898, 51, for benedenii, 1887.

benedenii Stoss., 1887, 95, pl. 10, fig. 39 (in Mugil chelo); 1898, 51.—Braun, 1891d, 424; 1892a, 569, 698, 720.—Looss, 1894, 178; 1901, 439; 1902, 129, 130, 136, 137.—Mont., 1893, 82.—Sons., 1894, 254, 255.—[Type of Haploporus 1902.]

bergense Olss., 1868, 43–44, pl. 10, fig. 93 (in Anguilla vulgaris).—Ben., 1870, 83 (bergensis).—Odhn., 1905, 356, 357 (syn. of Lecithaster gibbosus (Rud.)).—Stoss., 1886, 24 to (Brachylaimus); 1902, 582.

bergensis Ben., 1870, 83, for D. bergense.

beroës Will., 1844, 343–344, pl. 10, figs. 10–13 (in Beroe rufescens; Triest).—Busch,
1851, 99 (in Saggita; Triest).—Dies., 1850a, 381 (renamed D. papillosum).—
Mont., 1893, 123.—J. Mueller, 1850, 497.—Pag., 1862, 298.

betencourti Mont., 1893, 33, 34, 35, 43, 52, 84, 85, 88, 91, 95, 96, 102, 106, 107, 114, 146, 171, 186, 190–193 (syns. D. luteum of Ben., 1870, 3; Stoss., 1886, 52; Mont., 1890, 432) (in Scyllium; Boulogne), pl. 8, fig. 121; 1896, 152.—Ariola, 1899, 8 (D. luteum Ben. as syn.).—Linst., 1903, 354.—Looss, 1894, 219; 1899b, 622.—Stoss., 1899, 10 (to Pleurogenes; syn. luteum Ben., in Scyllium canicula, S. stellare at Boulogne); 1904, 193–197.

bicoronatum Stoss., 1883, 113–114, pl. 1, figs. 1–3 (in Ciconia nigra); 1886, 34, 113 to (Echinost.); 1887, 186; 1890, 42 (=D. cesticillus Mol.); 1892, 64; 1898, 54.—Braun, 1892a, 571, 583; 1893a, 911; 1893b, 184 (in Zeus faber).—Carus, 1884, 127.—Looss, 1901, 599, 600.—Mont., 1888a, 14; 1893, 160, 161.—Sons., 1891, 258.—[1901 to Stephanochasmus.]

bifurcatum Wedl, 1861, 477–478, pl. 3, fig. 38 (in Crocodilus vulgaris; Egypt).—Brand., 1888a, 15; 1888, 57, to Diplost.

bifurcum Braun, 1899, 631 (in Flussschildkröten); 1901b, 13, 18 (to Telorchis).—Luehe, 1899, 529 (to Telorchis).

bilharzii Herff, 1894a, 415, for Bilharzia hæmatobia.

biliosum Leidy, 1858a, 111 (in a fish; America); 1904a, 112.—Dies., 1859c, 430-431.

bilis (Braun, 1789) Zed., 1803a, 214–215 (in Falco melanaëtus; Germany).—
 Braun, 1892a, 632.—Dies., 1850a, 376 (in Falco chrysaëtos, F. albicilla).—
 Rail., 1898, 412 (to Campula).—Rud., 1809a, 408 (renamed D. crassiusculum).

bilobum Rud., 1819a, 114, 416 (in Tantalus falcinellus; Mus.Vien.) to (Echinost.),—Braun, 1892a, 584.—Dies., 1850a, 385; 1858e, 347 (in Ibis falcinellus, Platalea leucorodia, Fulica atra); 1859c, 434.—Duj., 1845a, 431—432.—Mol., 1858, 291 (in Ibis falc.; Patavia); 1861, 218–219, 247, pl. 3, figs. 5, 8.—C. Par., 1887, 332.—Stoss., 1891, 216; 1892, 171 (to Echinost.).—Wedl, 1857, 246–247, pl. 1, fig. 7.—Also reported for Plegadis falcinellus.

binode (Mueller, 1776) Zed., 1803a, 215 (in a fish).—Dies., 1850a, 379.—Rud., 1809a, 439-440; 1819a, 123.

bipartitum (Wedl, 1855) Mont., 1893, 150.—[1902 to Didymostoma.]

blanchardii Cobbold, 1860a, 8, Brachylæmus erinacei Blanchard, 1847, renamed (in Erinaceus europæus; Paris).—Stoss., 1892, 35 (syn. of D. linguæforme Dies.).

blennii (Mueller, 1776) Zed., 1803a, 211–212.—Dies., 1850a, 344 (syn. of D. divergens Rud.).—Harz, 1881c, 11.—Rud., 1809a, 371 (renamed D. divergens Rud.).

bliccæ Linst., 1877, 185–186 (in Blicca bjoerkna).—Braun, 1893a, 871.—Stoss., 1886, 46.

bolodes Braun, 1902b, 11, 17, 18, 19, 74, figs. 12–13 (in Fulica atra; Rossitten (Kurische Nehrung)).

bonnieri Mont., 1893, 40, 42, 43, 49, 61, 82, 83, 84, 85, 86, 88, 89, 90, 91, 95, 102, 180–184, pl. 1, fig. 8, pl. 6, figs. 76–80 (in Trigla gurnardus; Wimereux), syn.

D. varicum Mueller of Mont., 1889, 492, pl. 22, fig. 20.—Looss, 1894, 170;

1899b, 641, 642 (type of Liopyge).

bosci Cobbold, 1859d, 364, pl. 63, figs. 6-7 (includes Fasc. colubri Bosc, Dist. colubri americani Rud.) (in Coluber sp.); 1860a, 19; 1861e, 119; 1864, 22; 1879b, 455 (boscii).—Braun, 1893a, 872, 876 (boscii).—Staff., 1905, Apr. 11, 692 (belongs to Zeugorchis).—Stoss., 1895, 226.—Volz, 1899, 235, 238.—West, 1896, 322.

boscii Cobbold, 1879b, 455, for bosci, q. v.—Sons., 1892, 92.

bothryophoron Braun, 1892a, 700, for botryophoron.

botryophoron Olss., 1868, 42–43, fig. 92 (in Cyclopterus).—Ben., 1870, 51 (batryophoron).—Braun, 1889a, 367; 1892a, 643, 700 (bothryophoron), 721.—Lint., 1901, 415 (in Clupea harengus. Pomolobus pseudoharengus), 419, 437, 439, 444, 485, figs. 355, 356 (bothryophoron); 1905, 328, 334, 378, 397, 411, figs. 174–175 (bothryophoron) (in Micropogon undulatus, Orthopristis chrysopterus, Paralichthys dentatus).—Looss, 1899b, 729 (bothryophoron) (to Hemiurus).—Odhn., 1905, 356 (syn. of Lecithaster gibbosus (Rud.)), 359 (type of Lecithophyllum).—Stoss., 1886, 21 (in Cyclopterus lumpus, Argentina silus.

Molva abyssorum); 1896, 128 (bothryophoron); 1898, 40 (both.).

brachysomum Crep., 1837a, 314 (in Hæmatopus ostralegus); 1846. 134, 136, 142; 1849a, 1, 68.—Braun, 1892a, 578, 583, 710, 720, 721, 723, 728, 733, 736; 1893a, 865, 874, 879; 1900, 234; 1902b, 155.—Cobbold, 1879b, 438.—Dies., 1850a, 397; 1858e, 354 (in Hæmatopus ostralegus).—Gamb., 1896a, 71.—Jægers., 1898, 14; 1900, 738, 739; 1901, 982 (type of Levinsenia).—Linst., 1878, 133 (in Hæmatopus ost.), 136 (in Ægialites hiaticula), 142 (in Ardea cinerea); 1882, 20–21; 1887, 104; 1889, 48 (in Tringa alpina), 50 (in Æg. hiat.), 55 (in Fuligula cristata), 56 (in Glaucion clangula); 1894b, 336 (in Actitis hypoleucus; Seeburger See).—Looss, 1894a, 175; 1899b, 620, 621, 622 (? type of Levinsenia): 1901b, 207; 1902m, 703, 704, 705, 706 (type of Levinsenia according to Luehe), 826.—Luehe, 1899k, 537, 538.—Mont., 1888, 94; 1893, 43, 83, 86, 95, 102.—Nicoll. 1906, 524.—Odhn., 1900, 13; 1905, 317.—Stiles & Hass., 1902d, 20 (type of Levinseniella).—Stoss., 1892, 148 (includes D. calidris Rud.).—Villot, 1875, 475, 476; 1878, 22–24 (in Anas glacialis, Ardea cinerea), 27, 28, pl. 5, fig. 7 (in Strepsilas interpres, Tringa variabilis).—Ward, 1901, 176, 184 (type of Levinsenia Stoss.).—Also reported for Actitis hypoleucus, Anthura gracilis, Bucephala clangula, Charadrius calidris, C. hiaticula, Harelda glacialis.

bramæ (Mueller, 1776) Zed., 1803a, 218.—Dies., 1850a, 341 (syn. of D. globiporum

Rud.).—Harz, 1881c, 11.—Rud., 1809a, 365.

brevicolle Crep., 1829b, 54–55 (in Hæmatopus ostralegus; Greifswald).—Braun, 1901, 944: 1902b, 11, 12 (to Psilost.), 14, 15, 23, 26.—Cobbold, 1859d, 364; 1860a, 14.—Dies., 1850a, 363–364.—Duj., 1845a, 445.—Linst., 1887, 104.—Mehlis, 1831, col. 176.—Mueller,•1897, 19, pl. 3, fig. 2.—Nicoll, 1906, 514 (in Hæm. ostr.).—Stoss., 1892, 35 D. (Dicrocælium), 177.

brusinæ Stoss., 1889, 25–26, pl. 14, fig. 60 (in Oblata melanura; Triest); 1898, 32; [1904, 197 (type of Diphtherost.)].—Braun, 1892a, 587, 700, 736; 1893a, 874, 910.—Looss, 1899b, 622, 623, to Pleurogenes.—Mont., 1893, 85, 86, 102.

brusinai Looss, 1901d, 399, for brusinæ (in Labrus merula, L. maculata, Crenilabrus pavo, C. griseus, C. quinquemaculatus), 400–401; 1902h, 129.—Barbagallo & Drago, 1903, 410 to (Brachycæcum) (in Oblata melanura; Catania).—Stoss., 1904, 195, 196, 197 (type of Diphtherost.).

buccini Fil., 1855b, 23.

buccini mutabilis Fil., 1855b, 17, pl. 2, figs. 17–18 (in Buccinum mutabile; Gulf of Gênes); 1857b, 433, pl. 2, figs. 17–18.—Dies., 1858d, 266 to Cerc. (Acanthocephala)).—Par., 1894, 160.—Villot, 1875, 479.

bufonis Linst., 1877, 185 (in Bufo vulgaris).—Braun, 1893a, 870.—Stoss., 1889, 69.

bursæ fabricius Podwyssozki, 1890, 157 (in Gallus domesticus).

bursicola Crep., 1837, 310, 311, 313 (in Ardea cinera): 1846, 134.—Braun, 1893a, 876; 1893d, 467 (in A. cin.); 1901, 259 (syn. of Echinost. cloacinum Braun) (types in Creplin's collection, Zool. Museum, Greifswald); 1902b, 74.—Dies., 1850a, 397.—Hass., 1896a, 2 (syn. of Cephalogonimus ovatus (Rud.)).—Stiles, 1901k, 593; 1901l.—Stoss., 1892, 144 (syn. of Cephalog. ov. (Rud.) Stoss.): 1898, 23 (syn. of Cephalog. ov.).

buski Bl., 1888, 622-625, for D. buskii, q. v.

buskii Lankester, E., 1857b, 437 (in Homo; from Asia).—Aitken, 1866, 804, syn. of D. crassum (Busk).—Bl., 1888a, 622 (includes D. crassum Busk, 1859 [nec Sieb., 1836], 623–625; 1891, 610 (buski) (syn. D. rathouisi Poir.).—Braun, 1895b, 141 (includes D. crassum); 1889a, 348; 1902b, 95; 1903, 3 ed., 153 (to Fasciolopsis); 1906, 159, 160, fig. 90 (buski).—Brunet, 1902a, 125 (buski).—Cobbold, 1858b, 167; 1860a, 5; 1866, 6; 1876, 303; 1879b, 20.—Gamb., 1896a, 63.—Huber, 1896a, 575–576.—Ijima, 1889, 131.—Kholodk., 1898, 26.—Leuck., 1863a, 587.—Linst., 1878, 3 (syn. of D. crassum).—Looss, 1899b, 557 (see Fasciolopsis).—Moniez, 1896, 86, 114–117, 118.—Rail., 1893a, 363, to (Dicrocœlium).—Simon, 1897, 209, 223.—Tyson, 1903, 3 ed., 1180.—Ward, 1895, 328 (in Homo); 1903, 867 (to Fasciolopsis); 1904, v. 39, 260.—Weinland, 1858, to Dicrocœlium.—Wood, 1904, v. 39, 260.

buteonis (Gmelin, 1790) Zed., 1803a, 221.—Dies., 1850a, 396 (in Falco buteo).—Duj., 1845a, 442.—Rud., 1809a, 430; 1819a, 119.—Stoss., 1892, 178.

[cæruleum Sluiter, 1898, 4, 9, 14-16, pl. 2, fig. 4; pl. 3, fig. 11 (a tunicate).]

cahirinum Looss, 1896, 119–121, pl. 8, figs. 83–84 (in Bagrus bayad; Cairo); 1893, 752 (to Haplorchis).

calceolus Mol., 1858, 129 (in Conger conger, at Patavii); 1861, 210–211.—Braun, 1892a, 586.—Carus, 1884, 132.—Cobbold, 1860a, 25.—Dies., 1858, 342.—Stoss., 1886, 28 to (Dierocœlium).

calidris Rud., 1819a, 120 (in Scolopax calidris).—Dies., 1850a, 323 (syn. of Monost. mutabile Zed.).—Duj., 1845a, 447.—Stoss., 1892, 148 (syn. of D. brachysomum Crep., 1846).

callionymi Ben., 1870, 53, pl. 4, fig. 8 (in Calliomynus dracunculus).—Stoss.,

1886, 46.

calyptrocotyle Mont., 1891, 110; 1892, Oct. 7, 172, 176; 1893, 1–138, 171, 175, to (Accacelium), pl. 1, fig. 14; pl. 2, figs. 1–11; pl. 3, figs. 12–15, 17–21, 24–28, 30, 31; pl. 4, fig. 50; pl. 7, fig. 104; pl. 8, fig. 136 (in Beroë ovata Dch.); 1896, 151, 152.—Darr, 1902, 688.—Looss, 1894, 131, 142, 145, 165, 166, 189, 219; 1902, 639 (to Orophocotyle).

campannlatum Rivolta, 1884, 27, for campanulatum.

campanula Duj., 1845a, 435 (in Esox lucius; Rennes) to (Crossodera).—Cobbold, 1860a, 32 (=D. nodulosum).—Dies., 1850a, 381 (syn. of D. nod. Zed.); 1858e, 354; 1859e, 435, 436 (cf. Gasterost. fimbriatum Sieb.).—Kroyer, 1846–53a, 253; 1852–53a, 1221.—Moul., 1856a, 219 (in Cyprinus idus; Rennes).—Stoss., 1886, 42, to (Echinost.).—Tennent, 1906, 638 (syn. of Gasterost. fimb. Wag.) (in Esox).—Wagener, 1858, v. 1, 250, pl. 9; 1860, 165 (syn. of Gasterost. fimb.).—Wedl, 1857, 233–244, pl. 1, fig. 3.—Ziegler, 1883, 538 (in Cyprinus idus).

campanula Linst., 1886a, 125, for D. campula Cobbold.—Stoss., 1892, 16 (syn. of D. oblongum (Cobbold) Stoss.; includes Campula oblonga Cobbold, 1858), (in Phocena communis at Warnemünde; and Platanista gangeticum Cuv.).

campanulatum Erc., 1874, 432–439, pl. 1, fig. 7 (in Canis familiaris); 1875b, —; 1875c, —; 1875d, —; 1875e, —, 39–40 (in Canis domesticus); 1875f, 278; 1875g, —; 1876a, —.—Braun, 1893a, 875; 1893e, 348, 349, 351; 1893f, 386, 387, 389, 424 (=D. truncatum Rud.); 1893g, 802, 803.—Cobbold, 1879b, 300.—Jong, 1887a, in dog; 1887b, 223–224 in cat; 1890a, 140; 1896a, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, fig. 2.—Rail., 1890, 131 (in cat).—Schneidemuehl, 1896, 302 (kampanulatum).—Sons., 1889, 276, 277, 280, 281 (syn. of D. conus Crep.).—Ward, 1895, 341.—Zwaardemaker (1887), 265; 1888, 679.—Zuern, 1882, 220.

campula Cobbold, 1876q, 40, pl. 10, fig. 2, Campula oblonga renamed; 1879b, 418, 419, 423, fig. 68.—Braun, 1900g, 250.—Hoyle, 1890. 537.—Jackson, 1888, 648.—Looss, 1899, 558, 559, 560.—Mont., 1893, 44, 82.—Stiles, 1901r, 203.—Stiles & Hass., 1899, 102.—Stoss., 1892, 16 (syn. of D. oblongum Cobbold).

canaliculatum Rud., 1819a, 676 (in Sterna sp.=galericulata teste Dies.; Brazil).— Braun, 1901f, 561, 562; 1901g, 947; 1902b, 59, 140, 142, 144 (to Bilharziella).— Dies., 1850a, 346 (in Sterna galericulata).—Duj., 1845a, 449.—Stoss., 1892, 36, 178.

canaliculatum Mehlis, in Crep., 1846, 138 (in Colymbus cristatus).

capense Harley, 1864a, 55–72, figs. 1–16 (in Homo; Cape of Good Hope); 1864b,
173–175; 1864c, 156–157; 1864d, 161–163; 1864e, 515–517; 1864f, 142–143;
1864g, 302; 1864h,293; 1865a, 68.—R. Bl., 1888a, 636 (syn. of Bilharzia hæmatobia).—Bourel-Roncière, 1888a, 130 (capensis).—Braun, 1903, 3 ed., 169.—

Chassaniol & Guyot, 1878a, 66.—Cobbold, 1864g, 157; 1879b, 39; 1885a, 499 (syn. of Bilh. hæm.).—Colloridi, 1891a, 856 (syn. of B. hæm.).—Almeida Couto, 1872, 4.—Crevaux, 1874a, 173 (capensis).—Duffek, 1902a, 774.—Hoyle, 1890, 538.—Huber, 1896a, 580 (syn. of Bilh. hæm.).—Leuck., 1876, v. 2 (3), 873.—Simon, 1897, 99.—Stiles, 1898a, 58.—Ward, 1895, 253 (syn. of Gynæcophorus hæm.) (in Homo).

capitellatum Rud., 1819a, 99, 379–380 (in Uranoscopus scaber; Arimini, Naples).—
Braun, 1883a, 41; 1892a, 672; 1893a, 875.—Carus, 1884, 131.—Cobbold, 1860a, 26.—Dies., 1850a, 356–357.—Duj., 1845a, 456.—Jacoby, 1900, 2.—Looss, 1901, 656, 658.—Luehe, 1900, 504, 506.—Mont., 1888a, 17; 1893, 18, 32, 40, 41, 42, 43, 65, 66, 83, 84, 85, 95, 97, 102, 173–176, pl. 1, fig. 13, pl. 6, figs. 69–75, pl. 7, fig. 103.—Sons., 1890, 142 (in Uran. sc. L.).—Stoss., 1886, 46; 1886, 47; 1892, 64; 1898, 38–39.—Will.-Suhm, 1870, 8; 1871, 182.

capsulare Dies., 1858e, 355, based on Wedl. 1857, v. 26, 247, pl. 1, fig. 8 (in Ardea purpurea, A. nycticorax, A. cinerea, Gallinula crex, Podiceps nigricollis).—

Braun, 1893a, 870.—Stoss., 1892, 176 (to Agamodist.).

carinariæ della Chiaje (1841a), 139, pl. 109, figs. 29–30.—Carus, 1884, 133.—Mont., 1888a, 195, 196.

carinatum Zed., 1803a, 217 (in Cyprinus; Perca fluviatilis) (includes Fasc. longicollis. Dist. cyprinaceum, F. lagena, F. jesis).—Dies., 1850a, 341 (syn. of D. globiporum), 365 (pars, syn. of D. inflexum).—Looss., 1902, 763 (and Sphærost. globiporum).—Rud., 1809a, 365 (=D. globiporum), 366, 395 (pars =D. inflexum).—Stiles, 1901, 168.

carnosum (Hass., 1891) Leuck., 1892b, 798.—Braun, 1892a, 674; 1892, 563, Dist. (Fasc.) carnosa.

carnosum Rud., 1819a, 93, 366, 676 (in Sparus dentex; Naples).—Carus, 1884, 130.—Cobbold, 1860a, 22.—Dies., 1850a, 337 (in Dentex vulgaris).—Duj., 1745a, 458.—Mol., 1859, 833 (in Dentex vulgaris: Padua).—Stoss., 1885, 161: 1886, 23; 1898, 39; 1886, 59, to (Brachylaimus).—Also reported for Corvina nigra.

carolinæ Stoss., 1889, 26–27. pl. 13. fig. 55 (in Alausa finta; Triest); 1898, 29.—
 Braun, 1891d, 423.—Looss, 1899b. 641 (=Pronopyge ocreata).—Mont., 1891, 17 (syn. of Apoblema ocreatum); 1891, 510, 512 (syn. of Ap. ocr. Rud.).—
 Also reported for Alosa vulgaris.

caryocatactis Zed., 1800a, 163, 168-169 (in Gorvus caryocatactes); 1803a, 210.— Braun, 1901, 562; 1902b, 123, 125 (syn. of Harmost. caudale).—Dies., 1850a, 362 (syn. of Dist. caudale).—Rud., 1809a, 382.

catervarium Looss, 1896, 118-119, pl. 8, figs. 81-82 (in Alosa finta; Cairo).

caudale Rud., 1809a, 382–384 (caryocatactis Zed., renamed) (in Corvus caryocatactes); 1819a, 103.—Braun, 1893a, 874; 1901, 561, 562, 564; 1902b, 123, 124 (to Harmost.), 125 (of Mueller, 1897, 16=Urogonimus macrostomus); 1902b, 128 (of Rud., 1809=Harmost, mesostomum).—Cobbold, 1860a, 14.—Crep., 1837, 317; 1846,—..—Dies., 1850a, 362.—Duj., 1845a, 442.—Giebel, 1857, 266.—Leuck., 1863a, 477.—Looss, 1899b, 703.—Mont., 1893, 149.—Mueh., 1898, 99; 1898, 16.—Mueller, 1897, 16–17, pl. 2, fig. 7.—Olfers, 1816, 44.—Sieb., 1836, 234.—Stoss., 1892, 178.—Reported also for Ampelis garrulus, Coracias garrula, Corvus glandarius, C. pyrthocorax.

caudale of Mueller, 1897, in Coracias garrula.—Braun, 1902b, 125 (syn. of Urogoni-

mus macrostomus).

caudale of Will.-Suhm, 1870, 97 (in Pyrrhocorax alpinus).—Braun, 1902b, 124–125 (nec Rud.).

caudatum Polonio, 1859.—Par., 1894, 147 (in Natrix torquata, Tropidonotus viperinus: Padua), 627; 1896, 9–11. fig. 4, to (Opisthorchis).

caudatum Linst., 1873, 103–104, pl. 5, fig. 3 (in Erinaceus europæus); 1900, 85 (claims priority for caudatum over leptostomum).—Blochmann, 1892b, 649–652 (raised experimentally from a Cercariæum in kidney of Helix hortensis).—Braun. 1892a, 570, 723; 1893a, 831, 856, 857; 1901e, 338 (=D. leptostomum Olss.).—Cobbold, 1879, 295.—Gamb., 1896a, 71.—Hofmann, 1899a, 189, 196, 201.—Looss, 1894, 2, 236; 1899b, 652 (to Heterolope).—Mont., 1893, 149.—Roewer, 1906, 185.—Stoss., 1892, 17 (syn. of D. leptostomum Olss.); 1898, 24.

caudatum (Bosc, 1802) Steenstrup, 1859, 183.

caudiporum Rud., 1819a, 96, 370 (in Zeus faber: Arimini).—Dies., 1850a, 342, 371.—Duj., 1845a, 422 to ? (Apoblema).—Wagener, 1860, 181 (in Zeus faber).

caviæ Sons., 1890, 100 (in Cavia cobaya); 1897, 250.—Braun, 1903, 3 ed., 147 (syn. of Fasc. hepatica); 1906, 150, fig. 83.—Rail., 1893a, 342 (syn. of Fasc. hep.).—Stiles, 1898a, 48.—Stoss., 1892, 7 (syn. of Cladoccelium hep.).—Ward, 1895, 246 (syn. of Fasc. hep.).
328 (in Homo), 332 (in Bos taurus), 335 (in Ovis aries); 1903, 865 (syn. of Fasc. hep.).

centra appendiculatum Leidy, 1904a, 277, misprint for centrappendiculatum.

centrappendiculatum Leidy, 1891a, 416 (D. appendiculata Leidy, 1877, not Rud., renamed): 1904a, 237.

cercatum Mont., 1893, 40, 42, 43, 83, 95, 102, 157, host and locality not known, specimen in British Museum, to (Urogonimus).

cerebrale Yamagiwa, 1890, 457 (in Homo), for D. pulmonum, see Paragonimus westermanii.

cesti veneris Vogt, ——, 299 (in Cestum veneris).—Braun, 1893a, 852.—Dies., 1858e, 356 (in Cestum veneris).—Mont.. 1893, 124.

cesticillus Mol., 1858, 131 (in Lophius piscatorius; Patavii); 1861,——.—Braun, 1883a, 41; 1892a, 584, 672; 1901a, 34.—Carus, 1884, 127.—Dies., 1858e, 351—352.—Jacoby, 1900, 2.—Linst., 1903, 354.—Looss, 1899, 576 (type of Stephanost), 581; 1901, 600.—Luche, 1900, 505.—Mol., 1861, 221–223.—Mont., 1893, 41, 42, 61 (cesticillum), 160, 161.—Stoss., 1886, 34; 1890, 42 (D. bicoronatum Stoss., 1883).—Will.-Suhm, 1870, 8.

cestoides Ben., 1870, 17, pl. 6, fig. 9 (in Raja batis).—Braun, 1893a, 873.—Linst., 1903, 354.—Lint., 1901, 415, 432.—Mont., 1893, 53.—Stoss., 1886, 46.

cglindraceum Looss, 1894a, 36, misprint for cylindraceum.

characis Stoss., 1886, 26 to (Brachylaimus) (in Charax puntazzo; Triest): 1887, 91; 1898, 37.

chefrenianum Looss, 1896, 73–76. pl. 5, figs. 50–51 (in Rhinopoma microphyllum Geoff.; Ghizeh); 1898, 453, 454, 456, 457, 458, fig. 1 iii; 1899, 716, 717 to Lecithodendrium.—Stiles, 1901, 200.

chefresianum Looss, 1896. pl. 5 (misprint for chefrenianum).

cheloniæ atræ Braun, 1899e, 629, see D. pachyderma.

chelydræ Staff., 1900, 406–407, fig. 5 (in Chelydra serpentina); 1905, Apr. 11, 690, type of Auridist.

chilostomum Mehlis, 1831, 186–187 (in bats).—Braun, 1892a, 577; 1900, 221, 222, 223; 1900, 387, 388.—Crep., 1837, 328.—Cobbold, 1860a, 7; 1879b, 294.—Dies., 1836, 240; 1850a, 349 (includes D. noctulæ Rud., 1819a, 119).—Kolenati, 1857, 11.—Linst., 1878, 225–226, fig. 8.—Mueh., 1898, 26.—Sieb., 1836, 234.—Stoss., 1892, 20–21 (in Plecotus auritus, Vespertilio daubentonii, V. nattereri, V. murinus, V. mystacinus, Vesperugo discolor, V. leisleri, V. noctula, V. serotinus.

chilostomum Mehlis of Ben., 1873b, 27, pl. 6, figs. 7, 8, 19.—Sieb., 1835, 56.—Stoss., 1892, 14 (syn. of D. aristotelis).

chinense Cobbold, 1876, 97 (D. sinense, renamed).

chloropodis Zed., 1800a, 164, 198–199 (in Fulica chlorops).—Dies., 1850a, 384 (syn. of D. uncinatum Zed.).—Rud., 1809a, 420.

choledochum Linst.. 1883a, 306–307. pl. 9, fig. 49 (in Anas sp.); 1886, 30.—Braun, 1892a, 699; 1893a, 875; 1893, 353; 1893i, 426; 1894i, 602, 605.—Looss, 1896b, 58, 59; 1899b, 675.—Stoss.. 1892, 161 (in Anas sp.; Turkestan).

chrysaëti Rud.. 1819a, 119 (in Falco chrysaëtus).—Duj., 1845a, 441–442.—Stoss., 1892, 151 (syn. of D. crassiusculum Rud.).

chrystallinum Hannover, 1864a, 3, for crystallinum.

cignoides Mont., 1888a, 40, 80, for cygnoides.

cimbiforme Mont., 1896, 165, for cymbiforme.

cinctum (Rud., 1803) Rud., 1809a. 422–423, to (Echinost.); 1819a. 116.—Ben., 1858a, 1861a, 85.—Cobbold, 1860a. 35, to Echinost. (in Vanellus cristatus, V. melanogaster).—Crep., 1837. 311, 312, 316.—Dies., 1850a. 386 (includes D. tringæ helveticæ Rud., Fasc. ci. Rud.) (in V. crist., V. mel.).—Duj., 1845a, 431.—Giebel. 1857, 265.—Linst., 1887, 104.—Mueller, 1897, 19–20, pl. 3, fig. 3 to (Echinost.).—Nord., 1832a, 90.—Olfers, 1816, 47.—Olss., 1876, 21.—Stoss., 1892, 172 to (Echinost.); 1898, 53.

[circumvallata Sluiter, 1900, 8 (a tunicate).]

cirratum (Rud., 1802) Rud., 1808a, xxvi. pl. 6. fig. 7; 1809a, 376; 1819a, 100-101; spelled F. cirrosum, Rud.. 1808a, 296.—Braun, 1893a, 874; 1901, 561, 563; 1902b, 37, 38 (syn. of D. elegans), 39, 43 (to Plagiorchis), 40, 44, 45, 47, 48, 49.—Cobbold, 1860a, 12.—Crep., 1837, 317, 322, 326.—Dies., 1850a, 350 (includes Fasc. cirrata Rud.).—Duj., 1845a, 413 to (Brachylaimus).—Fil., 1855b, 8 (cirrhatum).—Jacoby, 1900, 20.—Looss, 1899, 703.—Luehe, 1899, 530, 531; 1901, 487.—Mueh., 1896, 589; 1896, 262-266, figs. 5, 12; 1898, 16, 27, 92.—Olfers, 1816, 44.—Stoss., 1892, 153 (includes D. elegans Rud., 1819a, D. globocaudatum Duj., 1845a); 1904, 2 (to Plagiorchis).—Reported also for Ægiothus linarius, Corvus cornix, C. corone, C. monedula, C. pica, Larus marinus, Sturnus vulgaris.

cirrhatum (Rud., 1802) Fil., 1855b, 8, for cirratum.

cirrhigerum Nord., 1840, 616, for cirrigerum 1827, q. v.

cirrigerum Baer, 1827, 553 (in Astacus fluviatilis).—Brand.. 1891b. 264.—Braun, 1892a. 749, 751; 1893a, 859. 870 (cirrhigerum).—Crep., 1837, 325, 326.—Dies., 1836, 240; 1850a, 363 (syn. of D. isostomum Rud.).—Harz, 1881a; 1881b; 1881c. 1. 2. 3, 4, 7, 8, 9, 10, 11, 14.—Hoyle, 1890, 537.—Jackson, 1888, 648, 652.—Kampmann, 1894b, 452, 454, 457, 458, 459, 460, 462, pl. 20, figs. 11–13.—Linst., 1903, 281, 282 (cirrhigerum); 1904, 254.—Lint., 1892, 69-70.—Looss, 1893b, 814; 1894a, 230.—Marshall & Gilbert, 1905a, 480.—Mont., 1888a, 64.—Moul., 1856, 217 (in Astacus fluviatilis).—Much., 1898, 14.—Nord., 1840, 616.—Reinhard, 1871, 8 pp.—Schneidemuchl, 1896, 302.—Sieb., 1835, 56, 59, 64, 67.—Steenstrup, 1842, 56.—Warren, 1903, Dec., 273–301, pls. 24–26 (anat., development); 1904, 15 June, 311.—Wierzejski, 1888, in 20 pp., 1 pl.—Zaddach, 1881, 893; 1881, 1 Aug., 398–404; 15 Aug., 423–431.

cirrosum Rud., 1808a, 296. pl. 6, fig. 2 [=7] (for D. cirratum).

cladocalium Dies., 1858e, 354 (in Ardea minuta) [based on "Distome (Cladocalium) du foie du Blongias, Pontallié, 1853, 103"].—Braun, 1893a, 875.—Stoss., 1892, 178.

clathratum Deslongchamps, in Lamouroux 1824a, 563 (in Cypselus apus).—Braun, 1893a, 875, 910; 1899, 714; 1902b, 109 (=D. refertum Mueh.).—Cobbold 1860a, 15.—Dies., 1850a, 396.—Duj., 1845a, 393-394.—Jacoby, 1900, 10, 11.—Locss, 1899, 634 (to Dicrocœlium).—Mueh., 1898, v. 1, 22, 26, 84-85, 86, 87, pl. 4, figs. 7, 17.—Olss., 1876, 24.—Stoss., 1892, 155.

clathratum Deslongchamps of Olss.—Rail., 1900, 239 (syn. of Dicrocelium olssoni Rail.).—Braun. 1902b, 100, 109.

clara Dies., 1850a. 356 (in Eunectes scytale. Hydroscopus plumbeus, Coluber flaviventris, Clœlia fasciata: Brazil): 1855, 66, pl. 3, figs. 7–8; 1858, 339.—Braun, 1892a, 568, 587; 1901b, 13, 58: 1901i, 58.—Cobbold, 1860a, 19.—Luehe, 1899, 524, 529 (type of Telorchis); 1900b, 151.—Sons., 1892, 92.—Stoss., 1895, 229–230.

clavatum (Menzies, 1791) Rud., 1808a, 260; 1809a, 387, 391–393 (includes Hirudinella Garsin, 1730; Fasc. scombri pelamidis Tilesius), 437; 1814a, 102; 1819a, 106, 122, 394–395, 682–683, 685.—Baird, 1853a, 59, 60 includes Hirudinella clav. (Menzies) Blainv.; D. clav. Owen=H. ventricosa.—Barbagallo & Drago, 1903, 410 (in Pelamys sarda: Catania).—Ben., 1870, 37.—La Billardière, (1801), 46.—R. Bl., 1891r, 692–693, syn. D. ingens Moniez (in Requim).—Brand., 1891d, 16; 1898a. 208.—Braun. 1892a, 568. 571, 576, 588, 591, 592, 593, 597, 603, 605, 608, 609, 610, 611, 623, 624, 625. 630, 631, 632, 635, 636, 637, 638, 640, 641, 642, 645, 647, 664, 665, 669, 673, 675, 677, 682, 683, 684, 685, 686, 687, 688, 690, 698, 699, 700, 701, 705, 712, 713, 717, 719, 724, 726, 731, 733; 1893a, 873, 878; 1893b, 184; 1893d, 466.—Buttel-Reepen, 1900a, 585, 586, 589, 590, 592, 594, 598; 1902, 166, 167, pl. 6, figs. 7, 11a, 11b, 12; 1904, Jan. 26, 24–25; 1905, July, 52–53.—Carus. 1884, 131.—Chatin, 1887d, 1003.—Cobbold, 1860a, 21; 1867k, 200–205 (from sword-fish); 1879b, 458, 460, 461.—Crep., 1837, 316, 327.—Darr, 1902, 662, 664, 665, 666, 667, 689.—Dies., 1850a, 366–367 (includes Hirudinella Garsin: Fasc. fusca Bosc; F. cl. Menzies; F. scombri pelamidis Tilesius; D. coryphænæ Rud.) (in Coryphænæ hippurus: Brazil; Pelamys sarda. Thynnus vulgaris): 1859c, 431.—Duj., 1845a, 459–460.—Goto, 1891a, 181 (Distonum).—Jackson, 1888, 644, 646, 647–648.—Jægers., 1900, 72.—Jourdan, 1881a, 438–448. pls. 7–8, figs. 1–10 (anatomy); 1881b, 12 pp., pls. 7–8, figs. 1–10; 1881c, 604–605.—Juel, 1889, 13, 17, 18, 19.—Kroyer, 1838–1840a, 596 (in Thynnus vulgaris).—Lander, 1904a, 8.—Lint., 1898, 539–540, pl. 53, figs. 8–11; 1901b, 415, 421, 445, 448 (in Thunnus thynnus, Xiphias gladius).—

Looss, 1849a, 9, 113, 145, 147, 151, 152, 198, 202; 1895c, 69; 1896b, 204; 1899b, Looss, 1849a, 9, 113, 145, 147, 151, 152, 198, 202; 1895c, 69; 1896b, 204; 1899b, 556, 646, 739.—Moniez, 1892, 108–118.—Mont., 1888a, 23, 26, 30, 31, 38, 39, 42, 44, 47, 48; 1893, 7, 15, 20, 22, 26, 27, 33, 34, 44, 65, 66, 67, 68, 69, 73, 83, 86, 88, 89, 104, 106, 113, 135, 152.—Olfers, 1816, 46.—Osbeck, 1765, 392.—Owen, 1835, v. 1, 381–384, pl. 41, figs. 17–20; 1835, 72–73; 1837, 271–273, pl. 2.—Par., 1887, 334–335.—Poir., 1885, 4, 5, 6, 7–8, 9, 10, 14, 15, 16, 18, 22, 23, 25, 26, 27, 29, 32, 33, 37, 38, 39, 43, 45, 46, 47, 48, 51, 52, 54, 55, 56, 57, 58, 60, 62, 63, 70, 71, 73, 75, 77, 78, 79, 81, 83, 84, 88, 93, 95, 96, 97, 102, 103, 105, 107, 109, 110, 113, 116, 117, 118, 120, 121, 127, 130, 131, 133, 134, 138, 139, 142, 143, 144, 146, 147, 149, 150, 151, 152, pl. 23, fig. 1; pls. 24, 25, 26, figs. 1–3; pl. 27, fig. 1; pls. 28, 29, figs. 1, 3, 4; pl. 30, figs. 1–3; pl. 31, figs. 1–5.—Sieb., 1835, 68.—Stiles, 1901, 194 (type of Hirudinella).—Stoss., 1886, 46.—Wagener, 1860, 182, pl. 9, figs. 11–12 (in Scomber pelamys).—Ziegler, 1905, 39.—Also reported for Scomber thynnus. Scomber thynnus.

claviforme Brand., 1888, 247–251, pl. 17, fig. 1 (in Tringa alpina).—Braun, 1892a, 568, 578, 586, 662, 700; 1893a, 874, 910.—Jacoby, 1900, 11.—Jægers.. 1900, 739, 740.—Looss, 1899b, 618, 619, 620; 1902m, 704.—Luehe, 1899, 537.—Nicoll, 1906, 524.—Odhn., 1900, 13.—Staff., 1905, Apr. 11, 692.—Stoss., 1892, 148–149 to (Brachycœlium).—Ward, 1901, 184.

clavigerum Rud., 1819a, 103, 389–391, 589 (in Bufo viridis at Berlin, and B. cinereus, Rana esculenta, R. temporaria, Hyla arborea).—Baird, 1853a, 52 (Fasc. ranæ Froelich).—Bellingham, 1844, 424.—Ben., 1858a, 1861a. 93, 96-97.—Bettend., 1897a, 4, 5, 8, 15; 1897, 308, 309, 312, 319.—Biehringer, 1888a, 234.—Brand., 1891b, 265.—Braun, 1892a, 635, 707, 715, 736, 750, 751; 1893a, 865, 879.—Cobbold, 1858b, 162, pl. 33, figs. 52-53 (in Rana temporaria); 1860a, 16; 1879b, 454.—Crep., 1837, 310, 311, 312, 316, 322, 325, 328, 329; 1839, 387.—Dies, 1850a, 352, 388 (includes Fasc. graps Froelich); D. C. Rud 1839, 287.—Dies., 1850a, 352, 388 (includes Fasc. ranæ Froelich; D. c. Rud., ex parte, as syn. of D. retusum Duj.); 1858, 429; 1858e, 338 (Cerc. (Acanthocephala) ornata), adult in Rana temporaria, Pelophylax esculentus; larva in Planorbis corneus, Hydrachus concharum); 1859c, 429 (Cerc. armata major, in Lymnæus stagnalis, L. ovatus, Planorbis corneus; Belgia), 434 (Cerc. ornata).—Duj., 1845a, 404.—Duncker, 1881a, 23; 1882, 188; 1884a, 40.—Erc., 1881e, 77, 78, 82, 83, 84, 85, 89; 1882a, 313, 314, 318, 319, 320, 321, 325.—Fil., 1857c, 32.—Florance, 1866a, 11.—Gamb., 1896a, 72.—Hoyle, 1890, 537, 540.— Jackson, 1888, 648.—Juel, 1889, 37.—Kampmann, 1894b, 446, 448, 454, 457, 459, 460, 461, 462, pl. 20, figs. 14–18.—Kath., 1894a, 152.—Kowal., 1894d, 3; 1895, 350–351; 1902d, 9 (to Pleurogenes).—Lamouroux, 1824a, 562.—Leuck., 1899, 350–351; 1902d, 9 (to Pleurogenes).—Lamouroux, 1824a, 562.—Leuck., 1863a, 477, 520.—Linst., 1873, 1 (Cerc. ornata La Valette is young stage); 1882, 18; 1887, 97, 98, 99, 105; 1890f, 184; 1904, 254.—Looss, 1885b, 24, 39, 40, pl. 23, fig. 15; 1893b, 810, 811 (includes D. neglectum Linst., 1887); 1894a, 1, 2, 82, 83, 84, 85, 87, 91–101 (1 text fig.), 102, 103, 104, 105, 106, 107, 119, 124, 126, 137, 140, 158, 159, 165, 167, 173, 176, 179, 181, 191, 192, 197, 202, 205, 206, 208, 209, 210, 211, 212, 219, 227, 228, 230, 234, 264, 270, 273, pl. 2, figs. 30–32; pl. 8, figs. 165–166, 170–175; pl. 9, fig. 189 (includes D. neglectum Linst., D. medians Olss., e. p.); 1894d, 5, 50; 1896, 94, 95; 1898, 461; 1899b, 611, 614, 617 (type of Pleurogenes), 622, 623.—Luehe, 1899, 536; 1901, 169.—Mol., 1859, 846–848, pl. 3, fig. 3 (in Pelophylax esculentus; Padua).—Mont.. Mol., 1859, 846–848, pl. 3, fig. 3 (in Pelophylax esculentus; Padua).—Mont., 1888a, 15, 65; 1893, 71, 95, 102; 1896, 152.—Much., 1898, 23.—Nickerson, 1900, 814, 815.—Noack, 1892, --.—Nord., 1840, 616.—Olfers, 1816, 44.— Pag., 1857, 17, 18, 39-40, 52, pl. 4, figs. 8-14 (in brown and green frogs).—Par., 1887, 490.—Poir., 1885, 101.—Schwarze. 1885, 76.—Sieb., 1835, 64, 65; 1836, 233, 239.—Sons., 1893, 187, 189 (in Rana esculenta).—Staff., 1905, Apr. 11, 684.—Stiles, 1901, 197, 198, 199, 201.—Stoss., 1889, 64, 1896, 128; 1898, 43-44.—Wolf, 1903, 612, 613, 619, 621.—Zuern, 1882, 220.

clavigerum Rud., 1819a, of Duj., 1845a.—Looss, 1894, 101 (renamed D. confusum Looss); 1896, 88, 91, 93, 94, 95.

clupeæ (Schrank, 1788) Zed., 1803a, 218 (in Mayfische; Rhine).—Baird, 1853a, 54 (syn. of D. appendiculatum Rud.).—Dies., 1850a, 371 (syn. of D. app. Rud.).—Rud., 1809a, 437.

clupeæ rhenanæ Rud., 1809a, 437-433 (includes Fasc. alosæ Hermann, 1783, F. clupeæ Schrank, 1788, D. clupeæ Zed., 1803); 1819a, 110 (syn. of D. app. Rud.).—Baird, 1853a, 54 (syn. of D. app. Rud.).—Dies., 1850a, 371 (syn. of D. app. Rud.).

cochlear Dies., 1850a, 357–358 (in Sterna cantiaca, S. minuta; Brazil) (D. cochleariforme [sternæ] Rud., 1819a, 681, renamed).—Braun, 1901, 561, 563: 1901, 895

1902b, 56 (includes D. diesingi Cobbold), 58 (to Microlistrum).—Stoss., 1892, 37, 179.

cochleariforme Rud., 1819a, 681-682, 687 (in Pelecanus aquila, ?Sterna cantiaca [see D. cochlear], S. minuta, S. sp.; Brazil).—Braun, 1901, 561, 563; 1901, 895; 1902b, 55 (type of Microlistrum), 56, 58 (of Stoss., 1892, 37; Duj., 1845a, 449; Rud., 1819a, 681; pars, syn. of Microlistrum cochlear).—Cobbold, 1860a, 13.—Dies., 1850a, 357.—Duj., 1845a, 449.—Stoss., 1892, 37, 179.

cochleariforme sternæ Rud., 1819a, 681, of Dies., 1850a, 357 (renamed D. cochlear

Dies.).

colebs Linst., 1875, 192-193, pl. 3, fig. 15 in Fringilla collebs.—Braun. 1893a,

870.—Stoss., 1892, 176 (to Agamodist.).

cwlomaticum Giard & Billet, 1892a. 614-615 (in cattle; Tonkin).—Gomy, 1897a, 374, 375.—Looss, 1899b, 634; 1907, Feb. 1, 126 (in ox; Cao Bang, Tonkin).—Rail. & Marotel, 1898, 30, 31, 32.—Stiles, 1898a. 57 (syn. of Dicroccelium pancreaticum).—Ward, 1895, 332 (in Bos taurus).

coleostomum Looss, 1896b, 101–106, 154, pl. 7, figs. 66–68 (in pelican: Egypt); 1899b, 578, 581, 585 (type of Ascocotyle).—Braun, 1901b, 34.—Vaullegeard,

1901, 143 (colostomum).

collurionis (Schrank, 1790) Zed., 1803, 211.—Cobbold, 1860a, 15.—Dies., 1850a, 396.—Rud, 1809a, 430; 1819a, 119.—Stoss., 1892, 179.

colostomum Vaullegeard, 1901, 143 (for coleostomum).

colubri (Bosc, 1802) Rud., 1809a. 434—35 (in Coluber sp. americana).—See bosci. colubri americani Rud., 1819a. 121 (colubri, 1809, renamed).—Cobbold, 1859d, 364 (syn. of D. bosci).—Dies., 1850a, 398 (includes Fasc, colubri Bosc).—Sons.,

1892, 92. colubri murorum Rud., 1819a, 121 (in Coluber murorum).—Dies., 1850a, 367 (syn.

of D. allostomum Dies.).—Duj., 1845a, 452.—Stoss., 1895, 230.

colubri natricis intestinale Rud., 1809a, 433–434; 1819a, 103 (syn. of D. mentulatum Rud.) (in Coluber natrix).—Dics., 1850a, 355 (syn. of D. ment.).

cotubri natricis pulmonale Rud., 1809a, 434; 1814a. 103; 1819a, 99, 377 (syn. of D. naja Rud.) (in Coluber natrix).

colubri tesselati Rud., 1819a, 121 (in Coluber tesselatus).—Dies., 1850a, 355 (syn. of D. mentulatum Rud.).

columbæ Mazzanti, 1889a, 161–165 (in Columba livia; Pisa), teste Par., 1894, 145; 1890, 139.—Braun, 1902b, 117.—Hass., 1896a, 2, 3 (syn. of Mesogonimus commutatus (Dies.).—Sons., 1889, 283.

commune Olss., 1867, 31–32 (in Labrus).—Braun, 1891d, 424 (in Crenilabrus griseus).—Lint., 1901, 415, 485, 486.—Looss, 1901d, 399.—Odhn., 1901, 485, 486, 487, 494, 495, 499, 500, 503, 508; 1905, 321.—Sons., 1891, 257, 258 (in Crenilabrus griseus, Labrus mixtus).—Stoss., 1886, 32.—Also reported for Anguilla yulgaris

Cottus bubalis, C. scorpius, Crenilabrus melops, Labrus maculatus, L. sp. commune Hausmann, 1896?, 24.—Looss, 1899b, 571 (possibly identical with Creations)

dium angusticolle).

commutatum Dies., 1858e, 339-340 (D. dimorphum, 1852, 555, pl. 16, 1 [not Dies.] renamed) (t. h. Phasianus gallus; Pisa).—Anacker, 1887c, 513.—Braun, 1893a, 874; 1900, 25; 1900h, 19, pl. 1, figs. 1, 2; 1902b, 116.—Caruccio, 1886, 293.—Cobbold. 1860a, 15.—Hass., 1896a, 2, 3 (to Mesogonimus).—Landois, 1882, 23.—Looss, 1899b, 650 (perhaps a Clinost.).—Mont., 1893, 155.—Rail., 1890, 131.—Schneidemuehl, 1896, 303 (kommutatum).—Sons., 1889, 11; 1889, 282-283; 1890, Feb. 15, 138.—Stoss., 1892, 175 (to Mesogonimus).

compactum Cobbold, 1859d, 363. pl. 63, figs. 1–3 (in lung, Viverra mungos); 1861,
118; 1864, 16, fig. 2; 1879, 299, 419; 1884g, 976.—Braun, 1892a, 663; 1893a,
876; 1899g, 492, to Paragonimus: 1901e, 329, 330, 333, 334.—Kerbert, 1878a,
272.—Mont., 1893, 83, 155.—Sons., 1884, 19.—Stiles & Hass., 1890a, 605, to
Paragonimus.—Stoss., 1892, 31 (to Mesogonimus).—Also reported for Mungos

fasciatus.

complanatum Rud., 1814a, 103-104 (in Ardea cinerea: Berlin): 1819a, 98, 376-377, 680.—Braun, 1893a, 873; 1899g, 465, 485, 486, 489 (specimens from Pavia and Cagliari, as syn. of Clinost. heterost.), 490; 1900h, 14, 15, 17, 18, 19, 23 to Clinost., 24, 25, 26, 27, 28, 29, 30; 1900, 141; 1901, 561.—Cobbold, 1860a, 9.—Dies., 1850a, 338 (to Dist.), (t. h. Ardea cinerea; Berlin), 354.—Duj.,

1845a, 399-400 to (Dicrocœlium).—MacCallum, 1899, 697, 706.—Olfers, 1816, 44.—Par., 1887, 23 (syn. of D. hians) (to Clinost.); 1887, 489.—Stoss., 1892, 160.

complanatum from Ardea cinerea and Nycticorax griseus, labeled complanatum
 (=hians) of Genueser Sammlung, from Pavia and Cagliari, are according to
 Braun, 1900h, 19, identical with Clinost. heterost.

complanatum Erc. of Par., 1894, 144, and Ward, 1895, 341, from Canis fam., misprint for campanulatum.

complexum Stiles & Hass., 1894e, 425–426, pl. 1, figs. 3–4, to (Dicrocœlium) (in Felis catus dom.; U. S. A.); 1894f, 89–91.—Braun, 1900h, 16.—Looss, 1896, 58; 1899b, 565 (to Metorchis).

complicatum Mehlis, 1846, 141 (in Haliaëtus carbo).—Crep., 1846, 141.

concavum Crep., 1825a, 45-47, 83, figs. 7-8 (in Colymbus rufogularis); 1837, 310, 314, 318; 1846, 138, 141, 145, 146.—Braun, 1893a, 874, 879; 1900h, 6; 1901, 564.—Cobbold, 1860a, 11.—Dies., 1850a, 340–341.—Duj., 1845a, 448.—Jacoby, 564.—Cobbold, 1860a, 11.—Dies., 1850a, 340–341.—Duj., 1845a, 448.—Jacoby, 1900, 22, 23.—Jægers., 1898, 9, 10, 12, 14, 16.—Kowal., 1894, 2; 1895, 350.—Looss, 1899b, 586 (to Tocotrema); 1900, 607, type of Cryptocotyle.—Luehe, 1899, 539 (type of Cryptocotyle).—Mueh., 1898, 4, 19–21, 27, 80–83, figs. 6, 20, 26.—Stoss., 1892, 158; 1897, 10; 1898, 42.—Reported for Alca torda, Anas clangula, A. fusca, A. glacialis, A. hornschuchii, A. marila, Bucephala clangula, Colymbus cristatus, Fuligula marila, Harelda glacialis, Larus glaucus, Mergus merganser, M. serrator, Podiceps nigricollis.

confusum Looss, 1894a, 2, 17, 83, 84, 91, 92, 93, 95, 96, 99, 101–104, 105, 106, 108, 113, 126, 135, 137, 149, 150, 159, 167, 177, 179, 180, 181, 182, 191, 192, 197, 210, 211, 212, 214, 216, 217, 227, 228, 230, 234, 256, 263, pl. 2, figs. 33–35, pl. 8, figs. 164, 167 (D. clavigerum of Duj., 1845a, 404 (in Rana esculenta; Paris, Rennes), renamed; includes also D. clavigerum, ex parte, and endolobum e. p., of Pag., and clavigerum of Pachinger); 1893b, 810, 811; 1894d, 17; 1896b, 88, 91, 92, 93, 94, 95; 1898a, 460, fig. 2; 1899b, 556, 614, 616, 622, 623 (type of Prosotocus).—Luehe, 1901p, 169.—Mueh., 1898, 23.—Nickerson, 1900, 814, 815.—Stiles, 1901, 107

Stiles, 1901, 197.

conicum Polonio, 1859, teste Par. 1894, 627 [see also Diplost. conicum].—Anacker, 1892c, 94.

conis Crep. (misprint for D. conus Crep., in Perroncito, 1886, 250).

consunctum Rivolta, 1884, 26, for conjunctum, 1860.

conjunctum a Cobbold, 1860a, 8 (t. h. American Canis fulvus); 1860, 349; 1861, 118; 1864, 20–22, pl. 2; 1873, 81; 1876, 210; 1879b, 28, 30–34, 299, fig. 56; 1879b, 28, 30–34, 299, fig. 56; 1883, 401, fig. 16.—R. Bl., 1888a, 612-614 (two (all to date) human infections, review), figs. 314–315, 631; 1891, 610 (syn. of D. conus).—Braun, 1883a, 65–66; 1892a, 642, 663, 748; 1893, 349, 352; 1893a, 874, 882; 1893f, 386, 389, 425; 1894i, 1892a, 642, 663, 748; 1893, 349, 352; 1893a, 874, 882; 1893f, 386, 389, 425; 1894i, 605; 1895b, 147–148, 151, fig. 65; 1903, 3 ed., 163 (of Cunningham and McConnell, renamed Opisthorchis noverca), 164, fig. 113 (from Canis fulvus).—
Brunet, 1902a, 125.—Dunglison, 1893, 338.—Fitz, 1876b, 517.—Gamb., 1896a, 63.—Hackley, 1886, 519.—Hahn & Lefèvre, 1884a, 545–546 (= sinense).—Huber, 1896a, 578 (= O. noverca).—Ijima, 1889b, 146.—Jamison, 1897a, 74.—de Jong, 1887a, 57; 1896a, 2.—Ikamensky, 1900a, 17, 23.—Kholodk., 1898, 26, 29–30, pl. 11, fig. 13.—Kuech. & Zuern, 1881, 335.—Leuck., 1889, 355, figs. 162–163.—Lewis & Cunningham, 1872, 168.—Linst., 1873, 100.—Looss, 1899b, 565 (to Metorchis); 1905, 90 (of Lewis & Cunningham, syn. of O. noverca).—Maddox, 1867, 95.—Manson, 1901, 540 (in Homo); 1903, 3 ed., 635, fig. 98 (McConnell in an East Indian 1874, apparently of but slight paths fig. 98 (McConnell in an East Indian, 1874, apparently of but slight pathological importance).—McConnell, 1875, Aug. 21, 271-274, figs. 1-3; 1875, Oct., 772-780, figs. 1-3; 1876, Mar. 4, 343-344, figs. 1-3; 1876, Apr., 242-246, figs. 1-3 (case in man); 1876, July, 314-317; 1878, Mar. 30, 343, 476.—Moniez, 1896, 86, 122–125, 137.—Mont., 1893, 83 (conjunctum), 95.—Mosler & Peiper, 1894, 178.—Neumann, 1892, 529-530, fig. 284.—Perroncito, 1882, 285; 1886, 250, 297.—Rail., 1886, 297; 1893a, 365 to (Brachylaimus).—Schneidemuehl, 1896, 302 (kongenitum).—Simon, 1897, 209, 223—224.—Sons., 1889, 276, 277, 278, 280; 1889, 281 (syn. of D. conus Crep.).—Stiles, 1904i, 34 (to Metorchis).—Stiles & Hass., 1894e, 429—430, pl. 3, figs. 13–15 (consider form from man

 $[^]a$ All references to "Distoma conjunctum Cobbold" as a parasite of man are based upon the findings of Cunningham, McConnell, and Lewis, and refer in reality to Opisthorchis noverca, q. v.

distinct).—Stoss., 1892, 24-25 (syn. of D. conus).—Tyson, 1903, 3 ed., 1180.—Ward, 1895, 238 (man, dog), 328 (Homo), 341 (in Canis familiaris).—Wood & Fitz, 1897, 335.

conjunctum of Lewis and Cunningham, in Homo, India; see Opisthorchis noverca.

conoideum (Bloch, 1782) Rail., 1886, 297; 1893a, 366 (syn. of D. echinatum Zed.; includes excavatum Rud., 1819, echiniferum La Val., 1855, ? militare Rud., 1809; cf. Cerc. echinatoides Fil., C. echinifera, C. spinifera); in ducks, geese, swan, etc., and dog.—Ward, 1895, 341 (in Canis familiaris).

- conostomum Olss., 1876, 17–18, pl. 3, fig. 37 (in Coregonus oxyrrhynchus).—Braun, 1892, 45; 1892a, 663, 699, 711, 722; 1893a, 872; 1901b, 9, 10, 12.—Looss, 1899b, 605, thinks it a Spathidium; 1901l, 558.—Luehe, 1900aa, 507.—Mont., 1893, 95, 102.—Odhn., 1902, 65, 67.—Stoss., 1886, 22, 62 (in Coregonus oxyrrhynchus).
- constrictum Mehlis, in Creplin, 1846, 142, 143 (in Anas mollissima, A. nigra).— Crep., 1846, 142, 143.—Dies., 1850a, 397.—Stoss., 1892, 180.—Reported also for Anas fusca.
- constrictum Leared, 1862, 271–273, fig. 21 (in edible turtle).—Braun, 1892a, 569, 763, 764; 1893a, 876 (in Chelone mydas); 1893d, 467; 1899b, 715 (= D. mistroides).—Cobbold, 1879b, 35.—Laveran & R. Bl., 1895, 104.—Leuck., 1861, 79.—Looss, 1899b, 554, 656 (type of Hapalotrema).—Moniez, 1896, 143.—Mont., 1896, 144, renamed mistroides.—Stoss., 1895, 230.
- constructum Staff., 1900, 407, misprint for D. constrictum Leared.
- continuum Ariola, 1899, 6–8, pl. 1, fig. 6 (in Carcharias rondeletti; Genova).—Linst., 1903, 354.
- contortum Rud., 1819a, 118, 424–425 (in Orthagoriscus mola; Naples), 595.—Bellingham, 1844a, 427.—Braun. 1892a, 578; 1893a, 872, 910.—Cobbold, 1879b, 458.—Crep., 1837, 310, 311.—Dies., 1850a, 394; 1858e, 353 (in Orth. mola).—Drago, 1902, v. 15, 20; 1904, v. 79, 4 pp.. 1 fig.; 1904, v. 17, 720.—Drumond, —, 240.—Duj., 1845a, 469.—Kroyer, 1852–53a, 745 (in Orth. mola).—Lint., 1898, 528–530, pl. 48, figs. 3–7; 1901, 415 (in Mola mola), 422, 466.—Looss, 1894a, 165, 190; 1899b, 631 (type of Accacelium).—Luehe, 1900, 487.—Mont., 1888a, 7, 8, 12; 1888, 198, 199; 1891, 110; 1893, 13, 14, 19, 22, 23, 27, 29, 30, 32, 33, 34, 36, 37, 40, 44, 46, 47, 61, 64, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 90, 93, 94, 95, 97, 102, 107, 108, 113, 115, 119, 120, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 150, 152, 169, pl. 1, fig. 12, pl. 3, figs. 16, 22, 25, 26, 32, pl. 4, figs. 33–37, pl. 8, figs. 123 (type of Accacelium); 1896, 151, 153.—Olss., 1876, 17.—Sons., 1890, 140 (in Orth. mola L.); 1891, 265.—Stoss., 1896, 190, to (Podocotyle).
- conum Huber, 1896a, 578 (refers to Stoss., syn. of D. conjunctum=O. noverca); see conus Crep.
- conus Crep., 1825, 50–53 (in Felis catus dom., Canis vulpes): 1837, 310; 1839, 286, 288.—Baillet, 1866b, 105 (includes Amphist. truncatum Rud.).—R. Bl., 1891, 610 (syn. D. conjunctum).—Braun, 1892a, 580, 613, 663, 699; 1893a, 875; 1893f, 382, 383, 384, 386, 387, 388, 389, 390, 391, 424 (of Sons. in dogs and cats in Italy, syn. of D. felineum); 1893g, 802 (= Amphist. truncatum Rud.).—Cobbold, 1875n, 819 (Distoam); 1884g, 976.—Dies., 1850a, 404 (syn. of Amphist. truncatum); 1858e, 349–350 (in Canis vulpes; Felis catus dom.) (to Dist.).—Duj., 1845a, 331, 440.—Hahn & Lefèvre, 1884a, 538.—Hilgendorf, 1870, 566–567.—de Jong, 1896a, 2, 3, 4, 6.—Kamensky, 1900a, 5.—Leuck., 1863a, 589.—Linst., 1878a, 43, 44.—Moniez, 1896, 136, 139.—Sons., 1889, 273, 276, 277, 278, 279, 280, 281 (syns. Amphist. truncatum Dies., D. campanulatum Erc., D. conjunctum Cobbold, D. felineum Rivolta, D. truncatum Erc.); 1890, v. 7 (3), 15 Feb., 134–138.—Stoss., 1892, 24, 25 (includes D. truncatum Erc., D. conjunctum Cobbold, Amphist. truncatum Dies.) (in Vulpes vulgaris; Canis fulvus, America; Canis familiaris, Pisa; Felis domestica).—Wagener, 1857, 102, pl. 22, figs. 1, 2; 1858, 102.—Ward, 1895, 341 (see D. truncatum) (in Canis familiaris).
- conus of Gurlt, 1831, 193, 373-375, pl. 8, figs. 34-36 (nec Crep., 1825).—Braun, 1893f, 383, 384, 385, 386, 387, 389, 390, 422 (syn. of D. felineum); 1903, 3 ed., 157 (syn. of Opisthorchis felineus).—Dies., 1858e.—de Jong, 1896a, 10.—Rail., 1893a, 361 (= D. fel.).—Ward, 1895, 243 (syn. of D. fel.), 328 (in Homo), 341 (in Canis fam.); 1903, 869 (syn. of Opisth. fel.).

conus of Sons., 1889 (sub D. truncatum), see felineus.—Braun, 1892, 580, 613, 663, 699; 1893, 874; 1893e, 348, 349, 350, 352; 1893f, 386, 424.

- convolutum Brand., MS. in Braun, 1901f, 565, in Platalea ajaja; 1902b, 24 (syn. of Mesaulus grandis).
- cornifrons Leidy, 1878, 382–383 (in Donax fossor).
- cornu Zed., 1800a, 180–181, pl. 2, fig. 1–3 (in Ardea cinerea; Europe).—Dies., 1850a, 327 (to Monost.).—Nord., 1840, 627 (to Amphist.).—Rud., 1809a, 346 (to Amphist.).
- coronarium Cobbold, 1861e, 119 (in Alligator mississippiensis; died in England);
 1864, 17; 1879b, 455 (All. miss.).—Braun, 1892a, 699; 1899, 630; 1901b, 34.—
 Looss, 1896b, 114, 117, 118; 1899b, 575, 581, to Acanthost.—Luehe, 1899, 532.—
 Par., 1896, 7-9, fig. 3, to (Echinost.).—Pavesi, 1881, 294.—Stoss., 1895, 230;
 1899.—Also reported for Crocodilus americanus.
- coronatum Rud., 1819a, 686 (in Didelphis? virginiana; Brazil).—Braun, 1900,
 27; 1901e, 318, 319, 320, 323.—Cobbold, 1879b, 432.—Dies., 1850a, 400 (to Rhopalophorus).—Duj., 1845a, 425.—Looss, 1899b, 581.—Stiles & Hass.,
 1898a, 93, 96 (type of Rhopalias).—Stoss., 1892, 30-31 (to Echinost.).
- coronatum Wagener, 1852, 567-569, figs. 4-6 (in Corvina nigra; Nice).—Braun, 1892a, 579, 583, 643; 1901b, 34, 1901, 568; 1902b, 28, 30, 31.—Carus, 1884, 127.—Dies., 1856, 64; 1858e, 352 (in Corvina (Sciæna) nigra).—Looss, 1899b, 578, 581, 582, 583, type of Anoiktost.—Mont., 1893, 84 (? of Dies. or of Wag.).—Stoss., 1885, 157; 1886, 37.
- coronatum Rentsch, 1860, 38, pl. 11, fig. 2a, 41 (in Gasterosteus spinachia).
- corpulentum Lint., 1905, 327, 334, 378, 382, figs. 180–182 (in Lagodon rhomboides, Orthopristis chrysopterus; Beaufort, N. C.).
- corrugatum Duj., 1845a, 409–410 (in Sorex tetragonurus; Rennes), to (Brachylaimus).—Braun, 1901e, 341, 342.—Cobbold, 1879b, 296.—Dies., 1850a, 360.—Stoss., 1892, 13.
- corvinæ Stoss., 1886, 46–47, pl. 7, fig. 29 (in Corvina nigra; Triest); 1886, 36; 1898, 56; 1899, 15.—Braun, 1892a, 579; 1902b, 31.—Looss, 1899b, 578, 581.—Mont., 1893, 83.
- coryphænæ (Bosc., 1802) Rud., 1809a, 436–437 (includes Fasc. fusca Bosc., 1802; F. cory. Bosc., 1802; F. caudata Bosc., 1802; F. coryphænæ hippuridis Tilesius); 1819a, 122, 685 (in Coryphæna hippuris).—Baird, 1853a, 60 (syn. of Hirudinella ventricosa).—Cobbold, 1879b, 460 (coryphenæ), 461 (coryphænæ).—Dies., 1850a, 367 (syn. of D. clavatum Rud.), 373 (syn. of D. tornatum Rud.).—Poir., 1885, 6.

coryphenæ Cobbold, 1879b, 460, for coryphænæ.

coryphana Cobbold, 1879b, 461, for coryphana.

[costæ D. Valle (a tunicate).]

cranum Huber, 1896a, 575, misprint for crassum=buskii.

crassicaudatum Busch, 1851, 99–100, pl. 15, fig. 13 (in Saggita).—Wagener, 1860, 166.
crassicolle Rud., 1809a, 378–379 (in Salamandra atra) (Fasc. salamandra Freelich, renamed); 1819a, 102, 385–386.—Baird, 1853a, 52.—Braun, 1892a, 603, 681; 1893a, 881; 1895b, 128, fig. 45.—Cobbold, 1860a, 18.—Crep., 1837, 316; 1846, 147, 148.—Dies., 1850a, 356; 1858, 339 (in Triton marmoratus).—Duj., 1845a, 404–405, to (Brachycœlium).—Kampmann, 1894b, 454, 457.—Kerbert, 1881a, 536, 573.—Linst., 1887, 97, 98.—Looss, 1894a, 1, 82, 83, 84; 1899b, 611, 612, 613, 614, 618; 1902m, 705, 769, 773, 774, 816, 821, 822 (type of Brachycœlium).—Luehe, 1899, 536; 1900, 562, 563.—Minot, 1878a, June 11, 1–12, pl. 1, figs. 1–17.—Mont., 1888a, 25, 38.—Olfers, 1816, 45.—Par., 1896, 2; 1896, 13–16 to (Dicrocœlium) (includes D. enterarchos Fil., D. salamandrinæ perspicillatæ Sons.).—Pontallié, 1853, 211–219.—Stiles, 1901, 196, 198, 199, 200, 201, 202.—Stiles & Hass., 1898a, 83, 97 (type of Brachycœlium).—Stoss., 1889, 63; 1895, 214–215; 1897, 9; 1898, 32.—Ziegler, 1883, 545.—Also reported for Anguis fragilis, Rana temporaria, Salamandra maculosa, Salamandrina perspicillata, Triton alpestris, T. cristatus, T. punctatus, T. tæniatus.

crassiusculum Rud., 1809a, 373, 408–409, 429 (in Falco melanaetus) (Plan. bilis Braun renamed); 1819a, 112.—Braun, 1891d, 424 (in Circus æruginosus); 1892a, 583, 584, 610, 663; 1893a, 875; 1893, 353; 1893f, 426; 1901, 561; 1902b, 8, 10.—Cobbold, 1860a, 15.—Crep., 1837, 326; 1846, 129.—Dies., 1850a, 376 (includes D. felleum falconis chrysaéti Viborg); 1858e, 353 (in Falco buteo).—Duj., 1845a, 441–442.—Linst., 1873, 99.—Looss, 1896b, 58; 1899b, 677 (to

Metorchis .—Mont., 1893, 15.—Much., 1898, v. 1, 21, 23–24, 87–89, fig. 1.— Olfers, 1816, 46.—Stoss., 1890, 50; 1892, 151.—Wedl, 1857, 244–245, pl. 1, fig. 4.—Also reported for Aquila chrysactos, Archibuteo lagopus, A. vulgaris, Buteo vulgaris, Circus æruginosus, C. rufus, Colymbus septentrionalis, Falco albicilla. Mergus serrator, Nyctea nivea.

crassum Sieb., 1836, 234 (in Hirundo urbica) ("Hoden — zwischen dem Porus anticus und posticus verborgen"); 1848, 143.—Braun, 1893a, 874; 1902b, 46, 94–95.—Crep., 1837, 327, 328; 1846, 133.—Dies., 1850a, 397.—Leuck., 1863a, 587.—Looss, 1899b, 557.—Poche, 1907, 126.—Stoss., 1892, 156.—Also reported for Hirundo demostical demostrical demostri for Hirundo domestica.

 crassum Cobbold, 1860a, 5–6 (buskii Lankester, renamed; in Homo); 1864a,
 192–194, fig. 42; 1866, 6; 1875i, 423; 1876g, 285–296, 1 fig.; 1876h, 210, 211;
 1876i, 297–305, 1 fig.; 1879b, 17, 20–28, 421, 480, fig. 2; 1883, 401; 1884g, 976.— 1876i, 297–305. 1 fig.: 1879b. 17, 20–28, 421. 480, fig. 2; 1883, 401; 1884g, 976.— Aitken, 1866, 804. 839; 1872, 146, 205; 1874, 58.—Anders, 1903, 6 ed., 1245.— Aschoff, 1892, 495.—Bassi, 1875b. 507.—Biermer, 1863a, 395 (grassum).— Braun, 1883a, 62–63; 1902b. 95; 1903. 3 ed., 153 syn. of Fasciolopsis buski; 1906, 159, fig. 90.—Brunet, 1902a, 125.—[Budd, 1845, 399, fig. 20; 1846, 390, fig. 20; 1846, 444; 1852, 484, fig. 20; 1857, 494, fig. 20.]—Busk, 1859.—Clarke, 1894b, 244–245, fig. 21.—Dav., 1877a, lxxvii, 258–259.—Dunglison, 1893, 338, 820.—Eichhorst, 1901a, 301.—Gamb., 1896a, 63.—Hackley, 1886, 518, fig. —, 882.—Hahn & Lefevre, 1884a, 541–542, 544.—Harley, 1864a, 62.—Hoyle, 1890, 538.—Huber, 1896a, 575 (cranum).—Jackson, 1888, 653.—Jacob, 1891a, 1287.—Jamieson, 1897a, 73.—Kholodk., 1898, 30, pl. 11, fig. 15.—Leidy, 1891, 234; 1904a, 239–241 includes Fasc, magna,—Leuck., 1863a, 526, 586–588, fig. 196; 1876, v. 2 (3), 870, 871.—Looss, 1899b, 557, to Fasciolopsis; 1905, 110 isyn. of Fasciolopsis buski; 1907, Feb. 1, 124.—Manson, 1880, 1905, 110 syn. of Fasciolopsis buski : 1907, Feb. 1, 124.—Manson, 1880, May. 66: 1901. 440. 543: 1903. 3 ed., 631. 639.—Moniez, 1896, 114. 115.— Mosler & Peiper, 1894, 176.—Oppenheim, 1900, 181.—Packard, —, 522.— Rail., 1893a, 363–364 includes D. buskii Lankester, 1857; D. Dicrocœlium) buski).—Roberts. 1888. 673.—Simon. 1897, 223 [cranum].—Sinciair, 1891, Dec. 26, 1468.—Smith. 1881. July. 14 [=? Tænia solium or T. saginata].—Stoss., 1892, 27 [syn. of D. rathouisi Poir.].—Verrill, 1870, 171.—Vogt. 1878, 10. 14.—Wagner. 1883, 121.—Walker. 1891, 5 Oct.. 1205. figs. 1—2.—Ward, 1895, 328 [syn. of D. buskii]; 1903, 867 [syn. of Fasciolopsis buski].—Wood & Fitz, 1897, 335.

crassum Olss., 1876, 25, pl. 4, figs. 27–28 in Hirundo urbica) [nec Sieb., 1836].— Braun, 1902b, 46 (? syn. of Plagiorchis maculosus), 94, 95.—Looss, 1899b, 557 (? syn. of maculosum Rud.).

 crassum Cobbold, of Leidy, 1891, p. p., 234; 1904a, 239-241 in Cervus virginianus.
 Bos taurus .—Stiles, 1898a, 51 syn. of Fasc. magna .—Ward, 1895, 253, 332, 335. 338 (in Bos taurus. Ovis aries, Equus caballus) (p. p. syn. of Fasc. magna; 1903. 866.

crenatum (Rud., 1802 [nec Frælich, 1802]) Rud., 1809a, 387 401, 404-405, pl. 5, fig. 1 (in Gasterosteus aculeatus): 1810a, 376; 1819a, 110 (syn. of D. appendiculatum Rud.).—Dies., 1850a, 371 (syn. of D. app.).—Lander, 1904a, 1 (to Hemiurus.—Luehe, 1901, 399 extensive synonymy, 401 (not Rud., 1810, 376, from Salmo salar), 402.—Mont., 1891, 497, 522.—Odhn., 1905, 352 (type of Brachyphallus).—Oliers, 1816, 46.—Wagener, 1860, 166, 183 (in Gast. ac.).

crenatum Mol., 1859, 840-842, pl. 1. fig. 3 (in Centrolophus pompilius: Padua .-Braun, 1893a, 873.—Carus, 1884, 125.—Looss, 1899b, 640 (to Hemiurus).— Luehe, 1901, 399 (not Rud.), 402 (cf. D. excisum Rud.).—Odhn., 1905, 352 (to Brachyphallus).—Stoss., 1886, 14; 1898, 27.

[cristallinum Ren. | a tunicate |.]

cristatum Rud., 1819a, 117-118, 422-423 (in Stromateus fiatola: Arimini).—Carus, 1884, 128.—Cobbold, 1860a, 37.—Crep., 1837, 311.—Dies., 1850a, 393.—Duj., 1845a, 432.—Mont., 1893, 193.—Stoss., 1886, 47.—Also reported for Hippocampus brevirostris.

croaticum Stoss., 1889, 183-184, pl. 4, figs. 5-7 (in Carbo graculus; Fiume, Croazia); 1890. 131: 1892, 165 to Echinost. .—Braun. 1891d. 421: 1892a, 763, 765: 1893a.

911.—Reported for Carbo cormoranus.

crocodili Poir., 1886, 30–32. pl. 1. figs. 4–5 in Crocodilus siamensis.—Braun, 1892a,
 722.—Looss. 1899b, 575, 578.—Mont., 1893, 43, 83, 86, 95, 102.

crotali Humboldt [an arachnoid. now in Porocephalus].—Dies., 1836, 21: 1850a, 612 (syn. of Pentast. proboscideum).—Leidy, 1850b, 97 (syn. of Pentast.

prob.); 1904a, 32.—Nord., 1840, 645 (to Porocephalus).—Rud., 1809a, 433.—Stiles, 1891a.

crotali durissi Rud., 1809a, 433 (for crotali), 434; 1814a, 103; 1819a, 124 (syn. of Pentast. proboscideum).

crucibulum (Rud., 1819) Duj., 1845a, 435, to (Crossodera) (in Muræna conger; M. cassini).—Dies., 1850a, 322 (to Monost.).—Tennent, 1906, 640 (to Gasterost.).

cryptobothrium Ben., 1870, 1871a, 31, pl. 5, fig. 16 (in Trigla gurnardus; Belgium). crystallinum Rud., 1819a 100, 380–382, 595 (in Rana esculenta, R. temporaria, Bufo viridis, B. igneus, Vipera berus; Berlin).—Bettend., 1897, 308, 311, fig. 28, fig. 4; 1897a, 4, 7, pl. 1, fig. 4.—Brand., 1888a, 14.—Braun, 1893a, 875.—Dies., 1850a, 352 (to Dist.); 1858e, 338.—Duj., 1845a, 453–454.—Hannover, 1864a, 3 (chrystallinum).—Pag., 1857, 39, 46, pl. 4, fig. 6 (in Frosch).

cteniceps Leidy MS. in Stiles & Hass., 1894d, 249 (manuscript name found in bottle containing parasite of Muskrat, published as D. echinatum; see Leidy,

1888, 126–127) (in Fiber zibethicus).

cucumerinum Rud., 1809a, 360–361 (in Avis riparia; gen. sp. incert.); 1819a, 94.— Braun, 1899f, 465–468 (is a Monost.); 1901, 561.—Dies., 1850a, 338 (to Dist.).— Olfers, 1816, 44.—Stoss., 1902, 9, 32 (to Typhlocœlum).

Cuneatum Rud., 1809a, 358–359 (in Otis tarda); 1819a, 93 (Greifswald).—Anacker, 1892c, 94.—Braun, 1893a, 877; 1893d, 467; 1901, 13, 15; 1901, 561; 1902b, 69, 72, 73, 74, 79 (=D. ovatum autt. (nec D. ov. Rud.); Fasc. ov. Rud. part) (to Prosthogonimus).—Cobbold, 1860a, 9.—Crep., 1846, 134.—Dies., 1850a, 336 (to Dist.)(in Otis tarda; (Gryphiæ, Rud., intestine)(in Pavo cristatus, Gurlt).—Duj., 1845a, 445.—Looss, 1899b, to Prosthogonimus.—Luehe, 1905, 159.—Olfers, 1816, 44.—Schneidemuehl, 1896, 303 (kuneatum).—Stoss., 1892, 180.

cuspidatum Looss, 1896b, 97-101, 104, pl. 7, figs. 64-65 (in Milvus parasiticus; Matarieh); 1899b, 578, 581, 584 (type of Centrocestus).—Braun, 1901b, 34.

cyclopteri Rud., 1809a, 438 (in Cylopterus lumpus); 1819a, 121.—Dies., 1850a, 373 (syn. of Dist. reflexum), 398.—Stoss., 1886, 47.

cygni oloris Dies., 1858e, 344 (syn. of D. echinatum), based on Bellingham, 1844a,

cygnoides Zed., 1800a, xxiv, 163, 175-176 (in Rana esculenta); 1803a, 213.—Baird, 1853a, 51.—Ben., 1858a, 1861a, 189, 203, 214.—Bensley, 1897a, 326-331, figs. 1-6 (2 forms); 1897, 293.—Braun, 1883a, 56; 1892a, 569, 634, 635, 698, 711, 712, 1-6 (2 10rms); 1897, 293.—Brauth, 1885a, 56; 1892a, 509, 654, 659, 698, 711, 112, 761, 762, 764, 768, 773, 776, 780, 782, 783, 785, 788, 790, 797, 798, 806; 1893a, 854, 865, 876, 882; 1893d, 467; 1895b, 18, 129; 1899g, 492 to Phyllodist.; 1901b, 9, 10, 12; 1902b, 22.—Cobbold, 1858b, 162; 1860a, 15; 1872b, 91; 1879b, 49, 454.—Crep., 1837, 310, 323, 324, 326.—Desmonceaux, 1868, 21.—Dies., 1850a, 342 (includes D. hylæ Rud.); 1858d, 272; 1858e, 334 (to Dist.) (Cerc. (Acanthocephala) macrocerca) (adult in Pelophylax esculentus; Bombinator igneus; Dendrohyas viridis; Rana temporaria; R. pipiens; R. palustris; R. halecina; Salamandra maculata; S. (Amblystoma) rubra; S. salmonea; young in Pelophylax esculentus; larva in Cyclas cornea).—Duj., 1838a, 7 (embryo); 1838c, 47; 1845a, 396–397 to (Dicroccelium).—Fil., 1855b, 24, 25; 1857c, 32.— Florance, 1866a, 6, 12.—Fraip., 1880c, 418.—Gamb., 1896a, 72.—Hahn & Lefèvre, 1884a, 516.—Kampmann, 1894b, 446.—Kerbert, 1881a, 566, 575.— Kowal., 1894, 3.—Leidy, 1851b, 207; 1856, 44; 1904a, 48-49 (in Rana pipiens, Salamandra maculata, S. (Amblyostoma) rubra), 87 (in R. palustrus, R. halecina, S. salmonea).—Leuck., 1863a, 511; 1886d, 40.—Linst., 1873, 1 (young= Cerc. macrocerca Fil.); 1887, 97, 101.—Looss, 1885b, 24; 1892a, 125; 1893b, 813, 815; 1894a, 2, 23, 56–64, 82, 110, 125, 127, 136, 137, 138, 153, 154, 159, 162, 168, 169, 174, 178, 182, 184, 189, 200, 201, 204, 206, 211, 215, 216, 227, 230, 244, 251, 253, 256, 269, 275, pl. 1, figs. 23–26, pl. 6, figs. 124–132 (in Bombinator igneus; Hyla arborea; Rana esculenta; R. temp.); 1896b, 46; 1899b, 605–606, type forgodera, 607; 1902m, 784, 807, 810, 846, 851.—Loschge, 1785, 10–14, pl. 1, figs. a-e.—Luehe 1900, 558; 1901, 58.—Miescher, 1840c, 39–42 (in R. esc.).—Mol., 1859, 833 (in Pelophylax esculentus; Padua).—Mont., 1888a, 40, 80 (cignoides); 1893, 82, 148, 149; 1896, 166.—Moul., 1856, 48, 99, 161 (Cerc. macrocerca is larva of; teste Fil.).—Mueh., 1898, 27–28.—Nord., 1840, 617 (sygnoides, misprint).—Odhn., 1902, 64, 65.—Olfers, 1816, 44.—Osborn, 1903, 256.—Pachinger, 1883, 46 pp.—Pag., 1857, 28, 30, 44–46, 47, 52, pl. 6, fig. 1 (in green frog).—Par., 1887, 60; 1887, 332–333; 1887, 490.—Rossbach, 1906, 377, 388.—Rud., 1809a, 367-368; 1819a, 96, 370-371.—Schuberg, 1875, 169.—Sieb.,

1835, 66, 82; 1842, 298.—Sons., 1890, 136; 1893, 187 (cygnoidis) (in Rana esc.); 1893, 217.—Ssinitzin, 1905, 33, 34; 1906, 682 (in Rana temporaria, R. esculenta; Warschau) (of Looss, named Gorgodera loossi; of Pag., renamed G. pagenstecheri), 684.—Staff., 1900, 405, 409; 1902, 18 Dec. (American), 411, 412, 421, 482, to (Gorgodera) (in frogs and toads), 895; 1903, 15 Dec., 901; 1905, July, 51-52.—Stiles, 1896, 205.—Stoss., 1889, 60.—Valentin, 1841, 51-54.—Will.-Suhm, 1870, 4.—Wagener, 1857, 26, 27, 29–30, 34, 35, 40, 42, 43, 44, 45, pl. 30, fig. 1, pl. 31, fig. 1, pl. 32, fig. 36A, 7.—Walter, 1866, 64.—Reported also for Dendrohyla viridis, Bombinator sp.; Bufo igneus, B. vulgaris, Cyclas, Limnæa sp.; Piscidium; Rana catesbiana, R. clamitans, R. virescens, Salamandra maculosa.

cygnoides var. A. of Bensley, 1897a.—Staff., 1902, 419-421 (svn. of Gorgodera amplicava).

cygnoides var. B. of Bensley, 1897a. —Staff., 1902, 417 (syn. of Gorgodera simplex). cygnoides ranæ Wagener, 1857, 19-24.

cygnoidis Sons., 1893, 187, for cygnoides.

cylindraceum Zed., 1800a, 164, 188-190, pl. 4, figs. 4-6 (in Rana esculenta); 1803a, nuraceum Zed., 1800a, 164, 188-190, pl. 4, ngs. 4-6 (In Rana esculenta); 1803a, 217.—Baird, 1853a, 51.—Bellingham, 1844a, 424.—Ben., 1858a, 1861a, 193.—Bettend., 1897a, 4, 5, 7, 10, pl. 1, fig. 6; 1897, 308, 309, 311, 312, 314, pl. 28, fig. 6.—Blochmann & Bettend., 1895a, 217, fig. 2.—Brand., 1891c, 730; 1891d, 7, 12.—Braun, 1890d, 568; 1892a, 568, 586, 589, 593, 597, 599, 602, 608, 635, 640, 641, 643, 671, 677, 682, 683, 684, 685, 698, 703, 707, 717, 719, 726, 729, 733, 737, 751, 754, 759, 760, 762, 763, 768, 773, 780, 783, 785, 788, 790, 806; 1893a, 865, 876, 882, 910.—Cobbold, 1858b, 162 (in Rana temporaria); 1860a, 17.—Crep., 1857a, 230, 236, 1858a, 266, 268 (included Exception of the company of th 1837, 310, 323, 326.—Dies., 1850a, 368 (includes Fasc. subclavata); 1858e, 351 (in Rana oxyrrhinus; R. platyrrhinus).—Duj.,1845a, 395–396.—Galli-Valerio, 1898n, 371–373; 1899b, 186–187; 1901c, 365 (in Rana esculenta).—Gamb., 1896a, 72.—Hausmann, 1897b, 14.—Hofmann, 1899, 196.—Jackson, 1888, 650 (reproductive system).—Knoch, 1862, 101.—Kowal., 1894, 3; 1895, 372–390, pl. 8, fig. 9.—Linst., 1887, 97; 1890f, 173–191, 2 pls., figs. 1–39; 1891a, 241–242; 1904, 254.—Looss, 1885b, 6; 1893b, 811, 815; 1894a, 1, 2, 7, 37 (cglindraceum), 64–71, 72 (of Pachinger, 1888, syn. of D. variegatum), 73, 74, 75, 76, 77, 85, 86, 87, 115, 119, 125, 127, 135, 139, 144, 146, 148, 159, 163, 167, 175, 180, 181, 191, 192, 194, 199, 206, 215, 220, 221, 227, 230, 242, 268, pl. 2, figs. 39–42, pl. 7, figs. 147–152, pl. 8, 163 (includes Fasc. subclavata) (in Hyla arborea; Rana esc.; 141-152, pl. 8, 165 (Includes Fasc. subclavata) (In Hyla arborea; Rana esc.; R. temp.); 1899b, 556, 559-600 (type of Haplometra); 1902m, 620.—Luehe, 1899, 533.—Mont., 1888a, 73; 1893, 25, 27, 33, 36, 42, 43, 48, 61, 83, 91, 102, 106, 107, 108.—Moul., 1856a, 49.—Mueh., 1898, 26.—Nord., 1840, 617, 620 (to Fasc.).—Olfers, 1816, 46.—Olss., 1876, 14; 1893, 10.—Pag., 1857, 43, pl. 5, figs. 3, 4 (in brown frogs).—Rhumbler (1895), v. 4, 553-554.—Rud., 1809a, 336, 349, 393-394; 1819a, 106.—Schauinsland, 1882, 496, 497.—Sieb., 1835, 82; 1852, 116.—Sons, 1893, 187 (in Rana en).—Staff, 1002, 000.—Stage, 1893, 187 (in Rana en).—Staff, 1002, 000.—Stage, 1893, 187 (in Rana en).—Staff, 1002, 000.—Stage, 1893. 1852, 1, 16.—Sons., 1893, 187 (in Rana sp.).—Staff., 1902, 909.—Stoss., 1889, 65.—Swammerdam, 1737, 315.—Volz, 1899, 232.—Walter, 1866, 64.—Ziegler, 1883, 488; 1883, 546, pl. 33, fig. 26.—Reported for Bufo yulgaris; Ilybius fuliginosus; Limnæa ovata; Rana fusca; R. oxyrhinus; R. palustris: R. platyrhinus.

cylindraceum Zed. of Pachinger, 1888.—Looss, 1894a, 71, 72 (syn. of D. variegatum Rud.).

cylindricum (Gœze, 1782) Mayer, 1841a, 26, 34.—Looss, 1894a, 71 (syn. of D. variegatum).

cylindricum ranæ Mayer, 1841a, 18–19, pl. 3, figs. 13, 17.

biforme Rud., 1819a, 96, 371 (in Testudo mydas; Arimini).—Braun, 1893a, 876 to Phyllodist.; 1893d, 467; 1899b, —, 715, 720-721; 1899g, 492 (to Phyllodist.); 1901b, 9, 10, 11, 12; 1902b, 19.—Carus, 1884, 129.—Cobbold, 1860a, 18.—Dies., 1850a, 342 (in Halichelys atra).—Duj., 1845a, 451.—Looss, 1899b, 605 (to Spathidium); 1901b, 202, 203, 204, 205, 207 (resembles Spathidium—Phyllodist., but lacks pharynx, hence was made type of a new genus, Plesiochorus); 1901, 557; 1902m, 411, 469, 476, 477, 478, 481 (to Plesiochorus).—Mont., 1896, 165 (cimbiforme).—Odhn., 1902, 37; 1902, 64, 65, 67.—Par., 1846.—Sons., 1893, Feb. 5, 2; 1893, 183, 184, 185 (in Chelonia caretta).—Stoss., 1895, 38, pl. 4, fig. 1; 1895, 215-216; 1897, 9; 1898, 34-35.—Reported for Chelone caretta; C. mydas; Chelonia midas; Thalassochelys caretta; T. corticata). cymbiforme Rud., 1819a, 96, 371 (in Testudo mydas; Arimini).—Braun, 1893a, 876

cymbuliæ delle Chiaje (1841a), 109, fig. 29?.—Pag., 1862, 298.

cyprinaceum Zed., 1800a, 164, 181–183 (intest. Cyprinorum).—Dies., 1850a, 341 (syn. of D. globiporum).—Looss, 1894a, 41 (syn. of D. glob.); 1902m, 763 (and Sphærost. glob.).—Rud., 1809a, 365, 367.—Stiles, 1901r, 168.

cyprini idi (peritonei) Dies., 1858e, 367 (based on Duj., 1845a, 463) (as ? syn. of Tetracotyle echinata) (in Leuciscus idus, Acerina cernua).

cysticola phalangii opilionis Dies., 1855, 64, based on Crep., 1846, 156, see cystidicola.

cysticum Crep., 1846a, 159 (in Planorbis sp.), refers to Henle, 1835, 597.—Also reported for Planorbis marginatus, Nephelis vulgaris.

cystidicola Crep., 1846a, 156 (in Phalangium opilio); 1851, 304.—Dies., 1858e, 356 (in Phal. opilio).

dactyliferum Braun, 1892a, 568, for dactylipherum.

daetylipherum Poir., 1885, 10, pl. 23, fig. 3 (in Argonauta).—Braun, 1889a, 395 (daetyliferum); 1892a, 568 (daetyliferum); 1893a, 873.—Buttel-Reepen, 1902, 167, 172, pl. 6, fig. 14.

deflectens Rud., 1819a, 677–678, 745 (in Sylvia sp., Motacilla sp.; Brazil).—Braun, 1901, 561, 563; 1902b, 101 (to Dicrocœlium), 102.—Cobbold, 1860a, 12.—Dies., 1850a, 347 (in Thryothorus hypoxanthus; Brazil).—Duj., 1845a, 433-444.—Stoss., 1892, 180; 1892, 38.

delicatulum Rud., 1809a, 373–374 (in Anas sponsa); 1819a, 99.—Blainv., 1828a, 585 (to Fasc.).—Braun, 1893a, 875; 1901, 561.—Cobbold, 1860, 10.—Dics., 1850a, 352–353.—Duj., 1845a, 449.—Kowal., 1897b, 1, 2 [reprint]; 1898h, 132.—Olfers, 1816, 44.—Odhn., 1900, 19.—Stoss., 1892, 180.—Also reported for Cygnus olor.

deliciosum Olss., 1893, 10, pl. 1, figs. 16-18 (in Larus argentatus).—Odhn., 1900,

12, 13; 1905, 311 (to Gymnophallus).

delphini Poir., 1886, 34–36, 37, pl. 4, figs. 1–3 (in Delphinus delphis).—Braun, 1892a, 673; 1893a, 874, 910; 1893, 354; 1900g, 251.—Looss, 1894a, 204; 1899b, 560 (to Brachycladium).—Mont., 1888a, 12, 38; 1893, 44, 45, 83, 86, 102, 107.—Stiles, 1895l, 219, pl. 8, figs. 1–3.—Stoss., 1892, 10 (to Cladocœlium).

dendricum Ben., 1870, 36, for dendriticum.

dendriticum Rud., 1819a, 93, 364–365 (in Xiphias gladius).—Ben., 1870, 36.— Carus, 1884, 131.—Cobbold, 1860, 22.—Darr, 1902, 698.—Dies., 1850a, 336.—Duj., 1845a, 460.—Jacoby, 1900, 10, 11.—Kroyer, 1838—40a, 597 (in Xiphias gladius).—Looss, 1899b (to Dicrocœlium).—Par., 1896, 16–18, fig. 6 to (Dicrocœlium).—Stoss., 1886, 47.

dentatum Lint., 1900a, 269, 283, 289, 294, pl. 39, figs. 64–67 (in Paralichthys dentatus); 1901b, 415, 421, 451, 483; 1905, 328, 334, 369, 372, 374, 396, 411, 413, 415 (in Coryphæna equisetis, Lophopsetta maculata, Micropogon undulatus, Paralichthys albiguttus, P. dentatus, Pomatomus saltatrix, Rachycen-

tron canadus; Beaufort, N. C.).—Odhn., 1905, 331.

denticulatum (Rud., 1802) Rud., 1809a, 424–425, pl. 5, fig. 3 (in Anas clypeata, Sterna hirundo) to (Echinost.); 1819a, 116, 419.—Crep., 1837, 311.—Dies., 1850a, 391–392.—Duj., 1845a, 430.—Olfers, 1816, 47.—Stoss., 1892, 170 to Echinost.—Also reported for Anas sponsa.

depressum Polonio, 1859 (in Triton cristatus; Padua) teste Par., 1894, 149.— Braun, 1892a, 579, 663.

depressum Stoss., 1883, 118, pl. 3, fig. 10 (in Dentex vulgaris; Triest); 1886, 24; 1898, 39-40.—Carus, 1884, 130.

detruncatum (Braun, 1899) Braun, 1900h, 28 (in Mycteria americana), 33 to Clinest., 35 (from Tantalus loculator) to (Clinost.), 39.

dicorynum Dies., 1850a, (359, D. affine), 680, affine renamed (in Lampris guttatus).— Ben., 1870, 34.—Kroyer, 1838–40a, 598.—Stoss., 1886, 47.

dictyotus Mont., 1893, 156 (includes D. reticulatum Looss, 1885 [nec Wright]).— Braun, 1900, 26.—Looss, 1894a, 171.—MacCallum, 1899, 705.—Stiles & Hass., 1898a, 86 (D. reticulatum Looss, 1885 [nec Wright, 1879], (type of Mesogo-

didelphidis Par., 1896, 3–5, fig. 1a (in Didelphys azaræ).—Luehe, 1899, 532.—Stoss., 1904, 2 (didelphydis).

didelphydis Stoss., 1904, 2, for didelphidis.

diesingi Braun, 1901, 561, 563, for diesingii.

- diesingii Cobbold, 1860a, 14 (D. cochlear Dies., renamed; in Sterna cantiaca, S. minuta; Brazil).—Braun, 1901, 561, 563 (diesingi); 1902b, 56, 58 (syn. of Microlistrum cochlear (Dies., 1850a)).—Stoss., 1892, 179 (syn. of D. cochlear).
- diffusocalciferum Gastaldi, 1854, 5–6, pl. 1, figs. 4–5 (in Rana esculenta).—Braun, 1893a, 870.—Cobbold, 1860a, 18.—Dies., 1855, 64 footnote (D. diffusocalciferum ranæ esculentæ); 1858e, 336 (in Pelophylax esculentus).—Stoss., 1889, 69.
- diffusocalciferum ranæ esculentæ Gastaldi of Dies., 1855, 64 footnote (see D. diffusocalciferum Gastaldi).
- dilatatum Fischer de Waldheim, 1840, 158 (in Gallus communis; Vilnæ).—
 Anacker, 1887c, 513.—Baillet, 1866b, 105.—Braun, 1893a, 874.—Caruccio, 1886b, 293.—Cobbold, 1879b, 440.—Crep., 1846, 134.—Dies., 1850a, 383-384.—Hahn & Lefèvre, 1884a, 516 (of Miram.).—Hass., 1896a, 2, 3 (syn. of Echinost. echinatum (Zed.)).—Landois, 1882, 23.—Linst., 1873, 106.—Stoss., 1892, 167 (syn. of Echinost. ech. (Zed.)); 1898, 52.—Verrill, 1870, 179.

dilutatum Schneidemuehl, 1896, 303, misprint for dilatatum.

- dimidiatum Crep., 1829, 55–56 (in Acipenser sturio).—Dies., 1850a, 372 (syn. of D. grandiporum Rud.).—Duj., 1845a, 470.—Mehlis, 1831, col. 176–177.—Mol., 1859, 826 (D. grandiporum Rud.).—Odhn., 1905, 360 (syn. of Derogenes varicus (Mueller) Odhn.).
- dimorphum [see below] Dies., 1850a, 353–354 [contains D. marginatum Rud., 1819a, 680, and Duj., 1845a, 446] (in Salmo carapus, Geophagus poppoterra, Chætobranchus flavescens, Crenichichla johanna, C. lepidota; Ardea, Ciconia americana, C. mycteria, Ardea cocoi; Brazil); 1855c, 55–56, pl. 3, figs. 1–6; 1855, 363; 1858e, 338.—Braun, 1892a, 699, 735; 1893a, 865, 871, 872, 873, 874; 1893, 354; 1899g, 485, 486, 490; 1900c, 25, 26, 28–29; 1900h, 4, 18, 25, 32, 33, 34, 36, 42; 1902b, 116.—Cobbold, 1860a, 10.—Gamb., 1896a, 72 (in Ardea ciconia; Brazil).—Looss, 1899b, 651 (from Ardea cocoi; Brazil).—MacCallum, 1899, 697, 706, 707.—Mont., 1893, 155 (in Gallo comune).—Sons., 1889, 283.—Stoss., 1886, 25; 1892, 181.—Also reported for Carapus brachyurus, Hydrolichus scomberoides.
- dimorphum Dies., 1850a, 354, pars (in Ardea cocoi, A. sp., Mycteria americana; in Brazil, collected by Natterer).—Braun, 1899g, 490, is Clinost. marginatum (Rud., 1819a); type is no. 1493, Berlin Museum; and nos. 343, 750, and 831, Vienna Museum; 1900h, 25.
- dimorphum Dies., 1850a, 353, pars, in Tantalus loculator; in Brazil, collected by Natterer.—Braun, 1899g, 490, is Clinost. sorbens Braun n. sp.; type is no. 878, Vienna Museum; 1900h, 34.
- dimorphum Dies., 1850a, 354, pars, in Ciconia americana, Mycteria americana; in Brazil, collected by Natterer.—Braun, 1899g, 490, is Clinost. detruncatum Braun n. sp.; types are nos. 750 and 882, Vienna Museum; 1900h, 32.
- dimorphum Dies., 1850a, 353, 354, pars, in Ardea cocoi; in Brazil, collected by Natterer.—Braun, 1899g, 490, is Clinost. dimorphum (Dies.) Braun; types are nos. 832 and 879, Vienna Museum. [Braun erroneously says n. sp.]; 1900h, 36.
- dimorphum of Wagener, 1852, 555-557, pl. 16, fig. 1 (from chicken) [nec Dies.].—Braun, 1900c, 25; 1900h, 18 (= D. commutatum Dies.).—Dies., 1858e, 339 (renamed D. commutatum; in Phasianus gallus); 1855, 64, footnote 4.—Hass., 1896a, 3 (syn. of Mesogonimus commutatus (Dies.).—Rail., 1893a, 371 (to Mesogonimus).
- distichum (Mueller, 1776) Zed., 1803a, 215 (in fish).—Cobbold, 1860a, 29.—Dies., 1850a, 378–379.—Rud., 1809a, 402, 440–441; 1819a, 123.
- divergens Rud., 1809a, 371–372, 395 (in Blennius viviparus), Fasc. blennii Mueller, renamed; 1819a, 97, 372–373, 676.—Braun, 1892a, 636, 637, 640, 643, 646.—Carus, 1884, 129.—Cobbold, 1860a, 23.—Dies., 1850a, 337, 344.—Duj., 1845a, 461.—Fraip., 1880b, 106, 107, 265 (in Zoarces viviparus); 1881b, 2–4, 24; 1883a, 37.—Harz, 1881c, 5.—Kampmann, 1894b, 447, 449, 459, 460, 461.—Knoch, 1894a, 10.—Kroyer, 1838–40a, 138, 368, 583 (in Zo. vivip. Linn., Cottus scorpius Linn.).—(Lamouroux, 1824, 863.)—Looss, 1885b, 20.—Mont., 1888a, 42, 43, 44, 45, 46; 1893, 62.—Nord., 1840, 619.—Olfers, 1816, 45.—Stoss., 1886, 47.—Also reported for Blennius cornutus, B. gattorugine, B. tentaculatus.
- dubium (Leidy, 1856) Dies., 1858e, 336.—Stoss., 1892, 181 (includes D. leidyi Cobbold).

dujardinii Cobbold, 1860a, 29 (D. soleæ Duj., renamed) (in Solea vulgaris).

duplicatum Baer, 1826a, 124; 1827b, 558-570, pl. 29, figs. 1-15 (in Anadonta ventricosa, A. anatina).—Braun, 1892a, 769, 770; 1893a, 838; 1893b, 184.—Cobbold, 1879b, 453, 454.—Crep., 1837, 310, 325, 326.—Dies., 1836, 240; 1850a, 293 (renamed Rhopalocerca tardigrada; Regiomontii); 1855a, 380, 384; 1858d, 241, 271.—Desmonceaux, 1868, 22.—Erc., 1881e, 37, 45; 1882a, 273, 281.—Fil., 1854a, 9, 26; 1855b, 7, 20, 22, 24; 1856a, 263, 265; 1857c, 25, 27, 29, 31, 32.—Florance, 1866a, 12.—Keber, 1851a, 90.—Kuech., 1856c, 269, 277.—Lampert, 1894.—Laveran & R. Bl., 1895, 105.—Leuck., 1863, 509, 765.—Levin., 1881a, 81.—Looss, 1894a, 17, 23, 63, pl. 4, fig. 78 (syn. of Cerc. Dist. folii, C. folii, young of D. folium).—Moul., 1856a, 14, 21, 51, 56, 78, 82, 120, 129, 139, 143 (to Cerc.), 177.—Mueh., 1898, 11.—Nord., 1832a, 57; 1840, 548, 617, 630, 631.—Pag., 1857, 6, 7, 8, 9, 28-30, 46, 52, pl. 6, fig. 14 (in Anodonta cygnea); 1862, 299, 300, 301, 302, 304.—Reuss, 1902, May 19, 375-377; 1902, July, 405-406; 1903, 458-477, pl. 23, figs. 1-21; 1903, Nov. 26, 206; 1905, July 3, 63-64.—Rossbach, 1906, 420, 431.—Sieb., 1835, 56; 1854, 109, note 2.—Steenstrup, 1842, 51-55, 56.—Tennent, 1906, 651, 653.—Wagener, 1857, 22, 36, 45, 51, pl. 24.—Ziegler, 1883, 540.

echeneidis remoræ Rud., (1821-28) 163 (in Echeneis remora).—Braun, 1893a

echeneidis remoræ Rud., (1821–28)
 163 (in Echeneis remora).—Braun, 1893a,
 875.—Dies., 1855, 64, footnote 16; 1858e, 356 (in Echeneis remora).—Stoss.,

1886, 47.

echinata (Sieb., 1837) Ben., 1861, 89–92, pl. 11, figs. 1–8, in Limneus auricularis; L. ovatus; L. stagnalis; Physa; Cyclas cornea.

echinatoides Dies., 1858d, 263, syn. of Cerc. (Nephrocephala) megacotyla, for echinatoides anodontæ.

echinatoides anodontæ Pag., 1857, 32 (t. h. Anodonta cygnea).—Dies., 1858d, 263 (syn. of Cerc. (Nephrocephala) megacotyla).

echinatum Zed., 1803a, 220-221 (D. anatis Gmelin, 1790) Zed., 1800, renamed; made to include Cucullanus conoideus 1782, Festucaria anatis 1788, F. made to include Cucullanus conoideus 1782, Festucaria anatis 1788, F. boschadis 1790.—Anacker, 1887c, 513.—Baillet, 1866b, 96, 105.—Baird, 1853a, 55.—Bellingham, 1844a, 426.—Ben., 1858a, 1861a, 89–92, pl. 11, figs. 1–8 (syn. Cerc. brunnea, C. echinata); 1868, 296.—Braun, 1883b, 28; 1883a, 70, fig. 14; 1892a, 585, 595, 633, 722, 797, 807; 1893a, 821, 861, 863, 865; 1893b, 182.—Bremser, 1824c, 134; pl. 10, fig. 4–5; 1828, 134 (of birds).—Crep., 1837, 310, 311, 312, 316, 326; 1839a, 289; 1846, 134, 141, 142, 144, 145.—Cobbold, 1860a, 33.—Dies., 1836, 240; 1850a, 344, 345, 346, 383, 398 (includes Cucullanus conoideus, D. anatis; D. excavatum (Nycticoracis) Rud.; D. gruis; D. radiatum Duj.?; Fasc. anatis; F. gruis; Festucaria boschadis; Plan. teres poro simplici); 1858d, 261, 263 (larval stage—Cerc. ech.); 1858e, 344–345 (adult in Anas boschas dom. & fera: A. apser: A. evgnus fer 1858e, 344-345 (adult in Anas boschas dom. & fera; A. anser; A. cygnus fer.; A. clangula; A. clypeata; A. ferina; A. fuligula; Ardea comata; A. gardeni; A. grus; A. marila; A. moschata; A. nycticorax; A. nyroca; A. olor; A. pavonia; A. penelope; A. strepera; A. tadorna; Ciconia nigra; Carbo cormoranus; C. pygmæus; Podiceps cristatus; P. minor; Stat. juven. in Fringilla domestica; C. pygmæus; Podiceps cristatus; P. minor; Stat. juven in Fringilla domestica; larva in Paludina vivipara; Lymnæus stagnalis; Planorbis corneus) (syn. Cerc. (Nephrocephala) ech.; 1859c, 433 (includes as hosts: Cyclas cornea; Belgia; Lymnæus auricularis; L. ovatus; L. stagnalis; Physa).—Desmonceaux, 1868, 22.—Duj., 1845a, 426-427.—Erc., 1881e, 29, 31, 48, 52, 88; 1882a, 265, 267, 284, 288, 324 (syn. Cerc. ech.).—Fil., 1854a, 19; 1855b, 25.—Florance, 1866a, 12.—Fraip., 1880c, 418.—Gamb., 1896a, 63, 72.—Generali, 1881a, 614-615 (in dogs) [Erc.]; 1882a, 70-71.—Giebel, 1857, 265.—Hahn & Lefèvre, 1884a, 516 (of Ben.) (in Canara).—Harz, 1881c, 4.—Hass., 1896a, 2, 3 (to Echinost.).—Kastenbaum, 1899, 244, fig. 33, 5, 34, 7.—Kowal., 1894, 3; 1895, 351-352, pl. 8, figs. 1-3, 12a, 13; 1896d, 7 (257) (in Corvus cornix; Anas crecca; Dublany).—Leidy, 1888, 126; 1904a, 211.—La Valette, 1855, 32.—Leuck., 1863, 82, 498, 526, figs. 143, 144, 178, 180; 1879, 15, 102, 107; 1886d, 11, 76, 80.—Linst., 1873, 1, 98, 101, 105-106 (young stage=Cerc. ech. Sieb.) (in Anas 80.—Linst., 1873, 1, 98, 101, 105–106 (young stage=Cerc. ech. Sieb.) (in Anas boschas); 1884, 139; 1894b, 333 (in Bythinia ventricosa, Physa fontinalis, Doschas), 1884, 139, 1894b, 353 (In Bythinia Ventricosa, 14); 1893b, 813, 814; 1894a, 2, 119, 120, 124, 159, 168, 169, 170, 172, 215, 230, pl. 6, figs. 114, 115–117, pl. 9, figs. 191–192; 1895c, 74, 75; 1896b, 10; 1899b, 556; 1902m, 804, 805, 810, 811, 822.—Lutz, 1892a, 785.—Mégnin, 1884, 53.—Mol., 1858, 290; 1861, 216.—Mont., 1888a, 14, 24, 40.—Moul., 1856a, 190 (larva is Cerc. ech., teste La Valette).—Nathusius, 1835, 53.—Nord., 1832a, 47, 90, 98 (in Barsch); 1840, 621 (syn. of Fasc. trigonocephala).—Pag., 1857, 244–251 (experimental

MA—Continued.
infections); 1858, 5–6; 1859, 58, 60.—Par., 1887, 489.—Rail., 1893a, 366.—
Rud., 1809a, 418–420 to (Echinost.), 431, 432; 1814a, 102; 1819a, 115, 121,
416–417, 685–686.—Schneidemuehl, 1896, 303.—Schlotthauber, 1860, 129.—
Sieb., 1835, 57; 1854, 23.—Sons., 1890, 134.—Stiles & Hass., 1898a, 87, 96
(type of Echinost.).—Stoss., 1890, 51; 1891, 111; 1892, 167 (to Echinost.);
1898, 52.—Tasch., 1879, 71.—Verrill, 1870, 179.—Ward, 1895, 341 (in Canis familiaris; duodenum).—Wedl, 1857, 245-246, pl. 1, figs. 5-6.—Zuern, 1882, 207.—Also reported for Anas clypeata; A. querquedula; Anser cinereus dom.; Ardea grus; A. nycticorax; Aythya americana; Colymbus minor; Cygnus musicus; C. olor ferus; Fiber zibethicus; Fuligula ferina; F. marila; F. nyroca; Hausente; Limnæa; Paludina vivipara; Phasianus gallus.

echinatum (Zed.) of Wedl, 1858, 245, pl. 1, fig. 5, from Phalacrocorax carbo is,

according to Looss, 1899b, 681, a misdetermination.

echmiferum (La Valette, 1855) La Valette, 1855, 32.—Braun, 1892a, 772.—Cobbold, 1879b, 454.—Dies., 1858d, 261, 262, 263 (larval stage=Cerc. (Nephrocephala) 1878, 454.—Dies., 1858d, 201, 202, 203 (talvar stage=Cerc. (Nephrocephrata) echinatoides); 1858e, 345–347 (in Stat. juven. in Fringilla domestica); 1859c, 433 (syn. D. militare Ben.. 1858a, 84, in Scolopax gallinago; Mergus merganser; Podiceps minor; Anas boschas).—Erc., 1881e, 29, 31; 1882a, 265, 267.—Moul., 1856a, 201.—Rail., 1893a, 366 (syn. of D. echinatum).—Stoss., 1892, 171 (to Echinost.).—Ward, 1895, 341 (syn. of D. echinatum) (in Canis familiaris).—Zuern, 1882, 207.

echiniferum paludinæ Pag., 1857, 30-32, 52, pl. 2, figs. 5-6 (in Paludina vivipara);

on 55, descr. of pl. 2, echiniferum.

[echinocephala Crep., 1837a, 326, as name of a group of distomes; quotes Nitzsch.] echinocephalum Rud., 1819a, 115, 418 (in Falco milvus) to (Echinost.) (contains D. milvi Rud., 1809a, 429).—Braun, 1893a, 874.—Dies., 1850a, 385 (syns. D. milvi; Fasc. milvi; Plan. latiuscula Gœze).—Duj., 1845a, 425.—Stoss., 1892, 173 to Echinost.

echiuri Greef, 1879a, 130 (in Echiurus pallasi).—Braun, 1893a, 869; 1893d, 468.

elegans (Rud., 1802) Rud., 1809a, 375-376; 1819a, 100.—Braun, 1901, 561, 564, 567; 1902b, 38-39, 40 (Vienna no. 362 from Fringilla cœlebs = pars Urog. mac.; no. 363 from F. montana = pars Plagiorchis = D. cirratum Rud.), 41, 42, 45, 47, 18. 48, 49 (to Plagiorchis) (syn. of Urogonimus macrostomus).—Cobbold 1860a, 12.—Crep., 1829, 59–63; 1837, 313, 316.—Dies., 1836, 240; 1850a, 349–350 (includes D. erraticum Rud.).—Duj., 1845a, 414–415 to (Brachylaimus).—Fil., 1855b, 8.—Mehlis, 1831, 179.—Mueh., 1896, 589.—Olfers, 1816, 44.—Schlotthauber, 1860, 129.—Sieb., 1835, 56.—Stoss., 1892, 153; 1892, 11 to (Brachylaimus); 1904, 2.

elephantis Dies., 1858, 354 (based on Jackson, 1847, 317; in Elephas indicus) (to Cladocœlium); 1858e, 354 (in Elephas indicus).—Cobbold, 1860a, 8-9 [see Fasc. jacksoni Cobbold]; 1879b, 394.—Fitz, 1876b, 513.—Stoss., 1892, 9 (to Cladocœlium).—Jackson, 1847, 317 [see Fasc. jacksoni Cobbold].

ellipticum Mol., 1858, 130 (in Acipenser nasus; Patavii); 1861, 216-217, pl. 3, figs. 1, 4.—Carus, 1884, 126.—Dies., 1850a, 351; 1858e, 351 (in Ac. nasus).—

Stoss., 1886, 41, to (Echinost.).

elongatum Mehlis, 1831, 177 (in Larus argentatus, L. marinus, L. ridibundus).— Crep., 1837, 316; 1846, 139.—Dies., 1850a, 397.—Stoss., 1892, 181.—Also reported for Chroocephalus ridibundus.

embryo Olfers, 1816, 110, fig. 16 (in Perca vulgaris, P. cernua).—Cobbold, 1860a, 26; 1879b, 458.—Dies., 1850a, 365 (includes D. longicolle Crep.).—Kroyer,

1838–40a, 579 (in Acerina cernua).—Štoss., 1886, 48.

[endemica Sons., 1884, 17-21(not as specific name but refers to Dist. ringeri ende-

mic in Japan and Formosa).]

endemicum Bælz, 1883, 235 (sub. D. hepatis).—Braun, 1893b, 187; 1893, 349, 352; 1893f, 386, 425 (syn. of D. japonicum R. Bl.).—Corlette, 1897a, 146.—Dunglison, 1893, 338.—Hoyle, 1890, 538.—Huber, 1896a, 577.—Ijima, 1886a, 1890, 1893, 538.—Hoyle, 1890, 538.—Hublet, 1890a, 547.—Jamie, 1890a, 47–59, pl. 7, figs. 1–9; 1887a, 596; 1888a, 213–215; 1889b, 137.—Jamieson, 1897b, 147 (syn. of D. sinense).—Katsurada, 1891c, 151–155.—Laspeyres, 1904a, 6.--Lockwood, 1901, 2 ed., 821.—Looss, 1907, Feb. 1, 141–147.—Mont., 1888a, 25, 36, 52, 58, 60.—Rail., 1890, 143.—Simon, 1897, 223.—Sons., 1889, 278, 279, 280, 282 (syn. D. japonicum Bl.) (in Felis dom.).—Stiles, 1904; 35 (syn. of Opisthorchis sinensis).—Stoss. 1892, 23–24 (syns. D. sinense 1904i, 35 (syn. of Opisthorchis sinensis).—Stoss., 1892, 23-24 (syns. D. sinense, D. spatulatum Leuck.).—Reported also for Homo sapiens, China, Giappone,

Bengal, Tonkin, Corea, Formosa; Felis domestica).—Tyson, 1903, 3 ed., 1180 (syn. of D. sinense).—Ward, 1903, 870 (syn. of Opisthorchis sinensis).

endemicum hepatis St. Remy, 1883, 528–529, fig. 3.—La Clínica de Málaga, 1883, 309.—Cf. hepatis endemicum.

endoboluun Mont., 1891, 110, for endolobum.

endolobium Gurlt, 1845a, 288, 290 (for endolobum).

endolobum Duj., 1845a, 397 (in grenouilles vertes et rousses et la salamandre; Rennes).—Brand., 1892, 506.—Braun, 1889a, 355; 1892a, 598, 642, 700, 701, 715, 718, 719, 797, 806; 1893a, 818, 824, 860, 861, 865, 916; 1893b, 182.—Buetschli, 1872, 234-236, pl. 8.—Cobbold, 1860a, 17; 1879b, 454.—Dies. 1850a, 388-389; 1858d, 252 (larva is Cerc. (Acanthocephala) armata Sieb.); 1858e, 348, 349 (adult in Salamandra maculosa, Rhedoni; Pelophylax esculentus, Heidelbergæ; Rana temporaria; larva in Planorbis corneus, Paludina impura, Lymnæus stagnalis); 1859c, 434 (Cerc. armata).—Engel, 1866a, 8.—Erc., 1855a; 1881e, 21, 82, 84; 1882a, 257, 318, 320.—Fil., 1857c, 32.—Florance, 1866a, 11.—Gamb., 1896a, 72.—Harz, 1881c, 4.—Juel, 1889, 15.—Kampmann, 1894b, 446-454, 457, 461, 462, pl. 20, figs. 19-23.—Kath., 1894a, 146.—Kerbert, 1881a, 572.—Kowal., 1894, 3.—Lander, 1904a, 16.—Linst., 1873, 1 (young is Cerc. armata); 1887, 97-101, 102, pl. 2, figs. 1-2 (syn. D. rastellus); 1894b, 332-333 (larva in Phryganea flavicornis, Limnophilus rhombicus, L. griseus, Anabolia nervosa, Ephemera vulgata, Chloëon dipterum).—Looss, 1885b, 24; 1892, 66; 1893b, 815; 1894a, 2, 19, 26, 82, 83, 84-90, 94, 95, 122, 123, 124, 137, 140, 149, 150, 158, 159, 179, 181, 187, 188, 189, 190, 191, 202, 206, 207, 208, 214, 215, 216, 230, 231, 240, 245, 249, 259, 264, 270, pl. 2, figs. 27-29, pl. 7, figs. 153-156, pl. 8, figs. 157-162, pl. 9, figs. 176-186 (syns. D. rastellus; D. retusum Duj. of Ben.; Fasc. ranæ); (reported for Bufo vulgaris, B. variabilis, B. calamita, Rana esculenta, R. temporaria, Triton cristatus); 1896b, 95; 1899b, 589, type of Opisthioglyphe; 1905, 21 (Fasc. ranæ).—Mont., 1891, 110 (endoboluun); 1893, 28, 83.—Mueh., 1898, 28.—Olss., 1893, 11.—Pag., 1857, 19, 41, 52, pl. 5, fig. 1 (in green frogs).—Schwarze, 1885, 67.—Sons., 1893, 188 (in Rana esc. L.).—Staff., 1900, 404, 406; 1902, 724.—Ziegler, 1883, 557.—Also reported for: Gammarus pulex, Limnophilus rhombicus, Triton alpestris.

enterarchos de Fil.—Par., 1896, 13, syn. of D. crassicolle Rud.

epatico Galli-Valerio, 1893a, 178, 181, pl. 2, figs. 3, 4.

ercolani Mont., 1893, 40, for ercolanii.

ercolanii Mont., 1893, 40, 42, 43, 83, 86, 95, 98, 102, 188, pl. 6, fig. 67.—Braun, 1901b, 13 (includes D. signatum Erc. [not Duj.]), 15, 16.—Looss, 1899b, 567 (in all probability identical with Telorchis linstowi).—Luehe, 1899, 528, 529, 530.—Rizzo, 1902, 28.—Stoss., 1895, 223–224, to (Dicroccelium); 1904, 1 (includes D. signatum Erc. [not Duj.], in Tropidonotus viperinus, 5–6 (to Telorchis).—Volz, 1899, 235, 237.—Reported also for Tropidonotus natrix.

erinacei (Bl., 1847) Braun, 1889, 343.

erinaceum Poir., 1886, 37–38, pl. 4, fig. 6 (in Delphinus delphis).—Braun, 1892a, 643; 1893a, 870.—Looss, 1899b, 590 (probably an Astia).—Mont., 1893, 83, 86, 105, 106, 107.—Stoss., 1892, 22, to (Dicrocœlium).

eriocis (Mueller, 1788) Zed., 1803a, 212.—Dies., 1850a, 363 (syn. of D. hyalinum Rud.).—Rud., 1809a, 389.

erraticum Rud., 1819a, 120 (in Fringilla linaria, Motacilla alba, Parus cœruleus, P. major, P. palustris, P. pendulinus).—Braun, 1902b, 38 (of Linst., 1894, 335, syn. of Plagiorchis elegans Rud.), 42.—Dies., 1850a, 350 (syn. of D. elegans Rud.), 361 (syn. of D. macrostomum Rud.).—Duj., 1845a, 443.—Erc., 1881e, 73; 1882a, 309.—Kowal., 1896d, 3 (253) (in Picus major; Dublany).—Linst., 1894b, 335–336 (in Parus major).—Luehe, 1899, 531 (probably a Plagiorchis).—Stoss., 1892, 183 (syn. of D. macrost. Rud.).

esocis lucii Rud., 1809a, 438-439 (in Esox lucius), based on Rud., 1803, 29-30.

eurystomum Linst., 1877, 183-184 (in Anas clangula).—Stoss., 1892, 159, to

(Dicrocœlium) (in Bucephala clangula).

exasperatum Rud., 1819a, 117, 421–422 (in Sorex eremita) to (Echinost.).—Braun,
1901b, 22; 1901, 341–344, pl. 19, figs. 6–7 (includes D. exasperatum Duj.,
1845a; Dies, 1850a; D. (Brachylaimus) exasperatum Stoss., 1892; D. (Brachylaimus) rubens Duj., 1845a; D. rubens Dies., 1850a).—Cobbold, 1879b,

296.—Crep., 1837, 311.—Dies., 1850a, 391.—Duj., 1845a, 411.—Stoss., 1892, 15 (in Crossopus fodiens, Sorex tetragonurus) (includes D. rubens).

excavatum (Rud., 1803) Rud., 1809a, 399-400, 404 (in Ardea ciconia): 1819a, 109 (in A. ciconia, A. nycticorax), 402-403.—Dies., 1850a, 383 ([nycticoracis] syn. of D. echinatum Zed.).—Lamouroux. 1822a, 194.—Nord., 1840, 628 (to Holost.).—Rail., 1893a, 366 (syn. of D. ech.).—Ward, 1895, 341 (in Canis familiaris) (syn. of D. ech.).—See next entry.

excavatum (Rudolphi, 1803) Blainv., 1824a, 518-519 (a holostome).—Dies., 1850a, 310 (to Hemist.).—Duj., 1845a, 375.—Mehlis, 1831, 175.—Nathusius, 1837, 53.—

See foregoing entry.

excisum Rud., 1819a, 112, 411–412, 685 (in Scomber colias, at Naples; Scomber scomber, at Arimini).—Bellingham, 1844, 425.—Ben., 1870, 37.—Braun., 1892a, 654, 682.—Bremser, 1824, 134, pl. 9, figs. 19–20.—Carus, 1884, 128.—Cobbold, 1860a, 25.—Cohn, 1902k, 54, to (Lecithocladium).—Crep., 1837, 326; 1839, 289.—Dies., 1850a, 375–376; 1858e, 342, 343; 1859c, 432.—Duj., 1845a, 436.—Harz, 1881c, 5.—Kroyer, 1838–40a, 236, in Scomber scombrus.—Looss, 1899b, 640 (to Hemiurus).—Luehe, 1901, 397, 398.—Mol., 1858, 290; 1859, 841; 1861, 211–212.—Mont., 1891, 497, 500, 502.—Olss., 1868, 51; 1876, 21.—Sons., 1891, 259 (in Scomber scombrus).—Stoss., 1883, 116; 1886, 15.—Wagener, 1860, 165, 166, 172–176, pl. 8, figs. 8–14.

exiguum Mueh., 1898, 17, 25, 89-90, fig. 11 (in Circus rufus).—Looss, 1899b, 564

(type of Holometra Looss).

expansum (? Creplin, 1842) Mont., 1892, 714 [speaks of M. expansum on same page], probably lapsus for expansum Crep., 1842.

exspinosum Hausmann, 1896a, 391; 1897b, 4, 6, 20, 22, pl. 1, figs. 4–5 (in Barbus

fluviatilis).—Looss, 1899b, 598 (to Asymphylodora).

fabenii Mol., 1859, 289 (in Cantharus vulgaris: Batavii); 1858, 289; 1861, 202–203.—Carus, 1884, 130.—Dies., 1859c, 428–429.—Sons., 1890, 142–143 (in Cantharus lineatus Mont.).—Stoss., 1886, 27.

factum Mont., 1893, 32, misprint for fractum.

falcolacx Leidy, MS., in Stiles & Hass., 1894d, 250 (name found in bottle in Leidy collection; is probably D. trapezium Leidy) .

falconis chrysaëti Rud., 1809a, 429 (=D. felleum Falconis chrysaëti Viborg).

falconis milvi Rud., 1809a, 429-430, for F. milvi Gmelin.

falconis rufi Rud., 1819a, 119 (in Falco rufus; Cat. Ent. Vien.).—Dies., 1850a, 346 (syn. of D. lineola).—Duj., 1845a, 442.—Stoss., 1892, 183 (syn. of D. lin.).

fallar Rud., 1819a, 117, 120 (in Uranoscopus scaber; Naples).—Carus, 1884, 126.—Cobbold., 1860a, 37. to Echinost.—Dies., 1850a, 392.—Duj., 1845a, 432.—Looss, 1899b, 576, 581, 582; 1901, 655, 658; 1902m, 789, 804, 805, 811, 822, type of Anisogaster 1901 [not 1863] and Anisocladium, 1902.—Stoss., 1886, 34; 1890, 41.—Wagener, 1860, 170.

farionis (Mueller, 1788) Bl., 1891, 481–483, fig. 38 (includes F. truttæ Frælich,
 D. laureatum Zed.).—Braun, 1893b, 179, 184.—Hausmann, 1897b, 4, 6, 17,

20, 22, 35 (in Thymallus vulgaris).

fasciatum Rud., 1819a, 97, 373—374, 595 (in Labrus tinca, L. merops, Perca marina; Naples).—Barbagallo & Drago, 1903, 410 to (Dicroccelium) (in Crenilabrus cœruleus: Catania; Serranus scriba).—Braun, 1891d, 421; 1892a, 720, 765; 1893a, 910; 1893b, 184 (in Labrus merula).—Carus, 1884, 130.—Cobbold, 1860a, 23.—Dies., 1850a, 344 (includes D. labri Rud., 1819a, 122).—Duj., 1845a, 456.—Lint., 1900, 291.—Looss, 1899b, 571 (this form as described by Stoss., 1885, belongs to Creadiinæ, possibly to Creadium); 1901, 399, 401.—Mont., 1888a, 66; 1893, 192, pl. 1, fig. 15.—Odhn., 1901, 484, 485, 486, 487, 488, 490, 491, 493, 496, 497; 1902, 160.—Olss., 1868, 32.—Sons., 1890, 141 (in Serranus scriba L.); 1891, 257.—Stoss., 1885, 160; 1886, 32; 1892, 64; 1898, 46; 1902, 578.—Will.-Suhm, 1870, 8; 1871, 182, pl. 11, fig. 7.—Also reported for Anguilla vulgaris, Crenilabrus melops, C. tinca, Ctenolabrus rupestris, Labrus mixtus.

felineum Rivolta, 1884, 20–29, 1 pl. (in cat and dog: Italy).—Askanazy, 1900b, 491 (cats and dogs; Italy), 494 (felinum); 1900c, 72–80; 1901, 72, 73, 74, 75, 76, 77, 78, 79, 80 (cases in man); 1906, 127–131 (in Idus idus, Leuciscus rutilus).—Braun, 1893, 349, 351, 352, 353; 1893f, 386, 387, 388, 390, 391, 392, 422, 423, 424, 425 (includes D. conus Gurlt, Sons. D. lanceolatum Crep. p.p., Sieb., van

Tright; in dogs and cats), 426, fig. 3 (extensive account of early synonymy); 1894, 129; 1894i, 602, 605, 606; 1895b, 148-151, figs. 66-68; 1900g, 250; 1901e, 314, 315, 338; 1902b, 5.—Kholodk., 1897, v. 27 (1), 185–186 (in man).—Huber, 1896a, 576 (syn. of D. sibiricum).—de Jong, 1887a, 57ff (in dog; Holland); 1886–87; 1887b, 223, 224; 1896a, 1, 2, 3, 4, 9, 10, 11, fig. 4.—Kamensky, 1900a, 1–23 (syn. of Opisthorchis tenuicollis Rud., Mueh.); 1900b, 1–23; 1901a. 323–324.—Kholodk., 1898, 26, 29, pl. 9, figs. 10–12; 1899a, 152.—Kowal., 1898g, 74; 1898h, 144 (41).—Laspeyres, 1904a, 513 (felinum).—Linst., 1903, 279.— 174, 163811, 144 (41).—Laspeytes, 1904a, 313 (leftitum).—Liffist., 1903, 279.—Looss, 1896b, 58, 59; 1899b, 530, 674, 675, 676; 1905, 89 (to Opistorchis).—Moniez, 1896, 86, 123, 136–141, fig. 26.—Mueh., 1898, 15, 16, 21 (syn. of D. tenuicolle), 24.—Rail., 1893a, 361.—Ratz, 1898, 67–69, fig. 1 (felinum); 1900, 250; 1900, 141–142; 1900, 532.—Rindfleisch, 1904, March., 346 (in man); 1904, 8.—Roth, 1904, 93-94 (in man, infection from Leuciscus rutilus).-Schneidemuehl, 1896, 302.—Shaw, 1901, 1027.—Simon, 1897, 223.—Sons., 1889, 276, 280; 1889, 281 (syn. of D. conus Crep.); 1897, 254, 255, 256–257 (in Canis fam.; Pisa); 1897, Apr., 221.—Stiles, 1894n, 358–360.—Stiles & Hass., 1894e, 426–427, figs. 5–8; 1898a, 85 (type of Opisthorchis).—Tyson, 1903, 3 ed., 1181 (in cats and man).—Ward, 1895, 152; 1895, 238 (man, cat, dog; includes D. lanceolatum of cats and dogs), 243-244 (includes as syns. D. conus Gurlt nec Crep.; D. lanceolatum Sieb., 1836; D. truncatum Rud. pars.); cat, dog, and glutton; Europe; man; Siberia), 328 (in Homo; biliary ducts), 341 (in Canis familiaris; biliary ducts); 1895, (3), Mar., 152–158, 1 fig.; 1895, 8 pp.; 1895, 304–309; 1901, 180 (of Ward syn. of Opisthorchis pseudofelineus).—Weichselbaum, 1898, 315.—Also reported for Gulo borealis.

felineum of Ward, 1895, 152-158, 1 fig.; 1895 (nec Riv.), 239, fig. 1, 240, fig. 2, 242, fig. 3 (=Opisthorchis pseudofelineus); 1896, 5 Nov., 709; 1901, 180.

felinum (sibiricum) Askanazy, 1900b, 491–502, for felineum, cases in man; 1900c, 711, 712.

felinum (see D. felineum; misprint in Ratz, 1898, 67).

felleum falconis chrysaeti Viborg, 1795, 243 (renamed D. Falconis chrysaeti by Rud.).—Dies., 1850a, 376 (syn. of D. crassiusculum).

fellis Olss., 1868, 44–46, pl. 5, fig. 94 (in Anarhichas lupus).—Braun, 1893a, 875.— Jacoby, 1899c; 1900, 1, 12–16, 20, figs. 8–12.—Lint., 1900, 291.—Luehe, 1900, 504.—Odhn., 1905, 307, 309, 310.—Stoss., 1886, 24 (in Anarrhichas lupus).

ferocis Mont., 1888a, 14, apparently misprint for ferox.

ferox (Rud., 1795) Zed., 1803a, 221.—Baird, 1853a, 56, includes Echinorhynchus t (Rud., 1799) Zed., 1803a, 221.—Battl, 1855a, 30, lifetides Echilolitylicitis ardeæ nigræ Braun.—Ben., 1868, 295, 296–299, pl. 1, figs. 1–5.—Braun, 1892a, 568, 583, 584, 721; 1893a, 877, 881.—Bremser, 1824, pl. 10, figs. 6–11.—Crep., 1837, 311, 312, 316, 326.—Dies., 1850a, 387–388 (syns.: Plan., D. ardeæ Zed.; D. ardeæ stellaris Rud.; Echinorhynchus ardeæ nigræ Braun; Fasc. ardeæ Gmelin).—Duj., 1845a, 429–430.—Gamb., 1896a, 63.—Giebel, 1857, 265.—Kowal., 1896d, 253 (3) (in Ciconia alba; Dublany).—Leck., 1866d, 11; 1879, 144.—Lipet. 1872, 166 (in Ciconia alba; Dublany).—Leck., 1866d, 11; 1879, 1841a. A.—Linst., 1873, 106 (in Ciconia alba).—Looss, 1892a, 14.—Mayer, 1841a, 4.—Mol., 1858, 130; 1861, 219.—Mont., 1888, 144.—Nathusius, 1837, 53.—Nord., 1832a, 90.—Olfers, 1816, 47.—Olss., 1876, 22.—Rud., 1803, 90; 1809a, 426–427, to (Echinost.), 433; 1814a, 102; 1819a, 116–117, 120, 419–420.—Villot, 1875, 473.—Also reported for Ardea ciconia, A. nigra, A. stellaris, Ciconia

ferruginosum Linst., 1877, 184–185, pl. 14, figs. 25–27 (in Barbus fluviatilis).—
Braun, 1892a, 587, 729, 766.—Hausmann, 1897b, 30.—Looss, 1894a, 24, 25, 26, 31, 32, 123 (syn. of D. perlatum).—Stoss., 1886, 48.

filicolle (Rud., 1819a) Ben., 1858a, 1861a, 104–107, 111, 187, 189, 195, 199, 200, 201, pl. 10, figs. 1–10 (includes D. okenii); 1870c, 137.—Braun, 1892a, 698–727.—Carus, 1884, 128.—Cobbold, 1860a, 31 (in Brama raii), to Köllikeria as type.—Leuck., 1879, 137; 1886d, 106.—Mont., 1888a, 9, 18, 52, 62, 93, 104; 1893, 150.—Sons., 1890, 143 (of Wagener).—Stoss., 1886, 17.

filiferum Sars, 1885, 222, pl. 38, figs. 19–21 (in Nematoscelis megalops S., Thysanoëssa gregaria S.; South Atlantic Ocean).—Braun, 1893a, 836, 837, 853.—Linst., 1888, 17.—Mont., 1893, 40, 42, 43, pl. 5, fig. 61.

filiforme Rud., 1819a, 112, 411, 772 (in Cepola tænia at Arimini; Cepola rubescens).—Braun, 1883a, 41; 1892a, 672.—Carus, 1884, 131.—Cobbold, 1860a, 28.—Dies., 1850a, 375.—Duj., 1845a, 460–461.—Stoss., 1886, 48.—Will.-Suhm, 1870, 8; 1871, 182.

filum Duj., 1845a, 418 (in moineaux at Rennes). to (Brachylaimus).—Cobbold, 1860a, 15 (in Fringilla dom.).—Dies., 1850a, 376.—Hausmann, 1897b, 28.—Stoss., 1892, 152.

fimbriatum Busch, 1851, 99, pl. 15, fig. 12 (in Sagitta).

flagellatum Moniez, 1891, 27 (in Gymnotus electricus); 1896, 144.

flavescens Pag., 1857, 34–35, pl. 3, fig. 14 (in Bulimus radiatus).—Braun, 1892a, 642.—Dies., 1858d, 278 (to Cercariæum).

flavescens Ben., 1870, 47, pl. 5, fig. 4, (in Gobius jozo; G. minutus).—Stoss., 1886, 21. flavocinctum Linst., 1879, 183–185 (in Anguis fragilis).—Stoss., 1895, 230; 1897, 9 (= D. crassicolle); 1898, 32.

flexum Lint., 1892, 98-99, pl. 6, figs. 36-44 (in Oedemia americana).

flexuosum Rud., 1808a, 346; 1809a, 50, 389–391 (in Talpa europæa): 1819a, 105.— Bellingham, 1844a, 424.—Braun, 1891d, 421; 1901a, 33.—Cobbold, 1860a, 7; 1879b, 296.—Dies., 1850a, 360; 1858e, 341 (in Talpa europæa).—Duj., 1845a, 398–399.—Hausmann, 1897b, 28.—Looss, 1899b, 562, type of Omphalometra.—Mueh., 1896, 244–248, figs. 1–8; 1896, 588; 1898, 16, 28.—Olfers, 1816, 44.—Stoss., 1892, 26 (in Talpa europæa; Greifswald, Copenhagen, Rennes).

facundum Lint., 1900, 269, 282, 289–290, pl. 36, figs. 27–35, pl. 37, figs. 36–37 (in Lopholatilus chamæleonticeps); 1901, 415, 419, 432, 472.

foliaceum Mol., 1859, 288 (in Gobius paganellus; Batavii); 1861, 199.—Carus, 1884, 131.—Dies., 1859c, 428.—Stoss., 1886, 48.

foliatum Lint., 1898, 532–534, pl. 49, figs. 3–5, pl. 50, figs. 1–3, pl. 51, figs. 1–4 (in Mola mola); 1900, 282; 1901, 415, 422, 466; Looss, 1902m, 644, to Orophocotyle.

foliiforme Crep., 1846, 149 (in Squalus griseus); 1837, 317.

folium Olfers, 1816, 45, fig. 15 (t. h. Esox lucius).—Bell, 1887a, 117.—Braun, 1892, 44; 1892a, 642, 663, 674, 697, 710, 716, 720, 723, 737, 746, 783, 785; 1892g, 461–463; 1893a, 876; 1893d, 467; 1899, 2; 1899g, 490, 492, type of Phyllodist.; 1899b, 721; 1901b, 9, 10, 12.—Cobbold, 1860a, 23.—Cohn, 1896, 240.—Darr, 1902, 688.—Dies., 1850a, 343 (in Esox lucius; Berlin); 1858e, 335.—Duj., 1845a, 464.—Fil., 1855b, 25.—Hausmann, 1897b, 15.—Kroyer, 1846–53a, 253.—Looss, 1893b, 813, 814, 815; 1894a, 2, 18–24. D. duplicatum [Rhopalocerca tardigrada], viewed as young form, 28, 29, 31, 58, 62, 63, 64, 110, 125, 136, 137, 138, 142, 159, 161, 167, 174, 179, 181, 183, 185, 186, 194, 197, 200, 204, 206, 211, 215, 217, 227, 230, 269, 270, 275, pl. 1, figs. 19–22, pl. 4, figs. 75–80 (reported for Acerina cernua; Cottus gobio; Salmo umbla; Thymallus vulgaris; Trutta variabilis); 1899b, 605, type of Spathidium; 1902m, 476, 477.—Mont., 1888a, 7, 55; 1893, 83, 102, 107.—Much., 1898, 11, 15, 28.—Odhn., 1902, 64, 65.—Rossbach, 1906, 377, 388.—Rud., 1819a, 96–97, 371–372, 588.—Spengel, 1892.—Ssinitzin, 1901, v. 24, 25 Nov., 689–694 (in Dreissenia polymorpha); 1902, 359–360; 1904, 768, figs. d, e.—Staff., 1902, 481, to (Spathidium), in Ameiurus nebulosus.—Stoss., 1886, 48.—Wagener, 1855, 1857, 26, 27.—Will.-Suhm, 1873.—Zschokke, 1884.

formosum Sons., 1890, 134–136 (in Grus cinerea Bechst.); 1891, 2 Mch., 291.— Braun, 1892a, 698: 1901, 944; 1902b, 21, 22.—Mont., 1893, 82, 148; 1896, 166.—

Ofenheim, 1900, 160.—Stoss., 1892, 143 (to Polyorchis).

fractum Rud., 1819a, 107, 397 (in Sparus salpa; Naples).—Braun, 1893a, 910.—Carus, 1884, 124.—Cobbold, 1860a, 28.—Dies., 1850a, 377-378.—Duj., 1845a, 458.—Jacoby, 1900, 4.—Looss, 1894a, 170, 204, 219.—Luehe, 1900, 487.—Mont., 1892, Oct. 7, 174, 175; 1893, 13, 23, 24, 32 (factum), 33, 40, 41, 42, 43, 61, 79, 82, 83, 85, 87, 88, 91, 94, 96, 98, 102, 105, 106, 107, 114, 120, 141, 146, 167-172, 175, 186, pl. 1, fig. 9, pl. 5, fig. 62, pl. 6, figs. 81-86, pl. 7, figs. 110-111, pl. 8, figs. 112, 114.—Nord., 1832a, 35.—Odhn., 1905, 328.—Sons., 1890, 139-140 (in Box salpa Cuv.); 1891, 262.—Stoss., 1886, 49, to (Podocotyle); 1887, 184; 1892, 65.

fragile Lint., 1900, 269, 282, 295, pl. 39, figs. 68-70 (1n Mola mola); 1901, 415, 420,

466.—Type of Stenocollum 1904.

fraternum Looss, 1894d, 42–48, pl. 2, figs. 13–15 (in Pelecanus onocrotalus); 1895a, 1896b, 60–63, 101, 154, 156, pl. 4, figs. 36, 37 (in pelican, Alexandria); 1899b, 535, 550, 556, to Cœnogonimus; 1902m, 804, 805, 886, 887.—Braun, 1901e, 334, 336, 338.—Jacoby, 1900, 23.—Jægers., 1898, 9, 12.—Luehe, 1899, 539, to Cotylogonimus (Cotylogonimus).—Moniez, 1896, 143–144.—Mueh., 1898, 81, 82 (considers D. concavum a Mesogonimus and more or less related to fraternum and heterophyes).—Sons., 1896, 314.—Stoss., 1898, 42.

 $\it fralichii$ Kowal., 1894, 3, host not given (to Echinost.); 1895, 353–355, pl. 8, figs. 4–8, 12 b; 1895, 372–390, pl. 8, figs. 1–6.

fuligulæ ferinæ Dies., 1858e, 355 (in Anas ferina; Ireland) based en Bellingham, 1844, v. 13, 430.—Stoss., 1892, 167 (to Echinost. echinatum).

fulvum Rud., 1819a, 98, 374–375 (in Gadus molva at Naples; G. mediterraneus).—Bellingham, 1844a, 423.—Carus, 1884, 131.—Cobbold, 1858b, 157, pl. 31, figs. 6–8 (in Moletta quinquecirrata) (includes D. simplex Rud.; Fasc. bramæ Mueller); 1860a, 23.—Dies., 1850a, 345 (in Lota molva; Naples); 1858e, 335 (in Raja batis).—Duj., 1845a, 466.—Kroyer, 1843–45a, 166 (in Lota molva).—Linst., 1903, 354.—Stoss., 1886, 49.—Also reported for Gadus mustela Linn.; Molva vulgaris; Motella mustela.

furcatum Bremser in Rudolphi, 1819a, 107, 396–397, 683–684 (in Mullus surmuletus, M. rubescens at Arimini; Gadus molva at Naples); 1824, 133.—Braun, 1892a, 576, 584; 1893a, 910.—Carus. 1884, 124.—Cobbold, 1860a, 29.—Crep., 1837, 310.—Dies., 1850a, 378 (in Mullus barbatus; M. rubescens; Armini; Lota molva, Naples; Coryphæna hippurus, Brazil).—Duj., 1845a, 402.—Eysenhardt, 1829, 145.—Hausmann, 1897b, 35.—Kroyer, 1838–40a, 581; 1843–45a, 166 (in Mullus surmuletus; Lota molva).—Looss, 1902m, 771, 772 (undoubtedly represents a new genus).—Luehe, 1900, 487, 488, 490, 491, 492.—Mont., 1888a, 12; 1893, 192, pl. 1, fig. 18.—Odhn., 1905, 325, 326.—Stiles & Hass., 1898a, 92.—Stoss., 1883, 116, 1886, 16.—Also reported for Box salpa; Molva vulgaris.

furcigerum Olss., 1868, 26, pl. 4, fig. 72 (in Pleuronectes limanda, P. limandoides).—
Braun, 1892a, 643, 728, 752; 1893a, 910.—Levin., 1881a, 12, 61-64, 67, 76, 77, pl. 2, figs. 5-6 (in Cottus scorpius, at Egedesminde; Pl. limanda; P. limandoides).—Linst., 1890f, 179.—Mont., 1893, 61, 95, 105, 193.—Odhn., 1905, 302, 305 (type of Steringophorus).—Staff., 1904 (type of Leioderma n. g., nec Suhm, 1873).—Stoss., 1886, 28 to (Dicroccelium).

[fusca Sluiter, 1900, 7 (a tunicate).]

fuscatum Rud., 1819a, 101, 384–385 (in Tetrao coturnix; Ancona).—Braun, 1901, 561, 564; 1901, 941; 1902b, 114, 115 (to Harmost.).—Cobbold, 1860a, 11; 1879b, 440.—Dies., 1850a, 353.—Duj., 1845a, 445.—Stoss., 1892, 182; 1892, 40.

fuscescens Rud., 1819a, 113, 413 (in Sparus dentex; Arimini).—Carus, 1884, 130.—Dies., 1850a, 377 (in Dentex vulgaris; Arimini).—Luehe, 1900, 490.—Mol., 1859, 833, 838–840, 845, pl. 1. fig. 1 (in Dentex vulgaris; Padua).—Mont., 1893, 40, 41, 42, 43, 82, 83, 95, 102, 177–179, pl. 5, fig. 63.—Stoss., 1885, 159; 1886, 27; 1886, 59 to (Dicrocœlium); 1898, 45 (in Dentex vulgaris; Triest).—Also reported for Caranx trachurus.

fuscum (Bosc, 1802) Poir., 1885, 6, 12–13, pl. 23, fig. 7.—Braun, 1893a, 872, 873.—
 Buttel-Reepen, 1902, 167, pl. 6, fig. 17.—Reported for Doras sp.

fusiforme Zed., 1800a, 163 (in Upupa ep.), 171–173, Fasc. upupæ Schrank, 1790, renamed; 1803a, 210.—Braun, 1901, 561.—Dies., 1850a, 351 (syn. of D. involutum).—Rud., 1809a, 377.

gadi Dies., 1855, 64, based on Bellingham, 1844a, 428, t. h. Gadus æglefinus; Ireland.

gadi æglefini Dies., 1858e, 341, based on Bellingham, 1844, 428 (renamed D. anonymum Dies.)

galactosomum Leidy, 1888i, 166–167 (in Labrax lineatus; U. S. A.): 1889, 611; 1904a, 216–217.—Braun, 1893a, 872; 1900h, 43.—MacCallum, 1899, 704, 707.

gammari Rentsch, 1860, 18, 35–50, pl. 12, figs. 7–12, 15–16 (in Flohkrebs, Gammarus ornatus).

gammari Linst., 1877, 186, n. sp.; 1878a, 315 (in Gammarus pulex).

gammari ornati Rentsch, 1860, in explanation of fig. 2, pl. 12 (D. gammari Rentsch, 1860, renamed).

gastrocolum Leidy, 1891a, 414–415 (in Trichiurus lepturus; U. S. A.); 1904a, 235–236.—Stiles & Hass., 1894, 414.

gelatinosum Rud., 1819a, 102, 386–387 (in Testudo mydas; Arimini).—Braun, 1899b, 715, 716–717; 1901b, 16, 18, 19, 29–34, figs. 6, 12, 19; 1902b, 26.—Carus, 1884, 129.—Cobbold, 1860a, 19.—Dies., 1850a, 356.—Duj., 1845a, 451.—Looss, 1899b, 570, 579, 580; 1901l, 563, 564, 565; 1902m, 445 (to Rhytidodes), 446, 451, 452, 454, 455, 456, 457.—Mont., 1888a, 38; 1892, 715; 1896, 165; 1903a, 86, 102.—Par., 1846.—Sons., 1890, 4 May; 1893, 5 Feb.; 1893, 183, 184

(in Chelonia caretta L.).—Stoss., 1895, 37; 1895, 226–227; 1898, 43; 1904, 3.—Also reported for Chelone mydas, Emys lutraria, Halichelys atra, Podocnemis expansa, Thalassochelys caretta L., T. corticata.

gelatinosum Rud. of Poir., 1886, 33-34, pl. 3, fig. 6 (nec Rud.) (in Cistudo lutraria).—Braun, 1899b, 716 (syn. of D. poirieri Stoss.); 1901, 13, 19.—Looss, 1899b, 567 (possibly identical with Telorchis linstowi).—Luehe, 1899, 528.— Stoss., 1904, 3 (syn. of D. poirieri Sons., 1893, Telorchis poirieri Stoss.).

geminum Looss. 1896b, 50-52, 54, 58, 59, pl. 4, figs. 25-27 (in Milvus parasiticus;

Cairo); 1899b, 675 (to Opistherchis).

geniculatum Dies., 1850a, 373-374(in Physophora tetrasticha: Naples) (D. physophoræ Fil., renamed).—Cobbold, 1860a, 30.—Mont., 1888a, 77; 1888, 195, 196; 1893, 124.—Moul., 1856a, 217.

genu Rud., 1819a, 107-108, 397-398 (in Labrus luscus; Naples).—Braun, 1893a, 873.—Carus, 1884, 131.—Cobbold, 1860a, 27.—Dies., 1850a, 368–369.—Duj., 1845a, 462.—Looss, 1901d, 399.—Nord., 1832a, 36.—Odhn., 1901, 484, 487, 490, 496.—Stoss., 1886, 49.

giardi Stoss., 1898, 50-51, for giardii.

giardii Stoss., 1889, 25(3), pl. 13, fig. 56 (in Naucrates ductor; Triest); 1898, 50-51 (giardi).—Braun, 1892a, 583, 673; 1893a, 910.—Buttel-Reepen, 1902, 202.—Mont., 1893, 29, 43.

gibbosum (Rud., 1802) Rud., 1809a, 399, pl. 6, fig. 8 (in Esox belones); 1819a, 107, 395–396.—Bellingham, 1844a, 424.—Carus, 1884, 124.—Cobbold, 1860a, 28.—Crep., 1837, 326.—Dies., 1850a, 378; 1858e, 343 in Gadus æglefinus, Ireland; 1859c, 433.—Duj., 1845a, 402.—Harz, 1881c, 5, 11.—Kroyer, 1846–1853a, 273 (in Belone rostrata Fabricius).—Luehe, 1900, 487, 492; 1901, 480.—Mol., 1858, 290 (in Belone acus; Patavii); 1861, 213.—Odhn., 1905, 356, 357 (to Lecithester). Olion, 1816, 46.—Stilos, 1901, 193.—Stilos, 6, Hars, 1898a, 92. thaster).—Olfers, 1816, 46.—Stiles, 1901, 193.—Stiles & Hass., 1898a, 92.—Stoss., 1886, 49.—Also reported for Colymbus cristatus.

giganteum Dies., 1858e, 331–332 (gigantica, 1855, renamed).—Bassi, 1875b, 508.— Braun, 1892a, 650 (gigantea), 568, 674; 1893d, 466; 1903a, 875, 910.—Leuck., 1863, 530.—Stiles, 1898a, 49.—Stoss., 1892, 9 (to Cladocœlium).

1863, 530.—Stiles, 1898a, 49.—Stoss., 1892, 9 (to Chadoccenium).

gigas Nardo, 1827. 68-69 (in Proctostegus proctostegus, teste Braun); 1833, 523; 1874-1875 or (1876), 265-266.—R. Bl., 1888a, 543; 1891, 479-481, fig. 37.—Braun, 1892a, 632; 1893a, 873; 1893b, 184 (in Proct. prototyphus).—Buttel-Reepen, 1902, 167, 168, 169, 171, 172, pl. 6, figs. 10, 10a.—Carus, 1884, 126.—Cobbold, 1858, 167; 1860a, 28; 1879b, 460.—Crep., 1837, 310.—Darr, 1902, 664, 665, 666, 669, 671, 697.—Dies., 1850a, 373.—Hoyle, 1890, 540.—Luche, 1901, 483, "D. gigas" (from Luvarus imperialis), to Accacelium raynerianum, 485.—Mont., 1889, 322; 1891, 500, 520; 1893, reprint, 7-9.—Par., 1902, 6 (in L. im.; Portoferrajo).—Poir., 1885, 6.—Setti, 1894, (19 pp.), pl. 28; 1894, 17 pp., pl. 28, 6 figs.; 1895, 316; 1895, 306; 1895, Feb. 28, 270; 1895, 367.—Stoss., 1886, 50.—Also reported for Ansonia cuvieri.

glabrum Crep., 1846, 148 (in Salamandra maculosa).—Dies., 1850a, 398.—Stoss.,

1889, 70.

glandulosum Looss, 1896b, 64-68, 69, 71, 72, 76, pl. 5, figs. 41-44 (in Taphosus nudiventris; Ghizeh); 1898, 453, 454, 455, 456, 457, 458, figs. 1, v; 1899b, 547, 716, 717 to Lecithodendrium.—Stiles, 1901, 200.

glauci Bergh, 1884, 18, pl. 10, figs. 5-17 (in Glaucus atlanticus, etc., teste Braun). -Linst., 1888, 17.

globicaudatum Crep., 1849a, 64.

globiferum Lamarck, 1816, 182 for D. globiporum.—Nord., 1840, 618 (syn. of Fasc. globifera).

globiparum Ehrenb., 1837b, 199, for globiporum.

globiporum (Rud., 1802), Rud., 1809a, 364-367 (includes Fasc. bramæ Mueller; F. lanceolata (bramæ) Schrank, 1790; F. tincæ Modeer, 1790; F. longicollis F. lanceolata (braine) Schrank, 1790; F. thice Modeer, 1790; T. longicollis (carpionis) Fredich, 1791; D. cyprinaceum Zed., 1800; D. carinatum Zed.), 410, 414, 441; 1819a, 96.—Ben., 1858a, 1861a, 100, 191, 197, 203.—Braun, 1883a, 52; 1892a, 635, 695, 699, 711, 746, 747, 756, 779, 780, 783, 785, 788, 790; 1893a, 865, 879, 883.—Burm., 1835b, 187–194, pl. 2, figs. 1–6.—Cobbold, 1860a, 21.—Condorelli-Francaviglia, 1898, 2, 4, 5–7.—Crep., 1837, 310, 313, 322, 323, 324, 327, 328, 329; 1846, 152, 153, 154.—Dies., 1850a, 341 (includes Fascbramæ Mueller, F. lanceolata Schrank, F. longicollis Frælich, F. globipora

Rud., D. cyprinaceum Zed., D. carinatum Zed., D. bramæ Zed.); 1858e, Rud., D. cyprinaceum Zed., D. carinatum Zed., D. bramæ Zed.); 1858e, 334 (larva in Lymnæus stagnalis).—Duj., 1845a, 417.—Ehrenb., 1837b, 199 (globiparum); 1837, 151–180; 1838b, 15.—Gamb., 1896a, 72.—Hausmann, 1897b, 4, 6, 8, 10, 17, 18, 20, 21, 22, 39–40 (in Scardinius erythropthalmus, Chondrostoma nasus, Barbus fluviatilis, Squalius cephalus, S. leuciscus, Cobitis barbatula, Abramis brama, A. blicca, A. vimba, Alburnus lucidus, Acerina cernua, Cyprinus carpio, Esox lucius, Leuciscus rutilus, L. meidingeri, L. scardapha, Phoxinus lævis, Perca fluviatilis, Tinca vulgaris, Thymallus vulgaris).—Kampmann, 1894b, 446.—Kroyer, 1838–40a, 20; 1846–53a, 367, 387, 434; 1852–1853a, 1222, 1223, 1224, 1225 (in Abramis brama, A. blicca, A. vimba, Aspins alburnus, Barbus fluviatilis, Leuciscus erythrophthalmus, L. rutilus Aspius alburnus, Barbus fluviatilis, Leuciscus erythrophthalmus, L. rutilus, Perca fluviatilis, Phoxinus aphya, Tinca vulgaris).—Lamarck, 1816, 182 (globiferum).—Lamouroux, 1824, 563 [Distome à pores globuleux].—Leuck., 1863a, 500, 501, 503, 504.—Linst., 1882, 19, fig. 24; 1885, 251.—Lint., 1901, 415 (in Pseudopleuronectes americanus), 420, 486, fig. 347; 1905, 328, 334, 356, 378, 393, figs. 159, 173, 198, 199 (in Fundulus majalis, Leiostomus xanthurus, Orthopristis chrysopterus).—Looss, 1885b, 22, 23, 24, pl. 23, fig. 1894a, 41 (includes Fasc. bramæ Mueller; F. longicollis Fasc. b 1894a, 41 (includes Fasc. bramæ Mueller; F. longicollis Fredich; D. cyprinaceum Zed., 1800; Fasc. globipora Rud., 1802), 2, 24, 41–48, 49 (of Olss., syn. of D. isoporum Looss), 50, 52, 53, 54, 58, 98, 123, 124, 136, 137, 141, 150, 151, 157, 159, 161, 162, 179, 191, 192, 195, 197, 214, 215, 216, 231, 239, 242, 243, 249, 256, pl. 1, figs. 11–14; pl. 5, figs. 95–101 (in Abramis brama, A. blicca, A. vimba, Alburnus lucidus, Barbus fluviatilis, Chondrostoma nasus, Cyprinus carpio, Esox lucius, Leuciscus erythrophthalmus, L. jeses, L. rutilus, Perca fluviatilis, Phoxinus lævis, Squalius cephalus, Thymallus vulgaris); 1896b, 208; 1899b, 646, 647, 648; 1902m, 757 (and Fasc. bramæ), 758, 760, 762, 764, 769.—Mol., 1858, 129; 1861, 199.—Mont., 1888a, 69, 72; 1892, Oct. 7, 187.—Moul., 1856a, 48 (embryo).—Much., 1898, 24, 28.—Nord.; 1832a, 88; 1840, 616, 618 (syn. of Fasc. globifera).—Olfers, 1816, 45.—Olss., 1876, 16; 1893, 11.—Par., 1896, 2, to (Dicroccelium).—Poir., 1885, 1,101.—Schauinsland, 1882, 496, 497.—Sieb., 1836, 233, 236, 237, 238, 239; 1836, v. 1, 218, pl. 6; 1838, 301.—Sramek, 1901, 95 (in Abramis vimba, C. V., A. brama), 96 (in Squalius dobula Heck.), 107–108 in Squalius lepusculus Heck.) (syn. (in Squalius dobula Heck.), 107–108 in Squalius lepusculus Heck.) (syn. Fasc. bramæ Mueller).—Stiles, 1901, 168, 194.—Stiles & Hass., 1898a, 95 (type of Sphærostoma).—Stoss., 1883, 116; 1886, 29; 1902, 582.—Wagener, 1857, 26, 44, pl. 23, fig. 1 (in Lymnæus stagnalis); 1860, 170; —, v. 9, 88, 89. pl. 1, fig. 5.—Also reported for Abramis alburnus, Anguilla chrysypa, A. vulgaris, Blicca björkna, Cobitis fossilis, Cyprinus balerus, C. barbus, C. blicca, C. brama, C. alburnus, C. dobula, C. erythrophthalmus, C. phoxinus, C. nasus, C. rutilus, C. tinca, C. vimba, Idus melanotus, Leuciscus phoxinus, Limnæa ovata, Physa fontinalis, Planorbis marginatus, Succinea pfeifferi, S. putris.

globiporum tincæ Rud. of Dies., 1850a, 395 (syn. of D. perlatum Nord.).

globocaudatum Crep., 1825, 49–50 (in Corvus cornix); 1849a, 64.—Braun, 1902b, 44 (not Linst., 1883).—Cobbold, 1860a, 13.—Dies., 1850a, 351; 1858e, 337 (in Corvus glandarius).—Duj., 1845a, 413—414.—Fil., 1855b, 8.—Linst., 1878, 99; 1883, 307–308; 1886, 31.—Stoss., 1892, 153 (syn. of D. cirratum Rud.).

globosum Ben., 1858a, 1861a, 193 (refers to Sieb.).—Braun, 1901c, 311 (as a name

of Dies. for D. orbiculare), 312.—Jackson, 1888, 647.

globulus Rud., 1814a, 104 (in Anas fuligula); 1819a, 109, Greifswald, 401–402.—
Bellingham, 1844a, 425.—Braun, 1893a, 879; 1901, 561, 564; 1902b, 19, 152
(includes D. gl. Braun, 1901, 564; Crep., 1846, 142, 143, 145, 146; Dies., 1850a, 365, and 1858e, 341; Duj., 1845a, 450; Rud., 1819a, 109, 401; Stoss., 1892, 40), 155 (to Psilost., Looss) [does not use name globulus in combination with P.].—Cobbold, 1860a, 11.—Crep., 1846, 142, 143, 145, 146.—Dies., 1850a, 365; 1858e, 341 (in Anas Cygnus ferus).—Duj., 1845a, 450.—Olfers, 1816, 46.—Stoss., 1892, 182.—Also reported for Alca torda, Anas fuligula, Amarila, A. sponsa, Dafila acuta, Fuligula cristata, F. marila, Harelda glacialis, Mergus merganser, M. serrator.

gobii Stoss., 1883, 116–117, pl. 2, figs. 6–7 (in Gobius jozo; Triest); 1886, 29; 1890,
41; 1898, 45; 1904, 12 (to Helicometra).—Braun, 1891d, 421; 1892a, 711,
765.—Carus, 1884, 124.—Condorelli-Francaviglia, 1898, 7.—Looss, 1901d,
399.—Mont., 1893, 83, 95, 102.—Odhn., 1901, 494, 495; 1902, 160.—Sons.,

1891, 258.

gobii Rentsch, 1860, 43–50, pl. 11, figs. 3–5, 8–9b–d; pl. 12, figs. 1, 5c–e (in Gobius minutus) [called also D. gobii minuti in description of plates].—Also reported for Gasterosteus spinachia.

gobii minuti Rentsch, 1860, in descr. of plates (for D. gobii).—Odhn., 1901, 495.
goliath Ben., 1858b, 95–97, 1 pl., figs. 1–5 (in liver of a Balæna); 1858f, 282; 1870, 365.—Braun, 1891d, 423; 1892a, 586, 721; 1893a, 875; 1893d, 466; 1902, Nov. 17, 800–803, 1 pl., to Lecithodesmus; 1902, Dec. 30, 841–842; 1905, July, 51, 53.—Cobbold, 1860a, 8.—Dies., 1858e, 336–337 (in Balæna borealis [Balænoptera rostrata Fabricius]).—Hahn & Lefevre, 1884a, 516.—Jægers., 1891, 132.—Lænnberg, 1891, 71–73.—Odhn., 1905, 344, 347 (to Lecithodesmus).—Stoss., 1892, 34–35 (in Balæna mysticetus, Balænoptera rostrata).—Villot, 1878, 2.

gracile (Leidy, 1856) Dies., 1858e, 336 (in Esox spec., Pomotis vulgaris).— Braun, 1893a, 871; 1900h, 43, 44, 45.—MacCallum, 1899, 704.—Mont., 1893, 155.—Stoss., 1886, 50.—Also reported for Apogon anullorum.

gracile of Wright, 1879, 9 (in Perca flavescens).—Braun, 1899g, 491 (probably syn.

of Clinost. marginatum Rud.); 1900h, 43, 45.

gracile of Lint., 1898, 523-524, pl. 46, figs. 6-8 (in Lepomis auratus. Eupomotis pallidus, Chænobryttus gulosus).—Braun, 1899g, 491 (probably syn. of Clinost. marginatum Rud.); 1900h, 44, 45 (thinks Poirier's form, 1886, 39, pl. 3, fig. 8, from Axinurus dugesi, may belong here).

gracile of MacCallum, 1899, 704, pl. 39, fig. 7.—Braun, 1900h, 44, 45, probably syn.

of Clinost. marginatum.

gracilescens Rud., 1819a, 111, 409 (in Lophius piscatorius; Triest).—Braun, 1893a, 886.—Bremser, 1824c, pl. 9, figs. 17–18.—Cobbold, 1858b, 161, pl. 32, figs. 33–37 (in Lophius piscatorius); 1879b, 462.—Crep., 1837, 326.—Dies., 1850a, 374–375; 1858e, 361 (in Loph. pisc.) (to Rhipidocotyle).—Duj., 1845a, 462.—Kroyer, 1838–40a, 471.—Linst., 1878a.—Maddox, 1867, 97.—Stoss., 1898, 61.—Tennent, 1906, 638 (to Gasterost.)—Wagener, 1860, 189.

grande Rud., 1819a, 676–677 (in Platalea ajaja; Brazil).—Braun, 1901g, 561, 564–565; 1902b, 24 (of Braun, 1901g, 564; Dies., 1850a, 346 (of Duj.); 1845a, 446 (of Rud.); 1819a, 676; and of Stoss., 1892, 2; to Mesaulus), 26, figs. 16–19.—Cobbold, 1860a, 12.—Dies., 1850a, 346–347.—Duj., 1845a, 446.—Odhn., 1902,

32, 34, 35.—Stoss., 1892, 182.

grandiporum Rud., 1819a, 110–111, 407–408 (in Muræna helena; Naples).—Braun, 1893a, 873.—Cobbold, 1860a, 23.—Dies., 1850a, 342, 371–372 (includes D. varium Eysenhardt, D. dimidiatum Crep.).—Duj., 1845a, 421.—Johnston, 1901, 337.—Kroyer, 1852–53a, 778 (in Acipenser sturio).—Lint., 1898, Jan. 20, 520–521, pl. 44, fig. 9; 1901, 415, 418, 436, 486 (in Anguilla chrysypa, Pseudopleuronectes americanus); 1905, 328, 334, 351 (in Leptocephalus conger).—Looss, 1899b, 640 (to Hemiurus).—Luehe, 1901, 476–477.—Mol., 1859, 826–828, pl. 2, fig. 5 (syns. D. varium Eysenh., D. dimidiatum Crep.) (in Accipenser sturio, Greifswald; Anguilla vulgaris, Padua; Muræna helena, Naples).—Odhn., 1905, 360.—Olss., 1876, 20.—Stoss., 1886, 14; 1902, 582.

granulum Rud., 1809a, 394–395 (in Cottus scorpius) (includes Fasc. scorpii Mueller, 1776); 1819a, 106.—Cobbold, 1860a, 27.—Dies., 1850a, 366.—Duj., 1845a, 457.—Kroyer, 1838–40a, 138, 368 (in Zoarces viviparus, Cottus scorpius).—Nord., 1840, 620 (syn. of Fasc. scorpii).—Olfers, 1816, 46.—Stoss.,

1886, 50.

grassum Biermer, 1863a, 395 (for crassum).

gruis (Gmelin, 1790) Zed., 1803a, 221–222.—Baird, 1853a, 55 (syn. of D. echinatum Zed.).—Dies., 1850a, 383 (syn. of D. ech. Zed.).—Rud., 1809a, 432; 1819a, 115.

gulosum Lint., 1901, 415, 418, 454, figs. 315–317 (in Rhombus triacanthus).— Nicoll, 1907, 69.

gyrini Linst., 1884, 141–142, pl. 10, figs. 27–28 (in tadpole of Rana temporaria).— Braun, 1893a, 870.—Stoss., 1889, 68.

hæmatobe Ben., 1858a, 219 (for D. hæmatobium).

hæmatobium Bilharz, 1852a, 72–76 (in Homo; Egypt); 1853a, 454–456, figs. a–k;
1853 [in Sieb., 1853], 59–62, pl. 5, figs. 11–15 (in Homo; Egypt); 1856a,
49–52, 65–68.—Agnew, 1881, 709.—Aitken, 1866, 841, fig. 14a (to Bilharzia);
1872, 206, fig. 37.—Albarran, 1897b, 1098, 1104, 1105, 1106.—Almeida Couto,

MA—Continued.
1872, 4, 6, 7, 13, 24, 42.—Anders, 1903, 6. ed., 1245–1246.—Batho, 1872b, 502;
1872, 331.—Belleli, 1885b, 54–56, figs. 19–20.—Ben., 1858a, 1861a, 188, 199, 200, 201; 1878a, 276.—R. Bl., 1888a, 636 (includes Bilh. hæm. Cobbold).—Bomford, 1887a, 53–55, pl. 11 to (Bilh.), reported for cattle in Calcutta.—de Bonis, 1876, 158, pl. 2, figs. 14–15; 1882, 174, pl. 2, figs. 14–15.—Bouchut, 1879a, 874–877 (chyluria).—Bowlby, 1891a, 194–195 (eggs in lungs and urinary organs).—Braun, 1883a, 41, 66–68, 181, fig. 17; 1903, 3. ed., 169 (to Schistosomum).—Brooks, 1897a, 492–493 (case in N. Y.); 1897, 617.—Burghart, 1904, XI, 6 (eggs); 1904, XII, 13, 2250.—Cantani, 1886, 9 (hematuria); 1886, 73.—Cabbold, 1859d, 364: 1860, 31: 1864b, 34, 197–204, figs. 44–45; 1865, —; 1865, Cobbold, 1859d, 364; 1860, 31; 1864b, 34, 197–204, figs. 44–45; 1865, —; 1865, 617; 1866, 6; 1871h, 359; 1871g; 1871k; 1873, 145; 1879b, 39.—Crevaux, 1874a, 173, 177.—Da Costa, 1884, 935.—Damaschino, 1882a, 949; 1882, 150; 1883a.—Dav., 1860, 312; 1877a, pp. lxxvii, lxxix, fig. 41; 318–321, fig. 13; 939, 940–944, 946, 973.—Delafield & Prudden, 1897, 130.—Dunglison, 1893, 142, 338, 506, 820, 1174.—Ebstein, 1884a (vescicle calculi).—Eichhorst, 1901a, 142, 338, 500, 820, 1174.—Ebstein, 1884a (Vescicle calculi).—Elfanforst, 1901a, 301.—Fritsch, 1867a, 752.—Goubert, 1878a, 132–133, fig. 61.—Griesinger, 1854a, 561–575; 1866a, 96; 1866, 381 (hämatobium); 1872a, 472.—Guès, 1879a, 168.—Günther, 1858a, 208–209.—Hackley, 1886a, 519, fig. 885.—Handford, 1887b, 240–245, pl. 7, figs. 1–3; 1889a, 424–425.—Harley, 1865; 1869, 394; 1869, 379–387; 1871, 47; 1871, 359.—Haupt, 1878a, 19–20.—Hillmantel, 1893a, 230–233, figs. 1–3; 1893b, 4 pp., 3 figs.—Hoek, 1859, 42 (D. hæm. venæ portarum).—Huber, 1894, 294, 295, 296, 297, 298, 299; 1896, 580 (to Bilharzia).—Ijima, 1889b, 155.—Joel, 1866a, 308.—Jourdan, 1877a.—Kartulis, 1885a, 139–145. figs. 1–4 (eggs in abdominal organic): 1885b, 188–189; 1808b. 453, 455, 466, 488, 526, 617–632, figs. 145, 208–216; 1868, v. 2 (2), 458; 1876, v. 2 (3), 628, 629, 630, 633, 637, 638, 873; 1879, 10, 58, 59, 61, 163, 165, 166, 188, mum).—Mantey, 1880.—Meckel, 1856, 46–47.—Meinecke, 1897a, 209–211, 1 pl. (bladder case).—Meissner, ——, 84–85.—Mosler & Peiper, 1894, 179–185, figs. 71–73.—Moul., 1856a, 25, 48, pl. 4, fig. 25.—Nachtigal, —.—Nitze, 1891, 40, 1891, 692.—Ogden, 1900, 267–268, figs. 49–50.—Paul, 1860, 20.—Purdy, 1900; 208–210, figs. 32–33.—[Renoult, 1808, 366–370.]—Reyer, 1856, 214.—Rieder, 1899, 84, fig. 19.—Rindfleisch, 1884, 203, 210–211.—Roberts, 1865, 493.—Rochard, 1871, 298.—Roger, 1901, 95.—Ruetimeyer, —.—Sachs-Bey, 1880, 1253–1255.—Sandwith, 1901, 690.—Schiess-Bey, —, 303.—Schneidemuehl, 1896, 300–301.—Shaw, 1901, 645.—Sieb., 1852, 59–62, 71; 1852, 537; —, 454.—Simon, 1897, 99, 209, 224, 510.—Simpson, 1872, 320–321.—Sondern, 1897, 554–557, figs. 1–6.—Sons., 1875, 9; 1876, —.—Stiles, 1898a, 58–60 (to Schistosoma), figs. 41–44, 48.—Stiles & Hass., 1898a, 93, 98 (type of Schistos 1891, 504-507, 198. 1-6.—Sons., 1875, 9; 1876, —.—Stiles, 1898a, 58-60 (to Schistosoma), figs. 41-44, 48.—Stiles & Hass., 1898a, 93, 98 (type of Schistosoma).—Stoss., 1892, 5 (to Gynæcophorus).—Swart, 1862, 33, 36-37.—Tedeschi, 1886, 73-75.—Thacher, 1893, 826.—Wagener, 1857, 26.—Wagner, 1883, 121-122.—Ward, 1895, 253 (to Gynæcophorus), 328; 1903, 872 (to Schistosoma).—Weichselbaum, 1898, 315.—Wood & Fitz, 1897, 9, 335.—Virchow, 1891, —.—Zancarol, 1882, 144; 1883, 45; 1884, 306.—Zuckerlandl, 1880, 1853, 270, 233 1253.—Zuern, 1882, 220, 223.

hæmatobium hominis Dies., 1855, 63, footnote (for D. hæmatobium).

hæmatobium venæ portarum Hoek, or Pag., 1859, 42 (for D. hæmatobium).

hæmatoma Braun, 1891d, 426, see hematoma.

halecis (Gmelin, 1790) Zed., 1803a, 222.—Dies., 1850a, 372 (syn. of D. ocreatum Rud.).—Mont., 1891, 496.—Rud., 1809a, 398.

halosauri Bell, 1887a, 116-117 (in Halosaurus macrochir; Cape St. Vincent).-Braun, 1892a, 580, 665; 1893a, 876; 1893d, 467.—Hoyle, 1890, 540.—Mont., 1889, 322; 1893, 83.

helicis Leidy, 1847, 220–221 (in Helix alternata).—Dies., 1855a, 398 (renamed Cercariæum Helicis alternatæ); 1858d, 278 (syn. of Cercariæum vagans).

helicis asperæ Dies., 1850a, 302-303 (based on Duj., 1845a, 472, in liver of Helix aspersa) (to Heterost.).

helicis pomatiæ Dies., 1850a, 303 (to Heterost.), see also Mueller's Arch., v. 5, p. 71, pl. 1, fig. 7.—Vaney & Conte, 1899, 196.

hematoma Semprum, 1890, 596 (in Homo; Cuba).—R. Bl., 1891p, 611-612.—Braun, 1891d, 426 (hæmatoma).

hemicyclum Mol., 1859, 829–830 (in Belone acus; Padua).—Carus, 1884, 127.—Stoss., 1886, 43 to (Echinost.).

tticum (Linn., 1758) Abildg. —.—Abela, 1883, 47.—Aitken, 1866, 804, 839 (to Fasc.).—Alexander, [1833a, 319–323; 1833b, 405].—Anacker, 1888b, 314; 1892c, 94.—Andral, [1829c, 504, 520]; 1829d, 633.—Appenheim, 1899, 104–105 (lungs of sheep).—Armatage, 1895, 429.—Aschoff, 1892, 495.—Askanazy, 1901, 75.—Assénova, 1899, 21, 28, 31, 90–93, 118.—Baer, 1828f, 197–198.—Baillet, 1866b, 99–104.—Baldi, [1900a, 223].—Bellingham, 1844a, 423.—Ben., 1855a, 1861b, 190, 170, 170, 170, 1805. hepaticum (Linn., 1758) Abildg. -1858a, 1861a, 100, 170, 197.—Bettend., 1897a, 4, 7, 8, 9, 12, 15, 17, 18, 26, 27, 31, 1858a, 1861a, 100, 170, 197.—Bettend., 1897a, 4, 7, 8, 9, 12, 15, 17, 18, 20, 27, 31, 33, 34, 36, 39, 42, 43, 44, pl. 2, fig. 8, 9, 10, 13, 14, pl. 4, fig. 26, 27, 28, 29, pl. 5, fig. 33, 34, 35, 36, 37, 38, 39, 40, 41, 42; 1897b, 308, 311, 312, 313, 316, 319, 321, 322, 330, 331, 335, 337, 338, 343, 346, 347, 348, pl. 29, fig. 8–10, 13, 14, pl. 31, fig. 26–29, pl. 32, figs. 33–42.—Bilharz, 1856a, 49.—Billet, 1893a, 506, 508, 509.—Bitting, 1895b, 83–85, pls. 1–2, figs. 1–12.—R. Bl., 1886, 306; 1888a, 543 (syns. Fasc. hepatica Linn., F. humana Gmelin, F. Lapacelleta Bud. 1803). 543, 669, 669, 205, 211, 609, (of Zod. 1800). Pud. 1810. lanceolata Rud., 1803), 543–602, figs. 295–311, 602 (of Zed., 1800; Rud., 1810, syn. of Dist. lanceolatum), 603, 606, 609, 623, 631.—Blochmann, 1892b, 650.— 629, 631, 635, 638, 641, 644, 645, 669, 674, 675, 678, 681, 682, 684, 688, 690, 695, 701, 703, 705, 712, 717, 719, 724, 725, 728, 730, 733, 745, 747, 748, 755, 766, 768, 781, 783, 798, 806, 814, 815; 1892f, 44; 1893a, 861, 865, 875, 878, 880, 882, 910; 1893f, 424; 1894i, 606; 1895b, 11, 12, 126, 132, 133, 134, 138–141, figs. 43, 46–51, 53–56; 1897a, 1387, 1467, 1468, 1522; 1900a, 1668; 1903, 147 (to Fasc.); 1906, 134, 140, 150 (to Fasc.), figs. 69, 75, 83.—Brunet, 1902, 119–128 (Tunisie).—Brusina, (1898a), 227–228.—Burm., 1837a, 529.—Buttel-Reepen, 1902, 178, 184, 185, 2, 7, 14.—Desmonceaux, 1868, 6.—Dewitz, 1892, 117–125, figs. 62, 66, 68, 70, 75, 77, 78.—Dies., 1836, 240; 1850a, 332–333 (syns. Fasc. hepatica Linn., Dist. hepaticum Zed.), 333-334 (of Zed., 167, syn. of Dist. lanceolatum Mehlis); 1858e, 331; 1859c, 427.—Duncker, 1881a, 24.—Dunglison, 1893a, 338, 424, 820, 875, 1174.—Eichwald, 1829a, 248.—Eiss, 1838a, 21-22.—Encycl. méthodique, Par., 1824, v. 2, 316.—Encycl. metropolitana, 1845, v. 18, 141.—Erc., 1881e, [58, 64, 69, 90, 91]; 1882a, 294, 300, 305, 326, 327.—Eschricht, 1840a, 1881e, [58, 64, 69, 90, 91]; 1882a, 294, 300, 305, 326, 327.—Eschricht, 1840a, 73; 1855, 1.—Fagge & Pye Smith, 1902, 4. ed., 475 (to Fasc.).—Fenger, 1854, 173.—Fischer, 1883a, 24, 25, 29, 40.—Fischder, 1903h, 507.—Fitz, 1876a; 1876b, 513, 514.—Florance, 1866a, [22].—Fraip., 1880a, 398; 1880c, 418, 426, 428; 1881b, 36, 39; 1883a, 35, 41.—Francis, 1891c, 127–130, pls. 1–2; 1892b, 608; 1894a, 450.—Fritsch, 1885a, 408; 1888a, 212.—Gaede, (1817a), 8.—Gaffron, 1883a, 509.—Gamb., 1896, 4, 63, 64, 68, 72.—Ghose, 1869a, 210–211 [evidently D. crassum].—Giard & Billet, 1892a, 614.—Gomez, 1879a, 81–89.—Gomy, 1897a, 372.—Gosse, (1857a), 124.—Grall, 1887a, 460, 469.—Griffith & Henfrey, 1883a, 268.—Gronkowski, 1902a, 511, 512, 514, 515–517, 518, 519, 521, 522, 523–529, 530, 531, 532, 533 [4, 5, 7, 8–10, 11, 12, 14, 15, 16–22, 23, 24, 25, 26], figs. B, C, pl. 13, figs. 1, 5, 16.—Günther, 1858a, 205–207.—Gurlt, 1831, 193, 370–372, pl. 8, figs. 29–33.—Hackley, 1886, 518, figs. 880.—Hahn & Lefèvre, 193, 370–372, pl. 8, figs. 29–33.—Hackley, 1886, 518, fig. 880.—Hahn & Lefèvre, 1884a, 516-537, 538, 539, 541 (syn. Fasc. hep.); 1884, 805.—Harley, 1864a, 62.—Harris, 1899a, 900.—Henneguy, 1906, 47-88, pl. 3, figs. 1-24; 1906, 46-52.—Harris, 1893, 900.—Helliegly, 1906, 47–38, pl. 3, figs. 1–24, 1906, 44–88, pl. 3, figs. 1–24.—Hoeven, 1859, 211.—Huber, 1896, 576 (Bassi's magna).—Hutcheon, 1903m, 551–554 (pathology).—Jamieson, 1897a, 73.—Janson, 1893c, 261.—Joy, 1835a, 504, 505, 518–519; 1845a,—.—Juel, 1889, 13, 14, 16, 19, 22, 26, 29, 36, 41.—Kampmann, 1894b, 445.—Kastenbaum, 1899, 243–248, fig. 33, 34.—Kath., 1894a, 143.—Kerbert, 1881a, 530, 536, 538, 539, 540, 547, 551, 565, 566, 567, 569, 570.—Kholodk., 1898a, 24, 25, 26, 27, pl. 10, fig. 5, 22;

1899a, 149–165, figs. 171–174, 177f, 178.—Kitasato, (1884a), May 10.—Knoch, 1894a, 4, 11, 12, 13, 16.—Kowal., 1894a, 220; 1895c, 372–390, pl. 8, fig. 8; 1895g, 20, [60]; 1896d, 7, [257] (in Cervus capreolus; Dublany); 1896i, 9, [353].—Krabbe, 1865, 60.—Kuech., 1855, 183–207, 208, 210, 241, 481, pl. 5, figs. 1–10; 1857, 247–272, pl. 5, figs. 1–10.—Kuech. & Zuern, 1881, 290, pl. 7, fig. 1, 2, 4, pl. 8, fig. 14.—Lamouroux, 1824a, 560, 561, 562, 563.—Lampert, 1898a, 1xxxii–1xxxiii.—de Lanessan, 1882, 221–238, figs. 193–203.—Laspeyres, 1904a, 5, 7.—Leidy, 1874b, 364–365.—Lejtenyi, 1881a, 5, 7, 8, 11, 16, 18.—Leuck., 1863, 13, 14, 48, 100, 456, 457, 458, 463, 464, 465, 469, 471, 472, 475, 476, 479, 480, 483, 484, 487, 488, 490, 517, 520, 522, 526, 527, 529, 530–586, 587, 588, 589 $\begin{array}{c} 479, 480, 483, 484, 487, 488, 490, 517, 520, 522, 526, 527, 529, 530 - 586, 587, 588, 589, \\ 590, 591, 593, 594, 595, 596, 597, 599, 601, 602, 603, 604, 605, 606, 607, 608, 609, 612, \end{array}$ 626, 740, 765, fig. 153, 156, 160, 165, 182, 184–191, 193; 1868, 405; 1876, 868–870, 871, 872; 1879, 16, 17, 59, 74, 164, 186, 208, fig. 39, 68, 69, 70; 1882f, 524–528; 1882g, 320–322; 1886, 12, 46, 56, 73, 81, 128, 146, 147, 161, 164, 589, fig. 35, 39, 1882g, 320–322; 1886, 12, 46, 56, 73, 81, 128, 146, 147, 161, 164, 589, fig. 35, 39, 68, 69, 90; 1892b, 797, 799.—Lindquist, 1882, 180; 1887, 391.—Linst., 1873, 98, 99, 101, 102; 1883, 308; 1890f, 175; 1901, 280; 1906, 175 (syn. Fasc. Linn., 1746, which included Dist. hep., Dendroccelium lacteum, and Schistocephalus solidus).—Looss, 1885b, 6, 7, 10, 12, 15, 17, 19, 25, 26, 31, 34, 38; 1892, 126, 132–136, 141, 179; 1894, 1, 19, 65, 93, 115, 116, 117, 118, 120, 122, 125, 131, 138, 142, 144, 153, 159, 171, 172, 180, 181, 206, 212, 215, 220, pl. 6, figs. 118–120; 1895, 75, 85; 1896, 33–36; 1898, 459; 1899, 556; 1905, 88 (to Fasc.).—Love, 1896a, 48–49.—Ludwig, 1886a, 852–853, figs. 803, 805, 809.—Lutz, 1893a, 128.—Macé, 1882, 91 pp., 3 pls.—Manson, 1903, 3 ed., 639.—Mégnin, 1882s, 221 (in lungs of Bos taurus).—Michalik, 1891, 57.—Minort, 1884a, 418.—Mirra 1889, 317.—Minort, 1882, 144.—Mol., 1859, 825.—Mos. Peiper, 1894, 169–175, figs. 65, 66.—Moul., 1856a, 12, 17, 18, 19, 21, 23, 43, 102, 267.—Muehl., 1898, 9, 21.—Nord., 1832a, 47, 55; 1840, 547, 616, 617.—Ofenheim, 1900, 152.—[Organesyantz, 1873–74, 205].—[Oken, 1835, 550–551].—[Olfers, 1816, 4; 1816, 30, 44].—Oppenheim, 1899, 104–105.—Owen, 1843, 56, fig. 27.—Packard, ——, 522.—Padrone, 1904, 489.—Pag., 1857, 53.—Parker, E. A., 1891, 79, 124, 266, 268.—Parkes, L. C., 1889, 71, 310.—Paul, 1860, 20.—Poir., 1885, 1, 4, 20, 23, 32, 37, 40, 46, 50, 51, 55, 56, 66, 67, 70, 77, 80, 90, 92, 93, 94, 101, 102, 113, 114, 115, 116, 117, 118, 120, 131, 133, 144, pl. xxvii, fig. 2, 3, pl. xxix, fig. 5, pl. xxx, fig. 4; 1887, 203–208, 210.—Prunac, 1884, 14 pp.—Rail., 1886, 286, 293, figs. 180–189; 1890, 836–839; 1890, 143.—Rail. & Marotel, 1898, 31, 32.—Ratz, 1898, 298; 1899, 617; 1900, 532.—Rindfleisch, 1884, 210.—Risso, 1826, no. 34; 1826, 262.—Rivolta (1881), 68; (1887), 391.—Roberts, 1888, 686.—Roewer, 1906, 187.—Rossbach, 1906, 381, 387, 392, 394, 402, 414, 419, 428, 429, 432, pl. 19, figs. 47–48.—Rud., 1803, 62; 1809a, 50, 214, 349, 352–357 (syns. Fasc. hep. Linn, Plan., latiuscula Gœze, Fasc. humana Gmelin, Dist. hep. Zed., Fasc. lanceolata Rud.), 378; 1819a, 92–93, 363–364, 576–577, 583, 588, 616, 617.—Ruser, 1892, 548 (in lungs).—St. Peiper, 1894, 169–175, figs. 65, 66.—Moul., 1856a, 12, 17, 18, 19, 21, 23, 43, 102, 363-364, 576-577, 583, 588, 616, 617.—Ruser, 1892, 548 (in lungs).—St. Remy, 1891, 220.—Schaeffer, 1721, 1719—1721.—Schauinsland, 1882, 497.—Schneidemuchl, 1896, 296—300.—Schubart, 1853, 28—31.—Schuberg, 1894, 185.—Seeger, 1852, 51.—Shaw, 1901, 619, fig. 220.—Sidekrift. Vet. Med. (1882), (see also Amer. Vet. Rev., 1887, 391).—Sieb., 1835, 57, 64, 65; 1836, 233, 237; 1850, 644, 672, 673; 1854, 7.—Signol, 1884, 232.—Simon, 1896, 182, 190, fig. 51; 1897, 209, 222—223, fig. 52.—Sjobeck, 1830, 10.—Steenstrup, 1842, 27.—Stevenson, 1892, 274, 498.—Stiles, 1898a, 29, 49; 1905z, 14.—Stiles & Hass., 1892a, 89.—Stoss., 1892, 7 (to Cladoccelium).—Tasch., 1878, 176.—Taylor, 1885, 58—60, 1 fig.—Tennent, 1906, 666.—Theobald, 1900, 52.—Thomas, 1883, 329.—Vogt, 1878, 9, 38, 42, fig. 28, 35.—Vogt & Yung, 1888, 226—248, figs. 99—108.—Wagener, 1860, 174.—Wagner, 1883, 109, 120—121.—Wallenstedt, 1847, 7.—Walter, 1858, 269—297, pls. 11—13.—Ward, 1895, 246 (to Fasc.), 253 (in part Curtice, syn. of Fasc. magna); 1903, 865 (syn. Fasc. hep.).—Weichselbaum, 1898, 314.—Weinland, 1859, 280.—Wheeler, 1894, 117—118 (New Orleans).—Will.-Suhm, 1870, 1; —, 175—179.—Wolf, 1903, 612, 613, 616.—Wood & Fitz, 1897, 336.—Ziegler, 1883, 542, (543), 545, 546, Remy, 1891, 220.—Schaeffer, 1721, 1719–1721.—Schauinsland, 1882, 497. 117–118 (New Orleans).—Will.-Suhm, 1870, 1; ——, 175–179.—Wolf, 1903, 612, 613, 616.—Wood & Fitz, 1897, 336.—Ziegler, 1883, 542, (543), 545, 546, 553, 556, pl. 33, fig. 13, 14, 15; 1883, 488.—Zuern, 1882, 204, 208, 209, 211, 216, pl. 4, figs. 5-9.

hepaticum, anatomy of: Havet, 1900b, 351–381, pls. 1–4, figs. 1–28 (nervous system).—Macé, 1882, 91 pp., 3 pls., 18 figs.; 1882, 7–9.—Marcinowski, 1903a,

544-550; 1903b, 477; 1903c, 2391; 1903d, 738-739; 1905a, 21-22 (Schlundganglion).—Mehlis, 1825, 42 pp., pl. 1; 1826, 627.—Prenant, 1904, 522-525 (intest. epithelium).—Sommer, 1880, 539-640, pls. 27-32.—Veratti, 1900, 115-125 (nervous system).

- hepaticum, embryo of: Coe, 1896a, 561-570, pl. 42, figs. 1-5; 1897a, 30.

- hepaticum, geographic distribution of: Brunet, 1902a, 119–128 (Tunisie).— Mol., 1859, 825 (Ovis aries, Bos taurus domesticus, Equus caballus; Padua).— Saito, 1906, Aug. 7, 822.

hepaticum, intermediate host of: Agric. J., Cape Town, 1896, Apr. 30, 221.

Cherry, 1896a, 183 (Bulimus, in Victoria).—Studer, 1882, 10-11.

Cherry, 1896a, 183 (Bulimus, in Victoria).—Studer, 1882, 10–11.

- hepaticum, in various animals: Lungs of cattle: Barbagallo, 1903–4, 165 (Catania).—Cope, 1887a, 385–386.—Curtice, 1887a, 390–392.—Hedley, 1881a, 374–375; 1881b, 399–400; 1881c, 27–28.—Littlewood, 1887a, 546.—Mégnin, 1882, 221.—Morot, 1889, 159.—Murray, 1882, 100–103.—Schmidt, 1887, 361–362 (Europe).—Vet. J. & Ann. Comp. Path., Lond., 1899, v. 49, Sept., 179–180.—In Man: R.Bl., 1888a, 589–595; 1891p, 604–606 (Sagarra's case in 1890 in Spain).—Carter, 1862a (India).—Lockwood, 1901a, 2 ed., 821.—Manson, 1901, 539, 540 (also life cycle).—Preuss. Militäräztl. Ztg., 1863, v. 3 (2), 15.—Tyson, 1903, 3 ed., 1180.—Ward, 1895, 328.—In sheep: Brett, 1881a, 139–142; 1881b.—Curtice, 1890c, 16–17, 127–134, pl. 16.—Friedberger, 1878a, 145–166.—Lessona, 1812a.—Lydtin, 1890a, 373 (in bronchi).—Trollip, 1893, 424–425.—Sclavo, 1900–01, (41); 1902, 221–222.—Miscellaneous: Anders, 1903, 6. ed, 1245 (horse, goat, ass, sheep, rabbit, man).—Hutcheon, 1900i, 497 (duiker antelope).—Joel, 1866a, 308 (Bos taurus; liver).—Piana, 1882, 12 pp. (domestic ruminants; liver).—Ward, 1895, 332 (Bos taurus), 335 (Ovis aries), 338 (Equus caballus).—See also Fascioliasis. - hepaticum, life history of: Erc., 1881a, 123–130; 1881b, 320–326; 1881c, 11 pp.;

hepaticum, life history of: Erc., 1881a, 123-130; 1881b, 320-326; 1881c, 11 pp.; epuctum, the fistory of Eur., 1801a, 125–150, 1801b, 320–320, 1881c, 11 pp.; 1881d, 229; 1881, v. 7, 443–447.—Gulliver (1840c), 30–31 (eggs); 1841a, 507–508; 1842a, 95.—Henneguy, 1902a, 1235–1238 (egg, maturation, fecundation); (1902b), 128–131; 1905, July, 49.—R. Leuck., 1881c, 641–646; 1882a; 1882b, 80–119; pl. 8, figs. 1–9; 1882c–e; 1882f, 524–528; 1882g; 1883a; 1884c.—Lutz, 1892a, 783–796, figs. 1–5; 1892b, 436–437; 1892c, 301–306; 1893b, 320–328; 1893c, 389–393; 1893d, 24–25.—Minot, 1884, 418.—Schauinsland, 1882, 4944.—Weinland, 1882, 20–08

land, 1882, 494.—Weinland, 1883, 89-98.

hepaticum, method of feeding of: Bossuat, 1902, 186-187; Rail., 1890, Mar., 88-92; 1890, June 30, 271-272; 1890, Aug. 22, 277.

hepaticum ægyptiaca (also egyptiaca) Looss, 1896b, 10, 33-36, 151, 183, 192, 204, 205, pl. 3, fig. 16, pl. 11, figs. 117-118 (in "buffles, bœufs, moutons;" Alexandria); 1898a, 459, 460.—Linst., 1901, 420 (in Bos zebu, Ovis aries).

hepaticum angusta (Rail., 1895) Looss, 1898a, 459.

hepaticum hominis Cobbold, 1884g, 976.

hepaticum (perniciosum) Taylor, 1884, 52-53, fig. 2.—See hepatis perniciosum.

hepatis endemicum Bælz, 1883, 234-236, fig. 1 (in Homo; Japan).—RBl., 1888a, 618 (syn. of D. japonicum), 619-621, figs. 320-322 (description and infection); 1891, 607 (syn. of D. sinense Cobbold).—Braun, 1903, 3. ed., 161 (syn. of Opisthorchis sinensis).—Hahn & Lefèvre, 1884a, 542-544.—Kamensky, 1900a, 18.—Katsurada, 1900, 479.—Looss, 1905, 90 (syn. of Opisthorchis sinensis); 1907, Feb. 1, 140.—Rail., 1893a, 362 (syn. of D. sinense).—Ward, 1895, 328 (in Homo) (syn. of D. sin.); 1903, 870 (syn. of Opisthorchis sinensis).—See Clonorchis.

hepatis innoccuum Caræs, 1888a, 41, pl. 1, for innocuum.

hepatis innocuum Bælz, 1883, 236, fig. 2.—Billet, 1893a, 509 (syn. of D. sinense Cobbold).—R.Bl., 1888a, 618 (syn. of D. japonicum), 621-622, figs. 323-324; 1891, 607 (syn. of D. sinense).—Braun, 1903, 3. ed., 161 (syn. of Opis. sin.).-Caræs, 1888a, 41, pl. 1 (innoccuum).—Cobbold, 1884g, 976.—Grall, 1887a, 460, 468, 469, 1 fig.—Hahn & Lefèvre, 1884a, 544 (syn. of D. sin.).—Hoyle, 1890, 538.—Kamensky, 1900a, 18.—Katsurada, 1900, 479.—Looss, 1905, 90 (syn. of Opisth. sinen.); 1907, Feb. 1, 140.—Rail., 1893a, 362.—Ward, 1895, 328 (in Homo) (syn. of D. sin.); 1903, 870 (syn. of Opisth. sin.).—See Clonorchis.

hepatis perniciosum Bælz, 1883, 234.—Billet, 1893a, 509 (syn. of D. sinense Cobbold).—Caræs, 1888a, 41.—Cobbold, 1884g, 976.—Grall, 1887a, 460, 469.— Katsurada, 1900, 479.—Looss, 1905, 90 (syn. of Opisth. sin.).—See Clouorchis.

hepatium Rivolta, 1884, 27, for hepaticum.

heteroclitum Mol., 1859, 289 (in Perdix coturnix; Batavii); 1861, 203.—Braun, 1893a, 874; 1902b, 116.—Dies., 1859c, 430.—Looss, 1899b, 650 (thinks this may be a Clinost.).—Mont., 1893, 155.—Stoss., 1892, 174 (to Mesogonimus).

hcterolecitho-les Braun, 1899a, 3 (in Porphyrio porphyrio; from Madagascar, Africa); 1899, 632; 1899, 133–135, 300 (in Gallinula chloropus, Porphyrio porphyrio); 1902b, 109.—Loos, 1899b, 635 (type of Athesmia); 1902m, 790.—Jacoby, 1899a, 133–135; 1899b, 300 (in Gallinula chloropus, Porphporph.); 1899c, 1–30, 2 pls.; 1900, 1–11, 12, figs. 1–5.

heteromorphum Crep., 1837a, 317 (in Trigonocephalus sp. dub.); 1846a, 147 (in Trigonocephalus sp. dub.)

Trigonoc. sp. dub.).

heterophies Perroncito, 1879, 6, for heterophyes.

heterophies Perroncito, 1879, 6, for heterophyes.
heterophyes Sieb., 1852, 62-64, pl. 5, figs. 16-17 (in Homo: Egypt); 1853, 455, pl. 5, figs. 16-17.—Aitken, 1866, 804, 839; 1872, 146, 205; 1874, 58.—Arschoff, 1892, 495.—Bilharz, 1856a, 50.—R. Bl., 1888a, 625, 627, fig. 325, 631; 1891s. 791; 1891p, 609, 610, 611.—de Bonis, 1876, 163; 1882, 179.—Braun, 1883a. 66; 1892, 50; 1892a, 581, 642, 707, 735, 738; 1895b, 143-144, fig. 59; 1900h, 3, 6; 1901e, 334, 336, 337; 1903, 3. ed., 164 (to Cotylogonimus).—Cobbold, 1860a, 6; 1864, 194; 1866, 6; 1876, 210, 211; 1879b, 34-35, fig. 4; 1883, 401 (hyterophytes).—Dav., 1877a, 1xxvii.—Dies., 1858e, 32.—Dunglison, 1893, 338. \$20, 1174.—Eichhorst, 1901, v. 1, 301.—Goubert, 1878, 101.—Günther, 1858, 207-208.—Hahn & Lefèvre, 1884a, 546.—Harley, 1864a, 62.—Hoek, 1859, 42 (or Pag. id.).—Hoyle, 1890, 538.—Huber, 1896a, 579.—Ijima, 1889b, 147.—Jacoby, 1900, 22, 23 (D. fraternum and D. heterophyes with D. lingua and D. concavum together form one group).—Jægers., 1898, 7, 8, 9, 12 (com-147.—Jacoby, 1900, 22, 23 (D. fraternum and D. neterophyes with D. lingua and D. concavum together form one group).—Jægers., I898, 7, 8, 9, 12 (compares D. het., D. fraternum, D. lingua).—Jamieson, 1897a, 74.—Janson & Tokishige, 1892, 350.—Janson, 1893c, 265.—Kholodk., 1898, 26, 30, pl. 11, fig. 14; 1899a, 152.—Kuech., 1855, 210–212, pl. 4, figs. 11–12.—Kuech. & Zuern, 1881, 338.—Leuck., 1863a, 526, 613–616, 619, fig. 267; 1889, 399.—Looss, 1894d, anatomy, 1–42, pl. 1, figs. 1–8, pl. 2 (figs. 9–12); 43, 44, 45, 46, 47, 48, 49, 51, 52, 55; 1894a, 168; 1895a; 1896b, 60, 61, 63–64, 156, pl. 5, figs. 28, 10, 1896b, 863–864, 1899b, 533, 539, 550, 556, 585 (type of Cenogonimus). 38-40; 1896h, 863-864; 1899b, 533, 539, 550, 556, 585 (type of Comogonimus), 700; 1900, 607; 1902m, 804, 805, 886.—Luehe, 1899, 538, 539 (type of Cotylogonimus).—Manson, 1901, 541 (in man in Egypt; apparently no morbid symptoms); 1903, 3. ed., 664 (to Mesogonimus).—Moniez, 1896, 86, 133, 141–144, fig. 28.—Mont., 1893, 95, 155, 157; 1896, 168.—Mosler & Peiper, 1894, 177, fig. 69.—Mueh., 1898, 81, 82 (compares D. het. and D. fraternum with D. concavum, which he looks upon as Mesogonimus).—Paul, 1860, 20.—Perroncito, 1879, 6 (heterophies).—Rail., 1890, 138 (to Mesogonimus).—Roberts, 1888, 673.—Sandwith, 1899, 591; 1899, Sept. 30, 888 (case).—Schneidemuehl, 1896, 302.—Simon, 1897, 209, 223–224.—Sons., 1883, v. 1, 155 (found by Sons., Egypt); 1896, 295, 299, 314.—Stiles, 1904i, 44 type of Heterophyes.—Stiles & Garrison, 1906a, 76.—Stiles & Hass., 1900a, 563 (type of Heterophyes).—Stoss., 1892, 31–32 (to Mesogonimus); 1898, 42.—Swart, 1862, 37.—Verrill, 1870, 171.—Vogt, 1878, 10, 13, 14.—Wagner, 1883, 121.—Ward, 1895, 328 (in Homo); 1903, 870.—Wood & Fitz, 1897, 335.—Also reported for Canis familiaris. 700; 1900, 607; 1902m, 804, 805, 886.—Luehe, 1899, 538, 539 (type of Cotylo-Also reported for Canis familiaris.

heterophyes hominis Dies., 1855, 64, for heterophyes.

heteroporum Duj., 1845a, 402–403 (in Vespertilio pipistrellus; Rennes) to (Brachycelium).—R. Bl., 1891, 467–468.—Brand., 1888, 249, 250, pl. 17, fig. 4.—Braun, 1891d, 421; 1892a, 568, 579, 715, 766; 1893a, 911; 1893b, 185 (in Vesperugo pipistrellus); 1900, 224, 227; 1900, 388.—Cobbold, 1860a, 8; 1879b, 294.—Dies., 1850a, 382–383.—Kolenati, 1857, 12.—Linst., 1884, 139.—Looss, 1892, 66; 1896b, 86; 1899b, 547, 611, 614, 618, 718 (type of Pycnoporus); 1902m, 772.—Luehe, 1899, 536, 537.—Staff., 1903, 828.—Stiles, 1901, 197, 199, 200, 201, 202, 203.—Stoss., 1892, 12 (in Vannago pipistrellus; Hameln, Bennes) 202, 203.—Stoss., 1892, 12 (in Nannugo pipistrellus; Hameln, Rennes).

heterostomum Rud., 1809a, 50, 381–382 (in Ardea purpurea; t. l. apparently Europe); 1819a, 102–103, 388, 680.—Braun, 1892a, 578, 674, 721; 1893a, 872, 873, 910; 1899, 1 (Dicrocœlium); 1899g, 465, 484, 485, 486 (of Rud., 1809a, to Clinost.); 1900, 140–141; 1900h, 4, 9, 13, 14, 15. 16, 17, 18. 19 (includes the following as syns. of Clinost. heterostomum: Rud., 1809a, 1819a; Duj., 1845a; Dies., 1850a; Stoss., 1892; Par., 1896) (in Ardea cinerea, A. purpurea, Nycticorax griseus), 20, 24, 25, 26, 28, 29, 30, 31, 42, 43, pl. 1, figs. 1–2; 1901, 561.—Cobbold, 1860a, 10.—Dies., 1850a, 353 (includes Fasc. epatica of Rosa, in Ardea purpurea) — Duj. 1845a, 400.—Leuck, 1863, 503.—Leoss, 1890b, 650, 651. purpurea).—Duj., 1845a, 400.—Leuck., 1863, 503.—Looss, 1899b, 650, 651.—

MacCallum, 1899, 697, 704, 705, 706, 707.—Mont., 1893, 95.—Olfers, 1816, 44.—Par., 1896, 2.—Stiles & Hass., 1898a, 86 (includes C. gracile Leidy, 1856, type of Clinost. Leidy).—Stoss., 1892, 64; 1892, 159; 1898, 42.—Wright, 1879, 3, pl. 1, fig. 1.—Also reported for Ardea herodias.

heterostomum from Ardea purpurea in Turin collection ("C. No. 43 resp. C. No. 361") and in collection of Par. (collected at Geneva).—Braun, 1899g, 491

(Clinost. foliiforme); 1900h, 30.

heterostomum Rud. of Linst., 1883, 306; 1886, 30, fig. 49.—Braun, 1899g, 491 (not Clinost. heterostomum but perhaps Cl. complanatum).

heterostomum Rud. of Wright, 1879, and MacCallum, 1899.—Braun, 1899g, 491 (not Clinost. heterostomum but probably Cl. marginatum); 1900h, 29, 43.

heurteli Poir., 1885, 9, 10, pl. 23, fig. 2 (in Thynnus vulgaris).—Buttel-Reepen, 1902, 167, 172, pl. 6, fig. 13.

hians Rud., 1809a, 359-360 (in Ardea nigra; Greifswald); 1814a, 101; 1819a, 94, 366, 680.—Baird, 1853a, 51 (includes D. œsophagi ardeæ nigræ Viborg).—Ben., 1858a, 1861a, 171, 202; 1868, 295, 296, 298, 299–300, pl. 1, figs. 6–7.— Braun, 1891d, 424 (in Ardea cinerea); 1892a, 584, 662, 699, 764, 768, 776, 784, 785, 786; 1893a, 873; 1893, 354; 1895b, 17; 1899g, 485, 486, 489 (from Ardea cinerea, in Payia, and from Nycticorax griseus, in Cagliari, as syn. of Clinost. heterostomum); 1899, 465; 1900, 24, 25; 1900h, 14, 15, 17, 18, 19, 23; 1901b, 33; 1901, 896; 1901, 561; 1902b, 4.—Cobbold, 1860a, 10.—Condorelli, 1897c, 33; 1901, 896; 1901, 501; 1902b, 4.—Cobbold, 1800a, 10.—Condorelli, 1897c, in 118–124 (in Hydrocolæus minutus).—Crep., 1837, 311, 316, 324.—Dies., 1836, 248; 1850a, 337–338; 1858e, 333.—Duj., 1845a, 399.—Gamb., 1896a, 63.—Gurlt, 1845, 276.—Kuech., 1855, 192.—Leuck., 1879, 14, 15; 1886d, 11.—Looss, 1892a, 14; 1899b, 562–563 (type of Cathæmasia).—MacCallum, 1899, 706.—Mueh., 1896, 588–589; 1896, 252–257, figs. 3, 10; 1898, 28.—Mueller, 1897, 15–16, pl. 2, fig. 6.—Nathusius, 1837, 65.—Nord., 1832a, 90, 93; 1840, 617.—Olfers, 1816, 44.—Par., 1887, 331, pro parte (in Nycticorax griseus) (syn of Clinost. heterostomum teste Braun, 1900h, 19).—Sieb., 1835, 66, 73, 82.—Stoss, 1891, 111; 1892, 160 (syn. D. complanatum).—Waggener, 1857. 82.—Stoss., 1891, 111; 1892, 160 (syn. D. complanatum).—Wagener, 1857, 26.—Will.-Suhm, 1876, 337, 339.—Also reported for Ardea nycticorax, A. purpurea, Ciconia alba, C. nigra.

hippopodii Vogt, 1854, 97–98, 99. pl. 15, fig. 3 (in Hippopodius leteus; Quoy et Gaimard, Mediterranean).—Braun, 1893a, 852.—Graeffe, 1860a, 13.—Mont., 1888a, 77; 1888, 195; 1893, 124.

hirndinis Brand., 1888a, 13, misprint for hirudinis.

hirsutum Looss, 1896b, 68–73, 76, 78, 81, 98, pl. 5, figs. 45–49 (in caméléon; Alexandria); 1899b, 547 to Lecithodendrium.—Luehe, 1899, 536.—Stiles, 1901, 200.

hirudinis Henle.—[Crep., 1841a, 79 (Distome in Hirudo vulgaris), does not use combination].—Dies., 1850a, 418 (syn. of Heptast. hirudinum Schomburgk).

hirundinum Zed., 1800a, 163, 169-171 (for hirundinis Frælich) in Hirundo apus, H. urbica; Europe; 1803a, 210.—Braun, 1901, 566; 1902b, 46 (syn. of Plagiorchis maculosus).—Dies., 1850a, 349 (syn. of D. maculosum).—Rud., 1809a, $\bar{3}74.$

hispida ventriculi accipenseris sturionis Viborg, 1795, 243, see hispidum.

hispidum Abildg. in Rud., 1819a, 118, 423—424 (in Accipenser sturio; Arimini, Berlin); 1809a, 435 (syn. of D. sturionis for D. hispida accipenseridis sturionis).—Ben., 1870, 83.—Braun, 1892a, 567, 583, 594, 729, 737;—Carus, 1884, 126.—Cobbold, 1858b, 162, pl. 32, figs. 47—48, pl. 33, figs. 49, 50 (in Acipenser sturio); 1860a, 36.—Crep., 1829, 73—76; 1837, 311, 312, 325.—Dies., 1850a, 392—393.—Duj., 1845a, 470.—Kroyer, 1852—53a, 21, 778 (in Acipenser sturio; Osmerus eperlanus)—Leidy, 1887, 24.—Lint, 1901b, 414, 439, 478, 66, 291 Osmerus eperlanus).—Leidy, 1887, 24.—Lint., 1901b, 414, 422, 478, fig. 321, 322, 323 (in Phycis tenuis); 1905, 328, 334, 364, 400, 478 (includes as hosts: Menticirrhus americanus; Seriola Ialandi).—Looss, 1894a, 218; 1899b, 576, 581; 1901, 634.—Mehlis, 1831, 187–190.—Nord., 1832a, 90.—Odhn., 1902, 154.— Stoss., 1885, 156; 1886, 41; 1891, 216.—Also reported for Acipenser glaber, A. ruthenus, A. stellatus.

histrix Dies., 1850a, 393–394 for D. hystrix; 1858d, 268.—Duj., 1845a, 433.—Mol.,

1858, 131; 1861, 223.—Mont., 1888, 198.

holostomum Rud., 1819a, 94-95, 368 (in Rallus aquaticus; Vien. Mus.).—Braun, 1892a, 772; 1901, 561, 565; 1902b, 136 (syn. of Urogonimus macrostomus Rud.) (includes Dies., 1850a, 339; Duj., 1845a, 446; Rud., 1819a, 94, 368; Sieb., 1848, 144; 1853, 433; Walter, 1866, 14; Zeller, 1874, 574), 137.—Cobbold,

1860a, 11.—Dies., 1850a, 339, includes D. ralli; 1858d, 277.—Duj., 1845a, 446.—Fil., 1855b, 25.—Moul., 1856a, 21, 183 (in Rallus aquaticus; Gallinula porzana; G. chloropus).—Par., 1896, 2.—Sieb., 1836, 234.—Stoss., 1892, 145, to Cladocœlium.—Wagener, 1857, 24, 45.—Walter, 1866, 64.

homoeostomum Dies., 1858e, 343 (D. Triglæ pini Bellingham, 1844a, 428 renamed) (t. h. Trigla (Pini) cuculus; in ventriculo, Hibernia, Bellingham).—Cob-

bold 1860a, 30.—Stoss., 1886, 50.

homolostomum Linst., 1887, 104–105, pl. 2, figs. 5, 6, 17a (in Limnæa stagnalis).— Braun, 1893a, 873.

horridum Leidy, 1850, 303-304, pl. 43, fig. 1 (in Boa constrictor); 1850f, 118; 1856b, 44; 1904a, 41, 87, 227.—Braun, 1893a, 876; 1893d, 467.—Cobbold, 1860a, 20.— Dies., 1858e, 355.—Looss, 1894a, 196.—Luehe, 1899, 531, 532.—Sons., 1890,—; 1893, 499; 1893, 215–216; 1893, 2 pp.; 1893, 28 Oct., 566.—Stoss., 1895, 220 (in Python molurus); 1904, 2.—Volz, 1899, 235, 237.

hospitale Staff., 1900, 403, fig. 3 (in Diemyctylus viridescens); 1902, 481 (in Pletho-

don) to (Brachycœlium).

hyalinum Rud., 1809a, 389 (in Salmo eriox) Fasc. eriocis renamed; 1819a, 105.— Cobbold, 1860a, 26.—Dies., 1850a, 363.—Duj., 1845a, 465.—Harz, 1881c, 5 (syn. Fasc. eriocis Mueller).—Kroyer, 1843–45a, 624 hyatinum (in Salmo eriox).—Nord., 1840, 620 (syn. of Fasc. eriocis).—Olfers, 1816, 46.—Stoss., 1886,

hyans Moul., 1856a, 49, misprint for hians.—Mehlis, 1831, 190.

hyatinum Kroyer, 1843-45a, 624 misprint for hyalinum.

hylæ Rud., 1819a, 121 (in Hyla arborea).—Dies., 1850a, 342 (syn. of D. cygnoides).

hystrix Duj., 1845a, 433 (in Pleuronectes maximus, P. platessa).—Braun, 1892a, 655; 1893a, 865, 871, 873.—Carus, 1884, 128.—Dies., 1850a, 393 (histrix); 1858e, 353 (in Merlangus carbonarius, Lepidoleprus trachyrhynchus; Lophius piscatorius; Rhombus maximus).—Fil., 1855b, 18, 19.—Gamb., 1896a, 72.—Kroyer, 1838–40a, 610, 612 (in Platessa vulgaris; Rhombus maximus).—Looss, 1899b, 581, 696 (to Stephanost.).—Mol., 1861, 223.—Moul., 1856a, 212 (Filippi believes Cerc. echinocerca is the young form), 219 (in Pleuronectes maximus; P. platessa).—Olss., 1868, 52.—Stoss., 1886, 37.

hyterophytes Cobbold, 1883, 401 (for heterophyes).

[illotum Sluiter, 1898 (a tunicate).]

imbutiforme Mol., 1859, 839, 844–845 (in Labrax lupus; Padua).—Carus, 1884, 127.—Looss, 1899b, 576, 578, 581, 582; 1901, 631.—Mont., 1893, 177.—Stoss., 1883, 115; 1886, 36.

imitans Mueh., 1898a, 17–18 (in Abramis brama; Kænigsberg i. Pr.); 1898b, 25, 91, fig. 8b.—Looss, 1899b, to Asymphylodora.

imparispine Lint., 1905, 327, 334, 371, figs. 189-194, in Rachycentron canadus; N.C. incerta Cobbold, 1885g, 177–178, 1 fig. (in Coluber; Maldonado, Rio Plata).—Braun, 1893a, 872.—Stoss., 1895, 230.—West, 1896, 322.

incistidata Erc., 1881e, 96, pl. 2, fig. 10, 11, 12; 1882a, 332 (probably incistidata simply means incysted and is not used as a specific name).

incisum Rud., 1809a, 361, 435 (D. anarrhichæ lupi Rathke, renamed); 1819a, 94, 122.—Ben., 1870, 48, pl. 4, fig. 5.—Braun, 1893a, 875.—Cobbold, 1860a, 22.—Dies., 1850a, 339.—Duj., 1845a, 461–462.—Jacoby, 1900, 12, 13.—Kroyer, 1838–40a, 380 (in Anarrhichas lupus).—Linst., 1903, 278.—Odhn., 1905, 310.— Olfers, 1816, 45.—Type of Fellodist., 1904.

incivile Leidy, 1856b, 44 (in Leiostomus obliquus); 1904a, 87.—Cobbold, 1860a, 20.—Dies., 1858e, 350 (in Leiost. obliq.; Philadelphia).—Stoss., 1886, 40; 1895,

231.—Also reported for Sciæna obliqua.

inclusum Polonio, 1859, see Par., 1894, 149 (in Triton punctatus; Padova).

incommodum (Leidy, 1856) Leidy, 1891a, 414 (includes D. oricola Leidy, 1884); 1904a, 235.—Mont., 1892, 715.

incomptum Stoss., 1886, 51, for incomtum.

incomtum Rud., 1819a, 683 (in Chætodon sp., Brazil).—Cobbold, 1860a 27.— Dies., 1850a, 367.—Duj., 1845a, 459.—Stoss., 1886, 51 (incomptum).

inconstans Lint., 1905, 327, 334, 400, fig. 183-187, in Chætodipterus faber; N. C.

 $\begin{array}{l} incrassatum \ {\rm Dies.,\ 1850a,\ 390-391\ (in\ Lutra\ solitaria;\ Brazil);\ 1855,\ 68,\ pl.\ 3,\ figs.} \\ 22-25;\ 1858e,\ 350\ (in\ L.\ sol.).--Braun,\ 1901e,\ 317-318,\ 328,\ fig.\ 5,\ 8,\ 9;\ 1902b,\ 27.--Cobbold,\ 1879b,\ 298.--Mont.,\ 1888a,\ 14.--Stoss.,\ 1892,\ 30\ (to\ Echinost). \end{array}$

increscens Olss., 1868, 36–37, pl. 4, fig. 83 (in Scomber, Merlucius, Hippoglossus).—
Ben., 1870, 37.—Braun, 1892a, 699.—Lint., 1901, 415, 479.—Mont., 1893, 95.—Odhn., 1905, 332 (in Scomber), 338 (syn. of Lepodora rachiæa, type).—
Stoss., 1886, 42, to (Echinost.) (in Hippoglossus maximus, Merlucius vulgaris, Scomber scombrus).—Also reported for Gadus merlangus, G. morrhua.

inerme Nitzsch, MS., in Rud., 1819a, 375 (in Anas boschas fera) (syn. of D. oxycephalum).—Dies., 1850a, 346 (syn. of D. oxyc.).

inerme Fil., 1857c, pl. 2, fig. 16, for D. inerme paludinæ impuræ.—Erc., 1881e, 35; 1882a, 271.—Harz, 1881c, 4.

inerme Linst., 1879, 183 (in Petromyzon fluviatilis).

inerme paludinæ impuræ Fil., 1857c, pl. 2, fig. 16.—Erc., 1881e, 33; 1882a, 269.

inflatum Crep., 1849, 64 (in Alauda arvensis).

inflatum Mol., 1859, 826 (in Anguilla vulgaris; Padua).—Braun, 1892a, 699.—
Looss, 1899b, 576, 581, 582; 1901, 634.—Mont., 1888a, 14 (? Creplin or ? Molin); 1893, 83, 95.—Odhn., 1902, 154.—Olss., 1868, 37.—Par., 1887, 335, pl. 6, fig. 37.—Stoss., 1885, 157; 1886, 42; 1902, 582.

inflexum (Rud., 1802) Rud., 1809a, 395–396 (includes Fasc. jesis Gmelin), host Cyprinus jeses; 1819a, 106.—Cobbold, 1860a, 26.—Crep., 1837, 326.—Dies., 1850a, 365–366 (includes Bloch, 1782a, 11, pl. 2, figs. 10–11; Fasc. jesis Gmelin, 1790a, 3058; F. inflexa Rud., 1802, 82–83; D. carinatum Zed., 1803a, 217).—Duj., 1845a, 463.—Erc., 1881e, 56; 1882a, 292.—Nord., 1832a, 88.—Olfers, 1816, 46.—Stoss., 1886, 51.—Wagener, 1857, 44.—Also reported for Idus melanotus.

ingens Moniez, 1886b, in 531–543, pl. 15, figs. 1–10 (host unknown); 1887d,100–102; 1887e, 271; 1887f, 242–243; 1896, 143.—Bl., 1888a, 543; 1891r, 692–693 (syn. of D. clavatum Rud.).—Braun, 1889a, 397 (host unknown); 1892a, 586, 690; 1893b, 184.—Buttel-Reepen, 1900a, 585, 586, 587, 588, 590, 591, 592; 1902, 167, 171, 172, 173, 176, 196, 202, 206, pl. 6, fig. 18.—Darr, 1902, 666, 677, 678, 685.—Hoyle, 1890, 540.—Mont., 1888a, 39, 47; 1893, 22, 26, 27, 34, 36, 65, 72, 73.

innocuum Bælz of Taylor, 1884, 53 (for D. hepatis innocuum).—Caraes, 1888a,
 36ff.—Grall, 1887a, 460.—Ijima, 1899b, 139.—Laspeyres, 1904a, 6, 12
 (inocuum).—Looss, 1907, Feb. 1, 141, 148 (syn. of Clonorchis sinensis).

innocuum hepatis La Clínica de Málaga, 1883, 309 (for D. hepatis innocuum).— St.-Remy, 1883, 529.

inocuum Laspeyres, 1904a, 6, 12, for innocuum.

insigne Dies., 1850a, 347 (in Echinorhinus spinosus) (D. scimna Risso, 1826, renamed); 1858e, 335.—Ariola, 1899, 7, 9 (syn. of D. veliporum Crep.).—
Braun, 1892a, 586, 591, 592, 593, 603, 608, 617, 623, 624, 625, 628, 635, 647, 655, 666, 669, 677, 682, 686, 690, 708, 712, 717, 719, 724, 731, 733; 1903a, 873.—Buttel-Reepen, 1902, 170, 171, 202, pl. 6, fig. 23.—Crep., 1851, 1, 296.—Darr, 1902, 683.—Fischer, 1883a, 29.—Jægers., 1900, 72.—Jourdan [1881b], 5, 9.—Kerbert, 1881a, 551, 557, 574.—Lander, 1904a, 7.—Linst., 1903, 354.—Looss, 1885b, 5, 10, 17, 26; 1894a, 152, 198, 199.—Mont., 1893, 27, 33, 34, 52, 77, 95, 102, 107, 192.—Olss., 1896, 508 (cf. D. veliporum Crep.).—Poir., 1885, 4, 21, 25, 26, 30, 32, 33, 37, 40, 42, 44, 48, 49, 51, 53, 61, 62, 70, 74, 75, 80, 82, 83, 87, 88, 94, 97, 105, 106, 107, 109, 117, 118, 120, 127, 129, 142, 150, pl. 33.—Villot, 1876, 1345; 1878, 2, 3–18, pl. 5, fig. 8, pl. 6, figs. 1–9, pl. 7, figs. 1–3, pl. 8, figs. 1–10; 1882, 506, 507.—Wolf, 1903, 619.—Ziegler, 1883, 545.

instabile Duj., 1845a, 412 (in Sorex fodiens; Rennes), to (Brachylaimus).—Braun, 1901, 342.—Cobbold, 1860a. 8; 1879b, 296.—Dies., 1850a, 387.—Stoss., 1892,

16 (in Crossopus fodiens; Rennes).

intermedium Mehlis, in Crep., 1846. 138, 139 (in Colymbus cristatus, C. subcristatus).—Dies., 1850a, 397.—Stoss., 1892, 183.

[intersectus Lænnec, 1807, 1812c, 9-12 (Distomus n. g.), in Homo.]

[intestinale Rud., 1819a, 119, see D. aluconis intestinale.]

intestinale Taylor, see Carter, 1862a, xxxi, in int. of Homo; District of Dacca, India.

intestinalis ardex nigræ Viborg, 1795, 242.

intestinalis testudinis mydæ Viborg.—See Rud., 1809a, 433 (syn. of D. testudinis mydæ Rud.).

intestinalis vulpis Viborg, 1795, 242.

invaginatum Mayer, 1841a, 17, D. appendiculatum renamed.

involutum Rud., 1809a, 377–378 (in Upupa epops), includes Fasc. upupæ Schrank,
1790, and D. fusiforme Zed., 1800a, from Upupa epops; 1819a, 101 (July).—
Braun, 1901, 561.—Cobbold, 1860a, 13.—Dies., 1850a, 351 (includes Fasc. upupæ Schrank, 1790, 123; D. fusiforme Zed., 1800a, 171, and 1803a, 210).—
Olfers, 1816, 44.—Stoss., 1892, 183.

irroratum Rud., 1819a, 105, 393–394 (in Testudo mydas; Arimini, May).—Braun, 1893a, 873; 1893d, 466; 1899b, 715, 717–718, 722; 1901b, 20, 36–38, figs. 27, 30, 32.—Carus, 1884, 129.—Cobbold, 1860a, 20.—Dies., 1850a, 364 (in Halichelys atra; Arimini).—Duj., 1845a, 451–452.—Looss, 1899b, 590 (belongs to or is closely related to Astia judging from Braun, 1899, 717); 1901l, 558, 559, 560 (ex parte syn. of Pachypsolus lunatus (Looss, type of genus); 1902m, 485 (to Pachypsolus; includes P. lunatus (Rud.) Looss, 1901, 558), 486, 487, 493, 494, 496, 497, 499, 503, 504, type of Pachypsolus, 505.—Mont., 1892, 715; 1896, 165.—Par., 1894, 147 (in Thalassochelys caretta; Rimini).—Stoss., 1895, 231.—Also reported for Thalassochelys corticata.

ischnum Leidy, 1890, 415 (in Saurus feetens; Beach Haven, N. J.).

isoporum Looss, 1894a, 2, 49–56, pl. 1, figs. 15–18, pl. 5, figs. 102–112 (in Cyprinus carpio, Phoxinus leevis, Leuciscus rutilus, Abramis brama, Squalius cephalus, Tinca vulgaris, Esox lucius), 58, 59, 97, 124, 127, 136, 137, 140, 144, 153, 154, 157, 159, 162, 167, 179, 191, 192, 208, 209, 212, 214, 215, 218, 231, 251, 264; (includes Fasc. longicollis Fredich, e. p.; D. globiporum Rud. of Olss.); 1894, 17; 1896b, 46; 1899b, 570 (type of Creadium); 1900 (type of Allocreadium).— Braun, 1900, 232; 1901b, 33.—Hausmann, 1897b, 4, 6, 17, 20, 22, 27 (in Barbus fluviatilis).—Kowal., 1896d, 3 (253) (in Cyprinus carpio; Dublany).—Linst., 1903, 281.—Odhn., 1901, 483.—Staff., 1902, 481 (in Lemotilus bullaris).— Stoss., 1901, 94 (6).

isoporum armatum MacCallum, 1895, 401–406, figs. 1–4 (in Aplodmotus grunniens, Lepomis gibbonus, Accepenser rubicundus; Lake Erie and Grand River of Ontario).—Looss, 1902, 785 (not Allocreadium isoporum, but allied to D. sophiæ).—Type no. 6856 U. S. Nat. Mus.

sopmæ).—Type no. 0850 °C. S. Nat. Mus.

isostomum Rud., 1819a, 105, 392–393 (in Astacus fluviatilis).—Baer, 1827, 553.—

Bettend., 1897a, 29, 30; 1897, 333, 334.—Braun, 1892a, 632, 683, 684, 685, 686.—

Cobbold, 1860a, 30.—Crep., 1829b, 64–66; 1837, 310, 325, 326.—Dies., 1850a, 363 (includes Carus, 1818, 51; D. cirrigerum Baer?).—Duj., 1845a, 471–472.—

Gaffron, 1883a, 508, 509.—Harz, 1881a-b; 1881c, 1.—Haswell, 1887a, 294.—

Jackson, 1888, 643–644.—Kampmann, 1894b, 452, 453, 454, 455, 457, 458, 459, 462, pl. 19, figs. 1–5.—Knoch, 1894a, 5, 13, 15, 16, 17.—Linst., 1903, 281.—

Looss, 1893b, 809; 1894a, 8, 10, 20, 143, 145, 146, 149.—Mehlis, 1831, column 184.—Mont., 1888a, 47, 48; 1893, 65, 66, 67, 69, 74.—Moul., 1856a, 217.—

Mueh., 1898, 14.—Poir., 1885, 147.—Zaddach, 1881, 893.

italicum Stoss., 1893, 135 (6) (in Lichia amia; Triest, Sept.); 1898, 51–52.— Jacoby, 1900, 17.

jacksoni Braun, 1892, 44; 1893d, 466, for jacksonii.

jacksonii (Cobbold, 1869) Braun, 1892a, 567, 650, 674, 710, 875, 910; 1892, 44 (jacksoni); 1893d, 466.

japonicum Bl., 1888a, 596, 618-621, figs. 320-322 (in Homo) (includes D. hepatis endemicum sive perniciosum; D. hepatis innocuum), 631; 1891, 607 (syn. of D. sinense Cobbold).—Billet, 1893a, 509 (syn. of D. sinense Cobbold).—Braun, 1893f, 386, 425; 1903, 3 ed., 161 (syn. of Opisthorchis sinensis).—Brunet, 1902a, 125.—Gamb., 1896a, 63.—Huber, 1896a, 577 (syn. of D. spatulatum Lkt.).—Katsurada, 1900, 479.—Looss, 1905, 90 (syn. of Opisth. sin.).—Simor, 1897, 223.—Sons., 1889, 278 (syn. of D. endemicum Bælz).—Ward, 1895, 328 (in Homo) (see D. sinense); 1903, 870.

kampanulatum Schneidemuehl, 1896, 302, for campanulatum.

kölikerii Cobbold, 1860a, 30 (D. pelagiæ Kælliker renamed) (in Argonauta argo, Pelagia noctiluca).—Mont., 1893, 122, 124.

köllikerii Mont., 1893, 122, 124, corrected form of kölikerii.

kommutatum Schneidemuehl, 1896, 303, for commutatum.

kongenitum Schneidemuehl, 1896, 302, for conjunctum.

krassum Schneidemuehl, 1896, 302, for D. crassum.

kuneatum Schneidemuehl, 1896, 303, for D. cuneatum.

labiatum Rud., 1819a, 108, 400 (in Syngnathus pelagicus; Naples, July).—Braun, 1893a, 875.—Carus, 1884, 129.—Cobbold, 1860a, 27.—Dies., 1850a, 370.—Duj., 1845a, 468.—Pag., 1862, 305.—Stoss., 1886, 51.

labii Linst., 1889, 79, for labri Stoss.

labracis Duj., 1845a, 398, to (Dicroccelium), (in Labrax lupus; Rennes).—Barbagallo & Drago, 1903, 410 (in Labrax lupus; Catania).—Ben., 1858a, 1861a, 179; 1870, 24, to (Echinost.).—Dies., 1850a, 399.—Fraip., 1880c, 417.—Kroyer, 1838-40a, 578 (in Lab. lup.).—Looss, 1901, 399.—Luehe, 1900, 487.—Mol., 1859, 843.—Odhn., 1901, 513, 514.—Stoss., 1898, 46-47 (in Lab. lup.; Triest).—Also reported for Labrus maculatus.

labri Rud., 1819a, 122 (in Labrus rupestris; C. E. V.).—Dies., 1850a, 344 (syn. of

D. fasciatum).

labri Ben., 1870, 45, n. sp. (in Labrus maculatus).—Looss, 1901d, 399.

labri Stoss., 1886, 30 (in Labrus mixtus) (nec Rud.); 1887, 91–92; 1887, 186; 1898,
45.—Linst., 1889, 79 (labii).—Odhn., 1901, 493, 494, 495; 1902, 160 (syn. of Helicometra pulchella).—Sons.. 1891, 257, 258.

labri rupestris Olss., 1876, 20–21.—Mont., 1891, 500.—Reported for Ctenolabrus rupestris, Labrus rupestris.

lacertæ Rud., 1819a, 121 (in Lacerta cærulescens; C. E. V.).—Dies., 1850a, 355
 (syn. of D. mentulatum) (in Lacerta agilis; Berlin).—Stoss., 1895, 218.—
 Also reported for Lacerta viridis.

laciniatum Duj., 1845a, 437 (in Simia maimon; Paris), based on Fasc. de Brongniart, Alaire de Brongniart of Blainv., Dict. sc. nat., pl. 41, fig. 3; 1824, 518; atlas, pl. 2, fig. 8; 2. ed., pl. 14, fig. 15; type of Alaire, not Alaria Schrank.—Braun, 1893a, 875; 1893d, 467 (in Cynocephalus maimon); 1901e, 311.—Cobbold, 1860a, 8; 1879b, 289.—Dies., 1850a, 374 (includes Fasc. maimonis Blainv.—Gurlt, 1845, 224.—Stoss., 1892, 35 (in Papio mormon.)

lære Lint., 1898, 517-518, pl. 43, figs. 5-8, pl. 45, fig. 1 (in Macrourus bairdii), type U. S. N. M. 4852; 1901, 415, 418, 481.—Looss, 1899b 641, to Hemiurus.

lagena Brand., 1888, 249, 250 (ascidia Ben., 1873, nec Rud., 1819, renamed).—
Braun, 1900, 388.—Looss, 1899b, 556, 609 (type of Lecithodendrium), 618, 715, 716; 1902m, 827; 1907, Mar. 5, 484 (of Looss, 1899b, 715, syn. of Lecithodend. granulosum).—Stiles, 1901, 200.

lagena (Gmelin, 1790) Rud., 1809a, 366.

lageniforme Lint., 1898, 524-525, pl. 47, fig. 12 (in Remora remora); 1901, 415, 421, 473.

lancea Dies., 1850a, 334 (in Delphinus tacuschi; bil. duct, Barra do Rio Negro, Brazil); 1855, 64, pl. 2, figs. 17-19; 1858e, 333.—Ben., 1870, 362.—Braun, 1892a, 574; 1893a, 875; 1893, 354; 1900g, 250; 1901e, 314.—Cobbold, 1860a, 6: 1876, 35, pl. 10, fig. 1; 1879b, 416, 417, fig. 67.—Linst., 1886, 125.—Looss, 1902m, 790 (amphitypie).—Mont., 1888a, 8.—Stoss., 1892, 26-27 (in Orcella brevirostris; Delphinus tacuschi; Brazil).—Weski, 1900, 4 May, 579-583, 1 fig.

lanceatum (Stiles & Hass., 1896) Looss, 1899b, 556.

lanceolatum (Rud., 1803) Mehlis, 1825, 42 pp. [see lanceatum].—Aitken, 1866, 804, 839; 1872, 146, 205; 1874, 58.—Anacker, 1892c, 94.—Anders, 1903, 6 ed., 1245 (in man).—Anglas & Ribacourt, 1901, 313–352; 1902, 313–354, 38 figs. (anat. hist.); 1902, Dec. 30, 840–841; 1905, July, 50.—Aragon, 1896, 452.—Armatage, 1895, 429.—Aschoff, 1892, 493–496, pl. 13, fig. 4, case in Homo; 1893, Aug. 25, 256.—Askanazy, 1900b, 491.—Baillet, 1866b, 18, 90, 104–105.—Baldi, 1900a, 222–224 (in horse; Milan) [224 lanceulatum]; 1900b, 123.—Bettend., 1897a, 25.—Biermer, 1863a, 395.—Bilharz, 1856a, 49.—E. Bl., 1847, 292–295, pl. 12, fig. 1.—R. Bl., 1888a, 590, 597, 602 (includes Fasc. hepatica Bloch, 1782; D. hep. Zed., 1800; Rud., 1810; Plan. latiuscula Gœze, 1782), 612 (description, eggs, embryo, sporocysts, redia, cercaria, adult; cases in man [five authentic cases to date reviewed], distribution), figs. 312–313, 615, 623, 631; 1891, 466–467; 1891, 610.—Bolotoff, 1890a, 695–696.—de Bonis, 1882, 172, pl. 2, fig. 13.—Bos, 1894, 245.—Bossuat, 1902, v. 6 (2), 62.—Brand., 1888a, 29; 1890a, 558.—Braun, 1883a, 62, 63–64, 70, 71, figs. 12, 16; 1895b, 11, 65, 144–145, figs. 60–62 (says 7 cases in man); 1892a, 586, 592, 598, 601, 602, 622, 635,

MA—Continued.
638, 644, 661, 664, 669, 672, 673, 677 (anceolatum), 678, 682, 692, 700, 704, 705, 706, 712, 714, 717, 719, 724, 727, 728, 730, 732, 733, 745, 757, 758, 762, 763, 764, 766, 778, 780, 784, 785, 787, 789, 791; 1893, 347, 348, 349, 350, 351; 1893a, 857, 875, 878, 910; 1893b, 184, 185 (in Lepus variabilis, Anodonta sp.); 1893f, 384, 385, 386, 389, 390, 424; 1894i, 605, 606; 1899, 3; 1901b, 702; 1903, 3 ed., 157 (of Sieb., 1836, Tright, 1829, not Mehlis, sp., of Opisthorchis felineus), 166 (Mehlis, 1825, to Dicroccelium).—Brunet. 1902a, 125.—Buchholz, —, 64.—Bettel-Reepen, 1902, 203.—Caraes, 1888a, 41.—Chatin, 1886b, 244; 1887a, 597.—Cobbold, 1855b, 4; 1864a, 184–191, fig. 39–40; 1866, 6, 7; 1872b, 91, 92; 1876, 210, 211, 303; 1879b, 17–20, 25, 28, 35, 36, 49, 318, 331, 404, figs. 1, 18; 1883, 401.—Cornil & Petit, 1901a, 178.—Cosmovici, 1891, 15 Oct., 247–248.—Crep., 1837, 313, 318, 321, 322, 323, 325, 326; 1839, 288.—Creutzberg, 1890a, 6, 7, 21.—Darr, 1902, 680.—Dav., 1877a, 1xxvi, 240–257, figs. 2, 7, 39–40.—Delafield & Prudden, 1897, 130.—Delafond, 1854a, 30.—Dies., 1850a, 333–334; 1858, 58; 1858e, 332; 1859c, 427.—Duffek, 1902a, 774.—Duj., 1845a, 391–392 to (Dicroccelium).—Dunglison, 1893, 338, 820, 1174.—Eiss, 1838, 22.—Erc., 1881e, 69, 90, 91; 1882a, 305, 326, 327.—Fischer, 1840, 158.—Fleming, 1881a, 90, 91.—Florance, 1866a, 5, 22.—Fraip., 1880a, 398; 1880e, 418, 426, 428; 1881b, 39; 1883a, 35.—Friedberger, 1878a, 160; 1880a, 254.—Galli-Valerio, 1893a, 174, 175, 178, 181, pl. 2, figs. 1, 5, 6.—Gamb., 1896a, 63.—Gomy, 1897a, 372, 374.—Grall, 1887a, 469.—Gronkowski, 1902a, 515, 519 (300), 497 (in Duiker antelope).—Jima, 1889b, 134.—Jackson, 1888, 69, 63.—Gomy, 1897a, 37, 374.—Grall, 1887a, 469.—Gronkowski, 1902a, 515, 519 (300), 497 (in Duiker antelope).—Jima, 1889b, 134.—Jackson, 1888, 69, 63.—Asatsenbaum, 1899, 73, 74.—de Jong, 1896a, 1, 9, 10 (of van Tright), 10 (of Sieb.) (in Felis catus), 10, 11 (of Mehlis).—Kamensky, 1900a, 5, 6.—Kastenbaum, 1893, 434; 1895e; 1900.5, 9–10, 11, figs. 6-7.—Jami 633; 1905, 88 (to Dicrocelium).—Macé, 1882a, 8, 9, 12, 21, 29, 60.—Manson, 1901, 540 (supposed intermediate host Planorbis marginatus).—Moniez, 1892, 77-79; 1896, 86, 102, 103, 104, 108, 118, 122, 136, 138, fig. 24.—Mont., 1888a, 35, 40, 56, 57, 72; 1891, 110; 1891, 117 (Distomun): 1893, 28, 83, 88, 95, 99, 102, 106, 107; 1896, 162.—Mosler & Peiper, 1894, 171, 175-176, figs. 67-68.—Moul., 1856a, 18, 32, 39, 41, 43, 44, 47, 50.—Nicoll, 1906, 521.—Nord., 1832a, 47; 1840, 547, 616.—Packard, —, 522.—Padrone, 1904, 489 (Distomi lanceolato).—Pag., 1857, 53.—Perroncito, 1885.—Piana, 1882, —,—Poir., 1886, 29.—Rail., 1890, 143; 1893a, 361 (felis cati Sieb., 1836, and van Tright, 1889=D. felineum); 1897, 1134.—Rail. & Marotel, 1898, 30, 33, 38.—Ratz, 1898, 67, 68; 1898, 298; 1900, 141.—Raum, 1883, 10.—? Rayer, 1846, 20-23, 4 figs.—Reynes, 1869, 30-36.—Rindfleisch, 1884, 210.—Roewer, 1906, 192, 193, 196, 218.—Schmalz, 1831, 24.—Schneidemuehl, 1896, 300.—Schuberg, —, 88; 1895, 168, 181, 184, 185, 186, 187, pl. 10, figs. 1-7.—Shaw, 1901, 619, fig. 220; 1901, 1027.—Sieb., 1835, 64, 65.—Signol, 1884, 232, fig. 229.—Simon, 1896, 182, 192, fig. 52; 1897, 209, 223, fig. 53.—Sons., 1889, 275; 1893, 215, 216; 1896, 332.—Stewart, 1898, 328.—Stiles, 1898a, 55.—Stoss., 1892, 22-23 (reported for Antilope dorcas; Auchenia lama; Capra hircus; Bos taurus, Europa, America sett.; Cervus elaphus; Dama vulgaris; hircus; Bos taurus, Europa, America sett.; Cervus elaphus; Dama vulgaris; Equus asinus; Felis domestica; Homo sapiens, Weimar, Kaplitz in Bohemia; Lepus cuniculus; L. timidus; L. variabilis, Briançon; Ovis aries, Normandia, America sett.; Sus scrofa, America sett.).—Swart, 1862, 35–36.—Tyson, 1903, 3 ed., 1180.—Valentin, ——, 147.—Verrill, 1870, 171, 175, 176, 177, 178, 219.—Vogt, 1878, 9, 38, 42, fig. 29.—Wagner, 1883, 121.—Walter, 1858, 268–297, figs.—Ward, 1895, 238–244 (from cat and dog, syn. of D. felineum), 328 (in Homo; biliary ducts), 332 (in Bos taurus; biliary ducts),

335 (in Ovis aries; biliary ducts), 341 (in Canis familiaris) (error for D. truncatum and D. felineum).—Weichselbaum, 1898, 315.—Will.-Suhm, 1870, 1, 4, 5.—Wolf., 1903, 617.—Wood & Fitz, 1897, 335.—Zschokke, 1892, 500.—Zuern, 1882, 208, 209, 211, 218, pl. 4, figs. 1—4.

lanceolatum of Sieb., 1836, 113; 1850, 672-673 (in Felis catus dom. at Danzig), and of van Tright, 1889 (in Canis fam. at Utrecht). syn. of D. felineus= Opisthorchis felineus. See Braun, 1893f, 424; 1903, 3 ed., 157.—Rail., 1893a, 361.—Stiles & Hass., 1894e.—Ward, 1895, 243; 1903, 869.—See also Dies., 1850a, 404 (? syn. of Amphist. truncatum); 1858e, 332.

lanceolum Gronkowski, 1902a, 519 (12) (for lanceolatum).

lanceulatum Baldi, 1900a, 222-224 (for lanceolatum).

lasium Leidy, 1891a, 415-416 (in Ilvanassa obsoleta; Beach Haven, N. J.).

laticolle Rud., 1819a, 117, 421 (in Caranx trachurus: Naples, Arimini). to (Echinost.).—Braun, 1892a, 575, 576.—Carus, 1884, 126.—Crep., 1837, 311.—Dies., 1850a, 386.—Duj., 1845a, 432.—Kroyer, 1838—40a, 597 (in Caranx trachurus).—Looss, 1899b, 580.—Mont., 1890, 422; 1893, 191, pl. 1. fig. 17.— Stoss., 1886, 35.

laticolle Mueh., 1896, 590 (in Anas glacialis; East Prussia), not Rud., 1819, see

platyurum.

laureatum Zed., 1800a, 164, 192-194 (in Salmo trutta; Europe); 1803a, 219.reatum Zed.. 1800a. 164, 192–194 (in Salmo trutta; Europe); 1803a. 219.—Blainv., 1824a, 518, type of "lobostome."—Braun, 1892a, 735, 736, 784, 785, 786; 1893b. 184; 1900, 231, 232; 1900, 389; 1901b. 31.—Dies.. 1850a. 380 (includes Fasc. farionis Mueller; F. truttæ Frælich).—Duj., 1845a. 435, to (Crossodera).—Kroyer, 1838–40a, 615; 1843–45a, 644; 1846–53a. 54 (in Thymallus vulgaris Cuv.; Salmotrutta Linn.; S. fario Linn.).—Linst.. 1903, 279.—Mont.. 1893, 85, 86, 94.—Nord., 1840, 621 (to Fasc.).—Olfers, 1816, 46.—Olss., 1876, 24; 1893, 12.—Risso, 1826, 262.—Rud., 1809a. 413–414; 1814a. 102; 1819a, 113, 413.—Sieb., 1850, 645.—Stoss.. 1886. 18 (in Coregonus oxyrrhynchus, Salmo salvelinus, Thymallus vulgaris, Trutta fario, T. trutta).—Wagener, 1860, 165.—Also reported for Coregonus lavaretus, Salmo fario. 1860, 165.—Also reported for Coregonus lavaretus, Salmo fario.

[laysani Sluiter, 1900, 9 (a tunicate).]

[leachii Stevens, 1827 (a beetle), Distomus.]

leidyi Cobbold, 1860a, 10 (Clinost. dubium renamed).—Stoss., 1892, 181.

lentis (Gescheidt, 1833) Dolley, 1894a, 989.

leptosomum Crep., 1829, 57-58 (in Tringa variabilis); 1831, 177; 1837, 311, 316, 318; 1846, 136.—Braun, 1892a, 650; 1893b, 183 (in Synapta inhærens).—Cobbold, 1879b, 438.—Dies., 1850a, 384.—Duj., 1845a, 428.—Mehlis, 1831, column 177.—Mont., 1888a, 14; 1893, 95.—Stoss., 1892, 169 (to Echinost.).—Villagoria (1998). Villot, 1875, 475, 476; 1878, 24-26, 33, pl. 5, figs. 4-6 (in Tringa variabilis, Calidris arenaria).

leptosomum Roewer, 1906, 185, 186, for leptostomum.

leptostomum Olss., 1876, 18–19, pl. 3, figs. 38–40 (in Meles taxus).—Braun, 1892a, 699, 722, 723, 734, 735; 1899g, 492 (to Harmost.); 1900h, 5, 6, 11, 12; 1901e, 338, type of Harmost., 341.—Hofmann, 1899a, 174–204, pls. 11–12 (development from Cercariæum helicis).—Looss, 1894a, 2, 120, 125, 136, 140, 168, 169, 170, 171, 173, 179, 184, 197, pl. 3, fig. 50, pl. 6, fig. 113; pl. 7, fig. 133 (in Erinaceus europæus); 1899b, 556, 652, 746 (type of Heterolope).—Roewer, 1906, 185, 186 (leptosomum).—Stoss., 1892, 17 (in Erin. europ., Me. tax.) (includes D. caudatum Linst.), to (Brachylaimus); 1898, 24, syn. of Mesogonimus lingusoformis nimus linguæformis.

leucochloridii Leuck., 1858a, 115, see Leucochloridium paradoxum.

ligula Ben., 1870, 1871a, 17 (in Scymnodon ringens; Portugal).—Linst., 1903, 354. lima Rud., 1809a, 37, 408, 427–429 (in Vespertilio auritus, V. murinus; Europe) (includes Fasc. vespertilionis Mueller, 1788; ? Plan. vesp. Gœze, 1782; Fasc. picta Rud., 1802; D. vesp. Zed., 1803); 1819a. 117, 119.—Ben., 1873, 25.—Brand., 1888, 249, 250.—Braun, 1892a. 663; 1900, 221, 223, 229, 234, 236; 1900, 388, 389, 390.—Cobbold, 1860a, 7; 1879b, 294.—Crep., 1829, 70-72; 1837, 311, 322, 326.—Dies., 1836, 240; 1850a, 349, 387.—Duj., 1845a, 437-438.—Kolenati, 1857, 11, 12.—Lamouroux, 1824, 563 (Distome lime).—Linst., 1885, 248; 1887, 103, —Locs, 1890b, 500; 1007, Mar. 5, 482, 487 Linst., 1885, 248; 1887, 103.—Looss, 1899b, 590; 1907, Mar. 5, 483 (of Ben., 1872, 25, figs. 1-6, 18, possibly belongs to Parabascus).—Luehe, 1899, 530,

532 (type of Plagiorchis); 1901n, 487.—Mehlis, 1831, 185-186.—Mueh., 1898,

MA—Continued.
29.—Nord., 1840, 616.—Olfers, 1816, 46.—Sieb., 1835, 56, 64, 65.—Staff., 1905, Apr. 11, 693 (syn. of Plagiorchis vespertilionis (O. F. Mueller)).—Stoss., 1892, 14 (in Plecotus auritus; Rhinolophus ferrum-equinum in Vienna and Belgium; R. hippocrepis, Belgium; Vespertilio dasycneme, Belgium; V. daubentonii, Belgium; V. murinus, Greifswald, Maestricht; V. nattereri, Maestricht; V. emarginatus, Belgium; V. mystacinus, Maestricht; Vesperugo discolor, Vienna; V. noctula, Belgium; Vesperus serotinus; Nannugo pipistrellus, Lauvain; Molossus rufus, Brasil; M. nassatus); 1904, 2.—Also reported for Vespertilio discolor, V. ferrum-equinum, V. nattereri, V. noctula, ported for Vespertilio discolor, V. ferrum-equinum, V. nattereri. V. noctula, Vesperugo pipistrellus.

limacis Dies., 1850a, 302 (to Heterost.; Rhedoni), based on Duj., 1845a, 472, 473.

limatulum Braun, 1900f, 389-390 (in Molossus sp., from Brazil); 1900b, 233-234, pl. 10, fig. 12.—Looss, 1907, Mar. 5, 488 (probably belongs to Parabascus).

limnææ ovatæ Linst., 1885, 251, pl. 15, fig. 28.

limnæi Dies., 1850a, 302 (to Heterost.), based on Duj., 1845a, 473 (in Lymnæus palustris; Rhedoni).

limnophili Linst., 1879, 185–186, pl. 12, figs. 32–33 (larva form in larva of Limnophilus (?) rhombicus; apparently Germany); 1887, 100.—Stoss., 1889, 66,

syn. of D. endolobum.

 lineare (Rud., 1793), Zed., 1803a, 219.—Anacker, 1887c, 513.—Baillet, 1866b,
 105.—Braun, 1892a, 874; 1901, 561.—Caruccio, 1886, 293.—Cobbold, 1860a, 32 (to Crossodera); 1879b, 440.—Dies., 1850a, 379–380.—Duj., 1845a, 444.—Hahn & Lefèvre. 1884a, 516.—Hass.. 1896a, 2 (to Crossodera).—Landois, 1882, 23.—Magalhāes, 1899, 258.—Olfers, 1816, 46.—Rail., 1893a, 368.—Rud., 1809a, 50, 414–415; 1819a, 113–114, 414–415, 685.—Schneidemuehl, 1896, 303.—Sieb., 1835, 52.—Stoss., 1892, 146 (to Crossodera).—Verrill, 1870, 179.—Reported for Anas anser, A. boschas dom.

lineola Dies., 1850a, 346 (in Falco rufus) (includes D. falconis rufi Rud., 1819a, 119; C. E. V.).—Braun, 1901, 561, 565.—Cobbold, 1860a, 12.—Stoss.,

1892, 183.—Also reported for Circus rufus.

lingua Crep., 1825a, 47-48 (in Larus marinus v. maximus; Dec., apparently Europe); 1837, 310; 1846, 139.—Braun, 1892a, 569, 699, 721.—Cobbold, 1860a, 11.—Dies., 1850a, 343 (L. argentatus).—Duj., 1845a, 448.—Jacoby, 1900, 23.—Jægers., 1899a, 16 pp., 1 pl., figs. 1-4; 1901, 982; 1903a, 1, 5.—Kowal., 1896d, 252 (2) (in Dominicanus marinus: Livon.).—Looss, 1899b, 586 (type of Tocotrema).—Luehe, 1899, 539.—Mont., 1893, 94.—Mueh., 1898, 21-22, 29, 94-96, fig. 16 (in Larus ridibundus; see D. mühlingi; Pillau, Mar.).—Olss., 1876, 15; 1893, 11.—Stoss., 1892, 158; 1896, 129; 1898, 41-42.

linguæforme Dies., 1850a, 335 (Brachylæmus erinacei Blan. renamed) (in Erinaceus europæus; Paris).—Cobbold, 1879b, 295.—Stoss., 1892, 35 (syn. of D. blanchardii) (in Erin. europ.); 1898, 24, to Mesogonimus.

linguatula Rud., 1819a, 100 (host not known; Brazil, Aug.), 383 (Rana n. sp.; Brazil), 679 (R. musica; Brazil).—Cobbold, 1860a, 18.—Dies., 1850a, 353 (in Cystignathus pachypus; Docydophryna agua; Ceratophris varia; Brazil).—Duj., 1845a, 454.—Par., 1896, 11—12 to (Brachylaimus).—Stoss., 1889, 70 (in Cystignathy collected). Cystignathus ocellatus).—Reported also for Bufo agua.

linstowi Mont., 1893, 102, for linstowii.

linstowii Stoss., 1890, 42–43, pl. 16, figs. 67–69 (Monost. aculeatum Linst., from Testudo græca, renamed); 1895, 224–225; 1898, 43; 1904, 5.—Braun, 1891d, 1881, 1891, 1891, 1891, 1892, 1892, 1893, 1893, 1893, 1894, 1894, 1891,

lobotes MacCallum, 1895a, 406—410, figs. 5–7 (in Anguilla chrysopa, Perca flavescens, Stegostedion vitreum) [type U. S. Nat. Mus. 6857].—Type of Centrovarium, 1904.

loliginis (Gmelin, 1790) Zed., 1803a, 222–223.—Baird, 1853a, 115 (=Tetraboth-riorhynchus barbatus).—Dies., 1850a, 573 (syn. of Tetrabothriorhynchus migratorius).—Rud., 1809a, 385.

longicauda Rud., 1809a, 372-373 (in Corvus cornix); 1819a, 98, renamed D. macrourum.—Braun, 1899b, 714; 1901, 561, 562, 565; 1902b, 99 (p. p. of Stoss., 1892, 14, and Much., 1896, 248, as syn. of Lyperosomum albicolle), 106-109, to

Lyperosomum, type.—Dies.. 1850a. 345 (syn. of D. macrourum).—Jacoby, 1900, 10. 11.—Looss, 1899b. 634 (to Dicrocœlium, also Lyperosomum).—Mueh.. 1896, 248–252, figs. 2. 9: 1896, 589.—Olfers, 1816, 44.—Volz, 1900, 149.—Wolffhuegel. 1900, 9. 20, 21, 23, 24, 25, 26, 27, 28, 29, 31, 35, 36, 41.—Also reported for Corvus corone, C. frugilegus.

longicolle Crep., 1825a, 57-59 (includes D. embryo Olfers; in Perca cernua, P. vulgaris): 1837, 310.—Ben., 1858a, 1861a, 203.—Braun, 1892a, 784, 785.—Dies., 1850a, 365 in Acerina vulgaris; Berlin (syn. of D. embryo).—Duj., 1845a, 455.—Leblond, 1835c, 103 (in Murcena conger Linn.); 1836f, 4.—Moul., 1856a, 48, 219 (in Perca cernua, P. fluviatilis).—Vaullegeard, 1899, 82.—Wagener, 1857, 26.

longicolle Freelich, 1791 Harz, 1881c, 5, 11.

longicolle Crep., 1846, 154, 155 (in Cottus gobio).—Sieb., 1848, 142 (in Cottus gobio).

longicolle (Abildg., 1788) Cobbold. 1860a, 18-19 (in Tropidonotus naja).

longissimum Linst., 1883, 308-309, pl. 9, fig. 50 (in Ardea stellaris; Turkestan);
1886, 32.—Braun, 1892a, 567, 699, 711, 722; 1893a, 875; 1893, 353; 1893f, 426;
1894i, 602, 605; 1899, 4.—Looss, 1896b, 58.—Mont., 1893, 83, 95, 102.—Stiles & Hass., 1894e, 418; 1896c, 151-155, figs.—Stoss., 1892, 161 (Botaurus stellaris);
1904, 12.

longissimum Poir., 1886, 29–30, pl. 1, fig. 6 in Delphinus tursio .—Braun, 1892a, 699.—Mont., 1893, 41.—Par., 1896, 1–3 /= D. tursionis Marchi).—Stoss., 1892, 25–26 (in Delph. tur.).

longissimum corvinum Stiles & Hass., 1894e. 418 (in Corvus americanus, C. ossi-fragus): 1896c, 151-155, figs.—Looss, 1899b, 564, to Opisthorchis.

longissimum eorvinum Stiles & Hass., 1896c. 155. misprint for corvinum.

longum Leidy, 1851, v. 5, 206, v. 8, 44, 340 (in Esox estor; Cleveland, Ohio).—
 Braun, 1893a, 872.—Cobbold, 1860a, 24; 1879b, 458.—Dies., 1855, 64, footnote 13; 1858, 340.—Stoss., 1886, 52.—Wolf, 1903, 605, fig. 1.—Type of Megadistomum 1904.—Also reported for Esox reticulatus.

lorum Duj., 1845a, 407 (? syn. Monost. ocreatum) (in Talpa europæa; Rennes), to (Brachylaimus).—Braun, 1892a, 567, 579, 597, 642, 662, 663, 671, 699, 700, 705, 723, 737; 1893a, 893; 1900, 13; 1900h, 6; 1901, 897; 1902b, 135, syn. of Ityogonimus ocreatus.—Dies., 1850a, 326 syn. of Monost. ocreatum).—Kuech., 1855, 181.—Looss, 1899b, 653 (type of Dolichosomum); 1900, 608 (type of Dolichodemas); 1901m, 192.—Luche, 1899, 538, syn. of D. ocreatum Zed., which is taken as type of Ityogonimus.—Melnikow, 1865a, 49-55, pl. 3, figs. a-b.—Mont., 1892, 41; 1893, 155.—Stoss., 1892, 17-18 (includes Monost. ocreatum; M. acreatum) (in Talpa europæa, Vienna, Offenbach, Rennes, Denmark).—Velz, 1899, 232.—Will.-Suhm, 1870, 12.

loxia Rud., 1819a, 120 (in Loxia chloris; L. coccothraustes; L. pyrrhula; C. E. V. ..—Duj., 1845a, 443.—Stoss., 1892, 149 (syn. of D. mesostomum).

lucii Rud., 1819a, 122 (in Esox lucius; Greifswald, Jan.) for D. esocis lucii Rud., 1809a, 438.—Dies., 1850a, 399.

lucii Mueller, 1776) Zed., 1800a, xvii, 163, 173-175; 1803a, 213, pl. 3, fig. 3.—
Ben., 1858a, 1861a, 100 (syn. of D. tereticolle).—Dies., 1850a, 358 (luzii, syn. of D. teret.).—Harz, 1881c, 5 (syn. D. teret. Rud.).—Looss, 1894a, 5 (syn. of D. teret.).—Rud., 1809a, 380 (syn. of D. teret.).—Schmalz, 1831, 25.—
Sramek, 1901, 105 (syn. of D. teret.).

lucioperea (Mueller, 1776)
 Zed., 1803a, 216.—Dies., 1850a, 381 (syn. of D. nodulosum)
 —Rud., 1809a, 411, 412 (syn. of D. nod.)
 —Sramek, 1901, 106 (syn. of D. nod.)

D. nod. Zed.).

lucipetum Rud., 1819a, 94, 367 (in Larus glaucus, L. fuscus; Mus. Vien.).—
Braun, 1893a, 877; 1893d, 468; 1897c, 2-3; 1901, 561, 565-566; 1902b, 31 (to Philophthalmus), 32 (includes Bremser, 1824, pl. 9, figs. 1-2; Rud., 1819a, 94, 367; Duj., 1845a, 400; Braun, 1897, 2; 1901, 565; Dies., 1850a, 338; Stoss., 1892, 15, 37).—Cobbold, 1860a, 11.—Crep., 1837, 317.—Dies., 1850a, 338.—Duj., 1845a, 400-401, to (Dicrocœlium).—Looss, 1899b, 701 (to Philophthalmus).—Mol., 1859, 844.—Moul., 1856a, 50.—Nord., 1832a, 17.—Stoss., 1892, 157.

lutea Ben., 1870, 3, pl. 4, fig. 9 (in Scyllium canicula): 1878, 28.—Ariola, 1899, 8 (syn. of D. betencourti).—Braun, 1892a, 700, 705, 720, 721, 736.—Jacoby, 1900, 11.—Linst., 1903, 354 (syn. of D. betencourti).—Looss, 1902, 129.—Mont., 1890, 424.—Stoss., 1886, 52 (in Sc. can., Mugil chelo).

luteum Baer, 1826a, 125 (Paludina vivipara); 1827, 610-611, pl. 29, figs. 20-22.—Braun, 1892a, 632.—Crep., 1837, 310, 325, 326.—Dies., 1850a, 302 (renamed Heterost. ovatum); 1858d, 279, 280 (of La Valette and Wagener, syn. of Cercariæum ovatum) (in Esox lucius) "Wagener: n. sp."—Erc., 1881e, 45, 46, pl. 2, fig. 8; 1882a, 281, 282.—Fil., 1854a, 25.—Fraip., 1880c, 445, 446.—Gamb., 1896a, 62, fig. 30.—Leuck., 1863a, 466, figs. 149, 154, 155; 1879, 10, fig. 1 (of La Valette); 1886d, 6 (of La Valette), fig. 1.—Mont., 1893, 60, 82, 190 (syns. Heterost. ovatum Dies.; Dist. luteum La Valette, 1854, 86, Wagener, 1853, 103, Pag., 1857, 34; Cercariæum ovatum Dies., 1858, 279; Dist. ovatum Cobbold, 1860a, 30), 191 (of Ben., 1870, 3; Stoss., 1886, 52; Mont., 1890, 432, renamed betencourti).—Moul., 1856a, 213, 215 (Heterost. ovatum Dies. as syn.), 216 (in Paludina vivipara), 229, 230.—Mueh., 1898, 11.—Pag., 1854, pl. 3, fig. 15, in Paludina vivipara; 1857, 6, 34, pl. 3, fig. 13 (in Paludina vivipara).—Sons., 1897, 252.—Wagener, 1857, pl. 23, fig. 3 (in liver of Paludina vivipara and in int. of a Hecht, at Nice).

luzii Dies., 1850a, 358, misprint for lucii (Mueller) Zed. (syn. of D. tereticolle). lydix (Stoss., 1896) Looss, 1899b, 581 (to Stephanost.).—Type of Dihemistephanus,

1901.

lymnæi auricularis Fil., 1854, 25–26, 30, pl. 2, fig. 32 (in Lymnæus auricularis); 1854, 280–282, pl. 2, fig. 32; 1855b, 23.—Dies., 1855a, 398 (to Cercariæum).—Wagener, 1857, pl. 36.

lymphaticum Linst., 1903t, 353–354, figs. 3–4 (in Mustelus vulgaris).—Odhn., 1906,

64 (syn. of D. megastomum).

macconnelli Cobbold, 1876, 97, D. sinense renamed.

macrobothrium Ben., 1870, 70, pl. 4, fig. 1 (in Osmerus eperlanus).—Stoss., 1886, 52.
macrocotyle Dies., 1858e, 342 (in Orthagoriscus mola, intestine; Ireland), based on Bellingham, 1844a, 429.—Braun, 1892a, 569; 1893a, 873, 910; 1901b, 27.—Lint., 1898, 522, 523, pl. 45, figs. 8-10, pl. 46, 1-5; 1900, 282; 1901b, 415 (in Mola mola, Myliobatis freminvillei), 421, 434, 466.—Looss, 1894a, 165; 1899b, 631; 1900, 487.—Mont., 1893, 23, 27, 29, 30, 32, 40, 47, 80, 81, 83, 93, 94, 95, 96, 102, 108, 113, 115, 119, 126, 127, 128, 132, 133, 134, 135 (= D. megninii), 136, 137, pl. 1, fig. 7, pl. 3, fig. 27, pl. 4, figs. 43-47; 1903, 10.—Olss., 1868, 24.—Par., 1902, 6 (in Lophius piscatorius, Orthagoriscus mola; Tonnara d'Enfola).—Sons., 1891, 258 (in Lophius piscatorius).—Stoss., 1886, 20; 1887, 185; 1896, 190, to (Podocotyle); 1898, 52.

macrolaimus Linst., 1894b, 334–335, fig. 9 (in Vesperugo pipistrellus).—Braun, 1900, 227–228; 1900, 388.—Looss, 1899b, 618, to Lecithodendrium; 1902m,

774.—Luehe, 1899, 536.

macrophallos Linst., 1875a, 190–192, pl. 2, figs. 12–13 (in Totanus hypoleucos); 1877, 183; 1887, 104 (macrophallus).—Braun, 1892a, 585, 736, 737; 1893a, 838, 839, 849; 1900, 234; 1902b, 50, 155.—Jægers., 1900, 738, to (Levinsenia).—Looss, 1894a, 135; 1899b, 620; 1902m, 704.—Luehe, 1899, 537.—Nicoll, 1906, 524.—Odhn., 1900, 13; 1905, 317.—Stiles & Hass., 1902d, 20.—Stoss., 1892, 147 (in Actitis hypoleucos at Ratzeburg, Totanus fuscus at Hameln).—Ward, 1901, 184.

macrophallus Linst., 1887, 104.

macropoculum Cobbold, 1860a, 25 (in Orthagoriscus mola) (new name for D. macrocotyle Dies.; D. orthagorisci molæ Bellingham).—Mont., 1893, 127.

macroporum Mont., 1893, 133–138, pl. 4, figs. 48–49 (in Lophius piscatorius).—Stoss., 1898, 52 (syn. D. macrocotyle Stoss.) (in L. pisc.; Triest).

macrostomum (Rud., 1803) Rud., 1809a, 382, 383, 386–387, 393 (in Motacilla luscinia); 1819a, 104.—Bavay, 1902a, 200.—Bettend., 1897a, 7, 15, 39; 1897, 311, 319, 343.—Bremser, 1824, 133.—Braun, 1883a, 56; 1891c, 215, 219; 1892a, 593, 597, 598, 603, 607, 608, 638, 640, 641, 642, 675, 676, 677, 682, 684, 685, 687, 693, 700, 701, 703, 712, 713, 718, 736, 773, 775, 777, 778, 779, 780, 784, 785, 798, 801, 803, 806, 807, 808, 814; 1893a, 818, 819, 820, 823, 824, 825, 829, 830, 831, 841, 855, 856, 860, 862, 863, 865, 868, 874, 879; 1893b, 177; 1895b, 134, 135; 1901, 561, 566, 568; 1902b, 42, 43 (coll. Vienna, 469, from Parus major, as syn. of Plagiorchis elegans), 137.—Cobbold, 1860a, 14; 1879b, 436.—Creutzberg, 1890a, 11.—Dies., 1850a, 361 (includes D. erraticum et philomelæ Rud., 1819a, 120; D. ringens Rud., 1819a, 101, 385).—Duj., 1845a, 443.—Gamb., 1896a, 62, 64–67, 72, figs. 32, 33.—Heckert, 1887a, 456ff; 1889a, 66 pp., 4 pls., monograph,

life history, Leucochloridium; 1889b, 357–362; 1890a, 42–43; 1891a, 189; 1892a, 189.—Hoyle, 1890, 540, fig. 4 A. B. (has remarkable larva known as Leucochloridium paradoxum).—Jackson, 1888, 648 (rep. system), 651 (= Leucochloridium Kowal., 1896i, 353 (9), to (Urogonimus); 1896d, (4), 254 (in Turtur aurita; Dublany).—Leuck., 1879, 95; 1886d, 72.—Looss, 1892, 101, 116, 122, 159; 1894a, 151, 179, 181, 202, 206, 236, 240, 252, 262; 1896b, 139, 140; 1899b, 536, 646, 647.—Mont., 1888a, 92; 1892, Oct. 7, 187; 1892, 713; 1893, 157.—Mueh., 1898, 101.—Nicoll. 1906, 521.—Olfers, 1816, 44.—Stiles, 1901, 194.—Stoss., 1892, 183; 1898, 23.—Westhoff, 1890, 337–339.—Wolf, 1903, 617, 618.—Wolffhuegel, 1900, 9, 24, 40, 41, 43, 44.—Also reported for Aegithalus pendulinus, Apternus tridactylus, Budytes flavus, Corvus corone, Dryocopus martius, Garrulus glandarius, Picus major, Succinea putris).

macrostomum Schlotthauber, 1860, 130 (in Petromyzon fluviatilis).

macrourum Rud., 1819a, 98 (in Corvus cornix), D. longicauda, 1809, renamed.—Baird. 1853a, 50, includes D. clathratum Deslongchamps and D. albicolle Rud.—Braun, 1891, 101; 1892a, 764 (macrurum); 1893a, 875; 1901, 561, 562, 565; 1902b, 99 (pp. of Stoss., 1892, 13, syn. of Dicrocœlium albicolle), 106 (syn. of Lyperosomum longicauda), 107 (includes Dies., 1850a, 345; Linst., 1883, 309; 1886, v. 2, v. 34; Rud., 1819a, 98; Stoss., 1892, 13 pp.).—Cobbold, 1860a, 11.—Crep.. 1837, 324; 1846, 132.—Dies., 1850a, 345 (in Corvus cornix, Anthus arboreus, Turdus merula; Rhedoni) (includes D. attenuatum Duj. ?).—Linst., 1883, 309–310; 1886, 34.—Nord., 1832a, 93.—Rail., 1900, 240.—Stoss., 1892, 156.—Also reported for Alauda arborea, Aquila pennata, Cypselus apus.

macrurum Braun, 1892a, 764 (for macrourum).

maculatum Looss, 1901d, 402–404, fig. 3 (in Labrus merula, Crenilabrus pavo, C. griseus; Triest).

maculosum (Rud., 1802) Rud., 1809a, 374–375 (in Hirundo apus; H. rustica; H. urbica) (includes Fasc. hirundinis Freelich, D. hirundinum): 1814a, 101; 1819a, 100, 382–383.—Baird, 1853a, 52.—Braun, 1892a, 642; 1901, 561, 566; 1902b, 37, 46, 47, 48, 49 (includes Dies., 1850a, 349, 412; Olss., 1876, 14; Stoss., 1892, 11) (to Plagiorchis).—Cobbold, 1860a, 13.—Dies., 1850a, 349 (includes D. hirundinum Zed.; Fasc. hirundinis Freelich); 1858d, 260 (larva=Cerc. (Acanthocephala) virgula); 1858e, 337 (adult in Cypselus apus, Hirundo riparia, H. rustica, H. urbica, Caprimulgus europæus; larva in Valvata piscinalis; Paludina impura).—Duj., 1845a, 412–413.—Erc., 1881e, 23; 1882a, 259.—Fil., 1855b, 8.9.—Giebel, 1857, 266.—Harz. 1881c, 4,7.—Kowal., 1896d, (3), 253 (in Hirundo rustica).—Linst., 1901, 195–197, pl. 9, fig. 24.—Looss, 1899b, 557.—Luche, 1899, 532.—Moul., 1856a, 43 (des Hirondelles), 152, 163.—Mont., 1893, 95.—Olfers, 1816, 44.—Olss., 1876, 14.—Pag., 1857, 53.—Par., 1896, 2.—Stoss., 1892, 13.—Walter, 1892, 250.—Wolffhuegel, 1900, 9, 18.—Also reported for Chelidon rustica, C. urbica, Cotyle riparia, Drusus trifidus McLach.

magnum Bassi, 1875b, 497-515, pls. 1-3, figs. 1-10, 1-7, 1-3 [in Hirsch; Mandria].—Braun, 1893a, 875, 877, 882, 910; 1897a, 1468.—Curtice, 1887, Nov., 390-392.—Dinwiddie, 1892, 4.—Erc., 1881e, 90; 1882a, 326.—Francis, 1894, 450.—Gamb., 1896a, 4, 68.—Huber, 1896a, 576.—Leuck., 1982b, 797-799; 1893c, 435-436.—Looss, 1899b, 556.—Par., 1894, 50.—Sons., 1889, 275 (to Fasc.).—Stiles, 1892e, 148; 1892g; 1892m, 464-466; 1894c; 1898a, 49.—Stoss., 1899, 9 (syn. of Cladocœlium giganteum).—Ward, 1895, 253 (to Fasc.), 332 (in Bos taurus), 338 (in Equus caballus); 1903, 866 (to Fasc.).

marculentum Braun, 1901g, 948 (in Emberiza citronella; Vien. Mus. no. 620);

1902b, 148–150, fig. 91.

margaritarum Dubois, 1901, 603–605 (in Mytilus edulis); 1901, 149–150.—Seurat, 1901, 700–702.

marginatum Rud., 1819a, 680 (in Ardea sp.; Brazil, May).—Braun, 1892a, 578, 663, 734; 1899g, 485, 486, 490, to Clinost.; 1900h, 18, 25, 26, 27, 28, 29, 30, 45; 1900, 141; 1901, 561.—Dies., 1850a, 354 (syn. of D. dimorphum).—Duj., 1845a, 446.—Looss, 1899b, 650.—Mont., 1893, 155.—Stoss., 1892, 175 (to Mesogonimus).—Wagener, 1852, 555–557.

marginatum Mol., 1858, 128-129 (in Anas crecca; Batavii); 1861, 198-199, pl. 1,
 figs. 13-14.—Cobbold, 1860a, 10.—Dies., 1858e, 333 (in A. cr.; Patavii).—

Stoss., 1892, 149.

m'connelli McConnell, 1878a, 406, for macconnelli.

medians Olss., 1876, 25–26, pl. 4, figs. 59–63 (in Bufo vulgaris).—Braun, 1892a, 583, 642, 721, 736, 737.—Looss, 1893b, 811; 1894a, 2, 82, 83, 84, 91 (e. p., syn. of D. clavigerum), 92, 95, 96, 99, 105–108, 119, 126, 137, 158, 159, 167, 173, 181, 182, 191, 192, 197, 210, 211, 212, 214, 219, 221, 230, 256, 264, 273, pl. 2, figs. 36–38, pl. 8, figs. 168, 169; pl. 9, figs, 187–188 (in Bufo calamita; B. variabilis; B. vulgaris; Rana esculenta; R. temporaria); 1894, 5, 50; 1896b, 91, 94, 96; 1898, 461; 1899b, 616, 622, 623 (to Pleurogenes).—Luehe, 1900, 558; 1901, 169.—Mont., 1893, 86, 95, 102; 1896, 151, 152.—Mueh., 1898, 23.—Nickerson, 1900, 813, 814.—Staff., 1900, 412, 414; 1902, 724.—Stoss., 1889, 70.

medians Olss. of Staff., 1900, Aug. 30, 412.—Staff., 1905, Apr. 11, 683=D. arcanum

Nickerson, 1900, Oct., to Loxogenes as type.

megacotyle Dies., 1850a, 379 (negacotyle), 659, D. velellæ Fil., renamed (in Velella spirans; Naples) (negacotyle).—Cobbold, 1860a, 30.—Linst., 1904a, 14 (in Stenodus nelma; Siberia).—Mont., 1893, 123.—Moul., 1856a, 217 (in Vel. sp.).
 megalocotyle Mont., 1893, 52, 53, pl. 8, fig. 124.—Linst., 1903, 354 (megatocyle,

misprint).

megaloon Linst., 1879, 337-338 (in Lacerta agilis).—Stoss., 1895, 231 (in L. ag.).

megastomum Rud., 1819a, 102, 387 (in Squalus galeus; Arimini, May).—Ariola, 1899, 7 (includes D. soccus Mol.).—Ben., 1858a, 1861a, 170; 1870, 6, pl. 4, fig. 7; 1870, 7.—Braun, 1892a, 647, 679, 701, 702, 728, 734, 738, 761, 784, 786, 873; 1893b, 176.—Bremser, 1824, 133.—Carus, 1884, 129.—Cobbold, 1860a, 24; 1872b, 91; 1879b, 49, 299.—Crep., 1837, 310, 326.—Crety, 1892, 21–24, fig. 1; 1892c, 399; 1892, (92–97); 1893a, 380–382, 383.—Darr, 1902, 667.—Dies., 1850a, 357 (includes Rud., 1819a, 102, 387; Eisenhardt, 145; Bremser, 1824c, pl. 9, figs. 5–8; Kuhn, 1829, 463, pl. 11, figs. 4–5; Duj., 1845a, 471); 1858, 339 (in Scyllium catulus).—Duj., 1845a, 471.—Eisenhardt, —.—Jacoby, 1899c, 1900, 16–24, figs. 13–16.—Jægers., 1900b, 68–74, figs. 1–4 (copulatory organs).—Kowal., 1895g, 20 (60).—Kroyer, 1852–53a, 851 (in Galeus vulgaris).—Kuhn, 1829, 463.—Looss, 1894a, 128; 1899b, 570 (thinks that this form may belong to Azygia); 1902m, 829, 830 (type of Ptychogonimus).—Linst., 1903, 354.—Luehe, 1900, 490.—Mont., 1888a, 64, 72; 1890, 426; 1892, Oct. 7, 189; 1893, 6, 7, 33, 34, 35, 53, 71, 77, 78, 79, 82, 95, 102, 192, 193, pl. 1, fig. 2, pl. 7, figs. 105–109, pl. 8, figs. 115, 116, 133–135.—Odhn., 1906, 64 (includes D. lymphaticum Linst.).—Orley, 1885, 218.—Stoss., 1886, 23; 1890, 43; 1898, 38.—Villot, 1878, 2, 9, 11, 13 (in Carcharias; Scyllium catulus; Galeus canis).—Wagener, 1860, 167.—Wedl, 155, 383, pl. 2a, fig. 16.—Will.-Suhm, 1870, 5–7; 1871, 179–181, pl. 11, figs. 4–5.—Also reported for Squalus mustelus, Mustelus vulgaris.

megastomum Grobben, 1878a, 89 (in Portunus depurator).

megastomum leporis Kuhn, 1829c; 1830a; 1830b.—Stiles & Hass., 1905a, 941.

megatocyle Linst., 1903, 354, for megacotyle.

megnini Poir., 1885, 4, 5, 14–15, 22, 25, 31, 32, 33, 39, 41, 45, 49, 51, 54, 64, 65, 66, 70, 76, 88, 94, 109, 112, 116, 117, 118, 120, 129, 130, 133, 144, 150, pl. 23, fig. 8, pl. 29, fig. 2, pl. 31, figs. 5–7, pl. 34 (in a fish).—Braun, 1892a, 569, 597, 603, 608, 623, 624, 625, 631, 666, 671, 673, 674, 677, 690, 712, 717, 724, 731, 733.—Buttel-Reepen, 1900a, 590; 1902, 184, 192, 202.—Darr, 1902, 683, 687, 693.—Jackson, 1888, 644.—Lander, 1904a, 7.—Looss, 1894a, 145 (megninii).—Mont., 1893 (megninii), 23, 27, 29, 30, 32, 33, 34, 36, 37, 40, 47, 48, 64, 80, 81, 93, 94, 95, 104, 113, 119, 125, 126, 127, 134, 135 (megninii), 102, 106, 107; 1896, 154 (megninii).—Wolf, 1903, 619, 621.

megninii Mont., 1893, 102, for megnini.

melis (Gœze, 1782) Zed., 1800a, 164, 194–196.—Dies., 1850a, 382 (syn. of D. trigo-nocephalum).—Rud., 1809a, 415 (syn. of D. trig.).

mentolatum Mueh., 1898, 19, for mentulatum.

mentulatum Rud., 1819a, 103, 388–389, 681 (in Coluber natrix, Lacerta agilis, L. maculata; Berlin, Arimini, Apr. and July).—Braun, 1891, 99; 1892a, 766, 780, 784, 785; 1901b, 15.—Cobbold, 1860a, 19.—Dies., 1850a, 355 (includes D. colubri natricis intestinale Rud., D. colubri tessellati Rud.; D. lacertæ Rud.); 1858e, 339 (in Tropidonotus natrix).—Duj., 1845a, 415, to (Brachylaimus).—Erc., 1881e, 73, 74, 75, 76, 77, 78, 79, 82, 89 (in Tropidonotus natrix); 1882a, 309, 310, 311, 312, 313, 314, 315, 318, 325.—Holstein-Beck.—Jackson, 1888, 650 (rep. system).—Kampmann, 1894b, 451, 454, 456, 457, 462, pl. 20, figs. 6-9.—Linst., 1879, 184.—Looss, 1899b (=Lepoderma).—Luehe, 1899,

529 (of Erc., syn. of D. nematoides), 530; 1901, 487.—Mol., 1859, 831, 837–838 (in Lacerta agilis, Berlin; Podarcis merremii, Ariminii; P. muralis, Padua; Tropidonotus natrix, Berlin; T. tesselatus).—Mont., 1893, 187.—Mueh., 1898, v. 1 (1), 10, 19 (mentolatum), 29, 92–93, fig. 13.—Par., 1894, 147, to (Brachylaimus) (in Lacerta merremii; Rimini).—Schauinsland, 1882, 496.—Sons., 1893, 186.—Stoss., 1889, 61; 1895, 217–218; 1898, 33–34; 1904. 2.—Volz, 1899, 235, 236, 238.—Wedl, 1855, 401–409, pl. 2 b, fig. 9.—Reported for Lacerta muralis, L. viridis, L. vivipara, Rana temporaria.

mentulatum Rud., of Erc., according to Luehe, 1899, 529, is D. nematoides.

mergi Rud., 1819a, 121 (in Mergus albellus; C. E. V.).—Dies., 1850a, 391 (renamed D. baculus).

merlangi carbonarii Dies., 1858e, 341, renamed D. anonymum, based on Bellingham 1844a, 428.

merlangi rulgaris Dies., 1858e, 341, renamed D. anonymum, based on Bellingham, 1844a, 428.

merlangorum Dies., 1855, 64 footnote 15, based on Bellingham, 1844a.

meropis Rud., 1819a, 120 (in Merops apiaster; C. E. V.).—Braun, 1901, 568; 1902b,
51, 52, 54 (includes Par., 1896, 5, as syn. of Plagiorchis triangularis).—Dies.,
1850a, 351 (renamed D. triangulare).—Duj., 1845a, 444.—Looss, 1899b, 531,
532, 725.—Par., 1896, 5–7 to (Brachylaimus), fig. 2a.—Stiles, 1901, 172.—
Stoss., 1892, 186.

mesosternum Linst., 1873, 101, see mesostomum.

mesostomum (Rud., 1803) Rud., 1809a, 387–388 (in Turdus iliacus); 1819a, 104.—
Braun, 1901, 561, 566–567; 1902b, 116, 117, 126–129, fig. 76 (includes Dies., 1850a, 361; Stoss., 1890, 5; 1896, 128; 1892, 7; 1898, 33).—Cobbold, 1860a, 14.—Dies., 1850a, 361 (includes D. turdi) (in Turdus iliacus; Greifswald; Pyrrhula vulgaris, Fringilla coccothraustes, C. chloris).—Linst., 1873, 101 (mesosternum).—Much., 1898, 16.—Olfers, 1816, 44.—Stoss., 1890, 43; 1892; 149 (D. loxiæ, 1819); 1896, 128; 1898, 33.—Wolffhuegel, 1900, 9, 45, 46.—Also reported for Coccothraustes vulgaris, Columba livia, Merula vulgaris.

metacus Braun, 1900f, 389 (in Vespertilio lasiopteras; V. noctua; Vien. Mus.);

1901b, 31.

micracanthum Stoss., 1889, 29, pl. 14, fig. 58 (in Pagellus erythrinus; Triest); 1898, 48.—Barbagallo & Drago, 1903, 410, to (Dicroccelium) (in Pagellus erythrinus; Catania).—Braun, 1892a, 583, 720.—Mont., 1893, 86, 95, 102.

microcephalum Baird, 1853a, 58. pl. 2, fig. 2 (in Acanthias vulgaris; Falmouth Harbor); 1853, 22; 1855, 73–74.—Ariola, 1899, 7 (syn. of D. veliporum Crep.).—Braun, 1893a, 873.—Cobbold, 1860a, 24; 1886, 52.—Dies., 1855, 64; 1858e, 335 (in Acan. vulg.).—Linst., 1903, 354 (syn. of D. veliporum).—Lopez, 1888a, 137.—Mont., 1888a (microcephalum); 1889, 132–134; 1889, 322 (=D. veliporum Crep.); 1893, 52.—Stoss., 1886, 52.

microcephalum Crep., 1837, 311; 1849, 64 (in Corvus cornix).

micrococcum Rud., 1819a, 101, 383–384 (in Glareola austriaca; Arimini, April).— Braun, 1901, 561, 564, 567; 1902b, 41, 49, 62 to Phaneropsolus, 63, 64 (thinks identical with Phaneropsolus sigmoides), 155.—Cobbold, 1860a, 13.—Dies., 1850a, 351.—Duj., 1845a, 447.—Linst., 1887, 104.—Looss, 1902m, 823.—Stoss., 1892, 150.

microcotyle Dies., 1858e, 340 (D. pleuronectis maximi Bellingham, 1844a, 428) (in Rhombus maximus; Hibernia).—Cobbold, 1860a, 25.—Stoss., 1886, 52.

micropharyngeum Luehe, 1898g, 623–624, 625 (in flamingo; Berberei); 1899, 537.— Jægers., 1898, 16; 1900, 738.—Looss, 1899b, 619.—Odhn., 1900, 12, 13, 17.

microphylla Ben., 1870, 1871a, 70, pl. 4, fig. 2 (in Osmerus eperlanus; Belgium).—Braun, 1889a, 369.—Linst., 1887.—Stoss., 1886, 52.

microporum Mont., 1889, 322, pl. 33, fig. 1 (in Plagyodus ferox; Madeira); 1891, 520 (Plag. ferox).—Looss, 1899b (to Hemiurus).

microsoma Rud., 1819a, 109, 401 (in Perca marina; Naples, June).—Carus, 1884, 125.—Cobbold, 1860a, 27.—Duj., 1845a, 456.—Dies., 1850a, 370 (microsomum).—Sons., 1890, 141 (in Serranus scriba).—Stoss., 1886, 52.

microsomum Dies., 1850a, 370 (for microsoma Rudolphi).

microstomum Rud., 1809a, 50, 388–389 (in Pleuronectes solea; Paris, May); 1819a, 104.—Cobbold, 1860a, 26.—Dies., 1850a, 362.—Duj., 1845a, 467.—Kroyer,

1843-45a, 488 (in Solea vulgaris).—Mont., 1893, 193.—Olfers, 1816, 46.— Risso, 1826, 262.—Stoss., 1886, 53.—Wagener, 1860, 187.

miescheri Zschokke, 1890, 764, 765, 767, 781-784, pl. 11, fig. 2 (in Trutta salar) to (Cladoccelium).—Braun. 1892a. 642, 699, 700, 721, 733, 735.—Jacoby, 1900, 11.—Looss, 1894a, 173, 175; 1899b, 627.—Mont., 1893, 95 (miescherii) 153.

miescherii Mont., 1893, 153, for miescheri.

migrans Duj., 1845a, 407–409 (in Sorex araneus; S. leucodon) (=Brachylaima advena renamed, hence type of Brachylaima).—Braun, 1892a, 772; 1893a, 831, 864; 1899g. 492; 1901e, 341, 342.—Cobbold, 1860a, 9; 1879b, 296.— Dies., 1850a, 389.—Erc., 1881e, 64; 1882a, 300.—Gamb., 1896a, 71.—Moul., 1856a, 167.—Stiles & Hass., 1898a, 83, 84 (syn. of Brachylaima advena Duj.).—Stoss., 1892, 19 (migras) (in Crocidura aranea at Rennes; Sorex leucodon).

migras Stoss., 1892, 19, misprint for migrans.

miliaris Brown, 1881a, 329, misprint for militare.

militare (Rud., 1803) Rud., 1809a, 421-422 (in Scolopax arquata); 1819a, 115, tare (Rud., 1803) Rud., 1809a, 421–422 (in Scolopax arquata); 1819a, 115, 418.—Bellingham, 1844a, 426.—Ben., 1858a. 1861a, 84–89, 90, 168, 180, 193, 195, 215, 218, 219, 222, pl. 9, figs. 1–11 (includes Cerc. echinata; C. fallax; C. pacifica).—Braun, 1892a, 585, 865.—Chatin, 1880a, 308 (seminal receptacle).—Cobbold, 1879b, 436 to (Echinost.).—Crep., 1837, 311, 312, 316, 317.—Dies., 1836, 240; 1850a. 384–385; 1858d, 261; 1858e, 347 (in Numenius arquatus).—Duj., 1845a, 429.—Erc., 1881e, 29, 31; 1882a, 265, 267.—Fil., 1854a, 18, 19, 21.—Gamb., 1896a, 72.—Hahn & Lefèvre, 1884a, 516.—Hemprich & Ehrenberg.—Hoyle, 1890, 540.—Kowal., 1896d, 253 (3) (in Limnocryptes gallinula; Dublany) to (Echinost.).—Leuck., 1863a, 456.—Linst., 1873, 106 (larva=Cerc. echinifera) (in Scolopax gallinula).—Mégnin, 1884, 53.—Mont., 1888a, 24.—Moul., 1856a, 190.—Nord., 1832a, 69 (in Numenius arcua-Mont., 1888a, 24.—Moul., 1856a, 190.—Nord., 1832a, 69 (in Numenius arcuatus).—Olfers, 1816, 46.—Sieb., 1835, 57; 1854, 23.—Sons., 1897, 252.—Stoss., 1892, 163 (to Echinost.).—Ward, 1895, 341 (in Canis familiaris) (perhaps D. echinatum).—Also reported for Ascolopax gallinago, A. gallinula, Paludina vivipara.

milvi (Gmelin, 1790) Zed., 1803a, 209-210.—Dies., 1850a, 385 (syn. of D. echinocephalum Rud.).—Rud., 1809a, 429 (includes D. falconis milvi Rud.); 1819a, 115 (renamed D. echinocephalum).

minima Ben., 1870, 67 (in Clupea sprattus).—Mont., 1891, 498, 514 (minimum).

minimum Mont., 1891, 498, 514 (for minima).

minutum Cobbold, 1859d, 364, pl. 63, figs. 4–5 (in Hæmatopus ostralegus); 1860a,
14 (London); 1861, 118.—Braun, 1892a, 579; 1902b, 155.—Mont., 1893,
83.—Mueller, 1897, 18, pl. 3, fig. 1.—Nicoll, 1906, 514 (in Hæm. ostr.).— Stoss., 1892, 184.

mistroides (Mont., 1896 [144 (D. constrictum Leared, 1862 [nec Mehlis, 1842] renamed) Braun, 1899b, 715.

[modestum Sluiter, 1898, (a tunicate).]

moleculum Linst., 1880, 51-52 (in Rallus pygmæus); 1887, 104.—Braun, 1892a, 579, 586; 1902b, 157.—Stoss., 1892, 150.

molini Polonio, 1859, see Par., 1894, 148 (molinii) (in Rana esculenta; Padova). molinii Par., 1894, 148 (for D. molini).

molle (Leidy, 1856), Stiles & Hass., 1894h, 162–163, fig. 2; 1895a, 737–741, pl. 3, figs. 1–3 to (Polyorchis); 1895b, 316; 1895c, 700–701; 1896d, 426.—Braun,

1902b, 23.—Mont., 1896, 165, 166 to (Polyorchis). mollissimum Levin., 1881a, 59-61, pl. 2, fig. 4 (in Cottus scorpius; Egedesminde).— Braun, 1892a, 699, 711; 1893a, 911.—Johnstone, 1907, 185–186, fig. 16 (in Belone vulgaris; Foulney Island).—Odhn., 1905, 356, 357 (syn. of Lecithaster gibbosus (Rud.)), 359 (of Stoss., 1889; syn. of Lecithaster confusus Odhn.).-

Stoss., 1886, 11 (in Cott. scorp.; Greenland); 1889, 23 (in Alausa finta; Triest).—Also reported for Alosa vulgaris.

monas Rud., 1819a, 679 (in Amphisbæna sp.; Brazil).—Cobbold, 1860a, 19 (in Siphonops annulatus; Brazil).—Dies., 1850a, 350.—Duj., 1845a, 453 to (Brachylaimus).—Par., 1896, 12–13, fig. 5 to (Brachylaimus).—Stoss., 1889, 71.

monorchis Stoss., 1890, 40-41(2-3), pl. 15, fig. 62 (in Cantharus orbicularis; Triest).— Braun, 1892a, 579, 581, 698, 720, 721, 728, 729; 1895b, 129.—Jacoby, 1900,

23.—Looss, 1894a, 178, 218; 1899b, 599.—Mont., 1893, 82, 86, 87, 95, 149, 151 to [(Monorchis)].—Stiles & Hass., 1898a, 91, 98 (type of Monorchis).

monticellii Lint., 1898c, 518–520, pl. 44, figs. 2–8 (in Remora remora; Woods Holi, Mass., July, Aug.); 1901, 415, 418, 449, 451, 473, 482 (in Rem. rem.; Paralichthys dentatus, Pomatomus saltatrix); 1905, 327, 328, 334, 354, 360, 363, 364, 371, 373, 374, 376, 381, 386, 388, 393, 396, 399, 405, 411, 413, figs. 154, 155, 158 (in Bairdiella chrysura, Centropristes striatus, Coryphæna equisetis, C. hippurus, Cynoscion nebulosus, Lagodon rhomboides, Leiostomus xanthurus, Menidia menidia, Menticirrhus americanus, Micropogon undulatus, Paralichthys albiguttus, P. dentatus, Pomatomus saltatrix, Prionotus tribulus, Rachycentron canadus, Scomberomorus regalis, Seriola lalandi, Synodus foetens, Trachinotus carolinus).—Johnston, 1901, 337.—Looss, 1899b (to Hemiurus).

mormyri Stoss., 1885, 160, pl. 6, fig. 26 (in Pagellus mormyrus; Triest, Feb.); 1886, 26; 1890, 41; 1898, 36.—Barbagallo & Drago, 1903, 410 to (Brachylaimus) (in Pagellus mormyrus).—Braun, 1892a, 721, 736, 737.—Lint., 1900, 290.—Looss, 1899b, 571 (judging from Stoss,'s description, this form belongs to the

Creadiinæ, perhaps to Creadium).—Mont., 1893, 94.

mormyris Hausmann, 1897b, 28 (for mormyri). motellæ Ben., 1870, 63 (in Motella mustela).

[mucosum v. Dr. (a tunicate).]

mühlingi Jægers., 1899a, 7 (in Larus ridibundus) (lingua of Mueh. renamed as n. sp.; apparently Koeningsberg).—Looss, 1899b, 586 (muehlingi) (pro tem. to Tocotrema).

mülleri Levin., 1881a, 56–58, pl. 2, fig. 3 (in Cottus scorpius, Gadus ovak; Egedesminde) [spelled also müllerii, muellerii].—Braun, 1892a, 643 (muelleri), 673, 720, 721, 735.—Looss, 1896b, 136; 1899b, 643 (type of Progonus).—Mont., 1888a, 34, 37 (mullerii); 1893, 43, 61 (müllerii), 82 (muelleri), 85, 86, 95, 182, 183.—Odhn., 1905, 365, 366 (mülleri to Genarches).—Stoss., 1886, 21, 58, to (Brachylaimus).

mullerii see mülleri.

mulli Stoss., 1883, 114–115 (in mullini; Triest, January); 1886, 39 (in Mullus barbatus; Triest); 1898, 48.—Carus, 1884, 127.

muris Erc., 1882c, 103 [pl. 3, figs. 16–21].—Hoyle, 1890, 538 (refers to Erc.'s experiments with Cerc. armata).

musculi Rud., 1819a, 119 (in Mus musculus; Cat. Vien.).—Braun, 1901e, 340 (of Dies., 1850a, Duj., 1845a) (syn. of D. recurvum Duj., 1845).—Dies., 1850a, 395.—Duj., 1845a, 441.—Stoss., 1892, 35 (in Mus musculus; Vienna).

musculorum Braun, 1893a, 871 [D. musculorum percæ Waldenburg, renamed].

musculorum percæ Waldenburg, 1860, 16 (in Perca fluviatilis).

musculorum suis Duncker, 1896a, 279–282, 8 figs.; 1897a, 197–198; (1881a, 23–25, figs. 1–6, 55, 159–160, 1 fig.; 1881d, 141; 1884a, 39–40, figs. 1–6).—Stiles, 1898a, 28. See Agamodist. suis.

musculum Brett, 1880a, 453; 1881a, 139–142 (in Ovis aries).—[=Sarcosporidia?] mutabile Mol., 1859, 833–834 (in Podarcis muralis; Padua).—Braun, 1893a, 875; 1901, 563.—Hollack, 1902a, 869.—Klein, 1905, 20; 1905, 78.—Looss, 1902m, 818, 819, 823, 839.—Luehe, 1900, 563–566; 1901, 172, 173; 1901, 487.—Odhn., 1902, 41, 42.—Rizzo, 1902, 27 (to Anchitrema).—Sons., 1893, 185, in Podarcis muralis Wagl.—Stoss., 1895, 224.

mystacidis Linst., 1896i, 379, larva (in Mystacides nigra).

naia Duj., 1845a, 387, for naja.

naja Rud., 1819a, 99, 377–378, 589 (in Coluber natrix; Berlin) (includes Fasc. longicollis Abildg., 1788, 34, pl. 151, figs. A 1–2, from Coluber natrix renamed and D. colubri natricis pulmonale).—Braun, 1891, 99; 1892a, 766, 780, 784, 785; 1893a, 876.—Cobbold, 1860a, 18 (D. longicolle).—Crep., 1837, 326.—Dies., 1850a, 348.—Duj., 1845a, 395 to (Dicroccelium).—Erc., 1881e, 79, 80, 81, 82; 1882a, 315, 316, 317, 318.—Giebel, 1857, 266.—Kampmann, 1894b, 454, 457.—Lamouroux, 1824a, 562.—Looss, 1899b, 603–604 (type of Macrodera); 1902, 732 (type of Saphedera).—Luehe, 1899, 533.—Mont., 1893, 187.—Mueh., 1898, 29.—Par., 1894, 147 (in Natrix torquata; Padova).—Rizzo, 1902, 28.—Schauinsland, 1882, 496.—Sons., 1893, 185 (naia, in Tropidonotus natrix).—Stoss., 1895, 221–222.—Volz, 1899, 234, 237.—Also reported for Zamenis viridiflayus.

nanum (Rud., 1802) Rud., 1809a, 376–377; 1819a, 101.—Braun, 1893a, 874; 1901, 561, 567; 1902b, 47, 48, 49 (to Plagiorchis), 155.—Cobbold, 1860a, 12.—Dies., 1850a, 350.—Duj., 1845a, 446.—Linst., 1887, 104.—Olfers, 1816, 44.—Stoss., 1892, 184; 1892, 41; 1904, 2.

nardoi Polonio, 1859, see Par., 1894, 627 (in Lacerta muralis; Padova).

nassæ mutabilis Fil., 1855b, 22.

negacotyle Dies., 1850a, 379, for megacotyle.

neglectum Linst., 1887d, 101–102, pl. 2, fig. 3 (in Rana temporaria).—Braun, 1892a, 699, 721, 736, 737; 1893a, 881.—Kowal., 1894, 3; 1902d (9), 27 (syn. of Pleurogenes claviger Rud.).—Looss, 1894a, 82, 83, 84, 91, 92 (syn. of D. clavigerum); 1896b, 94, 95; 1899b, 617 (=clavigerum=type of Pleurogenes).—Mont., 1893, 83, 86.—Stoss., 1889, 65.

nematoides Mueh., 1898, 18 (in Tropidonotus natrix; East Prussia); 1898, 11, 29, 93–94, fig. 22.—Braun, 1899, 631; 1901b, 13, 15, 16.—Looss, 1899b, 567–568 (identical with D. ercolanii).—Luehe, 1899, 528, 529, 530.—Stoss., 1904, 5.—Volz, 1899, 236, 238.—Also reported for Lacerta agilis.

nephrocephalum Dies., 1850a, 391 (in Turdus saxatilis; M. C. V., spring and summer) (D. turdi renamed).—Stoss., 1892, 174, to Echinost.

neuronaia monroi Maddox, 1867, 97, 98, pl. 8.—Cobbold, 1872b, 92 (neuronaii monroii); 1879b, 52.

neuronaii monroii Cobbold, 1872b, 92 (for neuronaia monroi).

nigrescens Olss., 1876, 19–20, pl. 3, fig. 41 (in Lophius piscatorius).—Braun, 1892a, 587; 1893a, 872, 873.—Stoss., 1886, 53 (in Loph. pisc., Molva vulgaris).—Also reported for Hippoglossus maximus, Lota molva.

nigrofavum Rud., 1819a, 118-119, 425-427 (in Orthagoriscus mola; Naples, July) to (Echinost.).—Bellingham, 1844a, 427.—Ben., 1855c, 2, 520; 1856, 84; 1858a, 1861a, 111.—Bettend., 1897a, 17; 1897, 321.—Brand., 1898a, 209 (17).—Braun, 1892a, 578, 684, 687; 1893a, 878.—Carus, 1884, 126.—Cobbold, 1860a, 29.—Crep., 1837, 310, 311.—Deslongchamps, 1829, 238 (nigrostavum).—Dies., 1850a, 394 (includes Schisturus paradoxus); 1858e, 353 (in Orthag. mola).—Drummond, —, 240.—Duj., 1845a, 469.—Kerbert, 1881a, 547.—Knoch, 1894a, 11, 12.—Kroyer, 1852-53a, 745 (in Orthag, mola).—Lint., 1898, 530-531, pl. 48, figs. 8-11, pl. 49, figs. 1-2; 1900, 282; 1901, 415, 422, 466 (in Mola mola); 1905, 328, 334, 374 (in Coryphæna equisetis).—Looss, 1894a, 145, 165; 1899b, 528, 631; 1902m, 721, 764, 765, 778.—Luehe, 1900, 487.—Mont., 1888a, 47; 1893, 13, 22, 23, 29, 30, 32, 40, 46, 47, 64, 65, 66, 67, 68, 80, 81, 83, 84, 93, 94, 95, 96, 102, 108, 113, 115, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 169, pl. 1, fig. 11, pl. 3, fig. 23, pl. 4, 38-42; 1891, 110; 1899, 103.—Olss., 1868, 25.—Par., 1887, 333-334.—Risso, 1826, 262.—Sons., 1890, 140-141 (in Orthag, mola); 1891, 265.—Stiles, 1901, 195, 196.—Stiles & Hass., 1898a, 92, 93 (?includes Schisturus paradoxus).—Stoss., 1886, 40.—Wagener, 1860, 174.—Ziegler, 1883, 552.

nigrostavum Deslongchamps, 1829, 238; misprint for nigroflavum.

nigrovenosum Bellingham, 1844a, 429.—Braun, 1893a, 873; 1900, 233; 1901b, 37.—Dies., 1858e, 343 (in Tropidonotus natrix; England).—Looss, 1894a, 219; 1902m, 504, 820, 821, 822, 831.—Luehe, 1899, 533, 534, 535 (thinks this is a Lecithodendrium); 1900, 561–562.—Mont., 1893, 24, 32, 33, 34, 40, 41, 42, 43, 82, 83, 84, 85, 88, 89, 91, 95, 98, 102, 105, 106, 107, 113, 116, 117, 185–189, pl. 6, figs. 88–90, pl. 8, figs. 117–120, 132; 1896, 151, 152, 154.—Mueh., 1898, 29.—Odhn., 1902, 42.—Rizzo, 1902, 28 (to Lecithodendrium).—Stoss., 1895, 222–223.—Volz, 1899, 235, 236, 237.—West, 1896, 323.

nigrovenosum natricis torquatæ Dies., 1855, 64, based on Bellingham, 1844.

nigrum Linst., 1883, 307 (in Corvus cornix); 1886, 31.—Stoss., 1892, 185.

nitens Lint., 1898c, 534–535, pl. 51, figs. 5–6, pl. 52, fig. 1 (in Tylosurus caribbæus; Woods Holl, July); 1901, 415, 422 (in Tyl. acus).—Looss, 1899b, 710.

[nitidum Sluiter, 1898, 17 (a tunicate).]

noctulæ Rud., 1819a, 119 (in Vespertilio noctula; C. E. V.).—Dies., 1850a, 349 (syn. of D. chilostomum Mehlis).—Kolenati, 1857, 11.

nodulosum (Frœlich, 1791) Zed., 1800a, 164, 190–192; 1803a, 219.—Baird, 1853a, 56–57.—Ben., 1858a, 1861a, 100, 171, 177, 181, 191, 203.—Braun, 1892a, 575, 584, 587, 642, 654, 699, 747, 768, 772, 780, 784, 785, 786; 1893a, 865, 879, 911; 1900, 231, 232.—Bremser, 1824, 134; 1824, pl. 10, figs. 1–3.—Crep., 1829,

67-69; 1837, 323, 324, 326, 327, 328, 329; 1839, 289.—Dies., 1836, 248; 1850a, 380-381 (syns. Fasc. luciopercæ, F. percæ cernuæ Mueller, F. percina Schrank, F. nodulosa Frælich, D. campanula Duj., D. luciopercæ Zed., Planaria lagena Braun); (includes Crep., 1829a, 289; Mueller, 1836, 233, 238; Sieb., 1836, 217); 1858d. 282; 1858e, 343.—Duj., 1845a, 434-435 to (Crossodera).—Erc., 1881e, 56; 1882a, 292.—Fil., 1854a, 21; 1857c, 14 to (Crossodera).—Fraip., 1880c, 416.—Gamb., 1896a, 72.—Harz, 1881c, 5.—Hausmann, 1897b, 4, 6, 8, 14, 18, 20, 22, 27, 35 (in Perca fluviatilis).—Jackson, 1888, 652 (in Acerina cernua).—Kowal., 1894, 2; 1894, 222.—Kroyer, 1838-40a, 20, 41, 54; 1846-53a, 333; 1852-53a, 1221 (in Acerina vulgaris, Barbus fluviatilis, Esox lucius, Lucioperca sandra, Perca fluviatilis).—Lint., 1892c, 69, 70.—Linst., 1873a, 1-7, pl. 1, figs. 1-11 (development); 1873b, 328-331; 1873c, 230-231; 1873d, 345-347; 1873e, 101.—Looss, 1894a, 2, 33-41, 47, 49, 50, 123, 124, 133, 136, 150, 161, 164, 181, 191, 192, 197, 199, 208, 215, 216, 264, 268, pl. 1, figs. 8-10, pl. 5, figs. 92-94 (syns. Fasc. luciopercæ, F. percæ cernuæ, F. percina, Plan. lagena) (in Acerina cernua, Aspro vulgaris, A. zingel, Esox lucius, Lucioperca sandra, Perca fluviatilis); 1902m, 452.—Mehlis, 1831, 184-185.—Mueh., 1898, 22.—Moul., 1856a, 19, 24, 46, 47, 99.—Nord., 1832a, 88; 1840, 617, 621 (to Fasc.).—Olfers, 1816, 46.—Olss., 1876, 23.—Rud., 1809a, 366, 410-413; 1819a, 113.—Schauinsland, 1882, 496, 497.—Sieb., 1835, 66, 69, 70, 73, 82; 1836, 217, 233, 235, 236, 237, 238, pl. 10, fig. 1; 1850, 645.—Sramek, 1901, 95, 106, 107, fig. 60 (syns. Fasc. luciopercæ Muell., F. percæ cernuæ Muell., D. luciopercæ Zed.) (in Acerina cernua, Perca fluviatilis).—Staff., 1902, 481 (to Bunodera) (includes Fasciola luciopercæ Mueller).—Wagener, 1857, 26, 27, pl. 21, figs. 3-7; 1860, 165.—Wright, 1884, 429,*430.—Reported also for Abramis vimba, Acipenser rubicundus, Barbus communis, Bithynia tentaculata; Cyprinus).

nodulosum partim Dies., 1850a, 380; 1858e, 354 (in Esox lucius) (syn. of D.

(Crossodera) campanula Duj.).

[non coronata Crep., 1837a, 326, as name of a group of distomes.]

notidobiæ Linst., 1896i, 379, a larva in Notidobia ciliaris and Sialis lutaria "aus der Gärte."

obesa Ben., 1870, 33 (Cottus scorpius).

obesum Dies., 1850a, 361–362 (in Leporinus friderici, Salminus brevidens, Xiphostoma cuvieri; Brazil); 1855, 67, pl. 3, figs. 11–13; 1858e, 341.—Braun, 1892a, 579; 1893a, 872, 875.—Cobbold, 1860a, 24; 1879b, 458.—Odhn., 1902, 152.—Stoss., 1886, 53.

oblongum (Cobbold, 1858) Braun, 1891, 99; 1892a, 579, 583, 635, 636, 674, 692; 1893a, 875, 894, 910; 1893, 354; 1900g, 250, to Campula.—Mont., 1893, 44.—Stiles, 1895l, 219–220, pl. 7, figs. 2, 3; 1895m.—Stoss., 1892, 16–17 (includes Campula oblonga, D. campula, D. campanula) (in Phocæna communis,

Platanista gangeticum).

obovatum Mol., 1858, 288–289 (in Chrysophris aurata; Patavii); 1861, 201–202.—Braun, 1892a, 736, 737.—Carus, 1884, 130.—Dies., 1859c, 428.—Lint., 1900, 290.—Looss, 1899b, 571 (thinks this form, judging from the description given by Stoss., 1890, belongs to Creadiinæ possibly to Creadium).—Mont., 1893, 94.—Stoss., 1885, 159; 1886, 27; 1889, 26; 1898, 35–36; 1901, (7) 95 (to Allocreadium).

obtusum Looss, 1896b, 78–81, 83, 85, 87, 88, 89, pl. 6, figs. 53–56 (in cameleon, Alexandria, Egypt); 1899b, 547 to Lecithodendrium.—Luehe, 1899, 534; 1900, 562.—Stiles, 1901, 200.

"ocreatum Zed." i. e. (Goze, 1782) Luche, 1899k, 538 (= Dist. lorum Duj.) type

of Ityogonimus.—Looss, 1900, 607.

ocreatum (Rud., 1802) Rud., 1809a, 387, 397–398 (Clupea harengus; Griefswald) (includes Leuwenhoek, Epist. 97, 47; Fasc. halecis Gmelin, 1790a; Fasc. ocreata Rud., 1802; Dist. halecis Zed., 1803a, 222); 1819a, 107.—Braun, 1892a, 711; 1893a, 853.—Dies., 1850a, 298, 372.—Duj., 1845a, 422–423 to (Apoblema).—Kroyer, 1846–53a, 174 (in Cl. har.).—Looss, 1899b, 641 (—D. carolinæ Stoss.) type of Pronopyge.—Mont., 1891, 496, 497, 499, 500, 501, 502, 506, 512; 1891, 16 (to Apoblema).—Reported also for Clupea alosa, C. pilchardus.

ocreatum Rud. of Mol., 1858, 289 (in Clupea alosa; Padua), 290; 1861, 209–210, pl. 3, fig. 7 (in ——).—Dies., 1859c, 432 (of Mol., p. 289, in Alausa vulg.;

Padua).—Johnstone, 1907, 177–180, fig. 13 (in Conger vulgaris; England).—Linst., 1878a, 269 (syn. of D. ventricosum Rud., in Alosa vulg.).—Mont., 1891, 497, 498, 499, 500, 501, 520.—Sons., 1891, 259 (syn. of Dist. ventricosum Rud.).—Stoss., 1886, 12 (in Ammodytes lancea, A. tobianus [see next entry sub Odhner]; Clupea harengus, Trutta salar).—Also reported for Alosa finta,? A. sardina.

ocreatum Mol., of Olss., 1868, 48, pl. 5, figs. 96-98 (in Ammodytes lancea, A. tobianus; —).—Braun, 1891d, 423.—Lander, 1904a, 1, syn. of Hemiurus crenatus (Rud.).—Looss, 1896b, 125; 1899b, 640, 641 (to Hemiurus).—Luehe, 1901n, 399, 400 (syn. of Hemiurus crenatus Rud.).—Mont., 1891, 501, 505.—Odhn., 1905, 352 (syn. of Brachyphallus crenatus) (fig. 98 from Ammodutes leaves A. & A. & Brachyphallus crenatus) dytes lancea, A. tobianus, syn. of Hem. communis).—Stoss., 1886, 12.

ocreatum Mol., of Lint., 1898c, 514-515, pl. 42, fig. 13 (in Pomatomus saltatrix; Woods Hole); 1900a, 269, 282 to (Apoblema), 288 (in Merluccius bilinearis, Pollachius; Woods Hole); 290, pl. 35, figs. 16-24; 1901b, 415, 418, 472, 474, 475, 478 (in Lopholatilus chamæleonticeps, Merluccius bilinearis, Pollachius virens, Urophycis chuss; Woods Hole).—See Hemiurus lintoni, Pratt.

ocreatum of Stoss., 1888.—Stoss., 1898, 30 (syn. of Apoblema stossichii Mont.) (in Alosa sardina; Triest).—Luehe, 1901n, 398 (of Mont., 1887; Stoss., 1888, 1898) in stomach of Clupea pilchardus at Triest, syn. of Hemiurus stossichii Mont., 1891).—Mont., 1887, 86; 1891, 16, 21; 1891, 514 (syn. of Apoblema stossichii).

ocreatum (miscellaneous).—Ben., 1870, 64.—Dies., 1855a, 399.—Hausmann, 1897b, 4, 6, 20, 22 (in Trutta salar; near Basel).—Linst., 1873, 99.—Mont., 1888a, 8, 56.—Jackson, 1888, 648.—Johnstone, 1901, 337.—Juel, 1889, 7.—Olfers, 1816, 46.—Pag., 1857, 4.—Pratt, 1898, 6 (in Pseudocalanus elongatus at Kiel) det. by Mœbius as ocreatum, by Mont., 1891, 13 as Apoblema append. Rud.; 1898, 354.—Wagener, 1860, 166, 183.—Will.—Suhm., —, 142–143; 1871, 382-383.

octopodis delle Chiaje, 1841, 139.—Carus, 1884, 133.

ocular de Bonis, 1882, 180.—Stiles, 1902s, 29 (see Agamodist. ophthalmobium).

oculare Moquin-Tandon, 1860, 347.—Stiles, 1902s, 29.

oculatum Levin., 1881a, 64, 67, pl. 2, figs. 7–8 (in Cottus scorpius; Egedesminde).—
Braun, 1892a, 579, 693, 728, 729.—Johnstone, 1907, 188.—Looss, 1894a, 204, 218.—Mont., 1888a, 10, 38, 49, 78; 1891, 121; 1893, 41, 70, 82, 88, 95, 96, 105, 107, 108, 109, 160.—Odhn., 1905, 328, 331 (type of Acanthopsolus).—Stoss., 1886, 38 (in Cottus scorpius; Greenland).—Also reported for Clupea harengus.

1830, 38 (In Cottus scorpus, Greenand).—Also reported for Chipea harengus.

oculi humani Gescheidt, 1833, 434–435 (in Homo).—Almon, 1833.—Assenova,
1899, 29.—R. Bl., 1888a, 542, 630–631; 1891, 610.—de Bonis, 1882, 180.—
Braun, 1895b, 145; 1903, 3. ed., 151 (perhaps F. hepatica); 1906, 156.—Cobbold, 1866, 7.—Dies., 1850a, 334 (syn. of D. ophthalmobium).—Dunglison,
1893, 338, 820.—Eiss, 1838, 22–23.—Gamb., 1896a, 63.—Hackley, 1886, 519.—
Ijima, 1889b, 123.—Kuech. & Zuern, 1881, 328–329, pl. 8, fig. 12.—Moniez,
1896, 86, 152–153.—Mont., 1892, 713.—Schneidemuehl, 1896, 302.—Sieb.,
1839, 164.—Stiles, 1898a, 48; 1902s, 27, 29, 32, 33.—Fischer von Waldheim,
1840a, 149.—Wallenstedt, 1847, 7.—Ward, 1895, 328 (in Homo) (see D. ophthalmobium): 1903, 866 (syn. of Agamedist, ophthalmobium, Dies).—Weinland mobium); 1903, 866 (syn. of Agamodist. ophthalmobium Dies.).—Weinland, 1859, 281 to Dicrocœlium.

æsophagi ardeæ nigræ Viborg, 1795, 242.—Baird, 1853a, 51.—Dies., 1850a, 337 (syn. of D. hians).—Rud., 1809a, 359.

okeni Ariola, 1906, 185, for okenii (Dist.).

okenii Kcelliker. 1849c, 55-58, pl. 1, fig. 7 (in Brama raji).—Ariola, 1906, 185 (okeni) (to Köllikeria).—Ben., 1858a, 1861a, 105 (syn. of D. filicolle); 1870c, 137.—Braun, 1892a, 572, 573, 574, 647, 672, 698; 1893a, 878, 894, 912; 1906, 184 (okeni).—Dies., 1850a, 359, includes Monost, filicolle; M. ? molæ); 1859c, 423, 429-430.—Gamb., 1896a, 70, fig. 35.—Goldb., 1855, 16.—Kroyer, 1838-40a, 594; 1852–53a, 745 (in Brama rayi Bl.; Orthagoriscus mola).—Looss, 1892, 81.—Mont., 1893, 137, 150.—Moul., 1856a, 25 (trouvé par Kœlliker, dans l'intestin d'un cyprin).—Sons., 1890, 143.—Tasch., 1879, 608, 613.—Wagener, 1852, 566.

okuli humani of Schneidemuehl, 1896, 302 for oculi humani.

oligoon Linst., 1887d, 103 (in Gallinula chloropus).—Braun, 1892a, 167; 1902b, 16, fig. 11 (includes D. spiculigerum Mueh.).—Mueh., 1898, 97.—Stoss., 1892, 149.

oloris Dies., 1855, 64, based on Bellingham, 1844a, 427.

onycephalum Galli-Valerio, 1898m, 923 (for oxycephalum).

opacum Ward, 1894, v. 15, 173–182, 1 pl. (in Amia calva, Ictalurus punctatus, Perca flavescens, Cambarus propinquus; New Baltimore; Lake St. Clair; August); 1895, 15 Feb., 218–219; (1895), 633–634; 1901, 176–184.—Braun, 1900, 234.—Jægers., 1900, 736, 738, 740.—Looss, 1899b, 621.—Odhn., 1905, 317.—Stiles & Hass., 1902d, 20 (type of Microphallus).

ophthalmobium Dies., 1850a, 334 (in Homo; Dresden) [D. oculi humani Gescheidt renamed]; 1858e, 333.—Aitken, 1866, 804, 839; 1872, 146, 205; 1874, 58.—R. Bl., 1888a, 630.—de Bonis, 1876, 164; 1882, 180.—Braun, 1883a, 64-65; 1893a, 870; 1903, 3. ed., 151 (perhaps F. hepatica); 1906, 156.—Cobbold, 1860a, 6; 1864a, 191-192, fig. 41; 1866, 7; 1876, 211. "D. ophthalmobium or Monost. lentis;" 1879b, 36, fig. 5.—Dav., 1877a, lxxix, 820, 822.—Dunglison, 1893, 338, 820, 1174.—Gamb., 1896a, 63.—Goubert, 1878, 130.—Gunther, 1858, 209.—Harley, 1864a, 62.—Hæk, 1859, 42.—Hoyle, 1890, 538.—Ijima, 1889b, 123.—Jamieson, 1897a, 74.—Kholodk., 1898, 26, 32, 34, pl. 11, fig. 28; 1899a, 152.—Kuech., 1855, 181, 222-223, pl. 4, figs. 13-15.—Leuck., 1863a, 526, 610-613, 633, fig. 205; 1889, 440, fig. 191.—Linst., 1878, 3.—Moniez, 1896, 152-153.—Moquin-Tandon, 1861, 373.—Mosler & Peiper, 1894, 177.—Schneidemuehl, 1896, 302.—Stiles, 1898a, 48; 1902s, 25, 29, 34.—Verrill, 1870, 171.—Vogt, 1878, 10, 13.—Wagner, 1883, 121.—Ward, 1895, 328 (perhaps young D. lanceolatum); 1903, 866 to Agamodist.—Wood & Fitz, 1897, 335.

opisthotrias Luetz, 1895a, 181–188, pl. 2, figs. 1–4 (in Didelphys aurita); 1895b, 189–193; 1896a, 623.—Braun, 1899g, 492 (to Harmost.); 1900h, 12; 1901e, 338–339 (to Harmost.).—Looss, 1899b, 748 (to Heterlope).—Luehe, 1899, 532.—

Par., 1896, 3, 5.

orbiculare Dies., 1850a, 349 (in Cebus trivirgatus; Brazil); 1855, 64, pl. 2, figs. 20–22; 1858e, 335–336.—Braun, 1901e, 311, 312, 313, fig. 14.—Cobbold, 1860a, 7; 1879b, 289.—Stoss., 1892, 36 (in Nyctipithecus trivirgatus; Brazil).

oricola Leidy, 1884a, 47 (in Alligator mississippiensis; Florida); 1891a, 414 (syn. of D. incommodum); 1904a, 180.—Braun, 1893a, 872.—Mont., 1892, 715 (oricula).—Stoss., 1895, 219 oricula (syn. of D. pseudostomum).

oricula Mont., 1892, 715, for oricola.—Stoss., 1895, 219.

ornithorhynchi Johnson, 1901a, Nov. 7, 334–338, pl. 22, figs. 1–4 (in Ornithorhynchus anatinus Shaw).

orthagorisci molæ Dies., 1855, 64; 1858e, 342 (in Orthagoriscus mola; Ireland) (renamed D. macrocotyle).

osculatum Looss, 1901e, 654-656, fig. 11 (in Motella vulgaris).

oratum (Rud., 1803) Rud., 1809a, 50, 357–358, 420, 432; 1819a, 93, 121, 674.—
Anacker, 1887c, 513.—Baillet, 1866b, 105.—Baird, 1853a, 51 (includes Hirudo fasciolaris Mueller, Fasc. ovata Rud.).—R. Bl., 1888a, 643.—Braun, 1892a, 714, 734, 736, 765; 1893a, 876, 877, 879, 893; 1893d, 467, 468; 1901, 12, 13, 14, 15, 16; 1901, 258–259; 1901, 561; 1902b, 68 (to Cephalogonimus by Sons., 1890; type of Prosthogonimus Luehe, 1899, 539; and of Prymnoprion Looss, 1899b, 628), 69, 72 (in Corvus corone) 73, 74, 75, 76, 77, 78, 79, 80, 82, 83, 85.—Caruccio, 1886, 293.—Cobbold, 1860a, 9; 1879b, 440.—Crep., 1837a, 317, 318, 322, 326, 329; 1846, 130, 131, 134–137, 139, 141–143, 146; 1851, 284.—Dav., 1877a, lxxix.—Dies., 1850a, 335–336 (includes: Igeln in Hühnereiern Hanow; Hirudo fasciolaris Mueller, Fásc. ov. Rud., D. anatis domesticæ Rud.); 1858e, 333 (in Fringilla montana, Scolopax gallinago, Ardea grus, Fulica atra).—Duj., 1845a, 394–395.—Giebel, 1857, 265.—Hahn & Lefèvre, 1884a, 516.—Hass., 1896a, 2 (to Cephalogonimus).—Hugi., 1827, 48.—Juel, 1889, 36.—Kowal., 1894, 2; 1894, 221; 1895, 350; 1895, 372–390, pl. 8, fig. 7; 1896d, (7), 257, to (Cephalogonimus) (in Corvus cornix, Gallinago scolopacina; Dublany).—Landois, 1882a, 13; 1882, 23 (in hen's egg).—Linst., 1873, 95, 96, 97, 98, 100, 101, 103.—Lint., 1887, 367–369 (in hen's egg).—Linst., 1873, 95, 96, 97, 98, 100, 101, 103.—Lint., 1887, 367–369 (in hen's egg).—Sinst., 1873, 95, 96, 100.—Morth., 1898, 29–30, 100.—Nord., 1840, 616.—Olfers, 1816, 44.—Sambon, 1900, 219.—Schneidemuehl, 1896, 303.—Sieb., 1835, 64, 65; 1836, 233.—Sons., 1890, 134.—Spencer, 1889, 109 (in hen's egg).—Staff., 1902, 719.—Stoss., 1892, 144 (to Cephalogonimus); 1898, 23.—Tschudi, —, 75.—Verrill, 1870, 179.—Wedl, 1857, 241–242, pl. 1, fig. 1.—Wolffhuegel, 1900, 9, 24, 26, 32.—Also reported for Anas boschas dom., A. clangula, A. clypeata, A.

ferina, A. glacialis, A. marila, Anser cinereus dom., Ascolopax gallinago, Astur nisus, Aegolius brachyotus, Buteo lineatus, Ceryle rudis, Chelidon rustica, Colymbus subcristatus, Corvus americanus, C. corax, C. cornix, C. corone, C. frugilegus, C. glandarius, C. monedula, C. pica, Crax pratensis, Cygnus musicus, Falco buteo, F. nisus, F. subbuteo, Fringilla cœlebs, Fulica atra, Fuligula ferina, F. marila, Gallinula chloropus, Grus cinerea, Hirundo urbica, Lanius minor, Larus canus, Numenius arcuatus, N. arquata, Otis tarda, Parus major, Phasianus gallus, Rallus porzana, Scolopax gallinago, S. rusticola, Strix brachyotus, Turdus viscivorus, Uria grylle, Vanellus cristatus.

ovatum of Matsubara (in chickens; Yedo, Japan).—Braun, 1902b, 81-82 (=Prosthogonimus japonicus).

ovatum of Wedl, 1858, 241, pl. 1, fig. 1 (in Scolopax gallinula, Grus cinerea, Fulica atra) and of Linst., 1873, 96 (in Corvus corone).—Braun, 1902b, 72–73, 77–78 (=Prosthogonimus cuneatus).

oviforme Poir., 1886, 26–27, pl. 2, figs. 7, 8, 9 (in Nycticebus javanicus).—Braun, 1892a, 700, 721, 735, 737; 1901e, 311, 313.—Looss, 1899b, 608, 618, 714, 715 (to Phaneropsolus).—Luehe, 1899, 536 (thinks it is a Lecithodendrium).—Mont., 1888a, 38; 1893, 43, 83, 95, 102, 106, 107.—Stoss., 1892, 12–13 (in Nyct. iov.) Nyct. jav.).

Nyct. Jav.).

ovocaudatum Vulpian, 1859, v. 5, 150–152 (in Rana viridis).—Brand., 1898a, 215 (23).—Braun, 1892a, 603, 765, 774, 775, 784, 786, 787, 798, 803, 805, 808, 810, 814, 815; 1893a, 857, 865, 872, 881; 1893b, 181; 1895b, 11.—Creutzburg, 1890a, 32 pp. (anat. development).—Darr, 1902, 654, 657.—Gamb., 1896a, 72.—Heckert, 1887a, 460.—Kampmann, 1894b, 446.—Leuck., 1863, 490.—Looss, 1885b, 24; 1892a, 74, 101, 123; 1894a, 2, 109–111, 120, 133, 136, 137, 154, 168, 170, 173, 179, 184, 197, 199, 200, 201, 207, 210, 211, 212, 214, 275, pl. 3, fig. 49, pl. 6, fig. 123; 1893b, 813; 1896b, 215, 226, 227; 1899b, 645 (type of Halipegus).—Luehe, 1900, 558.—Mont., 1888, 68, 72.—Mueh., 1898, 29.—Schuberg, 1894, 169.—Sons., 1892, 95; 1893, May 7, 215–217; 1893, 187, 188 (in Rana esculenta, R. temporaria); 1893, 28 Oct., 566; 1893, Apr. 30, 63–64; 1894, 2 May, 172 (Distomomum).—Stoss., 1889, 71.

ovocaudatum of Nickerson, 1898, 261–264.—Staff., 1900, 405, 409–410; 1905, Apr. 11, 687-688 (syn. of Halipegus occidualis).

ovofarctum Odhn., 1902, 153-154 (in gallbladder of Synodontis sp.; Omdurman, Apr.); 1902, 42.

ovum Crep., 1846, 159 (in Planorbis marginatus).

oxycephalikum Schneidemuehl, 1896, 303, for oxycephalum.

oxycephalum Rud., 1819a, 98, 375-376 (in Anas boschas, A. clypeata).—Anacker, 1887c, 513.—Baird, 1853a, 56 (var. of echinatum in Gallus dom.).—Baillet, 1866b, 105.—Bellingham, 1844a, 423.—Braun, 1892a, 595; 1893a, 879.—Cobbold, 1860a, 12; 1879b, 440.—Crep., 1837, 311, 312; 1846, 142, 144, 145.—Dies., 1850a, 345–346 (syns. Fasc. appendiculata boschadis, D. inerme); Dies., 1850a, 345–346 (syns. Fasc. appendiculata boschadis, D. inerme); 1858e, 335 (in Anas clyp.; A. crecca).—Duj., 1845a, 448.—Galli-Valerio, 1898m, 923 (onycephalum) (syn. of Echinost. conoideum).—Giebel, 1857, 265.—Hahn & Lefèvre, 1884a, 516 (syn. Fasc. appendiculata Freeh.).—Hass., 1896a, 2, 3 (syn. of Echinost. echinatum).—Landois, 1882, 23.—Linst., 1873, 105.—Leuck., 1863a, 471.—Loos, 1885b, 18.—Mont., 1888, 40.—Nord., 1832a, 47.—Schneidemuehl, 1896, 303 (oxycephalikum).—Stoss., 1892, 167 (syn. of Echinost. ech.); 1899, 13.—Verrill, 1870, 179.—Reported for Anas boschas; A. boschas dom.; A. boschas fera; A. clypeata; A. crecca; A. ferina; A. marila; A. querquedula; A. tadorna; Anser albifrons; A. cinereus; Fuligula ferina; Mergus merganser).

oxyurum Crep., 1825, 48–49 (in Anas marila; Feb.); 1837a, 316; 1846, 141, 142, 143.—Braun, 1891, 100; 1892a, 570; 1901, 944; 1902b, 11, 15, 23, 26.—Cobbold, 1860a, 12.—Dies., 1850a, 345 (A. marila at Greifswald, A. clangula, A. fuligula, A. glacialis, A. nigra, A. tadorna).—Duj., 1845a, 450.—Linst., 1882, 19.—Poir., 1885, 101 (oxyurus).—Sieb., 1836, 239.—Stoss., 1892, 185.—Reported also for Bucephala clangula; Fuligula cristata; F. marila; Harelda

glacialis).

pachisomum Par., 1894, 155 (for pachysomum).

pachyderma Braun, 1899e, 629 (in Chelone atra =? Thalassochelys caretta): 1901b, 26–27, fig. 23.

pachysoma Eysenhardt, 1829, 144 (in Mugil auratus).—Braun, 1892a, 578, 614, 672, 698, 727, 737; 1893a, 910; 1895b, 129.—Cobbold, 1860a, 27.—Dies., 1850a, 366 (in Mug. aur.).—Looss, 1894a, 178; 1902, 129.—Luehe, 1900, 487.—Mont., 1888a, 12; 1893, 43, 82, 129, 151 to [(Monorchis)].—Par., 1894, 155 (pachisomum).—Sons., 1891, 253, 255.—Stiles & Hass., 1898a, 91.—Stoss., 1886, 15, 47-48 (in Mugil); 1886, 16 (in Mugil auratus, M. cephalus); 1898, 27 to (Podocotyle).— Also reported for Mugil chelo, Mergil auratus.

pacifica Steenstrup, 1842, 74-75, 135, pl. 2, figs. 2-8 (of Cerc. echinata Sieb.).—
Braun, 1881a, 329.—Dies., 1850a. 297 (syn. of Cerc. fallax); 1855a, 390-391 (syn. of Cerc. (Hormost.) echinata) (includes Sieb., 1843, 6, 54-57).—Erc., 1881e, 46, 47, pl. 2, fig. 7; 1882a, 282, 283 (in Paludina vivipara).—Fil., 1854a, 17, 18.—Moul., 1856a, 187, 189, 201, 215, 230.

pagelli Ben., 1870, 1871a, 43, pl. 4, fig. 17 (in Pagellus centrodontus at Louvain, Belgium; Cantharus brama, Belgium).—Jacoby, 1900, 16.—Stoss., 1886, 20 (in Cantharus brama, Pagellus centrodontus).

palæmonis Linst., 1877b, 186 (in Palæmon serratus); 1878a, 315.

paliatum Braun, 1892a, 608 (misprint for palliatum).—Gronkowski, 1902a, 523, 531 (16, 24) (for palliatum).

pallasii Poir., 1885, 13-14, pl. 23, fig. 6 (in Delphinus phocæna).—Braun, 1892a, 569; 1893a, 873.—Buttel-Reépen, 1902, 167, 171, 172, 176, pl. 6, fig. 16.— Darr, 1902, 666, 671, 698.—Linst., 1886, 125.—Stoss., 1892, 27 (in Phocena communis; India).

palleniscum Shipley & Hornell, 1905, 53-54, 55, fig. 11 (in Balistes sp.; Ceylon).

pallens Rud., 1819a, 111, 408, 676 (in Sparus aurata; Naples; August).—Carus, 1884, 130.—Cobbold, 1860a, 26.—Dies., 1850a, 337, 348.—Duj., 1845a, 457–458.—Lint., 1898, 526–527, pl. 47, figs. 8–9; 1901, 415 (in Alutera schæpfii), 420, 464.—Looss, 1902m, 762.—Stoss., 1886, 31; 1887, 93; 1898, 47.

420, 404.—L00ss, 1902iii, 762.—Stoss., 1886, 31, 1887, 95, 1895, 47.

palliatum Looss, 1885a, 390—427, pl. 23, figs. 1–5, 7–14, 30; 1885b, 3–40, 46, pl. 23, figs. 1–5, 7–14, 30 (in gall ducts of Delphinus delphis); 1894a, 234; 1899b, 556, 558 (type of Brachycladium), 560; 1902m, 711, 776, 777.—Bettend., 1897a, 15; 1897, 319.—Biehringer, 1888a, 230, 231, 232, 233, 234.—Braun, 1892a, 569, 591, 593, 594, 597, 602, 607, 608 (paliatum), 618, 629, 640, 641, 662, 666, 671, 673, 674, 675, 677, 682, 688, 699, 701, 704, 711, 712, 713, 717, 719, 726, 727; 1893a, 875, 910; 1893a, 354; 1900g, 250, 251, 252, 253.—Buttel-Reepen, 1902, 185, 202.—Gronkowski, 1902a, 523, 531 (16, 24) (paliatum).—Jackson, 1888, 644.—Juel, 1889, 36, 37.—Kath., 1894a, 142.—Kowal., 1898h, 124 (21).—Lander, 1904a, 16.—Linst., 1886, 125; 1890f, 174, 183.—Mont., 1888a, 26, 36, 52, 54, 58; 1893, 27, 33, 41, 42, 65, 66, 67, 75, 83, 84, 85, 88, 90, 91, 95, 99, 102. 52, 54, 58; 1893, 27, 33, 41, 42, 65, 66, 67, 75, 83, 84, 85, 88, 90, 91, 95, 99, 102, 106, 107, 108.—Poir., 1886, 37.—Schuberg, 1895, 178.—Stiles, 18951, 219, pl. 7, fig. 1; 1901, 203, 204.—Stoss., 1892, 10-11 (to Cladocœlium).—Volz, 1899, 232.—Wolf, 1903, 615, 618.

palmatum Rentsch, 1860, 38 (in Seestichling, Gasterosteus), 41.

paludinæ impuræ Fil., 1854, 279, pl. 2, figs. 28–31 (in Paludina impura); 1854a, 24–25, pl. 2, figs. 28–31; 1855b, 23; 1857c, 9.—Dies., 1855a, 399 (to Cercariæum).—Erc., 1881e, 33, 46; 1882a. 269, 282.—Leuck., 1863a, 500.—Looss, 1894a, 32, 362 (see perlatum).—Moul., 1856a, 96 (to Cercaria).

paludinæ impuræ Baer, v. 13, 2, 655.—Dies., 1855a, 399 (to Cercariæum (tenta-

culorum)).

paludinæ impuræ armatum Fil., 1857c, 207-211, pl. 2, figs. 14-16; 1857c, 9-13.— Dies., 1858d, 281 (to Cercariæum).—Looss, 1894a, 32

paludinæ impuræ inerme Fil., 1857c, 207–211. pl. 2, figs. 16–18; 1857c, 9–13.—Dies., 1858d, 280 (to Cercariœum).—Erc., 1881e, 33, 34; 1882a, 269, 270.—Looss, 1894a, 32.—Sons., 1884, 58; 1897, 252.

[pancerii D. Valle (a tunicate).]

pancreaticum Rail., 1890, 143; 1893a, 360 (in sheep; Japan); 1897, 371-377, 1 fig.— Gomy, 1897a, 371, 372.—Jacoby, 1900, 10, 11.—Janson, 1893c, 261.—Looss, 1907, Feb. 1, 124–127, of Janson, (from Hong-Kong).—Stiles, 1898a, 57.— Ward, 1895, 332 (in Bos taurus).

pancreaticum Katsurada & Saito, 1906, 501-506 (in cattle; Japan).—See Eurytrema. pancreatinum Ward, 1895, 335 (in Ovis aries) "probably error for D. lanceolatum." papillatum Rud., 1814a, 105, Fasc. appendiculata Freelich, p. 56, pl. 2, figs. 8-9 (in Anas boschas fer.; Germany), renamed.

papilliferum Mol., 1859, 290 (in Belone acus; Batavii); 1861, 213.—Braun, 1892a, 567, 576; 1900, 231.—Carus, 1884, 128.—Dies., 1859c, 433.—Stoss., 1886, 19.

[papillosa Crep., 1837a, 326, as name of a group of distomes.]

papillosum Dies., 1850a, 381 (in Beroë rufescens; Triest) (= D. beroës Will, 1844, 343, pl. 10, figs. 10–13, renamed).—Mont., 1888, 198, 199; 1893, 123.

paronæ Mont., 1893i, 43, 191, pl. 1, fig. — (in Seriola dumerilii).—Par., 1896 (in Lichia amia).

[parva Sluiter, 1900, 6, a tunicate.]

patellare Sturges, 1897, Sept., 57-69, figs. 1-5 (in Triturus (Molge) pyrrhogaster Boie).—Braun, 1899b, 721; 1899g, 492 (to Phyllodist.); 1901b, 9, 10, 12.— Looss, 1899b (to Spathidium).—Odhn., 1902, 64, 65.

pectinatum Lint., 1905, 327, 334, 366, 389, figs. 200–203 (in Trachinotus carolinus,

Bairdiella chrysura; Beaufort, N. C.).

pedicellatum Stoss., 1887, 184–185, pl. 12, fig. 52 (in Chrysophrys aurata; Triest) (according to Stoss., 1898, this form belongs in Podocotyle).—Braun, 1892a, 578, 720; 1893a, 874, 910.—Looss, 1899b, 571 (thinks it belongs to the Creadiinæ possibly to Creadium).—Luehe, 1900u, 487 (pedicillatum).—Mont., 1888a, 12; 1893, 86, 94, 105, 107.

pedicillatum Luehe, 1900u, 487 (for pedicellatum).

pedocotyle Leidy, 1890, 282 (in Mola rotunda; Beach Haven, N. J.).; 1904a, 231.— Mont., 1893, 135.

pelagiæ Kælliker, 1849, 53-55, pl. 2, figs. 5-6 (in Pelagia noctiluca; Naples) to (Accaccelium).—Ben., 1858a, 1861a, 178.—Braun, 1892a, 673, 682; 1893a, 852.—Buttel-Reepen, 1902, 202.—Crep., 1849a, 78.—Dies., 1850a, 395.—Mont., 1888, 198, 199; 1893, 29, 30, 40, 47, 64, 122, 123, 124, 126, 135, 136.—Moul., 1856a, 217 (in Argonauta argo; Pel. noct.).—Pag., 1862, 298 (in Arg. argo).

pelagicum Staff., 1900, 399–401, fig. 1 (free form) at Passamaquoddy Bay, St. Andrews, New Brunswick; 1902, 481, to Hemiurus (Apoblema).

pellucidum Linst., 1873, 95-103, pl. 5, figs. 5-6 (in Gallus domesticus).—Anacker, 1887c, 513.—Braun, 1892a, 678, 711, 725, 728, 734, 736; 1893a, 873, 893; 1901, 13; 1902b, 68, 72, 79 (Prosthogonimus).—Hass., 1896a, 2 (to Cephalogonimus).— Juel, 1889, 39.—Landois, 1882, 23.—Looss, 1899b, 539 (=Prosthogonimus), 628 (to Prymnoprion), 721.—Luche, 1899, 539.—Mont., 1893, 157.—Neumann, 1892, 374 (to Mesogonimus).—Rail., 1890, 138 (to Cephalogonimus); 1893a, 369.—Schneidemuchl, 1896, 303.—Staff., 1902, 719.—Stoss., 1892, 157.

pelophylacis esculenti Wedl, 1849, 197.—Braun, 1893a, 870.—Dies., 1858e, 355 (in

Pelophylax esculentus).—Stoss., 1889, 71 (in Rana esculenta).

peregrinum Braun, 1900f, 389 (in Rhinolophus ferrum-equinum); 1900b, type of Mesotretes.

pericardium Crep., 1849a, 78, D. helicis Leidy, renamed.—Dies., 1858d, 278

(syn. of Cercariæum vagans).—Leidy, 1857, 44 (=D. vagans).

perlatum Nord., 1832, 88–98, 101, pl. 9, figs. 1–9 (in Cyprinus tinca); 1840, 617, 620 (syn. of Fasc. eriocis).—Ben., 1858a, 1861a, 193.—Braun, 1892a, 747, 768, 780, 784; 1893a, 856; 1893b, 183.—Cobbold, 1860a, 29.—Crep., 1837, 311, 313, 314, 316, 317,318, 321,322, 323, 324, 328, 329.—Dies., 1850a, 394-395 (includes Fasc. tincæ; 317,318,321,322,323,324,328,329.—Dies., 1850a, 394–395 (includes Fasc. tincæ; D. globiporum tincæ); 1858d, 280 (larva=Cercariæum paludinæ impuræ inerme).—Duj., 1845a, 401 to (Podocotyle).—Erc., 1881e, 72; 1882a, 308.—Fil., 1857c, 12, 13.—Harz, 1881c, 5.—Hausmann, 1896a, 390, 391; 1897b, 4, 6, 18, 20, 22, 29, 30, 31, 32, 33 (in Tinca vulgaris; Barbus fluviatilis).—Klein, 1905, 15; 1905, 73.—Kowal., 1894, 2; 1894, 222.—Kroyer 1846–53a, 367 (in Tinca vulgaris).—Linst., 1873, 1 (larva=Cercariæum paludinæ impuræ inerme).—Looss, 1893b, 815; 1894a, 2, 24–33, 42, 48, 86, 113, 122, 123, 124, 125, 126, 127, 135, 136, 137, 140, 148, 150, 157, 158, 166, 167, 173, 177, 178, 191, 192, 195, 207, 208, 211, 212, 213, 215, 218, 231, 232, 239, 249, 257, 262, 264, 266, 269, 270, 273, 274, 276, pl. 1, 16es, 4–7, pl. 4, figs. 81–91, pl. 6, fig., 122, pl. 9, fig., 190 (in 274, 276, pl. 1, figs. 4–7, pl. 4, figs. 81–91, pl. 6, fig. 122, pl. 9, fig. 190 (in Abramis brama; Tinca yulgaris) (includes Fasc. tincæ, 1790; D. ferruginosum, 1877); 1896, 83; 1899b, 598 (type of Asymphylodora); 1902m, 770, 771.—Luche, 1900, 487.—Moul., 1856a, 50.—Much., 1898, 25, 91, fig. 8a.— Odhn., 1905, 322.—Sieb., 1835, 65, 66, 82.—Sons., 1897, 252.—Stiles & Hass., 1898a, 92, 93.—Stoss., 1886, 41; 1886, 62 to (Echinost.).—Wagener, 1857, 44.

perlatum var. exspinosum Hausmann, 1896a, 390-391 (in Barbus fluviatilis; Switzerland); 1897b, 29–34.

permixtus (Braun, 1901) Stoss., 1904, 2.

 perniciosum Taylor, 1884, 53.—Cares, 1888a, 35ff.—Corlette, 1897a, 146.—Dunglison, 1893, 338.—Jamieson, 1897b, 147 (syn. of D. sinense).—Laspeyres, 1904a, 12.—Lockwood, 1901.
 2. ed., 821.—Tyson, 1903, 3. ed., 1180 (syn. of D. sinense). nense).-See Clonorchis.

personatum Poir., 1885, 11-12, pl. 23, fig. 5, pl. 26, fig. 4, host unknown, Gulf of Mexico.—Buttel-Reepen, 1902, 167, 172, pl. 1, fig. 15.—Darr, 1902, 666, 671.—

Lænnberg, 1891, 71.

petalosum Lander in Looss. 1902m, 454 (probably identical with "D. auriculatum Wedl of Linton, 1897, 521." from Acipenser rubicundus and probably belongs to Bunodera).—See also lintoni.

phasiani galli Dies., 1855, 64 (D. dimorphum Wagener).—Braun. 1892a, 735; 1893a, 874.

philocholum Crep., 1845, 330 (in Delphinus phocæna, liver).

philodryadum West, 1896, 28 Feb., 322-324, pl. 11 (in Philodryas schottii; England).—Johnston, 1901, 337.—Luehe, 1900ee, 743.

philomelæ Rud., 1819a. 120 (in Motacilla philomela; C. E. V.).—Dies., 1850a, 361 (syn. of D. macrostomum).—Duj., 1845a, 443.—Stoss., 1892, 183 (syn. of D. macrost.); 1898, 23.

phoxini Linst., 1896i, 378-379, fig. 9, a larva in Phoxinus lævis.

phryganeæ Linst., 1877b, 185 (in Phryganea grandis); 1878a, 296 (in Ph. gr.); 1887,

physæ fontinalis Dies., 1855a, 400 (to Cercariæum) based on Baer, 1827, 656 (in Physa fontinalis).

physophoræ Kælliker, 1849c, 53, based on Fil., 1843, 66, pl. 5, fig. 11 (in Physophora tetrasticha; Naples).—Dies., 1850a, 374 (syn. of D. geniculatum).—Mont., 1893, 124.

pictum Crep., 1837, 313, 316 (in Ciconia alba); 1846, 135.—Ben., 1868, 298.-Braun, 1893a, 874; 1901, 260; 1901, 896; 1902b, 86, 87 88, fig. 49 (includes D. singulare Mol., 1858; Stomylus Looss, 1899b, 629; Stomylotrema Looss, 1900, 602) (to Stomylotrema).—Dies., 1850a, 397.—Stoss., 1892, 185.

pinnarum Wagener, 1857, 26 (in Gasterosteus).

pinnatum Will.-Suhm. (1873), 341.—Thomas, 1882, 447.

piriforme Crep., 1846, 142 (in Anas fusca, A. glacialis), (see D. pyriforme Crep., 1837).—Braun, 1901, 259–260.

pittacium Braun. 1901, 947-948 (in Strepsilas interpres); 1902b, 146-147.—Nicoll, 1906, 521, 522.

planicolle Rud., 1819a, 682, 686-687 (in Pelecanus sula; Brazil) to (Echinost.).-Brand., 1892, 506 to (Echinost.).—Braun. 1901, 561, 567–568; 1902b. 28, 29, 30, 31 (to Anoictost.) (includes Brand., 1892, 506; Duj., 1845a, 430).—Dies., 1850a, 326 (syn. of Monost. echinostomum).—Duj., 1845a, 430–431.—Mont., 1892, 710.

planorbis carinati Fil., 1857c, 13-14, pl. 2, figs. 12-13; 1857 or 1859, 211-212, pl. 2, figs. 12-13.—Dies., 1858d, 282 (to Cercariæum).—Linst., 1873, 7; 1877, 14.— Looss, 1894a, 41.

planorbis cornei Linst., 1877, 187.

platessæ (Mueller, 1784) Zed., 1803a, 216.—Dies., 1850a, 352 (syn. of D. areolatum).—Rud., 1809a, 402 (syn. of D. areolatum).

platyurum Mueh., 1896, 267-270, figs. 6, 13-14 (in Harelda glacialis) [D. laticolle Rud. of Mueh.: East Prussia]: 1897. v. 243–279. pl. 4; 1897, 127–128; 1897, 478–479; 1898. 16. 25. 96, 97. 98.—Braun, 1901b, 33; 1902b, 11, 15, 18.—Looss, 1899b, 573–574 (type of Psilost.).

pleroticum Braun, 1899e, 631 (in host?: Brazil); 1901b, 13, 17.—Luehe, 1899, 529.

 plesiostomum Linst., 1883, 305–306, pl. 10, fig. 48 (in Perdix græca; Turkestan);
 1886, 29.—Braun, 1892a, 578, 700, 711, 734; 1899b, 714; 1901, 946; 1902b,
 109, 112, 119.—Jacoby, 1900, 10, 11.—Looss, 1899b, 634 (to Dicrocœlium).— Stoss., 1892, 152,

- pleuronectis maximi Dies., 1855, 64 based on Bellingham, 1844a; 1858e, 340 (see Bellingham, 428), renamed D. microcotyle.
- [plumbeum D. Valle, (a tunicate).]
- poirieri Stoss., 1895, 227 (D. gelatinosum of Poir., 1885); 1904, 3.—Braun, 1899e, 631 (in Emys europæa); 1899b, 716; 1901b, 13, 15, 16, 17, 19, 30.—Looss, 1899b, 567.—Luehe, 1899, 528, 529.
- polonii Mol., 1859, 291 (in Caranx trachurus; Batavii); 1861, 219.—Braun, 1892a,
 576.—Carus, 1884, 126.—Dies., 1859c, 435.—Mont., 1893, 191.—Olss., 1868,
 29; 1876, 15.—Sons., 1890, 142 (in Caranx trachurus L.).—Stoss., 1886, 38;
 1899, 16.
- polyclinorum Pag., 1862, 306, pl. 29, fig. 4 (in Polyclinum; Golf von Spezia).
- polymorphum Fil., 1837a, 337–338, figs. 8–14 (in Planorbis submarginata); 1855b, 23.—Dies., 1850a, 296 (renamed Cercaria brachyura).—Moul., 1856a, 213 (syn. of Cercaria brachyura).
- polymorphum (Rud., 1802) Rud., 1809a, 363–364 (in Muræna anguilla), includes Fasc. anguillæ 1790; 1814a, 101; 1819a, 95, 369–370.—Braun, 1891d, 421; 1892a, 642, 736, 765.—Carus, 1884, 132.—Cobbold, 1860a, 23 (in Anguilla vulgaris); 1879b, 458 (in Esox lucius).—Dies., 1850a, 340 (includes D. anguillulæ Abildg. ?, D. anguillulæ Zed., Fasc. anguillæ); 1858e, 333 (in Anguilla vulgaris) (includes Dies., 1850a, 340; Wedl, 1855, 383, 394, pl. 2a, 17, 400, 408, pl. 1b, 7; Stein, pl. 7, fig. 23).—Duj., 1845a, 468.—Juel, 1889, 39.—Kroyer, 1846–53a, 641 (in Anguilla migratoria).—Leeuwenhoek, —, 344.—Linst., 1873e, 99.—Nord., 1840, 618 (syn. of Fasc. anguillæ).—Olfers, 1816, 45.—Risso, 1826, 262.—Stoss., 1902, 562.—Wedl, 1855, 383–384, pl. 2a, fig. 17; 400, pl. 1b, fig. 7.
- polyoon Linst., 1887, 103–104 (in Gallinula chloropus).—Braun, 1902b, 155–156, fig. 98.—Dies., 1850a, 340.—Stoss., 1892, 186.
- polyorchis Stoss., 1889, 24–25, fig. 61 (in Corvina nigra; Triest).—Braun, 1892a, 583, 673, 698, 700, 728; 1895b, 129; 1902b, 22.—Buttel-Reepen, 1902, 202.—Lint., 1901, 415, 417, 460, figs. 363, 364, 365 (in Cynoscion regalis); 1905, 328, 334, 366 (in Cyn. reg.), 385.—Looss, 1894a, 178.—Mont., 1893, 82, 84, 85, 86, 95, 148, 149; 1896, 166, to (P.).—Ofenheim, 1900, 160.—Sons., 1890, 136.—Stiles & Hass., 1895a, 737–742; 1895h, 162–163; 1898a, 92, 97 (type of Pleorchis).
- pontaliei Stoss., 1892, 178, for pontalliei (Dist.).
- pontalliei Cobbold, 1860a, 10 (in Ardea minuta) (D. cladocalium Dies., renamed).—Stoss., 1892, 178 (pontaliei) (syn. of D. cladocalium).
- porrectum Braun, 1899b, 714 (in Saurophaga saurophaga; New Guinea); 1902b, 109.—Jacoby, 1900, 10, 11.—Looss, 1899b, 634.
- pristiophori Johnston, 1902, 326–330, pl. 13, figs. 1–7 (in Pristiophorus cirratus Lath.).—Linst., 1903, 354.
- pristis Deslongchamps, in Lamouroux, 1824, 563 (in Marsouin); 1824, 281.—
 Braun, 1892a, 579, 583, 584, 728.—Cobbold, 1860a, 36 (to Echinost.).—Dies., 1850a, 390 (in Merlangus communis; Normandy).—Duj., 1845a, 433.—
 Kroyer, 1838-40a, 606 (in Merlangus vulgaris Cuv.).—Lamouroux, 1824a, 563.—Looss, 1894a, 204, 218; 1899b, 581 (to Stephanost.).—Mont., 1888a, 14; 1893, 86, 88, 105, 107, 108, 109.—Stoss., 1886, 36, 45-46, pl. 8, fig. 33.—
 Vaullegeard, 1901, 143-146, 1 fig.; 1901, v. 8, 640.
- pseudoechinatum Olss., 1876, 21–22, pl. 4, figs. 45–49 (in Larus marinus; Scandinavia).—Braun, 1893a, 874.—Kowal., 1896d, 253 (3) (in Dominicanus marinus), to (Echinost.).—Mont., 1888a, 14; 1893, 86.—Stoss., 1892, 166 (to Echinost.).
- pseudostoma Will.-Suhm, 1870, 11–12, pl. 11, fig. 2 (in Alligator lucius); 1871,
 185–186, pl. 11, fig. 2.—Brand., 1888a, 15.—Mont., 1892, 34; 1892, 715.—
 Pavesi, 1881, 294.—Poir., 1886, 334 (to Diplost.).—Stoss., 1895, 219 (in Alligator missisipiensis; Florida).
- pudens Lint., 1900a, 269, 283, 290–291, pl. 37, figs. 40–47 (in Paralichthys dentatus; Woods Hole, Sept.); 1901, 415, 420, 482; 1905, 328, 334, 372, 413 (in Paralichthys albiguttus, Rachycentron canadus).
- pulchellum Rud., 1819a, 94, 367–368, 595 (in Labrus cynædus; Naples).—Barbagallo & Drago, 1903, 410 (in Gobius jozo; Catania), to (Dicrocœlium).—Carus, 1884, 131.—Cobbold, 1860a, 22.—Dies., 1850a, 338–339.—Duj., 1845a,

462.—Looss, 1901, 399.—Mol., 1859, 844.—Odhn., 1901, 484, 493, 494; 1902, 160-161 (to Helicometra).—Sons., 1891, 257-258 (in Tinca vulgaris).—Stoss., 1886, 54; 1898, 45-46.—Also reported for Blennius ocellaris, Labrus mixtus.

pulcherrimum (Weyenbergh, 1876) Weyenbergh, 1878, v. 3, 31-38 (in Hypostomus plecostomus; Argentina); 1878, 354–361; 1878, 554–561; 1880, 31–39.– Braun, 1893a, 871.—Looss, 1885b, 55.

pulicis Linst., 1892, 333-334, pl. 15, fig. 19 (in Gammarus pulex).—Braun, 1891b, 182.

pulmonale Bælz, 1883a, 236–237, fig. 3 (in Homo).—Anders, 1903, 6 ed., 1245.—Bl., 1888a, 627 (syn. D. ringeri).—Brand., 1888a, 50; 1890a, 577; 1891b, 265.—Braun, 1892a, 664, 758; 1893a, 876, 882; 1893b, 185, 186; 1901, 332; 1903, 3 ed., 155 (syn. of Paragonimus westermani).—Cobbold, 1884g, 976.—de Does, 1903, 409–412.—French, 1896a, 644.—Gamb., 1896, 63, 70.—Hahn & Lefèvre, 1884a, 546 (syn. of D. ringeri Cobb.).—Huber, 1896a, 576–577.—Janson, 1897a, 454–455 (in dogs, cats, swine).—Janson & Tokishige, 1892, 349, 351.—Katsurada, 1899a, 8–29; 1900, 506.—Katsurada & Fijiki, 1899a, 1–18, 8–29 (pathology).—Katsurada & Saki, 1899a, 141–185 (treatment).—Kholodk (pathology).—Katsurada & Saki, 1899a, 141–185 (treatment).—Kholodk., 1898, 26, 31, pl. 11, figs. 18–21; 1899a, 152.—Kurimoto & Ijima, 1892a, 718– 1898, 26, 31, pl. 11, figs. 18–21; 1899a, 152.—Kurimoto & Ijima, 1892a, 718–720.—Leuck., 1889, 404–440, figs. 181–190.—Linst., 1890f, 178 (syn. of D. westermani).—Looss, 1894a, 118, 142, 159, 180, 181, 186, 206; 1899b, 560; 1905, 84 (syn. of Paragonimus westermani); 1905m, 280.—Manson, 1895, 54–55; 1901 (Gibson), 541 (syn. of D. ringeri); 1903, 3 ed., 631 (syn. of Paragonimus westermanni).—Miyake & Matzui, 1894, 1–6, pl. 1.—Moniez, 1896, 144.—Mont., 1888, 54; 1893, 13, 33, 35, 38, 39, 83, 87, 95, 102, 105, 106, 107, 155, 157.—Mosler & Peiper, 1894, 178–179.—Otani, 1888.—Rail., 1890, 143.—Remy, 1883, 525–527, fig. 2.—Rev. sci., 1890, 189.—Schneidemuehl, 1896, 301–302.—Shaw, 1901, 600.—Simon, 1896, 225; 1897, 209, 224, 259.—Stiles & Hass., 1900a, 567.—Stoss., 1892, 32 (to Mesogonimus).—Taniguchi, 1903, Dec., 100, 105.—Taylor, 1884, 44–48, 51–52, fig. 1.—Vierordt, 1898, 158.—Vincent, 1889, 184; 1890. 189–190; 1890, 80.—Ward, 1895, 244 (syn. of D. westermanni), 328 (in Homo), 341 (in Canis familiaris); 1903, 867 (syn. of Paragonimus westermanii).—Weichselbaum, 1898, 315.—Wolf, 1903, 612.—Wood & Fitz, 1897, 336.—Yamada, 1899, (pathology); 1900 (prophylaxis).—Yamagiwa, 1890a, 457 (seu cerebrale); 1891a; 1891b; 1891c. (prophylaxis).—Yamagiwa, 1890a, 457 (seu cerebrale); 1891a; 1891b; 1891c.

pulmonale colubri natricis Viborg, 1795, 243.—Dies., 1850a, 348 (syn. of D. naja).—Rud., 1809a, 434.—Ward, 1895, 244 (syn. of D. westermanni).

pulmonar La Clínica de Málaga, 1883, 308, for pulmonale.

pulmonis Kiyona, Suga & Yamagata, 1881, teste Ijima, 1889b, 148.—Braun, 1903, 3 ed., 155 (syn. of Paragonimus westermani).—Ijima, 1889b, 148.—Inouye, 1903, 120.—Katsurada, 1900d, 506 (syn. of D. westermanni).—Ward, 1895, 328 (in Homo), 341 (in Canis familiaris); 1903, 867 (syn. of Paragonimus westermanii).

pulmonum (Bælz, 1880) Tomono Hidekata, 1883 (autopsy).—Inouye, 1903, 120.—Inouye & Katsurada, 1893, 798; 1897, 995 (in brain).—Looss, 1905, 84 (syn. of Paragonimus westermani).—Riusai, 1884 (treatment).—Tomono, 1883, no. 96.—Yamagiwa, 1890.—Yamagiwa & Inouye, 1890a, 20 Sept., -5 Oct., —; 20 Oct., —; 1890, 5 Nov., 30-40; 20 Nov., 26-38.

pulvinatum Braun, 1899, 630 (in Flussschildkreeten; Brazil); 1901b, 24-26, fig. 18.—Looss, 1902m, 515, 516.—Odhn., 1902, 42; 1902, 153.

punctatum (misprint for D. punctum Zed., 1800a, 184) Dies., 1850a, 329 (syn. of Monost. cochleariforme).—Harz, 1881c, 11.

Bonost. Cochieganorime.—Harz, 1881c, 11.

punctum Zed., 1800a, 164, 183–184 (in Cyprinus barbus; Europe); 1803, 217.—

Bremser, 1824, 134.—Cobbold, 1860a, 28 (in Barbus communis).—Crep., 1837, 326; 1839, 289.—Dies., 1850a, 377 (includes Bremser, 1824c, pl. 9, figs. 21; 22; Duj., 1845a, 463; Rud., 409, 1819a, 112).—Duj., 1845a, 463.—Harz, 1881c, 5, 10, 11 (punctatum), 152, 154.—Hausmann, 1896a, 391; 1897b, 30.—Kroyer, 1846–53a, 333 (in Barbus fluviatilis Ag.).—Nord., 1832a, 88.—Olfers, 1816, 46.—Rud., 1809a, 327, 366, 409–410; 1819a, 112–113.—Stoss., 1886, 54.

pungens Linst., 1894, 333-334, fig. 10 (in Podiceps minor; Seeburger See). pusillum (Braun, 1790) Zed., 1803a, 210.—Braun, 1893a, 870.—Cobbold, 1860a, 7 (in Erinaceus europæus); 1879b, 295.—Crep., 1825, 55–57; 1837, 310, 326; 1841, 78.—Dies., 1850a, 360 (to Fasc.).—Duj., 1845a, 438–439.—Harz, 1881c, 3 (pussilum).—Olfers, 1816, 30, 44.—Rud., 1809a, 163, 384–386; 1819a, 104, 119 (pars, syn. of D. aluconis thoracicum).—Stoss., 1892, 33, (to Agamodist.).

pussilum Harz, 1881c, 3 (for pus'llum).

putorii Mol., 1858, 131 (in Mustela putorius; Patavii); 1861, 224, pl. 5, fig. 4.—
Braun, 1893a, 870 (in Fœtorius putorius).—Cobbold, 1860a, 9.—Dies., 1858e, 354 (in Mustela putorius).—Erc., 1881e, 56; 1882a, 292.—Linst., 1875, 192, pl. 2, fig. 14.—Stoss., 1892, 34 (to Agamodist.).—Also reported for Putorius vulgaris.

putorii (Gmelin, 1790) Rail., 1893a, 366.

pygmæum Levin., 1881a, 73-75, pl. 3, fig. 3 (in Somateria mollissima; Egedesminde).—Braun, 1892a, 578, 583, 586, 642, 720, 721, 736; 1900, 234.—Jægers., 1901, 982 (type of Spelotrema).—Looss, 1899b, 620; 1902m, 704, 705, 706, 784 (in Somateria mollissima).—Luehe, 1899, 537.—Mont., 1893, 43, 61, 83, 95.—Nicoll, 1906, 522, 523, 524.—Odhn., 1900, 13; 1905, 314 (to Spelotrema).—Stiles & Hass., 1902d, 20 (type of Spelotrema).—Stoss., 1892, 147.—Ward, 1901, 184.

pygmæum similis (Jægers., 1900) Looss, 1902m, 784.

pyramidum Looss, 1896b, 76–78, pl. 6, fig. 52 (in Rhinolopus hippocrepis Bonap.; Ghizeh); 1898, 453, 454, 456, 457, 458, figs. 1, 4.—Stiles, 1901, 200.

pyriforme Crep., 1837a, 316 (in Eisente); 1847, 142 (piriforme).—Dies., 1850a, 397 (in Anas fusca, A. glacialis).—Stoss., 1892, 186.

pyriforme Lint., 1900, 269, 279, 292–293, pl. 38, figs. 52–59 (in Palinurichthys perciformis; Woods Hole, Aug.); 1901, 415 (in Cynoscion regalis, Monticirus saxatilis, Pal. percif.), 422, 453, 454, 457, 458, 460, 462, 483; 1905, 328, 334, 352, 360, 380, 382 (in Brevoortia tyrannus, Lagodon rhomboides, Menidia menidia).

pyxidatum Bremser in Rud., 1819a, 678–679 (in Crocodilus sclerops; Brazil).—Cobbold, 1860a, 19.—Dies., 1850a, 358 (in Champsa sclerops; Brazil).—Duj., 1845a, 452.—Leuck., 1863a, 460.—Rud., 1819a, 678–679.—Stoss., 1895, 231–232.—Also reported for Caiman sclerops, Jacare sclerops.

quietum Staff., 1900, 403–406, fig. 4 (in frogs; apparently Canada).—Type of Glypthelmins.

rachion Cobbold, 1858b, 158, pl. 31, figs. 9–10 (in Morrhua ægelfinus); 1860a, 25.—Braun, 1892a, 642.—Lint., 1898, 538–539, pl. 53, figs. 3–7; 1901, 415 (in Gadus callarias), 422, 476.—Mont., 1893, 82, 85, 86.—Nicoll, 1907, 77, 79 (syn. of Lepodora rachiæa).—Odhn., 1905, 332 (type of Lepodora) (rachiæa).—Stoss., 1886, 43 (in Gadus morrhua).

radiatum Duj., 1845a, 427-428 (in Carbo cormoranus or Pelecanus carbo), to (Echinost.), from Vien. Mus.—Dies., 1850a, 383 (syn. of D. echinatum).—Stoss.,

1892, 167 (syn. of Echinost. ech.); 1898, 52.

radula Duj., 1845a, 433–434 (in Lymnæus palustris; Rennes), to (Echinost.).—Dies. 1850a, 395.—Fil., 1854a, 17.—Moul., 1856a, 203.

ragazzi Linst., 1903t, 354, for ragazzii.

ragazzii Setti, 1897, 8–12, pl. 8, figs. 6–7 (in Squalus; Eritrea), to (Polyorchis).—Ariola, 1899, 7.—Linst., 1903, 354 (ragazzi).—Looss, 1899b, 642, 644, 731, 737 (type of Syncœlium).—Par., 1896, 3, nomen nudum (in Squalus sp.; Red Sea).

rajæ Rud., 1809a, 435 (D. rajæ intestinale Viborg).

rajæ intestinale Viborg, 1795, 242.—Rud., 1809a, 435 (D. rajæ).

ralli Rud., 1819a, 120 (in Rallus aquaticus; M. C. V.).—Dies., 1850a, 339 (syn. of D. holostomum).—Duj., 1845a, 447.—Stoss., 1892, 145 (syn. of Cladoccelium holost.).

ramlianum Looss, 1896b, 36–44, pl. 3, figs. 17–19 (in Caméléon; Ramleh); 1898, 461; 1899b, 589–590 (type of Lepoderma).—Luehe, 1899, 531.—Stoss., 1904, 2.

ranæ esculentæ Dies., 1850a, 388 (syn. of D. retusum), based on Valentin, 1841, 54.
 rarum Ben., 1858a, 1861a, 178 (in Cyprinus dobula).—Fraip., 1880c, 417.

rastellus Olss., 1876, 16–17, pl. 3, figs. 31–36 (in Rana temporaria).—Braun, 1892a, 736; 1893a, 881.—Linst., 1887, 97, 98, 101, 102 (syn. of D. endolobum Duj.).—Looss, 1894a, 82, 84, 85 (syn. of D. end. Duj.).—Mont., 1893, 94, 102 (rastellum).—Stoss., 1889, 66.

rathonisi Simon, 1896, 182, 192, misprint for rathouisi.

rathonisii Huber, 1894a, 2 (contents), 1896a, 575 (misprint for rathouisi; syn. of D. buskii).—Simon, 1896, 182, 192; 1897, 223.

rathouisi Poir., 1887, 203-211, pl. 13, figs. 1-7 (in Homo); 1888, 186-187; 1888, 635, 638, 682, 684, 699, 704, 717, 719, 733; 1893a, 875, 882; 1895b, 141–142 (rathouisi), fig. 57 (probably identical with D. buski); 1903, 3 ed., 154, fig. 100 (rathouisi); 1906, 157–160, fig. 91.—Gamb., 1896a, 63, 70 (rathouisi).—Hoyle, 1890, 538.—Ijima, 1889b, 133 (rathouisi).—Kholodk., 1898, 26, 30, 31, pl. 11, fig. 17.—Leuck., 1889, 328-336, figs. 150, 152, 153.—Moniez, 1896, 86, 117-118.-Mont., 1893, 33 (rathousii), 83, 84 (rathousii), 102, 107.—Stoss., 1892, 27–28 (syn. of D. crassum) (in Homo; China; India).—Ward, 1895, 328 (in Homo).

rathouisii Braun, 1892a, 568, for rathouisi.

rathousii Mont., 1893, 33, for rathouisi.

raynerianum Nardo, 1827, 68-69 (in Luvarus imperialis; Venice); 1833, 524.— Carus, 1884, 125.—Cobbold, 1860a, 28; 1879b, 460.—Dies., 1850a, 376.—Mont., 1891, 500, 520.—Stoss., 1886, 15; 1891, 112.—Also reported for Proctostegus prototypus.

receptaculum Cobbold, 1860a, 29 (D. labracis Duj. renamed, in Labrax lupus).—

Odhn., 1901, 514.—Stoss., 1898, 46.

recurvatum Linst., 1873, 101, 105, 106, pl. 5, fig. 1 (in Anas marila).—Braun, 1892a, 569; 1893b. 183.—Kowal.. 1894, 3; 1895, 355–356, figs. 9–11, 12c: 1895g. 1, 15), 41, 55; 1896d, (7), 257 (in Anas boschas: Fuligula marila; Dublany), to (Echinost.).—Sons., 1897, 252.—Stoss., 1892, 162 (to Echinost.).—Villot, 1878, 25.

recurvum Duj. 1845a, 410 (in Mus sylvaticus; Rennes), to (Brachylaimus).—Braun, 1899g, 492; 1901e, 339–341 (includes D. musculi Rud.; Duj.; Dies.), 341.— Cobbold, 1860a, 9; 1879b, 316.—Dies., 1850a, 389.—Stoss., 1892, 19-20 (in Mus

sylvaticus; Rennes).

refertum Mueh., 1898, 18 (in Cypselus apus; East Prussia). 26, 84, 86-87, figs. 5, 12. -Braun, 1902b, 100, 109 (= clathratum Desl.).—Jacoby, 1900, 10, 11.-Looss, 1899b (to Dicroccelium).—Rail., 1900, 239 (syn. of Dicroccelium

clathratum Deslong.).

reflexum Crep., 1825a, 54 (in Cyclopterus lumpus; Mar. and Apr.).—Bellingham, 1844a, 425.—Ben., 1858a, 1861a, 178; 1870, 51, pl. 5, fig. 3.—Braun, 1892a, 578, 699, 700, 734, 737; 1893a, 873, 910.—Cobbold, 1860a, 24.—Dies., 1850a, 373 (includes D. cyclopteri in Cyclopterus lumpus; Greifswald); 1858e, 342 (in Cyclopterus lumpus).—Duj., 1845a, 467.—Hausmann, 1897b, 4, 6, 20, 22 (in Trutta salar).—Jacoby, 1900, 11.—Kroyer, 1838—40a, 613 (in Cyclopterus lumpus).—Mont., 1893, 95, 102.—Nicoll, 1907, 73, 74.—Odhn., 1901, 484, 485, 506, 508, 509, 512; 1905, 321.—Olss., 1868, 52.—Stoss., 1886, 28.

reinhardi Linst., 1903, 280-282, fig. 16 (in Astacus leptodactylus).

renale Fil., 1855b, 19-20, 23, pl. 2, fig. 21 (in Helix adspersa near Turin); 1857, 435–436, fig. 21, pl. 2.—Dies., 1858d, 266 to Cerc. (Gymnocephala).—Erc., 1881e, 64, 65; 1882a, 300, 301.—Moul., 1856a, 164, 216 (to Cerc.) (in Helix aspera at Turin).—Par., 1894, 160.

reniferum Looss, 1898, 461 (D. unicum Looss, not Mol., renamed); 1899b, 590 (type of Astia); 1900 (type of Astiotrema).—Braun, 1901b, 37.—Luehe,

1899, 531 (reniforme).

reniforme Luehe, 1899, 531 (syn. unicum Looss, nec Mol.), 532 for reniferum Looss.—Stoss., 1904, 2.

repandum Rud., 1819a, 681 (in Rana sp.; Brazil).—Dies., 1850a, 355.—Cobbold, 1860a, 19 (in Cystignathus pachypus; Brazil).—Duj., 1845a, 454.—Stoss., 1889, 71 (in Cystignathus ocellatus).

recticulatum Wright, 1879, 58-59, pl. 1, fig. 6 (in Ceryle alcyon); 1879, 7.—Braun, 1892a, 570, 698; 1893a, 876.—Looss, 1894a, 171; 1899b (= Fasc.).—Stoss., 1892, 154.

reticulatum Looss, 1885b, 40–57, 59 (retikulatum), pl. 23, figs. 16, 20–29 (in "Wels;" mlatum Looss, 1885b, 40-57, 59 (retikulatum), pl. 23, ngs. 16, 20-29 (in weis; Costa Rica); 1885a, 427-444; 1894a, 137, 138, 171; 1899b, 536, 585, 650, 651, (to Clinost.).—Biehringer, 1888a, 230, 231, 232, 233.—Bock, 1886a, 544.—Braun, 1892a, 603, 607, 608, 611, 615, 640, 650, 651, 664, 677, 688, 735, 741, 742; 1892, 46; 1893a, 871; 1899g, 484, 485, 486, 487, 491; 1900h. 2, 3, 4, 5, 6, 9, 13, 43, 44; 1900. 26, 31; 1902b, 129.—Jackson, 1888, 644, 645.—Linst., 1890f, 183.—MacCallum, 1899, 705, 707.—Mont., 1888a, 15, 30, 32, 38, 43, 54, 57, 92 (type of Mesogonimus); 1893, 18, 44, 65, 66, 83, 85, 88, 90, 91, 102, 106, 107, 155, 156.—Rossbach, 1906, 377.—Stiles & Hass, 1898a, 86 (see D. dietyotus) (type 156.—Rossbach, 1906, 377.—Stiles & Hass., 1898a, 86 (see D. dictyotus) (type of Mesogonimus).—Reported for Acipenser sp.

reticulatum Poir., 1886, 39, pl. 3, fig. 8 (in Axinurus dugesii).—Braun, 1899g, 491
 (syn. of Clinost. marginatum); 1900h, 44, 45.

retikulatum Looss, 1885b, 59, for reticulatum.

retroconstrictum Srámek, 1901, 95, 108, fig. 62 (syn. Monost. constrictum) (in Abramis brama Cuv.); 1902, 21 Apr., 173.

retroflexum Mol., 1859, 290 (in Belone acus; Batavii); 1861, 213.—Braun, 1893a, 910; 1893b, 184 (in Bel. vulgaris).—Carus, 1884, 124.—Dies., 1859c, 432.—Luehe, 1900, 488, 492; 1901, 480.—Stoss., 1883, 117 (in Bel. acus); 1886, 16; 1891, 216.

retusum Duj., 1845a, 405–406 (in Rana temporaria; Rennes), to (Brachycœlium).—Baillet, 1866b, 96.—Ben., 1858a, 1861a, 92–96, 178, 186, pl. 11, figs. 9–27.—Braun, 1892a, 767; 1893a, 860, 866, 881.—Cobbold, 1860a, 18 (in Rana esculenta; Valentin).—Dav., 1877a, lxxi, fig. 34.—Dies., 1850a, 388 (includes Dclavigerum; D. ranæ esculentæ); 1858, 348 (in Rana halecina); 1859c, 434 (cf. Cerc. armata minor Ben., 1858a, 98, pl. 11, figs. 9–27, in Lymnæus stagnalis; Belgium).—Erc., 1881e, 73, 82, 84, 85, 89; 1882a, 309, 318, 320, 321, 325.—Fraip., 1880c, 417.—Gamb., 1896a, 72.—Hahn & Lefèvre, 1884a, 516 (of "Rud.").—Leidy, 1851, 207; 1856, 44.—Linst., 1887, 97, 98, 99.—Looss, 1894a, 82, 84 (of Ben. is syn. of D. endolobum), 85, 208; 1899b, 611.—Luche, 1899, 536.—Mont., 1893, 43.—Pag., 1857, 41.—Staff., 1902, 724.—Stiles, 1901, 197, 199, 201.—Stoss., 1889, 63.—Also reported for Phryganeidæ larvæ.

rhatonisii Simon, 1897, 223, misprint for rathouisii.

rhizophisæ Mont., 1888, 199 (for rhizophysæ).

rhizophysæ Studer, 1878, 12–13, pl. 1, figs. 2, 7 (in Rhizophysa conifera).—Braun, 1893a, 869; 1893d, 468.—Mont., 1888, 199 (rhizophisæ); 1893, 123.

[rhodopyge Sluiter, 1898 (a tunicate).]

rhombi Ben., 1870, 72 (in Rhombus maximus).—Braun, 1893a, 872.

ricchiardii Lopez, 1888, 137–138.—Looss, 1899b, 552, 736; 1901, 204, 206; 1902m, 482, 791 (amphitypie), 846, 854.

richardii Brand., 1891b, 267 (for richiardii).

richiardi Mont., 1891, 500 (for richiardii).

richiardii Lopez, 1888a, 137–138 (in Acanthias vulgaris; Risso).—Ariola, 1899, 8.—Brand., 1891b, 267 (richardii).—Braun, 1892a, 698, 715.—Crety, 1892b, 373 (vitelline nuclei of); 1892c, 396, 399 (richiardi); 1892, 24–26, fig. 2; 1892, (92–97); 1893a, 380, 382–384 (richiardi).—Linst., 1888, 46; 1903, 354.—Looss, 1894a, 134, 145, 166, 179, 189, 190, 200, 204, 210, 211, 224; 1902m, 482 (ricchiardii), 791, 846, 854, 855 (type of Probolitrema).—Mont., 1889, 132–134; 1889, 612; 1891, 500 (richiardi); 1892, 5, 6, 7 (richiardi); 1892, Oct. 7, 188 (richiardi); 1893, 17, 19, 32, 33, 34, 40, 41, 42, 43, 49, 50, 61, 62, 65, 66, 67, 68, 69, 71, 72, 76, 78, 79, 82, 83, 84, 85, 86, 87, 88, 90, 91, 94, 95, 96, 97, 98, 99, 102, 103, 106, 107, 109, 110, 112, 114, 117, 120, 122, 135, 139–148, 209, pl. 1, fig. 10, pl. 5, fig. 51–55, pl. 6, fig. 87, pl. 7, figs. 91–102, pl. 8, figs. 125–130; 1899, 103; 1896, 147, 166 (richiardi).—Ofenheim, 1900, 147, 153, 156, 160, 163, 164, 169, 174.—Shipley & Hornell, 1905, 54, 55 (in Rhinodon typicus).—Sons., 1890, 135 (richiardi).—Stiles, 1896, 205.

rigens Linst., 1878a, 282 (in Scymnodon ringens) [apparently lapsus for Dist. ligula Ben., 1871a, 12], 360 (ringens).—Ariola, 1899, 8.—Mont., 1893, 53.

ringens Rud., 1819a, 101, 385 (in Picus tridactylus; Mus. Vien.).—Braun, 1893a, 877; 1893d, 467; 1901, 561, 568.—Dies., 1850a, 361 (syn. of D. macrost.).—Duj., 1845a, 444.—Linst., 1903, 354.—Stoss., 1892, 184 (syn. of D. macrost.); 1898, 23.—Schlotthauber, 1860, 130.

ringens Linst., 1878a, 360, see rigens.

ringeri Cobbold, 1880, 139–140, pl. 10, figs. 1–3 (Homo); 1884g. 976.—Anders, 1903, 6 ed., 1245 (syn. of D. pulmonale).—R. Bl., 1888a, 627–630, 631, fig. 326; 1891, 610 (syn. of D. westermanni).—Braun, 1893a, 876, 877, 882; 1901e, 332; 1903, 3 ed., 155 (syn. of Paragonimus westermanni); 1906, 161.—Brunet, 1902a, 125.—Chedan, 1886a, 241–244.—Coplin, 1898, 339–340.—Dunglison, 1893, 338.—Eyles, 1887a, 660.—Gamb., 1896a, 63.—Giard & Billet, 1892a, 614.—Hackley, 1886, 519.—Hahn & Lefèvre, 1884a, 546–548 (syn. of D. pulmonale Bælz).—Huber, 1896a, 576 (syn. of D. pulm.).—Ijima, 1889b, 148.—Inouye, 1897c, 1–4 (in brain); 1903a, 120–135; 1904a, 617–618.—Jackson, 1888a, 653.—Katsurada, 1900, 506.—Linst., 1889, 3; 1890f, 178.—Lockwood, 1901, 2 ed., 821.—Looss,

1905, 84 (syn. of Par. westermani).—Manson, 1880, Aug., 139–140, pl. 10, figs. 1–3; 1881, 8–9; 1881, July 2, 10–12, figs. 1–9; 1882, 55–62, figs. 1–25; 1882, July 8, 42–45; 1883, Mar. 31, 532–534; 1883, 1813; 1883, 134–138, 138–156, pls. 8–9; 1886, 241–244; 1893, 852–860, fig. 79; 1901, 541; 1903, 3 ed., 631 (syn. of Par. westermanni).—Moniez, 1896, 144.—Mont., 1893, 155.—Mosler & Peiper, 1894, 178.—Mouye (1904), v. 1 (1–2); 1904, v. 2, 144.—Packard, ——, 523.—Rail., 1886, 296.—Rail. & Marotel, 1898, 31.—Simon, 1897, 259.—Sons., 1884, 17, 18, 19, 20, 21.—Stiles & Tayler, 1902a, Apr. 19, 45.—Stoss., 1892, 32 (syn. of Mesogonimus pulmomale).—Taniguchi, 1903, Dec., 100.—Ward, 1895, 244 (syn. of D. westermanni), 328 (in Homo), 341 (in Canis familiaris); 1903, 867 (syn. of Par. westermanii).—Yamagiwa, 1891, 5 Jan., 36–41; 1892, 446.—In tiger.

ringers Rev. Sci., 1890, 189, misprint for ringeri.

robustum Lorenz, 1881a, 583–586, pl. 19 (in Elephas africanus), figs. 1–6.—Braun, 1892a, 593, 603.—Mont., 1893, 82, 105, 106, 107.—Stoss., 1892, 18 (in El. afr.).

rochebruni Poir., 1886, 36–37, pl. 4, figs. 4–5 (Delphinus delphis).—Braun, 1892a, 673 (rochebrunni); 1893a, 875, 910; 1893, 354; 1900g, 251, 252, 253.—Looss, 1894a, 204; 1899b, 560 (to Brachycladium).—Mont., 1893, 44, 107.—Stiles, 1895m, 219, pl. 8, figs. 4, 5.—Stoss., 1892, 11 (to Cladocœlium).

rochebrunni Braun, 1892a, 637, for rochebruni.

rosaceum Nord., 1832a, 82–88, pl. 8, figs. 1–5, 11 (in Gadus lota; October); 1840, 617, 620 (syn. of Fasc. eriocis).—Ben., 1858a, 1861a, 99, 100 (syn. of D. tereticolle).—Braun, 1892a, 653, 757.—Cobbold, 1860a, 26; 1879b, 458.—Crep., 1837, 313, 314, 318, 321, 323.—Dies., 1850a, 364.—Harz, 1881c, 5.—Kroyer, 1838–40a, 609 (in Lota vulgaris).—Looss, 1894a, 5, 6 (syn. of D. teret.).—Moul., 1856a, 49 (seems to be only a var. of teret.).—Stoss., 1886, 25; 1898, 38.—Wagener, 1857, 25.

rosarum Cobbold, 1860a, 21; misprint for rosaceum.

rosea Ben., 1870, 1871a, 90, pl. 4, fig. 10 (in Petromyzon omalii; Belgium).—Linst., 1878a, 290 (in P. fluviatilis).

rubellum Olss., 1868, 40, pl. 4, fig. 89 (in Labrus maculatus; Scandinavia).— Braun, 1892a, 579; 1893a, 910.—Looss, 1899b, 618; 1901d, 399.—Luehe, 1899, 537.—Odhn., 1902, 59, 61.—Staff., 1905, Apr. 11, 692.—Stoss., 1886, 20.

rubens Duj., 1845a, 411 (in Sorex fodiens, S. tetragonurus; Rennes) (to Brachylaimus).—Braun, 1901e, 342, 344 (syn. of D. exasperatum Rud.).—Cobbold, 1860a, 6; 1879b, 296.—Dies., 1850a, 334–335 (in Sorex daubentonii; Rhedoni). Stoss., 1892, 15 (syn. of D. exasp.).

[rubrum Savigny, ——, 176 (a tunicate).]

rude Dies., 1850a, 360–361 (in Lutra brasiliensis; Brazil); 1855, 66–67, pl. 3, figs.
9–10; 1858e, 341.—Braun, 1892a, 735; 1893a, 876, 877; 1899g, 492; 1901e, 327, 329, 332, figs. 12, 15, 17 (to Paragonimus).—Cobbold, 1860a, 7; 1859d, 363; 1879b, 298.—Kerbert, 1878a, 272.—Lamouroux, 1824, 563 [Distome rude].—Stiles & Hass., 1900a, 604 (to Paragonimus).—Stoss., 1892, 36 (in Lut. bras.).

rufoviride Rud., 1819a, 110, 406-407 (in Muræna conger; Naples, July).—Bellingham, 1844a, 425.—Ben., 1858a, 1861a, 178, 181, 189, 193, 195.—Braun, 1891d, 424 (in Uranoscopus scaber); 1892a, 705; 1893a, 879.—Carus, 1884, 125.—Cobbold, 1860a, 22 (in Conger vulgaris).—Dies., 1850a, 342, 371, 372 (includes D. varium Eysenhardt) (in Capros aper, Naples; Labrax lupus, Rhedoni; Saurus saurus, Naples); 1858e, 342 (in Cong. vulg.).—Duj., 1845a, 421.—Jackson, 1888a, 647.—Kroyer, 1846-53a, 615 (in Anguilla conger L.).—Lander, 1904a, 7 (to Lecithochirium).—Levin., 1881a, 60.—Lint., 1898, 515-517, pl. 42, 43, figs. 1-4: 1901, 415, 418, 455.—Linst., 1903, 354.—Looss, 1899b, 640 (to Hemiurus).—Luehe, 1901, 58; 1901, 474, 476.—Mol., 1858, 129; 1859, 827, 841, 844 (in Scorpæna porcus, S. scropha, Anguilla vulgaris, Labrax lupus, Trigla corax); 1861, 205-209, pl. 2, figs. 1, 2, 4, 5.—Mont., 1888a, 7, 8, 17; 1891, 497, 500, 502.—Olss., 1868, 49; 1876, 20.—Pag., 1862, 305, pl. 29, figs. 9-10.—Sons., 1891, 259, 261 (in Rhombus lævis, Trigla cuculus).—Stoss., 1883, 115 (in Anguilla vulg.); 1885, 159; 1886, 13 (= D. caudiporum Duj.); 1890, 40, 50; 1891, 112; 1902, 582.—Wagener, 1860, 166, 178-181, 183, pl. 9, figs. 6-10.—Also reported for Acipenser sturio, Centronotus glaucus, Cepola rubescens, Conger conger, Dactylopterus volitans, Roccus lineatus, Saurus lacerta.

saginatum Ratz, 1898, Oct. 15, 73-75, fig. 5 (in Ardea alba); 1900, 437-439.

salamandræ (Freelich, 1789) Zed., 1803a, 215 (in Salamandra nigra).—Dies., 1850a, 356 (syn. of D. crassicolle).—Rud., 1809a, 379 (syn. of D. crassicolle).

salamandrinæ perspicillatæ Sons., 1896, 1; 1896, 116 (in Salamandrina perspicillata Sav).

sanguineum Sons., 1894, 111; 1894, 1-4 (n. sp. in Chamæleo vulgaris; Gabes, Tunis);
1895, 124.—Braun, 1899b, 718; 1900, 225.—Jacoby, 1900, 11.—Looss, 1896b,
106-114, pl. 7, figs. 69-74, pl. 8, figs. 75-78 (in Taphosus nudiventris at Cairo;
caméléon in Egypt); 1899b, 556, 636, 637 (type of Anchitrema).—Ofenheim,
1900, 183.—Stoss., 1895, 217 (in Camæleo vulg.; Tunisia).

sauromates Poir., 1886, 24–26, pl. 2, figs. 4–6 (in lungs of Elaphis sauromates).—
Braun, 1892a, 699, 736, 737; 1893a, 876 (sauromatis).—Looss, 1894a, 204.—
Luehe, 1899, 532, 533; 1900, 561.—Mont., 1888a, 57; 1893, 83, 86, 95, 102, 107.—Sons., 1893, 185; 1893, 216.—Stoss., 1895, 220–221 (in Elaphis sauromates); 1904, 2.—Volz, 1899, 232, 234, 237.

sauromatis Braun, 1893, 876, for sauromates.

scaber Rud., 1819 of Odhn., 1905, 353, perhaps a Stephanochasmus.

scabrum (Mueller, 1788) Zed., 1803a, 215–216 (in Gadus barbatus).—Bellingham, 1844a, 427.—Ben., 1870, 61 to (Echinost.).—Carus. 1884, 126.—Cobbold, 1858b, 158; 1860a, 37.—Crep., 1837. 311.—Dies., 1850a, 393, 398 (in Morrhua barbata, Lota molva; Naples); 1858e, 351 (in Merlangus pollachius).—Duj., 1845a, 432–433.—Kroyer, 1838–40a, 605; 1843–45a, 41, 166 (in Gadus morrhua, Lota molva).—Looss, 1899b, 581, 582 (of Mueller, generically Hemiurus, specifically problematic) (of Rud., probably a Stephanost.).—Mont., 1891, 507, to (Fasc.).—Nicoll, 1907, 78.—Olfers, 1816. 46.—Rud., 1809a, 387, 406–408, 428, 435; 1810a, 376; 1819a, 118, to (Echinost.), 122, 124 (see also scaber).—Stoss., 1886, 34.—Also reported for Gadus pollachius. Molva vulgaris.

scimna Risso, 1826, 262 (also spelled scymna and scymni by various authors) (in Echinorhinus spinosus; Europe).—Ariola, 1899, 7 (scymni).—Dies., 1850a, 347 (syn. of D. insigne).—Mont., 1893, 52 (scymni).—Villot, 1876, 1344–1346; 1878, 3 (scymna).

scombrinum Linst., 1889a, 80, for sobrinum.

scorpænæ Rud., 1819a, 122 (in Scorpæna scrofa; C. E. V.):—Barbagallo & Drago, 1903, 410 (in Scorpæna lutea; Catania) to (Dicrocœlium).—Braun. 1892a, 569, 642, 728.—Dies., 1850a, 398.—Looss, 1899b, 571 (this form as described by Stoss., 1885. 3, 5, belongs to the Creadiinæ, possibly to Creadium).—Mont., 1893, 83, 84, 95.—Stoss., 1885, 158 (in Scorpæna scrofa; Triest); 1886, 33; 1898, 49.

scorpii (Mueller, 1776) Zed., 1803a, 216.—Dies., 1850a, 366 (syn. of D. granulum) (in Cottus scorpio).—Rud., 1809a, 395.

77:11 + 1070 0 2 :

scymna Villot, 1878, 3, for scimna.

scymni Mont., 1893, 52, for scimna.—Ariola, 1899, 7 (syn. of D. veliporum). scyphocephalum Braun, 1899e, 630 (in Testudo matemata; Brazil); 1901b, 34-36,

fig. 14. segmentatum Mueller, 1894, 125–127, pl. 7, fig. 10, to (Echinost.).

semiarmatum Mol., 1858, 131 (in Acipenser naccari; Patavii); 1861, 223.—Braun, 1892a, 584.—Carus, 1884, 127.—Dies., 1858e, 352.—Odhn., 1902, 159.—Stoss., 1886, 42.

semiflarum Linst., 1880, 50-51 (in Petromyzon fluviatilis).—Stoss., 1886, 54.

semisquamosum Braun, 1900b, 228–229. pl. 10. figs, 6, 7 (in Vesperugo noctula).—Looss, 1907. Mar. 7, 483 (belongs in Parabascus).

seriale Rud., 1808a, 351 (in Salmo alpinus; Greenland); 1809a, 368-370 (in Salmo alpinus), includes Fasc. umblæ, 1780; 1819a, 97.—Braun, 1893a, 876; 1893d, 467.—Cobbold, 1860a, 23; 1879b, 457 (in Salmo umbla).—Dies., 1850a, 343 (includes D. umblæ; Fasc. umblæ).—Duj., 145a, 464.—Olfers, 1816, 45.—Stoss., 1886, 54.

serpentatum Mol., 1859, 830-831 (in Sayris camperi: Padua).—Par., 1896, 18-19, fig. 7 (to Brachylaimus).—Also reported for Scomberesox rondeletti.

serpentulum Carus, 1884, 131 (for serpentatum).—Stoss., 1886, 54.

serratum Dies., 1850a, 385 (in Aranus scolopaceus; Brazil); 1855a, 67, pl. 3, figs. 14-17; 1858e, 347.—Braun, 1892a, 571.—Stoss., 1892, 171.

sialidis Linst., 1892, 334, pl. 15, fig. 20 (in Sialis lutaria).—Braun, 1893b, 183.

sibiricum Winogradow, 1892, 116-130 (in Homo); 1892, Nov. 30, 910-911; 1900, 617-618 (in Man).—Anders, 1903, 6 ed., 1245 (in man).—Askanazy, 1900, 491, 492, 494; 1901, 73, 77 (in Tomsk, in Homo, dog, cat).—Bossuat, 1902, v. 6 (2), 188 (syn. of Opisthorchis felineus).—Braun, 1894g, 128-129; 1894h, 755; 1894i, 602-606; 1903, 3 ed., 157 (syn. of Op. fel.), 159, fig. 106.—Huber, 1896a, 576.— Kamensky, 1900a, 5, 19.—Katsurada, 1900, 500.—Kholodk., 1899a, 152.—Looss, 1899b, 674; 1905, 89 (syn. of Op. fel.).—Moniez, 1896, 137, fig. 27.—Simon, 1897, 209, 223.—Ward, 1895, 328 (in Homo); 1903, 869 (syn. of Op. fel.).—Weichselbaum, 1898, 315.

siemersii Buttel-Reepen, 1900a, 586, 589, 596-598, figs. 8-9 (in Sphyræna barracuda; Atlantic Ocean); 1902, Dec. 8, 165–236, pl. 6, fig. 26, pl. 10, fig. 54, text fig. h; 1904, Jan. 26, 24–25; 1905, July, 52–53.—Darr, 1902, 668, 671.

signatum Duj., 1845a, 415-416 (in Coluber natrix; Rennes) to (Brachylaimus. Braun, 1891, 100; 1892a, 780, 784, 785; 1893a, 864, 866, 873; 1901b, 13, 16 (of Erc., syn. of D. ercolanii Mont.).—Cobbold, 1860a, 20.—Dies., 1850a, 390 (in Tropidonotus natrix); 1858e, 350 (in Tropidonotus).—Gamb., 1896a, 72.— Hoyle, 1890, 538.—Kampmann, 1894b, 454, 456, 457, pl. 20, fig. 10.—Linst., 1879, 185.—Looss, 1899b, 706.—Luehe, 1899, 528 (of Erc., not Duj.).—Mont., 1893, 187, 188.—Mueh., 1898, 30.—Schauinsland, 1882, 496.—Sons., 1893, 186.—Stoss., 1895, 216–217, 1898, 34.—Volz, 1899, 235, 237.—Wedl, 1855, 400–401, pl. 2b, fig. 8.—West, 1896, 323.—Also reported for Natrix torquata.

signatum of Erc., 1881c, 73, 74, 75, 76, 78, 79, 81, 82; 1882a, 309, 310, 311, 312, 314, 315, 317, 318, pl. 2, figs. 2–5 (in Tropidonotus natrix), renamed D. ercolanii by Mont., 1893, 187, 188, pl. 6, fig. 67.—Braun, 1901b, 13, 16.—Luehe, 1899, 528.—Par., 1904, 1.—Stoss., 1904, 1.

simile Sons., 1890, 105 (in Python molurus); 1893, 215, 216; 1893; 185 (in Py. mol.); 1893, 499.—Looss, 1899b, 602.—Luehe, 1899, 532.

simile Looss, 1899b, 602 (see Hæmatolæchus similis).—Stiles, 1901, 178.—Stiles & Hass., 1902d, 20 (renamed Hæmatolæchus similigenus).

simillimum Mueh., 1898, 18 (in Fuligula nyroca); 1898, 26, 96–97, 98, figs. 4, 19.— Braun, 1902b, 11, 18.—Looss, 1899b, 574 (to Psilost.).

simplex Rud., 1809a, 370–371 (in Gadus æglefinus), Fasc. æglefini Mueller, 1776, renamed; 1819a, 97.—Braun, 1892a, 642, 699, 711, 728, 736; 1893a, 866, 879.—Cobbold, 1858b, 157.—Dies., 1850a, 343–344.—Duj., 1845a, 466.—Kroyer, 1838–40a, 606 (in Gadus æglefinus).—Lamouroux, 1824a, 563 (distome simple).—Levin, 1881a, 18, 67–69, pl. 3, fig. 1 (of Olss., syn. Fasc. æglefini) (larva in Themisto libellula; adult in Cottus scorpius; Phobetor ventralis; Egedesminde); 1876, 15.—Lint., 1898, 525–526, pl. 47, figs. 3–7; 1900, 295; 1901, 415 (Hemitripterus americanus, Leptocephalus conger, Limanda ferruginea, Microgadus tomcod), 420, 436, 451, 468, 475, 482, 483, 485, 486, figs. 331, 332; 1905, 328, 334, 397 (in Micropogon undulatus).—Linst., 1889a, 97 (in Conger vulgaris).—Mont., 1893, 61, 83, 86, 94, 102, 105, 106, 107.—Nicoll, 1907, 70, 73 (of Olss., syn. of Podocotyle atomon).—Nord., 1840, 619 (syn. of Fasc. æglefini).—Odhn., 1901, 484, 485, 506, 508, 509, 510, 511, 512, 513; 1905, 320–321 (of Olss., syn. of Podocotyle atomon) (to Sinistroporus, designated type in letter to Stiles, May 15, 1905).—Olfers, 1816, 45.—Olss., 1868, 34; 1876, 15.—Staff., 1904, 484.—Stoss., 1886, 30–31 (in Anguilla vulgaris, Cottus scorpius, Gadus æglefinus, G. melanostomus, Lota vulgaris, Phobetor ventralis, Raniceps niger, Sebastes norvegicus); 1902, 582.—Also reported for Conger vulgaris.

simplex Rud. of Olss., 1868, 34; 1876, 15.—Levin., 1881, 67-69 (syn. Fasc. æglefini) (larva in Themistone libellula; adult in Cottus scorpius, Phobetor ventralis; Egedesminde).—Odhn., 1905, 320-321 (syn. of Podocotyle atomon).

simplex Polonio, 1859, teste Par., 1894, 147 (in Lacerta muralis; Padua).

simulans Looss, 1896b, 52–54, pl. 4, figs. 28–30 (in Pernis apivorus; Alexandria);

1899b, 673 (to Opisthorchis).—Jacoby, 1900, 7.

sinense Cobbold, 1875i, Sept. 18, 423 (in Homo); 1875, Oct., 780–781; 1876, 97 (same as chinense and macconnelli); 1877, Jan., 15–16; 1883, 401; 1884g, 976.—Bælz, 1883, 234.—Biggs., 1890a, 30–37, 1 fig. (in U. S. A.).—Billet, 1893a, 506-510, figs. 1-2 (syns.: D. hepatis perniciosum, D. hepatis innocuum, D. japonicum).—R. Bl., 1888a, 585, 596, 615-618 (syn. D. spathulatum Leuck.), 622 (similar to D. japonicum), 631; 1891, 605, 607-609, 610 (syns.:

D. japonicum Bl., D hepatis endemicum sive perniciosum, D. hepatis inno-D. japonicum Bl., D hepatis endemicum sive perniciosum, D. hepatis innocuum); 1901b, 209; 1901c, 586 (to Opisthorchis).—Braun, 1891d, 426; 1893, 349, 352, 353; 1893a, 875; 1893f, 386, 425, 427, 428; 1894f, 605; 1895b, 146–147, figs. 63–64; 1903, 3 ed., 161 (to Opisth.).—Bruce, 1897a, 211–212.—Brunet, 1902a, 125.—Caræs, 1888a, —, pl. 2 (liver flukes of man in Orient); 1888, 44 pp., 4 figs.—Chester, 1887a, 360.—Corlette, 1897a, 146–147 (Australia).—Delafield & Prudden, 1897, 130.—Gamb., 1896a, 63, 70.—Giard, 1904, 8.—Grall, 1887a, 468, 1 fig; 1887, xlviii.—Hackley, 1886, 518–519, fig. 883.—Hahn & Lefèvre, 1884a, 544–545 (syns. D. spathulatum Leuck., D. hepatis innocuum Balz).—Hori 1890.—Huber 1896a, 577 (syn. of D. spathulatum Leuck.) 883.—Hahn & Lefèvre, 1884a, 544—545 (syns. D. spathulatum Leuck., D. hepatis innocuum Bælz).—Hori, 1890.—Huber, 1896a, 577 (syn. of D. spatulatum Leuck.).—Ijima, 1889b, 145.—Jamieson, 1897a, 71–74, 1 fig. (jaundice); 1897b, 147—148.—Janson, 1893c, 265—266.—Katsurada, 1891; 1900, 479.—Kholodk., 1898, 26, 28, pl. 11, figs. 6–9; 1899a, 152.—Kuech., 1881, 333–335, pl. 8, fig. 10A.—Kurimoto, 1893a, 1–7, 21–24, 35–39, 1 pl. (in Saga, Japan); 1893b, 67–69, 85–87, 109–111.—Linst., 1903, 279.—Looss, 1905, 90 (to Opisth.); 1907, Feb. 1, 136, 137, 141, 142, 143, 144, 147.—MacGregor, 1877, 3–16, 1 pl.—McConnell, [1875, 271–274; 1876, 343 (sineuse); 1878, March 16 406 (m'connelli)]—Manson, 1901 (Gibson), 540; 1903, 3 ed. 631, 635–637. 16, 406 (m'connelli)]—Manson, 1901 (Gibson), 540; 1903, 3 ed., 631, 635–637, 639.—Moniez, 1896, 86, 123, 125–136, 137, fig. 25 (excellent discussion).—Mosler & Peiper, 1894, 177–178, fig. 70 (sinese).—Moty, 1893, March 3, 224–230 (pathology).—Nakahama, 1883, —..—Pfihl, 1884, 156.—Rail, 1890, 142, 143.—Remy, 1883, 513.—Schneidemuehl, 1896, 302.—Simon, 1897, 223.—Sons., 1889, 278, 279, 280; 1889, 7 July, —; 1896, 297, 302.—Staff., 1902, 483 to (Opisth.).—Stoss., 1892, 23 (syn. of D. endemicum).—Taylor, 1875, 772–780, figs. 1–3; 1884, 48–51, 52–53; 1885, 58–60.—Tyson, 1903, 3 ed., 1180.—Vallot, 1889, 382.—Ward, 1895, 238 (man, cat), 328 (in Homo); 1903, 869 (to Opisth.).—Weichselbaum, 1898, 315.—White, 1902, Dec., 523 (cases in U. S. A.).—Wood & Fitz, 1897, 335. 16, 406 (m'connelli)].—Manson, 1901 (Gibson), 540; 1903, 3 ed., 631, 635-637,

sinese Mosler & Peiper, 1894, 177-178, fig. 70 (for sinense).

sineuse of McConnell, 1876, 343 (misprint for sinense).

singulare Mol., 1859, 288 (in Ibis falcinellus); 1861, 200-201, pl. 2, fig. 6, pl. 3, fig. 3.—Braun, 1891d, 421; 1892a, 578, 700, 736, 737; 1901, 260; 1901, 896; 1902b, 86 (syn. of Stomylotrema pictum Crep.).—Dies., 1859c, 428 (in Ibis falcinellus; Patavii).—Looss, 1899b, 723 (type of Stomylus).—Stoss., 1892, 162.

singulare Mol. of Looss, 1899b, 723.—Braun, 1902b, 86 (renamed Stomylotrema

perpastum).

sinuatum Rud., 1819a, 97-98, 374 (in Ophidium imberbe; Naples).—Braun, 1883a, 41; 1891d, 421; 1892a, 672, 763, 765.—Carus, 1884, 131.—Cobbold, 1860a, 25.—Dies., 1850a, 344 (in Fierasfer imberbe).—Duj., 1845a, 468.—Odhn., 1901, 484, 490, 491; 1902, 160.—Par., 1902, 6 (in Ophidium barbatum; Portoferrajo).—Stoss., 1886, 55.—Will.-Suhm, 1870, 7-8; 1871, 181-182, pl. 11, fig. 6.—Also reported for Fierasfer acus.

siredonis Poir., 1886, 32–33, pl. 3, figs. 4–5 (in Siredon mexicanus, int.).—Looss, 1894a, 204, to Opisthioglyphe.—Mont., 1893, 83, 86, 102, 107.

sirenis Braun, 1893a, 870 for sirenis lacertinæ Vailliant.

sirenis lacertinæ Vailliant, 1863, 348-350, pl. 9, fig. 9 (in Siren lacertina).—Stoss., 1889, 69.

sluiteri (Brock, 1886) Braun, 1892a, 651.

sobrinum Levin., 1881a, 70-71 (in Cottus scorpius; Egedesminde).—Looss, 1899b, 581 (to Stephanost.).—Odhn., 1905, 331 (to Stephanochasmus).—Also re-

ported for Aspidophorus decagonus.

soccus Mol., 1858, 129 (in Mustelus plebejus; Patavii); 1861, 203.—Ariola, 1899,
7 (syn. of D. megastomum).—Braun, 1893a, 873.—Dies., 1858e, 351 (in Mustelus plebejus).—Jacoby, 1900, 17.—Linst., 1903, 354.—Mont., 1893, 53, 192,
193.—Stoss., 1883, 118; 1890, 131; 1898, 38.

sociale Luehe, 1901p, 171–173 (in Bufo melanostictus Schneider).—Braun, 1901h,

700.—Klein, 1905, 20, 78.—Odhn., 1902, 42.

solex Duj., 1845a, 417-418 (in Pleuronectes solea; Rennes) to (Brachylaimus).— Dies., 1850a, 399.—Kroyer, 1838–40a, 612 (in Solea vulgaris).—Mont., 1893, 193.—Stoss., 1886, 55.—Wagener, 1860, 184.

solexforme Rud., 1809a, 384 (in Trigla gurnardus) (D. triglæ gurnardi renamed); 1819a, 104.—Braun, 1893a, 873.—Cobbold, 1860a, 26.—Dies., 1850a, 362.— Duj., 1845a, 457.—Fraip., 1881b, 4 (in Tr. gur.).—Kroyer, 1838–40a, 100 (in Tr. gur.).—Olfers, 1816, 45.—Stoss., 1886, 55.

soleare Braun, 1899e, 629-630 (in Testudo midas); 1901b, 22-23, fig. 8.

somateriæ Levin., 1881a, 71–73, pl. 3, fig. 2 (in Somateria mollissima; Egedesminde).—Braun, 1892a, 578, 579, 586, 642, 735; 1893a, 893, 910.—Giard, 1903g, 1225.—Jægers., 1898, 15, 16; 1900, 738.—Jameson, 1902a, 140–166, 4 pls., 3 figs.—Linst., 1890, 179.—Looss, 1894a, 173; 1899b, 618, 619; 1901, 207.—Luehe, 1898, 624, 625; 1899, 537.—Mont., 1888a, 12, 38; 1893, 43, 61, 83.—Odhn., 1900, 12, 13; 1905, 311 (to Gymnophallus), 313.—Staff., 1905, Apr. 11, 692.—Stoss., 1892, 146.

sophiæ Stoss., 1886, 44-45, pl. 8, figs. 34-35 (in Pagellus mormyrus; Triest); 1898, 49-50.—Braun, 1892a, 644, 720, 728.—Looss, 1899b, 571 (thinks this belongs to Creadiinæ, possibly to Creadium); 1902m, 785 (cf. D. isoporum armatum).—Mont., 1893, 85, 94, 102.—Odhn., 1905, 328, 338.

soricis Dies., 1858e, 354 (in Sorex araneus) (based on Distome (Dicroccelium) de la Musraigne-Musette Pontaillié, 1853, 103).—Braun, 1893a, 875.—Cobbold, 1860a, 9.—Stoss., 1892, 36 (in Crocidura aranea).

soricis aranei Dies., 1855, 64, footnote 3, to (Dicroccelium).

spari Rud., 1819a, 122 (in Sparus erythrinus, S. smaris; C. E. V.).—Dies., 1850a, 399.—Stoss., 1886, 55 (in Pagellus erythrinus, Smaris vulgaris).

spathaceum Rud., 1819a, 109, 403 (in Larus glaucus; Mus. Vien.).—Baird, 1853a,
48.—Crep., 1837, 310; 1839, 287.—Dies., 1850a, 310 (to Hemist.).—Nitzsch,
1819, 400.—Nord., 1840, 628 (to Holost.).

1819, 400.—Nord., 1840, 628 (to Holost.).

spathulatum Leuck., 1876, 871–872 (nec Crep., 1849) see Opisthorchis sinensis (see also D. spatulatum) (in Homo); 1889, 336–355, figs. 154–161.—R. Bl., 1888a, 615 (syn. of D. sinense); 1901b, 209; 1901c, 586 (syn. of Op. sin.).—Brand., 1891b, 265; 1891c, 731; 1891d, 11.—Braun, 1883a, 65; 1892a, 598, 602, 641, 661, 669, 671, 682, 699, 700, 704, 707, 712, 714, 717, 719, 724, 725, 732, 733, 784, 785; 1893, 349; 1903, 3 ed., 161 (syn. of Op. sin.).—Cobbold, 1876, 97; 1883, 401.—Duffek, 1902a, 774.—Gamb., 1896, 63.—Hahn & Lefèvre, 1884a, 544 (syn. of D. sinense Cobb.).—Harz, 1881c, 5.—Huber, 1896a, 577–578.—Ijima, 1889b, 145.—Inoue, 1900a, 1–60, 18 charts; 1903, 107–146.—Jamieson, 1897b, 147 (spatulatum) (syn. of D. sinense).—Katsurada, 1891b, 1–12, 1 pl. (case of cystoma hepatis); 1898a, 165–167 (relation to gallstones); 1900b, 479–505, pl. 13, figs. 1–13; 1900c; 1901a, 169–174; 1902a, 50–52; 1904, v. 2 (3), 22; 1904, Dec., 148 (in cats and dogs).—Laspeyres, 1904a, 16 pp. (case).—Looss, 1894a, 180, 186, 206, 212, 214; 1899b, 564; 1905m, 280 (syn. of Op. sin.); 1905, 90 (syn. of Op. sin.); 1907, Feb. 1, 141 (syn. of D. sinense).—MacGregor, 1877, May 26, 775–776 (in Homo).—Mont., 1888a, 39, 52 (spatulatum), 57; 1877, May 26, 775–776 (in Homo).—Mont., 1888a, 39, 52 (spatulatum), 57; 1891, 110; 1893, 83, 84, 86, 95, 102, 105, 106, 107.—Mosler & Peiper, 1894, 177.—Rail., 1890, 143.—Saito, 1898, June 5, ——; 1906, Sept. 10, 133–138, figs. 1–10; 1906, Sept. 20, 1555.—Sons., 1889, 278 (spatulatum).—Vogt, 1878, 10, 14.—Ward, 1903, 870 (syn. of Op. sin.).—Yamagiwa, 1891, 5 March; 1901, xxx, 155–168, fig. 1.

spathulatum Rud. (nec Leuck.) of Crep., 1837, 310; 1839, 287; 1849, 66–67.—Braun, 1901b, 29.—Kowal., 1896d, (3), 253 (in Botaurus minutus; Dublany), to (Echinost.) for spatulatum; 1902d (9), 27 (to Sodalis).—Looss, 1902m, 462.

spathulatum hepatis Simmonds, 1901, 110-111.

spatula Duj., 1845a, 394 (in Accentor modularis; Rennes) to (Dicroccelium).-

Dies., 1850a, 336.—Stoss., 1892, 156.

spatulatum Rud., 1819a, 109, 403–404, 600 (in Ardea minuta; Vienn. Mus.).—
Bremser, 1824c, pl. 9, figs. 15–16.—Crep., 1829b, 4.—Dies., 1850a, 367 (to Holost.).—Duj., 1845a, 376.—Harz, 1881c, 5.—Nitzsch, 1819, 400.—Nord., 1840, 628 (to Holost.).—Stoss., 1892, 186 (in Ardetta minuta; Vienna).—Also reported for Botaurus minutus.

spatulatum Cobbold, 1879b, 28 (for spathulatum Leuck.).—Aschoff, 1892, 495.— Askanazy, 1900c, 712.—Hoyle, 1890, 538.—Huber, 1896a, 277–278.—Jamieson, 1897b, 147.—Mont., 1888, 52.—Simon, 1897, 209, 223.—Stoss., 1892, 23 syn. of D. endemicum Bælz).—Ward, 1895, 328 (in Homo).

spatulatum (endemicum) Anders, 1903, 6 ed., 1245 (in man).

species Barbagallo & Drago, 1903, 410 (in Mugil cephalus; Catania).

species Barbagallo & Drago, 1903, 410 (in Julis pavo; Catania).

species Barbagallo & Drago, 1903, 410 (in Trachurus trachurus; Catania).

species Duncker, 1881d, 141; 1884a, 39-42, figs. 1-6.

species Heymann, 1905, 96-98, figs. A. B.

species Johnstone, 1907, 186-188, fig. 17 (in Labrus mixtus; Morecambe Bay).

species Kellicott, 1894, 123-126.

species Nicoll, 1907, 70, 92 (in Cottus scorpius Bloch).

species Nicoll, 1907, 71, 91-92, pl. 3, fig. 12, pl. 4, fig. 13 (in Cottus bubalis)

species Nicoll, 1907, 71, 92 (in Gobius ruthensparri).

species Nicoll, 1907, 72 (in Pleuronectes microcephalus).

species Sons., 1890, 105, t. h. Python molurus.

sphærostomum Schlotthauber, 1860, 130 (in Corvus caryocatactes).

sphærula Looss, 1896b, 81–86, pl. 6, figs. 59–60 (in Rhinolophus hippocrepis Bonap.; Ghizeh); 1898, 454, 456, 457, figs. I, VI; 1899b, 547.—Stiles, 1901,

spiculator Duj., 1845a, 424-425 (in Mus decumanus; Rennes) to (Echinost.).—
Braun, 1901e, 340.—Cobbold, 1879b, 316.—Dies., 1850a, 382.—Erc., 1881e,
15; 1882a, 251.—Linst., 1886, 128-131, pl. 9, figs. 28-29; 1890, 184.—Stoss., 1892, 28 (to Echinost.).

 spiculigerum Mueh., 1898, 18–19 (in Fuligula nyroca); 1898, 4, 26, 97–100, fig.
 18.—Braun, 1902b, 11, 15 (to Psilost.), 16 (syn. of D. oligoon), 18, 155.— Looss, 1899b, to Psilost.

spiniceps Looss, 1896b, 114-118, pl. 8, figs. 79-80 (in Bagrus bayad; Cairo); 1899b, 578, 581 (type of Acanthost.).—Braun, 1899, 630; 1901b, 34.

spinosum Linst., 1880, 51 (in Sylvia rufa).—Braun, 1892a, 696, 733.—Looss, 1894a, 175.—Luehe, 1899, 531 (thinks this is perhaps D. cirratum).—Stoss., 1892, 162.

spinulosum Rud., 1808a, 458; 1809a, 425 (in Larus cinerarius, L. nævius, Colymbus septemtrionalis; Greifswald, July, August) to (Echinost.); 1819a, 116 (in Colymseptemtrionalis; Greitswald, July, August) to (Echinost.); 1819a, 116 (in Colymbus cristatus; Mus. Vien.), 419.—Baird, 1853a, 56.—Bellingham, 1844a, 426.—Braun, 1892a, 575; 1893a, 879.—Crep., 1837, 316; 1846, 141, 144, 146.—Dies., 1850a, 392; 1858e, 350, 351 (in Numenius arquatus; Anas clangula; Larus capistranus; L. ridibundus; Podiceps cristatus).—Duj., 1845a, 430.—Linst., 1877, 183, pl. 13, fig. 14.—Mol., 1858, 130–131; 1861, 220–221, pl. 3, fig. 6, pl. 4, fig. 2.—Mueller, 1897, 20–21, pl. 3, fig. 4.—Nord., 1832a, 90, 102.—Olfers, 1816, 47.—Stoss., 1892, 64 (in Larus ridibundus; Triest); 1892. 169 (to Echinost.).—Reported for Anas boschas fera, A. marila, A. querquedula, Carbo graculus, Ceryle rudis, Colymbus cristatus, C. septentrionalis, Halieus graculus, Uria gravlle). graculus, Uria grylle).

spinulosum Hofmann, 1899a, 184, 193, 201.—Braun, 1899, 492 (in Erinaceus europæus) to Harmost.; 1900, 12; 1901e, 338 to Harmost., 341.—Looss, 1899b, 689.

"spinulosum" of Mol., 1861, pl. 4, fig. 2 (is not E. spinulosum, see Looss 1899b, 689, but perhaps a Stephanost.).

spirale (Dies., 1850) Brand., 1892, 507.—Braun, 1899, 631-632; 1901b, 54.—Stoss., 1895, 232 (in Peltocephalus tracaxa, Testudo tabulata, Hypsilopus tubercu-

spirale Fil., see Par., 1896, 2 (in Dentex vulgaris).

squamata Kerbert, 1881a, 556 (in Rana temporaria) for squamula.

squamosum (Villot, 1878) Brand., 1890a, 577.—Braun, 1892a, 569, 735.

squamula Rud., 1819a, 103, 391 (in Mustela putorius; Mus. Vien.).—Baird, 1853a, 52.—Brand., 1892, 506.—Braun, 1890a, 439; 1892a, 567, 568, 571, 628, 636, 637, 638, 640, 641, 642, 647, 648, 655, 678; 1893a, 866, 871, 880.—Bremser, 1824, pl. 9, figs. 9–10.—Crep., 1837, 326; 1839, 288.—Dies., 1850a, 321 (to Monost.) (in Mustela putorius; Toulouse).—Duj., 1845a, 406, type of Eurysoma.—Fraip., 1880a, 398; 1880b, 106; 1880c, 419, 420–429, 430, 432, 433, 441, 442, 443, pl. 18, figs. 1–17 (in Rana temporaria, larva; Mustela putorius, adult); 1881b, 1, 3, 4, 14, 1883a, xyxyii —Gamb, 1896, 72 1881b, 1, 3, 4, 14; 1883a, xxxvii.—Gamb., 1896, 72.—Jackson, 1888, 645.— Kampmann, 1894b, 446, 448.—Levin., 1881a, 75.—Linst., 1873, 99.—Looss, 1885b, 20.—Maddox, (1867a), 87–99 (in Morrhua æglefinus).—Mont., 1888a, 7, 41, 43, 45; 1893, 62.—Poir., 1885, 102.—Stiles & Hass., 1898a, 88, 98 (type of Eurysoma).—Stoss., 1892, 20 (in Putorius putorius; Putorius vulgaris; Vienna, Tolosa).—Zeller, 1867, 21 Mar., 213–220, pl. 13, figs. 1–4 (in grass frog).—Also reported for Fœtorius putorius.

sternæ cantiacæ Dies., 1858e, 355 (in Sterna cantiaca), based on LaValette, 9, 37, pl. 1, figs. 15, 1. 2., and Moul., 1856, 102.—Stoss., 1892, 186.

stossichii (Mont., 1891) Braun, 1891d, 423 (=D. ocreatum Mont.) (in Clupea pilchardus); 1893a, 911.

stridulæ Reich, 1801, 371–386 (in Strix stridula).—Brand., 1888a, 9.—Dies., 1850a, 386.—Pulteney, ——, 371.—Rud., 1809a, 423 (renamed D. apiculatum).

sturionis Rud., 1809a, 435 (in Acipenser sturio; Arimini); 1819a, 118 (syn. of D. hispidum).—Baird, 1853a, 57.—Dies., 1850a, 392 (syn. of D. hispidum).

subclavatum (Goze, 1782) Zed., 1800a, 164 (in Rana), 185–188.—Ben., 1858a, 1861a, 81, 82 (to Amphist.).—Brand., 1888a, 9.—Dies., 1850a, 318 (to Diplodiscus).—Rud., 1809a, 348.

subflavum Sons., 1892, 91–92 (in Zamenis viridiflavus Lacep.); 1893, 184.—
 Braun., 1893b, 185 (in Z. virid.).—Stoss., 1895, 216 (in Z. virid.; Pisa).—Volz, 1899, 235, 237.

subtriquetrum Nord., 1840, 616 [the sequence of names is D. amphist., D. subtriquetrum, apparently a lapsus for Amphist. subtriquetrum].

sulcatum Linst., 1883, 309, pl. 9, fig. 51 (in Perdix graca; Turkestan); 1886, 33.—
 Braun, 1901, 941; 1902b, 119.—Stoss., 1892, 145 (to Cladocœlium).

suspensum Braun, 1901g, 948 (in Corvus sp.); 1902b, 11, 147.

sygnoides Nord., 1840, 617 for D. cygnoides.

tacapense Sons., 1894, 111–112; 1894, 2–6, 7 (in Ch. vulg., Rana esculenta, Bufo sp.; Gabes in Tunis) to (Brachycœlium); 1895, 124 (in Chamæleo vulgaris).—Looss, 1896b, 86–97, pl. 6, figs. 61–62, pl. 7, fig. 63 (in caméléon; Alexandria); 1898a, 460, 461, fig. 2; 1899b, 616, 617, 622 (syn. of D. medians Olss., based on reexamination of original slides).—Stoss., 1895, 215 (in Ch. vulg.; Tunisia).

talpæ Viborg, 1795, 242.—Rud., 1809a, 331, 390.

tarda Steenstrup, 1842, 75–76, pl. 3, figs. 1–6 (Cerc. armata Sieb., renamed) [name not mentioned in text].—Dies., 1850a, 298 (p. 138 of Steenstrup as syn. of Cerc. armata), 418 (p. 138, pl. 3, fig. 5e–g of Steenstrup as ? syn. of Heptast. hirudinum); 1855a, 381, 388 (syn. of Cerc. (Xiphidioc.) armata); 1858e, 366 (of 1850a, 418, syn. of Tetracotyle typica, new name.—Erc., 1881e, 21, 48, pl. 2, fig. 6; 1882a, 257, 284.—Fil., 1854a, 10, 15, 22.—Moul., 1856a, 150, 151, 201, 215, 226, 227, 230 (in Lymnæus stagnalis, Planorbis corneus).

tartinii Stoss., 1899, 6, pl. 1, fig. 13 (in Oblata melanura; Triest).

tectum Linst., 1873, 104, pl. 5, fig. 4 (in Osmerus eperlanus).—Odhn., 1905, 352, 353 (=crenata 1802, type of Brachyphallus).—Stoss., 1886, 55.

tenere Looss, 1898, 461 ("D. tacapense Sons," of Looss, 1896, 86, misdetermined) to Pleurogenes.

tenerum Looss, 1899b, 616 to (Pleurogenes), 622.

tenue Lint., 1898c, 535–536, pl. 52, figs. 2–8 (in Roccus lineatus); 1900, 294; 1901, 415, 455, 468, 469, 479; 1905, 328, 334, 365, 370, 373, 374, 376, 379, 386, 391, 396, 399 (in Caranx hippos, Centropristes striatus, Coryphæna equisetis, C. hippurus, Cynoscion nebulosus, Menticirrhus americanus, Micropogon undulatus, Orthopristis chrysopterus, Pomatomus saltatrix, Sciænops ocellatus).—Odhn., 1905, 331.

tenue tenuissime Lint., 1898c, 536-537, pl. 52, figs. 9-12 (in Morone americana); 1901, 415, 421, 456 (in Roccus lineatus, Opsanus tau).—Vaullegeard, 1901, 146.

tenuicolle Rud., 1819a, 93, 365–366 (in Phoca barbata, liver).—Braun, 1893, 353; 1893a, 875; 1893f, 426, fig. 4; 1901e, 314–315, 338; 1903, 3. ed., 157 (of Mueh., 1896, p. p., syn. of Opisthorchis felineus).—Cobbold, 1860a, 6–7; 1879b, 313.—Dies., 1850a, 336–337.—Duj., 1845a, 440, 444.—Jægers., 1898, 6.—Looss, 1899b, 530, 674.—Mueh., 1896, 589 (D. felineum as syn.); 1896, 257–262, figs. 4, 11; 1898, 16, 24, 87, 88, 89, 90, 95.—Odhn., 1905, 347 (Rud. of Olss).—Olss., 1893, 9.—Stiles & Hass., 1894e, 428.—Stoss., 1892, 18–19 (in Phoca barbata).—Ward, 1903, 869 (syn. Op. fel.).—Reported for Erignathus barbatus, Delphinus phocæna, Felis domestica, Halichærus grypus, Phocæna phocæna.

tenuissime (Lint., 1898) Vaullegeard, 1901, 145 (=D. tenue tenuissime).

tereticolie (Rud., 1802) Rud.. 1809a, 379–381, 405 (in Esox lucius, Perca lucioperca, Salmo trutta); 1819a, 102, 122, 386, pl. 2, fig. 5, 600.—Ben., 1852, 24–29, 33, pl. 2, figs. 1–3; 1858a, 1861a, 98, 104, 174, 177, 178, 186, 189, 193, 206, pl. 8,

figs. 1-17 (syns. D. rosaceum, D. lucii, Fasc. longicollis, F. lucii, Plan. lucii); 1870c, 140, 141, 142.—Bettend., 1897a, 31; 1897, 335.—R. Bl., 1888a, 544, 545, 547.—Brand., 1888a, 41; 1890a, 570.—Braun, 1883a, 70; 1892a, 568, 574, 575, 644, 653, 727, 745, 747, 757, 761, 763, 768, 778, 779, 780, 781, 782, 784, 786, 787, 788, 789; 1893a, 872, 873, 879, 883, 911; 1901b, 33; 1901e, 314, 315, 338.— Bremser, 1824c, pl. 9, figs. 5–6.—Cobbold, 1860a, 21.—Crep., 1837, 310, 311, 313, 314, 316, 317, 318, 321, 322, 323, 325.—Creutzberg, 1890a, 21.—Dies., 1850a, 358 (syns. D. luzii, D. truncatum Abildg., F. longicollis, F. lucii, Plan. lucii); 1858d, 272 (larva? Rhopalocerca tardigrada); 1858e, 340 (in Esox lucii); 1858d, 272 (larva? Rhopalocerca tardigrada); 1858e, 340 (in Esox estor); 1859c, 429.—Duj., 1845a, 419-420.—Fil., 1855b, 24, 25; 1857c, 29.—Fischer, 1840, 158.—Fraip., 1880c, 417, 443.—Harz, 1881c, 5 (syn. of D. lucii Zed.).—Hausmann, 1897b, 4, 6, 14, 15, 17, 20, 21, 22 (in Esox lucius, Thymallus vulgaris).—Jackson, 1888, 649.—Johnston, 1902, 329.—Jurine, —, 489.—Kowal., 1894. 2; 1894, 220; 1895, 372-390, pl. 8, fig. 10.—Kœlliker, 1843, 99-101, 130, 135, 137.—Kroyer, 1838-40a, 41, 615; 1843-45a, 644; 1846-53a, 253 (in Esox lucius, Lucioperca sandra, Salmo fario, S. trutta).—Kuech., 1855, 187.—Lander, 1904a, 19.—Leidy, 1851b, 206; 1856b, 44; 1904a, 48, 87 (terreticolle).—Leuck., 1863a, 457, 459, 460, 485, 490, figs. 147, 148, 159.—Looss, 1893b, 813, 815; 1894a, 2, 5-18, 20, 22, 25, 28, 29, 31, 109, 113, 115, 116, 117, 118, 120, 124, 125, 136, 137, 142, 144, 147, 148, 149, 150, 151, 152, 153, 154, 159, 165, 166, 167, 176, 179, 181, 184, 185, 187, 188, 189, 197, 200, 202, 205, 206, 212, 215, 219, 241, 245, 270, 272, 275, 276, pl. 1, figs. 1-3, pl. 3, figs. 53-63, pl. 206, 212, 215, 219, 241, 245, 270, 272, 275, 276, pl. 1, figs. 1-3, pl. 3, figs. 53-63, pl. 4, figs. 64-71, 74, pl. 6, fig. 121 (syns. D. lucii, D. rosaceum, Fasc. lucii, Plan. lucii) (in Esox lucius, Lota vulgaris, Lucioperca sandra, Salmo fario, S. hucho, 180 Alpinus, S. trutta, S. umbla, Trutta variabilis); 1894, 17; 1896b, 109; 1899b, 548, 549, 570; 1902m, 456, 457.—McIntosh, 1864, 150.—Mont., 1888a, 25, 27, 32, 43, 68, 69, 72; 1892, Oct. 7, 185, 187, 191; 1893, 16, 60, 115; 1896, 152, 153.—Moul., 1856a, 25, 31, 32, 49.—Mueh., 1898, 30.—Nord., 1840, 617, 192, 193.—Moul., 1890a, 25, 31, 32, 49.—Muen., 1898, 30.—Nord., 1840, 617, 619, (syn. of Fasc. longicollis Bloch).—Ofenheim. 1900, 152.—Olfers, 1816, 45.—Olss., 1876, 18; 1893, 11.—Poir., 1885, 102.—Schauinsland, 1882, 494, 496, 497,—Sieb., 1835, 59, 66, 82; 1836, 233, 237.—Sons., 1891, 264 (in Esox lucius).—Sramek, 1901, 96, 105, figs. 58-59 (syns. Fasc. lucii Mueller, F. longicollis Bloch, F. teret. Rud., Plan. lucii Gœze, D. lucii Zed.).—Stoss., 1886, 25; 1890, 50; 1891, 216; 1898, 38; 1904, 199.—Tennent, 1906, 670.—Wagener, 1857, 25, 28, 45, pl. 20, figs. 1-5 (in Esox lucius).—Wolf, 1903, 610.—Also reported for Esox reticulatus.

tereticolle lucii Mayer, 1841a, 18.

tereticolle rosaceum (Nord., 1832) Dies., 1850a, 364.

tereticollis Blainv., 1828a, 585 (for Fasc. tereticollis Rud.).

teretiusculum Mont., 1893, 40, 41, 42, 43, 82, 83, 94, 96, 98, 102, 118, 193, pl. 1, fig. 4; pl. 6, fig. 66 (in Solea klenii).—Jacoby, 1900, 4.

tergestinum Stoss., 1889, 28, pl. 13, fig. 57 (in Oblata melanura; Triest); 1898, 44.— Braun, 1892a, 579.—Mont., 1893, 84, 86, 95.

terreticolle Leidy, 1904a, 48, 87, for tereticolle.

testudinis Rud., 1819a, 121 (in Testudo orbicularis; C. E. V.).—Braun, 1899, 630.—Dies., 1850a, 325 (syn. of Monost. delicatulum).—Duj., 1845a, 451.— Stoss., 1895, 232 (in Emys orbicularis; Vienna).

testudinis midæ Braun, 1899, 629 (for mydæ).

testudinis mydæ Rud., 1809a, 433, for D. intestinalis testudinis mydæ.—Braun,

1899, 629 (midæ).

tetracystis Gastaldi, 1854, 4–5, pl. 1, figs. 1–3 (in Rana esculenta).—Braun, 1893a, 871.—Cobbold, 1860a, 17.—Dies., 1855, 390; 1858d, 253 (larva=Cerc. (Acanthocephala) microcotyla); 1858e, 348 (stat. juven. in Pelophylax esculentus; larva in Paludina achatina; P. vivipara).—Fil., —, 30, pl. 1. fig. 9.— Linst., 1873, 1 (larva=Cerc. microcotyla Fil.).—Staff., 1900, 405, 410–412.— Stoss., 1889, 68 (in Rana esculenta; Torino).—Also reported for Rana cates-

tetracystis ranæ esculentæ Dies., 1855, 64, footnote 9. See tetracystis.

texanicum Francis, 1891c, 135–136, pls. 3–8, 5 figs. (in Bos taurus; Texas); 1892, 426.—Huber, 1896a, 576 (syn. of D. magnum).—Leuck., 1892b, 797, 798 (texicanum).—Stiles, 1892e, 148; 1892g, 732–733; 1898a, 51.—Ward, 1895, 253 (syn. of Fasc. magna), 332 (in Bos taurus), 335 (in Ovis aries), 338 (in Equus caballus); 1903, 866 (syn. of Fasc. magna).

texicanum Leuck., 1892b, 797, 798 (for texanicum).

tincæ (Modeer, 1790) Rud., 1809a, 366.

tobiani Krœyer, 1846-53a, 592 (in Ammodytes tobianus).

todari delle Chiaje, 1841, 139.

tornatum Rud., 1819a, 684-685 (in Coryphæna equiselis; C. hippuris).—Braun, 1893a, 872.—Buttel-Reepen. 1902. 167, pl. 6, fig. 8.—Cobbold, 1860a, 28 (in Megalops cyprinoides); 1879b, 458, 461.—Cohn, 1902k, 47.—Dies.. 1850a. Megalops cyprinoides); 1879b, 458, 461.—Cohn, 1902k, 47.—Dies. 1850a, 372–373 (syns. F. caudata, F. coryphænæ, F. coryphænæ hippuridis, D. coryphænæ); 1859c, 431.—Duj., 1845a, 421–422.—Johnston, 1901, 337.—Lint., 1898c, 513–514, pl. 42, figs. 6–12; 1901, 415, 418, 442, 444, 452, 455, 469, fig. 310 (in Coryphæna hippurus, Fundulus heteroclitus, Menidia notata, Roccus lineatus); 1905, 328, 334, 355, 356, 372, 374, 398, 399, figs. 156–158 (in Coryphæna equisetis, C. hippurus, Menticirrhus americanus, Synodus fœtens, Tylosurus marinus).—Looss, 1896b, 125 to (Apoblema); 1899b, 640.—Mont., 1891, 497.—Stoss., 1886, 12.—Wagener, 1860, 166, 176–178, 181, pl. 9, figs. 1-5 (in Coryphæna sp.).

torosum (Setti, 1897) Linst., 1903. 354.

torulosum Rud., 1814a, 104-105 (in Silurus glanis); 1819a, 111 (in S. glanis: Greifswald, October), 410.—Cobbold, 1860a, 28.—Dies., 1850a, 375.—Dij.. 1845a, 464.—Kroyer, 1846-53a, 137 (in S. gl.).—Olfers, 1816, 46.—Stoss., 1886, 56.

totari E. Bl., 1847a, 309, for todari.

trachea (Montagu, 1811) Rud., 1819a, 114 (in Phasianus gallus: Gryphiæ), 798 (sub D. lineare).

tranversale (Rud., 1802) Rud., 1809a, 361–362 (in Cobitis fossilis); 1819a, 95, 368.— Braun, 1893a, 873.—Cobbold, 1860a, 22 (in Acanthopsis (Cobitis) fossilis, Cobitis pæmiæ).—Dies., 1850a. 339.—Duj., 1845a, 463–464.—Kroyer, 1846–53a, 563, 573 (in Cob. fos., Botia tænia).—Odhn., 1901, 484, 505.—Pavesi, 1881, 616.—Stiles & Hass., 1898a, 95.—Stoss., 1886, 56.

trapezium Leidy, 1891a, 414 (in Pandion carolinensis): 1904a, 235.—Stiles & Hass., 1894e, 414.

triangulæ Linst., 1878, 72, for triangulare.

triangulare Dies., 1850a, 351 (in Merops apiaster) (syn. D. meropis).—Braun, 1901, 561, 568; 1902b, 51, 52, 53, 93 (to Plagiorchis).—Cobbold, 1860a, 13.—Linst., 1878, 92 (triangulæ).—Looss, 1899b, 531, 631, 725 (type of Megacetes); 1900, 602 (type of Eumegacetes).—Stoss., 1892, 186; 1904, 2

tricolor Stiles & Hass., 1894h, 160–162, fig. 2; 1895a, 729–737, pls. 1–2, figs. 1–8 (in Lepus sylvaticus, L. americanus; Maryland); 1895c, 700–701.

trifolium Braun, 1901g, 947 (in Ardea cocoi; Brazil); 1902b, 150-151, figs. 92. 93. triganocephalum Kampmann. 1894b. 454, 457, misprint for trigonocephalum.

triglæ Rud., 1819a, 122 (in Trigla cuculus; C. E. V.).—Dies., 1850a, 399.

triglæ gurnardi Rathke, 1799. 68, 146, fig. 2a-c (in Trigla gurnardus).—Dies., 1850a, 362.—Rud., 1809a, 384 (syn. of D. soleæforme).

triglæ pini Dies., 1855, 64; 1858e, 343 (renamed D. homæostomum), based on Bellingham, 1844, 428.

trigonocephalum (Rud., 1802) Rud., 1809a, 415–418 (in Ursus melis, Erinaceus europæus, Mustela putorius, M. vulgaris) (includes Plan. putorii; P. melis; Fasc. putorii; F. melis: Dist. melis: F. armata; D. armatum); 1810a, 376; 1819a, 114.—Baird, 1853a, 55.—Bellingham, 1844a, 425.—Blainv., 1824a, 512 (type of festucaire).—Brand., 1891b, 265; 1891d, 16.—Braun, 1891, 100; 1892a, 618, 754, 784, 785, 786, 787, 797; 1893a, 866, 879, 911.—Cobbold, 1879b, 295, 298 (in Erinaceus europæus, Mustela vulgaris), 299.—Crep., 1837, 310, 295, 298 (in Erinaceus europæus, Mustela vulgaris), 299.—Crep., 1837, 310, 311, 317, 326; 1845, 327, 337, 349b.—Dies., 1850a, 381–382 (syns. D. armatum; Fasc. armata; F. melis; F. putorii; Plan. melis; P. putorii); 1858d, 267 (larva=Histrionella ephemera Ehrenberg); 1858e, 344 (in Erinaceus europæus; Mustela putorius).—Duj., 1845a, 423.—Fil., 1855b, 24.—Gamb., 1896, 72.—Giebel, 1857, 265.—Kampmann, 1894b, 454, 457 (triganocephalum).—Kowal., 1896d, (3) 253 (in Putorius fectidus; Dublany).—Leuck., 1863a, 460, 471.—Linst., 1873, 101, 106 (larva=Cerc. echinatoidcs Fil.) in Fœtorius putorius; 1879, 185, pl. 12, fig. 31.—Looss, 1885b, 13, 18, 21, 37, 38, 50, pl. 23, figs. 6, 17; 1894a, 227; 1902m, 804, 805.—Mol., 1858, 129-130;

1859, 825, 846 (in Erinaceus europæus; Padua); 1861, 213–216, pl. 3, fig. 2.—Mont., 1888a, 40, 44, 65; 1893, 75, 77.—Nord., 1840, 621 (to Fasc.).—Olfers, 1816, 46.—Rail., 1893a, 365.—Schuberg, 1895, 178.—Sieb., 1854, 23.—Sons., 1893, —..—Stoss., 1890, 51; 1892, 29–30 (to Echinost.).—Wagener, 1857, 24, 42.—Reported for Canis vulpes; Felis catus domesticus; Fœtorius lutreola; F. putorius; F. vulgaris; Lutra vulgaris; Meles europæus; Meles taxus; Mustela foina; Mustela martes; M. putorii; M. vulgaris; Plecotus communis; Putorius vulgaris; Ursus meles; Paludina vivipara.

trilobum Rud., 1819a, 104–105, 392 (in Pelecanus carbo; Mus. Vien.).—Dies.,

1850a, 310 (to Hemist.).—Duj., 1845a, 449.

tringæ helveticæ Rud., 1819a, 120 (in Tringa helvetica; C. E. V.).—Dies., 1850a, 386 (syn. of D. cinctum).—Duj., 1845a, 447.—Stoss., 1892, 187 (in Tr. helv.; Vienna).

troncatum Perroncito, 1882, 284, for D. truncatum [Erc.].

truncatum (Abildg., 1806) Rud., 1814a, 105–106; 1819a, 122 (in Perca lucioperca).—Dies., 1850a, 358 (?syn. of D. tereticolle).—Kroyer, 1838–40a, 41, 579 ("of Leuck.," in Lucioperca sandra Cuv.).—Nord, 1832a, 88.

truncatum (Rud., 1819) Rail., 1886, 296–297; 1893a, 364 (syns. Dist. conus Crep., teatum (Rud., 1819) Rail., 1886, 296–297; 1893a, 364 (syns. Dist. conus Crep., 1825, [not Gurlt, 1831]) to (Brachylaimus).—Askanazy, 1901, 72.—Braun, 1893e, 349, 350, 352, 353, 354; 1893f, 384, 385, 387, 389, 390, 392, 424, 425, 427 (syns. D. conus Crep., D. campanulatum Erc.); 1893g, 802, 803 (syn. D. campanulatum Erc., 1875); 1894g, 129; 1894i, 606.—de Jong, 1896a, 2, 3, 4, 5, 6, 7, fig. 3; 1897a, 245–246.—Kholodk., 1898, 29.—Looss, 1896b, 97; 1899b, 565 to (Metorchis).—Moniez, 1896, 137 to (Dicrocælium), 139 (=conus Crep.), 140, 141.—Mueh., 1898, 15, 24, 25.—Neumann, 1892, 529; 1892, 544.—Pease, 1898, 82.—Perroncito, 1886, 250.—Ratz, 1898, 68; 1900, 141 (in cat).—Stiles & Hass, 1894e, 421–434 pl. 3, figs. 11–12 to (Dicrocælium) (in Phoca, vitulina, 1895, 1894e, 421–434 pl. 3, figs. 11–12 to (Dicrocælium) (in Phoca, vitulina, 1895). Hass., 1894e, 421–434, pl. 3, figs. 11–12, to (Dicrocœlium) (in Phoca vitulina, Felis catus dom., Canis fam., C. vulpes, Halichærus fætidus, Gulo borealis; Germany, Holland, Italy, France) (syns. D. conus Crep.; D. lanceolatum of Mehlis, of Dies., 332; D. campanulatum Erc.).—Ward, 1895, 243 (in part, syn. of D. felineum Riv.), 341 (in Canis fam.).—Zwaardemaker, 1890, 197–203, pl. 3.

truncatum Erc., 1859a, 382 (in Canis familiaris).—Braun, 1893a, 875; 1893e, 349 ("Ercolani 1846"), 351; 1893f, 386, 424 ("Ercolani 1846, in dog; Bologna), 425, syn. of D. truncatum (Rud.).—de Jong, 1887a, 57; 1896a, 6, 7, 11, figs. 1a, 1b.—Leuck., 1889, 357.—Perroncito, 1882, 284 (troncatum) in dog.—Sons., 1889, 276, 277, 280; 1889, 281 (syn. of D. conus Crep.).—Stoss., 1892,

25 (syn. of D. conus Crep.).

truncatum Leuck., 1842, 34–35, pl. 1, fig. 8a-b (in Sorex fodiens).—Braun, 1893a, 876; 1893d, 467; 1901e, 342.—Cobbold, 1860a, 8; 1879b, 296.—Dies., 1850a, 374 (in Sorex daubentonii).—Duj., 1845a, 438 (in S. fodiens).—Sons., 1889, 276 (in S. fod.).—Stoss., 1892, 36–37 (in Crossopus fodiens).

truttæ Dies., 1858e, 356 (in Salmo trutta), refers to Moul., 1856, 217.—Braun. 1893a, 871.

tubarium Rud., 1819a, 111-112, 410 (in Sciæna umbra; Spezia, August).—Carus, 1884, 125.—Cobbold, 1860a, 28.—Dies., 1850a, 375.—Duj., 1845a, 457.—Stoss., 1886, 56 (in Umbrina vulgaris).

tubiporum Braun, 1900, 388–389 (in Vespertilio sp.); 1900, 232–233, pl. 10, figs. 8, 9. tubulatum Rud., 1819a, 675–676 (in Muræna sp.; Brazil).—Cobbold, 1860a, 22.—Dies., 1850a, 337.—Duj., 1845a, 468.—Stoss., 1886, 56.

tumidulum Rud., 1819a, 95, 369 (in Syngnathus hippocampus; Vienna).—Bellingham, 1844a, 423.—Ben., 1870, 89, pl. 5, fig. 5.—Carus, 1884, 129.—Cobbold, 1860a, 22.—Dies., 1850a, 339–340; 1858e, 333 (in Syngnathus acus; Ireland).— Duj., 1845a, 469.—Kroyer, 1846-53a, 704 (in Siphost. acus L.).—Odhn., 1901, 484, 503.—Pag., 1862, 305.—Stoss., 1886, 56 (in Hippocampus guttatus, Syngnathus acus).

turdi Rud., 1819a, 120 (in Turdus saxatilis; C. E. V.).—Dies., 1850a, 361 (syn. of

D. mesostomum), 391 (syn. of D. nephrocephalum).

turgidum Brand., 1888, 247–251, pl. 17, figs. 2–3 (in Rana esculenta).—Braun, 1892a, 568, 578, 579, 584, 700, 720, 721, 727, 737; 1893a, 877, 881, 893; 1895b, 11; 1900h, 6.—Galli-Valerio, 1898n, 373.—Kowal., 1894, 3.—Looss, 1892, 101; 1894a, 1, 173; 1899b, 623, 775 (type of Brandesia).—Mont., 1893, 84, 85.— Mueh., 1898, v. 1 (1), 23, 30, 102-105, 114, fig. 21, 28.

tursionis Marchi, 1873, 304, pl. 5, fig. b (in Delphinus tursio).—Carus, 1884, 125.—
 Linst., 1886, 125.—Par., 1896, 1–3 (syn. of D. (Dicrocœlium) longissimum Poir.).—Poir., 1886, 30.—Stoss., 1892, 37 (in Del. tur.).

umblæ (Fabricius, 1780) Zed., 1803a, 212.—Dies., 1850a, 343 (syn. of D. seriale) (in Salmo umbla; Greenland; kidneys).—Rud., 1809a, 369.

umbrinæ Stoss., 1885, 159, pl. 4, fig. 18 (in Umbrina cirrhosa; Triest); 1886, 26;
1887, 91; 1898, 36–37.—Barbagallo & Drago, 1903, 410 (in Umbrina cirrhosa; Catania).—Braun, 1892a, 728, 737.—Lint., 1900, 290.—Luehe, 1900, 487.—Mont., 1893, 94.

uncinatum Zed., 1803a, 221 (in Fulica chloropus).—Ben., 1858a, 1861a, 85.—Braun, 1891, 101; 1893a, 874.—Crep., 1837, 316.—Dies., 1850a, 384 (syns. D. chloropodis, Fasc. crenata) (in Gallinula chloropus).—Duj., 1845a, 428.—Mueller, 1897, 22-23, pl. 3, fig. 6.—Olfers, 1816, 46.—Rud., 1809a, 420-421 to (Echinost.); 1814a, 102; 1819a, 115, 417-418.—Sieb., 1854, 23.—Stoss., 1892, 172 (syn. of Echinost. cinctum); 1898, 53.—Also reported for Ascolopax gallinago.

unicum Looss, 1896b, 44–49, pl. 3, figs. 20–24 (in Trionyx nilotica; Cairo, Egypt); 1898, 461 (renamed D. reniferum Looss); 1899b, 590 (=renifera, type of

Astia).—Luehe, 1899, 531.

unicum Mol., 1859, 835–837, pl. 3, fig. 1 (in Centrolophus pompilius; Padua).— Braun, 1893a, 910.—Carus, 1884, 124.—Luehe, 1900, 488, 492.—Stoss., 1886, 17.

urcatum Luehe, 1900, 489 (for D. furcatum).

vagans Leidy, 1850, 304–309, pl. 43, figs. 2–16 (in Helix alternata, H. albolabris) (D. helicis Leidy, 1847, renamed); 1857, 44 (syns. D. helicis Leidy; D. pericardium Crep.; Cercariæum helicis alternatæ Dies.; C. vagans Dies.).—Ben., 1858a, 1861a, 175, 186, 205.—Dies., 1855a, 398, (to Cercariæum).—Fraip., 1880c, 417.—Mont., 1888a, 40.

valdein flatum Stoss., 1883, 114, pl. 1, fig. 4 (in Gobius jozo; Triest); 1886, 35.—
Braun, 1893a, 871.—Carus, 1884, 127.—Lint., 1898c, 527–528, pl. 47, figs. 10–14, pl. 48, figs. 1–2; 1901, 416 (in Alutera scheepfil, Menidia notata, Spheroides maculatus), 421, 444, 464; 1905, 328, 334, 359, 366, 372, 379, 386, 393, 396, 400, 401, 409, 414 (in Cynoscion nebulosus, Leiostomus xanthurus, Menticirrhus americanus, Micropogon undulatus, Monacanthus hispidus, Opsanus tau, Orthopristis chrysopterus, Paralichthys albiguttus, Rachycentron canadus, Siphostoma fuscum; Trachinotus carolinus).—Linst., 1903, 354 (syn. of D. cesticillus Mol.).—Looss, 1894a, 252 (syn. of Echinost. cesticillus); 1899b, 697 (young of Stephanost. cesticillus).—Mont., 1891, 121; 1893, 61, 158, 160, 161.—Nicoll, 1907, 69 (valdemflatum).

valdemflatum Nicoll, 1907, 69 (for valdeinflatum).

validum Linst., 1886, 124–128, figs. 23–27 (in Delphinus sp.; South Atlantic);
1889a.—Braun, 1892a, 591, 603, 608; 1893a, 873.—Buttel-Reepen, 1902, 169,
171, 172, pl. 6, fig. 24.—Stoss., 1892, 15–16 (in Delphinus sp.; Atlantic).

vallei Stoss., 1896, 129, fig. 2 (in Falco subbuteo); 1898, 33.—Looss, 1902m, 746.
variabile Leidy, 1856, 44 (in Tropidonotus sipedon).—Braun, 1893a, 876.—Dies., 1858e, 352-353.—Luehe, 1900, 559-561.—Pratt, 1903, 31.—Stoss., 1895, 232-233 (in Trop. sip.; Philadelphia).—Volz, 1899, 234, 235.—West, 1896, 323.

varica Zed., of Ben., 1870, 69, see varicum.

varicum (Mueller, 1784) Zed., 1803a, 217.—Bellingham, 1844a, 424.—Ben., 1870, 25.—Braun, 1892a, 643, 735.—Cobbold, 1860a, 24.—Dies., 1850a, 368; 1858e, 342 (in Salmo salar).—Duj., 1845a, 465.—Hausmann, 1897b, 4, 6, 20, 22 (in Trutta salar).—Juel, 1889, 34.—Kroyer, 1843-45a, 581; 1846-53a, 54 (in Salmo salar L., Thymallus vulgaris Cuv.).—Levin., 1881, 5; 1881a, 54-56, 58, 61, 69, pl. 2, figs. 1-2 (in Cottus scorpius, Gadus ovak; immature forms in Harmothoë imbricata, Cottus scorpius, Gadus ovak; immature forms in Hemiurus; 1901, 438.—Luehe, 1901, 479.—M'Intosh, 1864, 149-150 (of Rud.) (in Salmo salar).—Mont., 1890, 424; 1891, 513; 1893, 43, 61, 85, 88, 95, 106, 107, 180, 182, 183, 184.—Mueller, 1788, 43.—Nord., 1840, 620 (to Fasc.).—Olfers, 1816, 46.—Olss., 1868, 40; 1876, 17.—Rud., 1809a, 396-397; 1810a, 376; 1819a, 106.—Stoss., 1886, 22; 1902, 582.—Reported for Anguilla vulgaris; Conger conger; C. vulgaris; Coregonus wartmanni; C. oxyrrhynchus; Gadus euxinus; G. minutus; Hippoglossus maximus; Labrus maculatus; Lota molva; Merlangus vulgaris; Molva abyssorum; M. vulgaris.

Variegatum Rud., 1819a, 99, 378-379 (in Rana esculenta; Berlin, October).—
Baird, 1853a, 53.—Ben., 1870c, 141.—E. Bl., 1847, to Brachylemus.—Braun, 1897a, 1459; 1892a, 784 (varigatum); 1893a, 876, 881, 911.—Cobbold, 1860a, 17.—Crep., 1829, 58-59; 1837, 310, 322, 326; 1839, 288.—Darr, 1902, 654.—Dies., 1850a, 354-355 (to Brachylemus); 1858e, 339 (in Rana pipiens, Pelophylax esculentus).—Duj., 1845a, 416-417.—Gronkowski, 1902a, 511, 515, 518-519, 531-532 (4, 8, 11-12, 24-25), pl. 13, figs. 3, 11, 12.—Hausmann, 1897b, 27.—Kamensky, 1900a, 8.—Leuck., 1863a, 490.—Leidy, 1851, 207, 242; 1856, 44.—Linst., 1883, 309; 1886, 34; 1887, 97.—Looss, 1892, 65, 93 (varigatum); 1893b, 812, 815, 818, 819, fig. 3; 1894a, 2, 3, 65, 68, 71-82, 113, 114, 115, 116, 118, 119, 120, 121, 125, 127, 135, 136, 146, 150, 159, 163, 167, 173, 175, 179, 182, 190, 193, 199, 204, 208, 209, 210, 212, 214, 215, 217, 221, 224, 226, 227, 230, 242, 256, 266, 268, pl. 2, figs. 43-48, pl. 7, figs. 134-146 (syns. D. cylindricum, D. cylindraceum, Monost. bombynæ, M. ellipticum) (in Rana esculenta, Bombinator igneus; Bufo cinereus); 1896b, 19; 1899b, 556, 661 (type lenta, Bombinator igneus; Bufo cinereus); 1896b, 19; 1899b, 556, 661 (type fenta, Bombinator igneus; Bufo cinereus); 1896b, 19; 1899b, 55b, 661 (type of Hæmatolæchus), 602, 603; 1901, 192; 1902m, 732 (type of Pneumonœces), 810.—Luehe, 1899, 533; 1901, 58.—Mehlis, 1831, 177–179.—Mol., 1859, 828–829, pl. 3, fig. 2 (syn. Brachylemus v.) (in Pelophylax esculentus; Europe).—Mont., 1892, 715.—Mueh., 1898, 26.—Pag., 1857, 41–42, pl. 5, fig. 2 (in green frog).—Poir., 1885, 102.—Sons., 1893, 185, 188, 189 (in Rana esculenta).—Staff., 1900, 405, 409; 1902, 482 (in Canadian frogs and toads); 1902, 24 Nov., 895, 904 (American representatives); 1903, 15 Dec., 901; 1905, 190 July, 50–51.—Stiles & Hass., 1898a, 84; 1902d, 20.—Stoss., 1889, 62; 1896, 128; 1898, 35.—Wagener, 1857, 25, 102, pl. 21, figs. 1–2.—Wright, 1879, 8.—Also reported for Buío vulgaris, Rana halecina, R. temporaria, Triton alpestris.

varigatum Looss, 1892, 93, for variegatum.—Braun, 1892a, 784. [variolosus Gærtner in Pallas (a tunicate)].—Lamarck, 1816, 101.

varium Eysenhardt, 1829, 148-151 (in Gadus merluccius).—Dies., 1850a, 342 (syn. of D. caudiporum), 371 (syn. of D. appendiculatum), 372 (syn. of D. grandiporum and of D. rufoviride).—Mol., 1859, 826 (syn. of D. grandiporum).—Mont., 1891, 498.—Wagener, 1860, 166.

velellæ Filippi (1843), 66, pl. 5, fig. 12 (in Velella spirans; Naples).—Dies., 1850a, 379, 659 (renamed D. megacotyle).—Kælliker, 1849, 53.—Mont., 1888, 198,

199; 1893, 123.

veliporium Johnston, 1902, 329, for veliporum.

veliporum Crep., 1837a, 310 (in Squalus griseus), 318.—Ariola, 1899, 7, 9 (syns. D. insigne Dies., D. microcephalum Baird, D. scymni Risso).—Braun, 1892a, 567, 586, 624, 625, 647, 682, 686, 690, 735; 1893a, 873, 879, 910; 1893d, 466.— Carus, 1884, 129.—Cobbold, 1860a, 25 (in Prionodon milberti, Hexanchus griseus).—Darr, 1902, 666, 667, 669, 683.—Dies., 1850a, 347 (syn. Fasc. squali grisei) (in Prionodon milberti; Hexanchus griseus; Panormi).—Duj., 1845a, 471 (in Squalus griseus).—Jægers., 1900, 72, 73, 74; 1900, 736.—Johnston, 1902, 329 (veliporium).—Kerbert, 1881a, 541.—Linst., 1903, 279, 354.—Lint., 1898, 521-522; 1901, 416 (in Raja lævis), 421, 431.—Lænnberg, 1891, 71; (1898), 4.—Looss, 1894a, 152, 211; 1899b, 570 (thinks this form may belong to Azygia).—Mont., 1888a, 7; 1889, 69; 1889, 70 (in Raja sp.; Porto Huite, Chile); 1889, 321; 1892, 5, 6, 7; 1893, 9; 1893, 33, 34, 36, 43, 52, 82, 102, 107, 113, 120, 122, 147, 191, pl. 1, fig. 6, pl. 8, figs. 122-131.—Ofenheim, 1900, 164.—Olss., 1868, 22; 1876, 13; 1896, 508 (includes D. insigne, D. microcephalum).—Poir., 1885, 22, 37, 42, 44, 48, 49, 53, 74, 82, 106, 127, 142.—Stoss., 1886, 19; 1890, 51; 1901, (9) 97, pl. 6, fig. 1 (in Notidanus griseus; Triest); 1904, 198, 199.—Villot, 1878, 2 (in Hexanchus griseus, Prionodon milberti).—Wagener, 1852, 543; 1857, 28; 1860, 174.—Reported for Acanthias vulgaris, Carcharias milbertii, Centrolophus pompilus, Chimæra monstrosa, Echinorhynus spinosus, Heptanchus cinereus, Hexanchus griseus, Raja grisei) (in Prionodon milberti; Hexanchus griseus; Panormi).—Duj., 1845a, Echinorhynus spinosus, Heptanchus cinereus, Hexanchus griseus, Raja batis, R. clavata, R. fullonica, R. lævis, R. lineata, R. radiata.

ventricosa Ben., 1871a, 68, pl. 4, fig. 11.—Braun, 1891d, 423.—Mont., 1891, 510

(syn. of Apoblema ocreatum Rud.).

ventricosum (Pallas, 1774) Mont., 1893, 26, 27.—Braun, 1893b, 184.—Stoss., 1900,

8.—Reported for Pimelepterus sp.

ventricosum Rud., 1819a, 108, 398 (in Clupea alosa; Arimini, April).—Ben., 1858a, 1861a, 179, 193; 1870, 68.—Bì., 1891, 468-478, fig. 36.—Buttel-Reepen, 1902, 166, 167, 168, pl. 6, figs. 19-22.—Carus, 1884, 125.—Cobbold, 1860a, 27.— Darr, 1902, 666.—Dies., 1850a, 369 (in Alosa vulgaris; Arimini).—Duj., 1845a,

465.—Hausmann, 1897a, 4, 6, 20, 23 (in Al. vulg.).—Jackson, 1888, 647.—Kroyer, 1846–53a, 218 (in Alosa finta Cuv.).—Lander, 1904a, 1.—Mont., 1887, 87; 1891, 12 (syn. of Apoblema appendiculatum), 16, 17 (p. p. syn. of Apoblema ocreatum (Rud.) Mont.); 1891, 496, 497, 498, 499, 500, 501, 502.—Nicoll, 1906, 525 (in Gasterosteus aculeatus).—Sons., 1890, 12–13; 1891, 259 (syn. of D. ocreatum Mol.) (in Alosa sardina).—Stoss., 1886, 11; 1887, 90; 1898, 29; 1902, 582.—Wagener, 1860, 166–172, 173, 175, 179, pl. 8, figs. 1–7 (in Clupeiden).—Also reported for Anguilla vulgaris, Clupea sp., C. sprattus, Motella communis, M. mustela.

ventricosum Stoss., 1898, 29 (syn. of Apoblema ocreatum Rud.).

ventricosum Rud. of Wagener, 1860, 166–172, 173, 175, 179, pl. 8, figs. 1–7 (in Clupeidæ).—Braun, 1891d, 423.—Levin., 1881a, 59 (syn. of D. appendiculatum Rud., Mol.).—Luehe, 1901. 396, 397, 399.—Mont., 1891, 499, 501, 502, 505 (syn. of Apoblema appendiculatum Rud.).

ventricosum minor Shipley, 1900, 540 (in Pimelepterus sp.; New Britain).

vereticolle lucii Rud., 1809a, 440 (misprint for D. tereticolle lucii).

verrucosum Busch, 1851, 100, pl. 15, fig. 11 (in Ophidium barbatum).—Braun, 1893a, 873.—Carus, 1884, 128.—Mont., 1893, 27, 94, 102.—Stoss., 1886, 19 (in Oph. barb.).

verrucosum Lint., 1892, 96-97, pl. 6, figs. 33-35 (in Larus californicus).

verrucosum Mol., 1859, 842–844 (in Labrax lupus; Rhedoni, Padua) (includes D. labracis Duj., 1845a, 398).—Braun, 1892a, 579, 642, 720, 728, 737.—Lander, 1904a, 8.—Odhn., 1901, 513, 514.—Par., 1899, 5; 1902, 4 (syn. of Dicrocœlium labracis).—Stoss., 1886, 31; 1886, 48 (in Labrax lupus; Triest); 1898, 46.

verrucosum Poir., 1885, 10–11, 16, 29, 60, 74, 79, 81, pl. 23, fig. 4 (in Thynnus).—
Braun, 1892a, 576, 608, 609, 873; 1893d, 466.—Jægers., 1900b, 72.—Lander,

1904a, 8.

verrucosum Frælich.—Wolffhuegel, 1900, 136 (for Monost. ver.?).

vespertilionis (Mueller, 1784) Zed., 1803a, 214.—Dies., 1850a, 387 (syn. of D. lima).—Kolenati, 1857, 12.—Rud., 1809a, 427 (syn. of D. lima).—Stoss., 1904, 2.

vexans Braun, 1901, 947 (in Turdus merula); 1902b, 151-152, figs. 94-95.

vibex Lint., 1900a, 269, 281, 291–292, pl. 38, figs. 48–51 (in Spheroides maculatus); 1901b, 416 (in Sph. mac.; Woods Hole, Mass.), 419, 464; 1905d, 328, 334, 402, fig. 188.

viperæ Linst., 1877, 186 (in Pelias berus).—Braun, 1893a, 870.

virgula Fil., 1837a, 338, fig. 5 (in Paludina impura, Valvata piscinalis; Ticini); 1855b, 5.—Dies., 1850a, 296 (to Cerc.).—Moul., 1856a, 161 (to Cerc.).

vitellatum Linst., 1875, 189–190 (in Totanus hypoleucos); 1887, 104.—Braun, 1902b, 50 (to Plagiorchis).—Stoss., 1892, 154 (in Actites hypoleucos; Ratzeburg); 1892, 12, to (Brachylaimus); 1904, 2 (vitellatus).

vitellilobum Olss., 1876, 14 (in Rana temporaria).—Braun, 1893a, 873, 881; 1893d, 466.—Linst., 1887, 97, 98, 102.—Looss, 1894a, 56, 57, 62, 82; 1899b, 606-607 (whether a good species or only a variety of Gorgodera cygnoides can not be determined from Olsson's description); 1902m, 851 (perhaps a Gorgoderina), 857 (to Gorgoderina as possible).—Sons., 1893, 187 (in Rana esculenta).—Stoss., 1889, 67.

vitellosum Lint., 1900a, 269, 282, 290, pl. 37, figs. 38–39 (in Merluccius bilinearis; Woods Hole, Mass.); 1901b, 16, 420, 435, 436, 437, 439, 440, 445, 446, 449, 451; 458, 460, 462, 464, 471, 474, 481, 482, 483, 485, 486, figs. 333–340; 1905d, 329, 335, 348, 351, 357, 366, 369, 378, 382, 385, 388, 390, 393, 396, 397, 399, 401, 404, 405, 409, 413, figs. 176–178.—Johnstone, 1907, 182–185, fig. 15 (in Pleuronectes flesus; Piel).—Reported for Anguilla chrysypa, Bairdiella chrysura, Brevoortia tyrannus, Clupea harengus, Cynoscion regalis, Dasyatis centrura, Decapterus macarellus, Lagodon rhomboides, Leiostomus xanthus, Leptocephalus conger, Limanda ferruginea, Menticirrhus americanus, M. saxatilis, Merluccius bilinearis, Micropogon undulatus, Monacanthus hispidus, Opsanus tau, Orthopristis chrysopterus, Paralichthys albiguttus, P. dentatus, Pomatomus saltatrix, Pomolobus pseudoharengus, Pseudopleuronectes americanus, Prionotus scitulus, P. tribulus, Sciænopš ocellatus, Sarda sarda, Scomber scombrus, Spheroides maculatus, Stenotomus chrysops, Tautoglabrus adspersus, Trachinotus carolinus, Tylosurus marinus).

[vitreum Sars (a tunicate).]

vitrinæ Targioni Tozzetti, 1873, 335-336 (in vitrine).

vitta Duj., 1845a, 418 (in Mus sylvaticus: Rennes) to (Brachvlaimus).—Braun, 1901e, 340.—Cobbold, 1879b, 316.—Dies., 1850a, 396.—Stoss., 1892, 13 (in Mus sylv.; Rennes).

viverrini Poir., 1886, 27–29, pl. 3. figs. 1–3 (in liver of Felis viverrinus).—Braun, 1892a, 699; 1893a, 875; 1893f, 425; 1893, 350, 352, 353.—Looss, 1899b, 675.—Mont., 1888a, 58; 1893, 43, 83, 86, 95, 102, 105, 106, 107.—Stiles & Hass., 1894e, 427–428 to (Dicrocœlium).—Stoss., 1892, 24.

vivipara Ben., 1870, 1871a, 28, pl. 3, figs. 3-4 (in Mugil chelo; Belgium).-Braun, 1891d, 424 (in Mugil cephalus); 1892a, 784.—Hahn & Lefèvre, 1884a, 516 (viviparum).—Looss, 1902, 129, 137.—Sons., 1891, 253, 254–255 (in Mugil cephalus).—Stoss., 1898, 51 (in Mugil chelo; Triest).

viviparæ fasciatæ Linst., 1877b, 186 (in Vivipara fasciata); 1878a, 322.

 viviparum Olss., 1868, 28–29. pl. 4. figs. 73–75 (in Pleuronectes microcephalus;
 Scandinavia).—Braun, 1892a, 785, 786; 1893a, 873.—Looss, 1901, 439, 442.—
 Mont., 1893, 82.—Nicoll, 1907, 82.—Odhn., 1902, 59, 61.—Sons., 1891, 255 (in Pleuronectes microcephalus).—Stoss., 1886, 38.

volvens Nord., 1833, 321, pl. 19, for Diplost. volvens.

vulpina Abildg., 1790, 63-64, pl. 5, figs. 6 a-c (syn. Alaria).—Baird, 1853a, 46.— Brand., 1888a, 60 (syn. of Hemist. alatum).—Dies., 1850a, 308 (syn. of Hemist. alatum).—Nitzsch, 1819, 399 (syn. of Holost. alatum).—Rud., 1809a, 403.

wachniæ Rud., 1819a, 122, 427, based on Tilesius, 1810. 363, 374, pl. 19. figs. 8-10
(in Gadus wachnia).—Cobbold, 1858b, 157 (= D. simplex, D. fulvum).—Dies., 1850a, 398 (in Gadus wachnia).—Rud., 1819a, 122, 427.

wedlii Cobbold, 1860a, 18 (D. pelophylacis esculenti renamed) (in Rana esculenta).-Stoss., 1889, 71.

westermani Kerbert, 1881a, 529-578, pl. 26-27, for westermanii.

westermanii Kerbert, 1878a, 11 Nov., 271-273 (in Kœnigstiger; Amsterdam).—
Bettend., 1897a, 7 (westermanni): 1897b. 311 (westermanni).—R. Bl., 1891a, 610-611 (westermanni) (syn. D. ringeri): 1895. 34-39, 188: 1895, 649-932, figs. 47-116.—de Bonis, 1882, 148 (westermanni).—Brand., 1888a, 50 (westermanni); 1890a, 577 (westermanni); 1891d, 7 (westermanni).—Braun, 1892a, 586, 588, 589, 597, 602, 604, 605, 607, 622, 628, 644, 645, 654, 664, 665, 666, 668, 669, 671, 675, 677, 679, 682, 699, 700, 701, 703, 705, 707, 711, 712, 713, 715, 717, 718, 719, 723, 724, 725, 727, 728, 730, 733, 735, 736, 752, 753, 758, 784, 785 (westermanni); 1893b, 185 (westermanni); 1893b, 142-143, fig. 58 (westermanni); 1893c, 491, 492 (type. of manni); 1895b, 142–143, fig. 58 (westermanni); 1899g, 491, 492 (type of Paragonimus) (westermanni); 1900h, 3, 5, 6 (westermanni); 1901e, 329, 330, 332, 333 (westermanni); 1903, 3 ed., 155 (westermani) (to Parag.).—Buttel-Reepen, 1902, 185 (westermani).—Cobbold, 1880l, 139–140, pl. 10, figs. 1-3 (westermanni); 1884g. 976 (westermanni).—de Does, 1903, 409-412 (westermanni); 1905, 278-279 (case in dog) (westermanni).—Dubler, 1892a, 78 (in panther) (westermanni).—Fischer, 1883a, 8, 29 (westermani).—Gamb., 1896a. 63 (syn. of D. pulmonale) (westermanni).—Gronkowski, 1902a, 522 (15) (westermanni).—Huber, 1896a. 576, 577 (syn. of D. pulmonale) (westermanni).—Inouve, 1893a, 79-86: 1897a, 175-178 (2 brain cases): 1903a, 120, manni).—Inouve, 1893a, 79–86: 1897a, 175–178 (2 brain cases): 1903a, 120, 122, 123 (westermanni).—Jackson, 1888, 643 (westermanni).—Juel, 1889, 14, 33, 37 (westermani).—Kath., 1894a, 143 (westermanni).—Katsurada, 1900d, 506–523. figs. 1–3, pl. 14–15, figs. 1–8 (Japan) (westermanni): 1902b, 50–52: 1904, v. 2 (3), 22; 1904f, 148, 157 (occurs in cats and dogs) (westermanni).—Kiyono, Suga & Yamagata. —, 261, 262, 263.—Lander, 1904a, 10, 16 (westermanni): 1904, April 9, 580.—LeCount, 1904, 580 (eggs).—Linst., 1890f. 178 (westermanni).—Looss, 1885b. 6, 7, 10, 14, 15, 16, 17, 26, 27, 36, 37 (westermanni); 1894a, 118, 180, 198 (westermanni); 1899b, 560, 561 (type of Polysarcus) (westermanni); 1905, 84 (to Parag.) (westermanni).—MacCallum, 1899, 708 (westermanni).—MacCallum, 1899, 708 (westermanni).—MacCallum, 1899, 1135 (case MacCallum, 1899, 708 (westermanni).—Mackenzie, 1904a, 1133-1135 (case in Portland, Oreg.): 1904b-e; 1904, May 7, 745, 758, 901: 1904, June 15, 193; 1905, Jan. 10, 790 (case) (westermanni).—Manson, 1901, 541 (syn. of D. ringeri) (westermanni).—Moniez. 1896, 86, 144-152 (westermanni).—Mont., 1888, 18, 24, 32, 33, 36, 39, 54, 56, 64 (westermani); 1893, 1–229, pl. 1–6; 1893, 6, 26, 36, 38, 155 (westermanni).—Poir., 1885, 26, 27, 149 (westermanni)

mani).—Pratt, 1898, 361.—Shaw, 1901, 1027 (westermanni).—Simon, 1897, 259 (syn. of D. pulmonale) (westermanni).—Sons., 1895, v. 9, 291–292; 1896, 297, 302 (westermanni); 1896, July, 534–535.—Stiles, 1894i, 57–58, figs. 1–4 to (Mesogonimus); 1894k, 5 pp., 4 figs.; 1894l, 107–110, figs. 1–4 (Ward's case in U. S. A.); 1894m, 756; 1898, 96.—Stiles & Hass., 1898a, 96 (westermanni); 1900a.—Stoss., 1892, 32 (to Mesogonimus) (westermanni).—Taniguchi, 1903, Dec., 100, 102 (westermanni).—Tyson, 1903, 3 ed., 1181 (westermanni).—Ward, 1894, 355–357, 358–360 (westermanni); 1894, 362–364 (westermanni); 1895, Feb., 87–89 (westermanni in U. S. A.); 1895, Mar. 2, 236–239, figs. 1–3 (westermanni); 1895, 304–309 (westermanni); 1895, 238 (man, tiger, cat, dog), 244–246, figs. 4–6 (westermanni); 1895, 328 (in Homo), 341 (in Canis familiaris); 1896, 5 Nov., 709 (westermanni); 1903, 867 (to Parag.).—Weber, 1891, lxxxiii—lxxxiiv (tiger, Sumatra) (westermani).—Ziegler, 1883, 544, 556, 557 (westermanni).

westermanni Leuck., 1889, 404, 408, 409, for westermanii.

winogradoffi Jaksch, 1897a, 219 for sibiricum.

xanthosomum Crep., 1846a, 138 (in Colymbus septentrionalis; Germany).—Braun, 1892a, 679, 699; 1893a, 875; 1893, 353; 1893f, 427; 1902b, 7 (to Metorchis), 8, 9.—Cobbold, 1860a, 15.—Dies., 1858e, 355 (in Podiceps minor).—Linst., 1873, 101.—Looss, 1896b, 58.—Stoss., 1892, 187.—Wagener, 1857, 22, figs. 3, 4 (in Pod. mi.).

zschokkei Volz, 1899, 231–234, 237, pl. 20 (in Heterodon platyrhinus).—Braun, 1901, 944; 1902b, 65.—Luehe, 1899, 532, 533; 1900, 561.—Pratt, 1903, 25.

DISTOMACEA Weinland, 1859, 280 (family of Trematodes).—Stiles & Hass., 1898a, 87, 90.

DISTOMATA Zed., 1800a,163, pl. of Distoma.—Bojanus, 1817b, 275, 276.—Jackson, 1888, 653.—Loomis, 1885, 394.—Wyman, 1851, August, 65–66 (development). hominis Taylor, 1884, 27 issue, 44–54, figs. 1–5, apparently used as "distomes of man."—Ward, 1895, 246 (syn. of Fasc. hepatica).

DISTOMATIDÆ Gamb., 1896a, 73, family name (Distoma).

[DISTOMATINÆ Senn, 1903 [flagellates].]

DISTOMATOSIS (= any infection with distomes, s. lat.): Anacker, 1885e, 438; 1889a, 22.—d'Avrilly, (1836), 208–212 (pourriture in ruminants).—Benion, 1874a, 632–646.—Bouley, —, 21–30.—de Does, 1903, 407–409.—Fenger, 1854, 173.—Gerlach, 1854, 289–292 (liver fluke).—Grassi, (1884b); 1885a, 229–234.—Hilgendorf, 1885a, 127–130.—Hurtrel D'Arboval, 1838, 255 (cachexie); 1874a, 224–230, figs. 95–98.—Hurzard & Tessier, 1817a, 14 pp. (bovine).—Jamieson, 1897b, 147–148.—Le Roy de Mericourt, 1870a, 391–400.—Luciano, (1846a).—Mégnin, 1884, 52–55 (ruminants).—Montel, 1906, 258–262 (pulmonary).—Moulé, 1885, 501.—Perroncito, (1874), 35–36, 1 pl.; (1885), v. 28, 83–86.—Rail., 1896, 159.—Raynaud, 1859, v. 33, 488 (ruminants).—Reynal, 1856.—Schell, 1855–56.—Semmer, 1885, 438.—Signol, 1884, 33–34, figs. 16–18.—Velzen, 1891, Jul., 141.—Veterinarian, 1836, v. 9, 227–228; 1880, Aug., 548–549.—Zuendel, 1874, 224–230, figs. 95–98; ?1880, 30.

——, hepatic: Anacker, 1885d, 380–382.—Deutsche tierærztl. Wchnschr., Hannover, 1904, Jun. 4, 226.—Gómez, 1879a, Mar. 1, 81–89.—Matsumoto, 1891, 9–12.

—, —, hepatic: Brusaferro, 1887c, 296–304.—Garcia, 1884a, 129–135.

——, geographic distribution of: Akanuma, 1892, 22–27 (hepatic, Saitama district); 1894, 6 (Province of Toyama).—Corlette, 1897a, 146–147 (Sydney).—Gaide, 1905, 568–579 (hepatic, Tonkin).—Kermorgant, 1905, Feb. 7, 104 (Tonkin); 1905, Feb. 8, 88; 1905, Apr. 11, 731; 1905, May 20, 463–464; 1906, Jan. 31, 64 (Tonkin).—Monsarrat, 1898a, 82 (Hawaiian Islands).

-----, in various animals: Bolotoff, 1890a, 695–696 (domesticated).—Delafond, 1854a,
 56 pp., figs. 12–13.—Didry, 1832a, 139–147.—Léger, 1897h, 957–958 (des pélécypodes).—Mangin, 1832, 420.—Sauer, 1898, Nov. 8, 421.

DISTOMATOSIS—Continued.

- -, IN CATTLE: Aragon, 1896, 451–456, figs. 1a–3a (hepatic).—Bilhuber, (1791a), (and sheep).—Blakeway, 1879a, 260.—Buuck, 1865a, 33–35.—Collyer, 1893a, 347.—Cornil & Petit, 1901a, 178–179; 1901b, 614; 1901c, 375; 1902a, 114–115; 1902b, 117; 1902c, 57–58 (atrophic cirrhosis).—Curtice, (1891b), Jan. 24.— Jæger, 1906, v. 32 (4–5), 456–476, 2 fig. (hepatic); 1906, Aug., 382.—Lucet, 1890b, 548–549 (lungs); 1890c, 549–550 (rate).—Pease, 1895, May, 326–328; —, 5–8 (and buffaloes; India); —, 590–594.—Repiquet, 1899, May 31, 271–272, figs. 2–3; 1899, Nov., 816 (abdominal wall).—Veterinarian, 1880, Feb., 115–116; 1881, May, 317–318.—Watson, 1902, Mar., 145–151 (Connaught).
- —, pulmonary in cattle: Mackh, 1889, 308–309.—Meltzer, 1894, n. 48; 1895, 137.—
 Morot, 1887, 38–39; 1887, 64–69; 1890, 407–409.—Rail., 1885, Aug., 285–286
 (Dist. hep.).—Rivolta, (1868), 9; (1868), 267; (1868), 296–300; 1869, 473; 1869, 73–75.—Tapken, 1891, 241.
- ———, bronchial in cattle: Morot, 1893, 141–144; 1893, 249.
 - —, heart, cattle: Covu, 1898a; 1898, 98.—Muscle, cattle: Willach, 1892, 239.
 - ---, IN DOG (Canis fam.): de Jong, 1896a, 1-12, pl. 1, figs. 1-4 (hepatic) (and cat);
 1897a, 245-246; ---, 479 (and cat; hepatic). ---Kurimoto, 1894a, 45-49. -Zwaardemaker, 1888, Oct. 15, 679; 1890, Feb., 134-140 (and cat); 1890, May, 197-203, figs. 1-2; 1890, Aug., 276-277; 1890, 139-140.
 - —, IN DEER: Bassi, 1875; 1876, 235.
 - —, IN HORSE: Adams, 1876, Nov., 764–765 (hepatic).—Galli-Valerio, 1893a, 173–182; 1895c, 266 (gall duct); 1895e, 558.—Willach, 1892, v. 18, 118 (lungs).
 - -, IN MAN: Cobbold, 1884g, 976;—Florance, 1866a, 36 pp.—Grall, 1887a, 459—470, 4 figs.
 - -, —, hepatic (see also fascioliasis): Allen, 1881b, 257.—Caraoes, 1888a, 44 pp., 4 figs. (Asia).—Cobbold, 1875i, 423; 1875k, 780-781.
 - ——, hepatic, Japan (see also opisthorchiasis): Akanuma, 1892, 22–27.—Aoyama, 1891a, Apr. 5.—Hori, 1890a, Feb. 5.—Inoue, Oaknishi & Katsurada, 1891a.—Kajama & Nanba, 1892a, 32–36.—Katsurada, 1890a, 38–41 (cirrhosis); 1891a, 20–32, 2 figs., 26–41, 2 figs. (Okajamaken); 1892a, 9–32, 28–32, 31–40, 27–32, 27–33, 18–21 (pathology); 1893a, 73–79; 1897a, 11–19, 3 figs. (pathology); 1899e, 38–45, 1–16 (pathology); 1900a, 1–9, 1–10, 1–5 (pathology).—Kurimoto, 1893a, 1–7, 21–24, 35–24, 35–39, 1 pl. (Provinz Saga); 1893b, 67–69, 85–87, 109–111.—Nochezuki & Tsutsumi, 1899a, 13–24.—Otani, 1892, 1–17 (Nagasaki district).—Winoue, 1892, Jun. 20, art. 2.
 - —, —, intestinal: Ghose, 1869a, 210–211.
 - - -, ----, venal (see also bilharziosis): Duval, 1842a, 769-772; 1842, 9.
- ——, subcutaneous: Malherbe, 1898, Jan. 22, 49–50, figs. 4–11; 1898, Mar. 12, 476; 1898, Dec. 24, 1289.
- ———, IN RABBIT (*Lepus*): Rail., 1887, Jul., 324–325.
 - —, IN SHEEP [nearly all articles on distomatosis in sheep refer to fascioliasis, q. v.] [liver rot, rot, etc.]: Agric. J., Cape Town, 1894, v. 7, 232–233.—Agric. J., Cape Town, 1891, v. 4, 161; 1892, Jan. 14, 160.—Alexander, 1906, Feb. 16, 3-4.—Am. J. Micr., 1880, Jun., 135.—Bass, 1893a.—Bidloo, (1697a).—Boissou, 1878a, 183–189 (Peyrat-le-Château).—Brown, 1861a, 347–379; 1861b, 617–619; 1881a, 326–337; 1886b, 613–615.—Bruce, 1895a, 486–490.—Cobb, 1897a, 453–481, figs. 1–18, 1 pl.; 1897b.—Cobbold, 1880c, 257–258; (1880d), Apr. 7.—Dickens, 1830a, 645–647.—Edin. Vet. Rev., 1863.—Exper. Station Rec., Wash., 1903, Nov., 304.—Fonssagrives, 1868a, 299–315, figs. 1–2.—Halse, 1887a; 1888a, 147–149.—Hamilton, 1880a, 290.—Hamont, 1834a, 537–546, 587–592 (Egypt).—Harrison (1804a).—Hedwig & Reutter (1790a); (1790b), 342.—Hering, 1852a, 117–128.—Hoefnagel & Reeser, 1905, Jun.; 1906, Jan. 1, 50.—Hogo, 1903a, 603.—Hutcheon, 1903n, 603–604; 1905, Jan., 33–48.—Jones, 1894a, 274 (Australia).—J. Comp. M. & S., 1880, Jul., 173.—Joy, 1846a, 297–298.—Karkeek, 1831a, 573.—Kingdon, 1861, Apr., 224–225.—Leeuwenhoek, 1704, 1522–1527; 1705, 1522.—Lessona, (1846a).—Marshall & Hurst, 1887, 25–35.—Moodie, 1903, Jun., 726–727.—Nosotti, 1906, (5); 1906, Jun., 141.—Oekon. Nachr. d. Ges. in Schlesien, —, B. 1, 242.—Rail., 1890, 422.—Ratz, 1899, 616–618 (in spleen).—Rolleston, 1880, Apr. 14.—Rowe, 1872, Sept. 7; 1873, Feb., 120–128 (in Australia).—Salvagnoli-Marchetti, 1856,



Samml. v. Natur.- u. Med.-Geschichten, Breslau, 1718a, 694.—Schaeffer, 1753; 1762.—Schiller, 1844, 285–288 (Hungary).—Sheldon, 1881, 107–109 (England).—Simonds, 1861, 274; 1862.—Steel, 1890, 135–172, figs. 29–48.—Stewart, 1900, Oct., 10 pp.—Veterinarian, 1836, v. 9, 539–540; 1863, v. 36, 100; 1863, Mar., 157; 1872, Aug., 542; 1873, Feb., 155–161; 1879, May, 359–360; 1880, Mar., 201–202; 1880, Feb., 136–137; 1880, Jun., 402–403 (Sussex); 1881, Mar., 178–179; 1881, Jan., 29–30; 1882, Nov., 362; 1883, Feb., 130–132.—Wells, 1883, Apr., 171.—Williams, 1895, Dec., 92–93 (Australia).—Zambelli, 1874.—Zuendel, 1880, Apr. 15, 326–336; 1880, 30 np. 30 pp.

MISCELLANEOUS: Harz, 1881c, 1-15, pl. 1 (in Astacus).—Rail., 1887, Aug., 559-560 (pulmonary); 1887, 392.—Vallada, 1882, 35-39 (crayfish and crabs).-Zuendel, 1881, 459-465; 1881, Oct., 833-848, 1 pl., figs. 1-3; 1882, Mar.,

196-202, figs. 1-3 (des écrevisses).

196-202, figs. 1-3 (des écrevisses).

PATHOLOGY AND LOCATION of: R. Bl., 1901b, 204-212, 218-219; 1901c, 581-589.—Bossuat, 1902, 161-206 (hepatic); 1902, 56 pp.; 1903, Jan. 17, 12; 1903, Feb. 28, 115-116; 1903, Aug. 14, 533-534; 1905, July, 76.—Bonvicini, (1881a), 133-134 (hepatic); 1882a, 114.—Colucci, 1882, 535-568.—Hamont & Fischer, (1834b), 129.—Inoue, 1897b, 1-5; 1898a, 8-32, 25-40, 16-40, 17-28 (Japan); (1899a), 20, 15, 1, 1, 1; 1900b, 1-23 (pulmonary); 1900c, 515, 664, 739.—Inoue & Katsurada, 1891a, 8-12, 1 pl. (hepatic).—Katsurada, 1892.—Macgregor, 1877.—Miura, 1889, 310-317, pl. 7, figs. 1-3.—Morot, 1889, 37 (cattle); 1890, 732-734.—Pastor, 1897; 1900, 617-618 (hepatic).—Perroncito, 1887, Mar. 3, 97-112.—Pilavios, 1894, 407 (lungs, cattle); 1894, 579; 1895, Apr., 133.—Sodero, 1889, Nov. 30, 484-490, figs. 1-2 (hepatic); 1892, 161.—Spengel, (1892), 146-147.—Taniguchi, 1904, 100-121; 1905, July 29, 508-509 (cerebral).—Toichiro, 1883, Feb. 23.—Yamagiwa, 1899; 1890 (cerebral, man).—Zacharias, 1892, Dec. 3, 752-753 (cysts).

-, mortality from: Veterinarian, Lond., 1836, Apr., 225–228; 1880, Apr., 246–247;

1880, Sept., 650–664.

-, symptoms of: Aitken, 1866, 42; 1872, 207.—Anders, 1903, 6 ed, 1245.—Inoue & Kodama, 1896a, 14–21.—le Ray, 1897a, 372–386.

prevention of: Agric. J., Cape Town, 1894, Sept. 20, 442.—Bennett. 1876a, 677–680.—Manson, 1901, 543.—Stiles, 1902, pp. 220–222, figs. 197–203; 1905dd, 220–222, figs. 119–128.—Tierarzt, Wetzlar, 1904, May, 99–100.—Veterinarian,

1837, Aug., v. 10, 442–444.

-, treatment of: Agric. J., Cape Town, 1898, Aug. 4, 138-139.—Anders, 1903, 6 ed., 1245.—Armatage, 1895, 445-454.—Bekker, 1893, Jan. 26, 23.—Fegliata, 1887a), 61-62.—Khouri, 1904, 80 (rhubarb).—Riomsai, 1883, Sept. 5 (pulmonary); 1884, Feb. 16.—Romagli, 1903, Feb. 25; 1903, May 31, 301 (salol, sheep); 1903, June, 245; 1903, July 1, 513-519; 1904, Jan. 1, 9; 1904, Apr., 89-90.—Tappemer, 1900, 97-105; 1900, Dec. 11, 1729-1731.—Universal Mag., 1838, (1748).—Vallada, 1857; 1859, Aug., 538-595.—Verhandl. ü. Arb. d. Oekon. Patriot. Soc. d. Fürstenth. Schweidnitz u. Janer, 1818, 323.—Veterinarian, 1838, Mar., 112 (salt); 1873. Oct., 698-699.

DISTOMEA Mont. [not Distomea (Protozoa)]. See Braun. 1890a, 516.—Olss., 1893.— Stiles & Hass., 1898a, 87, 90. See also Leuck. (1856), f. name.

DISTOME.E Leuck., 1863a, 527 (fam. name).—Brand., 1888a, 49; 1891c, 730.—Braun, 1883a, 38, 58; 1893a, 886.—Jackson, 1888, 644.—Schneidemuehl, 1896, 296.— Stiles & Hass., 1898a, 87, 90.—Tasch., 1879, 234.—Zuern, 1882, 113.

DISTOME.E Mont., 1888a, 7, 8, 9, 11, 12, 14, 15, 16, 18, 22, 27, 31, 34, 35, 36, 37, 38, 41, 47, 48, 49, 51, 52, 53, 54, 56, 57, 60, 73, 84, 90, 92, 102, 104, 108.

DISTOMIDA Manson, 1903, 3 ed., 606 (probably for Distomidæ).

DISTOMID.E Cobbold, 1860a, 1-56 (synopsis of tribe); 1877f, 326; 1879b, 4.—Brand., 1890a, 576.—Braun, 1883a, 59; 1890a, 515, 538; 1893a, 886, 890, 895, 900, 907; 1895b, 136.—Caullery, 1902, 21–24 (ascidian).—Haswell, 1887a, 286.—Haymann, 1905, 81–100, 1 pl. (from chelonians); 1906, Mar., 178.—[Herdman, 1890, 617 (family of Ascidiæ compositæ)] —Hoyle, 1890, 539 (includes: Dist., Fasc., Bilharzia. Echinost., Amphist., Gastrodiscus, Homalogaster, Gastrothylax, Euryccelium).—Jackson, 1888, 643, 646, 647, 648, 654 (key).—Jacoby, 1899c, 1–30, 2 pls.—Kholodk., 1898, 25.—Klein, v. 22, 59–80, 1 pl. (from Rana); 1906, Mar., 178.—Leuck., 1886d, 30, 90, 442.—Levin., 1881a, 67.—Looss, 1899b, 541, 542, 543, 659 (contains Amphistominæ Looss, Fasciolinæ Looss) Inot Distomidæ ascidian see Herdman, 1900, 821, 1900d, 601, 602, 603 Looss) [not Distomidæ, ascidian, see Herdman, 1900, 82]; 1900d, 601-608

DISTOMIDÆ-Continued.

(genera, nomenclature); 1901, 191, 196; 1902b, 120; 1902m, 838; 1904i, 21–22.—Maclaren, 1904, 598, 605, 608, 610, 611, 612, 613.—Montgomery, 1906, Feb. 12, 16.—Mont., 1888a, 7, 15, 17, 18, 20, 24, 28, 30, 43, 52, 56, 60, 68, 90, 92, 104 (subf.); 1891, 449, 516; 1892, Oct. 7, 214 (fam. of Malacocotylea); 1893, 153, 154, 155; 1893, 229; 1894, June 2, 872–874; 1894, Nov. 1, 16–21.—Much., 1898, 21.—Nickerson, 1902, 610.—Schneidemuchl, 1896, 295, 296.—Stiles, 1905z, 14.—Stiles & Hass., 1898a, 87, 89, 90; 1900a, 558–611, 2 pls., 6 figs.—Stoss., 1898, 22.

DISTOMIDEA Ben.,——.—Carus, 1863, 478.—Mont., 1888a, 84.

DISTOMIDÉS Bl., 1888a, 541, 542, 543 (includes Dist., Amphist., Bilharzia), French name.

DISTOMIDI Mont., 1892, Oct. 7, 191.

DISTOMIDŒ Montgomery, 1906, Feb. 12, 17 (for Distomidæ).

DISTOMII Duj. of E. Bl., 1847, 277 (includes: Distomidæ, Amphistomidæ, Holostomidæ).

DISTOMINÆ Mont., 1892, Oct. 7, 214 (subf.).—Braun, 1893a, 890.—Looss, 1899b, 541.—Mueh., 1898, 21.—Stiles, 1905z, 14.—Stiles & Hass., 1898a, 90.—Stoss., 1898, 22.

DISTOMINEA, see Stiles & Hass., 1898a, 87.

DISTOMOMUM Sons., 1894, 172 (for Distomum).

DISTOMOSE, see distomatosis, Rail., 1896, 159, name of disease.

DISTOMOPSIS Rafinesque, 1815, 151 (new name for Dist. Zed., hence type Fasc. hepatica).

DISTOMULUM Brand., 1892, Oct. 7, 510 (see Agamodist.).—Stiles & Hass., 1898a, 82, 87 (syn. of Agamodist. Stoss., 1892).—Stiles, 1902s, 29; 1904i, 12.

DISTOMUM Dies., 1850a, 141, for Distoma.

DISTOMUS Gærtner, 1774, tunicate.

mamillaris Gærtner in Pallas, 1774, 20, also index.

variolosus Gærtner in Pallas, 1774, 35, 40, pl. 4, fig. 7, tunicate.—Lamarck, 1816, v. 3, 101.

DISTOMUS Lænnec, 1812c, 9–12 (m. intersectus).

intersectus Lænnec, 1812c, 9-12.

DISTONUM Goto, 1891a, 181.—Laspeyres, 1904a, 5, 12 (for Distomum).

DOLICHODEMAS Looss, 1900, 603, 608, Dolichosomum Looss, 1899 (not Dolichosoma Steph.,—, coleopt.; not Huxley, 1867, saur.) renamed, hence type lorum Duj., 1845); τὸ δεμας, body.

DOLICHOSOMUM Looss, 1899b, Dec., 652–653, 655 [not Dolichosoma Steph., ——, coleop., not Dolichosoma Huxley, 1867, saur.] m. lorum; δολιχός long, τὸ σῶμα body; 1900, 603, 608 (renamed Dolichodemas).—Braun, 1900h, 13; 1901i, 56.—Luehe, 1900, 557.—Ofenheim, 1900, 183. 1899: Dolichost. Looss, 1899b, 551, 652, misprint.

lorum (Duj., 1845a) Looss, 1899b, 653.

DOLICHOSTOMUM Looss, 1899b, Dec., 551, 652, misprint for Dolichosomum.

DYSTOMA Bojanus, 1817b, pl. ix (for Distoma).

amphistomoides Bojanus, 1817b, 270–277, pl. 9, figs. 1–6 (in Castor fiber).

hepaticum (Linn., 1758) Most, 1836, v. 1, 581, see sub Distoma.

krohnii Kælliker, 1849d, 65 (in cephalopod), see sub Distoma.

ECHINATA (Echinostoma) Rud., 1809a, 415 [the term Echinata is probably not used in a nomenclatural sense].

ECHINELLA Ben. & Hesse, 1864, 93–94 (not Achar., 1803, poly.; not Swains., 1840, mollusk) (m. hirundinis).—Braun, 1890a, 408, 410, 446, 511, 516, 517, 523, 531, 532; 1893a, 890.—Gamb., 1896a, 73.—Mont., 1888a, 86, 88, 98; 1891, 111; 1903, 336 (subf. Udonellinæ).—Pratt, 1900a, 646, 649 (key), 655, fig. 13.

hirudinis Mont., 1888a, 10 (for hirundinis).

hirundinis Ben. & Hesse, 1864, 94, pl. 8, figs. 17–19 (in Trigla hirundo).—Braun, 1890a, 418, 532, 548, 550, 552.—Mont., 1888a, 10 (hirudinis).—Pratt, 1900a, 655, fig. 13, 657.—Tasch., 1878, 573 (to Udonella).

ECHINOSTOMAa Rud., 1809a, 37-38, 415 [tld. echinatum].—Baird, 1853a, 55.— Blainv., 1828a, 587-588.—R. Bl., 1891, 609.—Brand., 1892b, 506.—Braun, 1892a, 567. 568. 578, 584, 699, 708, 772, 815; 1893a, 821. 884, 885, 890, 909. 911; 1895b, 121, 138: 1900h, 3; 1901b, 29, 33, 34: 1901e, 315, 328; 1901, 562; 1902b. 26, 117.—Cobbold, 1858b, 162; 1860a, 32; 1864b, 32; 1879b, 461—462.—Dies., 1850a; 1855c; 1858e; 1859c.—Duj., 1845a, 423.—Fil., 1855b, 25; 1857c, 28.— Fuhrmann. 1904. 62.—(Gœze. 1782a).—Hoyle, 1890, 539.—Jackson, 1888, 643, 654.—Kowal., 1897c, 554–565, figs. 1–8 (in chickens and ducks): 1897d, 37–38. 415; 1814a; 1819a.—Stiles, 1901. 197.—Stiles & Hass., 1894; 1898a, 87 (type echinatum). 88, 96.—Stoss., 1892, 4, 5, 28; 1898, 52; 1899, in 11–16.

acanthocephalum (Stoss., 1887) Stoss., 1898, 57 (in Belone acus: Triest); 1899, 1,6

(to Tergestia).

acanthoides (Rud., 1819) Cobbold, 1860a, 33.—Braun, 1901e, 315.—Duj., 1845a, 424 (in Phoca vitulina; Berlin).—Looss, 1899, 581.—Stoss., 1892, 29 (in Ph. vit.: Baltic): 1899, 14.

africanum Stiles, 1901k, 594 (E. bursicola Looss, renamed).

anceps (Mol., 1859) Stoss., 1892, 168 (in Fulica atra; Padova); 1899, 13.—Looss, 1899b, 581.

annulatum (Dies., 1850) Cobbold, 1860a, 36 (in Gymnotus electricus; Brazil).— Looss, 1899, 581.—Stoss., 1899, 14

apiculatum (Rud., 1803 Cobbold, 1860a. 35.—Loo ss, 1899b, 581, 696.—Stoss., 1892 173 (in Strix flammea L., Greifswald; Syrnium aluco L., Vienna); 1899, 14 armatum Fuhrmann, 1904, 61-63, fig. 3 (in Rostrhamus sociabilis).

asperum (Wright, 1870) Stoss., 1892, 164 (in Botaurus minor; North America); 1899, 13.—Loess, 1899b, 581, 695, 696; 1902m, 817.—Odhn., 1902, 38,

 baculus | Dies., 1850 | Cobbold, 1860a, 36 (in Mergus albellus ; 1860, 31.—Looss, 1899b, 581, 693.—Nicoll, 1906, 154.—Stoss., 1892, 163 (in Mer. alb.; Ratzeburg); 1899, 14.

beleocephalum (Linst., 1873) Stoss., 1892, 169 (in Ardea cinerea L.; Ratzeburg); 1899, 14.—Looss, 1899, 581, 687, 695.

bicoronatum (Stoss., 1883) Par., 1902. 5 in Corvina nigra, Umbrina cirrhosa; Portoferrajo).

bilobum (Rud., 1819) Cobbold, 1860a, 35.—Looss, 1899, 572, 581, 684-685, 687, fig. 8; 1901b, 201, 209 (with E. ramosum probably represents a subg. of Echinost.).—Odhn., 1902, 38.—Stoss., 1892, 171–172 (in Fulica atra. Plegadis falcinellus: Padova and Vienna; Platalea leucorodia; Venice); 1899, 13.

bursicola Looss, 1899b, 694-696, figs, 19-20 (in Milvus parasiticus at Adeleninsel, Heluan. Apr.: Falco tinnunculus at Dachschur, Jan.).—Braun, 1901, 18; 1902b, 74.—Stiles, 1901s, 593-594 (renamed africanum); 1902cc, 360.

cesticillus (Mol., 1858) Mont., 1893, 158–161, pl. 5, figs. 56–60, pl. 6, fig. 68.—Barbagallo & Drago, 1903, 11 (in Seriola dumerili, Torpedo ocellata, Umbrina cirrhosa; Catania).—Looss, 1894a, 252 (Dist. valdeinflatum as syn.); 1899b, type of Stephanost.—Stoss., 1898, 54 (in Zeus faber, Lophius piscatorius, Corvina nigra, Umbrina cirrhosa; Triest).

cinctum (Rud., 1803) Cobbold, 1860a. 35 (in Vanellus cristatus, V. melanogaster).—Looss. 1899b, 580, 683, 687.—MacCallum, 1904, 547.—Stoss., 1892, 172–173 (in Vanellus cristatus. V. melanogaster, Gallinula chloropus; Rostock); 1896, 126; 1897, 10; 1898, 53; 1899, 12.

cloacinum Braun. 1901d, 259 | D. bursicola Crep., renamed) (in Ardea cinerea); 1902b, 74 (Dist. bursicola Crep., as syn.).—MacCallum, 1904, 547.—Stiles, 1901k, 593-594 (nomenclature); 1902cc, 360.

a Rud, originally proposed Echinostoma as a genus, but in the same work he used it in such a way that we must also interpret it as a subgenus. For the original species see D. (Echinostoma).

ECHINOSTOMA—Continued.

columba livia Mueller, 1897, 26, pl. 3, fig. 7 (in Columba livia).

conoideum (Bloch, 1782) Kowal., 1896, 5; 1897c, 554-565; 1897c, 407-408; 1898h,
111; 1902d, 27 (in Anas boschas dom.; Dublany); 1904, 24 (in Anas boschas dom.; Dublany).—Galli-Valerio, 1898m, 923.—Looss, 1899b, 581.—Rail.,
1898, 412.—Stoss., 1899, 13 (in Anas boschas).

coronatum (Rud., 1819) Stoss., 1892, 30–31 (in Didelphys virginiana; Brazil); 1892, 37; 1899, 14.—Braun, 1901e, 319, 320 (to Rhopalias).—Looss, 1899b, 581.

coronatum (Wagener, 1852) Stoss., 1898, 56-57 (in Corvina nigra; Triest).—Type of Anoiktost. 1899.

cristatum (Rud., 1819) Cobbold, 1860a, 37 (in Stromateus fiatola).

croaticum (Stoss., 1889)
 Stoss., 1892, 165–166 (in Carbo graculus; Fiume); 1899,
 12.—Looss, 1899b, 580.—Par., 1902, 1, 4 (in Phalacrocorax graculus; Caproja).

crocodili (Poir., 1886) Stoss., 1895, 227–228 (in Crocodilus siamense); 1899, 12.— Looss, 1899b, 575, 580.—Odhn., 1902, 20.

denticulatum (Rud., 1802) Cobbold, 1860a, 36.—Looss, 1899, 581.—Stoss., 1892, 170–171 (in Sterna cantiaca, Vienna; S. hirundo, Greifswald; Hydrochelidon nigra, Anas sponsa); 1899, 14.

dilatatum (Fischer, 1840) Cobbold, 1860a, 35 (in Phasianus gallus).—Hass., 1896a, 3 (syn. of Echinost. echinatum (Zed.)).—MacCallum, 1904, 547.—Stoss., 1892, 167.

dujardini MacCallum, 1904, 547, for dujardinii.

dujardinii Cobbold, 1860a, 37 (in Pleuronectes maximus, P. platessa); see Dist. hystrix Duj.

echinatum (Zed., 1803) Blainv., 1828a, 588.—Baird, 1853a, 55.—Braun, 1906, 142 (in ducks and geese).—Cobbold, 1860a, 33.—Galli-Valerio, 1898m, 923.—Hass., 1896a, 3 (syns., D. ech., D. oxycephalum, D. dilatatum, D. armatum, Echinost. dilatatum) (in Gallus dom.).—Kowal., 1894, 221; 1896. 257 (in Corvus cornix, Anas crecca); 1898g, 70; 1898h, 111, 113, 115, 116, 117, fig. 27; 1902d, 27 (in Anas boschas dom.; Dublany); 1904, 24 (in Anas boschas dom.; Gallus dom.; Dublany).—Linst., 1893, 333 (larva in Bythinia ventricosa, Physa fontinalis, Valvata macrostoma, Limnæa palustris).—Leoss, 1899b, 575, 580, 679–684, 687, 693, figs. 5, 7, 10; 1901b 201, 209; 1902m, 823.—Luehe, 1898, 625; 1901, 175.—MacCallum, 1904, 547, 548.—Mueh., 1898, 21; 1898, 71 (in Œdemia fusca, Colymbus arcticus, Ciconia alba, Buteo vulgaris).—Nicoll, 1906, 154.—Nord., 1840, 621 (syn. of Fasc. trigonocephala).—Rail., 1898, 412.—Sons., 1897, 252.—Stiles & Hass., 1898a, 88.—Stoss., 1892, 167 (syns. Dist. anatis fuscæ, D. armatum, D. dilatatum, D. ech., D. fuligulæ ferinæ, D. oxycephalum, D. radiatum, Ech. dilatatum, Fasc. anatis); 1896, 127; 1898, 52; 1899, 12.—Wolffhuegel, 1900, 9, 57, 58, 59, 60, 61.—Reported for Anas anser, A. boschas, A. clangula, A. clypeata, A. cygnus, A. ierina, A. fuligula, A. marila, A. moschata, A. nyroca, A. olor, A. penelope, A. strepera, A. tadorna, Anser albifrons, Archibuteo vulgaris. Ardea comata, A. gardeni, A. grus, A. nycticorax, A. pavonia, Balæarica pavonia, Bucephala clangula, Carbo cormoranus, C. pygmæus, Canis familiaris, Chaulelasmus streperus, Ciconia alba, C. nigra, Colymbus arcticus, Coracias garrula, Fuligula cristata, Grus cinerea, Machetes pugnax, Mareca penelope, Pernis apivorus, Podiceps cristatus, P. minor. Larva in: Lymnæus stagnalis, Paludina vivipara, P. achatina, Planorbis corneus.

echiniferum (La Valette, 1855) Stoss., 1892, 171 (in Scolopax gallinago, Mergus merganaso schas, Podiceps minor); 1899, 14.—Looss., 1899, 581.—

Wolffhuegel, 1900, 9, 56.

echinocephalum (Rud., 1819) Cobbold, 1860a, 34–35 (in Falco milvus).—Looss, 1899b, 581, 687, 696.—Stoss., 1892, 173–174 (in Milvus regalis); 1899, 14.

elegans Looss, 1899b, 692-693, fig. 18 (in Phænicopterus roseus; Ghizeh).

curyporum Looss, 1896b, 144–146, pl. 9, fig. 93 (in Milvus parasiticus; Cairo; also in Ardea cinerea); 1899b, 580, 686–687, 690, 696, fig. 16, 17.—Fuhrmann, 1904, 62.—Stoss., 1899, 12.

fallax (Rud., 1819) Cobbold, 1860a, 37.—Looss, 1902m, 823.—Stoss., 1898, 56 (in Uranoscopus scaber; Triest).

ferox (Rud., 1795)
 Blainv., 1828a, 588.—Cobbold, 1860a, 35.—Kowal., 1902d,
 28.—Looss, 1899, 581, 687;
 1902m, 817.—Luehe, 1905, 176.—Mueh., 1898,

ECHINOSTOMA—Continued.

21, 23, 102–105, fig. 14.—Nord., 1840, 622 (as syn. of Fasc. trigonocephala).—Odhn., 1902, 38 (represents a new [unnamed] genus).—Stiles & Hass., 1898a, 88.—Stoss., 1892, 164–165 (in Botaurus stellaris, Ciconia alba, C. nigra; Padova, Vienna, Greifswald, Rennes); 1899, 13.

flexum (Lint., 1892) Stoss., 1899, 13 (in Edemia americana).—Looss, 1899b, 581.
 fralichii (Kowal., 1894) Kowal, 1894, 222; 1898h, 111, syn. of E. conoideum (Bloch).—Stoss, 1899, 13 (in Anas boschas).

gadorum Ben., 1870, 60 (in Merlangus carbonarius; Belgian coast).—Hoyle, 1890, 539.—MacCallum, 1904a, 547.

garzettæ MacCallum, 1904a, 541–548, 1 fig. (in Garzetta nigripes Temm.; Benakat); 1904, Oct., 532.

hispida of Ben., 1870, pl. 4, fig. 13.—Odhn., 1902, 157 (syn. of Deropristis inflata).
hispidum (Abildg., 1819) Cobbold, 1858b, 171, figs. 47–50; 1879b, 462, fig. 77 (in sturgeon).—Ben., 1870, 23, pl. 4, figs. 12–13.—Mont., 1893, 83.—Odhn., 1902, 155, 156, 159 (type of Deropristis), 157 (fig. 13 of Ben., 1870=D. inflata).—Stoss., 1898, 57–58 (in Acipenser sturio; Triest); 1899, 14.

imbutiforme (Mol., 1859) Stoss., 1898, 55-56 (in Labrax lupus; Triest).

incrassatum (Dies., 1850) Cobbold, 1860a, 33 (in Lutra solitaria; Brazil).—Braun, 1901e, 317, 328, pl. 19, figs. 5, 8, 9.—Fuhrmann, 1904, 64.—Looss, 1899, 581.—Stoss., 1892, 30 (in Lutra solitaria; Brazil); 1899, 14.

inerme Fuhrmann, 1904, 63-64, fig. 4 (in Lutra).

inflatum (Mol., 1859) Stoss., 1898, 58 (in Anguilla vulgaris; Triest).

labracis (Duj., 1845) Ben., 1870, 45.—MacCallum, 1904a, 547.

laticolle (Rud., 1819) Par., 1899, 5 (in Trachurus trachurus; Rimini, Naples); 1902, 4 (in Corvina nigra; Seriola dumerili; Elba).—To Tergestia in 1899.

leptosomum (Crep., 1829) Cobbold, 1860a, 35 (in Tringa variabilis, Chalidris arenaria).—Looss, 1899b, 580.—Nicoll, 1906, 517.—Stoss., 1892, 169: 1899, 12.

lidiæ Par., 1902, 6, for lydiæ (in Orthagoriscus mola; Tonnara d'Enfola).

liliputanum Looss, 1896b, 141–143, pl. 9, figs. 91–92 (in Pernis apivorus at Alexandria, Egypt; Milvus parasiticus at Matarijeh); 1899b, 575, 580, 696; 1901b, 200, 201, 202, 209; 1902m, 816, 817 (form from cat represents a new genus), 818.—Stoss., 1899, 12.

lydiæ Stoss., 1896, 190–191, pl. 8, fig. 1 (in Orthagoriscus mola; Triest); 1898, 55.—
 Barbagallo & Drago, 1903, 411 (in O. m.; Catania).—Looss, 1901e, 605, type of Dihemistephanus.—Par., 1902, 6 (lidiæ) (in O. m.; Tonnara d'Enfola).

magniovatum Stoss., 1898, 53–54 (in Puffinus kuhlii; Triest); 1899, 13.—Looss, 1899b, 581; 1901b, 209; 1901e, 596.

militare (Rud., 1803) Cobbold, 1860a, 34; 1879b, 436, 437.—Kowal., 1902d, 28.—Looss, 1899b, 581.—Stoss., 1892, 163–164 (in Numenius arquatus, Scolopax gallinula, S. gallinago, Ortygometra porzana); 1899, 13.—Also reported for Limnocryptes gallinula.

mordax Looss, 1899b, 688, 690, fig. 9 (in Pelecanus onocrotalus; Egypt).—Kowal., 1904, 19 (in Podiceps auritus; Dublany).

nephrocephalum (Dies., 1850) Cobbold, 1860a, 36 (in Turdus saxatilis).—Looss, 1899b, 581.—Stoss., 1892, 174; 1899, 14 (in T. sax.).

nigroflavum (Rud., 1819) Barbagallo & Drago, 1903, 410 (in Orthagoriscus mola; Catania).

oxycephalum (Rud., 1819) Rail., 1896, 160.

pendulum Looss, 1899b, 580, 688–692, 693, 697, figs. 13, 15b, c (in Recurvirostra avocetta; Adeleninsel, Egypt, Feb.).—Fuhrmann, 1904, 62.

perlatum (Rud., [=Nord., 1832]) Barbagallo & Drago, 1903, 410-411 (in Tinca vulgaris; Catania).

phænicopteri Luehe, 1898g, 625 (in Flamingo; Berberei); 1899e, 235.—Looss, 1899b, 580, 693 (in Phænicopterus ruber).—Stoss., 1899, 12 (Barbary).

pristis (Deslongchamps, 1824) Cobbold, 1860a, 36 (in Merlangus communis).— Looss, 1899, to Stephanost.—Stoss., 1898, 54–55 (in Gadus euxinus; Triest).

pseudoechinatum (Olss., 1876)
 Stoss., 1892, 166 (in Larus marinus; Scandinavia):
 1899, 13.—Kowal., 1902d, 27.—Looss, 1899b, 581, 685–686, 690, 691, 692,
 figs. 11, 12, 15.—Mueh., 1898, 21–22.—Also reported for Larus fuscus.

ECHINOSTOMA—Continued.

pungens (Linst., 1893) Stoss., 1899, 14 (in Podiceps minor).—Looss, 1899b, 581.— MacCallum, 1904a, 547.

ramosum Sons., 1895, 123–124 (in Babulcus ibis: Nile Delta).—Looss, 1899, 581; 1901, 201.—MacCallum, 1904a, 547.—Odhn., 1902, 38.

recurratum (Linst., 1873) Stoss., 1892, 162–163 (in Fulix marila: Ratzeburg); 1899, 12.—Looss. 1899b, 580, 693.—Kowal., 1894, 220; 1902d, 27: 1904, 24 (in Anas boschas dom., Gallus dom.; Dublany).—MacCallum, 1904a. 547.—Rail., 1898, 412.—Also in Fuligula marila.

revolutum (Frœlich, 1802) Looss, 1899, 680.

scabrum (Mueller, 1788) Cobbold, 1860a. 37 (in Lota molva. Gadus morrhua).

secundum Nicoll, 1906, 514, 515-518, pl. 12, figs. 1-3 (in Hæmatopus ostralegus, Larus argentatus, L. ridibundus).

serratum (Dies., 1850) Cobbold, 1860a, 35 (in Anas scolopaceus; Brazil).—Looss, 1899b, 581.—Stoss., 1892, 171; 1899, 14.

spathulatum (Rud., 1819) Kowal., 1898g. 69–71. 76, figs. 1–14, 37 (in Botaurus minutus by Bremser); 1898h, 106–121, 158, figs. 1–14, 37 (in Botaurus minutus; Dublany) (spatulatum).—Looss, 1899b, 575, 581.—Stoss., 1899, 13.

spatulatum (Rud., 1819) Kowal., 1898h, 106-121, 158. See also spathulatum.

spiculator (Duj., 1845) Cobbold, 1860a, 33.—Looss, 1899b, 581.—Stoss., 1892, 28 (in Mus decumanus; Hameln, Rennes): 1899, 13.

spinulosum (Rud., 1809) Cobbold, 1858b, 165. pl. 33, figs. 68–72 (in Larus glaucus);
1860a, 36 (in Colymbus septentrionalis. Podiceps cristatus. Anas querquedula, Uria grylle, Carbo graculus. Larus argentatus, L. capistranus. L. glaucus, L. ridibundus. Anas clangula. Numenius arcuatus; 1879b, 448.—Cohn, 1903, 41 (in Alca torda).—Condorelli, 1897, in 118–124 (in Hydrocolæus minutus).—Gilbert, 1905, v. 39. 925–927. 1 fig.—Looss, 1896b, 141; 1899b, 580, 687, 689, 690, 691.—Mueh., 1898, 22.—Nicoll, 1906, 154; 1906, 516, 517.—Par., 1899, 5; 1902, 4 (in Fulica atra; Elba).—Stoss., 1892, 169–170 (in Querquedula circia, Phalaerocorax graculus, Larus argentatus, L. capistranus, Uria grylle, Totanus fuscus, Numenius arquatus); 1896, 127 (in Numenius tenuirostris; Monfalcone); 1898, 52–53 (in Larus ridibundus, Numenius tenuirostris, Podiceps cristatus; Triest); 1899, 12.—Wolffhuegel, 1900, 9, 56 (in Machetes pugnax), 62.—Also reported for Bucephala clangula, Chroocephalus minutus, Fulica atra, Larus fuscus.

tabulatum Mueller, 1897, 21–22. pl. 3, fig. 5 (in Numenius arquatus).—MacCallum, 1904a, 547.

trigonocephalum (Rud., 1802) Cobbold, 1860a. 33.—Kowal.. 1902d, 27; 1904, 25 (in Putorius fœtidus; Dublany).—Looss, 1899b, 581; 1902m, 823.—Mueh., 1898, 22.—Stoss., 1892, 29-30 (syns. Fasc. putorii, F. trig., Dist. trig.) (in Erinaceus europæus at Padova, Vienna; Meles taxus at Warnemünde; Mustela foina; Lutra vulgaris; Putorius lutreola; P. communis at Padova. Vienna; Putorius vulgaris at Venezia, Greifswald; Vulpes vulgaris); 1899, 13.

umbonatum Odhn., 1902, 21-22 (in Krokodil; Nile).

uncinatum (Zed., 1803) Cobbold, 1860a, 35 (in Gallinula chloropus).—MacCallum, 1904a, 547.—Stoss., 1892, 172 (=E. cinctum) (in Vanellus cristatus, V. melanogaster, Gallinula chlor.; Rostock).

(ECHINOSTOMA) Rud., 1809a, 37–38, 415–429 (tld. echinatum); a. subg. of Dist. See also Echinost.

acanthoides Rud., 1819a, 114, 415—416 (in Phoca vitulina; Berlin). See Echinostoma.

annulatum (Dies., 1850) Stoss., 1886, 63. See Echinost.

apiculatum (Rud., 1803) Rud., 1809a, 423. See Echinost.

areolatum (Rud., 1809) Stoss., 1886, 60.

bicoronatum (Stoss., 1883) Stoss., 1886, 58.

bilobum Rud., 1819a, 114, 416 (in Tantalus falcinellus). See Echinost.

campanula (Duj., 1845) Stoss., 1886, 62.

cesticillus (Mol., 1858) Stoss., 1886, 57. See Echinost.

cinctum (Rud., 1803) Rud., 1809a, 422-423. See Echinost.

contortum Rud., 1819a, 118 (in Orthagoriscus mola; Naples).

(ECHINOSTOMA)—Continued.

coronarium (Cobbold, 1861) —— ? ——, 1896.

coronatum Rud., 1819a, 686 (in Didelphis virginiana). See Echinost.

coronatum (Wagener, 1852) Stoss., 1886, 58. See Echinost.

corvinæ (Stoss., 1886) Stoss., 1886, 58.

cristatum Rud., 1819a, 117 (in Stromteus fiatola; Arimini). See Echinost.

denticulatum (Rud., 1802) Rud., 1809a, 424-425. See Echinost.

echinatum (Zed., 1803) Rud., 1809a, 418-420. See Echinost.

echinocephalum Rud., 1819a, 115 (includes D. milvi.) (in Falco milvus). See Echinost.

elipticum Stoss., 1886, 64, for ellipticum (Mol., 1858).

exasperatum Rud., 1819a, 117 (in Sorex eremita).

fallax Rud., 1819a, 117 (in Uranoscopus scaber; Naples). See Echinost.

ferox (Rud., 1795) Rud., 1809a, 426-427. See Echinost.

hemicyclum (Mol., 1859) Stoss., 1886, 60.

hispidum (Abildg., 1819) Rud., 1819a, 118, in Accipenser sturio; Arimini, Berlin; includes D. sturionis. See Echinost.

hystrix Duj., 1845a, 433 (in Pleuronectes maximus, P. platessa).

imbutiforme (Mol., 1859) Stoss., 1886, 59. See Echinost.

incrassatum (Dies., 1850) Stoss., 1892, 30. See Echinost.

increscens (Olss., 1868) Stoss., 1886, 58.

inflatum (Mol., 1859) Stoss., 1886, 63. See Echinost.

labracis (Duj., 1845) Ben., 1870, 1871a, 24 (in Labrax lupus; Belgium). See Echinost.

laticolle Rud., 1819a, 117 (in Caranx trachurus; Naples and Arimini). See Echinost.

leptosomum (Crep., 1829) Duj., 1845a, 428. See Echinost.

lima Rud., 1809a, 427-429.

militare (Rud., 1803) Rud., 1809a, 421-422. See Echinost.

mulli (Stoss., 1883) Stoss., 1886, 59.

nigroflavum Rud., 1819a, 118 (in Orthagoriscus mola; Naples).

oculatum (Levin., 1881) Stoss., 1886, 58.

perlatum (Nord., 1832) Stoss., 1886, 62. [Type of Asymphylodora.]

planicolle Rud., 1819a, 686 (in Pelecanus sula; Brazil).

polonii (Mol., 1859) Stoss., 1886, 58.

pristis (Deslongchamps, 1824) Duj., 1845a, 433. See Echinost.

rachion (Cobbold, 1858) Stoss., 1886, 61.

radiatum Duj., 1845a, 385, 427–428 (in Carbo cormoranus or Pelecanus carbo).

radula Duj., 1845a, 433–434 (in Lymnæus palustris; Rennes).

scabrum (Mueller, 1784) Rud., 1819a, 118. See Echinost.

semiarmatum (Mol., 1858) Stoss., 1886, 64.

sobrinum (Levin., 1881) Stoss., 1886, 58.

sophiæ (Stoss., 1886) Stoss., 1886, 59.

spathulatum (Rud. [Crep., 1837]) Par., 1902, 4 (in Ardetta minuta; Portoferrajo). spiculator Duj., 1845a, 424–425 (in Mus decumanus; Rennes). See Echinost.

spinulosum Rud., 1809a, 425 (in Larus nævius, L. cinerarius and Colymbus septemtrionalis). See Echinost.

trigonocephalum (Rud., 1802) Rud., 1809a, 415–418. See Echinost.

uncinatum (Zed., 1803) Rud., 1809a, 420-421. See Echinost.

validein flatum (Stoss., 1883) Stoss., 1886, 58.

viviparum (Olss., 1868) Stoss., 1886, 60.

ECHINOSTOMIDÆ Looss, 1902m, 817.

ECHINOSTOMINÆ Looss, 1899b, 543, 579; 1902, 839.—Braun, 1901.—Nicoll, 1907, 80.—Pratt, 1902a, 888 (includes: Echinost., Stephanochasmus, Dihemistephanus, Stephanoprora; related genus Rhopalias), 895 (key).

ECHINOSTOMUM. See Echinostoma.

EMISTOMUM Sons., 1889, 192, apparently for Hemistomum.

alatum (Gœze, 1782) Sons., 1889, 192.

EMOLEPTALEA Looss, 1900d, 602, Leptalea Looss, 1899 [nec Klug, 1839] renamed, hence type exilis.—Pratt, 1902a, 889, 901 (key).

ENCHELYS Mueller, 1786.—Nitzsch, 1827, 68 (contains Cerc. podura, C.viridis).—
[Not Enchelys for Echelus Raf., 1810, fish.]

ENCOTYILLABIDÆ Mont., 1888, 108 (for Encotyllabidæ).

ENCOTYLABE Gamb., 1896a, 73 (for Encotyllabe).

ENCOTYLLABE Dies., 1850a, 290, 427–428 (m. nordmanni); 1858e, 313, 364.—Ben. & Hesse, 1864, 66, 79–81.—Braun, 1890a, 412, 415, 469, 477, 498, 511, 515, 516, 519, 523, 525, 526, 529; 1893a, 889.—Carus, 1863, 477.—Gamb., 1896a, 73 (Encotylabe).—Goldb., 1855a, 20.—Massa, 1906, 43, 48.—Mont., 1888a, 7, 10, 13, 16, 66 (Encotyllabe), 86, 87, 97; 1891, 122; 1892, Oct. 7, 213 (gen. of Encotyllabinæ); 1899, 98, 99; 1903, 335 (subf. Encotyllabinæ; fam. Tristomidæ).—Pratt, 1900, 646, 649 (key), 655, fig. 11.—Tasch., 1878, 566 (syn. of Trist. Cuv.).

1888: Encotyllahe Mont., 1888a, 66 (for Encotyllabe).

1896: Encotylabe Gamb., 1896a, 73 (for Encotyllabe).

nordmanni Dies., 1850a, 428 (syn. Trist. excavatum Nord. MS.) (in Brama mediterranea); v. 14, 70, pl. 1, figs. 1–14; 1858e, 364 (in Br. rayi).—Braun, 1890a, 418, 530, 547, 550 (normanni).—Kroyer, 1838–40a, 595 (in Br. rayi Bl.).—Mont., 1891, 122.—Pratt, 1900, 655, 657, fig. 11.—Tasch., 1878, 564, 565, 568 (to Trist.).

normanni Braun, 1890a, 550 (for nordmanni).

pagelli Ben. & Hesse, 1864, 80–81, pl. 7, figs. 1–11 (in Pagellus centrodonotus).— Braun, 1890a, 411, 418, 419, 530, 547, 551.—Cunningham, 1887a, 278.—Gamb., 1896a, 58, fig. 25A.—Mont., 1888a, 7, 15, 66; 1891, 122.—Tasch., 1878, 569 (to Trist.).

species Par. & Perugia, 1890, 6 (in Crenilabrus pavo).—Braun, 1890a, 530, 547, 550. ENCOTYLLABIDÆ Mont., 1888, 87, 88, 97, 108 (Encotyillabidæ); 1891, 108, 122.—

Braun, 1890a, 517.

ENCOTYLLABINÆ Mont., 1892, Oct. 7, 213 (subf. of Tristomidæ); 1903, 335 (fam. Tristomidæ).—Braun, 1893a; 889.—Pratt, 1900, 646 (Encotyllabe), 649 (key).

ENCOTYLLAHE Mont., 1888, 166, misprint for Encotyllabe.

ENODIA Looss, 1899b, 592–593, 633, 710 (m. megachondrus); ἐνόδιος, am Wege; [not Enodia Huebn., 1816, lepidopteron; not Enodia Dahlb., 1843, hymenopteron; not Enodius, Lap., 1836, coleopteron]; 1900, 602 (renamed Enodiotrema Looss, 1900); 1902, 515.—Braun, 1901a, 23; 1901b, 23.—Stiles, 1901r, 189.

megachondrus Looss, 1899b, 593, 709–710, fig. 30, in Testudo (græca?); 1902, 508 (to Enodiotrema).—Braun, 1901a, 23, 24.—Luehe, 1900, 561.

ENODIOTREMA Looss, 1900, Dec. 3, 602, Enodia Looss, 1899 [nec Huebn., 1816], renamed, hence type megachondrus; 1901l, 562, 563; 1902m, 508, 512, 514, 515, 516, 584, 788, 824, 838.—Braun, 1901a, 23; 1901b, 26.—Cohn, 1902, 882.—Heymann, 1905, 93.—Odhn., 1902, 38, 40, 41, 42, 153.—Pratt, 1902a, 888 (related to Plagiorchiinæ), 900 (key).

acariæum Looss, 1902m, 415, 417, 517–519, pl. 24, figs. 50–54 (in Thalassochelys corticata; Egyptian coast), 873, pl. 24, figs. 50–54.

instar Looss, 19011, 562, 563 (in Thalassochelys corticata); 1902m, 516–517, 873,
 pl. 24, figs. 45–46.—Braun, 1902b, 24, fig. 9.

megachondrum (Looss, 1899) Braun, 1901b, 23, 24, fig. 9 (for megachondrus) includes "Monost. cacouanæ Kollar," in Thalassochelys caouana=T. caretta.

megachondrus (Looss, 1899) Looss, 1901l, 561–562, 563 (in Chelone mydas, Thalassochelys corticata; Egypt); 1902m, 414, 415, 417, 508–516, pl. 24, figs. 41–44, 517, 788, 872, pl. 24, figs. 41–44.—Odhn., 1902, 153.

reductum Looss, 19011, 562–563 (in Thalassochelys corticata); 1902m, 517, 518, 519, 873, pl. 24, figs. 47–49.

ENTOBDELLA Audouin, 1828a, 455, "Il paraît que Blainv. a désigné ce genre sous le nom d'Entobdelle."—Braun, 1890a, 518.—Johnston, 1865, 30, 32.

hippoglossi (Mueller, 1776) Johnston, 1856, 32 (syns. Hirudo hip. Mueller, Phylline hip. Lam., Trist. hamatum), on Hippoglossus yulg.

EPIBATHRA Looss, 1902m, 576, 577, 581, 582, 584, 589, 594, 597, 600, 601, 602, 603, 609, 612, 614-615, diagnosis (m. crassa).

crassa (Looss, 1901) Looss, 1902m, 414, 416, 557, 575–578, 615, 876, pl. 26, figs. 83–88, 888, pl. 32, figs. 179–180.

EPIBDELLA Blainv., 1828a, 567 (m. hippoglossi).—Ben., 1858a, 1861a, 11, 18–21, 61, 167, 168, 169, 170, 176, 190, 194.—Ben. & Hesse, 1864, 64, 66, 68–70.—Brand., 1894a, 308.—Braun, 1890a, 412, 414, 415, 423, 429, 451, 453, 465, 466, 469, 473, 484, 490, 495, 511, 515, 516, 517, 518, 519, 522, 523, 526, 527; 1890e, 597; 1891d, 422; 1893a, 889.—Carus, 1863, 477.—Dies., 1850a, 426 (syn. Phylline Oken).—Fraip., 1880c, 416, 445; 1881b, 28.—Gamb., 1896a, 73.—Goto, 1893a, 798.—Haswell, 1892a, 458.—Jackson, 1888, 646 (excretory system), 647 (rep. syst.), 648.—Johnston, 1865a, 32.—Kroyer, 1843–45a, 404 (in Hippoglossus maximus Mind.).—Lint., 1901a, in 267–304, 11 pls.—Maclaren, 1904, 593.—Massa, 1906, 43, 48, 49.—Mont., 1888a, 10, 11, 13, 16, 19, 28, 35, 36, 42, 52 (Epihdella), 55, 57, 66, 84, 86, 87, 97; 1891, 100, 104, 105, 106, 107, 111, 114, 115, 116, 117, 118, 120, 124, 125 (Epidella), 130; 1892, Oct. 7, 172, 186, 187, 213 (gen. of Tristominæ) (syn. Phyllonella); 1893, 8, 210; 1899, 92, 96, 98, 100, 101; 1902b, 137–145, figs. 1–4; 1902c, July 1, 401; 1903, 335 [= Phyllonella] (subf. Ancyrocotylinæ) (f. Tristomidæ); 1905, 75.—Pratt, 1900a, 646, 648 (key) (on skin of marine fishes), 658.—Scott, 1901, 142; 1905, 117 (Epidella).—St.-Remy, 1898, 532.—Tasch., 1878, 566 (syn. of Trist. Cuv.).

1891: Epidella Mont., 1891, 125 (for Epibdella).—Scott, 1905, 117.

1888: Epihdella Mont., 1888, 52 (for Epibdella).

bumpusii Lint., 1900a, 267, 269, 275, 286–287, pl. 34, figs. 11–15 (in Dasyatis centrura; Woods Hole, Mass.); 1901, 414, 433, 461.—Heath, 1902, 112.—Mont., 1902, 137, 138, 140, 141, 142, 143, 145.—Pratt, 1900a, 655, fig. 5, 657, 658 (key).

diadema Mont., 1902, v. 15, 137–145, figs. 1–4 (in Trygon violacea), 143 to (Phylline); 1902, July 1, 401; 1905, 75.

hendorffii (Linst., 1889) Goto, 1894, 233 (in Coryphæna hippurus).—Braun, 1890a, 418, 429, 437, 441, 444, 453, 460, 469, 470, 472, 473, 474, 490, 527, 547, 550.—Heath, 1902, 112 to (Phylline).—Linst., 1904, 678.—Mont.. 1891, 101, 105, 107, 108, 115, 119, 120, 125, 126, 130, pl. 5, fig. 8, pl. 6, fig. 22; 1892, Oct. 7, 172 (hendorfii); 1893, 49; 1902, 137, 138, 142, 143, 144 (hendorfi).

hendorfi Mont., 1902, 144 (for hendorffii).

hendorfii Mont., 1892, Oct. 7, 172 (for hendorffii).

hippoglossi (Mueller, 1776) Blainv., 1828a, 567.—Ben., 1858a, 1861a, 21-23, 36-37, 167, 195, pl. 2, figs. 1-10, pl. 3, figs. 1-8 (includes Tertia pediculorum sp. Baster; Trist. hamatum Rathke) (in Pleuronectes hippoglossus).—Ben. & Hesse, 1864, 69 (in Hippoglossus vulgaris).—Braun, 1889k, 621; 1890a, 411, 419, 423, 438, 441, 444, 445, 453, 458, 475, 483, 488, 519, 527, 547, 551; 1890c, 597.—Dies., 1850a, 427 (to Phylline); 1859c, 437 (to Phylline).—Gamb., 1896a, 55, fig. 22c.—Heath, 1902, 111.—Linst., 1901, 280.—Massa, 1906, 49.—Mont., 1888a, 13, 19, 53 (hippoglossus), 87; 1891, 105, 106, 107, 111, 114, 115, 116, 118, 119, 120, 125, 126, 130, pl. 5, figs. 5, 6, pl. 6, figs. 24, 25; 1892, 0ct. 7, 172; 1899, 96; 1902, 137, 138, 140, 141, 142, 143, 144, 145; 1905, 75.—Odhn., 1905, 370-372 (syn. Trist. hamatum).—Par., 1896, 1 (in Hippog. vulg.; North Sea.)—Schoett, 1896, 253-265, 1 pl.—Scott, 1901, 142 (in Halibut; Scotland); 1905, 117 (Epidella).—Staff., 1904, May 3, 482 (on Hippog. hippoglossus; Canada).—St.-Remy, 1898, 532.—Tasch., 1878, 568 (to Trist.).—Also reported for Hippoglossus maximus.

hippoglossi var. pleuronectes Mont., 1889, 117 (MS. name in Leuck. Coll., renamed Trist. uncinatum).

hippoglossii Ben., 1858a, 1861a, 21 (for hippoglossi).

hippoglossus Mont., 1888, 53, for hippoglossi.

ishikawa Goto, 1894a. 234–235, pl. 26, figs. 1–3 (on Lethrinus: Hagi, Japan, August).—Mont., 1899, 96; 1902, 137, 142, 143.—St.-Remy, 1898, 532–533 (ishikawai).

ishikawai St.-Remy, 1898, 532-533, for ishikawa.

monticelli Mont., 1902, 143 for monticellii.

monticellii (Par. & Per., 1895) Par., 1896, 1 (in Mugil auratus; Italy) to (Phylline).—Mont., 1902, 137, 142, 143 (monticelli).—St.-Remy, 1898, 533–534.

ovata Goto, 1894a, 234–237, pl. 26, figs. 4–8 (in Anthias schegelii; Misaki, Japan, August).—Mont., 1899, 96; 1902, 137, 142, 143.—St.-Remy, 1898, 533.

EPIBDELLA—Continued.

producta Linst., 1903, 354-356, figs. 5-6 (in Solea vulgaris).—Odhn., 1906, 65-66 (syn. of E. soleæ).

sciænæ Ben., 1856, 502–508, 1 pl., figs. 1–4 (on Sciæna aquila; Italy); 1858a, 1861a, 23–37, 195 (in Sc. aq.).—Ben. & Hesse, 1864, 69–70 (E. sciænæ, syn. of Benedenia elegans).—Braun, 1890a, 411, 419, 518, 527, 547, 552.—Fraip., 1880c, 442 (scienæ).—Goto, 1894a, 233; 1899, 269–270, pl. 20, figs. 8–9.—Massa, 1906, 49.—Mont., 1888, 13, 87; 1891, 105, 107, 120, 125, 126, pl. 6, fig. 23; 1893, 4; 1902, 137, 141, 142, 143, 144, 145; 1905, 75.—Par., 1896, 1 (in Sc. aq.; Italy).—Sons., 1891, 263.—Tasch., 1878, 564, 565 (to Trist.).

scienæ Fraip., 1880c, 442 (for sciænæ).

sciænæ Ben. & Hesse, 1864a, 69-70 (for sciænæ, syn. Benedenia elegans).

soleæ (Ben. & Hesse, 1863) Mont., 1890, 419; 1891, 105, 106, 107, 109, 111, 115, 116, 118, 119, 120, 124, 125, 126, 130, pl. 5, fig. 7, pl. 6, figs. 26-30; 1892, Oct. 7, 172; 1902, 137, 138, 139, 140, 142, 143, 144, 145.—Linst., 1903, 356.—Odhn., 1906, 65-66 (syn. E. producta).—Par., 1896, 1 (in Solea vulgaris; France), to (Phyllonella).

squamula Heath, 1902a, 109–136, pls., 15, 16, figs. 1–19 (in Paralichthys californicus); 1902b, 843.—Massa, 1906, 49.—Mont., 1905, 75–76, to (Phylline).—Odhn., 1905, 371.

EPIDELLA Mont., 1891, 125, for Epibdella q. v.

hippoglossi (Mueller, 1776) Scott. 1905, 117.

EPIHDELLA Mont., 1888, 52, for Epibdella q. v.

ERPOCOTYLE Ben. & Hesse, 1863, 87; 1864a, 87 (m. lævis).—Braun, 1890a, 413, 415, 416, 511, 516, 517, 523, 538, 539, 546; 1893a, 890.—Gamb., 1896a, 73.—Hoyle, 1890, 539 (one species from gills of Mustelus lævis).—Mont., 1888a, 8, 11, 13, 86, 89, 100; 1892, Oct. 7, 213 (gen. of Polystominæ); 1903, 336 (subf. Diaphorocotylinæ).—Pratt, 1900a, 646, 651 (key, "on gills of Mustelus"), 656, fig. 24.—Tasch., 1879, 253–254, 255; 1879, 69.

circularis Linst., 1904, 493–494, figs. 18–20 or 1904, 17–18, figs. 1–3 (in Acipenser ruthenus).

lævis Ben. & Hesse, 1863, 1864a, 87–89, pl. 7 bis, figs. 1–9 (in Mustelus lævis).—
Braun, 1890a, 414, 418, 539, 548, 551.—Linst., 1903, 280.—Mont., 1888a, 8, 13.—Pratt, 1900a, 656, fig. 24, 657.—Tasch., 1879, 254 (in Must. læv.).

ERTOPDELLA Rathke, 1843, 238 (for Entobdella?).

ETEROCOTYLEA Mont., 1899, 81, 88, 107 (for Heterocotylea).

EUBUCEPHALUS Dies., 1855a, 395 (m. Bucephalus polymorphus), subg. of Bucephalus.

(EUCERCARIA) Dies., 1855a, 385–388 subg. of Cercaria; 1858d, 246 (renamed Acanthocephala).

brachyura (Dies., 1850) Dies., 1855a, 386; 1858d, 257.

brunnea (Dies., 1850) Dies., 1855a, 387; 1858d, 247.—See Dist. echinatum, type of Echinost.

chlorotica (Dies., 1850) Dies., 1855a, 386; 1858d, 253.

fallax (Dies., 1850 pars) Dies., 1855a, 387-388; 1858d, 248.

minuta (Nitzsch, 1817) Dies., 1855a, 385.

neglecta (Fil., 1854) Dies., 1855a, 386-387; 1858d, 246.

vesiculosa (Dies., 1850) Dies., 1855a, 385; 1858d, 254.

virgula (Fil., 1837) Dies., 1855a, 386; 1858d, 260.

EUCOTYLE Cohn, 1904, 238 (m. nephritica).

nephritica (Mehlis, 1846) Cohn, 1904, 237-238, fig. 4.

EUMEGACETES Looss, 1900, Dec. 3, 602, Megacetes Looss, 1899 [nec Thomas, 1859], renamed, hence type "triangularis" of Looss =emendatus Braun.—Braun, 1901f, 568; 1902b, 53, 92, 93, 94, 95, 96.—Pratt, 1902a, 889 (related to Dicrocœllinæ), 904 (key).

contribulans Braun, 1901f, Apr. 25, 568 (in Hirundo rustica); 1901g, 895, 943;
 1902b, 54, 93, 95, 96, fig. 54.—Poche, 1907, 126 (syn. of crassus).

crassus (Sieb., 1836) Poche, 1907, 126.

EUMEGACETES—Continued.

emendatus Braun, 1901f, 568, new name for Dist. meropis of Par. =Megacetes triangularis of Looss, not D. triangulare Dies.; also in Caprimulgus europæus; 1901g, 895; 1902b, 54, 93, 94, 97.—Looss, 1902m, 819, 823.

medioximus Braun, 1901g, 895-896 (in Galbula grandis Lath; Brazil); 1902b, 96,

figs. 55, 56.

[triangularis of Looss, see emendatus.]

EUPOLYCOTYLEA Dies., 1850a, 289, 416 (subtribe of Polycotylea).—Braun, 1890a, 515.—Cerf., 1899a, 351.—Goldb., 1855, 18.

EURYCŒLIUM Hoyle, 1890, 539, for Eurycœlum.

EURYCŒLUM Brock, 1886a, 543-547 (m. sluiteri); 1887a, 186-187 [not Eurycœlum Chaudoir, 1848].—Brand., 1891b, 266.—Braun, 1892a, 741, 742, 743, 744; 1893a, 886, 887, 894.—Hoyle, 1890, 539.—Looss, 1902, 839.—Luehe, 1901, 395, 481, 482, 484-488.—Mont., 1888a, 43, 92.—Pratt, 1902a, 889, 904 (key).—Spengel, 1892.—Stiles, 1901, 177.—Stiles & Hass., 1898a, 88, 90, 91 (type, sluiteri; syn. of Hemiurus).

1890: Eurycælium Hoyle, 1890, 539, for Eurycælum.

sluiteri Brock, 1886a, 543–547 (in Diacope metallicus; Java); 1887a, 186–187.—
Brand., 1891b, 265.—Braun, 1892f, 45–48; 1892h, 727–729, 741, 742.—Linst., 1887.—.—Looss, 1894, 234; 1899 (to Hemiurus).—Luehe, 1901, 484, 485.—
Mont., 1893, 149.—Stiles, 1901, 177.—Stiles & Hass., 1898a, 90.—Zool. Anz., 1892, v. 15, 225.

(EURYSOMA) Duj., 1845a, 388, 406 (m. squamula), subg. of Dist. [not Eurysoma Koch, 1840, arachnoid; not Gistl., 1829; not Eurysomus Young, 1866].—Baird, 1853a, 52.—Braun, 1893a, 885, 909.—R. Bl., 1891, 609.—Looss, 1899b, 535; 1902m, 750, 751.—Mont., 1888a, 92.—Stiles, 1901, 165.—Stiles & Hass., 1898a, 88, 98.

squamula (Rud., 1819) Duj., 1845a, 406 (type of Eurysoma) (to D. (Dicroccelium) by Stoss., 1892, 20).

EUSTEMMA Dies., 1850a, 287, 317 (m. caryophyllum), 1855, 172; 1858e, 312, 322-323.—Brand., 1888a, 12, 13.—Braun, 1893a, 887, 894.—Goldb., 1855, 17.—Hoyle, 1890, 539.—Mont., 1888a, 84, 91.

caryophyllum Dies., 1850a, 317 (in Falco pileatus; Rio Parana, Brazil); 1855, 172,
pl. 1, figs. 1-5; 1858e, 323.—Brand., 1888a, 5, 65 (syn. of Holost. eustemma);
1890a, 593 (syn. of Holost. eustem.).—Braun, 1893a, 903.

EURYTREMA Looss, 1907, Feb. 1, 127–134 (tod. pancreaticum Janson; Japan).

cælomaticum (Giard & Billet, 1892) Looss, 1907, Feb. 1, 132–134, fig. 2 (syn. Dicrocælium pancreaticum Rail. & Marotel, 1898).

pancreaticum (Janson, 1889) Looss, 1907, Feb. 1, 128–132, 133, fig. 1.—[Janson, 1889a, is not accessible to us; our earliest reference is Railliet, 1890.]

EXACOTYLE Mont., 1888a, 8 (apparently for Hexacotyle); 1891, 109.

thynni De la Roche, 1811a?—Crety, 1892c, 399 (tynni).—Mont., 1890, 195; 1891, 129. tynni Crety, 1892c, 399 (for thynni).

EXACOTYLINÆ Mont., 1905, 77.

FAICIOLA Mueller, 1774, 70, for Fasciola.

FASCINIA Raffinesque, 1815, 151 (nomen nudum; g. of Fasciolaria; to contain species of Fasciola, but these are not named).

FASCIOLA Linn., 1758a, 644, 648–649, includes hepatica (type by elimination, also by later designation, and also by first-species rule) and intestinalis.—Abildg., 1790, 35 (syns: Fasc. intestinalis Linn., Planaria intest. Mueller, Ligula Bloch, Fasc. Gœze), 36 (to Dist.), 58.—Ariola, 1900, 426 (of Linn., 1735, — Schistocephalus).—Biehringer, 1889a, 654.—Blainv., 1824a, 514; 1828, 585–587.—E. Bl., 1847, 278.—R. Bl., 1888a, 589; 1895, 730.—Bosc., [1802a], v. 1, 263–268.—Brand., 1888a, 8.—Braun, 1892a, 674; 1893a, 883, 885, 893, 894, 908; 1895b, 138; 1900a, 1643, 1668; 1903, 3 ed., 147.—Cobbold, 1879b, 15, 1883, 401 (syn. of Dist.); 1883x, 514; 1883v, 500.—Crep., 1837a, 309; 1845b, 20–22.—Cuv., 1817a, 40.—Deslongchamps, 1824o, 386.—Dies., 1850a, 307 (of Gmelin, syn. of Hemist.), 312 (of Gmelin, syn. of Holost.), 318 (of Schrank; syn. of Diplodiscus), 319 (of Gœze, syn. of Monost.), 331 (of Linn., syn. of Dist.), 400 (of Mueller, syn. of Amphist.), 411 (of Frælich, syn. of Notocotyle), 412 (of Gmelin, syn. of Polyst.), 573 (of Linn., syn. of Tetrabothriorhynchus), 577 (of Gœze, syn. of Caryophyllæus), 579 (of authors, syn. of Ligula), 583;

1858e, 332 (syn. of Dist. giganteum).—Fabricius, 1794, 26, 27.—Fischer, 1799, 96.—Hahn & Lefèvre, 1884a, 515.—Hémont, 1827, 9.—Herbst, 1787a, 14, 15, 21, 31, 36.—Hoyle, 1890, 539–540 (3 species, type hepatica).—Jackson, 1888, 644, 646, 654.—Joubin, [1892a], 29.—Lamarck, 1801a, 333; 1816, 180–181.—Lamouroux, 1824a, 559, 608; 1824b, 404; 1825a, 356.—Leblond, 1836f, 4.—R. Leuck., 1863, 530.—Looss, 1899b, 551, 557; 1901, 199, 208, 658; 1902, 746, 756.—Luehe, 1905, 146.—MacCallum, 1899, 707.—Macé, 1882, 25.—Montagu, 1811, 194–198, 199–200 (in poultry, see Syngamus trachealis).—Mont., 1893, 44, 153, 182; 1896, 168.—Mueller, 1774, 52, 70 (Faiciola).—Nitzsch, 1819, 397.—Nord., 1840, 613–618 (syn. Dist.).—Odhn., 1905, 339, 344, 346, 347.—Pallas, 1781c, 59.—Par., 1900, 190–197, 1 fig. (in Buenos Ayres).—Pratt, 1902, 883, 887 (key to).—Rafinesque, 1815, 151 (g. of Fasciolaria).—Rail. & Marotel, 1898, 32–33.—Ræderer, 1762, 537–539.—Ræderer & Wagler, 1762, 194–210.—Rud., 1793a, 25; 1801, 50, 54; 1802, 61–62; 1809a, 21.—St. Vincent, 1824, 608.—Schneider, 1866, 7, 9, 11.—Schrank, 1803, 185.—Slawikowski, 1819, 8, 55, 56.—Sons., 1896, 112–116.—Stiles, 1894c (s. str.); 1895l; 1895m; 1896, 159; 1898a, 22, 27, 29; 1901, 163, 165, 174, 180; 1906, 14.—Stiles & Hass., 1898a, 88–89, 90, 95, 97 (type hepatica).—Tasch., 1879, 251 (of Gmelin, syn. of Polyst.).—Vogel, 1772, 648.—Ward, 1903, 865.—Wolffhuegel, 1900, 129.

1774: Faiciola Mueller, O. F., 1774, 70, misprint.

1782: Planaria Gceze, 1782a, 168, [not Mueller, 1776, worm; not Brown, 1827, mollusk; not Lea, 1833, mollusk], includes Fasc. hepatica as Pl. latiuscula, hence type P. latiuscula Gceze, 1782.

(1782): Distoma Retzius (1782); 1790, 32 [not Distomus Gærtner, 1774, mollusk; not Steph., 1827, coleopteron; not Distoma Savigny, 1816, mollusk], for Planaria Gæze, 1782, hence same type.

1815: Distomopsis Rafinesque, 1815, 151, Distoma Zed., renamed.

1825: Distomum Crep., 1825, for Distoma.

1845: Distoma (Cladocœlium, m. hepaticum) Duj., 1845a, 382, 388, 389.

1845: Fasciolaria Anonymous, 1845. 141, in Encycl. Metropolitana, Lond. [not Lamarck, 1799, mollusk], for Fasciola Linn.

1850: "Distomum Retzius," 1782, in Dies., 1850a, 141.

1853: "Cladocalium Duj." of Pontallić, 1853, 103-105 (for Cladoccelium).

1863: "Distomum (Fasciola)" R. in Leuck., 1863, 530.

1876: Fascolia Adams, 1876, 764, misprint.

1892: Cladocœlium (Duj., 1845) as genus in Stoss., 1892, 7 of reprint.

1894: Phasciola Wilder, H. H., 1894, 24 [not Phaseolus Mont., 1875, mollusk] (for Fasciola).

1898: "Clacoccelium" of Stiles & Hass., 1898a, 89, misprint.

abdominalis Gœze. 1782a, 41, 169, 186–190, pl. 16, figs. 4–9, includes F. intestinalis, 1758, and Ligula piscium Bloch.—Baird, 1853a, 95 (syn. of Lig. simplicissima).—Dies., 1850a, 580 (syn. of Lig. diagramma).—Nord., 1840, 590 (syn. of Lig. cingulum; F. a. of Zed., syn. of Lig. contortrix).—Rud., 1810a, 20 (syn. of Lig. cingulum), 18 (syn. of Lig. contortrix), 29 (syn. of L. carpionis).

æglefine Nicoll, 1907, 73 (syn. of Dist. simplex) for æglefini.

æglefini Mueller, 1776, 224 (in Gadus æglefinus; intestine); 1777, 33, pl. 30, fig. 4
(in Ga. ægl.); 1779a, 65 [pl. 3, fig. 4].—Bosc, 1802a, 272 (ægrefini, misprint).—
Bruguière, 1791a, pl. 7a, fig. 15.—Cobbold, 1858b, 157.—Dies., 1850a, 343
(syn. of Dist. simplex).—Gmelin, 1790a, 3056.—Herbst, 1787a, 33.—Lamarck, 1816, 182; 1840, 619.—Lamouroux, 1824a, 563.—Levin., 1881a, 67 (syn. of Dist. sim. Rud.? Olss.).—Nord., 1840, 619 (syn. of Dist. sim.).—Odhn., 1901, 512.—Rud., 1809a, 370.—Schrank, 1788, 19.—Type of Sinistroporus 1904.

ægrefini Bosc, 1802a, v. 1, 272, misprint for æglefini.

agyptiaca (Looss, 1896)
Sons., 1896, 112; 1896, 1.—Braun, 1903, 3 ed., 152.—
Looss, 1899b, 557; 1902m, 782, 783 (syn. of F. gigantea).—Ward, 1903, 866.
alata (Geze, 1782)
Rud., 1793a, 31; 1795a, 15; 1802, 84–85; 1809a, 403 (to Dist.).—

Brand., 1888a, 9-60 (to Hemist.).—Dies., 1850a, 308 (to Hemist.).

alosæ Hermann, 1783a, 46, pl. 2, fig. 8a-b (in Maifisch im Rhein, Clupea alosa).—Baird, 1853a, 54, syn. of Dist. appendiculatum).—Dies., 1850a, 370 (syn. of Dist. appen.).—Rud., 1809a, 401, 437 (= Dist. clupeæ rhenanæ).

americana Hass., 1891c, 539 (carnosa renamed) (in Bos taurus, U. S. A.).—Leuck., 1892b, 797.—Stiles, 1898a, 49.—Ward, 1895, 253 (syn. of F. magna), 332 (in Bos taurus), 335 (in Ovis aries), 338 (in Equus caballus); 1903, 866 (syn. of

F. magna).

anatis (Schrank, 1788) Gmelin, 1790a, 3055 (based especially upon a parasite (Plan. teres pro parte Gœze, 1782a, 174, pl. 13, figs. 8–11) from Anas boschas domestica and made to include Cuculanus conoideus Bloch, 1782a, and Hirudo fasciolaris Mueller).—Bosc, 1802a, v. 1, 269–270.—Dies., 1850a, 383 (syn. of Dist. echinatum).—Rud., 1809a, 418, 431-432 (anatis Bruguière, 1791 = anatis domesticæ).—Stoss., 1892, 167 (syn. of Echinost. echinatum).

anguillæ Gmelin. 1790a, 3056 (refers to Leeuwenheek arc. nat., p. 316, fig. 6) (in Anguilla).—Bosc, 1802a. v. 1, 272.—Dies., 1850a. 340 (syn. of Dist. polymorphum) (in Anguilla vulgaris (Leeuwenheek), May, August; Greifswald).—Lamarck, 1816, 182.—Nord., 1840, 618 (syns. Dist. polymorphum and D. an-

guillæ).—Rud., 1809a, 363.

angulata Mueller, 1774, 58-59 (in fundo arenareo oceani).—Fabricius, 1798, 53 (to

Plan.).—Verrill, ——, —— (to Amphiporus).

angusta (Rail., 1895) Looss, 1899. 557.—Braun, 1903, 3 ed., 152, fig. 98 (Gouvea's case in French naval officer at Rio; had been in Senegambia) thinks = ægyptiaca.—Looss, 1899b, 557; 1902m, 782, 783 (syn. of gigantea).—Ward, 1903, 704; 1903, 864, 866.

- anseris Gmelin. 1790a. 3055 (F. verrucosa Frœlich, 1789, renamed; in Anser domesticus; Europe).—Baird, 1853a, 45.—Ben.. 1858a, 1861a. 78 (syn. of Monost. verrucosum).—Bosc. 1802a, v. 1. 270.—Dies., 1840, 234: 1850a, 411 (syn. of Notocotyle triseriale).—Nord., 1840, 602 (syn. of Nocotylus triserialis).—Odhn., 1905, 366 (syn. of Catatropis verrucosa [type]).—Rud., 1809a, 331.
- apiculata Rud., 1803, 31–32, Dist. stridulæ Reich, 1801, renamed; 1809a, 423, to (Echinost.).—Dies., 1850a, 386.
- appendiculata Rud., 1802, 78-79, pl. 2, figs. 6a-b (nec Frœlich, 1802) (in Clupea alosa; Europe); 1809a, 400.—Baird, 1853a, 54.—Dies., 1850a, 370.—Luehe, 1901, 396.—Stiles & Hass., 1898a, 90, 96 (type of Hemiurus).
- appendiculata Freelich, 1802, 56–58, pl. 2, figs. 8–9 (nec Rud., 1802) (in Anas boschas; Europe).—Dies., 1850a, 346 (syn. of Dist. oxycephalum).—Hahn & Lefèvre, 1884a, 516 (syn. of Dist. oxyceph.).—Rud., 1814a, 105, renamed Dist. papillatum.

apri, see sub F. hepatica.

ardeæ Gmelin, 1790a, 3055 (in Ardea stellaris, int.: Europe).—Dies., 1850a, 388 (syn. of Dist. ferox).—Bosc, 1802a, v. 1, 270.—Rud., 1809a, 432.

armata Rud., 1793a. 26, F. putorii, 1790, renamed; 1802, 88–90; 1809a, 416.—Dies., 1850a, 382 (syn. of Dist. trigonocephalum).

atomon Rud., 1802. 70 (in Pleuronectes flesus: Greifswald); 1809a, 362.—Dies., 1850a, 340 (to Dist.).—Odhn., 1901, 506; 1905, 320 (to Podocotyle [type]).

barbata Linn., 1761, 505 (in Loligo; Sweden).—Abildg., 1790, 38.—Baird. 1853a, 115 (to Tetrabothriorhynchus).—Braun, 1893a, 883.—Dies., 1850a, 573 (syn. of Tetrabothriorhynchus migratorius).—Fabricius, 1780, 328-329.—Herbst, 1787a, 32.—Luche, 1905, 334 (=Tetrarhynchus).—Mueller, 1776, 223; 1780, 203.—Pallas. 1781, 109.—Rud., 1809a, 441; 1810a, 385; 1819a 130 (syn. of Tetrarhynchus megabothrius).—Sieb., 1850, 232 (syn. of Tetrarhynchus macrobothrius).—Vaullegeard, 1899, 167.

bergi Par., 1900, 193–194, 1 fig. (in Rana platana; Montevideo).

[bilamellata Pallas, 1774, 20 ("Fasciolæ bilamellatæ Linnæi").]

bilis (Braun, 1790) Gmelin, 1790a, 3054 in Falco melanaëtus.—Bosc, 1802a, v. 1,
 269 (billis).—Dies., 1850a, 376 (syn. of Dist. crassiusculum).—Rud., 1809a,
 408.

billis Bosc, 1802a, v. 1, 269, for bilis.

binodis Mueller, 1776, 224 (in fish); 1779a, 69 [pl. 30, fig. 8]; 1788, 34–35.—Bosc, 1802a, 271–272.—Bruguière, 1791, pl. 79, fig. 24.—Dies., 1850a, 379 (to Dist.).—Gmelin, 1790a, 3056.—Herbst, 1787a, 33.—Rud., 1809a, 439.—Schrank, 1788, 20.

blennii Mueller, 1776a, 224 [in Blennius viviparus]; 1777, v. 1, 32, 33, pl. 30, fig. 5, v. 2, 53, pl. 78, figs. 9–12 (in Cottus scorpius and Blennius viviparus); 1779a, 64, 66 [pl. 30, fig. 5]; 1780, 217; 1784, 116, pl. 28, figs. 9–14.—Bosc, 1802a, v. 1, 272.—Bruguière, 1791a, pl. 79, figs. 16–18.—Dies., 1850a, 344 (syn. of Dist. divergens).—Gmelin, 1790a, 3057.—Herbst, 1787a, 33.—Lamarck, 1816, 183.—Nord., 1840, 619 (syn. of Dist. divergens Rud.).—Rud., 1793a, 30; 1809a, 371.—Schrank, 1788, 19.

boschadis Schrank, 1803, 209.

boum Gmelin, 1790a, 3054, see sub hepatica boum.

bramæ Mueller, 1776, 224, v. 1, 33, pl. 30, fig. 6 (in Cyprinus brama; intest. Norway, Denmark); 1779a, 66 [pl. 30, fig. 6].—Bosc, 1802a, v. 1, 274.—Bruguière, 1791, pl. 79, fig. 19.—Cobbold, 1858b, 157.—Dies., 1850a, 341 (syn. of Dist. globiporum).—Gmelin, 1790a, 3058.—Herbst, 1787a, 33.—Looss, 1894a, 41 (syn. of Dist. gl.); 1902m, 757 (and Dist. gl.), 758, 760 (perhaps an Allocreadium, Bunodera, or Asymphylodora), 763, 765.—Nord., 1840, 619 (syn. of Fasc. globifera).—Rud., 1809a, 364.—Schrank, 1788, 20.—Sramek, 1901, 107 (syn. Dist. gl.).—Stiles, 1901, 168, 169, 192.—Stiles & Hass., 1898a, 94, 95, 97 (= globipora, type of Sphærost.).

brunnea Mueller, 1774, 54.—Johnston, 1865, 9.

buchholzii "Jærdens, 1801," of Braun, 1889a, 320 (lapsus for "Fasc. hepatica, ovata plana, Buchholzii"=Dicroccelium lanceatum).—Stiles, 1898a, 55.

buteonis Gmelin, 1790a, 3054 (in Falco buteo) based on Gœze, 1782a, 85.—Bosc, 1802a, v. 1, 269.—Rud., 1809a, 430 (=Dist. buteonis (in Falco buteo)).

candida Mueller, 1774, 71 (in littore Grænlandiæ sub lapidibus).—Verrill, —— (?syn. of Tetrastemma candidum (F.) Oersted).

capitata Mueller, 1774, 70.

carnosa Hass., 1891a, 208–209, 1 fig. (in Bos taurus; U. S. A.); 1891b, 464–465;
1891c, 359 (syn. of F. americana).—Huber, 1896a, 575, 576.—Leuck., 1902b,
797, 798.—Stiles, 1898a, 49.—Ward, 1895, 253 (syn. of F. magna), 332 (in Bos taurus), 335 (in Ovis aries), 338 (in Equus caballus); 1903, 866 (syn. of F. magna).

caudata Mueller, 1774, 70.—Buttel-Reepen, 1902, 166, 170, pl. 6, fig. 6.—Schrank,

1803, 171 (=Plan. acuminata).—Slawikowski, 1819, 38.

caudata Bosc 1802a, v. 1, 271, pl. 9, fig. 6 (nec Mueller, 1774) (in dorade=? Chrysophys aurata: Coryphæna hippurus).—Dies., 1850a, 373 (syn. of Dist. tornatum); 1859c, 431 (syn. of D. tornatum).—Rud., 1809a, 436 (=Dist. coryphænæ); 1819a, 685.

cerri (Schrank, 1790) Zed., 1790, 65–74, pl. 3, figs. 8–11.—Dies., 1836, 247; 1850a, 401 (syn. of Amphist. conicum).—Fischder., 1903h, 504 (type of Paramphist.), 506 (in Cervus elaphus).—Rud., 1809a, 351.—Schrank, 1803, 214.—Stiles, 1898a, 64.—[Type of Paramphist. 1901.]

ciliata Mueller, 1774, 55 (free form).

cincta Rud., 1803, 31 (in Tringa vanellus); 1809a, 422, to D. (Echinost.).—Dies., 1850a, 386 (to Dist.) (in Vanellus cristatus, V. melanogaster).

[cinerea (anatomical term).]

cirrata [Rud., 1802 (see Dist. cirratum)]; 1809a, 376 to Dist.—Dies., 1850a, 350 (to Dist.) (Griefswald, June).

cirrhata Rud., 1802, v. 3 (1), 66-67, pl. 2, fig. 4 (in Corvus frugilegus).—Braun,

1902b, 43 (syn. of Plagiorchis cirratus).

clavata Menzies, 1791, 187–188, pl. 17, fig. 2 (in Scomber pelamys; Pacific) [type of Hirudinella, 1828]; 1794, 58–59, pl. 8, fig. 2.—Baird, 1853a, 59.—Blainv., 1824a, 518 ("hirudinelle"); 1828, 586.—Buttel-Reepen, 1900a, 586; 1902, 166, 168, 178, pl. 6, fig. 3.—Cobbold, 1879b, 459.—Darr, 1902, 663.—Dies., 1850a, 366 (to Dist.); 1859c, 431 (Dist. clavata Rud., partim).—Rud., 1809a, 391.—Stiles, 1901, 192.

clupex Schrank, 1788, 20 (in rheinischen Mayfische).—Baird, 1853a, 54.—Dies., 1850a, 370 (syn. of Dist. appendiculatum).—Rud., 1809a, 437.

collurionis Schrank, 1790, 123 (in Lanius collurio); 1803, 211.—Dies., 1850a, 396 (to Dist.) (in Lanius collurio).—Rud., 1809a, 430 (to Dist.).

8588—No. 37—08——17

colubri Bosc, 1802. v. 1, 271. pl. 9, figs. 1–3 (in couleuvre d'Amérique).—Cobbold, 1859d, 364 (syn. of Dist. bosci).—Dies., 1850a, 398 (syn. of Dist. colubri americani) (in Coluber sp. americana).—Rud., 1809a, 434 (to Dist.).

colymbi immeris Viborg. 1795, 241 ["Fasciola s. Ligula].—Dies., 1850a, 579.—

Rud., 1810a. 27 to Ligula .

coruphænæ Bosc, 1802a, v. 1, 271, pl. 9, fig. 5 in dorade).—Baird, 1853a (=Hirudinella clavata), 59.—Buttel-Reepen. 1902, 166, 168, 170, pl. 6, fig. 5.—Dies., 1850a. 373 syn. of Dist. tornatum; 1859c, 431.—Rud., 1809a, 436 to Dist.; 1819a. 685.

coryphænæ hippuridis Tilesius in litteris Rud., 1809a, 436 (syn. of D. coryphænæ), 437.—Baird. 1853a, 59 (=Hirudinella ventricosa).—Buttel-Reepen, 1902, 166, 168.—Cobbold, 1879b, 461.—Dies., 1850a, 373 (syn. of Dist. tornatum).

crassa Rud., 1793a, 27 = Fasc. melis Gmelin, renamed).

crenata Rud., 1802.76-78, pl. 2, fig. 5, a-b/in Gasterosteus aculeatus, Pleuronectes maximus: 1809a. 404 (in Gast. acul.. Pleur. max.).—Baird, 1853a, 54 (=Dist. appendiculatum).—Dies., 1850a, 371 (syn. of Dist. app.).—Lander. 1904a. 1 (to Hemiurus).—Luehe, 1901, 399, 400.—Odhn., 1905, 352 (type of

crenata Freelich. 1802a. 60–62. pl. 2, figs. 10–11 (nec Rud. (in Fulica chloropus)).—Dies.. 1850a. 384 (syn. of Dist. uncinatum) (in Gallinula chl., May & July).—Rud., 1814a. 102 (=Dist. unc.).

crenata Mueller, 1774, 64 / nec Rud., nec Frælich .

cucumerina Rud., 1804, 166 (in liver of Pleuronectes maximus).

cygnoides Schrank, 1803. 212.

cylindracea Zed., 1800 Rud., 1802, 83-84; 1809a, 393 in Rana esculenta, R. temporaria .—Lamarck, 1816, 183.—Looss, 1894a, 64 to Dist.).—Nord., 1840, 620 (to Dist.).—Stiles & Hass., 1898a, 84.

cyprini carassii Viborg, 1795, 242.

delicatulum Rud., 1809 Blainv., 1828a, 585.

denticulata Rud., 1802, 91-92, pl. 2, fig. 8, a-c (in Sterna hirundo): 1809a, 424.—

Dies., 1850a, 392 to Dist.).

disticha Mueller, 1776a, 224; 1788, 35, pl. 30, fig. 9; 1779a, 69, pl. 30, fig. 3 [sic]= fig. 9.—Bosc, 1802a, v. 1, 272.—Bruguière, 1791a, pl. 79, fig. 25.—Dies., 1850a, 378 (to Dist.).—Gmelin, 1790a, 3056.—Herbst, 1787a, 33.—Rud., 1809a, 367, 440 (to Dist.).—Schrank, 1788, 20; 1803, 212-213.

echinata Nord., 1840. 621 (syn. of F. trigonocephala).

elaphi Gmelin, 1790a, 3054 (Festucaria cervi, renamed) (in Cervus elaphus).— Bosc, 1802a, v. 1, 269.—Dies., 1836, 247; 1850a, 401 (syn. of Amphist, conicum).—Fischder., 1903h. 504 (syn. of Paramphist. cervi), 506.—Nord., 1840, 627 (syn. of Amphist. con.).—Řud., 1809a. 350.—Stiles. 1898a, 64.—Ward, 1895. 256 (syn. of Amphist. conicum). 332 (in Bos taurus). 335 (in Ovis aries).

elegans Rud., 1802, 65-66 (in Sperling, Greifswald),—Braun, 1902b, 38 to Plagiorchis.—Dies., 1850a, 350 to Dist. .—Rud., 1809a, 375.

epatica Rosa, 1794, 5, dell' Ardea purpurea.—Braun. 1900h, 19 (? syn. of Clinost. heterost.).—Dies.. 1850a, 353 (syn. of Dist. heterost.).

epatica Brera, 1809a, 92-98, pl. 1, figs. 24-25.—Sons., 1884, 21.—For hepatica.

equi Gmelin, 1790a, 3054. var. of Fasc. hepatica.

criocis Mueller, 1784, 92, pl. 72, figs, 4–7; 1788, 42, pl. 72, figs, 4–7 in Salmo eriox.—Bosc, 1802a, v. 1, 274.—Bruguière, 1791a, pl. 80, figs, 3–4.—Dies., 1850a. 363 (syn. of Dist. hyalinum).—Gmelin, 1790a. 3057.—Harz, 1881c, 5 (syn. of D. hyalinum).—Lamarck, 1816, 183.—Nord., 1840, 619–620 (includes D. hy., D. rosaceum, D. perlatum).—Rud., 1809a, 389.

excavata Rud., 1803, 28-29 (in Storch, Ardea ciconia; Greifswald); 1809a, 399.— Dies., 1850a, 309–310 to Hemist.).

eccisa Rud., 1819 Blainy., 1828a, 586.

farionis Mueller, 1784, 91. pl. 72. figs. 1–3 (in Salmo fario); 1788, 42. pl. 72. figs. 1–3.—Bosc, 1802a, v. 1, 274.—Bruguière, 1791a, pl. 80, figs. 1, 2.—Dies., 1850a, 380 (syn. of Dist. laureatum).—Frœlich, 1802a, 62–63, pl. 2, figs. 4, 5.—Gmelin, 1790a, 3058.—Knoch, 1862, 19.—Nord., 1840, 621 (syn. of F. laureata).—Odhn., 1905, 353.—Rud., 1809a, 413; 1814a, 102 (syn. of Dist. laureata). reatum).

feror Rud., 1795a, 15-17 (in Ardea ciconia; Greifswald); 1802, 90-91; 1809a, 426 to D. (Echinost, .—Dies., 1850a, 388 (to Dist.).—Nord., 1840, 622 syn. of

Fasc. trigonocephala.

fimbriata Gozze. 1782a. 180, pl. 15. figs. 4–5.—Abildg., 1790, 32 (cf. Phylline).—Baird, 1853a, 97 (syn. of Caryophyllæus mutabilis).—Ben., 1858a, 1861a, 115 (syn. of Car. mutab.).—Braun, 1894.—Dies., 1850a, 577 [Fasciolaria].—Nord., 1840, 633 (syn. of Car. piscium).—Rud., 1810a, 9 (ac. mut.).

flaccida Mueller, 1774, 57-58 free form.

furcata (Bremser, 1819) Blainv., 1828, 586,

fusca Pallas, 1774, 21-22. pl. 1. fig. 13. a-b (in aq. stag.: Europe).

fusca Bosc, 1802a. v. 1. 271. pl. 9. fig. 4 in dorade [not Pallas, 1774].—Baird, 1853a. 59 = Hirudinella ventricosa .—Buttel-Reepen. 1902. 166. 168, 170. pl. 6. fig. 4.—Cobbold. 1879. 459, 460. 461.—Darr. 1902. 664.—Dies., 1850a, 366 syn. of Dist. clavatum: 1859c, 431.—Herbst. 1787a. 36.—Poirier, 1885, (to Dist.).—Rud., 1809a, 436.

gibbosa Rud., 1802b, 81-82, pl. 2, fig. 7 in Esox belone : 1809a, 399 to Dist.-

Odhn., 1905. 356 to Lecithaster .

gigantea (Dies., 1858) Cobbold. 1858; 1860a. 4 in Camelopardalis giraffa; 1864, 161 (see F. gigantica); 1876. 303; 1879b. 25. 322, 421.—R. Bl., 1895. 733-734 includes F. hepatica var. angusta: 1900. 488.—Braun. 1893a.—Dies., 1858.—Fitz, 1876b. 514.—Gervais et Ben., 1858.—Hoyle, 1890. 539.—Leuck., 1863; 1879-93.—Looss, 1899b. 557; 1902m. 782, 783 (syns. gigantica, angusta, ægyptiaca).—Sons., 1889. 275; 1890; 1896. 113. 114.—Stiles, 1895. 139-143, pl. 5 (includes Dist. hepaticum pars Gervais & Ben.; 1898a, 49.—Stoss., 1892, 9 (to Cladocœlium).—Also reported for Bos taurus.

gigantica Cobbold, 1855a, 262-266, pl. 7. figs. 1-15 in liver of Giraffa camelo-pardalis; 1855b, 3-7, pl. 7. figs. A. B., 1-15; 1856a, 108-109; 1858b, 167; 1860a, 4.—Braun, 1903, 3 ed., 152 (places Gouvea's (1895) case here); 1906, 157 (in giraffe).—Dies., 1858c, 332 (to Dist.).—Stiles, 1868a, 23, 29, 48, 49, 50, 51, 137, 140, 141, fig. 27.—Ward, 1903, 866 (closely related to F. angusta).

glauca Mueller, 1774, 60.

globifera Lamarck. 1816, 182 for globipora 1802.—Baird, 1853a, 53.—Lamouroux, 1824a, 563.—Nord., 1840, 618-619 includes Dist. globiferum. D. globiporum, Fasc. bramæ.

globipora Rud., 1802, 72-74 (in Cyprinus erythrophthalmus); 1809a, 365.—Dies.,
 1850a, 341 (to Dist.).—Looss, 1894a, 41 (to Dist.); 1902m, 763.—Stiles, 1901,
 168, 169, 192.—Stiles & Hass., 1898a, 95 (type of Sphærostoma).

grossa Mueller. 1774, 67.—To Planaria in 1776.

gruis Gmelin, 1790a, 3055 (in gruis).—Baird, 1853a, 55 (=Dist, echinatum).—Bosc, 1802a, v. 1, 270.—Dies., 1850a, 383 (syn. of Dist, echinatum).—Rud., 1809a, 432.

gulo Mueller. 1774. 56 | free from .—Schrank. 1803. 166 | to Plan. .

halecis Gmelin, 1790a, 3058 (in halece).—Dies., 1850a, 372 syn. of Dist.ocreatum !— Mont., 1891, 496.—Rud., 1809a, 398 syn. of D. ocr. .

helluo Mueller, 1774, 64-65.—Johnston, 1865, 13.—Schrank, 1803, 169 to Plan.

hepatica Linn., 1758a, 648-649 (in aquis dulcibus ad radices lapidum, inque hepate pecorum. Diss. de Ovibus: Europe): 1761, 505; 1766, 1077; 1792, 3053-3054; 1801, 33-34.—Abildg., 1790, 36 (to Dist. —Adams, 1887, 318.—Adenot, 1863, 112.—Aitken, 1866, 804, 839; 1872, 146, 205; 1874, 58.—Aldrovandus, 1602).—Anaker, 1892c, 94.—Andral, 1829d, 615.—Andry, 1701, 121-122.—Ariola, 1900, 426 (of Linn., 1735, syn. of Schistocephalus dimorphus).—Armatage, 1895, 429, 430, 432, 433, 440.—Australian Pastorialists Rev., 1896, v. 5, 610-611.—Baillet, 1866b, 15, 18, 90, 99 (to Dist.).—Baird, 1853, 49 (pars—Schistocephalus dimorphus).—Barbut, 1783, 14-15, pl. 2, fg. 1.—Bauhin, 1677.—Ben., 1858a, 1861a, 172.—Bert & R. Bl., 1885, 585-586, fg. 539.—[Bidloo, 1698a.]—[Biggs, 1890a, 36-37.]—Blainv., 1828, 585.—[Blakeway, 1879a.]—E. Bl., 1847, 279-291.—R. Bl., 1888a, 543 (to Dist., 602 of Bloch, 1782, syn. of Dist. lanceolatum): 1890, 66-75, fgs., 28-33; 1894z, 461-462; 1894h; 1895, 730-733, fgs., 78-79; 1900g, 488.—Bloch, 1782a, 5-6, pl. 1, figs. 3-4.—Blumenbach, 1825a, 241.—[de Bonis, 1876a, 155.]—Borlase, 1758a.—Bosc [1802a], v. 1, 268.—[Bonvicini, 1881a, 133-134.]—Braun, 1893a, 883;

OLA—Continued.

1893b, 180; 1900b, 9; 1901e, 327; 1903, 3 ed, 147-151, figs. 92-97 (syns. Dist. hep. Retz., Fasc. humana Gmelin, Dist. caviæ, Cladocælium hep. Stoss), 150; 1906, 140, 142, 143, 145, 150-156, 157, figs. 74, 77, 81, 83-88.—Brett, 1881b, 139-142.—[Brown, 1881a; 1882b; 1886b.]—Bruguière, 1791, pl. 79, figs. 1-8.—[Bruck, 1865a, 33-35.]—de Bry, 1879.—Chabert, 1787a, 25 (63-64); 1791, 152.—Civini, 1842.—Clerc, 1907, 557, 558.—Cobbold, 1855a; 1855b, 3, 4, 6; 1866, 6; 1872b, 91; 1876, 210, 211, 303; 1879b, 14, 25, 28, 48, 50, 315, 317, 318, 322-331, 356, 404, figs. 17, 61; 1882, 699-704, pl. vi, figs. 2-8; 1883v, 500; 1884g, 976.—Cuvier, 1798, 633-634; —, v. 2, 339; 1817a, 41-42; 1830, 263-264; 1831a, 364.—Daldorf, 1793, 159 (hepatum).—Dandolo, 1806, 28 pp.—Darr, 1902, 649, 652, 657, 663, 684, 687.—[Daubenton, 1750-52.]—Dav., 1877a, 2 ed., 1xxv. figs. 36-38.—[Delafond, 1854a, 3-56, figs. 12-13.]—Delorme, 1861. 2 ed., lxxv, figs. 36-38.—[Delafond, 1854a, 3-56, figs. 12-13.]—Delorme, 1861, 241.—Desmars, 1762.—[Dickens, 1830a, 645–647.]—[Didry, 1832a, 139–147.]— Dies., 1850a, 332 (to Dist.), 333 (of Bloch, syn. of Dist. lanceolatum), 401 (of Dies., 1830a, 332 (to Dist.), 333 (of Bloch, syn. of Dist. lanceolatum), 401 (of Mueller, syn. of Amphist. conicum), 584 (pars of Linn., syn. of Schistocephalus dimorphus).—Dinwiddie, 1892a, 3.—Dowker, 1882a, 10, 11, 12, 13, pl. 1, fig. B. 1, pl. 2 E, figs. 1-2.—Drosse (1856a).—Dunglison, 1893a, 424, 821.—Eber, (1798a), 22.—[Ellis, 1749.]—[Faber, 1670, 147.]—Fabricius, 1780, 327–328 (in sheep); 1794, 29.—Fil., 1854a, 27.—Fischder., 1903h, 504 (of Mueller, syn. of Paramphist. cervi).—La Fosse, 1772, 157; 1774, 320; 1779, 139.—Freelich, 1802a, 55–56.—Gabucinus, 1597.—Galli-Valerio, 1898b; 1898c, 7, 8.—Gamb., 1896a, 67.—[Gemma, 1575a, 40.]—Gerlach, 1854; 1862.—[Gesnerus, 1551.]—Giard & Billet, 1892a, 613.—Gmelin, 1790a, 3053–3054.—[Gege, 1782a, 169–172, pl. 14, fig. 1.]—Gomy, 1897a, 374.—Gurlt, 1849a, 120.— [Gœze, 1782a, 169–172, pl. 14, fig. 1.]—Gomy, 1897a, 374.—Gurlt, 1849a, 120.— Hahn & Lefèvre, 1884a, 516 (to Dist.).—Haldemann, 1851, 46, pl. 77, fig. 30.— [Hamont & Fischer, 1834a; (1834b).]—Harley, 1876, 255–256, figs. 208, 209.— Harrop. 1870a, 12–16 (heptica).—[Heide, 1686–1688, 46–47 (in sheep).]—Herbst, 1787a, 31–32, pl. 4, figs. 1–2 [=Dicrocœlium lanceatum].—Hodgson, 1838a, 528–538.—Hoyle, 1890, 535–540.—Huber, 1896, 574.—[Huzard & Desplas, 1797a, 10.]—[Huzard & Tessier, 1817a.]—Jaksch, 1889a, 186.— [Joseph, 1883e, 171–172; 1883f, 322–323.]—Khouri, 1904, 78 (as cause of halzoun, at Liban).—King, 1836a, 95–101.—Kingsley, 1885. 191–194, figs. 168–174.—Klencke, 1844, 420 pp.—Kowal., 1902, 26, 8.—Kriwonogow, 1886.—[Kulmus, 1721, 596.]—Lamarck, 1801a, 333.—[Lamouroux, 1824a, 560, 561, 562, 563.]—Lander, 1904a, 10.—Laveran & R. Bl., 1895, 8–33, 186–188.—[Leeuwenhoek, 1715, 1–33, 14 figs.]—Leuck., 1879, 33; 1886, 25; 1892b, 798.—[Looss, 1899, 557, 742; 1902m, 782; 1905, 110; 1907, Feb. 1.—Luehe, 1901, 167; 1902, 224; 1905, 334.—Marshall, J. T., (1883), 10 (= a parasite of Limnæa truncatula).—Marshall, W., 1887.—Martens, 1824.—Mojkowski, (1888a); 1888b, Mar., 118.—Moniez, 1896, 86, 90–114, 115, 118, 119, 121, 150, 158, figs. 19, 20, 22, 23.—Mueh., 1898, 10.—Mueller, O. F., 1774, 52–53; Hahn & Lefèvre, 1884a, 516 (to Dist.).—Haldemann, 1851, 46, pl. 77, fig. 30.— (1888a): 1888b, Mar., 118.—Moniez, 1896, 86, 90–114, 115, 118, 119, 121, 150, 158, figs. 19, 20, 22, 23.—Mueh., 1898, 10.—Mueller, O. F., 1774, 52–53; 1776; 1782.—Mueller, P. L., 1775, 42–43.—Neumann, 1888, 507–564, figs. 229–238; 1892. 504–528, figs. 271–281; 1892, 517–543, figs. 271–281.—[Nichols, 1755, 246–248.]—Nord., 1840, 618.—Odhn., 1905, 338–339, 340, 344.—Olss., 1876, 13.—Otto, 1816, vii.—Pallas, 1760; 1768, 269–271; 1781, 79.—Parkes, 1891, 79.—Par., 1887, 489.—Perroncito, 1882, 273–276, figs. 115–116.—[Pecquet, 1668.]—Poir., 1883, 74.—Pratt, 1898, 357.—Rail., 1899, 345.—Rail. & Marotel, 1898, 34, 37.—[Redi, 1684, 133; 1729.]—Roger, 1901, 94, 95.—[Romberg, 1706, 69–70.]—Rozier, 1774.—Rud., 1793a, 28–29; 1802, 62–64 (cf. lanceolata); 1804, 165; 1805, 37–38, a hominis beguing family; 1809a, 350, 353. lata); 1804, 165; 1805, 37-38, a. hominis, b. equi, c. tauri; 1809a, 350, 353; Bata); 1804, 105; 1805, 37-38, a. hommis, b. equi, c. tauri; 1809a, 350, 353; 1810a, 57 (pars syn. of Bothriocephalus solidus).—Ruysch, 1737, 23.—[Schæffer, 1726, 57.]—Schrank, 1788, 17; 1803, 210.—Schwarze, 1885, 41.—Shaw, 1901, 1027.—Sieb., 1845, 223.—Simon, 1897, 222.—Simonds, (1862), 21.—Smith, Wm. A., 1863, 35-36, fig. 7.—Sons., 1878, 615 (epatica); 1884, 59, 61; 1889, 275; 1891, 254; 1896, 112-116; 1896, 4 pp., varieties of; 1896, 295, 302, 303, 315; 1897, 250, 251, 252.—Spengel, 1905, 270.—Steel, 1881, 204 (in cattle).—Stewart, 1898, 328-331, figs. 0-17. Stide, 1827, 52, 50, 12. Stillag, 1897, 1898 315; 1897, 250, 251, 252.—Spengel, 1905, 270.—Steel, 1881, 204 (in cattle).—Stewart, 1898, 328–331, figs. 9–17.—Stieda, 1867, 52–59, pl. 2.—Stiles. 1896, 205; 1898a, 22, 29, 30, 31, 32, 33, 34, 35, 37, 38, 41, 42, 43, 45, 51, 52, 53, 55, 56, 57, 137. 138, 139, 140, 141, 142, 143, figs. 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17; 1901, 177; 1901, 180; 1901d; 1901e; 1902, 204; 1902, 220, 221, 222; 1903, 884.—Stiles & Hass., 1898a, 88, 89, 97.—Stoss., 1892, 7 (to Cladocelium).—Sweiten [1758, 89].—[Thomson, 1855, 188.]—Turton, 1807.—Tyson, 1903, 3 ed., 1180 (to Dist.).—Verrill, 1870, 170, 176, 177, 178, 213, 214.—Veterinarian, 1879, v. 52, 738–741.—Walter, (1866), 63–64.—Ward, 1895, 246–252, pl. 1, figs. 1–13, 253 (in part of Dinwiddie, 1889, to Fasc. magna), 256 (syns. Planaria latiuscula Goze, Dist. hep. Abildg., Fasc. humana Gmelin. (syns. Planaria latiuscula Gœze, Dist. hep. Abildg., Fasc. humana Gmelin,

F. lanceolata Rud., 1803, Fasciolaria hep., Distomata hominis Taylor, Dist. caviæ Sons.); 1903, 704, 706; 1903, 863, 864, 865 (syns. Dist. hep. Retzius, Fasc. humana Gmelin, Dist. caviæ Sons., Cladocœlium hep.), 866 (syn. probably Hexathyridium venarum), 867, 870, 871.—[Wepfer, 1716.]—Wernicke, 1886, 304.—[Willius, 1674–1675.]—Ztschr. f. mikr. Fleischschau, Berl., 1880.—Zuern, 1882, 207, pl. 4, figs. 5–8.

-, hepatica, anatomy of: Jackson, 1888, 643-645, 649, 651, 652, 653.—Ramdohr, 1814, 128–131 (in sheep).—See also foregoing entries.

-, bibliography of: Hass., 1894b, 407–417, 457–462.

180–189; 1883, May, 338–343; 1883, June, 404–409, figs. 1–6; 1883, July, 469–470 (and prevention); 1883, Aug., 550–557; 1883, Sept., 621–633; 1883, Oct., 681–690, figs. 1–20; 1883, v. 17, Oct., 270–286, figs. 1–18; 1883, v. 19 (37), 276–305, figs. 1–20; 1883, Nov., 344–352, fig. 19.—Veterinarian, Lond., 1880, May, 332-333; 1881, June, 390-392.—See also life cycle of.

-, embryo of: Baillet, 1879a, 197–215, figs. 1–11; 1880a, 19 pp.

-, geographic distribution of: Rail., 1895, 338-340 (var. in Senegal).— Rolleston, 1880b, 400–405.

—, histology of: Harley, 1876a, 255–256, figs. 208–209.

_____, intermediate host of: Cobb, 1898c, 36–38, 2 figs.—Fielder, 1896a, 24–28, 139–140, 146–147, figs. 1–7.

in various animals: [Frommann, 1688c, 252–259; 1755a, 291–294].— — in various animals: [Frommann, 1688c, 252–259; 1755a, 291–294].— In cattle: Cocu, 1897a, 369–371 (heart); 1898b, 98 (heart).—Galli-Valerio, 1901c, 364.—[Hutcheon, 1893h, 347 (death).]—Mills, 1876.—Morot, 1890, 191–193 (erratic).—Rail., 1897, July 30, 369–371 (heart).—Rail. & Morot, 1885, 285 (lungs).—Ward, 1895, 332 (biliary ducts).—Horse: Adams, 1876, 764–765 (India).—Galli-Valerio, 1901c, 364.—Ward, 1895, 338 (biliary ducts).—Man: Ashford, King & Igaravidez, 1904, Dec. 1, 92 (Porto Rico).—Looss, 1905, 88.—Ward, 1895, 238, 246, 328 (biliary ducts).—Wilson, 1880, Nov., 413–417.—Sheep: Camper, 1762a, 304, pl. 4.—Dupuy, 1822a, 29–36.—Galli-Valerio, 1901c, 364.—Halsted, 1883a, 741–748, figs. 1–11.—[Hutcheon, 1889e, 235 (Capetown); 1892m, 141; 1893o, 403–404; 1893r, 425–426.]—Morton, 1839, Nov., 735–738, fig. 1.—Rhodesian Agric. J., Cape Town, 1906, v. 3 (5), June, 529.—Ward, 1895, 238–246.—Miscellaneous: [Cadéac, 1885a (Equus asinus, 529.—Ward, 1895, 238–246.—Miscellaneous: [Cadéac, 1885a (Equus asinus, female); 1885b (Equus asinus, female).]—Cobbold, 1858b, 167 (Lepus cuniculus).—Ward, 1895, 238 (man, cattle, sheep, goat, horse, ass, hog, rabbit, cat), 246 (stag, squirrel, elephant), 335 (Ovis aries: biliary ducts).

—, life cycle of: Fagge & Pye Smith, 1902, 4 ed., 475 (syn. Dist. hep.).— Jackson, 1883a, 248–250.—Thomas, 1883, Jan., 99–133, pls. 2–3, figs. 1–26; 1883, v. 23, 8.—See also development of.

hepatica ægyptica (Looss, 1896), Stiles, 1898a, 23, 48, 49, 50, 139, 140, figs. 25–26.— Braun, 1903, 3 ed., 152 (in buffalo, cattle, sheep, goats); 1906, 157 (=F. angusta=F. ægyptiaca).—Looss, 1902m, 782.—Sons., 1896, 115; 1896, 295.

hepatica angusta Rail., 1895, 338-340 (in cattle; St. Louis, Senegal, Africa); 1895, 702.—Ř. Bl., 1895, 733 (=F. gigantea).—Braun, 1903, 3 ed., 151 (cattle, liver); 1906, 156, 158, fig. 89.—Gomy, 1897a, 376.—Looss, 1902m, 782.—Rail. & Gomy, 1899, 349.—Stiles, 1898a, 23, 48, 49, 137, 140, figs. 23, 24.—Sons., 1896, 2; 1896, 113, 115.—Veterinarius, 1898, 328.

hepatica apri Gmelin, 1790a, 3054 (in Sus scrofa; from Le Clerc, 1715a, 119).

hepatica boum Gmelin, 1790a, 3054 (in Bos taurus; Europe) (=Planaria latiuscula Gœze, 1782a. pro parte, renamed—Fasc. hepatica).

hepatica cavix Sons., 1896, 112-116; 1896, 4 (in Cavia).—Stiles, 1898a, 48, 139.

hepatica cervi Gmelin, 1790a, 3054 (in Cervus).

hepatica equi Gmelin, 1790a, 3054 (in Equus caballus).

hepatica ovata plana Buchholzii of Jærdens, 1802a, 64-66, pl. 7, figs. 13-14 (see Dicrocœlium lanceatum).

- hepatica porcorum Gmeiin. 1790a, 3054 (in Sus scrofa domestica; Europe) (=Plan. latiuscula Gœze, 1782a, pro parte, renamed=F. hepatica).
- heterophyes (Sieb., 1852) Moquin-Tandon, 1860.—R. Bl., 1888a, 625–627 (to Dist.).—Ward, 1895, 328 (in Homo).
- hirundinis Freelich. 1791, 75–76 (in Thurmschwalbe).—Baird, 1853 (syn. of D. maculosum).—Braun, 1901, 566; 1902b, 45, 46 (syn. of Plagiorchis mac.).—Dies., 1850a, 349 (syn. of D. mac.).—Linst., 1901, 197 (syn. of D. mac.).—Rud., 1809a, 374.
- humana Gmelin, 1790a, 3053 (in Homo).—R. Bl., 1888a, 543 (syn, of D. hepaticum).—Braun, 1903, 3 ed., 147 (syn, of F. hepat.); 1906, 150, fig. 83 (syn, of F. hep.).—Cobbold, 1866, 6.—Dunglison, 1893, 424.—Rud., 1899a, 353.—Stiles, 1898a, 29.—Ward, 1895, 246 (syn, of F. hepat.), 328 (in Homo), 332 (in Bos taurus), 335 (in Ovis aries), 338 (in Equus caballus); 1903, 865 (syn, of F. hepat.).

inflera Rud., 1802a, 82–83 (in Cyprinus jeses): 1809a, 395.—Dies., 1850a, 365 (to Dist.) (in Leuciscus jeses: Apr., Greifswald).

intestinalis Linn., 1758a, 649 (in intestinis piscium); 1761, 505.—Abildg., 1790, 34.—Baird, 1853 (syn. of Ligula simplicissima).—Barbut, 1783, 15, pl. 2, fig. 2.—Blumenbach, 1825, 242.—Bosc, 1802a, v. 1, 277 (syn. of Lig. abdominalis).—Braun, 1893a, 883; 1900a, 1642.—Bremser, 1819.—Dav., 1877, 260.—Dies., 1850a, 580 (syn. of Lig. diagramma).—Gœze, 1782a, 41, 183 (=a "Gattung" of Fasc, Gœze).—Herbst, 1787a, 32, 41 (syn. of Lig. piscium).—Luehe, 1905, 334.—Montin, 1763a, 113-118; 1766a, 122-127 (in Homo) [evidently a tapeworm, det. as F. i. Linn.].—Nord., 1840, 590 (syn. of Lig. cingulum).—Pallas, 1781, 95.—Rauh, 1779, 11.—Rud., 1810a, 20 (syn. of Lig. cingulum), 25 (syn. of Lig. mergorum).—Steenstrup, 1857, 186-195 (in Stichling): 1858, 298-312; 1859, Dec., 475-485.—Wegelin, 1779, 8.

intestinalis Gueldenstædt. (21791a). 115 (in Sorex moschatus).—Dies., 1850a, 617 syn. of Cephalocotyleum mygales moschatæ Rud.).—Rud., 1810a, 24

=Ligula soricis moschati).

jacksonii Cobbold, 1869, 48–49, in Indian elephant: 1870, 47–49; 1873, 13; 1875, 733–743; 1879b, 393, 394, 395, 396, 397; 1882, 223–258, pls. 23, 24.—Braun, 1892a, 567, 650, 674, 710 (jacksoni); 1893a, 875, 910.—Fitz, 1876a, 513–518; 1876b, 513–518; 1876c, 854 (jacksoni).—Jackson, 1847.—.—Linst., 1878,—.,—Looss, 1899b (to Fasciolopsis).—Sons., 1889, 275 (jacksoni); 1896, 114, 115 (jacksoni).—Stiles, 1895, 144–147, pl. 6, 213–215; 1898a, 29.—Veterinarian, Lond., 1876, 854 (jacksoni).

jesis Gmelin, 1790a, 3058 (in Cyprinus jeses intest.; Europe).—Bosc, 1802a, v. 1, 275.—Dies., 1850a, 365 (syn. of D. inflexum Rud.).—Rud., 1793, 30; 1809a,

372, 395.

lactea Mueller, 1774, 61-62 (in rivo sub foliis nymphææ ac in Palustris frequens).— Schrank, 1803, 167 (to Plan.).—Johnston, 1865, 10.

lagena Braun, 1788 Gmelin, 1790a, 3057 (in Perca fluviatilis).—Baird, 1853a,
 57 (syn. of Dist. nodulosum Zed.).—Bosc, 1802a, v. 1, 273.—Rud., 1809a,
 411 (syn. of D. nod.).

lanceolata Rud., 1803a, 24–25 (nec Schrank, 1790) (in Homo; Europe); 1809a, 353 (syn. of Dist. hepaticum).—R. Bl., 1888a, 543 (syn. of D. hep.), 602 (syn. of D. lanceolatum).—Braun, 1903, 3 ed., 157, 166 (to Dicrocolium).—Dies., 1850a, 333.—Dunglison, 1893, 424 (lanceolatum).—Stiles, 1898a, 55.—Stiles & Hass., 1898a, 87 (=lanceatum, type of Dicrocolium).—Ward, 1895, 246, 328 (in Homo), 332 (in Bos taurus), 335 (in Ovis aries).

lanceolata Schrank, 1790, 123 (in Cyprinus brama): 1803, 213-214.—Dies., 1850a, 341 (syn. of D. globiporum).

off syn. of b. globiporum.

lanceolata (brama) [Schrank, 1790, 123] of Rud., 1809a, 364 (syn. of Dist. globiporum), see lanceolata 1790.

laureata (Zed., 1800) Rud., 1802a, 86-87; 1809a, 413 (to Dist.).—Blainv., 1824a,
 518 (type of lobostome).—Dies., 1850a, 380 (to Dist.).—Lamarck, 1816, 184.—
 Nord., 1840, 621 (includes F. farionis).

tinearis Rud., 1793a, 29 (in Phasianus gallus; Greifswald, Oct.); 1802a, 85–86; 1809a, 414.—Dies., 1850a, 380 (to Dist.).—Hass., 1896a, 2, 3 (to Crossodera).

linearis Mueller, 1774, 67-68 (in foveis palustribus sylvarum).—Schrank, 1803, 172-173 (to Plan.).

linearis longa Linn., -—.—Pallas, 1781a, 95 (in Cyprinus variis).

lineata Mueller, 1774, 60-61 (in littore maris Balthici).

loliginis Gmelin, 1790a, 3059 [or Fabricius, ——, 328] (includes F. barbata Linn., probably this renamed).—Baird, 1853a, 115 (=Tetrabothriorhynchus barbatus).—Dies., 1850a, 573 (syn. of Tetraboth. migratorius).—Rud., 1810a, 385.

longicollis Abildg. [1788], 34, pl. 151, fig. A, 1-2 (in Coluber natrix).—Dies., 1850a, 348 (syn. of D. naja).—Olfers, 1816, 46.—Rud., 1819a, 377 (syn. of D. naja).

longicollis Rud., 1809a, 380 (syn. of Dist. tereticolle), based on Bloch, 1782a, 6 (syn. Fasc. lucii in Esox lucius).—Ben., 1858a, 1861a, 100 (to D. ter.).—Dies., 1850a, 358 (syn. of D. ter.).—Nord., 1840, 619 (syn. D. ter., F. lucii).— Lamarck, 1816, 183.—Sramek, 1901, 105.

longicollis Fredich, 1791, 73-75, pl. 3, figs. 9-11 (in Cyprinus carpio).—Dies., 1850a, 341 (syn. of Dist. globiporum).—Harz, 1881c, 11 (to Dist.).—Looss, 1894a, 41 (syn. of D. gl.), 49 (e. p. syn. of D. isoporum); 1902m, 763 (and D. gl.).—Rud., 1809a, 365 (syn. of D. gl.); 1814a, 103.—Stiles, 1901, 168.

longicollis (carpionis), see longicollis (Freelich).

lucii Mueller, 1776, 224 (in Lucius, esoph. and stomach); 1779a, 67-68, pl. 30, i Mueller, 1776, 224 (in Lucius, esoph. and stomach); 1779a, 67–68, pl. 30, fig. 7, in Esox lucius; 1780, 136; 1784, 115, pl. 78, figs. 6–8; —, vol. 1, 33–34, pl. 30, fig. 7 (ventric. of Esox lucius); —, vol. 2, 52–33, pl. 78, figs. 6–8; 1780, 203.—Baird, 1853a, 53 (=D. tereticolle).—Ben., 1858a, 1861a, 100 (syn. of D. ter.).—Bloch, 1779a, 537, pl. 14, figs. 1–4; 1782a, 6–7.—Bosc, 1802a, v. 1, 274.—Bruguière, 1791a, pl. 79, figs. 20–23.—Crep., 1837, 316.—Dies., 1850a, 358 (syn. of D. ter.).—Gmelin, 1790a, 3058.—Harz, 1881c, 5 (to Dist.).—Herbst, 1787a, 32, pl. 4, figs. 1–4.—Jurine, 1823a, 145–153, figs. 1–5; 1824a, 489–497, figs. 1–5; 1830a, 784–785.—Knoch, 1862, 19.—Looss, 1894a, 5 (syn. of D. ter.).—Nord., 1840, 619 (syn. of F. longicollis).—Rud., 1809a, 379.—Schrank, 1788, 18.—Sramek, 1901, 105.

luciopercæ Mueller, 1776, 223; 1779a, 64, pl. 30, fig. 2; 1788, vol. 1, 32, fig. 30, fig. 2 (int. Perca lucioperca).—Baird, 1853a, 56 (=D. nodulosum).—Bosc, 1802a, v. 1, 273.—Bruguière, 1791a, pl. 79, fig. 13.—Dies., 1850a, 380 (syn. of D. nod.).—Gmelin, 1790a, 3057.—Herbst, 1787a, 33.—Looss, 1894a, 33 (syn. of D. nod.).—Nord., 1840, 621 (syn. of F. nodulosa).—Rud., 1809a, 410.—Schrank, 1788, 18; 1790, 123.—Sramek, 1901, 106 (syn. of D. nod.).—Stiles & Hass., 1898a, 84, 85 (syn. of D. nod.).

lucipetum (Rud., 1819) Blainv., 1828a, 585.

macrostoma Rud., 1803a, 26-27 (in Nachtigall); 1809a, 386.—Blainv., 1828, 585.— Dies., 1850a, 361 (to Dist.).—Poche, 1907, 125.—Stiles, 1901, 192.—Stiles & Hass., 1898a, 95, 96 (type of Urogonimus).

maculosa Rud., 1802a, 67–68 (F. hirundinis Frœlich renamed) (in Hirundo apus, H. rustica, H. urbica); 1809a, 374.—Braun, 1902b, 46 (to Plagiorchis).—Dies., 1850a, 349 (to Dist.).

magna (Bassi, 1875) Stiles, 1894c, 172–178, 225–243, pls. 1–11, figs. 4–7, a–g; 1895l, ma (Bassi, 1875) Stitles, 1894c, 172–178, 225–243, pls. 1–11, figs. 4–7, a–g; 18951, 277–280; 1894w, 91–94; 1896i, 202; 1898a, 22, 27, 29, 42, 49, 51, 52, 53, 54, 55, 140, 141, 142, figs. 28–35; 1902, 222.—Bossuat, 1902, v. 6 (2), 192.—Looss, 1899b, 557; 1902m, 783.—Pratt, 1898, 357, 365.—Rail., 1893a, 356.—Salmon & Stiles, 1901, 454.—Smith, F. C., 1881, 14 (Dist. crassum).—Sons., 1889, 275 (syn. Dist. magnum); 1896, 114, 115; 1896, 303.—Ward, 1895, 238 (in cattle), 252–253, 254–255, pl. 2, figs. 14–16 (in cattle and deer) (includes: D. hepaticum p. p., F. hepatica p. p., F. carnosa, F. americana, D. texanicum), 332 (in Bos taurus), 335 (in Ovis aries), 338 (in Equus caballus); 1903, 866 (includes: F. americana, F. carnosa, Cladocœlium giganteum p. p., D. crassum p. p., D. texanicum), 867.—Also reported for Boselaphus tragocamelus. Cervus p. p., D. texanicum), 867.—Also reported for Boselaphus tragocamelus, Cervus canadensis, C. dama, C. elaphus, C. virginianus, C. unicolor.

maimonis Blainv., 1828a, 586, 2 figs. (in Simia maimon).—Dies., 1850a, 374 (syn. of

D. laciniatum).

marmorosa Mueller, 1774, 71 (in fossis aquaticis rara).—Schrank, 1803, 175 (to Plan.).

melis Schrank, 1788, 17 (in Dachs, Meles), based on Geeze, 1782a, 176, pl. 14, figs. 9-10; 1803, 214.—Bosc, 1802a, v. 1, 268.—Dies., 1850a, 381 (syn. of D. trigonocephalum).—Gmelin, 1790a, 3053.--Rud., 1793a, 26-27; 1809a, 415.

mesostoma Rud., 1863a, 28 (in Krametsvogel); 1809a, 388 to Dist.—Braun, 1902b, 126.—Dies., 1850a, 361.

militaris Rud., 1803a, 30-31 (in Scolopax arquata); 1809a, 421 (to Dist.).—Ben., 1858a, 1861a, 86 (to Dist.).—Dies., 1850a, 384 (to Dist.).

milvi Gmelin, 1790a, 3054 (in Milvus).—Bosc., 1802a, v. 1, 269.—Dies., 1850a, 385 (syn. of D. echinocephalum).—Rud., 1809a, 429 (syn. of D. falconis milvi).

muris hepatica Reederer, 1762, 537-539 (Mus musculus; Germany).—Braun, 1894.—Luche, 1905, 146 (syn. of Cysticercus fasciolaris).—Stiles, 1906, 43.—Stiles & Stevenson, 1905a, 10.—Tschudi, 1837, 67.

mutabilis Schrank, 1803, 210-211.

nana Rud., 1802b, 68-69 (in Scolopax gallinula; Greifswald, July); 1809a, 377, to Dist.—Dies., 1850a, 350 (to Dist.).—Braun, 1902b, 47 (to Plagiorchis).

nigra Mueller, 1774, 54 (in rivo; free form).—Herbst, 1787a, 36.—Johnston, 1865,

9.—Schrank, 1803, 165 (to Plan.).

nodulosa Frælich, 1791, 76-80 (includes F. percæ cernuæ Mueller (in Perca cernua, P. fluviatilis).—Dies., 1850a, 380 (to Dist.).—Looss, 1894a, 33 (to Dist.).—Nord., 1840, 621 (includes F. luciopercæ, F. percæ cernuæ).—Lamarck, 1816, 184.—Rud., 1809a, 411.—Schrank, 1803, 213.—Type of Crossodera 1845 and Bunodera 1896.

obscura Mueller, 1774, 65 (in piscinis).—Schrank, 1803, 171–172 (to Plan.).

ocreata Rud., 1802b, 79-81 (F. halecis Gmelin, renamed) (in Clupea harengus); 1809a, 329, 398 (to Dist.).—Dies., 1850a, 372 (to Dist.).—Mont., 1891, 15 (to Apoblema); 1891, 496, 499.—Type of Pronopyge 1899.

ocreata Geze, 1782a, 182, pl. 15, figs. 6-7 (in Maulwurf).—Dies., 1850a, 326 (to Monost.) (in Talpa europæa).—Nord., 1840, 624 (to Monost.).—Rud., 1809a,

329 (to Monost.).

ocularis Moquin-Tandon, 1860, 347 (includes Dist. ophthalmobium, Dicrocœlium oculi humani); 1861; 1862.—R. Bl., 1888a, 630.—Stiles, 1902, 29.—Ward, 1895, 328 (in Homo).

oculis Moquin-Tandon (Hulme's), 1861, 375, for F. ocularis.—Stiles, 1902, 28, 29.

ovata Rud., 1803a, 25–26 (in Corvus frugilegus); 1809a, 357 (to Dist.).—Braun, 1901, 13; 1902b, 69, 77 (p. p. syn. of Prosthogonimus cuneatus; pp. syn. of P. cuneatus).—Dies., 1850a, 335 (to Dist.).—Hass., 1896a, 2, 3 (to Cephalogonimus).—Looss, 1899b, 629 (type of Prymnoprion).

percæ Gmelin, 1790a, 3057 (= F. percæ cernuæ renamed) (in Perca cernua).—Bosc, 1802a, v. 1, 273.—Baird, 1853a, 56 (syn. of Dist. nodulosum).—Bruguière, 1791a, pl. 79, fig. 14.—Rud., 1809a, 411 (syn. of D. nod.).

perca cernua Mueller, 1776, 224; 1779a, 65, pl. 30, fig. 1 [sic. = fig. 3]; 1788, v. 1, 32, pl. 30, fig. 3 (in Perca cernua).—Dies., 1850a, 380 (syn. of D. nodulosum).—Herbst, 1787a, 33.—Looss, 1894a, 33 (syn. of D. nod.).—Nord., 1840, 621 (syn. of F. nod.).—Rud., 1809a, 410.—Schrank, 1788, 18.

percina Schrank, 1790, 123 (in Perca asper, P. vulgaris); 1803, 213.—Dies., 1850a, 380 (syn. of D. nodulosum).—Looss, 1894a, 33 (syn. of D. nod.).—Rud., 1809a,

411.

picta Rud., 1802, 64-65 (F. vespertilionis renamed) (in Fledermaus); 1809a, 427 (renamed lima).—Dies., 1850a, 387 (syn. of D. lima).—Kolenati, 1857, 12.

platellæ Bosc, 1802a, v. 1, 273 (see platessæ).

platessæ Mueller. 1784, 114, pl. 78, figs. 1-5; 1788, v. 2, 52, pl. 78, figs. 1-5.-Bruguière, 1791a, 79, figs. 26–27.—Cobbold, 1858b, 160.—Dies., 1850a, 352 (syn. of D. areolatum).—Gmelin, 1790a, 3057.—Rud., 1809a, 401 (renamed D. areol.).

polymorpha Rud., 1802b, 70-72 (F. anguillæ Gmelin, renamed) (in Aal); 1809a, 363 to Dist.—Dies., 1850a, 340 (to Dist.).

punctata Mucller, 1774, 57 (in pratis inundatis primovere, free form).—Pallas, 1774, 23, pl. 1, fig. 14 a b (in aq., Belgii).—Schrank, 1803, 166 (to Plan.).

punctum (Zed., 1800) Blainv., 1828a, 586.

pusilla (Braun, 1790) Gmelin, 1790a, 3055.—Bosc, 1802a, v. 1, 269.—Dies., 1850a, 360, to Dist.—Rud., 1802b, 75–76; 1809a, 385 (to Dist.).

putorii Gmelin, 1790a, 3053 (in Mustela putorius).—Bosc, 1802a, v. 1, 268.—Dies., 1850a, 381 (syn. of Dist. trigonocephalum).—Rud., 1793a, 25–26; 1809a, 415.—Schrank, 1788, 17.—Stoss., 1892, 29 (syn. of Echinost. trigonocephalum). quadrangularis Pallas, 1774, 20–21, pl. 1, figs. 12a-c (in fossis; Hagæ comitum).

radiata Mueller, 1774, 66 (in aquis sylvestribus, rara).

ranæ Gmelin, 1790a, 3055 (for Plan. subclavata Gœze, 1782a, 178, which includes Fasc. subclavata, ore sessili Pallas pro parte; t. h. [based on Gœze] frog, Rana, intestine; Europe).—Bosc, 1802a, v. 1, 270.—Rud., 1809a, 348 (syn. of Amphist. subclavatum).

ranæ Frœlich, 1791, 69–73, pl. 3, figs. 7–8 (in Rana temporaria L.).—Baird, 1853a, 52 (syn. of Dist. clavigerum).—Braun, 1892a, 768.—Dies., 1850a, 352 (syn. of D. clavigerum).—Looss, 1894a, 84, 85, 88 (syn. of D. endolobum); 1905, 21, 22 (syn. D. endol.).—Olfers, 1816, 45.—Rud., 1809a, 348 (syn. of Amphist. subclavatum).

reticulata (Wright, 1879) Looss, 1899b, 557.

reroluta Freelich, 1802, 58-60, pl. 2, figs. 6-7 (in Anas boschas; Europe).—Looss, 1899b, 680 (syn. of Echinost. echinatum).—Rud., 1814a, 102 (sub Dist. echinatum).

rosea Mueller, 1774, 58 (in sinu Dröbachiensi).

rostrata Mueller, 1774, 65–66 (in paludosis, primo vere annorum).—Johnston, 1865, 15.—Schrank, 1803, 170–171 (to Plan.).

rubra Mueller, 1774, 59 (Fucorum frequens).—To Plan. in 1776.

saccata Gœze, 1782a, 221, based on Sack-Egel of Merrem, 1781, 169, pl. 1, figs. 3-7.—
Braun, 1894a, 4.—Raum, 1883, 20.—Stiles, 1906, 43.—Stiles & Stevenson, 1905a, 10 (syn. of Cysticercus fasciolaris).—Tschudi, 1837, 67 (syn. of Cyst. fasc.).—Zed., 1803a, 406 (syn. of Cyst. tæniæformis).

salamandræ Frœlich, 1789, 119–121, pl. 4, figs. 8–10 (in black salamander, Salamandra atra).—Bosc, 1802a, v. 1, 270.—Dies., 1850a, 356 (syn. of Dist. crassicolle).—Gmelin, 1790a, 3055.—Rud., 1809a, 378.—Stiles, 1901, 197.

salmonis Mueller of Gœze, 1782a, 173.—Luehe, 1901, 399 (? syn. of Hemiurus crenatus (Rud.) Luehe), 401.—Odhn., 1905, 353.

scabra Mueller, 1784a, 31–32, pl. 51, figs. 1–8, in Gadus barbatus; —, v. 2, 14, pl. 51, figs. 1–8 (in ventric. Gadus barbatus).—Bosc, 1802a, v. 1, 272.—Bruguière, 1791a, pl. 79, figs. 28–32, v–z.—Dies., 1850a, 393.—Gmelin, 1790a, 3056.—Knoch, 1894a, 3.—Mont., 1891, 507.—Odhn., 1905, 349 (=F. serrulata Mueller), 353.—Rud., 1809a, 406 (to Dist.).

scombri pelamidis Tilesius, in Rud., 1809a, 391 (syn. of Dist. clavatum), 437.—Buttel-Reepen, 1902, 166.—Cobbold, 1879b, 461.—Dies., 1850a, 366 (syn. of D.

clav.).

scorpii Mueller, 1776, 223; 1779a, 64, pl. 30, fig. 1; 1788, v. 1, 32, pl.30, fig. 1 (in Cottus scorpius, Blennius viviparus, int.).—Bosc, 1802a, v. 1, 273.—Bruguière, 1791a, 79, fig. 12.—Dies., 1850a, 366 (syn. of D. granulum).—Gmelin, 1790a, 3057.—Herbst, 1787a, 33.—Lamarck, 1816, 183–184.—Nord., 1840, 620 (syn. D. gran.).—Rud., 1809a, 394 (syn. of D. gran.).—Schrank, 1788, 18.

serrulata Mueller, 1776, see Goze, 1782a, 173.—Luehe, 1901, 399 (? syn. of Hemiurus crenatus (Rud.) Luehe), 400, 401.—Odhn., 1905, 349 (= F. scabra

Mueller), 353.

solæiformis Blainv., 1828a, 585 (for soleæformis Rud., 1809).

squali grisei Risso, —, 38, see Dies., 1850a, 347 (syn. of D. veliporum).

stagnalis Mueller, 1774, 53 (in stagnis; free form).—To Plan. in 1776.

strigata Mueller, 1774, 66-67 (in aquis paludosis).—To Plan. in 1776.

strigis (Schrank, 1788) Gmelin, 1790a, 3055.—Baird, 1853a, 47 (= Holost. macrocephalum).—Bosc, 1802a, v. 1, 269.—Dies., 1850a, 312 (syn. of Holost. variabile).—Linst., 1905, 191.—Nord., 1840, 626 (syn. of Amphist. macrocephalum).—Rud., 1809a, 340 (syn. of Amphist. macro.).—Type of Strigea, Amphist., Holost.

strigis stridulæ Braun, in Rud., 1809a, 347.

subclavata (Gœze, 1782) Schrank, 1788a, 19: 1803, 211-212.—Dies., 1850a, 318 (of Schrank to Diplodiscus), 368 (of Pallas syn. of Dist. cylindraceum).—Looss, 1894a, 64 (Pallas e. p. syn. of Dist. cygnoides); 1905, 21, 22.—Rud., 1809a, 348 (of Schrank syn. of Amphist. subclavatum), 393 (of Pallas syn. of Dist. cylindraceum).

subclavata ore sessile Pallas, 1760 or 1761, 271 (polynomial), in lungs of Rana and liver of Hungarian sheep.

tentaculata Mueller, 1774, 63-64 (in aquis palustribus).—To Plan.

tereticollis Rud., 1802b, 74–75 (F. lucii Mueller, renamed); 1809a, 380.—Ben., 1858a, 1861a, 100 (to Dist.).—[Blainv., 1828a, 585.]—Dies., 1850a, 358.—Looss, 1894a, 5; 1899b, 570 (type of Azygia).—Sramek, 1901, 105 (to Dist.).

terrestris Mueller. 1774, 68-69 (in asseribus muscisque humidis).—Claus. 1885a, 262 (to Rhynchodesmus, Geoplanidæ).—Metchnikov, 1866b, 433-447; 1866, 22 pp. (? syn. of Geodesmus bilineatus).

tetragona Mueller, 1774, 69 (in stagno ac foveis aquæ purioris nec vulgaris).—To Plan. in 1776.

tincæ Modeer, 1790, 127 (in Cyprinus tinca).—Dies., 1850a, 395 (syn. of D. perlatum).—Looss, 1894a, 24 (syn. of D. perl.).—Rud., 1809a, 365 (syn. of D. globiporum).—Stiles & Hass., 1898a, 92 (syn. of D. perlatum).

torva Mueller, 1774, 62-63 (in aquis).—Johnston, 1865a, 11.—Schrank, 1803, 167—169 (to Plan.).

trachea Montagu. 1811a, 194–198, pl. 7, fig. 4 (in poultry).—Hass., 1896a, 3, 6 (syn. of Syngamus trachealis); 1896b, 1, 2.—Rail., 1893a, 453 (syn. of Syng. trach.).

transversalis Rud., 1802a, 69 (in Cobitis fossilis); 1809a, 362; 1819a, 95 (Greifswald and Berlin).—Dies., 1850a, 339.—Odhn., 1901, 505.

tremellaris Mueller, 1774, 72 (in mari Balthico, Hafniam alluente).—Johnston, 1865a, 6.—To Plan, in 1776.

trigonocephala Rud., 1802b, 87-88 (F. melis Schrank, renamed) (in Schweinigel).— Dies., 1850a, 382.—Lamarck, 1816, 184.—Nord., 1840, 621-622 (includes D. echinatum, Echinost. echinatum, E. ferox, F. echinata, F. ferox, Plan. putorii).—Stoss., 1892, 29 (to Echinost.).

truitta Bosc, 1802a, v. 1. 274 for truttæ, Frælich).

truncata Mueller, 1806, 35, pl. 151, fig. B, 1-3 (in Perca lucioperca).—Rud., 1814a, 105.

truttæ (intestinalis)
 Rœderer, 1762, June 19, 537 (in forelle).—Bosc, 1802a, v. 1,
 274.—Dies., 1850a, 583 (syn. of Ligula nodosa).—Luehe, 1904, 146; 1905, 334
 (=Echinorhynchus).—Rud., 1809a, 262 (? syn. of Echinorhynchus fusiformis); 1810a, 18 (syn. of Lig. nod.).—Schrank, 1803, 209.

truttæ Frœlich, 1789, 126, 127, pl. 4, figs. 16–17 (in Salmo trutta; Europe).—Dies., 1850a, 380 (syn. of Dist. laureatum).—Gmelin, 1790a, 3058,—Rud., 1809a, 413

(syn. of D. laur.); 1814a, 102 (syn. of D. farionis Freelich).

umblæ Fabricius, 1780a, 329; 1794, 26–29, pl. 3, figs. 6–8 D (in Salmo umbla: Greenland).—Bosc, 1802a, v. 1, 274.—Dies., 1850a, 343 (syn. of Dist, seriale).—Gmelin, 1790a, 3058.—Herbst, 1787a, 36.—Rud., 1809a, 368, 369 (renamed Dist, seriale, in Salmo alpinus).

uncinata Dies., 1850a, 412 (syn. of Polyst. integerrimum) lapsus for uncinulata.—

Stoss., 1898, 10.

uncinulata (Braun, 1790) Gmelin. 1790a. 3056.—Bosc. 1802a. v. 1. 270.—Nord., 1840, 594 (syn. of Linguatula integerrima).—Rud., 1809a. 452 (syn. of Polyst. integerrimum).

upupæ Šchrank, 1790, 123 (in Upupa epops, rectum); 1803, 211.—Dies., 1850a,
 351 (syn. of Dist. involutum).—Rud., 1809a, 377 (syn. of D. involutum).

varica Mueller, 1784a, 93–94, pl. 72, figs. 8–11 (in Salmo salar); 1788, v. 2, 43, pl. 72, figs. 8–11.—Bosc, 1802a, v. 1, 273.—Bruguière, 1791a, pl. 80, figs. 5–8.—Dies., 1850a, 368 (to Dist.).—Gmelin, 1790a, 3057.—Lamarck, 1816, 184.—Levin., 1881a, 54 (to Dist.).—Nord., 1840, 620 (to Dist.).—Odhn., 1905, 360 (to Derogenes).—Rud., 1802b, 81; 1809a, 396 (to Dist.).

venarum (Treutler, 1793) Schrank, 1803. 210.

ventricosa Pallas, 1774, 17–18, pl. 1, figs. 9–10 (host not given; Amboyna).—
Baird, 1853a, 59 (to Hirudinella).—Buttel-Reepen, 1900a, 586; 1902, 166, 172, pl. 6, fig. 2.—Cobbold, 1879b, 459, 460.—Darr, 1902, 664, 665, 666.—Dies., 1859c, 431.—Herbst, 1787a, 36.

verrucosa Frœlich, 1789, 112–115, pl. 4, figs. 5–9 (in weidender Gänse) Ben., 1858a, 1861a, 78 (to Monost.).—Dies., 1839e, 234; 1850a, 411 (syn. of Notocotyle triseriale).—Hass., 1896a, 3 (to Notocotyle).—Mont., 1892, 26.—Nord., 1840, 602 (syn. of Nocotylus triserialis). 624 (to Monost.).—Odhn., 1905, 366 (type of Catatropis).—Rail., 1893a, 340 (to Notocotyle).—Rud., 1809a, 331.—Schrank, 1803, 209.

- vespertilionis Mueller, 1784a, 95-96, pl. 72, figs. 12-16; 1788b, v. 2, 43-44, pl. 72, figs. 12-16 (in Vespertilio auritus).—Bosc, 1802a, v. 1, 268-269.—Braun, 1900, 387.—Bruguière, 1791a, pl. 80, figs. 9–11.—Dies., 1850a, 387 (syn. of D. lima).—Gmelin, 1790a, 3053.—Kolenati, 1857, 12.—Lamouroux, 1824a, 563.—Rud., 1793a. 27-28; 1809a. 427.—Schrank, 1788, 18.
 - viridis Mueller, 1774, 59-60 (in radicibus fucorum; Greenland).
- vulpis Gmelin. 1790a, 3053 (in Canis vulpes; Europe).—Brand., 1888a, 9, 60 (syn. Hemist. alatum Dies.).—Dies., 1850a, 308 (syn. of Hemist. alatum).—Rud., 1793a, 30–32; 1809a, 402 (syn. of D. alatum).
- FASCIOLARIA Rafinesque, 1815, 151, fam. name, includes Anthostoma, Caryophylleus Zed., Fascinia, Fasciola L., Hexathyridia Treutler, Linguatula L., Lingula L., Polystoma Zed., Unicola Raf.
- FASCIOLARIA Encyclop. Metrop. or Universal Dict. of Knowledge. Lond., v. 18, 141 (as generic name) [not Fasciolaria Lamarck, 1799, mollusk].—Dies., 1850a, 577 (misprint for Fasciola).—Stiles & Hass., 1898a, 89 (syn. of Fasciola).
 - fimbriata (Gœze, 1782) Dies., 1850a, 577 (misprint for Fasciola) (syn. of Caryophyllæus mutabilis).—Sramek, 1901, 114 (syn. of C. mut.).
 - hepatica (Linn., 1758) Encycl. Metropolitana, 1845, v. 18, 141 (type of Fasc. .-Stiles, 1898a, 29.—Ward, 1895, 246.
 - ranæ (Gmelin, 1790) Nord., 1840, 627 syn. of Amphist. subclavatum (for Fasciola).

- FASCIOLIASIS, BIBLIOGRAPHY OF: Huber, 1890i, 77-79.

 ——, GEOGRAPHIC DISTRIBUTION: Biermer, 1863a, 381-395 (Switzerland); 1895a, 253 (Switzerland).—Chardin, 1876a, 277-314 (Orleansville).
 - -, in various animals: Burke, 1886b, 470; 1887a, 47.—Hass., 1894a, 162-167 (epizootics).
- -, in cattle: Bilhuber, 1791a and sheep.—Lucet, 1890b, 548-549 (lungs); 1890c, 549-550 (spleen).—Stiles, 1902b, 64-69, figs, 93-98, 126-134, figs, 99-109, 204–209, figs. 110–113, 312–319, figs. 114–118, 373–379, figs. 119–122, 558– 559, figs. 123-124.
- -, HEPATIC, IN MAN: Askanazy, 1900b, 491, 496 (Bostroem's 1883a case): 1901, HEPATIC, IN MAN: Askanazy, 1900b, 491, 496 (Bostroem's 1883a case); 1901, 75 (chronic choleportitis and fibrinous periportitis).—Biermer, 1863a, 381-395 (Switzerland); 1865a, 253 (Switzerland).—R. Bl., 1891p, 604-606, 616 [see correction in R. Bl., 1894g].—Bossuat, 1902 v. 6 (2), 187.—Bostroem, 1883a, 557-577, 1 fig.—Chabert, 1852a, 173-174 [a tapeworm]; 1852c, 174; 1852d, 195-196.—Chester, 1887a, 359-360 (must have been contracted in Dorsetshire, England).—Duffek, 1902a, 772-775; 1902b, 826; 1903a, 30: 1903, Aug. 15, 843.—Garcia Sola, 1884a, 129-135, fig. 5.—Huber, 1896, 574-575 (22 cases in 100 years).—Humble & Lush, 1881a, 75-76 (England).—Leichtenstern, 1885b, 502.—Prunac, 1878, 1147-1149; 1879, v. 27, 99-109; 1879, July 27, 429-434.—Sagarra, 1890, v. 14, 505-512; 1891, Apr. 18, 516-511.—Staff., 1905, Apr. 11, 694 (Canada).—Wyss, 1868, v. 9, 172-177, pl. 3, figs. 1-2.
- -, HEPATIC, SHEEP: Bass, 1893a, 93.—Cristini, (1845a).—[Dowling, 1894a, 232.]—[Dun, 1880a, 550-556; 1881a, 141-204.]—[Dupuy, 1838a, 511-523.]—Flemming, 1881a, 87-93; (1881b).—[Fonssagrives, 1868a, 299-315, figs. 1-2.]—[Friedberger, 1880a.]—[Frommann, 1688b, 245-252.]—General, (1883a), 86-87.—Halse, 1887a, 62 pp.; 1887b, 65; 1888a, 147-149.—Hutcheon, 1895h, 350-354, figs. 1-4.—King, 1836a, 95-101.—Morton, 1839, 735-738.—Waldinger, 1818.—Wernicke, 1888, 673 (Buenos Ayres).
 - -, pathology and location: Bossuat. 1902, 187.
- -, symptoms: Khouri, 1904, 78, 79, 80, 81 (halzoun).
- -, treatment: Bennett, 1876a, 677-680 (salt and potash: Australia).—[Hellier, 1895k, 635.]—[Hutcheon, 1891m, 86-87; 1891m, 140-141, sheep): 1893b, 23 (creolin)]; 1897b, 100 (sheep): [1897g, 142-146].—Syme, 1895, Dec. 26, 680-
- FASCIOLID. E Rail., 1895, 338.—R. Bl., 1895, 729-730.—Braun, 1900b, 387-391 (of Chiroptera): 1900c, 24-32 (Clinost., 1900d, 27-29 (Rhopalias); 1900f, 387-391 (of Chiroptera); 1900g, 250; 1901i, 55–58 (nomenclature); 1902a, 120 (nomenclature); 1902b, 1–162, figs. 1–99 (of birds); 1902c, 442–444; 1903, 3. ed., 147, 445; 1905, 59–60.—Darr, 1902a, 644–701, pls, 33–35, figs, 1–34; 1902b, 735–736; 1902, 58 pp.; 1905, July, 55–56.—Looss, 1901, 91; 1901, 191, 196; 1902m,

FASCIOLIDÆ—Continued.

838.—Luehe, 1901, 486-488.—MacCallum, 1899, 707.—Pratt, 1902, 887, 893 (key) (includes Anisocœlinæ, Bunoderinæ, Centrocestinæ, Cotylogoniminæ, Echinostominæ, Fasciolinæ, Haplometrinæ, Omphalometrinæ, Opisthorchiinæ, Philophthalminæ, Plagiorchiinæ, Psilostominæ, Reniferinæ, Anaporrhutinæ, Brachycœliinæ, Cephalogoniminæ. Dicrocœliinæ, Clinostominæ, Gorgoderinæ, Harmostominæ, Microphallinæ, Pleurogenetinæ, Hermiurinæ, Syncœliinæ, Telorchiinæ, Urogoniminæ, Zoogoninæ).—Roger, 1901, 94.—Shipley, 1905, v. 6, 4.—Stiles, 1898a, 22, 27, 28; 1901, 191, 192, 195; 1904i, 10–11, 12–45, figs. (of man).—Stiles & Hass., 1898a, 81–89 (type Fasciola; superspecific inventory).—Ward, 1903, 863, 864, 865.

FASCIOLINÆ Stiles & Hass., 1898a, 86, 90, 95, 96 (subf. of Fasciolidæ).—Looss, 1899b, 543, 556, 560, 561, 626; 1901, 196; 1902m, 839.—Luehe, 1900, 562; 1901, 486; 1901, 175.—Odhn., 1905, 339, 344, 346, 347.—Pratt. 1902, 887 (includes Campula, Fasc., Fasciolopsis; related genera: Paragonimus, Pleorchis).—Stiles, 1898a, 22, 28.—Ward, 1903, 867.

FASCIOLOPIS Odhn., 1902, 581, misprint for Fasciolopsis.

buski (R. Bl., 1888) Odhn., 1902, 581.

FASCIOLOPSIS Looss, 1899b, Dec., 557, 561 (tld. buskii), includes crassa Busk, jacksoni Cobbold.—Braun, 1903. 3. ed., 153; 1906, 158.—Odhn., 1905, 339. 344, 346.—Pratt, 1902, 887, 893.—Stiles, 1904i, 39-40.—Ward, 1903, 866.

1902: Fasciolopis Odhn., 1902, 581, misprint.

buski (R. Bl., 1888) Odhn., 1902, 573-581, figs. 1-3. See buskii.

buskii (Lankester, 1857) Stiles, 1901, 1542; 1904i, 41–42, figs. 63–66 (in Homo).— Braun, 1903, 3. ed., 153–154, fig. 99 (in Homo); 1906, 159, fig. 90 (buski) (syns. Dist. buski and D. crassum).—Conyngham, 1904 (IX, 17), 663.— Looss, 1905, 110. fig. 16; 1907, Feb. 1, 123 (buski) (from Hongkong).—Moore & Ferril, 1905a, Sept. 30, 1002–1003, figs. 1-7; 1905b, 8 pp., figs. 1-7 (in U.S.).— Ward, 1903, 864, 867; 1903, 704.

crassa (Cobbold, 1860) Looss, 1899b. 557.

jacksoni (Cobbold, 1869) Looss, 1899b, 557.

rathouisi (Poir., 1887) Ward, 1903, 864, 867; 1903, 704.—Looss, 1905, 111, fig. 17.— Stiles, 1904i, 42-43, fig. 67.

FELLODISTOMUM Staff., 1904, May 3, 486 (m. incisum).—Odhn., 1905, 310.

incisum (Rud., 1809) Staff., 1904, 486 (in Anarrhichas lupus).

FESTUCARIA Schrank, 1788, Gattung 9, 16–17 (type by elimination anatis, see Luehe, 1901; also type by first species rule); 1803, 185.—Blainv., 1824a, 512 Grant ("genre adopté de Schrank pour le Distoma trigonocephalum, qui paraît n'être qu'une espèce de monostome"), 518 ("ce sont les monostomes cylindriques et à bouche terminale de M. Rudolphi").—Brand., 1888a, 8.—Braun, 1893a, 884, 894.—Cuv., 1817, 40.—Dies., 1850a, 307 (syn. of Hemist.), 312 (syn. of Holost.), 319 (syn. of Monost.), 400 (of Zed., syn. of Amphist.), 411 (syn. of Notocotyle).—Hémont, 1827, 9.—Knoch, 1862, 30.—Lamouroux, 1822a, 194.—Looss, 1901, 193; 1902m, 746, 755.—Luehe, 1901p, 174, 175 (type anatis).—Rafinesque, 1815, 151 (renamed Monostomeus).—Rud., 1801a, 50, 54: 1809a, 21. 54; 1809a, 21.

alata (Gœze, 1782) Schrank, 1790, 118; 1803, 208–209.—Brand., 1888a, 60 (to Hemist.).—Dies., 1850a, 308 (to Hemist.).—Lamouroux, 1822a, 194.—Rud.,

1809a, 402, 403 (to Dist.).

anatis Schrank, 1788, 16. based on Gozze, 1782a, 174, pl. 13, figs. 8–11 (in Enten).— Dies., 1850a, 383 (syn. of D. echinatum).—Luehe, 1901p, 175 (type of Festucaria by elimination), species unidentifiable, possibly Echinost. echinatum.-Rud., 1809a, 418.

boschadis Schrank, 1790. 122–123 (in Anas boschas sylvestris, A. b. dom.): 1796,
 332, pl. 5, figs. 16–17; 1803. 207–208.—Dies., 1850a, 383 (syn. of D. echinatum).—Looss, 1899b, 680.—Rud., 1809a, 418, 419.

caryophyllacea Rud., 1810a, 353 (misprint for caryophyllina).

caryophyllina Rud., 1802a, 66-67, pl. 1, fig. 3 (in Gasterosteus aculeatus); 1809a, 325.—Dies., 1850a. 328 (to Monost.).—Rail., 1893, 339 (to Monost.).

cervi Zed., (1790), 65-74, pl. 3, figs. 8-11 (in Hirsch).—Dies., 1836, 246; 1850a, 401 (syn. of Amphist. conicum).—Fischder., 1901, 368 (to Paramphist.); 1902, 11 (to Paramphist. as type); 1903h, 504, 506 (of the Hirsch, Vormagen).—Looss, 1896b, 32 (to Amphist.).—Rud., 1899a, 350.—Stiles, 1898a, 64.—Ward, 1895, 256 (syn. of Amphist. conicum), 332 (in Bos taurus), 335 (in Ovis aries). FESTUCARIA—Continued.

cyprinacea Schrank, 1790, 122 (in Cyprinus barbus); 1803, 207.—Dies., 1850a, 329 (syn. of Monost. cochleariforme).—Nord., 1840, 624 (syn. of Monost. cochl.).—Rud., 1809a, 326 (renamed M. cochl.), 410.

lentis (Gescheidt, 1833) Moquin-Tandon, 1860, 349; 1861, 375; 1862, ——.— R. Bl., 1888a, 542, (to Monost.).—Stiles, 1902s, 25, 28.—Ward, 1895, 328.

oti Rud., 1819a, 354 (F. otidis Frælich, 1802, renamed).

otidis Freelich, 1802, 53–54 (changed to oti Rud., 1819, 354) (in Strix otus).—Rud., 1814a, 99–100.—Baird, 1853a, 47 (=Holost. macrocephalum Blainv.).

pedata Schrank, (1769), 335–340 (in Anas querquedula); 1803, 208 (includes F. anatis Schrank, 1788).—Baird, 1853a, 45 (=Monost. verrucosum).—Ben., 1858a, 1861a, 78 (syn. of Monost. verr.).—Dies., 1839, 234; 1850a, 411 (syn. of Notocotyle triseriale).—Odhn., 1905, 366 (syn. of Catatropis verrucosa).—Rud., 1809a, 331.

pileata Rud., 1802a, 65–66 (in Sterna hirundo); 1809a, 338 (to Monost.).—Dies.,

1850a, 314 (to Holost.).

strigis Schrank, 1788, 16–17 based on Goze, 1782a, 174, pl. 14, figs. 4–6 (in Weideneule).—Baird, 1853a, 47 (=Holost. macrocephalum).—Dies., 1850a, 312 (syn. of Holost. variabile Nitzsch).—Frælich, 1802, 51–52.—Linst., 1905, 191.—Luehe, 1901, 175.—Rud., 1809a, 340 (syn of Amphist. macrocephalum); 1814a, 99.—Type of Strigea, 1790.

ventricosa Rud., 1803, 20–21 (in Nachtigall); 1809a, 335 (to Monost.); 1819a, 86 (in Motacilla luscinia; Greifswald, May).—Dies., 1850a, 328 (to Monost.).

FESTUCARIOCREATA Rud., 1802, 67 (apparently lapsus for Festucaria ocreata).

FLUKES (vernacular name for Trematoda, esp. for Fasciola hepatica) Cobbold, 1883, 514–515.—Dugmore, 1895a, 491.—Feuerheerdt, 1896a, 272–273 (life history).—Hahn & Lefevre, 1884a, 515–549 (douves).—Johnson's New Univ. Cycl., N. Y., 1876, v. 2, 176.—Kuech., 1890, 186 (mode of alimentation).—Tyson, 1903, 3. ed., 1180–1181 (of liver).—Veterinarian, Lond., 1880, June, 397.

FRIDERICIANELLA Brand., 1894a, 305–311 (m. ovicola).—Maclaren, 1904, 583.—Mont., 1903, 336 (subf. Calceostominæ); 1905, 65.—St.-Remy, 1898, 524, 564.

ovicola Brand., 1894a, 303-310, pl. 19, figs. 1-4 (in Arius commersonii Lac.; Rio Grande do Sul, Brazil).—Bettend., 1897a, 16; 1897, 320.—Goto, 1899, 290.—Maclaren, 1904, 597.—St.-Remy, 1898, 565, fig. 5.

FURCOCERCA Lamarck, 1815, 446–448 (type?).—Braun, 1892a, 767; 1893a, 884.— Nitzsch, 1827, 66.—Pag., 1857, 5.—Bory de St. Vincent, 1823a, 355; 1825a, 83–84.

catellina (Mueller, 1786) Lamarck, 1815, 448.

catellus (Mueller, 1773) Lamarck, 1815, 448.

crumena (Mueller, 1786) Lamarck, 1815, 447-448.

luna (Mueller, 1786) Lamarck, 1815, 448.

lupus (Mueller, 1773) Lamarck, 1815, 448, based on Mueller, —, pl. 20, figs. 14–17.

orbis (Mueller, 1786) Lamarck, 1815, 448.

podura (Mueller, 1773) Lamarck, 1815, 447, based on Mueller, pl. 19, figs. 1–5.— Bory de St. Vincent, 1825a, 84 (syn. Cerc. podura Mueller).

serrata Bory de St. Vincent, 1825a, 84 (syns. Furcularia furcata Lamarck, Vorticella furcata Mueller) (in infusion de foin).

trilobata Bory de St. Vincent, 1825a, 84 (in infusions d'écorce de chêne).

viridis (Mueller, 1774) Lamarck, 1815, 447.

[cercarie Wagener], see Dies., 1858d, 276 (furcocerce cercarie Wagener, syn. of Bucephalopsis aculeatus Dies.).

FURCULARIA

furcata (Mueller, 1786) Lamarck, 1816, 39.—Bory de St. Vincent, 1825a, 84 (syn. Furcocerca serrata).

GALACTOSOMUM Looss, 1899b, 671 (m. lacteum) subf. Haplorchiinæ; 1902m, 512.—Pratt, 1902, 890, 910.

lacteum (Jægers., 1896) Looss [1899, 671]; 1902m, 512 (type).

GANEO Klein, 1905, June 9, 72 (m. glottoides); ganeo, der Prasser, Schwelger. glottoides Klein, 1905, June 9, 72–78, pl. 5, figs. 6–8 (in Rana hexadactyla, intestine); 1905, 14–20, figs. 6–8.

GASTERODISCINÆ Mont.

GASTEROSTOMA Mont., 1888a, 84, see Gasterostomum.

GASTEROSTOMATA Odhn., 1905, 295–296.

GASTEROSTOMATIDÆ Gamb., 1896a, 73.

GASTEROSTOMIDÆ Braun, 1883a, 59; 1893a, 887, 895, 900, 912; 1895b, 136.—
Hoyle, 1890, 539.—Jackson, 1888, 654, includes Gasterost., (Bucephalus, larval form).—Looss, 1899b, 541, 543; 1901, 196.—Luehe, 1901, 488.—Mont., 1888, 14, 52, 92, 105.—Odhn., 1905, 296.—Par., 1887, 335.—Pratt, 1902, 890 (Gasterost.).—Schneidemuehl, 1896, 295.—Sieb., ——.

GASTEROSTOMIDES R. Bl., 1888a, 541 (embraced in Distomiens).

GASTEROSTOMINÆ Mont., 1892, Oct. 7, 214 (subf. of Distomidæ).—Braun, 1893a, 890.—Looss, 1899b, 541.—Much., 1898, 30.—Stoss., 1898, 59.

GASTEROSTOMUM Sieb., 1848, 112, 129, 138 (m. fimbriatum); 1854, 20.—Ben., 1870c, 142.—Bettend., 1897a, 15; 1897, 319.—Biehringer, 1888a, 231, 232, 233.—Brand., 1891d, 12; 1892, 507.—Braun, 1890a, 520; 1890b, 127; 1892a, 568, 577, 583, 586, 589, 603, 605, 608, 614, 629, 635, 336, 640, 641, 644, 659, 660, 671, 672, 673, 682, 683, 696, 700, 704, 708, 709, 711, 713, 715, 718, 721, 726, 727, 732, 736, 757, 760, 773, 806, 807; 1893a, 823, 834, 867, 872, 879, 886, 887, 890, 895, 899, 913, 916, 918; 1895b, 121, 125, 136; 1899, 3.—Carus, 1863, 479.—Dies., 1858e, 313, 360–361 (mentions only fimbriatum); 1859c, 436–437.—Fil., 1855b, 25.—Gamb., 1896, 73.—Hoyle, 1890, 539 (8 sp. in fish, larva Bucephalus), 540.—Larker, 1888, 448, 644, 648, 654, 1889, 540, 1889, Jackson, 1888, 642, 644, 648, 651 (=Bucephalus), 654.—Kath., 1894a, 138.— Lacaze Duthiers, 1854a, 295, 301.—Leuck., 1863, 474, 477, 503, 524.—Lint., 1905, 327 (Bucephalus haimeanus).--Looss, 1893b, 819; 1894a, 105, 158, 179, 191, 202, 203.—Mont., 1888a, 7, 10, 11, 12, 15, 33, 34, 38, 43, 46, 60, 77, 79, 84 (Gasterost.), 92, 105; 1891, 110; 1892, Oct. 7, 214 (g. of Gasterostominæ); 1893, 3, 9, 28, 95, 147.—Moul., 1856a, 12, 175, 178.—Odhn., 1905, 296 (type fimbriatum), 297, 307.—Poche, 1907, 125.—Pratt, 1900, 647; 1902, 890, 891, 908.—School-bloom of the control of the c Schneidemuehl, 1896, 295.—Spengel, 1905, 258.—Ssinitzin, 1906, 688.—Stoss., 1898, 59. —Tennent, 1906, 639 (in Cyclopterus lumpus).—Wagener, 1852, 557-567, figs. 2-3.—Wallenstedt, 1847, 7.—Wolf, 1903, 615, 617.—Ziegler, 1905, 36.—Cf. Bucephalus.

arcuatum Lint., 1900, 277, 278, 297–298, pl. 41, figs 85–90 (in Sarda sarda; Woods Hole); 1901, 416 (in Sarda sarda, Carcharinus obscurus), 427, 446, 447; 1905, 329, 335, 363, 365, fig. 235 (in Caranx hippos, Scomberomorus regalis).—Ten-

nent, 1906, 640 (in Car. obs.).

armatum Mol., 1859, 291 (t. h. Conger conger; Padua); 1859, 820; 1861, 224–226, pl. 4, figs. 4–5, pl. 6, figs. 1, 3.—Braun, 1892a, 752; 1893a, 913.—Dies., 1859c, 436.—Levin., 1881a, 64, 76–78 (in Cottus scorpius; Egedesminde) [see below].-Mont., 1893, 95.—Odhn., 1905, 305 (of Mol., 1861, syn. of Prosorhynchus crucibulum Rud.), 307; 1905, 297, 298 (syn. of Pros. squamatus).—Staff., 1904, May 3, 488 (in Acanthocottus scorpius, Brosmius brosme, Hemitripterus americanus, Hippoglossus hippoglossus).—Stoss., 1885, 161: 1898, 60.—Tennent, 1906, 639, 640, 669, 679 (in Conger conger).—Ziegler, 1883, 538 to (Rhipidocotyle?), 539 (syn. of G. crucibulum Gerv. & Ben.), 548 (in Conger conger).

armatum of Olss., 1868, 56, pl. 5, figs. 107–108, and of Levin., 1881, 76, pl. 3, fig. 4.–

Odhn., 1905, 297, 298 (syn. of Prosorhynchus squamatus).

baculum Lint., 1905, 327, 329, 335, 362, figs. 233, 234 (in Scomberomorus maculatus; Beaufort, North Carolina), based on Gasterost. sp. Lint., 1901, 447, pl. 34, figs. 369-372.

blanchardi Stoss., 1898, 61 (in Labrax lupus; Triest).

clupex Ben., 1870, 67 (in Clupea sprattus; Belgium).—Braun, 1893a, 913.—Tennent, 1906, 639 (in Cl. spr.).—Ziegler, 1883, 539 (in Cl. spr.).

crucibulum (Rud., 1819) Gerv. & Ben., 1859b, 207; 1870, 82, pl. 3, fig. 18.—Braun, 1892a, 784, 789; 1893a, 913.—Dies., 1859c, 425 (to Monost), 437.—Linst., 1878a, 272 (in Conger cassinii).—Mont., 1893, 95.—Odhn., 1905, 305 (of Ben., 1870, syn. of Prosorhynchus aculeatus; of (Rud.) Olss., syn. of Pros. crucibulum.—Stoss., 1892, 66; 1898, 60–61 (in Conger vulgaris; Triest).—Tennent, 1906, 639, 640, 679.—Ziegler, 1883, 539, 548 (syn. of G. armatum Mol.), 563.

fimbriatum Mol., 1859, 819–821, pl. 2. fig. 1 (t. h. Anguilla vulgaris; Padua).— Braun, 1892a, 575, 595, 599, 673, 709, 712, 717, 718, 784; 1893a, 866, 871, 913.— Buttel-Reepen, 1902, 215.—Cobbold, 1879b, 462.—Gamb., 1896, 72.—Mad-

dox, 1867, 97.

GASTEROSTOMUM-Continued.

fimbriatum Sieb.. 1848, v. 1, 129 (in Perca fluviatilis, Lucioperca; [Europe]) as possible adult of Bucephalus polymorphus.—Badcock, 1875a, 145 (in Perca fluviatilis, Lucioperca, carp: larva Bucephalus polymorphus.—Biehringer, 1888a, 230.—Brand., 1891d, 7.—Braun. 1892a, 575, 595, 599, 673, 709, 712, 717, 718, 784: 1893a, 866, 871, 913.—Dies., 1858e, 361 (in Lucioperca sandra, Perca fluviatilis, Esox lucius); 1859c, 435, 436 (cf. Dist. campanula Duj.).—Giard, 1874e, 487.—Haarlem. —. v. 9, 89: —. v. 13, 103, pl. 24, figs. 1–7.—Hausmann, 1896a, 392 (in Luc. sandra); 1897b, 4, 6, 20, 22, 34–35, pl. 1, figs. 6–8 (in Luc. sandra).—Juel. 1889, 12, 36.—Kowal., 1894, 4: 1894, 221.—Linst., 1878a, 208 (in Perca fluviatilis); 1878. 230.—Looss, 1885b, 11, 14: 1892a, 122: 1894a, 203, 205, 206, 207.—Mont., 1888a, 34.—Mueh., 1898, 30.—Odhn., 1905, 296 (type of genus), 300, 301, 303.—Olss., 1893, 12.—Par., 1887, 336.—Poche, 1907, 125.—Stoss., 1883, 119: 1898, 59–60 (in Anguilla vulgaris: Triest).—Tennent, 1906, 637, 638, 641, 643, 665, 666, 677, 678, 682 (in Perca, Lucioperca).—Wagener, 1852, 565–567; 1857, 45, pl. 24, figs. 1–7: 1858, 250: 1860, 165 (syn. Dist. campanula).—Ziegler, 1883, 491; 1883, 537, 538, 539, 541–567, pl. 33, figs. 16, 18–22, 27, 28.—Also reported for Abramis alburnus, Blicca bjoerkna, Blicopsis abramo-rutilus, Gobio fluviatilis. Leuciscus erythrophthalmus. Lota vulgaris, Squalius leuciscus.

gadorum (Rathke, 1799) Linst., 1878a, 238 (in Gadus morrhua; G. virens).—Braun, 1893a, 871.

galeatum (Rud., 1819) Stoss., 1898. 62 (in Lichia amia; Triest).

gorgon Lint., 1905, 327, 335, 364, figs. 240–242 (in Seriola lalandi: Beaufort. North Carolina).

gracilescens (Rud., 1819) Wagener, 1852, 563, pl. 16, fig. 3.—Braun, 1892a, 673; 1893a, 866, 880, 913.—Cobbold, 1872b, 92; 1879b, 52, 462, fig. 78.—Dies., 1858e, 361 (to Rhipidocotyle) (in Lophius piscatorius).—Gamb., 1896, 72.—Hausmann, 1897b, 34.—Linst., 1878a, 223 (to Rhipidocotyle).—Lint., 1905, 327, 335, 353, 357, 360, 365, 370, 402, 410, 414, figs. 230-232, 236-239 (in Caranx hippos, Menidia menidia. Opsanus tau, Paralichthys albiguttus, Pomatomus saltatrix, Spheroides maculatus, Stolephorus brownii, Tylosurus marinus).—Maddox, 1867, 97.—Mol., 1859, 821 (Loph. pisc.; Padua).—Odhn., 1905, 296, 302, 303.—Sons., 1891, 258 (in Loph. pisc.).—Stoss., 1890, 44; 1898, 61-62 (in Loph. pisc.; Triest).—Tennent, 1906, 637, 638, 640, 664, 666, 679, 682 (larva = Bucephalus haimeanus), pl. 40, fig. 40, pl. 41, figs. 44-50, 52, 53, pl. 42, figs. 54-58 (in Loph. pisc.).—Ziegler, 1883, 538, 539.—Also reported for Gadus æglefinus, G. minutus, Esox. Molya.

illense Ziegler. 1883, 543 (in Esox lucius. Leuciscus erythrophthalmus: Strassburg).—Hausmann, 1896a, 392.

laciniatum Mol., 1859, 821 [apparently lapsus for fimbriatum].—Par., 1894, 167 (in Anguilla vulgaris; Padova, Triest, Cagliari).

minimum Wagener, 1852. 558–563, fig. 2 (t. h. Trigla microlepidota).—Braun, 1889a, 356; 1892a, 599. 673; 1893a. 880, 913.—Dies., 1858e. 361. 362 (to Rhipidocotyle) (in Tr. micr.).—Odhn., 1905, 296.—Tennent, 1906. 637, 638 (in Tr. micr.).—Ziegler, 1883, 538, 548 (in Tr. micr.).

minimum Stoss., 1887, 96, pl. 10, fig. 41 (in Labrax lupus: Triest); 1898, 61.— Hausmann, 1897b, 34.

ovatum Lint., 1900a, 269, 297 (in Lobotes surinamensis: Woods Hole, Mass.); 1901, 416, 457.—Tennent, 1906, 640, 679 (in Lob. sur.).

pusillum Staff., 1904, May 3, 494-495 (in Stizostedion vitreum; Canada).

species Lint., 1901, 447. See G. baculum.

species Lint., 1900, 269, 298, pl. 41, fig. 9.

species Ben., 1870, 51.

tergestinum Stoss., 1883, 119–120. pl. 2. fig. 5 (in Gobius niger, G. jozo; Triest); 1898, 61; 1901, 91–92 (3–4).—Braun, 1892a, 642; 1893a, 874, 913.—Mont., 1888, 14; 1893, 95.

triglæ Ben., 1870, 30, pl. 3, fig. 15 (in Trigla hirundo: Belgium).—Braun, 1893a, 913.—Tennent, 1906, 639, 679 (in Tr. gurnardus).—Ziegler, 1883, 539 (in Tr. gurn.).

riperæ Ben., 1870, 26, pl. 3, fig. 17 (in Trachinus vipera; Belgium).—Braun, 1892a,
568; 1893a, 913.—Tennent, 1906, 639, 640 (in Tr. vip. Cuv.).—Žiegler, 1883,
539 (in Tr. vip.).

vivæ Ben., 1870, 25 (in Trachinus draco; Belgium).—Braun, 1893a, 913.—Odhn., 1905, 296.—Tennent, 1906, 639 (in Tr. dr.).—Ziegler, 1883, 539 (in Tr. dr. L.).

GASTROCOTYLE Ben. & Hesse, 1863; 1864, 96, 117-118 (m. trachuri).-1890a, 414, 498, 511, 517, 523, 540, 541, 546; 1893a, 890.—Cerf., 1895h, 918; 1896, 514; 1899a, 391.—Cunningham, 1887a, 279.—Gamb., 1896, 73.—Hoyle, 1890, 539.—Mont., 1888a, 8, 11, 13, 66, 86, 89, 101; 1892, Oct. 7, 213 (gen. of Microcotylinæ); 1903, 336 (subf. Axininæ).—Pratt, 1900, 646, 653, 654, fig. 42.—Tasch., 1879, 69; 1879, 257–258.—Will.-Suhm, 1870, 9, 11.

trachuri Ben. & Hesse, 1863, 118; 1864, 118, pl. 13, figs. 1–8 (in Caranx trachurus).— Braun, 1890a, 407, 414, 541, 549 (in Car. tr.; Atlantic Ocean, Mittelmeer), 550.—Par., 1894, 595.—Par. & Perugia, 1890, 8.—Pratt, 1900, 657, fig. 42.—

Tasch., 1879, 258 (in Car. tr.).—Will.-Suhm, 1870, 10.

GASTRODISCINÆ Mont., 1892, Oct. 7, 214 (subf. of Amphistomidæ).—Braun, 1893a, 890.—Looss, 1899b, 541.

GASTRODISCUS Leuck., in Cobbold, 1877e, 233–239 (m. sonsinoii); 1877f, 326.— Braun, 1892a, 569, 581, 603, 614, 645, 669, 677, 682, 683, 688, 696, 699, 705, 715, 716; 1893a, 879, 886, 890, 892, 895, 904, 906, 918; 1895b, 136; 1903, 3 ed., 715, 716; 1893a, 879, 886, 890, 892, 895, 904, 906, 918; 1895b, 136; 1903, 3 ed., 146.—Cobbold, 1877f, 326.—Couzin, 1885a, 426.—Fischder, 1901, 374; 1902a, 7, 46; 1903h, 488, 489 (m. polymastos) (in horses; Egypt).—Gamb., 1896a, 73.—Hoyle, 1890, 539.—Jackson, 1888, 642, 644, 654.—Looss, 1894, 24; 1895, 11; 1896b, 13, 14, 18, 151, 173, 178, 184, 188; 1899b, 541, 770; 1902m, 440, 442, 638, 676.—Mont., 1888a, 8, 12, 15, 26, 27, 34, 35, 41, 42, 52, 53, 54, 56, 57, 91, 103; 1892, Oct. 7, 214 (gen. of Gastrodiscinæ).—Piana & Stazzi, 1900, 523.—Poir., 1883, 79.—Pratt, 1902, 887, 892.—Schneidemuehl, 1896, 295 (Gastrodiskus).—Shipley, 1905, v. 6 (1), 4, 8.—Sons., 1895, Aug.—Sept., 179, 180, 181, 183, 184, 185, 186, 1 pl., figs. 1–7.—Ward, 1903, 865 (in horses and cattle in Egypt; one (G. hominis) in man).

1883: Gasterodiscus Ziegler, 1883, 545.

1896: Gastrodiskus Schneidemuehl, 1896, 295.

1898: Gastrodiseus Kowal., 1898h, 158, misprint.

ægyptiacus (Cobbold, 1876) Rail., 1893, 379 (syns. Hemist. sp. Sons., 1876; Diplost. ægyptiacum Cobbold, 1876; Cotylegaster cochleariforme Sieb., 1877; G. sonægyptiacum Cobbold, 1876; Cotylegaster Cochleariforme Sieb., 1877; G. sonsinoii Cobbold, 1877; G. polymastos Lkt., 1880).—Bettend., 1897a, 38; 1897, 342.—Brand., 1898a, 205 (13).—Fischder., 1902a, 46 (syn. of G. polymastos Leuck); 1903h, 494, 497.—Gronkowski, 1902a, 514, 520 (7, 13).—Kowal., 1898h, 158 (55) (Gastrodiseus).—Looss, 1896, 13–32, 149, 177–185, pl. 1, figs. 4–8, pl. 2, figs. 9–15, pl. 11, fig. 115, pl. 12, figs. 122–124, pl. 13, figs. 135–139 (syns. G. polymastos, G. sonsinoi, Hemist. sp., Diplost. ægyptiacum); 1907, Feb. 1, 134.—Moore, 1898, July 15, 531; 1897, 183.—Sons., 1895, 180, 181, 183, 184, 187, fig. 3; 1896, 294, 298, 309, 310.—Ward, 1895, 338 (in Equus capallus).—Also reported for Equus asigus, E. mulus caballus.)—Also reported for Equus asinus, E. mulus.

hominis (Lewis & McConnell, 1876) Sons., 1896, 298.—Braun, 1903, 3 ed., 146-147, fig. 91; 1906, 149, fig. 82 (syn. Amphist. hominis).—Conyngham, 1904, IX, 17, 663.—Fischder., 1901, 374; 1902a, 46–47 (in Homo; India).—Looss, 1905, 110.—Shipley, 1905, v. 6 (1), 4 (thinks Equus is probably normal host).—Stephens, 1903, Feb., 7–12, figs. 1–4.—Ward, 1903, 704; 1903, 864 (in man), 865.

stephens, 1905, Feb., 7–12, Igs. 1–4.—ward, 1905, 704; 1903, 864 (In Man), 865. polymastos Leuck., 1880 (in Equus caballus; Egypt).—Brand., 1898a, 205, 214 (13, 23).—Braun, 1892a, 568, 580, 591, 593, 602, 605, 607, 608, 638, 640, 645, 646, 649, 654, 657, 661, 662, 663, 688, 703, 717, 733; 1893a, 874, 906; 1893d, 466.—Collin, 1891a, 86 (syn. of G. sonsinoi).—Fischder., 1901, 374; 1902a, 46 (syns. Diplost. ægyptiacum Cobbold, G. ægyptiacus Looss, G. sonsinoi Cobbold) (in Equus caballus, E. mulus, E. zebra).—Gronkowski, 1902a, 514 (7).—Lejtenyi, 1880a, 125–146 (anatomy), pls. 1–3; 1881a, 22 pp., 3 pls.—Lingard, 1905, 50.—Looss, 1885a, 392, 397 (Gasterodiscus); 1885b, 5, 10 (Gasterodiscus); 1894a, 146, 237; 1896b, 13 (syn. of G. ægyptiacus).—Mont., 1888a, 15, 26, 60.—Rail., 1887, 406–408 (in Sénégal), 494.—Richard, 1896.—Schneidemuehl, 1896, 303 (Gastrodiskus polymastus).—Sons. 1895, 124 (syn. Schneidemuehl, 1896, 303 (Gastrodiskus polymastus).—Sons., 1895, 124 (syn. of G. sonsinoi), 179.—Stephens, 1906, 9.—Ward, 1895, 338 (see G. ægyptiacus in Equus caballus).—Ziegler, 1883, 545 (Gasterodiscus).—Zuern, 1882, 222.

secundus Looss, 1907, Feb. 1, 134–136, figs. 3–4 (in mules; Assam).

sonsinoii Cobbold, 1877e, 233–239, 1 pl., 1 fig. (in Equus; Egypt), syn. Diplost. sonsinoii Cobbold 1879b, 38, 359, fig. 62).—
R. Bl., 1888a, 636.—Braun, 1893a, 906 (sonsinoi); 1893b, 186 (sonsinoi); 1906, 149.—Collin, 1891a, 86–88 (in zebra; Kataui Mbuga, Ostafrika.)—Fischder., 1902a, 46 (sonsinoi) (syn. of G. polymastos); 1903h, 488 (sonsinoi).—Girard, 1880a, 1xix–1xx (sonsinoi).—Lejtenyi, 1881a, 2 (sonsinoi).—Lingard,

GASTRODISCUS-Continued.

1905, 50 (syn. G. polymastos).—Looss, 1896b, 13 (syn. of G. ægyptiacus).—Mégnin, 1880, Aug. 15, 782–784, figs. 1–2 (in horse); 1881, v. 2, 250–252, figs. 2.—Moore, 1897b, 167; 1897a, 183–184 (sonsinoi); 1897c; 1898a, 531.—Poir., 1883, 74 (sonsinoi).—Sons., 1895, 124 (syn. G. polymastos).—Theobald, 1900, 50.—Ward, 1895, 338 (in Equus caballus).

sonsinoi Girard, 1880a, Ixviii, for sonsinoii.

sonsinoni Poir., 1883, 74, for sonsinoii.

sonsinonis Cobbold, 1879b, 359, 463, fig. 62, for sonsinoii.

GASTRODISEUS Kowal., 1898h, 158 (55), for Gastrodiscus.

ægyptiacus (Cobbold, 1876) Kowal., 1898h, 158 (55).

GASTRODISKUS Schneidemuehl, 1896, 295, 303, for Gastrodiscus.

polymastus Schneidemuehl, 1896, 303, for polymastos.

GASTROTHYLAX Poir., 1883, 76–79; γαστήρ, stomach; θῦλαζ, pocket, pouch; [type crumenifer].—Brand., 1891d, 17; 1898a, 193–225, pls. 8–9; 1898, 33 pp.—Braun, 1892a, 576, 663, 696, 699, 715, 720, 721, 739; 1893a, 879, 886, 890, 892, 895, 904, 907, 918; 1893d, 466; 1892, 49.—Darr, 1902, 651.—Fischder., 1901, 370; 1902a, 7, 26–27 (type crumenifer); 1903h, 488, 489, 491, 493, 496, 497, 498, 556 (diagnosis), 557 (type crumenifer), 557–591.—Gamb., 1896a, 73.—Gronkowski, 1902a, 515 (8).—Hoyle, 1890, 539.—Looss, 1894a, 146, 175; 1894, 17, 22, 24; 1896b, 5, 7, 8, 9, 31, 32, 171, 173, 176, 177, 178; 1898a, 459; 1899b, 541; 1902m, 638, 676.—Mont., 1888a, 7, 9, 12, 15, 52, 91, 103; 1892, Oct. 7, 214 (gen. of Amphistominæ) (Gastrotylax); 1893, 27.—Pratt, 1902, 887, 892.—Shipley, 1905, v. 6 (1), 8.—Sons., 1895, 184, 185, 186.—Ssinitzin, 1906, 688.—Stiles, 1898a, 24.

1892: Gastrotylax Mont., 1892, 7 Oct., 214, misprint.

xgyptiacus Looss, 1898a, 459, apparently lapsus for gregarius.

cobboldi Fischder., 1901, 372 (for cobboldii); 1902a, 31, 32, 33 (in Palonia frontalis, Java; Bos kerabau, Ceylon; B. taurus, China); 1903h, 542, 575, 579–583, figs. M, 63–66 (from Ceylon in Bos kerabau, from China in Bos taurus), 585, 586, 587, 588, 589, 591; 1904, 459, 463.

cobboldii Poir., 1883, 77–79, pl. 2, fig. 3 (in Palonia frontalis; Java).—Braun, 1892a, 568, 739; 1893a, 873, 907.—Fischder., 1901, 372; 1903h, 575, 579 (syn. of G. cobboldi).—Sons., 1895, 185.—Stiles, 1898a, 24, 67, 69, 139, fig. 63.—Also reported for Bos frontalis.

compressus Brand., 1898a, 197, 219-220, 222 (5, 27-28), pl. 9, figs. 9-11 (in Bos indicus; Vien. Mus.); 1898, 219-220.—Fischder., 1901, 371; 1902a, 28 (in Bos taurus indicus); 1903h, 560, 563-565, figs. 48-49 (in Bos indicus), 567.

crumenifer (Crep., 1847) Otto, 1896, 95–97, figs. 3, 16 (for G. crumeniferum).—
Bettend., 1897a, 38; 1897, 342.—Brand., 1898a, 195, 197, 199, 201, 204, 216–219,
220, 221 (3, 5, 7, 9, 12, 24–27), pl. 9, figs. 1–8.—Fischder., 1901, 371; 1902a,
25, 27–28, 30 (in Bos taurus indicus, Calcutta; Bos kerabau, Ceylon); 1903h,
557 (type), 557–563, 564, 566, 567, 568, 570, 571, 572, 574, figs. H, 44–47 (in
Bos zebu, B. kerabau).—Linst., 1906, 175 (in Bos bubalus).—Stiles, 1898a,
24, 67, 68, 69, 140, figs. 57–62.

crumeniferum (Crep., 1847) Poir., 1883, 77.—Braun, 1893a, 907.—Fischder., 1901, 371; 1903h, 557–563, figs. H, 44–47, 563, 564, 566, 567, 568, 570, 571, 572, 574 (syn. of G. crumenifer) (in Bos zebu, B. kerabau).—Linst., 1906, 175 (in Bos bubalus).—Rail., 1903a, 378.—Sons., 1896, 311.—Stiles, 1898a, 67.—Ward, 1895, 332 (in Bos taurus).

elongatum Poir., 1883, 76–77, pl. 2, fig. 2 (in Palonia frontalis; Java).—Braun, 1892a, 739; 1893a, 873, 907.—Fischder., 1903h, 574, 575 (syn. of G. elongatus) (in P. front.; Java).—Mont., 1888a, 7.—Sons., 1895, 185.—Stiles, 1898a, 24, 67, 70, 139, fig. 64.

elongatus [Poir., 1883, 73–79 (in Palonia frontalis; Java)].—Brand., 1898a, 197, 198, 204, 223–225 (5, 6, 31–33), pl. 8, figs. 1–14 (in Anoa depressicornis; Zool. Gart., Berlin).—Fischder., 1901, 372; 1902a, 30–31 (in P. front., Java; Anoa depr., Bos sp., Africa; Bos taurus, China); 1903h, 451, 512, 542 (in Bos taurus, Canton Fu-mui, China), 574–579, 580, 581, 587, figs. L, 59–62 (in P. front., Bos kerabau, Anoa depr.); 1904, 463.—Gronkowski, 1902a, 520 (13).—Mont., 1888a, 7.

GASTROTHYLAX—Continued.

gregarius Looss, 1896b, 5–13, 170–177, pl. 1, figs. 1–3, pl. 11, fig. 116; pl. 12, figs. 119–121 (in Buffle; Alexandria, Egypt); 1898, 459; 1902m, 638.—Bettend., 1897a, 38; 1897, 342.—Brand., 1898a, 197, 198, 199, 222–223 (5, 6, 7, 30–31), pl. 9, figs. 12–16.—Fischder., 1901, 371; 1902a, 28 (in Egyptische Buffel); 1903b, 533; 1903h, 565–566, 584.—Kowal., 1898h, 158 (55).—Otto, 1896, 89–94, figs. 1–2, 11, 14, 15; 1896, 10–13.—Rossbach, 1906, 401.—Sons., 1896, 298, 311.—Stiles, 1898a, 24, 67, 71, 139, figs. 65, 66.

mancupatus Fischder., 1901, 371 (in African cattle); 1902a, 31–32, 33 (in Bos taurus; E. Africa); 1903h, 566, 584, 588, 589, 590, 591, figs. 67–71 (in Bos taurus, Dinka; Denjarinder, Afrika; Bos taurus, Africa, Nossi-Bé).

minutus Fischder., 1901, 372 (in Antilope sp., Tragelaphus scriptus, Kamerun); 1902a, 33–34; 1903h, 588–591, fig. N; 1904, 463.

spatiosus Brand., 1898a, 197, 220–221 (5, 28–29), pl. 8, figs. 15–16 (in Bos taurus, Dschidda, Arabian coast of Red Sea).—Fischder., 1901, 371; 1902a, 29, 31 (in B. taurus, Dschidda); 1903h, 566–570, 571, 581, 583 (in B. taurus), 584, 585, 586, figs. 50-54.

synethes Fischder., 1901, 371 (in Bos kerabau); 1902a, 29–30, 31 (in B. ker.; Ceylon); 1903h, 496, 539, 570–574, 575, 577, 578, 579, 580, 581, 584, 585, 586, figs. 55–58,

J. K. (in B. ker.; Ceylon).

GASTROTYLAX Mont., 1892, Oct. 7, 214 (gen. of Amphistominæ) (for Gastrothylax). GENARCHES Looss, 1902m, 732 (Progonus Looss, renamed; type mülleri).—Odhu.,

1905, 364, 365, 366. mülleri (Levin., 1881) [Looss, 1902m, 732] Odhn., 1905, 363, 365–366, pl. 4, figs, 8–9.

GIRODACTILIDÆ Sons., 1890, 175, for Gyrodactylidæ.

GIRODACTYLIDÆ Mont., 1888, 8, 13, 15, for Gyrodactylidæ.

GIRODACTYLUS Mont., 1888, 83, for Gyrodactylus.

GLAPHYROSTOMIUM Pratt, 1902a, 889, misprint for Glaphyrostomum.

GLAPHYROSTOMUM Braun, 1901g, 942 (tod. adhærens); 1902b, 129.—Pratt, 1902a, 889 (Glaphyrostomium), 907.

1902: Glaphyrostomium Pratt, 1902a, 889, misprint.

adhærens Braun, 1901g, 942 (in Mycothera sp., Gallinula kioloides Puch.; Brazil); 1902b, 130, 133.

propinguum Braun, 1901g, 942 (in Dendrocalaptes scandens; Brazil); 1902b, $13\bar{2}$ -133.

GLENOCERCARIA Dies., 1858d, 244 (type?). [See p. 385.]

flava (La Valette, 1855) Dies., 1858d, 244–245 (includes Cerc. ephemera Sieb., Histrionella eph. Sieb., Cerc. flava La Valette (in Planorbis corneus; Berlin, Heidelberg); 1858e, 326, 327 (to Monost.)—Linst., 1873, 1 (young of Monost. flavum Mehlis).—See Typhlocœlum 1902 (type).

tophocerca (Fil., 1857) Dies., 1858d, 245 (in Paludina impura; Turin).

melanoglena (Dies., 1855) Dies., 1858d, 245 (in salt water).

GLOSSIDIUM Looss, 1899b, 591-592, 594 (tod. pedatum); ή γλῶσσα, tongue.—Braun, 1902b, 55.—Heymann, 1905, 93.—Odhn., 1902, 40, 42.—Pratt, 1902, 888 (related to Plagforchiinæ), 899.

pedatum Looss, 1899b, 592, 705–706, fig. 27 (in Bagrus bayad, B. docmac; Cairo).—

Luehe, 1900, 561.

GLOSSOCOTYLE Ben. & Hesse, 1863; 1864, 96, 102 (m. alosæ).—Braun, 1890a, 414, 477, 516, 517, 522, 546.—Cerf., 1895h, 918, 920; 1896, 514, 515.—Mont., 1888, 11, 16, 86, 89, 99; 1903, 336 (syn. of Octobothrium).—Tasch., 1879, 239.

alosæ Ben. & Hesse, 1863, 1864, 102–103, pl. 9, figs. 11–18 (in Alosa vulgaris).— Braun, 1890a, 408.—Tasch., 1879, 244 (to Octobothrium).

finta Mont., 1888, 13.

GLYPHICEPHALUS Looss, 1901l, 620-621 (tod. solidus); 1902m, 570-571 (type solidus), 576, 583, 584, 586, 589, 591, 593, 594, 596, 599, 603, 609, 610, 611, 612, 616 (diagnosis).—Pratt, 1902, 890, 909.

crassus Looss, 1901l, 568, 620 (in Thalassochelys corticata; Egypt); 1902m, 575 (to Epibathra as type), 876.

lobatus Looss, 19011, 619 (in Chelone mydas; Egypt); 1902m, 416, 417, 573-575, 576, 596, 876, 877, pl. 26, figs. 81–82, pl. 27, fig. 91.

- GLYPHICEPHALUS-Continued.
 - solidus Looss, 19011, 7. Nov., 619 (in Chelone mydas; Egypt); 1902m, 571–573, 574, 587, 591, 616, 876, 877, 888, pl. 26, figs. 79, 80, pl. 27, figs. 92, 93, pl. 32, fig. 178.
- GLYPTHELMINS Staff., 1905, Apr. 11, 686–687 (m. quieta); γλυπτός, carved; ἕλμινς, worm.
 - quicta (Staff., 1900) Staff., 1905, Apr. 11, 686 (in Hyla pickeringii, Rana catesbiana, R. virescens; Canada) (syn. Opisthioglyphe endoloba Duj. of Staff., 1900, 403).
- GORGODERA Looss, 1899b, 551, 605–606 (tod. cygnoides); $\gamma o \rho \gamma \delta \varepsilon$, motile; $\dot{\eta} \delta \varepsilon \rho \eta$, neck; 1901b, 202; 1901l, 557, 558; 1902m, 478, 480 (includes: cygnoides, amplicava, simplex), 785, 797, 844, 848, 850, 851, 854, 856, 857, 858, 859, 860, fig. 1, 862 (Gorgoderinæ, Gorgoderidæ).—Braun, 1901b, 9.—Odhn., 1902, 65.—Osborn, 1903, 257.—Pratt, 1902, 888, 901.—Ssinitzin, 1906, 685.
 - amplicava Looss, 1899b, 606, 607 ("D. cygnoides var. A" of Bensley, 1897, renamed);
 1901b, 202, 209; 1902m, 480, 798, 837, 844, 854, 857.—Staff., 1902, 412, 419–421 (syn. D. cygnoides var. A of Bensley); 1905, Apr. 11, 687 (in urinary bladder of American frogs and toads).
 - attenuata Staff., 1902, 418–419 (in Rana catesbiana, R. virescens; Canada).—1905, to Gorgoderina.
 - cygnoides (Zed., 1800) Looss, 1899b, 606; 1901b, 202, 209; 1902m, 444 (Cerc. macrocerca), 480, 797, 798, 837, 844, 851, 854, 857, 862.—Kowal., 1902d, (9) 27 (Cerc. macrocerca in Cyclas cornea).—Luehe, 1901, 54.—Osborn, 1903, 257.—Ssinitzin, 1905, 33 (of Looss syn. of G. loossi).—Staff., 1905, Apr. 11, 687 (in urinary bladder of American frogs and toads).
 - loossi Ssinitzin, 1905, 33–34, pl. 3, figs. 30–31 (G. cygnoides of Looss, renamed); 1906, 682.
 - opaca Staff., 1902, 416-417 (in Bufo lentiginosus; America).
 - pagenstecheri Ssinitzin, 1905, 34-36, pl. 2, figs. 21-27 (cygnoides of Pag., renamed); 1906, 682, 683 [=Dist. cygnoides Zed. of Pag.].
 - simplex Looss, 1899b, 606, 607 (D. cygnoides var. B of Bensley, 1897, renamed);
 1901b, 202, 209; 1902m, 480, 798, 844, 851 (type of Gorgoderina), 863.—Staff.,
 1902, 412, 417-418 (in Rana catesbiana; America).
 - *translucida* Staff., 1902, 413–416 (in Bufo lentiginosus, Rana virescens; America).—Osborn, 1903, 256, 257, 258.
 - varsoviensis Ssinitzin, 1905, 36–37, pl. 1, fig. 1, pl. 3, figs. 32–34; 1906, 682 (in frogs; Warschau).
 - vitelliloba (Olss., 1876) Ssinitzin, 1906, 682.
- GORGODERIDÆ Looss, 1901, 558; 1902m, 485, 785, 810, 824, 843–866, 844 (diagnosis), 857 (includes Gorgoderinæ, Anaporrhutinæ).—Ssinitzin, 1906, 682 (of frogs).
- GORGODERINA Looss, 1902m, 851 (tod. simplex), 857 (Gorgoderidæ, Gorgoderinæ), 858, 859, 860, 862¹, fig. 2.—Ssinitzin, 1906, 685 (in fish).—Staff., 1905, Apr. 11, 687.
 - attenuata (Staff., 1902) Staff., 1905, Apr. 11, 687 (American toads and frogs).
 - opaca (Staff., 1902) Staff., 1905, Apr. 11, 687 (in American frogs and toads).
 - simplex (Looss, 1899) Looss, 1902m, 857, 862, 863.—Staff., 1905, Apr. 11, 687 (American frogs and toads).
 - translucida (Staff., 1902) Staff., 1905, Apr. 11, 687 (in American frogs and toads). vitelliloba (Olss., 1876) Looss, 1902m, 857, as doubtful member of genus.—Ssinitzin, 1905, 37, pl. 1, fig. 2, pl. 3, figs. 35–37; 1906, 685.
- GORGODERINÆ Looss, 1899b, 604; 1901b, 202; 1901l, 558; 1902m, 478, 480, 485, 797, 844, 857 (includes: Gorgodera, Gorgoderina, Phyllodist., Catoptroides), 863 (diagnosis).—Braun, 1901b, 9.—Luehe, 1901, 488.—Odhn., 1902, 65, 67.—Pratt, 1902, 888 (Phyllodist., Gorgodera).
- GRUBEA Dies., 1858e, 315, 385 (m. cochlear); 1859c, 444 (syn. Pleurocotylus Gerv. & Ben.).—Braun, 1890a, 518.—Mont., 1888, 84.—Tasch., 1879, 248 (syn. of Pleurocotyle).
 - cochlear Dies., 1858e, 385 (in Scomber scombrus).—Hoyle, 1890, 539.—Tasch., 1878, 575; 1879, 248 (syn. of Pleurocotyle scombri).
 - scombri (Grube, 1855) Ben. & Hesse, 1864, 100 (to Pleurocotyle).

(GYMNOCEPHALA) Leidy, 1877, 201 (as subg. of Dist.) [not Gymnocephalus Bloch, Schneider, 1801, fish; Geoffry, 1809, birds]. Apparently for Cerc. (Gymnocephala) Dies., 1858, 246.

ascoidea Leidy, 1877, 201 (in Planorbis parvus).

(GYMNOCEPHALA) Dies., 1858d, 246, (type?) as subg. of Cercaria.

agilis (Fil., 1857).—Dies., 1858d, 248–249 (in Lymnæus stagnalis; Turin).

brunnea (Dies., 1850) Dies., 1858d, 247 (in Lymnæus stagnalis).—See Dist. echinatum.

coronata (Fil., 1855) Dies., 1858d, 249, 250.

fallax (Dies., 1850) Dies., 1858d, 247–248, 262 (in Lymnæus stagnalis, Paludina vivipara).—See Dist. militare.

minuta (Nitzsch, 1817) Dies., 1858d, 246 (in various fresh-water mollusks; Halle). proxima (Lespés, 1857) Dies., 1858d, 249–250.

renalis (Fil., 1855) Dies., 1858d, 265–266 (in Helix adspersa; Turin).

sagittata [Lespés, 1857] Dies., 1858d, 249 (in Buccinum (Nassa) reticulatum).

setifera (Mueller, 1850) Dies., 1858d, 250–251 (in sea; Triest).

tuberculata (Fil., 1857) Dies., 1858d, 248 (in Paludina impura; Turin).

GYMNOCEPHALÆ Dies., 1858d, 246, see Gymnocephala.

GYMNOPHALLINÆ Odhn., 1905, 314.

GYMNOPHALLUS Odhn., 1900, 12–23 (tod. deliciosus); 1905, 312, 313, 314.—Looss, 1901b, 200; 1902m, 839.—Luehe, 1900, 505, 506; 1901, 488; 1904, 79–82.—Marshall & Gilbert, 1905, 479.—Pratt, 1902, 889, 902.—Ward, 1901, 177.

bursicola Odhn., 1900, 14, 20–21, 22, fig. 4 (in Somateria mollissima; west coast of Sweden); 1905, 312, 313 (includes Leveithodendrium somateriae e. p. of Jameson).

choledochus Odhn., 1900, 14, 18–19, 22, fig. 3 (in Vulpanser tadorna; loc.?); 1905, 312, 313–314 (in Somateria mollissima at Eisfjord, W. Spitzbergen, and S. spectabilis at Franz-Joseph's Fjord, Greenland).

deliciosus (Olss., 1893) Odhn., 1900, 14–17, 18, 19, 20, 21, 22, figs. 1–2 (in Larus argentatus, L. canus, L. fuscus); 1905, 311 (in Larus glaucus, L. marinus).

micropharyngeus (Luehe, 1898) Odhn., 1900, 14, 17, 22.—Luche, 1900, 505; 1901, 57. somateriæ (Levin., 1881) Odhn., 1900, 14, 19–20, 21, 22; 1905, 311–313, pl. 2, fig. 8 (in Somateria mollissima, S. spectabilis).

GYN.ECOPHORA Weinland, 1858, 87 (family name); 1859, 281.—Stiles & Hass., 1898a, 90, 94.

GYN.ECOPHORUS Dies., 1858e, 312, 356 (type hæmatobius).—Braun, 1893a, 880, 885, 894, 912; 1903, 3. ed., 168 (syn. of Schistosomum).—Cobbold, 1879b, 39; 1855a, 498 (syn. of Bilharzia).—Dunglison, 1893, 502.—Hoyle, 1890, 539 (= Bilharzia).—Huber, 1896a, 580 (syn. of Bilharzia).—Leuck., 1863, 617.—Mont., 1888, 92.—Rail., 1893a, 371.—Simon, 1897, 99.—Stiles & Hass., 1898a, 90, 93, 94.—Stoss., 1892, 4, 5.—Ward, 1895, 253.

bovis (Sons., 1876) Rail., 1893, 375.—Dolley.—Stiles, 1898a, 60.

crassus (Sons., 1888) Stoss., 1892, 6 (syn. Bilharzia bovis) (in Bos taurus ægyptus, Ovis aries).—Rail., 1893a, 375.—Stiles, 1898a, 60.—Ward, 1895, 332 (in B. taurus), 335 (in O. aries).

hæmatobius (Bilharz, 1853) Dies., 1858e, 356-357 (in Homo); 1859c, 480.—Aitken, 1866, 804, 840; 1872, 205 (to Bilharzia).—R. Bl., 1888a, 636.—Cobbold, 1866a, 6.—Harley, 1864a, 62 (to Dist.).—Huber, 1894, 298.—Rail., 1892, 161-164 (embryo); 1893a, 371.—Stiles, 1898a, 58.—Stoss., 1892, 5-6 (in Homo; Egitto, Nubia, Tunisia, Natale, Capo, Costa d'oro, Arabia; Cercopithecus fuliginosus in Africa).—Ward, 1895, 253, 256, 328 (in Homo), fig. 7; 1903, 872.

magnus (Cobbold, 1859) Stoss., 1892, 6 (syn. Bilharzia magna) (in Cercopithecus fuliginosus; Africa).—Braun, 1901e, 311.

GYRADACTYLIDÆ Mont., 1888a, 66, 107, see Gyrodactylidæ.

GYROCOTYLE Dies., 1850a, 408 (m. rugosa); 1859a, 492; 1859c, 447.—Mont., 1889c, 228–230 (Amphiptyches G. & W.); 1890b, 327–329; 1896, 153. [Cestodaria.]

rugosa Dies., 1850a, 408 (in Antilope pyarga; Port Natal); 1859a, 492 (in Mactra edulis; Valparaiso); 1859c, 447.

urna (Grube & Wag., 1852) Mont., 1896, 151.

GYRODACTYLE Moul., 1856a, 10, for Gyrodactylus.

GYRODACTYLEÆ Cerf., 1899a, 452.

GYRODACTYLIDÆ Cobbold, 1877f, 326; 1879b, 4.—R. Bl., 1888a, 541 (embraced in Polystomiens).—Braun, 1890a, 463, 465, 469, 511, 516, 517, 519, 523, 533, 538, 542; 1890b, 127; 1893a, 890.—Cerf., 1899a, 452.—Gamb., 1896a, 53, 61, 73.— Goto, 1899, 291.—Jackson, 1888, 642, 654 (includes: Gyrodactylus, Dactylogyrus, Calceost., etc.).—Maclaren, 1904, 583, 597, 598, 599, 600, 601.—Mont., 1888a, 8, 13, 15 (Girodactylidæ); 1889, 116; 1891, 108, 109; 1905, 80.—Par. & Perugia, 1889, 745.—Scott, 1901, 141.—Sons., 1890, 175 (Girodactilidæ).— See also next entry.

GYRODACTYLIDÆ "Ben. & Hesse, 1863," ——,—Hoyle, 1890, 539 (includes: Gyrodactylus, Dactylogyrus, Tetraonchus, Diplectanum, Calceost., Sphyranura).—Mont., 1888, 7, 8, 10, 11, 14, 20, 24, 37, 66, 70, 86, 88, 90, 101, 107, 108; 1903, 336 (raised from subf. to family rank; subf.: Gyrodactylinæ (Gyrodactylus, Dactylogyrus); Tetraonchinæ (Tetraonchus = Ancyrocephalus = Amphibdella = Dactylodiscus); Diplectaninæ (Diplectanum); 1892, Oct. 7, 213 (fam. of Eterocotylea, includes subf. Calcostominæ, Gyrodactylinæ).— Par. & Perugia, 1890, 8.—Pratt, 1900a, 646, 653 (includes: Gyrodactylus, Dactylogyrus, Tetraonchus, Diplectanum, Calcoost., Amphibdella, Dactylodiscus, Fridericianella, Anoplodiscus).—Stoss., 1898, 16–17.—Tasch., 1879, 69; 1879, 235, 237, 238, 260.—See also foregoing entry.

GYRODACTYLIDEA "Ben."—Olss., 1893, 6.

GYRODACTYLIDES Tasch., 1879, 235.—Mont., 1888, 86.

GYRODACTYLINÆ Mont.,1892, Oct. 7, 213 (subf. of Gyrodactylidæ); 1903, 336 (f. Gyrodactylidæ); 1905, 65, 80.—Braun, 1893a, 890.—Gamb., 1896, 73.—

St.-Remy, 1898, 523, 564.

GYRODACTYLUS Nord., 1832a, 105–106 [type by elimination elegans]; 1840, 603.— DACTYLUS Nord., 1832a, 105–106 [type by elimination elegans]; 1840, 603.—Ben., 1858a, 1861a, 11, 63–66, 171, 177, 297.—Ben. & Hesse, 1864, 121.—Biehringer, 1884, 19, 23.—Braun, 1883a, 57; 1889i, 440; 1889k, 622; 1890a, 412, 416, 436, 438, 443, 444, 445, 448, 450, 451, 452, 468, 470, 478, 486, 492, 503, 508, 511, 515, 516, 517, 523, 542, 543; 1892a, 813; 1893a, 888, 889, 890; 1893b, 179, 184, 187.—Burm., 1856a, 251.—Carus, 1863, 478.—Crep., 1838, 84; 1839, 301.—Dies., 1850a, 290, 432, 433 (syn. of Dactylogyrus), 650, 651; 1858e, 314, 374–375; 1859c, 423, 439.—Duj., 1845a, 480–481.—Fraip., 1880c, 442.—Gamb., 1896, 55, 61, 63, 73.—Goldb., 1855, 20.—Haswell, 1892b, 150; 1893e, 114.—Hoyle, 1890, 539.—Jackson, 1888, 643, 646, 648, 654.—Kath., 1894a, 125–164; 1894b, 125–164; 1895a, 431; 1899a, 328–329.—Leuck., 1863, 48, 51, 489; 1879, 58, 62.—Looss, 1892a, 72.—Maclaren, 1904, 586, 587, 598, 599, 600.—Metschnikoff. Gytotactylme), 1303, 330 (sum: Gytotactylme).—Other, 1300, 05.—Fiell, 1905, 28–30 (treatment with ammonia).—Pratt, 1900a, 646, 654, 657, fig. 43, 661.—Roth, 1904, 60–61 (treatment).—St. Remy, 1898, 524, 565.—Sieb., 1839, 163; (1849) v. 1 (4), 347–363.—Tasch., 1879, 69; 1879, 233, 258, 260 (syn. Dactylogyrus), 261 (syn. of Dactylogyrus), 263 (of Wedl, syn. of Tetraonchus).—Wagener, 1857, 25, 26, 49, 50, 76.—Wallenstedt, 1847, 7.

anchoratus Duj., 1845a, 480-481, pl. 8, fig. j (t. h. "carpe").—Bradley, 1861a, 257.-Dies., 1850a, 432.—Kath., 1894a, 155, 156.—Tasch., 1879, 261 (syn. of Dactylogyrus auriculatus).—Wagener, 1857, 49.

auricularis Wedl, 1857, 254, 258, 259, 274, 277, pl. 3, figs. 27–31 [uses auricularis and auriculatus], in Cyprinus carpio.—Dies., 1858e, 376 (syn. of Dactylogyrus anchoratus).—Kath., 1894a, 157.—Tasch., 1879, 261 (syn. of Dact. auriculatus).

auriculatus Nord., 1832a, 108–109, pl. 10, figs. 4–9 (t. h. Cyprimus brama); 1840, 546, 603–604.—Ben., 1858a, 1861a, 64, 66–67, pl. 7, figs. 9–11 (in C. br.).—Braun, 1891d, 421.—Crep., 1838, 84, 85, 86, 87; 1839, 301.—Dies., 1850a, 432 (of Duj., 1845a, 480, syn. of G. dujardinianus), 433 (of Nord., type of Dactylogyrus); 1859c, 440 (of Ben., 1858, 66, syn. of Dact. duj.).—Duj., 1845a, 480, pl. 8, fig. H.—Kath., 1894a, 155, 156.—Mont., 1888, 10.—Sieb., 1835, 70.—Tasch., 1879, 260 (of Duj. syn. of Dact. duj.), 261 (of Nord., to Dactylogyrus).—Wagener, 1857, 49, 50.—Wedl, 1857, 258–274, pl. 3, figs. 27–31.

cochlea Wedl, 1857, 258-274, pl. 3, figs. 32-37 (in Esox lucius).—Cobbold, 1862m, 38; 1879b, 466.—Dies., 1858e, 380 (syn. of Tetraonchus monenteron).—

Kath., 1894a, 157.

GYRODACTYLUS-Continued.

crassiusculus Wedl, 1857, 258–274, pl. 4, figs. 38–40 (in Lucioperca sandra).—Cobbold, 1862m, 38; 1879b, 466.—Dies., 1858e, 381 (syn. of Tetraonchus unguiculatus).—Kath., 1894a, 157.—Tasch., 1879, 263 (syn. of Tetr. ung.).

cruciatus Wedl, 1857, 258–274, pl. 4. figs. 46–47 (in Cobitis fossilis).—Dies., 1858e, 381 (to Tetraonchus).—Kath., 1894a, 157.—Tasch., 1879, 264 (to Tetr.).

dujardinianus Dies., 1850a, 432 (auriculatus of Duj., 1845a, 480, pl., fig. H, renamed) (in Cyprinus carpio, Leuciscus rutilus); 1858e, 376 (to Dactylogyrus).—Kath., 1894a, 156.—Kroyer, 1852–53a, 1224 (in Leuc. rut.).

elegans Nord., 1832a, 106–108, pl. 10, fig. 1–3 (t. h. Cyprinus brama); 1840, 603.—
Ben., 1858a, 1861a, 11, 64, 67–68, 223, pl. 7, fig. 12 (in Cyp. br.).—Ben. & Hesse, 1864, 121.—Bradley, 1861b, 209–210 (on stickle-back in Hampstead ponds).—Braun, 1883a, 57, 71: 1890a, 417, 425, 426, 438, 454, 455, 508, 543, 549, 550, 551; 1893b, 179.—Cobbold, 1862m, 35–39: 1879b, 464, 465, 466, fig. 79.—Crep., 1838, 84, 85, 87; 1839a, 301.—Dies., 1850a, 432, 649, 651; 1852, 417, v. 13, 51–54; 1858e, 375 (in Cyp. carpio, Abramis brama, Gasterosteus aculeatus, G. pungitius, Phoxinus lævis, Cobitis barbatula): 1859c, 439–440.—Duj., 1845a, 480.—Gamb., 1896, 61, fig. 29.—Houghton, 1862a, 77 (in Shropshire).—Hoyle, 1890, 539, 540, fig. 3–G.—Jackson, 1888, 650.—Janicki, 1903a, 241–245, figs. 1–4 (egg segmentation): 1903b, 380.—Kath., 1894a, 127, 129, 130, 132, 133, 134, 140, 141, 144, 150, 155, 156, 157, 158, pl. 7, figs. 1, 2, 5, 6: 1899a, 328; 1904a, 519–550, figs. A–K, 1–26 (development); 1904b, 444–445; 1905a (1, 31), 18–19.—Kholodk., 1899a, 148.—Kowal., 1902d, 23 (5) (in Cyp. carpio); 1904, 328–329; 1904, 10, (25) (in Cyp. carpio; Galicia); 1905a, 18–19.—Kroyer, 1838–40a, 592, 593; 1852–53a, 1225, 1226 (in Phoxinus aphya L., Cob. barb. L., Gast. acul., G. pungitius).—Leuck., 1863a, 489.—Levin., 1881a, 79.—Mont., 1888a, 8, 70; 1891, 111; 1892, Oct. 7, 186.—Odhn., 1905, 372.—Olss., 1893, 6.—Pavesi, 1881, 616.—St.-Remy, 1898, 565–566.—Schneidmuehl, 1896, 303.—Sieb., 1839, 164; 1849, 347.—Sramek, 1901, 95, 110, fig. 65 (in Abramis brama).—Tasch., 1879, 260–261 (in Cyp. carpio, Abramis brama, Gast. acul., G. pungitius, Phoxinus lævis, Cob. barb.).—Wagener, 1857, 49, 50, 51–54, 57, 60, 63, 64, 76; 1860, 768–793, pls. 17–18; 1861, v. 1, 196–212 (reported also for Abramis vimba, Cyclopterus lumpus, Esox lucius, Gobius minutus, Leuciscus phoxinus).

falcatus Wedl. 1857, 258–274, pl. 4, figs. 48–50 (in Cyprinus sp.).—Dies., 1858e, 377 (to Dactylogyrus).—Kath., 1894a, 157.—Tasch., 1879, 261 (to Dact.).

gracilis Kath., 1894a, 129, 131, 139, 141, 143, 147, 150, 157, 158-159, pl. 7, figs.
 4, 7, (in Cobitis fossilis, Cyprinus carpio, Gobius fluviatilis, Leuciscus erythrophthalmus, L. rutilus) [=G. elegans of Ben.]—Odhn., 1905, 372.—St.-Remy, 1898, 566.

grænlandicus Levin., 1881a, 78–79. pl. 3, figs. 5–6 (in Cottus scorpius; Egedesminde).—Odhn., 1905, 372.—Pratt, 1900, 372; 1900, 657, 661, fig. 43.

medius Kath., 1894a, 129, 131, 137, 141, 143, 147, 150, 158, pl. 7, figs. 3, 8, pl. 8, figs. 9–16, pl. 9, figs. 17–24 (in Cobitis fossilis, Cyprinus carpio); 1899a, 328.—Odhn., 1905, 372.—St.-Remy, 1898, 566.

mollis Wedl, 1857, 258–274, 272, pl. 4. fig. 51 (in Cyprinus carpio).—Dies., 1858e, 379 (to Dactylogyrus).—Kath., 1894a, 157.—Tasch., 1879, 262 (to Dact.).

species Ben., 1870, 27, pl. 3, fig. 14, see Dactylogyrus benedeni.—St.-Remy, 1898, 566, 567.

species Ben., 1870, 64, pl. 3, fig. 13.

tenuis Wedl, 1857, 258–274, 270, 278, pl. 4, figs. 41–45 (in Perca fluviatilis).—Cobbold, 1862m, 38, 39.—Dies., 1858e, 379 (to Dactylogyrus).—Kath., 1894a, 157.—Tasch., 1879b, 262 (to Dact.).

GYRODAKTILIDÆ Schneidemuehl, 1896, 296, for Gyrodactylidæ.

HEMATOBIUM shortened form of Dist. hæmatobium Dunglison, 1893, 506; also Reichenback's term for a blood corpuscle.—Billings, 1890, 612.—Danilewsky, 1890c, 753 [=Plasmodium].

equi Burke, 1882, 320, fig. 1 (a blood filaria), 322, fig. 2 (egg of fluke).

HEMATOLŒCHUS Looss, 1899b, 600–601, 602, 603 (tod. variegatus Looss, 1899) [not Hæmatolæcha Stål, 1874, hemipteron] (renamed: Pneumonæces Looss, 1902m, 732) αίματολοιχός, blood sucking; 1902m, 732, 839.—Klein, 1905, 64.—Luehe, 1900, 557, 561.—Odlm., 1902, 41.—Ofenheim, 1900, 156, 164, 182.—Pratt, 1902, 888, 900; 1903, 37.—Staff., 1902, 418; 1902, 725; 1902, 895–912; 1905, Apr. 11 (=Pneumonæces).—Stiles, 1901r, 189.—Stiles & Hass., 1901d, 20.

HÆMATOLŒCHUS—Continued.

asper Looss, 1899b, 601, 603-604 (separated from variegatus).—Luehe, 1902, 238.— Staff., 1902, 896.

breviplexus Staff., 1902, 901, 904-905, pl. 33, fig. 2 (in Rana catesbiana, R. virescens; Canada).

longiplexus Staff., 1902, 901-903, pl. 33, fig. 1 (in Rana catesbiana; Canada).

medioplexus Staff., 1902, 901, 908-910, pl. 33, fig. 5 (in Bufo lentiginosus, Rana virescens; Canada).

similigenus Stiles & Hass., 1902d, 20 (H. similis [Distoma simile] renamed).

similiplexus Staff., 1902, 901, 907–908, pl. 33, fig. 4 (in Rana virescens Kalm, Bufo lentiginosus Shaw; Canada).—Seely, 1906, 252.

similis Looss, 1899b, 601, 602 ("Dist. simile Looss, 1899" nec Sons., 1890).— Staff., 1902, 896, 908.—Stiles, 1901, 178.—Stiles & Hass., 1902d, 20 (renamed similigenus).

variegatus (Rud., 1819) Looss, 1899b, 601, 602; 1902m, 429, 732 (type of Pneumonœces).—Luehe, 1900, 556.—Ssinitzin, 1905, 137-140; 1906, 686 (in Colopteryx virgo; Warschau).—Staff., 1902, 896, 906.—Stiles, 1901, 178.—Stoss., 1902, 5.

varioplexus Staff., 1902, 901, 906, pl. 33, fig. 3 (in Rana catesbiana; Canada).

HÆMATOTREPHUS Stoss., 1902, 8, 22–23 (tod. lanceolatus Wedl).

cymbius (Dies., 1850) Stoss., 1902, 27-28 (in Himantopus wilsoni; Brazil).

fasciatus Stoss., 1902, 25, pl. 6, figs. 21, 22 (in Numenius arguatus; locality?).

lanceolatus (Wedl, 1858) Stoss., 1902, 23-24, pl. 5, figs. 17, 18 (in Himantopus candidus; Rome; H. melanopterus).

phaneropsolus Stoss., 1902, 25-26, pl. 6, figs. 23, 24 (in Totanus sp.; Yeddo, Japan). similis Stoss., 1902, 24, pl. 5, figs. 19, 20, pl. 8, fig. 30 (in Himantopus atropterus: Cairo, Alexandria).

tringæ (Brand, 1892) Stoss., 1902, 26–27, pl. 7, fig. 26 (in Tringa variabilis; Tor, Sinai).

HALICOMETRA Pratt 1902a, 888 (subf., Psilostominæ) 896, for Helicometra.

HALIPEGUS Looss, 1899b, 645–646 (m. ovocaudatus); αλίς, enough; πηγός, well fed; 1902m, 839.—Odhn., 1905, 364.—Ofenheim, 1900, 183.—Luche, 1901, 485.—Pratt, 1902a, 889 (related to Syncoeliine), 905.

dubius Klein, 1905, 68 (in Coluber olivaceus); 1905, 10.

longispina Klein, 1905, 65-68, pl. 5, fig. 3 (in Rana hexadactyla); 1905, 7-10.

occidualis Staff., 1905, Apr. 11, 687-688 (in Rana clamata Daud., R. catesbiana Shaw; Canada) (syns.: Dist. ovocaudatum of Nickerson 1898, 261, and of Staff., 1900, 409).

ovocaudatus (Vulpian, 1858) Looss, 1899b, 645.—Klein, 1905, 7, 8, 9; 1905, 65.—Luehe, 1900, 507; 1900, 558, 559.—Odhn., 1905, 364.—Ssinitzin, 1905, 140–144; 1906, 686 (in Calopteryx virgo; Warschau); larva is Cerc. cystophora.—Staff., 1904, May 3, 484 (compared with Derogenes varicus (in mouth of frogs) sp. Luehe, 1900, 558–559.—See dubius Klein, 1905.

HALZOUN Khouri, 1904, 78, name of disease, see sub Fascioliasis.

HAPALOMETRA Pratt, 1902, 889, misprint for Hapalotrema.

HAPALOTREMA Looss, 1899b, 656–657 (m. constrictum=mistroides); 1902m, 415, 520, 521, 523, 524, 839.—Braun, 1902b, 23.—Luehe, 1901, 488.—Odhn., 1902, 41, 42.—Ofenheim, 1900, 183.—Pratt, 1902a, 889 (Hapalometra, misprint), 907. 1902: Hapalometra Pratt, 1902a, 889, misprint.

constrictum (Leared, 1862 [nec Mehlis, 1846]) Looss, 1899b, 656, 750-752, figs. 72-

75; 1902m, 417, 519-521 (in Thalassochelys corticata in Egypt).

mistroides (Mont., 1876).—Stiles & Hass., 1908, 279.

HAPLOMETRA Looss, 1899b, 599–600 (tod. cylindracea), 601, 602, 603; 1902m, 839.— Luche, 1900, 557, 561.—Odhn., 1902, 41.—Ofenheim, 1900, 182, 183.—Pratt, 1902, 888, 900.—Staff., 1905, Apr., 11, 691.

cylindracea (Zed., 1800) Looss, 1899b, 600.—Darr, 1902, 663, 678.—Kowal., 1902d, (8) 26; 1904, (9) 24, (in Rana temporaria; Dublany).—Luche, 1900, 556, 557.

HAPLOMETRINÆ Pratt, 1902a, 888, 900 (includes: Haplometra, Hæmatolæchus, Ostriolum, Macrodera; related genera: Opisthogonimus, Asymphylodora).

HAPLOPORINE Looss, 1902, 129-143, 14 figs., distome subf.

HAPLOPORUS Looss, 1902h, 134-135 (tod. benedeni).

benedeni (Stoss., 1887) Looss, 1902h, 135, 136-138, figs. 5-6.

lateralis Looss, 1902h, 138-139, figs. 7-8 (in Mugil auratus, M. chelo).

HAPLORCHIDINE Pratt, 1902, 890 (includes: Haplorchis, Galactosomum, Opisthotrema, Cyclocœlum, Notocotylus, Ogmogaster, Stictodora, Mesometra, Monost.).

HAPLORCHIINÆ Looss, 1899b, 671.

HAPLORCHIS Looss, 1899b, 670–671 (tod. pumilio); ἀπλοῦς, single; 1902m, 442, 512.—MacCallum, 1902, 636.—Pratt, 1902, 890, 910.

cahirinus (Looss, 1896) Looss, 1899b, 671, 752-754, fig. 89.

pumilio (Looss, 1896) Looss, 1899b, 671.

HAPLOSPLANCHNUS Looss, 1902i, July 26, 119-122 (m. pachysomus); 1902k; 1903n, 899; 1905h.—Odhn., 1905, 293.

pachysomus (Evsenhardt, 1829) Looss, 1902i, 129.

HAPLOSTOMUM Burm., 1856a, 250 ("Bei den Holostomiden (Acolytea Dies.) ist ausser der Sauggrube, worin sich der Mund befindet, keine zweite Sauggrube am Körper vorhanden.—Diplostomum und Haplostomum mit den davon abgezweigten Formen haben weit von einander getrennte Geschlechtsöffnungen; bei Holostomum stehen sie dicht neben einander").

HARMOSTOMINÆ Looss, 1900, 605.—Pratt, 1902, 889, 997 (includes: Harmost., Ityogonimus, Glaphyrostomum, Scaphiost.).

HARMOSTOMUM Braun, 1899g, 492 (tod. leptostomum; also places here D. spinulosum Hofm., D. opisthotrias Lutz); 1900h, 5, 11, 12, 13; 1901b, 34; 1901c, 338, 342 (syn. Heterolope); 1901, 562, 564, 567; Braun, 1901, 897; 1902b, 114, 115, 116, 120, 122, 125, 135 (syn. Heterolope).—Cohn, 1902, 880.—Looss, 1900, 605 (Marmost., misprint); 1901, 199; 1902m, 813, 839.—Luehe, 1900, 557; 1901, 488.—Odhn., 1902, 42.—Pratt, 1902, 889, 907.—Stiles, 1901, 183, 185.

1899: Heterolope Looss, 1899b, 551, 651-652, 653, 655 (tod. leptostoma) [not Heterolope Franzenau, 1884, protozoon].

1900: Marmostomum Looss, 1900, 605, misprint.

æquans (Looss, 1899) Braun, 1900h, 12; 1901e, 338.

caudale (Rud., 1809 Braun, 1901f, 562; 1902b, 123, 124 (syn. Dist. carvocatactis Zed.) (in Nucirraga caryocatactes).

centrodes Braun, 1901g, 941 (in Tinamus variegatus Lath.; Brazil); 1902b, 120, 121, 122, figs. 73, 74.

fuscatum (Rud., 1819) Braun, 1902b, 114, 115 (in Coturnix communis). 116, 118, 129, fig. 71.

leptostomum (Olss., 1876) [Braun, 1899g, 492]; 1906, 133, fig. 68 (in Helix hortensis). marsupium Braun, 1901g, 941 (in Perdix rufina Spix; Brazil); 1902b, 118, 121, fig. 72.

mesostomum (Rud., 1803) Braun, 1902b, 128 (syn. D. caudale Rud.).

mordens Braun, 1901g, 941 (in Rallus sp.; Brazil); 1902b, 122, 132, fig. 75.

opisthotrias (Lutz, 1895) Braun [1899g, 492] 1901e, 338-339.

[spinulosum (Hofman, 1899) Braun, 1899g, 492.]

HECTOCOTYLUS Cuv., 1829b, 147 (type octopodis).—This supposed worm is not a trematode, but a male mollusk. The name is written in various ways: Hectocotyle, etc.

HELICOMETRA Odhn., 1902, 160–161 (tod. pulchella); 1905, 327, 328.—Stoss., 1903, v. 7 (3), 373–376, 1 fig.; 1904, 199; 1905, Jan. 31, 23; 1905, Jan. 10, 789.

1902: Halicometra Pratt, 1902a, 888, 896, misprint.

fasciata (Rud., 1819) Odhn., 1902, 161, 162.—Stoss., 1903, 373, 375; 1904, 13.

flava Stoss., 1903, 373-376, 1 fig. (in Centropristis hepatus; Triest); 1904, 13.

gobii (Stoss., 1883) Stoss., 1904, 12-13 (in Gobius jozo; Triest).

mutabilis (Stoss., 1902) Stoss., 1903, 375, 376; 1904, 13.—Engler, 1904, 186.—Hellack, 1902a, 868.

pulchella (Rud., 1819) Odhn., 1902, 161–162, fig. 3 (syn. D. labri Stoss.).—Stoss., 1903, 373, 375; 1904, 13.

sinuata (Rud., 1819) Odhn., 1902, 162.—Stoss., 1903, 373, 375; 1904, 13.

HEMISTOMA Cobbold, 1876, 853, for Hemistomum.—Fischder., 1903h, 488.

HEMISTOME E Brand., 1890a, 585.

HEMISTOMIDÆ Brand., 1888a, 58.—Braun, 1893a, 887.—Heider, 1900, 19-22, 4 figs. (Braunina n. g.).

HEMISTOMINÆ Braun, 1893a, 890, 895, 902.—Mont., 1892, Oct. 7, 214 (subf. of Holostomidæ).—Mueh., 1898, 18.—Pratt, 1902, 890 (Hemist.).—Wolf, 1903, 621.

HEMISTOMUM Dies., 1850a, 287, 307-312 (type by inclusion alatum=Alaria vulpis; also type by first species rule) (syns.: Plan. Gozze, Festuc. et Alaria Schrank, Fasc. Gmelin, Strigea Abildg., Dist. et Amphist. Rud., Holost. Nitzsch), 397 [not Hemistomia Crosse, 1872, mollusk]; 1855, 60–61; 1858e, 312, 318–319.— Brand., 1888a, 8, 9, 12, 13, 14, 50, 59; 1892, 505.—Braun, 1892a, 599, 600; 1893a, 872, 879, 887, 890, 894, 895, 900, 902, 917; 1894, 166; 1895b, 132, 136.— 1893a, 812, 879, 887, 890, 894, 893, 900, 902, 917; 1894, 166; 1893b, 152, 156.—Carus, 1863, 479.—Cohn, 1904, 235.—Fischder., 1903h, 488.—Gamb., 1896, 73.—Goldb., 1855, 17.—Johntson, 1904a (in Australian birds).—Hoyle, 1890, 539 (3 sp., 1 in wild cat, 2 in birds).—Looss, 1896b, 13 (sp. of Sons., syn. of Gastrodiscus ægyptiacus).—Mont., 1888, 71, 84, 91; 1891, 105, 109; 1892, Oct. 7, 214 (gen. of Hemistominæ).—Moul., 1856a, 12, 15.—Pratt, 1902, 890, 908.—Rail., 1893a, 381; 1896, 160 (—Conchosomum).—Schneidemuehl, 1896, 295, 303.—Villot, 1878, 19; 1898, 538.—Wolf, 1903, 607.

1876: Hemistoma Cobbold, 1876, 853.

xgyptiaca (Cobbold, 1876) Cobbold, 1876, Dec., 853-854.

alatum (Gœze, 1782) Dies., 1850a, 307–308 (includes Plan. alata Gœze, Festuc. alata Schrank, Fasc. alata Rud., Dist. alatum Zed., Holost. alatum Nitzsch, Anacker, 1892c, 94.—Baillet, 1866b, 106 (to Holost.).—Brand., 1888a, 16, 60 (syns. Alaria vulpis Schrank, Dist. vulpina Abildg.).—
Anacker, 1892c, 94.—Baillet, 1866b, 106 (to Holost.).—Brand., 1888a, 16, 60 (syns. Alaria vulpis Scrhank, Dist. alatum Rud., D. vulpina Abildg., Fasc. alata Rud., Festucaria alata Schrank, Fasc. vulpis Schrank, Holost. alatum Nitzsch, Plan. alata Gœze) (in Canis familiaris, C. vulpis, Thoas cancrivorus); 1890a, 568, 569, 587, pl. 40, figs. 1–5.—Braun, 1883b, 25; 1892a, 569, 599; 1893a, 879, 902; 1896b, 3; 1896d, 583; 1897c, —; 1901e, 336.—Cobbold, 1879b, 432.—Dav., 1877a, 233.—Mol., 1858, 127; 1861, 193.—Mueh., 1898, 18.— Par., 1894, 202, 620, 809, 1050.—Rail., 1893a, 382; 1896, 160 (to Conchosomum).—Ratz, 1898, 397.—Schneidemuehl, 1896, 303.—Sons., 1889, 192 (Emistomum).—Stoss., 1891, 111 (in Vulpes vulgaris).—Ward, 1895, 341 (in Canis familiaris).—Zuern, 1882, 221.—Also reported for Canis azaræ, Canis lagopus, C. lupus, Megalotis cerdo.

attenuatum Linst., 1906, 11-12, pl. 1, fig. 13 (in Buteo vulgaris).

auritum (Duj., 1845) Dies., 1850a, 311–312 (in Strix flammea, Apr., Rhedoni).—
 Brand., 1888a, 62; 1890a, 589 (in St. flam.).—Braun, 1893a, 902.

clathratum Dies., 1850a, 308 (t. h. Lutra brasiliensis; Matogrosso, Brazil); 1855, 61, pl. 1, figs. 13–15; 1858e, 318, 319.—Brand., 1888a, 60–61; 1890a, 587–588, pl. 40, figs. 6–13 (in L. bras.).—Braun, 1892a, 569, 582, 599, 699; 1893a, 902.—Cobbold, 1879b, 298.—Wolf, 1903, 605, fig. 2.

commutatum Dies., 1850a, 311 (includes "Amphist. pileatum" of Bremser, pl. 8, figs. 28, 29, not of Rud., 1819) (t. h. Sterna caspica; June, M. C. V.).—Brand.,

1888a, 62; 1890a, 590 (in S. cas.).—Braun, 1893a, 902.

cordatum Dies., 1850a, 308–309 (t. h. Felis catus ferus; Nov., M. C. V.); 1855, 61, pl. 1, figs. 16–18; 1858e, 319.—Brand., 1888a, 24, 25, 61–62; 1890a, 552, 554, 589, pl. 40, figs. 18, 20 (in Felis catus); 1892, 505.—Braun, 1892a, 569, 579; 1893a, 902.—Cobbold, 1879b, 307.—Heider, 1900, 19, 21.—Schneidemuehl, 1896, 303.—Wolf, 1903, 605, 606, 609, fig. 3.

denticulatum (Rud., 1819) Dies., 1850a, 311 (in Alcedo ispida; Rhedoni).—Brand., 1888a, 62 (in A. isp.; Wien. Mus.); 1890a, 589.—Braun, 1893a, 902.—Villot, 1898, 538, 539, 540, 541, 542 (adult of Diplost. volvens; larva in Phoxinus

lævis).—Wolffhuegel, 1900, 9, 18.

ellipticum Brand., 1888a, 59-60, 67 (in Piaya cayana; by Natterer); 1890a, 586,

595.—Braun, 1893a, 902 (Brazil).

excavatum (Rud., 1803) Dies., 1850a, 309-310 (in Ciconia alba, July, Gryphiæ; C. nigra).—Braun, 1893a, 902; 1894, 167.—Brand., 1888a, 62; 1890a, 590 (in Ciconia alba, C. nigra).—Giebel, 1857, 265.—Mueh., 1898, 16, 18.—Also reported for Nycticorax griseus.

grande (Dies., 1850) Brand., 1890a, 576 (syn. of H. macropterum).

intermedium Johnston, 1904, 109-110, pl. 5, figs. 7-10 (in Cygnus atratus Lath.).

HEMISTOMUM-Continued.

kordatum, see cordatum.

- macropterum Wien. MS. in Brand., 1888a, 55 (syn. Diplost. grande Dies.); 1890a, 576, 581.
- pedatum Dies., 1850a, 309 (t. h. Didelphis myosurus and D. cancrivorus; Brazil, May, June, Dec.); 1855, 61–62, pl. 1, figs. 19–24; 1858e, 319.—Brand., 1888a, 57, 61 (in Did. cancr., D. my.); 1890a, 584, 588, pl. 40, fig. 14.—Braun, 1892a, 581; 1893a, 880, 902.
- pileatum Brand., 1888a, 59, 62, 64 (in Sterna caspica, Larus glaucus, Colymbus arcticus, Mergus merganser) (syn. Holost. erraticum of Linst., 1877c, 188, pl. 13, figs. 18-19); 1890a, 586, 589, 590, 591, pl. 40, fig. 21; 1892, 510.—Braun, 1892a, 582; 1893a, 902.—Kowal., 1904, (8), 23 (in Mergus merganser).—Linst., 1906, 12.—Mueh., 1898, 18.—Stoss., 1895, 37; 1898, 20.—Wolffhuegel, 1900, 9, 15, 54, 61, 62.—Reported also for Buteo vulgaris, Ciconia alba, Colymbus glacialis, C. septentrionalis, Larus marinus, L. ridibundus, Nyctea nivea, Podiceps cristatus.
- podomorphum (Nitzsch, 1819) Dies., 1850a, 311 (in Falco haliaetos; Halle).— Brand., 1888a, 62; 1890a, 589 (in F. hal.).—Braun, 1893a, 902.—Also reported for Circus cineraceus.
- spathaceum (Rud., 1819) Dies., 1850a, 310 (syns. Amphist. lari glauci Rud., Holost. spathaceum Duj., 1845a, 375) (in Larus argentatus, L. argentatoides, L. canus, L. marinus, L. tridactylus; London).—Brand., 1888a, 62 (same hosts); 1890a, 589.—Braun, 1891d, 424 (in Lestris buffonis); 1894k, 681; 1894, 166, 167.—Kowal., 1896d, 252 (2) (in Dominicanus marinus Bruch., Lwow).—Lænnb., 1891, 76.—Mueh., 1898, 16, 18–19.—Olss., 1876, 29 (to Diplost.).—Stoss., 1898, 20.—Villot, 1898, 542 (adult of Diplost. volvens).
- spathula (Crep., 1825) Dies., 1850a, 309 [also spatula, spatulum] (includes: Strigea falconis palumbarii Viborg, Amphist. falconis palumbarii Rud., A. striatum Rud., A. macrocephalum (Falconis milvi) Rud., A. macrocephalum Bremser non Rud.; Holost. spathula Crep.); 1858e, 319 (in Falco nisus).—Brand., 1888a, 54, 59, 62, 68 (to Dist.); 1890a, 574, 580, 585–586, 589, 596, pl. 40, figs. 15–17 (in Buteo vulgaris, Syrnium aluco).—Braun, 1891d, 424; 1893a, 879, 902; 1893b, 185 (in Circus æruginosus).—Cobbold, 1858b, 164 (in Strix otus, Falco milvus); 1879b, 447.—Kowal., 1896d, (2) 252 (in Buteo vulgaris Bechst.; Dublany).—Linst., 1903, 279 (spatula); 1906, 12 (in Buteo vulg.).—Mol., 1858, 127; 1861, 193–194, pl. 1, figs. 3–5.—Mueh., 1898, 19.—Par., 620, 1050.—Stoss., 1890, 50; 1892, 66; 1895, 36–37; 1896, 126; 1898, 20.—Wedl., 1857, 257–258, pl. 1, figs. 23–26.—Wolffhuegel, 1900, 9, 11, 12, 14, 15, 16, 17, 44.—Reported also for Accipiter nisus, Aegolius brachyotus, Aquila chrysaëtos, A. nævia, Archibuteo vulgaris, Ascolopax gallinago, Astur nisus, A. palumbarius, Botaurus stellaris, Brachyotus palustris, Bubo maximus, Buteo lagopus, Circaetus gallicus, Circus cyaneus, C. rufus, Falco albicilla, Milvus ater, Otus vulgaris, Picus sp.
 - species Sons., 1876.—Ward, 1895, 338 (syn. of Gastrodiscus ægyptiacus), in Equus caballus.
- triangulare Johnston, 1904, 108-109, pl. 5, figs. 1-6 (in Dacelo gigas Bodd.).
- trilobum (Rud., 1819) Dies., 1850a, 310; 1858e, 319 (in Carbo comoranus).—Brand.,
 1888a, 59; 1890a, 586 (in Pelecanus crispus).—Braun, 1893a, 903.—Stoss., 1897,
 9 (in Botaurus stellaris).—Wedl, 1857, 255–257, pl. 2, figs. 20–22.
- HEMIURIDÆ Luehe, 1901n, 394–403, 473–488; 1901o, 638–640.—Lander, 1904a, 3.— Looss, 1902m, 839, 848 (includes: Hemiurus, Eurycœlum, Accacœlium, Derogenes, Pronopyge, Liocerca (=Liopyge), Halipegus).
- HEMIURINÆ Looss, 1899b, 640; 1901, 438.—Luehe, 1901n, 481.—Nicoll, 1907, 84.—Odhn., 1905, 355–356, 357, 360, 364, 366.—Pratt, 1902, 889, 905 (includes: Hemiurus, Lecithocladium, Pronopyge, Lecithochirium, Lecithaster, Liopyge, Derogenes).
- HEMIURUS Rud., 1809a, 38 [type appendiculatus by Stiles & Hass, 1898a, 90] (nec Hemiurus Gervais, 1855, mammal; nec Hemiura Ridgway, 1888, Jan. 6, 511, bird).—Darr, 1902, 698.—Linst., 1904, 252.—Looss, 1899b, 527, 534, 551, 571, 582, 583, 638, 639, 640–641, 642; 1901, 194, 201, 207; 1901, 438; 1902m, 756, 781, 805, 830, 831, 839.—Luehe, 1900, 509; 1901, 394, 395, 396–401, 402, 474, 479, 480, 482, 484.—Nicoll, 1907, 84.—Odhn., 1905, 354, 355, 356, 359.—Pratt, 1902, 889, 906.—Stiles, 1901, 177, 178, 185, 193.—Stiles & Hass., 1898a, 90–91, 96 (includes: Dist. (Apoblema), Duj., Apoblema Duj., Eurycœlum Brock) (type Fasc, appendiculata Rud.).

HEMIURUS—Continued.

1845: Dist. (Apoblema) Duj., 1845a, 383, 389, 420 [tld. appendiculatum].

appendiculatus (Rud., 1802) Looss, 1899b, 641, 671; 1902m, 831.—Lander, 1904a, 4.—Luche, 1901, 395, 396, 398, 400, 475.—Nicoll, 1907, 69, 70, 71, 72, 84, 86, 87, 88 (in Anguilla vulgaris, Centronotus gunnellus, Cottus bubalis, Hippoglossus vulgaris, Pleuronectes limanda, Pl. platessa).—Odhn., 1905, 349, 350, 351, 352 (in Alosa finta).—Staff., 1904, May 3, 484 (in esoph., stomach, Canada, of Salmo salar L., Osmerus mordax Mit., Clupea harengus L., Gadus callarias L., Pollachius virens L., Ammodytes tobianus L., Anguilla anguilla L., Acanthocottus scorpius L., Hippoglossus hippoglossus L., Platysomatichthys hippoglossoides Walb.): 1905. Apr. 11, 682, abundant in conepods (Acartia) hippoglossoides Walb.); 1905, Apr. 11, 682, abundant in copepods (Acartia).— Stiles, 1901, 177.

bothryophorus (Olss., 1868) Looss, 1899b, 641, 728-729 (in Alosa finta), 741.

communis Odhn., 1905, 348, 350, 351 (in numerous Scandinavian marine fishes), 352, 353 (syn. Dist. appendiculatum Rud. of Olss., 1868).—Nicoll, 1907, 71,

86-88 (in Ammodytes tobianus, Gadus æglefinus).

crenatus (Rud., 1802) Luehe, 1901n, 395, 397, 398, 399-401 ("nec Looss, 1899") [the synonymy as given by Luehe is not altogether clear].—Lander, 1904a, 1–28, figs. 1–42 (anat.; syns. Fasc. cren. Rud., Dist. cren. Rud., Dist. ocreatum Olss., Apoblema ocreatum Juel, Hemiurus ocreatus Looss); 1904b-c; 1905a.—Nicolf, 1907, 84.—Odhn., 1905, 350, 352, 353, 356 (type of Brachyphallus).—Tennent, 1906, 666.

crenatus (Mol., 1859) Looss, 1899b, 641.—Luehe, 1901n, 399 (not of Rud.).

digitatus Looss, 1899b, 641, 729-731, fig. 48 (in Sphyræna vulgaris; Sawakin, Egypt, Jan.).—Darr, 1902, 659.

excisus (Rud., 1819) Looss, 1899b, 641.

grandiporus (Rud., 1819) Looss, 1899b, 641, 730.—Luehe, 1901, 401.—Nicoll, 1907, 84.

lævis (Lint., 1898) Looss, 1899b, 641.—Luehe, 1901, 401.—Nicoll, 1907, 84.

levinseni Odhn., 1905, 348–351, pl. 4, fig. 2 (in Gadus saida of East Greenland; also in G. morrhua s. ovak, G. melanostomus, Cottus scorpius, Phycis blennoides; West Greenland, North Denmark) (syns. Dist. appendiculatum Rud. of Olss., and Rud. of Levin.).

lühei Odhn., 1905, 351, 352 (syns. Dist. appendiculatum Rud. of Olss., 1868; H. stossichi Luehe, 1901 [nec Mont., 1891]) (in Clupea harengus, C. sprattus, and ? C. pilchardus).—Nicoll, 1907, 72, 85–86, 87 (syn. of H. stossichii) (in Clupea harengus).

microporus (Mont., 1889) Looss, 1899b, 641.

mollissimus (Levin., 1881) Looss, 1899b, 641.

monticellii (Lint., 1898) Looss, 1899b, 641.

ocreatus (Mol. of Olss., 1868) Looss, 1899b, 641 (nec Rud.).—Lander, 1904a, 1, syn. of H. crenatus (Rud.).—Luehe, 1901n, 399.

rufoviridis (Rud., 1819) Looss, 1899b, 641, 730.

sluiteri (Brock, 1886) Looss, 1899b, 641.

species Braun, 1902b, 125.

stossichi (Mont., 1891) Looss, 1899b, 641.—Luche, 1901n, 398-399, 400 (stossichii).— Nicoll, 1907, 84, 85, 88 (syn. of H. lühei).

stossichi Luehe, 1901n (nec Mont., 1891) of Odhn., 1905, 351, 352 (renamed H. lühei Odlın.) (in Clupea pilchardus).

stossichii Luehe, 1901n, 400, for stossichi.

tornatus (Rud., 1819) Looss, 1899b, 641.

varicus (Mueller, 1784) Looss, 1899b, 641.

(HEMIURUS) Rud., see Hemiurus.

scabrum (Mueller, 1788) Looss, 1899b, 582.

HEPASTOMUM Brand., 1888a, 15, for Heptostomum.

HEPHOSTOMUM Burm., 1856a, 251 ("In anderen Fällen fehlen die Sauggruben am Munde, so bei Diclidophora, Hexacotyle, Hephostomum").

HEPTASTOMUM (for Heptostomum Schomburgk, 1844, 136).—Brand., 1888a, 15 (Hepastomum), 52; 1890a, 578.—Braun, 1893a, 884.—Dies., 1850a, 289, 418 (syn. Dist. Henle); 1858e, 314, 369–370 (m. hirudinum).—Fil., 1854a, 23.—Goldb., 1855, 19.—Mont., 1888a, 84, 92.—Tasch., 1879, 234. HEPTASTOMUM-Continued.

hirudinum Schomburgk, 1844, 136 (in Nephilis vulgaris, Clepsine complanatum) (includes Dist. hirudinis .—Brand., 1888a. 13.—Braun, 1892a, 795.—Crep., 1846, 159.—Dies., 1850a. 418-419 (syn. Dist. hirudinis Henle, D. tarda: 1858e. 370. pl. 2 (in Nephilis vulgaris. Clepsine complanata).—Fil., 1854a, v. 15, 23.

HEPTOSTOMUM Schomburg, 1844, 136, see also Heptastomum.

HERONIMUS MacCallum, 1902, 25 Oct., 632-636 (m. chelydræ) (monostome).

chelydræ MacCallum, 1902, Oct. 25, 632–636, figs. 1–2 (in Chelydra serpentina; Dunnville, Ontario): 1902, Dec. 30, 843–844: 1905, Aug., 62.

HETERACANTHUS Dies., 1836, 307-310 (Axine 1794, renamed, hence type pedatus =bellones renamed) [not Heteracanthus Newberry, 1889, fish]; 1850a, 425 (syn. of Axine).—Braun, 1890a, 518.—Crep., 1838, 83; 1839, 291,—Nord., 1840, 598.—Tasch., 1879, 255 (syn. of Axine Abildg., 1794).

pedatus Dies., 1836c, 310–313, pl. 17, figs. 1–2 (Axine bellones Abildg. on Esox belone renamed: 1850a, 425 (syn. of Axine bellones).—Ben., 1858a, 1861a, 53 (syn. of Ax. bello. —Crep., 1838, 83 (syn. of Ax. platyura).—Goto, 1894, 196 (Ax. belones partim).—Kroyer, 1846–53a, 273 (in Belone rostrata Γab.).—Nord., 1840, 598 (syn. Ax. bell.) (in Esox belone).—Sieb., 1839, 163.—Stoss., 1898, 14.—Tasch., 1879, 256 (syn. of Ax. belones).

[politus fossil fish .]

sagittatus Dies., 1836, 313. pl. 17. figs. 10–12 (on Esox belone: [Europe] : 1850a, 425 (syn. of Axine bellones).—Ben., 1858a, 1861a, 53 (syn. of Ax. bell.).—Crep., 1838, 83 (syn. of Ax. platyura).—Goto, 1894, 196 (Ax. belones Abildg. partim).—Kroyer, 1846–53a, 273 (in Belone rostrata Fab.,—Nord., 1840, 598–599 (in Esox belone).—Sieb., 1839, 163.—Tasch., 1879, 256 (syn. of Ax. belones Abildg.)

HETEROBOTHRIUM Cerr., 1895m. 141, 142, 145-146 (m. tetrodonis): 1896, 548, 551-552.—Mont., 1903, 336 (subf. Diclidophorine).—St.-Remy, 1898, 552.—Zool, Anz., 1895, 327.

tetrodonis Goto, 1894 | Cerf., 1895m, 141, 142, 146 | in Tetrodon sp.: Japan : 1896, 548, 552.—Zool, Anz., 1895, 327.

tetrodontis St.-Remy, 1898, 554 = Diclidophora tetrodonis, for tetrodonis.

HETEROCOTYLE Scott. 1904, 279 m. pastinacæ).

pastinaca: Scott. 1904. 279. pl. 17, fig. 14 in Trygon pastinaca: Dornoch Firth. Oct. .

HETERECOTYLEA Mont., 1905c, 68, apparently for Heterocotylea,—Massa, 1906, 43.

HETEROCOTYLEA Mont., 1892; 1899, 81, 88, 107 [Eterocotylea]; 1903, 234; 1903c, 334-336; 1904 (II.10), 65-80, figs. 1-5 (remarks with reference to some species of); 1905c, 65-80, 5 figs.—Braun, 1893a, 889, 917; 1893b, 188; 1895b, 136.—Gamb., 1896a, 73.—Maclaren, 1904, 574, 579, 583, 586, 590, 591, 594, 596, 599.—Much., 1898, 17.—Pratt, 1900, 645, 646 (includes: Temnocephalidæ, Tristomidæ, Monocotylidæ, Polystomidæ, Gyrodactylidæ), 647-654, 655-657, pls. 1-50, 658-661 (key to American species), 661-662; 1902, 890 (key).—Ward, 1903, 864.

HETEROKOTYLEA Schneidemuehl. 1896, 295, for Heterocotylea.

HETEROLOPE Looss. 1899b. 551. 651–652. 653. 655 (not Heterolopa Franzenau, 1884. Protozoon) orig. species: leptostoma [tod.]. opisthotrias. æquans, caudatal ἔτερος=anders: ἡ λώπη=wall, because of the structure of the skin: 1900. 605.—Braun. 1900h. 6. 11. 13: 1901e. 338 (=Harmost.): 1902b, 114 (syn. of Harmost. Braun. 1899. 492).—Cohn, 1902, 880 (=Harmost.).—Luche. 1900. 557.—Ofenheim. 1900. 183.

æquans Looss. 1899b. 652, 746-748. fig. 70 in Gerbillus **ægyptius.—Braun, 1900, 12.

caudata (Linst., 1873 | Looss, 1899b, 652 (nec Mueller, Bosc, Polonio).

leptostoma (Olss., 1876) Looss, 1899b, 652, 746, 747, 748.

opisthotrias Lutz. 1895 Looss. 1899b. 652

HETEROLOPINE Looss, 1899b, 653, 655; 1900, 605.—Braun, 1900h, 13.

HETEROPHYES Cobbold, 1866a. 6 m. agyptiaca=heterophyes [this genus was probably published in some earlier paper].—Loss, 1902m, 786, 805, 808, 824; 1902n, 886-891 (revision of : 1903o, 899-900; 1905k, 56; 1907, Mar. 5, 488, 489.—Ward, 1903, 870.

1899: Cotylogonimus Luche 1899, 538-539 tod. heterophyes.

1899: Coenogonimus Looss, 1899b. 585 (tod. heterophyes).

agyptiaca Cobbold, 1866a. 6 Dist. heterophyes renamed .

xquatis Looss. 1902n. 888 in dogs and cats; Egypt ...

dispar Looss. 1902n. SSS-889 in dogs and cats .

fraternus (Looss, 1894) Looss, 1902m, 785, 808, 809, 838, 854; 1902n, 887; 1907, Mar. 5, 488 (in Pelecanus onocrotalus Cairo, Egypt).

heterophycs Sieb., 1852 Stiles & Hass., 1900a, 563.—Loss, 1902m, 782, 785, 786, 808, 809, 838, 854;
1902n, 889;
1905, 111 in man, cats, and dogs;
Egypt, fig. 18.—Ward, 1903, 704;
1903, 864 in Homo, 870 includes: Dist, het. Sieb., 1852;
Mesogonimus het. Rail., 1890;
Coenogonimus het. Loss, 1900;
Cotylogonimus het. Luche, 1900, 871.

inops Loose, 1902n, 887-888 in Pelecanus onocrotalus: Milvus agyptius s. M. parasiticus: Egypt).

pallidus Looss, 1902n, 889-891 (in Milvus ægyptius s. parasiti us: Egypt . persicus Braun, 1901 Looss, 1902m, 782, 785.

HETEROSTOMA Fil., 1837a, 338-340 m. echinatum (ετερος, diverse στόμα, bocca: 1857c, 9 (Heterostomum).—Burm., 1856a, 250 (Heterostomum).—Dies., 1850a, 287, 301-303 (syn., Dist. Baer (Heterostomum): 1855a, 396 (mentions only two species; echinatum, ovatum).—Erc., 1881e, 33; 1882a, 269.—Goldb., 1855, 16.—Moul., 1856a, 12, 16, 121.

1850: Heterostomum Dies., 1850a, 287, 301-303, for Heterostoma.

cchinatum Fil., 1837a, 338-339, 340, figs. 16-18 t. h. Paludina impura: Italy an Agamodist. —Dies., 1850a, 302 Heterostomum: 1855a, 396: 1858d, 280 to Cercariæum.—Leidy, 1858a, 110: 1877, 202.—Moul., 1850a, 216 "pourait bien être un individu libre provenant de la Cerc, paludidæ impuræ Fil.".

helicis aspera Dies., 1850a, 302-303, based on Duj., 1845a, 472; 1855a, 398 (to Cercariæum helicis aspersæ).

helicis aspersa Dies.. 1855a. 398. for helicis asperæ to Cercariæum.

helicis pomatia: Meckel, 1846 Dies., 1850a. 303; 1855a. 398 w Cercaria um .

limacis Dies., 1850a, 302 based on Duj., 1845a, 472-473; 1855a, 397 to Carcariæum.
 limaci Dies., 1850a, 302 based on Duj., 1845a, 473; 1855a, 399 syn. of Cercariæum lymnæi palustris.

oratum Dies., 1850a, 302 (Dist. luteum Baer, 1827, pl. 29, figs. 20-22, renamed (in Paludina vivipara: Regiomontii): 1855a, 396 (in Pal. vivip.): 1858d, 279 (to Cercariæum .—Mont., 1893, 190.—Moul., 1856a, 215 syn. of Dist. luteum Baer).

HEXABOTHRIUM Nord., 1840, 600 (m. appendiculatum t.—Ceri., 1899a, 351, 359, 371.—Dies., 1850a, 419, renamed Onchocotyle.—Stoss., 1898, 11.—Tasch., 1879, 252 (syn. of Onchocotyle Dies.).

appendiculatum Kuhn, 1829 Nord., 1840, 601 in Squalus catulus.—Dies., 1850a.
419 type of Onchocotyle in Scyllium catulus. Læmargus borealis.—Tasch., 1879, 28 to Onchocotyle.

HEXACOTYLA Blainv., 1828a, 570-571 (tod. thynni).—Dies., 1850a, 410 (tenamed Plagiopeltis).

ocellatum Rud., 1819 Blainv., 1828a, 571.—Baird, 1853a, 41 (= Polyst.).

thynni Delaroche, 1811 Blainv., 1828a, 571.—Dies., 1850a, 417 syn. of Plagiopeltis duplicata, type of Plagiopeltis in Thynnus branchypterus, ad insulas Balearicas .

HEXACOTYLE Blainv., 1828a, 570 (same as Hexacotyla; both are used by Blainv. Hexacotyle has been adopted as the Latin form by a number of authors).—
Braun, 1890a, 413, 516, 517, 518, 523, 534, 537, 540; 1890a, 890.—Burm., 1856a, 251.—Cerf., 1895h, 920; 1896, 515; 1899a, 391.—Dits., 1850a, 409, 416 of Blainv., syn. Hexathyridium Treutler, 421 of Nord., syn. of Diclib othrium Leuck, 1.—Gamb., 1890a, 73.—Goto, 1893a, 798, 799; 1894, 215.—Hoyle, 1890, 539.—Mont., 1888a, 89, 100; 1892, Oct. 7, 213 (subg. of Octocutyle); 1903, 336 (= Plagiopeltis (subf. Plagiopeltinæ, —Nord., 1840, 600.—Pratt, 1900a, 653, 657, fig. 38, on gills of marine fishes.—St.-Remy, 1898, 556.—Tasch., 1879, 69; 1879, 239, 249-250 (syns, Polyst. Roche, Plagiopeltis Dies., 254 (of Nord., 1840, syn. of Diplobothrium Leuck.).

HEXACOTYLE—Continued.

1850: Plagiopeltis Dies., 1850a, 416–417 (duplicata = thynni).

acuta Goto, 1894a, 217–220 (in Thynnus sibi; Hagi and Osatsubé, (Hokkaidō)).—St.-Remy, 1898, 557.

clegans Nord., 1840, 597, 600 (Diklibothrium crassicaudatum renamed) (in Acipenser stellatus).—Dies., 1850a, 421 (syn. of Diclibothrium armatum Leuck.).—Tasch., 1879, 254 (syn. of Diplobothrium armatum Leuck).

grossa Goto, 1894a, 220–222 (in Thynnus sp.; Misaki).—St.-Remy, 1898, 557–558, lapridis Sars., ——.—Nord., 1840, 600 (in Lampris gulfatus).

ocellatum (Rud., 1819) Blainv., 1828a, 571.—Dies., 1850a, 413 (to Polyst.).—Nord., 1840, 600 (in Testudo orbiculata).—Tasch., 1879, 252 (to Polyst.).

pinguicola (Treutler, 1793) Chiaje, 1833, 13 (= Polyst.).

thunninæ (Par. & Perugia, 1889) Goto, 1896, 352: 1899, 273–274, pl. 20, figs. 13–15.— Pratt, 1900, 657, fig. 38.—St.-Remy. 1898, 557.

thynni (Delaroche, 1811) Nord., 1840, 597, 600.—Braun, 1890a, 418, 537, 548.—Dies., 1850a, 417 (duplicata, type of Plagiopeltis).—Goto, 1899, 274.—Lint., 1901, 414 (in Sarda sarda), 446, figs. 296, 297, 298.—Par., 1894, 578; 1899, 3-4; 1902, 3 (in Pelamys sarda, Portoferraĵo: Thynnus thynnus, Elba).—Par. & Perugia, 1890, 7.—Ŝt.-Remy, 1898, 556–557.—Tasch., 1879, 250 (syns.: Polyst. th. Roche, P. duplicatum Rud., Hex. th. Blainv., Plagiopeltis dup. Dies.) (in Thynnus branchypterus, Pelamys sarda; Napoli).

venarum (Treutler, 1793) Blainv., see Dies., 1850a, 410 (to Hexathyridium).

HEXACOTYLIDÆ Mont., 1899. —; 1903, 336 (subf.: Diplobothrine (g. Diplobothrium); Plectanocotylinæ (g. Plectanocotyle, Phyllocotyle)); 1905, 77, 78.

HEXASTOMA Rud., 1809a, 38, 451–457 (subg. of Polyst. containing: integerrimum [type by inclusion], pinguicola, (genus in Cuv., 1836a, 264).—Dies., 1850a, 409 (syn. of Hexathyridium Treutler).—Kuhn, 1829, 358 (for Polyst. Rud.).—Thon, 1830, v. 7, 341.—Wallenstedt, 1847, 7.

integerrimum (Frælich, 1791) Rud., 1809a, 451.

pinguicola (Treutler, 1793) [Cuv., 1836a, 264].—Dies., 1850a, 410 (to Hexathyridium).—Dolley.

venarum (Treutler, 1793) [Cuv., 1836a, 264 (to Hexathyridium)].—Dies., 1850a, 410 (to Hexathyridium).

HEXOSTOMA Rafinesque, 1815, 151, n. n. for Polyst. Roche, hence type thynni.

HEXATHIRIDIUM Blainv., 1821a, 144-145 (see Hexathyridium); 1824a, 516; 1828a, 571-572.

venarum (Treutler, 1793) Blainv., 1828a, 572.—Joy, 1835a, 505.—See Hexathyridium.

HEXATHYRIDIA Rafinesque, 1815, 151 (genus of Fasciolaria with Polyst. Zed. as syn.) for Hexathyridium.

HEXATHYRIDIUM Treutler, 1793, iv. 19–22 (type by first-species rule, pinguicola; this seems to be taken as type by authors; see, for instance, Braun, 1889a, 317.—Blainv., 1821a, 144–145 (Hexathiridium): 1824a, 516 ("Je doute beaucoup que ce genre puisse être regardé comme véritablement intestinal; il est extrêmement rapproché du polystome de Delaroche; aussi pensais-je qu'il a été décrit à l'envers. ce que l'on voit dans ma définition des caractères de l'ordre"): 1828, 571–572.—Braun, 1890a, 518; 1893a, 887, 891, 892, 894.—Burm., 1856a, 251 (Hexatiridium).—Cerf.. 1899a, 351, 371.—Dav.. 1877a, lxxx.—Dies., 1850a, 288, 409–410, 412 (Blainv., syn. of Polyst. Zed.): 1858e, 314, 368.—Encycl. méthodique. Par., v. 2, 455.—Goldb., 1855, 17.—Hoyle, 1890, 539 (probably a syn. for Polyst.).—Jærdens, 1802, 66.—Lamouroux, 1825, 193.—Leuck., 1863, 526; 1867. v. 2 (1), 150; 1868, v. 2 (2), 264.—Looss, 1902m, 756.—Mont., 1888a, 84 (Hexathyridum). 90.—Nord., 1840, 601 (Hexathiridium).—Rud., 1801a, 58; 1809a, 22.—Stess., 1898, 11.—Tasch., 1879, 324, 251 (syn. of Polyst. Zed.).—Thon., 1830, v. 7, 341-342 (Hexathiridium).

1809: Hexastoma Rud., 1809a, 38, 451-457 (type by inclusion pinguicola).

1815: Hexathyridia Rafinesque, 1815, 151.

1821: Hexathiridium Blainv., 1821a, 144-145.

1888: Hexathyridum Mont., 1888a, 84.

1856: Hexatiridium Burm., 1856a, 261.

affine Dies., 1850a, 410 (t. h. Bombinator igneus; Berlin).—R. Bl., 1895, 104.—Schmitz, 1826, 15, figs. 1–13.

HEXATHYRIDIUM—Continued.

appendiculatum (Kuhn) Nord.—Tasch., 1879, 253 to Onchocotyle (Kuhn) [Nord. uses Hexabothrium app., not Hexathyridium app.].

integerrimum (Frælich, 1791) Blainv., 1828, 572.—Baird, 1853a, 41 (= Polyst.

Rud.).—Dies., 1850a, 412 (to Polyst.).—Stoss., 1898, 10.

pinguicola Treutler, 1793, iv, 19–22, pl. 3, figs., 7–11 (t. h. Homo).—Blainv., 1828, 572.—Aitken, 1866, 804, 841; 1872, 146, 207; 1874, 58.—R. Bl., 1888a, 597.—de Bonis, 1882, 180.—Braun, 1883a, 69–70; 1895b, 155.—Brena, —, 100, pl. 1, fig. 28.—Cerf., 1899a, 351.—Cobbold, 1866, 7; 1879b, 36.—Dav., 1877a, lxxxi.—Dies., 1850a, 409–410 (includes: Polyst. ping. Zed., Linguatula ping. Lamarck, Hexast. ping. Cuv.); 1858e, 368 (to Polyst.).—Dunglison, 1893, 529, 821.—Hackley, 1886, 519.—Hoyle, 1890, 538.—Jærdens, 1802, 66, pl. 6, figs. 3–5.—Kholodk., 1898, 26, pl. 10, fig. 24.—Kuech., 1855, 464.—Leuck., 1863a, 526, 585, fig. 195.—Moniez, 1896, 108.—Mosler & Peiper, 1894, 186.—Mont., 1888, 90.—Rud., 1809a, 455 (t. Polyst.).—Verrill, 1870, 171.—Ward, 1895, 328 (in Homo).—Weinland, 1859, 281.

venarum Treutler, 1793, iv, 23–25, pl. 4, figs. 1–3.—Aitken, 1866, 804, 841–842; 1872, 146, 206; 1874, 58.—Biermer, 1863a, 393.—Blainv., 1828a, 572 (Hexathiridium).—R. Bl., 1888a, 596, 597; 1891, 605.—de Bonis, 1882, 181.—Braun, 1883a, 69; 1895b, 139; 1903, 3 ed., 150 (from Vena tibialis antica, probably young F. hepatica. Duval found adult F. h. in Vena portarum and other veins at Rennes (1842), and Vital (1874) describes same kind of case from Constantine); 1906, 155 (in Homo sapiens).—Bremser, 1824, 327–329, pl. 9, fig. u.—Cerf., 1899a, 351.—Cobbold, 1866, 7; 1879b, 36.—Crep., 1839, 290.—Dav., 1877, lxxxi, 329–330, figs. 15, 42.—Dies., 1850a, 410 (includes Polyst. ven. Zed., Polyst. sanguicola delle Chiaje, Linguatula ven. Lamarck, Hexast. ven. Cuvier, Hexacotyle ven. Blainv.); 1858c, 368.—Duval, 1842a, 771, 772.—Duj., 1845a, 321.—Dunglison, 1893, 529, 821.—Hackley, 1886, 519.—Hahn & Lefèvre, 1884a, 540–541.—Hémont, 1827, 16.—L'Herminier, 1826, 13.—Hoyle, 1890, 538.—Joy, 1835a, 505.—Kholodk., 1898, 26, pl. 10, fig. 23.—Kuech., 1855, 464.—Leuck., 1863a, 526, 575, 585, fig. 194.—Moniez, 1896, 108.—Mont., 1888, 90.—Mosler & Peiper, 1894, 186.—Rud., 1809a, 456 (to Polyst., as sp. inq.).—Sieb., 1839, 164.—Stiles, 1898a, 48.—Verrill, 1870, 171.—Ward, 1895, 328 (in Homo); 1903, 866.—Weinland, 1859, 281.

HEXATHYRIDUM Mont., 1888, 84, for Hexathyridium.

HEXATIRIDIUM Burm., 1856a, 251 (see Hexathyridium).

HIRUDELLA Poir., 1885, 5, 7 [not Muenster, 1842, leech], see Hirudinella.

HIRUDINELLA [Garsin, 1730, 58–59, fig., prelinnæan (m. marina); 1735, 261, pl. 4, figs. 1–3].—Blainv., 1824a, 518 ("hirudinelle"; "J'admets ce genre pour les fascioles cylindriques qui ont quelque ressemblance avec les sangsues, et entre autres le fasc. clavata Linn."); 1828, 586 (type clavata) [not Hirudinella Gray, 1850, mollusk; not Hirudinella Muenster of Dies., 1850a, leech].—Baird, 1853a, 59.—Buttel-Reepen, 1902, 168.—Darr, 1902, 644, 648, 658, 662–699.—Dies., 1850a, 292.—Encycl. méthodique, Par., v. 2, 456.—Lamouroux, 1825, 241.—Planque, —, 993, 221.—Poir., 1885, 5, 7 (Hirudella) [not Hirudella Muenster, 1842, renamed Hirudinella by Dies., 1850].—Stiles, 1901, 194 (type Fasc. clavata).

1885: Hirudella Poir., 1885, 5, 7 [not Muenster, 1842], for Hirudinella.

[angusta Muenster, 1842, 98, pl. 1, fig. 5.—Dies., 1850a, 471; 1859, 511.—Pictet,

—, 457.—A leach.

clavata (Menzies, 1791) Blainv., 1828, 586.—Baird, 1853a, 59 (includes H. marina Garsin, Fasc. clav. Menzies, F. scombri pelamidis Tilesius, F. coryphænæ Bosc, Dist. clav. Rud.).—Buttel-Reepen, 1902, 167, 168, 171, 185, 190, 191, 193, 195, 198, 202, 206, 209, 212, 214, 218 (syn. of Dist. ventricosum).—Darr, 1902, 649, 652, 657, 658, 667, 668, 669, 670, 676, 678, 680, 682, 687, 689, 690, 692, 696, 698-699, 1 fig., pl. 34, figs. 16-25, pl. 35, figs. 26-34.

ingens (Moniez, 1886) Darr, 1902, 687, 688.

marina Garcin, 1730a, 387–394, pl. 1, fig. 2 (in Scomber pelamys); 1732a, 43–44,
1 pl., 3 figs.—Baird, 1853a, 59 (=clavata).—Buttel-Reepen, 1900a, 586; 1902,
166, 168, 194, pl. 6, fig. 1.—Cobbold, 1879b, 461.—Darr, 1902, 663, 664.

[tenuis Muenster, 1842, 99.—Dies., 1850a, 471 (a leach); 1859, 511.—Pictet, ——, 457.]

ventricosa (Pallas, 1774) Baird, 1853a, 59–60 (includes Fasc. ventr. Pallas, F. fusca Bosc, F. coryphænæ hippuridis Tilesius, Dist. coryphænæ Rud., D. clavatum Owen).

- HIRUDO Linn.. 1758a, 644, 649 (type by Linnæan rule, medicinalis).—Several species of this genus have been placed in the trematodes: the genus is now confined to the leeches.—Dies., 1850a, 318 (of Braun, syn. of ?Diplodiscus., 425 (of Abildg., syn. of Nitzschia Baer), 426 (of Mueller, syn. of Phylline Oken), 427 (of Kroyer, syn. of Udonella Johnston), 433 (of Braun, syn. of Astacobdella Vallot), 439 (of Linn.. syn. of Ichthiobdella Blainv.), 445 (of Mueller, syn. of Malacobdella Blainv.), 446 (of Mueller, syn. of Clepsine Savigny), 456 (auct.. syn. of Nephelis Savigny), 461 (auct., syn. Aulost. Moquin-Tandon), 465–471 (of Rai et Linn., syns. of Sanguisuga Savigny; Iatrobdella Blainv.), 567 (of La Martinière, syn. of Tetrarhynchus Rud.), 583 (of Linné. syn. of Schistocephalus Crep.).
 - fasciolaris Mueller, 1788b, pl. 54. figs. 1–3 (is included in Fasc. anatis Gmelin, 1790a, 3055.)—Dies., 1850a, 335 (syn. of Dist. ovatum Rud.).
 - grossa Mueller, 1788, 21–22. pl. 21.—Bruguière, 1791a, pl. 52, figs. 6–8.—Dies., 1850a, 445 (to Malacobdella).—Gmelin, 1790a, 3098.
 - hippoglossi Mueller, 1776a, 220 (t. h. Hippoglossus; Denmark); 1788a, 18, pl. 54,
 figs. 1-4.—Baer, 1827b, 676, pl. 32. figs. 5-6.—Dies., 1850a, 426 (to Phylline).—
 Fabricius, 1780a, 322. pl. 1. fig. 8.—Gmelin, 1790a, 3098.—Leuck., 1842a,
 11.—Oken, 1815 (to Phylline).
 - sturionis Abildg., 1794b, 55-56, pl. 6. fig. 1, a-c (in Stören).—Cuvier, ——. (to Trist.).—Dies., 1850a, 426 (syn. of Nitzschia elegans Baer).—Gmelin.
 - tuba Braun, 1805, 49, pl. 5, figs. 5–8 (see Rud., 1809a, 348 = Amphist, subclavatum).—Dies., 1850a, 318 (syn. of Diplodiscus subclavatus Dies.).

HISTRIOBDELLA Ben., 1858i, 2 pls.

- HISTRIONELLA Bory de St. Vincent. 1823b, 356: 1825b, 252-253.—Braun. 1893a, 884.—Burm., 1856a, 250.—Dadai. 1888í, 84, 85; 1888g, 107.—Dies., 1850a, 286, 294 (of Bory syn. of Malleolus Ehrenberg). 299-300 (of Bory & Ehrenberg, includes Cerc. Nitzsch, Brachionus Schrank): 1855a, 378 (eyes), 392, 393 (syn. Brachionus Schrank): 1858d, 267.—Goldb., 1855, 16.—Moul., 1856a, 121, 124.—Nord., 1840, 617, 631.—Pag., 1857, 5, 24.—Wagener, 1857, 24.
 - —— Hemprich & Ehrenberg, 1828a, Entozoa, not paged, n. g. cf Cercozoorum, type evidently ephemera; includes also alata.
 - alata Hemprich & Ehrenberg, 1828a (in water at Berlin, Germany).—Dies., 1850a, 300 (sp. inq.); 1855a, 392-393 (Hab. primitivum ignotum); 1858d, 268 (free; Berlin).
 - annulicauda Bory. 1825b, 253.—Dies., 1850a, 299 (syn. of Histrionella lemna Ehrenb.).
 - bilineata (Haldemann, 1840) Dies., 1850a, 300 (in Limnæus catascopium; Penn.); 1855a, 393 (in Lymnæus cat.); 1858d, 269 (in Lymn, cat.; Penn.).
 - echinocerca (Fil., 1857) Mont.. 1888, 193, 194.—Dadai. 1888f, 85: 1888g. 108.—Dies.. 1858d, 267-268 (larva of Dist. appendiculatum Rud.) (in Buccinum linnæi; Genoa).
 - elegans (Mueller, 1855) Dies., 1858d, 269 (syn. Cerc. elegans Mueller in Valette, 1855) (free).—Dadai, 1888f, 85, 86; 1888g, 108, 109.—Mont., 1888, 193, 194.
 - ephemera (?Nitzsch. 1807) Hemprich & Ehrenberg, 1828a. pl. 6. fig. 3.—Dies. 1850a. 299 (includes Cerc. V. Baer. 1827, 625. pl. 31) (in Paludina vivipara: Regiomontii: Planorbis corneus: April. Halle, Berlin): 1855a. 392 (in Pl. corn., Pal. vivip.): 1858d. 244 (of Sieb. syn. of Glenocerc. flava Dies.). 267 (syn. of Cerc. ephemera Wagener) (of Ehrenberg larva of Dist. trigonoce-phalum Rud.) (in Pl. corn.. Pal. vivip.).—Moul., 1856a, 204 (to Cerc.).—Sieb., 1843.
 - fissa Bory de St. Vincent, 1824, 456; 1825b, 252 [see Lamouroux, 1824a].—Dies., 1850a, 295 (syn. of Malleolus furcatus).—Moul., 1856a, 168 (is not syn. of Cerc. furcata).
 - inquieta (Mueller, 1786) [Bory de St. Vincent, 1825b, 253 (Histrionelle inquiete]].— Dies., 1850a, 300 (sp. inq.) (in aqua marina): 1855a, 393 (Hab. primitivum ignotum); 1858d, 268-269 (free, Hafniæ).—Pag., 1862, 298.
 - lemna (Mueller, 1773) Hemprich & Ehrenberg, 1828.—Dies., 1850a. 299-300 (includes: Brachionus proteus Schrank. Cerc. major Nitzsch, Histr. annulicauda Bory) (in Lymnæus stagnalis, Planorbis carinatus, P. corneus): 1855a, 392 (in Lymn. stag. et Pl. corn.); 1858d, 268 (in Pl. carinatus. P. corn., Lymn. stag.).

HISTRIONELLA-Continued.

melanoglena Dies., 1855a, 393 (Melanoglena bipunctata Eichwald renamed) (Hab. primitivum ignotum); 1858d, 245 (to Glenocerc.).

[pupulaBory de St. Vincent, 1825b, 252 (Histrionelle poupée, Enchelis pupula Mull.).]

setosicauda Dadai, 1888f, 84–86, pl. 3, figs. 11, 13: 1888g, 107–109, pl. 3, figs. 11, 13.—Mont., 1888, 77 (setosicaudata); 1888, 193, 194 (setosicaudata).

sctosicaudata Mont., 1888, 77; 1888, 195, 194 (for setosicauda).

HISTRIONELLINA Dies., 1858d, 269 (type?).

erythrops (Dies., 1855) Dies., 1858d, 270 (in Paludina impura; Regiomonti).

fissicauda Dies., 1858d, 269–270 (syn. Cerc. ocellata Valette) (in Lymnæus stagnalis; Berlin).—Linst., 1878.

melanops (Dies., 1855) Dies., 1858d, 270 (syns. Cerc. melanops Dies.) (in Paludina impura; Regiomonti).

HOLLOSTOMOM Erc., 1881e, 88, for Holostomum.

HOLOMETRA Looss, 1899b, 564 (tod. exigua): ὀλος, entire; ἡ μήτηρ, uterus.—Braun, 1902b, 5.—Pratt, 1902, 888, 896.—Rail., 1900, 242.

exigua (Mueh., 1898) Looss, 1899b, 564, 678–679, fig. 4 (in Circus rufus; Adelen Island, Nile, near Heluan).—Stoss., 1904, 11.

HOLORCHIS Stoss., 1901, 93 (5) (tod. m. pycnoporus).

pycnoporus Stoss., 1901, 92–93 (rep. 4–5), pl. 6, fig. 10 (in Sargus salviani; Triest). HOLOSTOMATIDÆ Gamb., 1896, 63, 67, 73.

HOLOSTOMEE Brand., 1888a, 1-68, 72 pp. (family); 1889d, 241-245; 1890a, 590.

HOLOSTOMIDÆ E. Bl., 1847, 317.—R. Bl., 1888a, 541 (embraced in Distomiens).—
Brand., 1888a, 63; 1890a, 549-604, pls. 39-41; 1891a, 415-416.—Braun, 1883a, 58; 1890a, 515; 1892a, 570, 696, 707, 774, 775; 1893a, 887, 890, 895, 899, 900; 1894l, 165-167; 1894m, 755; 1894k, 680-682 (development); 1895b, 136.—Heider, 1900, 21.—Hoyle, 1890, 539 (Holost., Hemist., Eustemma).—Jackson, 1888, 654 (Holost.; larval forms Tetracotyle, Diplost., Hemist.).—
Johnston, 1904a, 108-116 (from Australian birds), pls. 5-6; 1904b; 1904c, (VII, 12), 759-760; 1905 (I, 31), 13.—Looss, 1899b, 541, 543.—Mont., 1888a, 91; 1891, 109, 110; 1892, Oct. 7, 214 (f. of Malacocotylea).—Much., 1898, 18.—Pratt, 1902, 889-890, 908 (includes Cyathocotylinæ, Diplostominæ, Hemistominæ, Helostominæ).—Rettger, 1897, 224-225.—Schneidemuchl, 1896, 295.—Stilcs & Hass., 1898a, 87.—Stoss., 1898, 20.

HOLOSTOMINÆ Mont., 1892, Oct. 7, 214 (subf. of Holostomidæ).—Braun, 1893a, 890, 895, 903.—Mueh., 1898, 19.—Pratt, 1902, 890 (Holostomum).

HOLOSTOMA, see Holostomum.

HOLOSTOMUM Nitzsch, 1819, 399–401 (type by inclusion variabile=Planaria strigis, type of Strigea); 1819, 35.—Baillet, 1866b, 99, 106.—Blainv., 1824a, 518 (holostome); 1828, 583–584.—E. Bl., 1847, 318.—Brand., 1888a, 8, 9, 10, 12, 13, 14, 17, 49, 50, 63; 1888b, 424–426; 1888, 954; 1889a, 67–68; 1890a, 576, 579, 590 (of Rud.).—Braun, 1883a, 40; 1892a, 570, 581, 600, 681, 715, 748, 792, 793, 794, 795, 797; 1893a, 844, 872, 879, 884, 886, 887, 890, 894, 895, 900, 902, 903, 905, 917; 1894, 166; 1895b, 132, 136.—Burm... 1837, 530; 1856a, 250.—Carus, 1863, 479.—Crep., 1839, 286, 287.—Dav., 1877. lxxix.—Dies., 1836, 237; 1850a, 287, 305 (of Nord., syn. of Diplost.), 307 (of Nitzsch pars, syn. of Hemist.), 312–317 (of Nitzsch syrs.: Plan. Gœze, Festuc. Schrank, Fasc. Gmelin, Strigea Abildg., Amphist. Rud.): 1858e, 312, 319–322.—Duj., 1845a, 364–366.—Dunglison, 1893, 533.—Eichwald, 1829a, 248.—Fischer, 1840, 156.—Fischder., 1902a, 6, 7 = Strigea); 1903h, 487 (=Amphist. capite discreto), 490 (syn. of Strigea), 507.—Gamb.,1896, 73.—Goldb., 1855, 17.—Hoyle, 1890, 539.—Jackson, 1888, 643, 648, 652, 653, 654 (Tetracotyle, Diplost.: larval forms).—Johnston, 1904, 3 n. Australian sp. from birds.—Lamouroux, 1822a, 194, 297 (syn. of Amphist.).—Leuck., 1863, 452, 524.—Looss, 1902m, 438, 439.—Luehe, 1901, 175.—Macleay, 1886, 342.—Mont., 1888, 8, 34, 63, 68, 71, 72, 83, 84, 91, 95, 104; 1891, 101, 105, 109; 1892, Oct. 7, 214 (gen. of Helostominæ); 1892, 709.—Moul., 1856a, 12, 15.—Nord., 1840, 626, 627–628.—Par., 1887, 329.—Pratt. 1902, 890, 908.—Schneidemuehl. 1896, 295.—Stoss., 1898, 51.—Tasch., 1879, 233.—Villot, 1898, 528; 1878, 19.—Wallenstedt, 1847, 7.—Wolf, 1903, 607.—See Strigea.

HOLOSTOMUM—Continued.

alatum (Geze, 1782) Nitzsch, 1819, 399, 400, pl. 4, figs. 1–4 (syns. Plan. al. Rud., Plan. al. Geze, Dist. vulpina Abildg.).—Baillet, 1866b, 106–107 (syn. Hemist. al.).—Ben., 1858a, 1861a, 179.—Blainv., 1828, 583.—E. Bl., 1847, 318–320, pl. 10, fig. 1.—Brand., 1888a, 10, 12, 38, 39, 60 (to Hemist.).—Cobbold, 1879b, 300.—Crep., 1829b, 66; 1839, 287.—Dav., 1877a, lxxix.—Dies., 1850a, 308 (to Hemist.).—Duj., 1845a, 367, pl. 8, fig. D.—Dunglison, 1893, 533.—Gurlt, 1831a, 375, pl. 8, figs. 39–40.—Macleay, 1886, v. 10, 342.—Mehlis, 1831, 175.—Mont., 1891, 105.—Nord., 1840, 628 (syn. Dist. al. Rud.).—Stoss., 1890, 131.—Verrill, 1870, 173.—Ward, 1895, 341 (in Canis familiaris).

anatis nigræ Dies., 1858e, 322, sp. inq. (based on Bellingham, 1844a, 340) (in Anas (Oidemia) nigra).—Brand., 1888a, 68 (syn. of H. crenulatum Cobbold); 1890a, 596.—Braun, 1893a, 903.—See also crenulatum.

auritum Duj., 1845a, 370 (t. h. Strix flammea; Rennes).—Brand., 1888a, 12.—Dies., 1850a, 312 (to Hemist.).

bellinghamii Cobbold, 1860a, 45, falconum renamed (in Falco nisus, F. rufus).— Brand., 1888a, 68 (in F. nisus, F. rufus); 1890a, 596 (syn. H. falconum Dies.).

brevicaudatum Nord., 1832a, 52-53 (t. h. Barbus communis; Berlin); 1840, 618.-Braun, 1892a, 795.—Crep., 1839, 287.—Dies., 1850a, 306 (to Diplost.).—Duj., 1845a, 380.—Gescheidt, 1833a, 430.—Moul., 1856a, 220.—Steenstrup, 1842, 58.

bulbosum Brand., 1888a, 67; 1890a, 595 (in Geronticus albicollis, Nauclerus furcatus).—Braun, 1893a, 903.

bursarium Nitzsch, in Giebel, 1857, 265 (t. h. Falco peregrinus).

bursigerum Brand., 1888a, 65 (in Larus ridibundus; Wien. Mus.); 1890a, 592, pl. 41, figs. 15–18.—Braun, 1892a, 586; 1893a, 903.—Stoss., 1891, 216; 1892, 65; 1898, 22.

cinctum Brand., 1888a, 34, 67 (in Ardea sp.; by Natterer); 1890a, 564, 594, pl. 41, figs. 21–22.—Braun, 1893a, 903 (Brazil).

clavus Mol., 1858, 128 (t. h. Gadus merlucius; Patavii); 1861, 196–197, pl. 1, figs. 9–11.—Brand., 1888a, 67; 1890a, 595.—Braun, 1893a, 874, 903.—Cobbold, 1879b, 462.—Dies., 1858e, 322 (in G. merl.).

coniferum Mehlis in Crep., 1846, 138 (t. h. Colymbus cristatus).

cornu Nitzsch in Rud., 1819a, 89, 357 (t. h. Ardea cinerea) (to Amphist.).—Brand., 1888a, 66; 1890. 594 (in Ardea sp.).—Braun, 1893a, 879-903; 1894k, 682.— Dies., 1850a, 315 (syn. Amphist. cornu Rud nec Dies.); 1858e, 321 (Amphist. cornu Bellingham) (in Ardea cinerea, A. herodias, A. stellaris).—Duj., 1845a, 374.—Linst., 1877, 187.—Schlotthauber, 1860, 129.—Stoss., 1895, 37; 1897, 9; 1898, 22 (in Ardea cinerea, Rovigno; A. purpurea; Monfalcone).—Wedl, 1857, 253–254. pl. 1, fig. 19.—Westrumb, 1823, 394.—Wolffhuegel, 1900, 9, 12.—Reported also for Accipiter nisus, Ardea garzetta, A. purpurea, Ciconia alba.

cornucopia Mol., 1859, 287–288 (t. h. Strix flammea; Batavii); 1861, 196, pl. 1, fig. 8.—Brand., 1888a, 68 (in Strix otus); 1890a, 596 (in Otis vulgaris).—Braun, 1892a, 585, 610, 674, 784, 785, 786, 787, 792, 793, 794; 1893a, 903.—Dies., 1859c, 424 (cornucopiæ).—Jackson, 1888, 652.—Linst., 1877, 188—191, pl. 13, fig. 20, pl. 14, figs. 29–30; 1885, 253–254, pl. 15, fig. 31; 1890f, 186 (cornucopiæ); 1906, 14 (in Strix flammea).—Piana, 1898, 9.—Also reported for Ægolius otus.

cornucopiæ Dies., 1859c, 424, for cornucopia.

cornutum (Rud., 1809) Duj., 1845a, 372-373.—Brand., 1888a, 68 (syn. H. multilobum Cobbold) (in Charadrius pluvialis); 1890a, 596 (of Dies.).—Braun, 1893a, 903.—Dies., 1850a, 316-317.

corones Dies., 1858e, 322 (in Corvus corone; Ireland).—Brand., 1888a, 68 (syn. of H. dubium Cobbold); 1890a, 596.—Braun, 1893a, 903.

crenulatum Cobbold, 1860a, 47 (in Anas nigra; Ireland).—Brand., 1888a, 68 (syn. H. anatis nigræ Dies.) (in Anas oidemia); 1890a, 596 (in Œdemia nigra).

cuticola Nord., 1832, 43, 49-52, pl. 4, figs. 1-4 (in Cyprinus carpio, C. brama, C. blicca, C. idus, C. rutilus, C. erythrophthalmus), belongs to Cryptostomum; 1840, 618, 628.—Brand., 1888a, 11, 15.—Braun, 1892a, 795.—Crep., 1839, 287; 1846, 154.—Dies., 1850a, 306 (to Diplost.).—Duj., 1845a, 379–380.—Gescheidt, 1833a, 429.—Kroyer, 1838–40a, 21, 578; 1846–53a, 388, 399, 434, 446, 462; 1852-53a, index (cuticula, 1250) (in Perca fluviatilis L., Abramis brama L., A. blicca Bl., Leuciscus erythrophthalmus L., L. rutilus L., L. idus L.).—Moul., 1856a, 217–218, 220 (in Perch, cyprins).—Pavesi, 1881, 615, 616 (in Cobitis tænia).—Steenstrup, 1842, 58.—Villot, 1898, 541.—Waldenburg, 1860, 5, 11.—Also reported for Cyprinus vimba.

HOLOSTOMUM-Continued.

cuticula Kroyer, 1852-53a, 1250, for cuticola.

denticulatum (Rud., 1819) Duj., 1845a, 372, pl. 8, fig. a, 1, 2.—Brand., 1888a, 12.—Dies., 1850a, 311 (to Hemist.) (in Alcedo ispida; Rhedoni).—Villot, 1878, 19; 1898, 538.

dubium Cobbold, 1860a, 45 (in Corvus corone).—Brand., 1888a, 68 (syn. H. corones Dies.); 1890a, 596 (in C. corone).

ellipticum Brand., 1888a, 67 (in Bubo magellanicus; by Natterer); 1890a, 595.— Braun, 1893a, 903.—Linst., 1906, 15.

crraticum (Rud., 1809) Duj., 1845a, 373-374.—Baillet, 1866b, 107.—Brand., 1888a, 59 (of Linst., 1877c) (syn. of Hemist. pileatum), 63-64; 1890a, 571, 586, 591, pl. 41, figs. 3-4.—Braun, 1892a, 797; 1893a, 844, 874, 879, 903 (Brazil); 1894, 166.—Dies., 1850a, 313 (syns. Strigea anatis tadornæ Viborg, S. candida Abildg., Amphist. anatis tadornæ Rud., A. isost. Rud., A. erraticum Rud., H. macrocephalum Crep.); 1858c, 320 (syn. Amphist. isostomum Bellingham) (in Anas boschas fera, A. ferina).—Erc., 1881e, 48-54, 88 (Hollostomom); 1882a, 284-290, 324.—Kowal., 1894, 2.—Linst., 1877, 188, pl. 13, figs. 18-19; 1906, 14 (in Strix otus, Falco albicilla).—Mont., 1888a. 71.—Olss., 1893, S.—Par., 1899, 4; 1902, 4 (in Querquedula circia; Elba).—Piana, 1898, 4.—Also reported for Alca torda, Anas boschas, A. crecca, A. fusca, Ascolopax gallinago, Colymbus arcticus, C. balticus, C. septentrionalis, Fuligula cristata, F. ferina, F. marila, Harelda glacialis, Limosa melanura.

custemma Brand., 1888a, 65-66 (in Accipiter pileatus; Brazil); 1890a, 593, pl. 41,

fig. 25 (syn. Eustemma caryophyllum Dies.).—Braun, 1893a, 903.

excavatum (Rud., 1803) Nitzsch, 1819, 399, figs. 5–7 (in Ardea nycticorax).—Ben., 1868, 296, 300, pl. 2, figs. 6–8.—Blainv., 1828, 584.—Brand., 1888a, 12.—Braun, 1893a, 903; 1894k, 682.—Dies., 1850a, 309 (to Hemist.).—Duj., 1845a, 375.—Gamb., 1896, 63.—Leuck., 1879, 14; 1886d, 11.—Looss, 1892, 14.—Nord., 1840, 628.

excisum Linst., 1906, 12-15, pl. 1, figs. 14-16 (in Ægolius otus, Strix flammea).

exiguum Mehlis in Crep., 1846, 145 (t. h. Cygnus musicus).—Also reported for Mergus merganser, M. serrator.

falconum Dies., 1858e, 322, sp. inq. (based on Bellingham, 1844, 39) (in Falconisus et F. rufus; Hibernia).—Brand., 1888a, 68 (syn. of H. bellinghamii Cobbold); 1890a, 596.—Braun, 1893a, 903.—Linst., 1883, 311.—See also bellinghamii.—Also reported for Astur nisus, Λ. palumbarius, Circus rufus.

gracile (Rud., 1819) Duj., 1845a, 378.—Brand., 1888a, 68 (in Mergus merganser); 1890a, 595, pl. 41, fig. 26.—Braun, 1893a, 903.—Crep., 1846, 141, 142, 144, 145.—Dies., 1850a, 315; 1858e, 321 (in Colymbus glacialis, Anas nigra) (Amphist. gracile Bellingham).—Kowal., 1896d, (2) 252 (in Anas crecca, A. boschas dom.; Dublany); 1904, (8) 23 (in Mergus merganser).—Linst., 1877, 188, pl. 13, fig. 17.—Par., 1901, 6.—Also reported for Anas clangula, Λ. fusca, Anser albifrons.

hillii Johnston, 1904, 110-111, pl. 6, figs. 1-8 (in Larus novæ hollandiæ Stephens). isostomum (Rud., 1814) Duj., 1845a, 377.

lagena Mol., 1858, 127 (t. h. Strix passerina; Patavii); 1861, 195.—Brand., 1888a,
68 (in St. pass.); 1890a, 596 (in Glaucidium passerinum).—Braun, 1893a,
903.—Dies., 1858e, 320 (in St. pass.).—Linst., 1906, 15 (in Gl. pass.).

longicolle (Rud., 1819) Duj., 1845a, 374–375.—Brand., 1888a, 64–65 (in Botaurus stellaris; Wien. Mus.); 1890a, 592, pl. 41, fig. 19.—Braun, 1892a, 586; 1893a, 903.—Crep., 1849, 69.—Dies., 1850a, 316; 1858e, 321 (in Larus argentatus) (syn. Amphist. longicolle Bellingham).—Leuck., 1863, 481.—Mol., 1859, 818 (in Larus ridibundus; Padua).—Mueh., 1898, 19.—Par., 1887, 330–331.—Stoss., 1890, 50; 1891, 216; 1895, 37; 1897, 9; 1901, 91 (3) (in Botaurus stellaris; Albona); 1898, 21–22.—Reported also for Ardea alba, A. stellaris, Larus cachinnans, Xema ridibundum.

lyratum Schlotthauber, 1860, 129 (in Ardea cinerea).

macrocephalum (Rud., 1803) Blainv., 1828, 584.—Baird, 1853, 47.—Brand., 1888a,
68; 1890a, 596.—Braun, 1891d, 424 (in Aquila chrysaëtos, Astur palumbarius,
Circus æruginosus, C. cyaneus, Strix noctua); 1893b, 185.—Crep., 1839, 288;
1846. 130.—Dies., 1850a, 312, 313 (of Crep. and Duj., syn. of H. variabile
Nitzsch), 313 (of Crep. pars, syn. of H. erraticum Duj.); 1858e, 320 (of Crep.,
1849, 1, 64, syn. of H. variabile Nitzsch).—Duj., 1845a, 368–369.—Fischder.,

HOLOSTOMUM-Continued.

1901, 367; 1902a, 7 (Plan. strigis Geeze, 1782); 1903h, 490 (=strigis, type of Strigea, Amphist., Holost.).—Leuck., 1863, 460.—Linst., 1906, 14 (syn. H. variabile Nitzsch) (in Buteo vulgaris).—Shipley, 1905, v. 6 (1),7 (type of Strigea).—Stoss., 1890, 131; 1890, 50; 1891, 111; 1892, 66; 1896, 126; 1898, 21.—Also reported for Falco buteo, F. nisus, F. palumbarius.

megalocephalum Brand., 1888a, 67 (in Stomias sp.; by Natterer); 1890a, 595.—

Braun, 1893a, 903 (Brazil).

microstomum (Rud., 1809) Duj., 1845a, 371.—Brand., 1888a, 68 (in Corvus caryocatactes); 1890a, 596 (in Caryocatactes nucifraga).—Braun, 1893a, 874, 903.—Dies., 1850a, 314 (in Corv. caryoc.; Gryphiæ).

multilobum Cobbold, 1860a, 46-47 (in Charadrius pluvialis).—Brand., 1888a, 68;

1890a, 596 (syn. of H. cornutum Dies.).

musculicola Waldenburg, 1860, 12 (in Cyprinus, Perca fluviatilis).—Brand., 188a, 15, 52; 1890a, 578.—Villot, 1898, 542.—Reported for Abramis brama, Scardinius erythrophthalmus.

musculosum Johnston, 1904, 112-114, pl. 7, figs. 4-9 (in Sterna bergii Licht.).

mtidum Leidy, 1856, 45 (t. h. Rana pipiens).—Brand., 1888a, 14, 68 (in R. pip.); 1890a, 595.—Braun, 1893a, 903.—Dies., 1858e, 321–322 (in R. pip.; Phila.).—Staff., 1902, 724.

ochreatum Nitzsch, in Giebel, 1857, 265 (t. h. Falco haliaëtos).

patagiatum Crep., 1846, 135 (t. h. Ardea stellaris).

pellucidum Schlotthauber, 1860, 129 (in Petromyzon fluviatilis).

pileatum (Rud., 1802) Blainv., 1828, 584.—Duj., 1845a, 377.—Ben., 1858a, 1861a, 179.—Brand., 1888a, 12.—Braun., 1893a, 903.—Crep., 1846, 137.—Dies., 1850a,

314-315.—Reported for Sterna macrura.

platycephalum (Crep., 1825) Duj., 1845a, 376.—Baird, 1853a, 48.—Brand., 1888a, 63 (syn. of H. variegatum Duj.); 1890a, 591.—Braun, 1891d, 424 (in Larus ridibundus); 1893a, 876; 1893d, 467; 1894, 166.—Crep., 1839, 287; 1846, 129, 136, 139.—Dies., 1850a, 313.—Mégnin, 1891a, 323 (in Larus ridibundus).—Much., 1898, 16.—Reported also for Carbo cormoranus, Colymbus cristatus, C. rufogularis, Falco albicilla, Halieus carbo, Larus argentatus, L. canus, L. fuscus, Lestris pomarina.

podomorphum Nitzsch, 1819, 399–400, figs. 8–10 (in Falco haliaëtos; Europe).— Brand., 1888a, 12; 18—, 62.—Dies., 1850a, 311 (to Hemist.) (in F. hal.; Halle).—

Duj., 1845a, 370.

rotundatum Linst., 1877, 187–188 (in Lanius collurio).—Brand., 1888a, 66 (syn. of

H. spherula Duj.) (in Lanius collurio); 1890a, 593.

serpens Nitzsch, 1819, 400–401 (in Falco haliaëtos), figs. 17–22; in Rud., 1819a, 88, 353–354 (t. h. Falco haliaëtus) (to Amphist. by Rud., 1819a, 88, 353; returned to Holost. by Duj., 1845a, 371, and Dies., 1850a, 316); 1819, 399, 400–401, pl. 4, figs. 17, 22.—Ben., 1858a, 1861a, 197.—Blainv., 1828a, 584.—Brand., 1888a, 11, 68 (in F. hal.); 1890a, 596; 1891b, 265.—Dies., 1850a, 316 (syn. Amphist. ser.).—Braun, 1892a, 586, 746, 751; 1893a, 903.—Giebel, 1857, 265.—Mueh., 1898, 19.—Nord., 1840, 616, 628 (syn. Amphist. ser.).—Linst., 1904, 254.—Looss, 1893b, 810.—Sieb., 1835, 66.

simplex Johnston, 1904, 112, pl. 7, figs. 1-3 (in Ardea novæhollandiæ Lath.).

spathaceum (Rud., 1819) Duj., 1845a, 375-376.—Baird, 1853a, 48.—Brand., 1888a,
 12.—Crep., 1846, 139.—Dies., 1850a, 310 (to Hemist.).—Erc., 1881e, 54;
 1882a, 290.—Nord., 1840, 628.—Stoss., 1898, 20.

spatula Sieb., 1835, 57.—Dies., 1836, 240, for spathula.—Baird, 1853a, 48 (in

Accipiter nisus).

spatulatum (Rud., 1819) Duj., 1845a, 376.—Brand., 1888a, 12.—Cobbold, 1876,
 97.—Dies., 1850a, 367 (to Dist.).—Kowal., 1898 (to Echinost.).—Nord., 1840,
 628.—Stoss., 1892, 186.

spharocephalum (Westrumb, 1823) Dies., 1850a, 314 (in Coracina scutata; Brazil).— Brand., 1888a, 65 (syns. Amphist. sph., Holest. westrumbii) (in Anas moschata); 1890a, 592–593, pl. 41, fig. 20.—Braun, 1893a, 903.—Wolffhuegel, 1900, 9, 60.

HOLOSTOMUM—Continued.

sphærula (Rud., 1803) Duj., 1845a, 371–372.—Blochmann, 1892b, 650 (in Corvus cornix).—Brand., 1888a, 66 (syn. H. rotundatum) (in Corvus cornix, C. corone, Lanius collurio, Oriolus cristatus); 1890a, 593, 594, pl. 41, fig. 7.—Braun, 1893a, 904; 1894k, 681.—Crcp., 1846, 132.—Dics., 1850a, 314 (in Corvus cornix, C. corone, C. frugilegus); 1858e, 321 (in C. glandarius).—Kowal., 1896d, 252 (2) (in Corvus cornix L.; Dublany); 1904, 23 (8) (in Garrulus glandarius).—Linst., 1877, 187.—Mol., 1858, 128; 1861, 196.—Mueh., 1898, 19.—Wolffhuegel, 1900, 9, 26, 33, 41.

squamosum Villot, 1878, 20–21, pl. 5, figs. 2–3 (in Strepsilas interpres).—Brand., 1888a, 18; 1890a, 577.—Linst., 1889.—Mont., 1888a, 23, 71.

tenuicolle (Westrumb, 1823) Dies., 1850a, 316 (in Falco rufus).—Brand., 1888a, 66 (in F. rufus); 1890a, 594, pl. 41, fig. 23.—Braun, 1892a, 586; 1893a, 904.— Also reported for Circus rufus.

unciforme (Rud., 1819) Brand., 1888a, 66; 1890a, 594 (in Oriolus cristatus).— Braun, 1893a, 904.

urniceps Schlotthauber, 1860, 129 (in Falco pygargus, int.).

urnigerum (Rud., 1819) Duj., 1845a, 378-379.—Baird, 1853a, 49 (syn. Codonocephalus mutabilis Dies.).—Brand., 1888a, 13.—Dies., 1836, 241; 1858e, 323 (of Wedl syn. of Co. mut., type Dies.).—Hannover, 1864a, 5.—Moul., 1856a, 219 (in Rana esculenta).—Sieb., 1835, 57.—Sons., 1893, 190 (syn. of Co. mut. Dies.).—Wedl, 1857, 255.

vaginatum Brand., 1888a, 64 (in Cathartes sp. by Natterer); 1890a, 591, pl. 41, fig. 24.—Braun, 1893a, 904 (Brazil).

variabile Nitzsch, 1819, 400, pl. 4, figs. 11–16 (in owls; Germany), syn. Amphist. macrocephalum Rud.; in Giebel, 1857a, 265 (in Strix aluco, Scolopax gallinula).—Baird, 1853a, 47 (=H. macroc. Blainv.).—Brand., 1888a, 5, 32, 63, 68 (in Strix aluco); 1890a, 561, 590, 596, pl. 41, fig. 1.—Braun, 1892a, 699; 1893a, 879, 881, 904; 1894k, 682; 1895b, 11.—Crep., 1839, 288.—Dies., 1850a, 312–313 (syns. Planaria teres poro simplici Geze, Festuc. strigis Schrank, Fasc. strigis Gmelin, Strigea Abildg., Amphist. macrocephalum Rud., H. macrocephalum Crep.) (in Strix brachyotus, S. bubo, S. flammea, S. otus, S. aluco, S. passerina, S. tangmalmi, Falco tinnunculus, F. apivorus, F. albicilla, F. nævius, F. gallicus, F. lagopus, F. cineraceus, F. ruíus, F. haliaetos, F. buteo, F. peregrinus, F. pennatus, F. cyaneus, F. ruípes, F. imperialis); 1858e, 320 (syns. Amphist. macrocephalum Bellingham, 1844; H. macroc. Crep., 1849) (in Falco rufus, F. peregrinus, Ardea cinerea, Strix nyctea, S. pygmaa). (in Falco rulus, F. peregrinus, Ardea cinerea, Strix hyctea, S. pygmea).—Giebel, 1857, 265.—Hoyle, 1890, 539.—Kastenbaum, 1899, 244, fig. 33, 7.—Kowal., 1896d, (2) 252 (in Buteo vulgaris Bechst; Dublany); 1904, (8) 23 (in Otus brachyotus; Dublany).—Linst., 1877, 187; 1903, 279; 1905, 191; 1906, 14 (syn. of H. macrocephalum).—Mol., 1858, 127; 1859, 818 (in Strix otus); 1861, 194–195, pl. 1, figs. 6–7.—Much., 1898, 16, 19.—Nord., 1840, 626 (syn. of Amphist. macrocephalum).—Par., 1887, 329–330; 1899, 4; 1902, 3–4 (in Aesalon regalus, Asio otus, Circus cyaneus; Elba).—Stoss., 1892, 66 (in Falco peregrinus; near Triest); 1895, 37 (in Circus æruginosus; Accipiter nisus at Triest; Syrnium aluco in Dalmatia); 1896, 126 (in Cerchneis tinnumnisus at Triest; Syrnium aluco in Dalmatia); 1896, 126 (in Cerchneis tinnunculus; Staranzano); 1898, 21.—Villot, 1898, 542 (adult of Tetracotyle colubri).—Walter, 1866, 64 (A. macrocephalum Crep.).—Wedl, 1857, 252–253, pl. 1, fig. 18.—Wolffhuegel, 1900, 9, 11 (in Vultur calvus), 16, 17, 44.—Reported for Aegolius brachyotus, A. otus, Aquila albicilla, A. imperialis, A. nævia, A. pennata, Archibuteo vulgaris, Ardea cineria, Astur nisus, Brachyotus palustris, B. variabilis, Bubo maximus, Buteo lagopus, Circaetus gallicus, Circus cineraceus, C. cyancus v. hudsonius, C. rufus, Falco cyaneus, Nyctea nivea, Otus vulgaris, Pica caudata, Picus sp., Scolopax gallinula.

variegatum (Crep., 1825) Duj., 1845a, 376-377.—Brand., 1888a, 7, 63 (syn. H. platycephalum Duj.).—Baird, 1853a, 48.—Braun, 1893a, 844, 904; 1894, 166, 167; cepnaum Dul.).—Baird, 1853a, 48.—Braun, 1893a, 844, 904; 1894, 166, 167; 1894k, 681.—Crep., 1846, 139.—Dies., 1850a, 315.—Duj., 1845a, 376-377 (in Larus maximus).—Mueh., 1898, 16, 19-20, fig. 9.—Villot, 1898, 542 (adult of Tetracotyle ovata).—Wolffhuegel, 1900, 9, 62.—Reported for Alca torda, Anas boschas, Carbo cormoranus, Colymbus rufogularis, C. scptentrionalis, Falco albicilla, Fuligula nyroca, Harclda glacialis, Larus argentatus, L. marinus, L. ridibundus, Podiceps cristatus, Uria troile, Xema ridibundum). westrumbii Cobbold, 1860a, 45 (Amphist. sphærocephalum Westrumb, renamed) (in Coracina scutata; Brazil).

HOMALOGASTER Poir., 1883, 74–76, 79 (m. paloniæ)(ὁμαλός=flat; γαστήρ=stomach); 1885, 120.—Braun, 1892a, 568, 581, 645, 663; 1893a, 879, 886, 890, 892, 895, 904, 906, 918.—Fischder., 1901, 374; 1902a, 7, 46, 47; 1903h, 489 (in Palo nia frontalis); 1903h & i, 630–631 (diagnosis, type paloniæ).—Gamb., 1896, 73.—Hoyle, 1890, 539.—Looss, 1895, 11; 1896b, 17; 1899b, 541; 1902m, 440.— Mont., 1888, 7, 12, 14, 35, 91, 103; 1892, Oct. 7, 214 (gen. of Amphistominæ).—Piana & Stazzi, 1900, 523.—Pratt, 1902, 887, 892.—Shipley, 1905, v. 6 (1), 8.—Sons., 1895, 184, 186; 1896, 310.—Stiles. 1898a, 24.

paloniæ Poir., 1883, 74–76. pl. 2, figs. 1 a-b (t. h. Palonia frontalis; Java).—Braun, 1893a, 874, 907; 1893d, 466.—Fischder., 1901, 374; 1902a, (7) 47 (in Palcnia frontalis; Java); 1903h, 630 (Dickdarm, Palonia frontalis; Java).—Giard & Billet. 1892a, 615.—Linst., 1889a, 21.—Sons., 1895, 184.—Stiles, 1898a, 24, 37,

72, 139, fig. 67.

 poirieri Giard & Billet, 1892a, 615 (in cattle; Tonkin).—Fischder., 1901, 374;
 1902a, 47-48 (in Bos taurus; Tonkin); 1903h, 630-631 (in Tonkin cattle).—
 Rail., 1893, 379.—Sons., 1895, 185.—Stiles, 1898a, 24, 67, 140.—Ward, 1895, 332 (in Bos taurus).

HOMALOMETRON Staff., 1904, May 3, 487 (m. pallidum); 'ομαλός, regular; μέτρον,

pallidum Staff., 1904, May 3, 487 (t. h. Fundulus heterochitus L.; Woods Hole) (based on Lint., 1901, 422, pl. 32, fig. 354).

HOPLODERMA Cohn, 1903, 35-37 (m. mesocœlium).—Poche, 1907, 125 (Cohn not Michael, renamed Pintneria).

mesocalium Cohn, 1903, 35-37, figs. 1-2 (in Draco volans; Eastern Java).

(HORMOCERCARIA) Dies., 1855a, 390, 394 (subg. of Cerc.) (type species echinata, or echinatoides) (corpus infra os collari echinis percurso cinctum).—Renamed (Nephrocephala), 1858.

echinata (Sieb., 1837) Dies., 1855a, 390-391; 1858d, 261. [See Dist. echinatum,

type of Echinost.

echinatoides (Fil., 1854) Dies., 1855a, 391–392; 1858d, 262. [Apparently type by elimination.] See, however, below, p. 385, Addenda.

HYDRICUCULUS McCrady, 1874a, 178 (m. cuculus, see sub Bucephalus) generic name abandoned by McCrady at date of publication.

HYPOSTOMA (Rud., 1809a, 36–37, 325–327) (nec Hypostomus Lac., 1803, fish) (type species probably caryophyllinum by Blainv., 1828a, 581); *ὑπο*, below; δτομα, mouth; originally a subg. of Monost.; see also (Hypost.).—Blainv., 1824a, 518 (hypostome. "C'est un genre bien douteux, peut-être un degré de développement"); 1828a, 581.—Crep., 1839, 285.—Looss, 1902m, 830.-Nord., 1840, 623.—Olfers, 1816, 48.

caryophillinus (Rud., 1802) Blainv.. 1828. 581.—Dies., 1850a. 328 (to Monost.).—Duj., 1845a, 360 (to Monost.).—Nord., 1840, 623 (to Monost.).

(HYPOSTOMA) Rud., 1809a, 36–37, 325–327, subg. of Monost. See Hypostoma. caryophyllinum (Rud., 1802) Rud., 1809a, 325–326 (to Hypost. by Blainy., 1828, 581; returned to Monost. by Duj., 1845a, 360).

cochleariforme Rud., 1809a, 326-327 (= Festuc. cyprinacea Schrank, 1790, renamed)

(in Cyprinus barbus).

gracile Rud., 1809a, 326 (t. h. Salmo eperlanus).

HYPOSTOMATA Rud., 1809a, 325, plural of Hypostoma.

ICHTHYDIUM Hemprich & Ehrenberg, 1828a (m. Cerc. podura Mueller; Nubia; Berlin), n. g. Ichthydinorum.

ITYOGONIMUS Luehe, 1899k, 538 (tod. ocreata Zed. =lorum Duj.; "rvs, margin, because of marginal position of genital pore); 1900, 557; 1901, 488.—Braun, 1900h, 6, 13; 1901i, 56; 1901, 897; 1902b, 129, 134.—Looss, 1900, 607; 1901, 206; 1902m, 755 (= Cucullanus Schrank), 839.—Pratt, 1902, 889, 907.

ocreatus (Geze, 1782) Braun, 1902b, 135.—[Luehe, 1899, 538 (= Dist. lorum

KLEPSITROMIS Hammerschmidt in Leuck., 1835a, 88 (m. melolonthæ) "das aber viele Aehnlichkeit mit Distoma zu haben scheint."

melolonthæ Hammerschmidt in Leuck., 1835a, 88, a drawing in Vienna.

KÖLLIKERIA Cobbold, 1860a, 31 (m. filicolle) (not Agassiz, 1862; not Mingazzini, 1891, protozoon).—Braun, 1893a, 885, 886 (Kœllikeria), 894, 895, 908, 909, 911, 918; 1895b, 127, 136.—Gamb., 1896, 73.—Lint., 1901, 447.—Looss, 1899b, 536, 542, 543; 1901, 196.—Moniez, 1896, 83.—Mont., 1888, 92, 105; 1893, 149, 153, 154.—Pratt, 1902, 889, 908.—Schneidemuehl, 1896, 295.—Stiles & Hass., 1898a, 91, 98 (type Monost. filicolle Rud.).—Stoss., 1892, 4.—Tasch., 1879, 608.

1893: Kællikeria Braun, 1893a, 886, for Köllikeria.

filicollis (Rud., 1819) Cobbold, 1860a, 31; 1879b, 462.—Ariola, 1906, 186, to (Dist.) (in Brama raii).—Braun, 1893a, 912.—Sons., 1890, 143 (in Brama raji).

okeni Ariola, 1906, 186 (syn. Dist. okeni Koell.), for okenii.

[staurocephali Mingazzini, 1891, protozoon.]

(KÖLLIKERIA) as subg. of Distoma. See also as genus.

dicorynum (Dies., 1850) Stoss., 1886, 47, as doubtful member of subg. (in Lampris guttatus).

filicolle (Rud., 1819) Stoss., 1886, 58.

LATIUSCULA Geze, 1782a, 41, 169–173 (a "Klasse" of Plan. Geze, containing: F. hepatica, Fledermausplattwurm, F. lucii, F. scorpii, F. lucioperce, F. percæ cernuæ, F. serrulata, F. farionis, F. salmonis, F. varica, F. platessæ, F. blennii).

LECANE Nitzsch, 1827, 69 (contains Cerc. orbis, C. luna).

LECHRIORCHIS Staff., 1905, Apr. 11, 691 (m. primus) $\lambda \dot{\epsilon} \chi \rho \iota o s$, oblique; $\ddot{o} \rho \chi \iota s$, testicle) (resembles Saphedera, Plagiorchis and Haplometra).

primus Staff., 1905, Apr. 11, 691 (t. h. Eutenia sirtalis L.; Canada) (includes Dist. sp. of Staff., 1902, 482).

LECITHASTER Luehe, 1901n, 395, 396 (tod. bothryophorus Olss. of Looss), 479–480.—Odhn., 1905, 357, 358, 359, 360, 366 (type confusus Odhn. by Odhn.).—Pratt, 1902, 889, 906.—Staff., 1904, 484.

bothryophorus (Olss., 1868 of Looss) Luehe, 1901n, 396, 480.—Odhn., 1905, 357 (botryophorus).—Staff., 1904, May 3, 484 (syn. Apoblema mollissimum Lev.) (in Salmo salar L., Clupea harengus L.).

botryophorus Odhn., 1905, 357, for bothryophorus, renamed confusus, 1905.

confusus Odhn., 1905, 357, 358, 359 (made type of genus by Odhn.) (syns. L. bothryophorus of Luehe, Dist. mollissimum Levin. of Stoss., Apoblema mol. Levin.), in intestine of Alosa finta of Mediterranean, and Clupea harengus of North Sea.

gibbosus (Rud., 1802) Luehe, 1901n, 480, in intest. of Belone acus, in Greifswald.—Odhn., 1905, 356-359 (syns. Fasc. gib. Rud., 1802, 81, pl. 2, fig. 7; Dist. gib. Rud., 1809a, 404, pl. 6, fig. 8; D. bergense Olss., 1868, 43, pl. 5, fig. 93; D. mollissimum Levin., 1881, 59, pl. 2, fig. 4; Lecithaster gib. Luehe, 1901, 480; [not Dist. botryophoron Olss., 1868, 42, pl. 5, fig. 9; not D. mollissimum Levin., of Stoss., 1889, 1-2; not Apoblema mollissimum (Levin.) Looss, 1896, 121, pl. 9, figs. 85-87]).—Nicoll, 1907, 72, 89-90 (syn. L. mollissimus) (in Ammodytes tobianus).

mollissimus (Levin., 1881) Nicoll, 1907, 89 (syn. of L. gibbosus).

LECITHASTERINÆ Odhn., 1905, 360, 364 (includes Lecithaster and Lecithophyllum).

LECITHOBOTRYS Looss, 1902h, 134, 135 (tod. putrescens).

putrescens Looss, 1902h, 135, 143, fig. 14 (in Mugil auratus).

LECITHOCHIRIINÆ Luehe, 1901n, 481 (includes Lecithochirium, Derogenes, and Lecithaster).

LECITHOCHIRINÆ Nicoll, 1907, 89, for Lecithochiriinæ.

LECITHOCHIRIUM Luehe, 1901n, 395, 396 (tod. rufoviride), 397, 401, 473–478, 479, 480, 484.—Odhn., 1905, 360, 362, 364; 1906, 59–66 (syn. Synaptobothrium).
—Pratt, 1902, 889, 906.

caudiporum (Rud., 1819) Luehe, 1901n, 477-478.

conviva Luehe, 1901n, 474, 476 (in Conger conger; Coll. Berlin).

copulans (Linst., 1904) Odhn., 1906, 59-66 (syn. Synaptobothrium copulans

digitatum (Looss, 1899) Luehe, 1901n, 396, 401, 474, 475, 478.

LECITHOCHIRIUM—Continued.

fusiforme Luche, 1901n, 476, 480, 485, fig. 3 (in Conger conger; Coll. Berlin). Odhn., 1905, 349.

grandiporum (Rud., 1819) Luehe, 1901n, 474, 477.

physicon Luehe, 1901n, 478 (in Lophius piscatorius; Triest).

rufoviride (Rud., 1819) Luehe, 1901n, 396, 474–475, 476, 477, 478.—Nicoll, 1907, 72 (in Anguilla vulgaris).—Odhn., 1905, 350, 355; 1906, 63.

LECITHOCLADIUM Luehe, 1901n, 395 (tod. excisum), 401–403, 474, 481, 485.—Cohn, 1902k, 47–68, 1 pl., 9 figs.—Nicoll, 1907, 84.—Odhn., 1905, 355, 356 (type excisum).—Pratt, 1902, 889, 906.

barbatum Cohn, 1902k, 47-54, 55, pl. 3, figs. 1-7 (in Coryphæna).

excisiforme Cohn, 1902k, 54-55, pl. 3, fig. 8 (in Scomber scomber).

excisum (Rud., 1819) Luehe, 1901n, 395, 398, 402, 403, 485, fig. 1.—Cohn, 1902k, 48, 54.—Odhn., 1905, 350.

tornatum (Rud., 1819) Luehe, 1901n, 403, 485, fig. 2.—Cohn, 1902k. 47, 48.

LECITHODENDRIINÆ Looss, 1902m, 815.

LECITHODENDRINÆ Luehe, 1901, 173.—Stoss., 1904, 198.

LECITHODENDRIUM Looss. 1896b, 86 (includes: Dist. glandulosum, D. hirsutum, D. chefrenianum, D. pyramidum, D. obtusum, D. sphærula, D. ascidia Rud., D. ascidia Ben. (=lagena Brand.), D. ascidioides, D. heteroporum (probably); 1898, 453; 1899b, 547, 548, 551, 609-610, type ascidia Ben. = lagena Brand., 611, 612, 613, 617, 618, 619, 621, 622, 625, 636, 718; 1901, 194, 199, 200; 1902m, 768–775, 813, 814, 815, 816, 818, 820, 821, 822, 823, 824, 827, 831, 832, 835.—Braun, 1900f, in 387–391; 1901, 948; 1902b, 150.—Jægers., 1900, 740.—Odhn., 1900, 12.—Luehe, 1899, 535, 536, 537.—Pratt, 1902, 889, 904.—Staff., 1903, 827, 828; 1905, Apr. 11, 684.—Stiles, 1901, 183, 185, 196, 197, 200, 201, 202, 203.— Stoss., 1899, 7, 8.—Ward, 1901, 176.

anticum Staff., 1905, Apr. 11, 693 (t. h. Vespertilio subtilis Say; Canada) (anticus,

foremost in ref. to ovary in front of acetabulum).

ascidia (Ben., 1873 [not Rud., 1819]) Looss, 1899b, 609, or Stoss., 1899, S.—Braun, 1900, 224-225; 1900, 388.—Heymann, 1905, 85.—Kowal., 1902d, 26 (8).—Looss, 1907, Mar. 5, 484 (of Looss, 1899b, 715, syn. of L. granulosum).—Staff., 1905, Apr. 11, 692.

ascidioides (Ben., 1873) Looss, 1899b, 609, or Stoss., 1899, 8 (intestino di diversi chirotteri; Belgium, France).—Braun, 1900, 388.—Staff., 1903, 828.

chefrenianum (Looss, 1896) Looss, 1899b, 716, or Stoss., 1899, 8 (in Rhinopoma microphyllum: Egypt).—Braun, 1900, 225.—Staff., 1905, Apr. 11, 692 (syn. of L. glandulosum).

chilostomum (Mehlis, 1831) Braun, 1900, 220-223, 224, 225, 228, 230.—Staff., 1903, 828; 1905, Apr. 11, 692 (Mehlis of Staff, with Dist. ascidioides Ben., of Staff., 1903, 827, renamed L. posticum).—Stoss., 1904, 1.

claviforme (Brand., 1888) Stoss., 1899, 9 (in Tringa alpina; Halle).

cordiforme Braun, 1900f (23. Juli), 389 (in Molossus sp.; Brazil), 390, or 1900b, 225-

227, 228, 233, pl. 10, figs. 4, 11 (in Molossus sp.).

crassicolle (Rud., 1809) Stoss., 1899, 9 (in Anguis fragilis; Triest, Hameln; Salamandra atra. S. maculosa, Salamandrina perspicillata; Toscana).—Looss, 1902m, 822, 823.—Luehe, 1900, 562–563; 1901, 173.

glandulosum (Looss, 1896) Looss, 1899b, 609, 716-717, or Stoss., 1899, 8 (in Taphosus nudiventris: Egypt).—Braun, 1900, 388; 1900, 225, 226.—Staff., 1905, Apr. 11, 692 (syn. L. chefrenianum).

granulosum Looss, 1907, Mar. 7, 483–484. fig. 4a–b (in Vesperugo kuhli; Cairo, Egypt) (syn. L. ascidia of Looss, 1899b, 715).

heteroporum (Duj., 1845a) Stoss., 1899, 9 (in Vesperugo pipistrellus; Hameln, Rennes).

hirsutum (Looss, 1896) Looss, 1899b, 609, 625, 716, 717, or Stoss., 1899, 9 (intestino del camaleonte; Alessandria).—Braun, 1900, 225; 1900, 388.—Staff., 1905, Apr. 11, 692.

lagena (Brand., 1888) Looss, 1899b, 609, 625, 636, 715-716, 718 (i. e., ascidia Ben.) (type of Lecithodendrium).—Braun, 1900, 225.

macrolaimus (Linst., 1893) Stoss., 1899, 9 (in Vesperugo pipistrellus; Germany).

LECITHODENDRIUM—Continued.

nigrovenosum (Bellingham, 1844) Luehe, 1899, 535.—Rizzo, 1902, 28 (in Tropidonotus natrix).

obtusum (Looss, 1896) Looss, 1899b, 609, or Stoss., 1899, 8 (intestino del camale-onte; Alessandria).—Staff., 1905, Apr. 11, 692.

oviforme (Poir., 1886) Stoss., 1899, 8 (in Nycticebus javanicus).—Staff., 1905, Apr. 11, 692.

posticum Staff., 1905, Apr. 11, 692–693 (t. h. Vespertilio subtilis Say; Canada) (posticus=hinder) (syn. L. chilostomum Mehlis of Staff., 1903, 827) (=Distascidioides Ben. of Staff.).

pyramidum (Looss, 1896) Looss, 1899b, 609, or Stoss., 1899, 8 (in Rhinolophus hippocrepis; Egypt).—Braun, 1900, 225; 1900, 388.

rubellum (Olss., 1868) Stoss., 1899, 9 (in Labrus maculatus).

somateria (Levin., 1881) Stoss., 1899, 8 (in Somateria mollissima; Greenland).—
 Jameson, 1902, 151–161 (Levcithodendrium).—Nicoll, 1906, 149 (Leucithodendrium).—Odhn., 1905, 311 (to Gymnophallus).

sphærula (Looss, 1896) Looss, 1899b, 609, 610, 621, or Stoss., 1899, 8 (in Rhinolophus hippocrepis; Egypt).—Braun, 1900, 225, 227; 1900, 388, 390.—Ward,

1901, 180.

urna Looss, 1907, Mar. 5, 485–486, figs. 5a-b (in Vesperugo kuhli; Cairo, Egypt).

LECITHODENDRUM Pratt, 1902, 903 (misprint for Lecithodendrium).

LECITHODESMUS Braun, 1902, 803 (m. goliath).—Odhn., 1905, 339, 344, 346, 347, 348.

goliath (Ben., 1858) Odhn., 1905, 342, 343, 344–346, 348, pl. 3, figs. 7–9, in Balænoptera rostrata, B. borealis.

LECITHOPHYLLUM Odhn., 1905, 359–360 (tod. botryophorum).

botryophorum (Olss., 1868) Odhn., 1905, 359 (in Argentina silus). LECITHORCHIUM Lander, 1904a, 7, for Lecithochirium.

LEIODERMA Staff., 1904, May 3, 486 (m. furcigerum) (not Leioderma Will-Suhm, 1873, worm) ($\lambda \epsilon i \sigma s$, smooth; $\delta \epsilon \rho \mu \alpha$, skin); 1905, 310.

furcigerum (Olss., 1868) Staff., 1904, 486 (in Pseudopleuronectes americanus, Platysomatichthys hippoglossoides, Hippoglossoides platessoides, Cryptacanthodes maculatus).

LEPIDAPEDON Staff., 1904, May 3, 485 (m. rachion, from intest. of Melanogrammus agglefinus; Canada) (λεπίς, seale; δάπεδον, pavement). See Lepodora. rachion (Cobbold, 1858) Staff., 1904, 485.

LEPIDOPHYLLINÆ Stoss., 1904, 198.

LEPIDOPHYLLUM Odhn., 1902, 68-69 (m. steenstrupi).—Pratt, 1902, 889, 902.

steenstrupi Odhn., 1902, 68-69, fig. 3 (in Anarrhichas minor s. pantherinus; Coll. Copenhagen); 1905, 310 (in A. lupus, Zoarces anguillaris).—Staff., 1904, May 3, 487 (in urinary bladder, Anarrhichas lupus L.; Canada, Zoarces anguillaris Peck; Canada).

LEPOCREADIINÆ Odhn., 1905, 328, 337.

LEPOCREADIUM Stoss., 1904, 200–201 (tod. album).—Odhn., 1905, 328, 336, 337–338. album (Stoss., 1890) Stoss., 1904, 200 (in Cantharus orbicularis; Oblata mela-

nura).—Odhn., 1905, 336, 338.

pegorchis (Stoss., 1901) Stoss., 1904, 200.—Odhn., 1905, 338. LEPODERMA Looss, 1899b, Dec., 589–590, 592, 594, 595, 598, 599, 601 (tod. ramlianum) (orig. species: ramlianum, cirratum, mentulum, lima probably) (τὸ λέπος. scale; τὸ δερμα, skin); 1900, 558; 1900, 604, 607; 1907, Mar. 5, 483.—Braun, 1901i, 56, 58; 1901, 563; 1902b, 37 (syn. of Plagiorchis Luche, 1899, 531).—Luche, 1901, 487.—Stiles, 1901, 189.—Stoss., 1904, 2.

cirratum (Rud., 1802 [cirrhatum]) Looss, 1899b, 590.—Braun, 1902b, 43 (to Plagiorchis).

mentulatum (Rud., 1819) Looss, 1899b, 590.

ramlianum (Looss, 1896) Looss, 1899b, 589.

LEPODERMATINÆ Looss, 1899b, 588, 591, 594; 1901b, 207; 1902m, 839, 841.— Luehe, 1900, 561; 1901, 173.—Odhn., 1902, 40. LEPODORA Odhn., 1905, 332-337, 338 (m. rachiæa=rachion Cobbold).—See Lepidapedon.

rachiæa Odhn., 1905, 328, 332–338, fig. 3, pl. 2, figs. 12–15 (syns.: Dist. rachion Cobbold, D. increscens Olss., pars) (in Gymnocanthus ventralis on west coast of Spitzbergen; Gadus æglefinus, G. morrhua, on west coast of Sweden; Merluccius vulgaris).—Nicoll, 1907, 71, 77–80, pl. 1, figs. 3–4 (in Gadus æglefinus).

LEPTALEA Looss, 1899b, 627–628 (tod. exilis) (not Leptalea Klug, 1839, hymenopt.) (renamed Emoleptalea Looss, 1900, 602) ($\lambda \epsilon \pi \tau \alpha \lambda \acute{\epsilon} o \varepsilon$, thin, weak): 1900, 602.—Braun, 1902b, 69.—Stiles, 1901, 189.

exilis Looss, 1899b, 628, 719–720, figs. 37–38 (in Bagrus bayad; Cairo.)

LEPTOCOTYLE Mont., 1905, 70 (subg. of Pseudocotyle) tod. Pseudocotyle minor Mont.

LEPTOPHYLLUM Cohn, 1902h, 880 (m. stenocotyle).

stenocotyle Cohn, 1902h, 880-882, fig. 4 (in Herpetodryas fuscus; South America); 1903, 37.

LEPTOSOMA Staff.. 1904, May 3, 484 (m. obscurum from Lophius piscatorius: Canada); λεπτός=slender; σῶμα, body) (not Leptosoma Leach. 1819, coleopteron; Risso, 1826, crustacean; Nardo, 1827, fish; Boisd., 1833, lepidopteron). obscurum Staff., 1904, May 3, 484 (obscurus=unknown) (t. h. Lophius piscatorius;

Canada).

LEUCERUTHRUS Marshall & Gilbert, 1905, 477, 481–483, 484 (m. micropteri).

micropteri Marshall & Gilbert, 1905, 481–483, figs. 8–14 (in Micropterus salmoides, M. dolomieu; near Madison, Wis.).

LEUCITHODENDRIUM Shipley & Hornell (see under L. somateriæ), 1904, 77, for Lecithodendrium.

somateriæ (Levin., 1881) Shipley & Hornell, 1904, 77.

LEUCOCHLORIDIA Cobbold, 1876h, 211, plural of Leucochloridium.

LEUCOCHLORIDIUM Carus, 1835a, 86–100 (m. paradoxum).—Ben., 1858a, 1861a, 219.—Braun, 1883a, 56: 1891c, 215: 1892a, 769, 771, 775, 801, 814: 1893a, 855, 884.—Burm., 1856a, 250.—Crep., 1839, 301.—Dies., 1850a, 287, 303 (syn. Vermis dubius helicis putris Rud.): 1855a, 397; 1858d, 276.—Duj., 1845a, 479.—Fil., 1855b, 25.—Goldb., 1855, 16.—Jackson, 1888, 651 (= Dist. macrostomum: in Succinea amphibia), 652.—Leuck., 1879, 99: 1886d, 75.—Looss, 1892, 128: 1894a, 245, 252.—Mont., 1888, 92, 94.—Moul., 1856a, 65, 87.—Pag., 1857, 6, 7, 9, 53.—Spengel, 1905, 258.—Stiles, 1901, 176.—Stiles & Hass., 1898a, 91, 95 (syn. of Urogonimus Mont.).—Tennent, 1906, 647, 650.—Vogt, 1878, 39.—Zeller, (1874), 564–578, pl. 48.

1893: Urogonimus.

macrostomum (Rud., 1803) Poche, 1907, 125.

paradoxum Carus, (1833a); 1835a, 86–100c (in Succinia amphibia, Helix putris; Saxony), pl. 7, figs. 1—; —, 36.—[Ahrens, 1810, 292, pl. 9, figs. 12–19; 1818, 1467].—Bavay, 1902a, 200.—Braun, 1891c, 219; 1892a, 769, 772, 773, 806, 808; 1893a, 830, 855, 863; 1895b, 134.—Cobbold, 1879b, 436.—Crep., 1837, 325; —, 302.—Creutzberg, 1890a, 11.—Dies., 1850a, 303 (syn. Vermis dubius helicis putris Rud.); 1855a, 397 (in Succ. amph.); 1858d, 276–277 (syn. Cerc. exfoliata Moul.) (in Succ. amph.).—Duj., 1845a, 479.—Erc., 1881e, 55; 1882a, 291.—Gamb., 1896a, 65.—Heckert, 1887a, 456–461; 1887b, 603; 1888a, 49–50; 1889a, 66 pp., 4 pls.; 1889b, 357–362; 1890a, 42–43; 1891a, 189.—Hoyle, 1890, 540 (larva of Dist. macrostomum). fig. 4 C-D.—Leuck., 1863a, 521; 1879a, 95; 1886d, 72.—Looss, 1892, 120; 1894a, 236, 262; 1896, 140.—Mont., 1888, 76; 1892. Oct. 7, 187.—Moul., 1856a, 56, 65, 75, 87–90, 180 (Cerc. exfoliata), 181, pl. 5, bis, fig. 8; pl. 6, figs. 13, 14 (in Succ. amph.; Helix. Limax).—Nord., 1840, 548, 592, 631.—Piper, 1851, 313.—Poche, 1907, 125.—Rud., 1819a, 198, 568.—Schultze, (1871), 129.—Sieb., 1835, 77; 1835, 334; 1836, 49; 1853, 425–437, pl. 16 B; 1854, 14.—Steenstrup, 1842, 56; 1842, 103.—Stiles & Hass., 1898a, 95.—Tschudi, 1837, 75.—Vogt., 1852, 183, 191, fig. 66.—Wagener, 1857, 22, 23, pl. 33, figs. 1, 2–4.—Will.-Suhm, 1870, 4.—Zeller, 1874, 366–370; 1874, 20. Oct., 564–578, pl. 48; 1875, Feb., 162–164.

vogtianum Baudon, 1881, 145–147, pl. 5, fig. 5 (in Succinea baudoni Drouët).

LEUKOCHLORIDIUM Sieb., 1853, 425, for Leucochloridium.

LEVCITHODENDRIUM Jameson, 1902, 151, misprint for Lecithodendrium.

somateriæ (Levin., 1881) Jameson, 1902, 151–161, pls. 15–16.—Odhn., 1905, 311 (to Gymnophallus). LEVINSENIA Stoss., 1899, 7, 9-10 (tld. brachysomum by Looss, 1899b, 622, and Luehe, 1899, 538) (not Levinsenia Mesnil, 1897, vermes).—Braun, 1900h, 6; 1900, 234.—Jægers., 1901, 982.—Looss, 1899b, 617, 620; 1902m, 703, 704, 705, 706.—Luehe, 1899, 538; 1900, 508; 1901, 488.—Odhn., 1900, 13; 1905, 314, 317.—Stiles & Hass., 1902d, 19-20 (renamed Levinseniella, type brachysoma); 1905, July, 54.—Ward, 1901, May, 175, 176 (Levinseniella Stiles), 181.

brachysomum (Crep., 1846) Stoss., 1899. 10.—Jægers., 1900. 736, 739 (brachysoma); 1901, 982.—Looss, 1899b, 620; 1902m, 704.—Ward, 1901, 175.

macrophallos (Linst., 1875) Stoss., 1899, 10 (in Actites hypoleucos, Totanus fuscus; Germany).—Looss, 1899b, 620.—Ward, 1901, 175.

opacum (Ward, 1894) Stoss., 1899, 9 (in Amia calva, Cambarus propinguus; Lake St. Clair: Ictalurus punctatus, Perca flavescens).—Looss, 1899b, 620.—Ward,

1901, 175 (type of Microphallus).

pygmæum (Levin., 1881) Stoss., 1899, 9, 10 (in Somateria mollissima; Greenland).— Jægers., 1900c, 732–740, figs. 1–3 (pygmæa); 1901b, 982 (type of Spelotrema).— Looss, 1899b, 620; 1902m, 705, 706, 784.—Nicoll, 1906, 522, 524 (pygmæa).— Odhn., 1900, 19, 20; 1905, 314 (to Spelotrema).—Ward, 1901, 175, 176.

pygmæa similis Jægers., 1900c, 737, figs. 1-3.—Luehe, 1900, 508.—Nicoll, 1906, 522. similis (Jægers., 1900) Nicoll, 1906, 514, 519, 522-525, pl. 13, fig. 8 (syn. Spelo-

trema simile Jugers.) (in Hæmatopus ostralegus, Larus argentatus).

LEVINSENIELLA Stiles & Hass., in Ward, 1901, May, 176, 181, 182, 183, 184 (tod. D. brachysomum) (Levinsenia Stoss., 1899, not Mesnil, 1897, renamed); Stiles & Hass., 1902d, 19–20; 1905c, 54.—Jægers., 1901, 982; 1903a. 14, 15.—Looss, 1902m, 703, 704, 705, 706, 828, 830, 839.—Odhn., 1905, 314, 317, 318.—Pratt, 1902, 889, 902, 903.—Staff., 1905, Apr. 11, 684.

brachysoma (Crep., 1846) Stiles & Hass., 1902d, 20 (type).—Looss, 1902m, 705,

830, 831.

pygmæa (Levin., 1881) Staff., 1903, 824.—Type of Spelotrema 1901.

LINGUATULA Frœlich, 1789a, 148 (m. serrata) [an arachnoid, formerly placed among the trematodes.

caviæ (Bosc, 1811) Blainv., 1828a, 532.

crotali (Humboldt, 1808) Blainv., 1828a, 532.

denticulata Rud., 1805.—Dies., 1850a, 616 (to Pentast.).

integerrima Fredich, 1791a, 104–105, host unknown.—Dies., 1850a, 412 (to Polyst.).—Rud., ——, 93, pl. 2, figs. 9a-f.—Type of Polyst.
 lanceolata (Chabert, 1787a) Blainv., 1828a, 532.—Dies., 1850a, 610) syn. of Pentast.

tænioides Rud.).

pinquicola (Treutler, 1793) Nordmann, 1840, 594.—Dies., 1850a, 410 (to Hexathyridium).

serrata Frœlich, 1789a, 148-150, pl. 4, figs. 14-15 (in hares).—Braun, 1903, 3. ed., 338 (larva of L. rhinaria).—Dies., 1850a, 616 (to Pentast.).

venarum (Treutler, 1793) Nordmann, 1840, 594.—Dies., 1850a, 410 (to Hexathyridium).

LINGULA L. ——.—Rafinesque, 1815, 151 (genus of Fasciolaria).

LINTONIA Mont., 1904, Dec. 1, 117-124 (tod. papillosa); 1905, 70; 1905, 117.

papillosa (Lint., 1898) Mont., 1904, 118-122, 123, pl. 7, figs. 1-7; 1905, 116-123, pl. 7, figs. 1-7 (in Gadus callarias).

LIOCERCA Looss, 1902m, 732 (type bonnieri), 839 (Liopyge, renamed).

1899: Liopyge Looss, 1899b, 642 (tod. bonnieri), not Liopygus Lewis, 1891, insect. bonnieri (Mont., 1893) [Looss, 1902m, 732].

LIOLOPE Cohn, 1902h, 877 (m. copulans).

copulans Cohn, 1902h, 877-880, figs. 1-3 (in Cryptobranchus japonicus; died in Europe); 1903, 39, 40, 41, fig. 4a-c.—Linst., 1904, 254.

LIOPYGE Looss, 1899b, 642 (not Liopygus Lewis, 1891, 385, insect) (tod. bonnieri); λεῖος=smooth, ή πύγη=das Hintere; 1900, 604 (renamed Liocerca Looss, 1902m, 732, 839).—Luehe, 1901, 481, 485.—Pratt, 1902, 889, 901, 905.—Stiles, 1901, 189.

bonnieri (Mont., 1893) Looss, 1899b, 642; 1902m, 732.—Luehe, 1901, 481.

LOBORCHIS Luehe, in Stoss., 1902, 578–582 (tld. by Luehe mutabilis), subf. Allocreadiinæ.

fasciatum (Rud., 1819) Stoss., 1902, 582.

gobii (Stoss., 1883) Stoss., 1902, 582.

labri (Stoss., 1886) Stoss., 1902, 582.

mutabilis Stoss., 1902, 579-582, 1 fig. (in Anguilla vulgaris; Triest); 1903, 373.

LOBOSTOME Blainv., 1824a, 518 (type Dist. laureatum Rud.) ("Ce sont les fascioles cylindriques dont la lèvre de la ventouse antérieure est lobée, comme le dist. laureatum de Rudolphi"). Only the French form of the name was used. See Crossodera.

LOPHOCERCARIA Dies., 1858d, 243 (m. fissicauda).

fissicauda Dies., 1858d, 243 (syn. Cerc. cristata Valette) (in Lymnæus stagnalis; L. palustris, Valvata piscinalis, Paludina impura, Planorbis submarginatus; Berlin & Turin).—Linst., 1878a, 324.

LOPHOCOTYLE Braun, 1896b, 3, 7 (m. cyclophora), Monocotylidæ.—Mont., 1903, 336 (Anisocotylinæ subf.); 1905, 68–70.—Pratt, 1900, 646, 649, 650 (on skin of marine fishes), 655, fig. 19.—St.-Remy, 1898, 523, 540, 543.

cyclophora Braun. 1896b, 3, 7, figs. 1–3 (in Notothenia sp.; Navarin, Puerto Toro).—
 Pratt, 1900, 655, 657, fig. 19.—St.-Remy, 1898, 543–544, fig. 4.

 $\begin{array}{l} {\rm LOPHOTASPIS\ Looss,\ 1901l,\ 7\ Nov.,\ 624-625\ (m.\ L.\ adherens\ Looss = Aspidogaster\ vallei\ (Stoss.,\ 1899));\ 1902m,\ 415,\ 418-430.--Pratt,\ 1902,\ 887,\ 892.} \end{array}$

adhærens Looss, 1901l, 624–625 (in Thalassochelys corticata; Egypt); 1902m, 415, 418 (syn. of L. vallei).

vallei (Stoss., 1899) Looss, 1902m, 415, 418–430, 442, 471, 791 (amphitypie) 868, pl. 21, figs. 1–9.—Odhn., 1902, 44.

LORIFORMIS Geze, 1782a, 41 (= Fasc. intestinalis Linn., Fasc. abdominalis Geze, Ligula), a "Gattung" of Fasciola Geze.

LOXOGENES Staff., 1905, Apr. 11, 683 (m. arcanum); λοξός, bent sideways, oblique; γενάφ, to produce).

arcanum (Nickerson, 1900) Staff., 1905, Apr. 11, 683 (in Rana catesbiana Shaw; Canada) (syns.: Dist. medians Olss. of Staff., 1900, 412; Dist. arcanum Nickerson, 1900, 811; Pleurogenes arcanum of Pratt, 1902, 959; Brandesia medians of Pratt, 1902, 959), 685.

LOXOSOMA Keferstein, 1862a. 131–132 (m. singulare).—Ben., 1869a, 22 (syn. Cyclatella), a bryozoon, see also Braun, 1890a, 518.—Mont., 1888a, 88.—Tasch., 1879, 56.

annelidicola (Ben. & Hesse, 1863) Prouho, 1891, 91–116, pl. 5, figs. 1–16 (in Clyménien de Saint-Vaast, la Hogue).—Sons., 1895, 119.

singulare Keferstein, 1862a, 131–132, pl. 11, fig. 29 ($\lambda o \xi \delta s$, oblique; $\sigma \tilde{\omega} \mu \alpha$, body) (Capitella rubicunda; St.-Vaast).

LYPEROSOMUM Looss, 1899b, 635 (for porrectum, plesiostomum, longicauda, clathratum); $\lambda \dot{\upsilon} \pi \varepsilon \rho o \varepsilon$, thin; tld. longicauda; 1902m, 770; 1907, Feb. 1, 125.—Braun, 1901h, 702; 1901, 946; 1902b, 97, 106 (type longicauda), 119.—Odhn., 1902, 38.—Pratt, 1902, 889, 904.—Stiles, 1901, 190, 191.

corrigia Braun, 1901, 946 (in Tetrao tetrix); 1902b, 111, fig. 68.

lobatum (Rail., 1900) Braun, 1902b, 110, fig. 67.

longicauda (Rud., 1809) Braun, 1902b, 106, 111, fig. 65 (includes: Dist. longicauda Rud., 1809, 372; D. macrourum Rud., 1819a, 98; Linst., 1883, 309; 1886c; Dies., 1850a, 345; D. (Dicrocœlium) macrourum p. p. Stoss., 1892, 13; D. longicauda Mueh., 1896, 248, figs. 2, 9; Dicrocœlium longicauda Rail., 1900, 239).

rudectum Braun, 1901g, 946 (in Ibis cœrulescens Vieill.; Brazil); 1902b, 112, 113, fig. 69.

salebrosum Braun, 1901g, 946 (in Cypselus melba III.; Coll. Vien.); 1902b, 113, fig. 70.

squamatum Linst., 1906, 174, pl. 1, fig. 18 (in Dissura episcopus; Palatupana).

MACRASPIS Olss., 1868 or 1869, 2–3 (m. elegans) [nec McL., ante 1835, coleopteron].—
Braun, 1893a, 879, 886, 887, 888, 890, 891, 894, 896, 898, 917, 918; 1893b, 188.—
Gamb., 1896a, 73.—Looss, 1902m, 428.—Mont., 1888a, 84, 91; 1892, Oct. 7, 196, 198, 213 (gen. of Aspidobothridæ).—Nickerson, 1902, 602, 604, 606, 607, 612, 614, 615, 616, 617.—Odhn., 1902, 42, 43.—Pratt, 1902, 887, 891.—
Tasch., 1879, 255.

[calcarata Spinola, 1835, 130-132, insect.]

MACRASPIS—Continued.

elegans Olss., 1868, 2-3 (in Chimæra monstrosa; Skagerrack).—Braun, 1893a, 875, 898; 1893d, 467 (in Ch. mon.).—Jægers., 1899b, 197-214, figs. 1–9 (anatomy).—Looss, 1902m, 420.—Mont., 1892, Oct. 7, 196; 1893a, 37.—Nickerson, 1902, 614, 618 (in Ch. mon.; Europe).—Tasch., 1879, 255.

MACROCERCUS Hill, 1752a, 8.—Mueller, 1773, 64.—Nitzsch, 1827, 68 (contains Cerc. gyrinus, C. gibba).

MACRODERA Looss, 1899b, 604 (tod. naja) (not Macroderes Westwood, ante 1882, coleopteron) (renamed Saphedera Looss, 1902, 732); $\mu\alpha\kappa\rho\delta$ s, long; $\eta\delta\epsilon\rho\eta$, neck; 1900, 605; 1902m, 732, 839 (syn. of Saphedera).—Luehe, 1900, 557, 561.—Odhn., 1902, 41.—Pratt, 1902, 888, 900.

naja (Rud., 1819) Looss, 1899b, 604; 1901, 658; 1902m, 732.—Luehe, 1900, 556.— Rizzo, 1902, 28 (in Tropidonotus natrix; Catania).

MACRUROCHÆTΛ Costa, (1864), 86-91 (? m. acalepharum).—Braun, 1889a, 364.— Par., 1894, 244.

acalepharum Costa, (1864), 86-91 (larva in Acalephen).—Mont., 1888, 77.—Par., 1894, 165 (syn. of Cerc. setifera Muel.) (in Beroe sp.; Naples), 481.

MALACOBDELLARII Mont., 1888, 94.

MALACOBOTHRIA Braun, 1893b, 188 (for Malacobothrii).

MALACOBOTHRII Burm., 1837, 529; 1856a, 243, 250.—Braun, 1890a, 515; 1893b, 188 (Malacobothria).—Mont., 1888a, 83.—Tasch., 1879, 233.

MALACOCOTYLEA Mont., 1892, Oct. 7, 213 (subo. of Trematoda); 1896, 162.— Braun, 1893a, 890, 891, 892, 895, 898 (Malacotylea), 917; 1893b, 188; 1895b, 136.—Gamb., 1896a, 73.—Looss, 1899b, 543.—Luche, 1901, 488.—MacCallum, 1902, 636.—Maclaren, 1904, 579, 583, 599.—Much., 1898, 18.—Pratt, 1900, 645, 646, 647; 1902, 887, 891, 892 (includes: Paramphistomidæ, Fasciolidæ, Schistosomidæ, Holostomidæ, Gasterostomidæ, Didymozoonidæ, Monostomidæ).—Stiles, 1898a, 27.—Ward, 1903, 864.

MALACOTYLEA Braun, 1893a, 898, for Malacocotylea.

MALAKOKOTYLEA Schneidemuehl, 1896, 295, for Malacocotylea.

MALLEOLUS Ehrenberg, 1838a, 465 (m. furcatus).—Burm., 1856a, 250.—Dies., 1850a, 286, 294–295 (syns.: Vibrio Mueller, Cerc. Nitzsch, Histrionella Bory) (mentions only M. furcatus); 1855a, 379, 395; 1858d, 270, 274.—Fil., 1854a, 6; 1857c, 22.—Goldb., 1855, 16.—Moul., 1856a, 121, 123–124.—Pag., 1857, 4, 5.

furcatus (Nitzsch, 1817) Ehrenberg, 1838a, 465.—Dies., 1850a, 294–295 (syns.: Vibrio malleus Mueller, Zirkelthier Eichorn, Histrionella fissa Bory, Cerc. furcata Nitzsch, Cerc. 6 Baer); 1855a, 395; 1858d, 274 (syns. Cerc. furcata La Valette) (in Lymnæus stagnalis at Halle, Paludina vivipara at Berlin).— Moul., 1856a, 168 (to Cerc.).—Pag., 1857, 6.

MANODISTOMUM Staff., 1905, Apr. 11, 682–683 (m. occultum), $\mu\alpha\nu\dot{\delta}\varsigma = \text{rare}$.

occultum Staff., 1905, Apr. 11, 682-683 (in Diemyctylus viridescens Raf., Rana virescens; Canada) (=Dist. sp. Staff., 1902, 482); occultus=obscure.

MARMOSTOMUM Looss, 1900, 605 (misprint for Harmost. Braun).

MAZOCRAES Hermann, 1782a, 182 (m. alosæ).—Braun, 1890a, 518.—Cerf., 1896, 516.—Dies., 1850a, 421, 422.—Mayer, 1841a, 19.—Tasch., 1879, 239.—See Octostoma, Octobothrium.

1827: Octobothrium Leuck., 1827 or 28, 18 (lanceolata = alosæ, type by inclusion), not Octobothrium Dies. 1850a, cestode.

1829: Octostoma Kuhn, 1829, 358–363 (alosæ, type by inclusion).

1850: Octocotyle Dies., 1850a, 289, 421-422 (lanceolata = alosæ, type by inclusion).

1858: Octoplectanum Dies., 1858e, 382 (Octocotyle 1850 renamed hence alosæ type).

1894: Octocotyte Par., 1894, 594, misprint.

alosæ Hermann, 1782a, 182, pl. 4, figs. 13–14 (in Clupea alosa).—Ben., 1858a, 1861a, 46 (syn. of Octobothrium lanceolatum).—Nord., 1832a, 76 (syn. of Oct. lanc. Leuck., 1828).—St.-Remy, 1898, 545.—Tasch., 1879, 241, 244 (syn. of Oct. lanc.).

MEGACETES Looss, 1899b, 630-631 (tod. triangularis) (not Megacetes Thomas, 1859, coleopteron) (μεγακήτης=grossschlundig); 1900, Dec. 3, 602 (renamed Eumegacetes).—Braun, 1902b, 53, 92 (syn. of Eumeg.).—Odhn., 1902, 38,— Stiles, 1901, 189.

MEGACETES—Continued.

triangularis (Dies., 1850) Looss, 1899b, 596, 631, 725–727, figs. 44–46; 1902m, 819 (=Eumegacetes emendatus Braun, 1901).—Braun, 1901, 568; 1902b, 51, 54, 93, 97 (syn. of Eumeg. em.).—Luehe, 1900, 565.

MEGADISTOMUM Staff., 1904, May 3, 488 (m. longum) ($\mu \dot{\epsilon} \gamma \alpha \varsigma = \text{great}$), 489.

longum (Leidy, 1851) Staff., 1904, May 3, 488 (in Esox masquinongy Mit.; Canada).

MELANOGLENA Eichwald.—Dies., 1850a, 650.

bipunctata Eichwald, ——, 78, pl. 4, fig. 3.—Dies., 1850a, 650 (in aqua salsa, Revaliæ).

MERISTOCOTYLE Rossbach, 1906, 374 (for Merizocotyle).

MERIZOCOTYLE Cerf., 1894k, 936–948 (m. diaphanum) (μερίζω = je partage) Tristomidæ; 1894, 571–572; 1895, 697; 1895m, 130; 1896, 539, 540; 1898b, 329–366, pls. 13–14; 1899a, 448.—Brand., 1898a, 215 (23).—Mont., 1899, 98, 103; 1903, 336 (Anisocotylinæ subf.); 1905, 67, 68, 70.—Pratt, 1900, 646, 650.—Rossbach, 1906, 374 (Meristocotyle).—St.-Remy, 1898, 523, 540, 541–542.

1906: Meristocotyle Rossbach, 1906, 374, misprint.

diaphanum Cerf., 1894k, 936–948, figs. 1–6 (in Raja batis); 1894g, 949–954; 1895m, 129; 1895, 698–699; 1896, 539; 1898b, 329, 330–357, 358, 359, 360, 363, pl. 13, figs. 1, 3, 5, 8–11, pl. 14, figs. 1, 9 (in Raja batis).—Braun, 1896a, 1346.—Mont., 1899, 103.—Pratt, 1900, 655, 657, fig. 17.—St. Remy, 1898, 542, fig. 3.

minus Cerf., 1898b, 330, 357, 361, 363, pl. 13, figs. 2, 4, 6, 7 (in Raja sp.; Roscoff).—St.-Remy, 1898, 543.

MESAULUS Braun, 1902b, 23, 26 (m. grandis).

grandis (Rud., 1819) Braun, 1902b, 24, figs. 16–19 (syns.: Dist. grande Rud., 1819a, 676; Duj., 1845a, 446; Braun, 1901g, 564; Dies., 1850a, 346; Stoss., 1892, 2; D. convolutum (Brand.)).

MESOCOTYLE Par. & Perugia, 1889, Sept., 76–80 (m. squillarum), belongs between Dactylocotyle and Choricotyle.—Braun, 1890a, 546.—Cerf., 1895h, 920; 1896, 515.

squillarum Par. & Perugia, 1889, 76–80, 1 pl., figs. 1a–3a (in Bopyrus squillarum; Triest.).—Braun, 1890a, 549; 1891d, 421.—Cerf., 1898a, 302 (in Bop. sq.)—Stoss., 1898, 12.

MESOGONIMUS Mont., 1888, 15, 92, 105 (tod. Dist. reticulatum Looss, 1885; not Wright, 1879); 1892, Oct. 7, 214 (g. of Distominæ); 1893a, 82, 155–157, 177.— R. Bl., 1891, 609, 610; 1895, 730.—Braun, 1892a, 696, 735; 1893a, 885, 886, 890, 892, 909, 911; 1895b, 138; 1899g, 485; 1900, 25, 31; 1900h, 2, 3, 4, 5; 1902b, 68, 129.—Looss, 1894a, 171, 173, 174; 1896b, 59; 1899b, 536, 538, 539, 542, 585, 649, 650.—Luehe, 1899, 538, 539, 540; 1900, 557.—Moniez, 1896, 89.— Odhn., 1902, 42.—Stiles & Hass., 1898a, 86, 91 (type D. reticulatum Looss).— Stoss., 1892, 4, 5, 31; 1898, 24.

**xqualis* (Duj., 1845) Stoss., 1892, 174 (in Strix perlata, S. flammea; Rennes).—Mont., 1893, 156.

commutatus (Dies., 1858) Sons., 1889, 283; 1891, 201–202 (in Gallus dom., Meleagris gallopavo).—Hass., 1896a, 3 (syns.: Dist. dimorphum Wag., D. commutatum Dies., D. columbæ Mazzanti, Mesogonimus dimorphus (Wag.)) (in Gallus dom.).—Looss, 1894a, 174.—Mont., 1893, 156.—Rail., 1893a, 371 (=M. dimorphus (Wag.) Rail.).—Stoss., 1892, 175 (in Gallus dom., Pisa, Nizza; Meleagris gallopavo, Pisa).

compactus (Cobbold, 1859) Stoss., 1892, 31, 36 (in Mungos fasciatus).—Mont., 1893, 156.

constrictus (Leared, 1862) Mont., 1896, 32 pp., figs. 1–22; 1896, 141–172, pl. 7–8 (in Thalassochelys caretta).—Mingazzini, 1900, 156.—Mont., 1896, 141 pp., pls. 7–8 (in Th. car.).—Type of Hapalotrema, 1899.—See mistroides.

dictyotus Mont., 1893, 156, reticulatum Looss, renamed.

dimorphus (Wagener, 1852) Rail., 1890, 143; 1893a, 371.—Galli-Valerio, 1901c, 364 (in poule).—Hass., 1896a, 3 (syn. of M. commutatus (Dies.)).—Mont., 1893, 83, 156.

heteroclitus (Mol., 1858) Stoss., 1892, 174–175; 1892, 31–32.—Mont., 1893, 156.

heterophyes (Sieb., 1853) Rail., 1890, 143; 1890, 138; 1893, 370.—R. Bl., 1895, 737–739; 1900, 488.—Braun, 1903, 3 ed., 164 (to Cotylogonimus).—Manson, 1903, 3 ed., 664–665.—Mont., 1893, 89, 156.—Stoss., 1892, 31–32 (in Homo; Cairo).—Ward, 1895, 328 (in Homo); 1903, 870 (to Heterophyes).

MESOGONIMUS-Continued.

linguæforme (Dies., 1850) Stoss., 1896, 127–128 (includes Dist. leptosomum Olss., D. caudatum Linst.); 1898, 24 (in Erinaceus europæus; Triest).

lorum (Duj., 1845) Mont., 1893, 156.

marginatus (Rud., 1819) Stoss., 1892, 175 (in Ardea sp.; Brazil).—Mont., 1893, 156. pellucidus (Linst., 1873) Neumann, 1892, 374.—Dolley, 1894a.

pulmonale (Baelz, 1878) Stoss., 1892, 32–33 [type of Paragonimus Braun, 1899]
 (syns.: Dist. pulmonale, D. westermanni, D. ringeri) (in Homo; China, Japan, Korea, Formosa).

pulmonalis (Baelz, 1883) Rail., 1890, 143.—Looss, 1894a, 174.—Mont., 1893, 83, 89, 156.

[reticulatus (Looss, 1885).]

ringeri (Cobbold, 1880) Rail., 1890, 143 [=westermanii].

westermanni (Kerbert, 1878) Rail., 1890, 143; 1893, 369–370.—R. Bl., 1895, 739–740; 1895, 34–39; 1900, 488.—Braun, 1903, 3 ed., 155 (westermani) (to Paragonimus).—Mont., 1896, 168.—Vincent, 1890, 189.—Ward, 1895, 244, 328 (in Homo), 341 (in Canis familiaris); 1903, 867 (westermanii; to Parag.)

MESOMETRA Luehe, 1901d, 57-60 (tod. orbicularis), Monostomidæ.—Looss, 1902m, 442, 780, 813, 824, 838.—Pratt, 1902a, 890, 910.

brachycælia Luehe, 1901d, 51, 55–57, 59, 60, figs. 4–5 (Monost. orbiculare of Par., e. p.) (in Box salpa; Genoa).—Looss, 1902m, 819.

orbicularis (Rud., 1819) Luehe, 1901d, 51–55, 56, 57, 59, 60, figs. 1–3.—Looss, 1902m, 819.

MESOTRETES Braun, 1900a, 229–230 (m. peregrinus).—Pratt, 1902, 889.

peregrinus (Braun, 1900) Braun, 1900, 229–230, pl. 10, fig. 3.
 METASTATICA Leuck., 1889.—Braun, 1893g, 895, 900; 1893b, 188.—Gamb., 1896, 73.—Looss, 1899b, 543.—Ward, 1903, 865.

METORCHIS Looss, 1899b, 564–566 (includes: albidus (tod.), truncatus, complexus, conjunctus, crassiusculus, amphileucus, campula) (μετὰ, behind; ὄρχις, testicle); 1902m, 811.—Braun, 1902b, 5, 7, 9, 11.—Luehe, 1901, 474.—Pratt, 1902, 888, 896.—Rail., 1900, 242.

albidus (Braun, 1893) Looss, 1899b, 565.—Engler, 1904, 186.—Hollack, 1902a, 868.—Luehe, 1901, 171.—Stoss., 1904, 11.

amphileucus (Looss, 1896) Looss, 1899b, 565 (amphielucus); 1902m, 811.—Odhn., 1902, 152.—Stoss., 1904, 11.

campula (Cobbold, 1876) Looss, 1899b, 565.

caruleus Braun, 1902b, 11, fig. 8 (in Cairina moschata; Brazil).—Stoss., 1904, 11. compascua (Kowal., 1898) Kowal., 1902d, (5) 23 (in Anas querquedula; Dublany); 1904a, (8) 23.

complexus (Stiles & Hass., 1894) Looss, 1899b, 565.—Hollack, 1902a, 868.— Stoss., 1904, 11.

conjunctus (Cobbold, 1860) Looss, 1899b, 565.—Stoss, 1904, 11.

crassiusculus (Rud., 1809) Looss, 1899b, 565.—Braun, 1902b, 10.—Engler, 1904, 186.—Hollack, 1902a, 868.—Kowal., 1902d, 23 (5) (corrected to M. xanthosomus (Crep.) Braun, 1902) (in Anas boschas domestica).—Luehe, 1901, 171.—Stoss., 1904, 11.

poturzycensis (Kowal., 1898) Hollack, 1902a, 868.

tener Kowal., 1903, 517, fig. 1 (in Mergus merganser); 1904d, 19 (4) (in M. merg.).—Stoss., 1904, 11.

truncatus (Rud., 1819) Looss, 1899b, 565.—Braun, 1903, 3 ed., 160, fig. 108.—Engler, 1904, 186.—Hollack, 1902a, 868.—Odhn., 1905, 339.—Stoss., 1904, 11.—Ward, 1903, 864; 1903, 704.

xanthosomus (Crep., 1846) Braun, 1902b, 7, 10, figs. 4-7.—Kowal., 1903, 517; 1904d, 23 (8).—Stoss., 1904, 11.

MICROBOTHRIUM Olss., 1869, 3–5 (only positive, hence type species apiculatum), Tristomidae.—Braun, 1890a, 516, 518.—Cerf., 1898b, 362.—Looss, 1894, 9.— Mont., 1888a, 84, 88; 1891, 127, 128; 1903, 336 (Pseudocotyle); 1905, 70.— Pratt, 1900, 649, 655, fig. 16.—St.-Remy, 1891, 213–223 (genital organs).— Tasch., 1879, 236 (syn. of Pseudocotyle); 1879, 49.

MICROBOTHRIUM-Continued.

apiculatum Olss., 1869, 4, fig. 13 (in Acanthias vulgaris; Skagerrack).—Braun, 1891d, 422; 1893b, 176. 178.—Cerf., 1898b, 341, 342.—Looss, 1894, 9.—Mont., 1888a, 88; 1891, 127.—Pratt, 1900, 655, 657, fig. 16.—St.-Remy, 1891, 213–223, 1 fig. (in Ac. vulg.).—Tasch., 1879, 49.

fragile Olss., 1869, 4–5, as doubtful sp. (in Raja batis; Norway).—Mont., 1888a, 88; 1891, 128.—St.-Remy, 1891, 213 (in Raja batis).—Tasch., 1879, 49.

MICROCOTYLE Ben. & Hesse. 1863; 1864, 96. 112 (either donavini or labracis should be type).—Ariola, 1899a: 1899 [in 129–138]; ——, 299.—Braun, 1890a, 414, 416, 426, 428, 445, 451, 454, 458, 468, 472, 477, 484, 485, 486, 489, 490, 494, 498, 499, 500, 511, 517, 523, 540, 546; 1891d, 422; 1893a, 890; 1896b, 7.—Cerf., 1895h, 918; 1896, 514; 1899a, 403.—Cunningham, 1887a, 278.—Gamb., 1896a, 73.—Goto, 1891a, 161, 169, 170, 178, 184, 186, 187; 1891c, 103; 1893a, 798, 799, 800, fig. 1; 1900, in 351–352.—Haswell, 1892a, 459; 1892b, 150; 1893e, 114.—Hoyle, 1890, 537, 539.—Ijima, 1884c, 638, 639.—Jackson, 1888, 642, 644, 645, 646, 647, 648, 654.—Juel, 1889, 33.—Kerbert, 1881a, 573.—Lint., 1901, 414, 451 (sp.); 1905, 335, 370, 385, figs. 147–150 (sp. on Cynoscion regalis. Pomatomus saltatrix).—Lorenz, 1878a, 405–436, pls. 31–33.—Looss, 1885b, 5, 10, 15, 17, 18; 1892, 72.—Maclaren, 1904, 595.—Mont., 1888a, 7, 8, 11, 15, 34, 40, 52, 53, 55, 59, 60, 66, 86, 89, 101; 1892. Oct. 7, 213 (gen. of Microcotylinæ); 1893, 110, 111; 1903, 336 (subf. Microcotylinæ).—Par. & Perugia, 1890, 175–219, 3 pls.; 1890, 13; 1896, in 135–138, 2 figs; 1896 in 4 pp.; ——, 653.—Pratt. 1900, 646, 650, 653, 655, 657, fig. 17, 660.—St.-Remy, 1898, 558–559.—Sons., 1891, 2 pp. (n. sp. in Umbrina cirrhosa).—Stoss., 1898, 14–15.—Tasch., 1879, 257; 1879, 40, 45, 46, 56, 58, 61, 62, 66, 69.

acanthurum Par. & Perugia, 1896, 2 (on Brama rayi; Genova).—Ariola, 1899, 5.— St.-Remy, 1898, 562.

alcedinis Par. & Perugia, 1890, 7: 1890, 744 (in Smaris alcedo; Genova).—Ariola,
 1899, 4.—Braun, 1890a, 418, 541, 549, 552.—Goto, 1894a, 184.—Also reported
 for Mæna trachini, M. vulgaris.

canthari Ben. & Hesse, 1863; 1864, 113–114 (in Cantharus griseus).—Ariola, 1899,
4.—Braun, 1890a, 418, 541, 548, 550.—Lorenz, 1878a, 434.—Stoss., 1898, 15.—
Tasch., 1879, 257 (in C. gr.)—Also reported for Cantharus brama, C. lineatus.

caudata Goto, 1894a, 186–187 (in Sebastes sp.: Mitsugahama).—Ariola, 1899, 4.— Heath, 1902, 117.—St.-Remy, 1898, 559.

chiri Goto, 1894a, 193–194 (in Chirus hexagrammus; Hakodaté).—Ariola, 1899, 4.— Heath, 1902, 117.—St.-Remy, 1898, 561.

chrysophryi (Ben. & Hesse, 1863) Braun, 1890a, 410 (chrysophrii), 418, 541, 548, 550.—Ariola, 1899, 5 (chrysophrii).—Linst., 1878a.—Mont., 1888a. 8 (chrysophrii).—Par. & Perugia, 1890, 8 (chrysophrii); 1894, 137 (in Chrysophrys aurata; Triest. Genova, Venice).—Stoss., 1891, 110; 1898, 15 (in Chr. aur.; Triest).—Tasch., 1879, 257 (in Chr. vulgaris).

donavani Mont., 1888a, 16, for donavini.

donavini Ben. & Hesse, 1863; 1864, 114–115, pl. 12, figs. 1–11 (in Labrus donavini).—
Ariola, 1899, 4 (donavani).—Braun, 1890a, 410, 418, 498, 541, 548, 551.—
Mont., 1888a, 16 (donavani).—Scott. 1905, 116–117, pl. 6, fig. 21 (in Labbergylta Ascan).—Tasch., 1879, 257 (in Lab. don.).

draconis Briot, 1904, Jan. 29, 126–127 (in Trachinus draco L.; Manche, North Sea).
 elegans Goto, 1894a, 188–189 (in Scombrops chilodipteroides; Misaki).—Ariola, 1899, 4.—St.-Remy, 1898, 559–560.

erythrini
Ben. & Hesse, 1863; 1864. 115–116 (in Pagellus crythrinus).—Ariola,
1899, 4.—Braun, 1890a, 418 (erythrinis), 453, 498, 541, 548, 551.—Par. & Perugia, 1890. 8; 1894, 136 (in Box boops; Genova).—St.-Remy, 1898, 546.—Tasch., 1879, 257 (in Pag. ery.).

erythrinis Braun, 1890a, 418 (for erythrini).

fusiformis Goto, 1894a, 192–193, pl. 2, fig. 3, pl. 4, fig. 6, pl. 5, fig. 1 (in Centronotus rubulosus; Mitsugahama).—Ariola, 1899, 4.—St.-Remy, 1898, 561.

hiatulæ Goto, 1899a, 281–282, pl. 21, fig. 29 (in Hiatula onitis; Newport, R. I.).

labracis Ben. & Hesse. 1863; 1864, 112–113, pl. 12, figs. 12–18 (in Labrax lupus).—
Ariola, 1899, 4.—Braun, 1890a, 418, 443, 477, 488, 498, 514, 541, 548, 551.—
Gamb., 1896a, 58, fig. 25c.—Mont., 1888, 8, 16, 66; 1893, 111.—Par. & Perugia, 1890, 8, 12; 1894, 136–137 (in Labrax lupus; Genova).—Scott, 1905, 117, pl. 6, fig. 23.—Stoss., 1898, 15–16.—Tasch., 1879, 257 (in Labrax lup.).

MICROCOTYLE-Continued.

- lichiæ Ariola, 1899, 1-5, pl. 1. figs. 1-5 (in Lichia amia; Genova).
- longicauda Goto. 1899a. 282–283, pl. 21, figs. 30–31 (in Cynoscion regale; Newport, R. I.).—Pratt, 1900, 657, 661, fig. 39.
- mormiri Mont., 1888, 34, for mormyri.
- mormyri Lorenz, 1878a. 425-434 (21-30). pl. 3. figs. 1-6 (on Pagellus mormyrus; Triest).—Ariola, 1899. 4.—Braun. 1890a. 410, 418, 443, 452, 477, 488, 541, 548, 551.—Goto, 1894. 184.—Hoyle. 1890. 539-540. fig. 3c.—Juel. 1899. 36.—Linst., 1889a.—Par. & Perugia, 1890. 745 (in Pagellus mormyrus): 1890. 8; 1894, 137 (Triest; Genova).—Mont.. 1888, 10. 16, 30, 34 (mormiri), 66; 1893, 111.—Stoss., 1898, 16.—Tasch., 1879, 257 (in Pag. morm.; Triest); 1879. 35.—Ziegler, 1883, 546.
- mugilis Vogt. 1878, May 7, 327-332, pl. 14, fig. 3 (in Mugil cephalus: Roscoff).—
 Ariola, 1899, 4.—Braun, 1890a, 514, 541, 548, 551.—Goto. 1894a, 183 (in M. ceph.).—Par. & Perugia, 1890, 4, 8, 12; 1894, 136 (M. ceph.; Genova).—Sons., 1891, 253-254 (mugylis) (in M. ceph.).—Tasch., 1879, 257 (in M. ceph.; Roscoff).
- mugylis Sons., 1891, 253-254, for mugilis.
- pancerii Sons., 1891. 303–304 (in Umbrina cirrhosa; [?Pisa], Italy).—Ariola, 1899,
 5.—Braun, 1893b, 184 (in Umb. cirr.).—Goto, 1894a, 184.
- pomatomi Goto, 1899a. 278–279, pl. 21, fig. 27 (in Pomatomus saltatrix; Newport, R. I.).—Pratt, 1900, 660.
- reticulata Goto, 1894a, 189–191 (in Stromateus argenteus; Mitsugahama).—Ariola, 1899, 5.—St.-Remy, 1898, 560.
- salpæ Par. & Perugia, 1890, 207, pl. 5, fig. 34 (in Box salpa): 1894 (at Genova).— Ariola, 1899, 5.—Goto, 1894a, 184.—Sons., 1891, 262 (in B. sa.).
- sargi Par. & Perugia, 1890, 4, 7, 11 (sargii): 1890, 744 (in Sargus rondeletii; Genova); 1894 (in Sargus rondeletii, S. annularis, S. salviani; Genova).—Ariola, 1899, 4.—Braun, 1890a, 418, 514 (sargii), 541, 549, 552.—Goto, 1894a, 184 (in Sargus annularis, S. rondeletii, S. vulgaris).
- sargii see sargi.
- sciænæ Goto, 1894a, 194–196 (in Sciæna sina; Mogi).—Ariola, 1899, 4.—Heath, 1902, 117.—St.-Remy, 1898, 561–562.
- sebastis Goto, 1894a, 187–188 (in Sebastes sp.: Hakodaté).—Ariola, 1899, 4.—St.-Remy, 1898, 559.
- stenotomi Goto, 1899a, 279–281, pl. 21, fig. 28 (in Stenotomus chrysops; Newport, R. I.).—Pratt, 1900, 660.
- trachini Par. & Perugia, 1889, 744–745 (in Trachinus radiatus; Genova): 1890, 744–745; 1890, 7, 10.—Par., 1894, 136, 594, 595.—Ariola, 1899, 4.—Braun, 1890a, 418, 541, 549, 552.—Briot, 1904, 127 (in Tr. rad.).
- truncata Goto. 1894a, 191–192 (in Pristipoma japonicum: Mitsugahama).—Ariola, 1899, 4.—St.-Remy, 1898, 560.
- MICROCOTYLIDÆ Tasch.. 1879. 235, 237, 255; 1879. 69.—Braun. 1890a, 511, 516, 517, 523, 533, 540, 546.—Ceri.. 1899a, 365, 452.—Hoyle. 1890, 539 (includes: Axine, Microcotyle. Gastrocotyle, Aspidogaster. Cotylaspis. Aspidocotyle).—Jackson, 1888. 654 (includes: Axine. Microcotyle, Aspidogaster. Cotylaspis).— Mont.. 1888. 8. 10. 11, 13, 15. 16, 20. 30. 34, 37. 66. 86. 88. 89, 91, 101, 108; 1892. Oct. 7. 196. 197: 1903. 336 (raised from subf. to fam. rank; subf. Microcotyling (g. Microcotyle); 2. Axining (g. Axine. Pseudaxine. Gastrocotyle).—Par. & Perugia, 1890, 7.
- MICROCOTYLINE Mont., 1892. Oct. 7, 213 (subf. of Polystomidæ); 1903, 336.— Braun, 1893a, 890.—Gamb., 1896, 73.—Pratt, 1900, 646, 653 (includes: Microcotyle, Gastrocotyle, Axine, Pseudaxine).—St.-Remy, 1898, 558.
- MICROLISTRUM Braun, 1901f, 563 (tod. cochleariforme); 1902b, 55.—Pratt. 1902, 889.
 - cochlear (Dies., 1850) Braun, [1901f, 563:] 1902b, 58, 59, fig. 36 (syns.: Dist. cochleariforme p. p. Rud., 1819; Duj., 1845; Stoss., 1892, 37; D. cochlear Dies., 1850; Stoss., 1892; D. diesingi Cobbold, 1861).
 - cochleariforme (Rud., 1819) Braun, [1901f, 563;] 1902b. 56, 58, fig. 35 (syn. Dist. cochleariforme Rud., 1819; Duj., 1845; Dies., 1850; Stoss., 1892).
 - spinetum Braun, 1901f. 563 (in Rhynchops nigra L.: Brazil) (syn. Dist. attenuatum Brems. MS.); 1901, 895; 1902b, 56, 59, 60, figs. 37–39.
 - 8588-No. 37-08-20

- MICROPHALLINÆ Ward, 1901, 185.—Jægers., 1903a, 14.—Odhn., 1905, 318.— Pratt, 1902, 889, 903 (includes: Microphallus, Levinseniella).—Stoss., 1904, 198.
- MICROPHALLUS Ward, 1901, 175–185 (m. opacus); 1902, 17 June, 361; 1903, v. 3, [71–83], 175–187, pl. 26, figs. 1–5,—Jægers., 1903a, 14.—Looss, 1902, 426.—Odhn., 1905, 317.—Pratt, 1902, 889, 903.—Staff., 1903, 824—Stiles & Hass., 1902d, 20.

opacus (Ward, 1894) Ward, 1901, 184.—Staff., 1903, 824.

- MICROPHARYNX Jægers., 1897a, 707–714 (m. parasitica) [an ectoparasitic triclade turbellaria].
 - parasitica Jægers., 1897a, 707-714. figs. 1-3 (on Raja clavata, R. lævis; Kattegat) (? syn. Microbothrium fragile).—Braun, 1900, 70.—Graff, 1904, 457.—Staff., 1904, May 3, 482 (syn. Pseudocotyle fragile Olss.) (on Raja lævis; Canada).
- MICROSCAPHA Looss, 1899b, 668–669, 769 (tod. reticularis) [not Microscapha Le Conte, 1866, coleopteron] ή 6καφη=Kahn, Nachen; 1900, Dec. 3, 602 (renamed Microscaphidium).—Stiles, 1901, 189.
 - linguatula Looss, 1899b, 668, 766-767 (in Chelonia mydas; Egypt), 768, 772, 773,fig. 87.—Braun, 1901b, 38, 54, type of Polyangium 1902.
 - reticularis (Ben., 1859) Looss, 1899b, 668, 763–766 (in Chelonia mydas; Egypt), 767, 768, 770, fig. 81.
 - sagitta Looss, 1899b, 668, 770, 772–773, fig. 88 (in Chelonia mydas; Egypt).— Braun, 1901b, 38, 54, type of Octangium 1902.
- MICROSCAPHIDIINE Looss, 1900, 605; 1902m, 696, 699, 841.—Pratt, 1902, 890, 909 (includes Microscaphidium, Deuterobaris).
- MICROSCAPHIDIUM Looss, 1900, Dec. 3, 602 (Microscapha Looss, 1899, not Le Conte, 1866, renamed, hence type reticulare); 1901, 200; 1902m, 442, 630, 632, 634, 642, 643, 647, 649, 651, 652, 658, 660, 665, 666, 667, 668, 675, 677, 681, 684, 690-691, 694, 695, 696, 698, 699, 805, 811, 824, 879 (type reticulare).—Pratt. 1902a, 890, 909.
 - aberrans Looss, 1902m, 630, 677, 692–693, 697, 698, 879, 881, pl. 28, figs. 106, 117–121 (in Chelone mydas; Egypt).
 - linguatula (Looss, 1899) Looss, 1902m, 688 (to Polyangium as type), 694, 695, 696, 697, 698, 811.
 - parallelum Looss, 1901l, 622 (in Chelone mydas; Egypt); 1902m, 689 (type of Angiodictyum), 690, 696.
 - reticulare (Ben., 1859) Looss, 19011, 622; 1902m, 630, 632, 662, 677, 689, 690, 691–692, 693, 694, 695, 696, 697, 698, 879, pl. 28, figs. 105, 107–116.
 - sagitta (Looss, 1899) Looss, 1901l, 621; 1902m, 627, 685 (to Octangium as type), 694, 695, 696, 811.
- MICROSCAPHINE Looss, 1899b, 667; 1900, 605.
- MIMODISTOMUM Staff., 1904, May 3, 488–489 (m. augusticaudum) (utµo5=imitator).

 augusticaudum Staff., 1904, May 3, 488–489 (in Lota maculosa Le S., Stitzostedion vitreum Mit.; Canada).
- MIRACIDIUM Braun, 1892a, 776, refers to the trematode embryo; not used as generic name.
- MONOCECUM Staff., 1903, 822 (m. baryurum).
 - baryurum Staff., 1903, 822–824. figs. 1–3 (in Necturus maculatus Raf.; near Montreal); 1905, 682.—Linst., 1905, 418, 422 (brachyurum).
 - brachyurum Linst., 1905, 418, 422, misprint for M. baryurum.
- MONOCERCA Wedl, 1861, 478-479 (m. heterobranchi).
 - heterobranchi Wedl, 1861, 478–479, pl. 3, fig. 39 (in Heterobranchus anguillaris; Cairo, Egypt).—Brand., 1888a, 15, 52; 1890a, 578.—Braun, 1892a, 796.—Linst., 1879a.
- MONOCERCE Dies., 1855a, 384; 1858d, 243.—Mont., 1888a, 94.
- MONOCOECUM Linst., 1905, 418, 422, for Monocæcum Staff.
 - brachyurum Linst., 1905, 418, 422, for baryurum.
- MONOCOTYLA Blainv., 1828a, 556, Trematode fam. name. includes: Axine, Branchiobdella, Capsala, Epibdella, Erpobdella, Geobdella, Glossobdella, Hippobdella, Iatrobdella, Ichthyobdella, Malacobdella, Nitzschia, Palæobdella, Pontobdella, Pseudobdella,

MONOCOTYLE Tasch., 1878, 573, 574 (m. myliobatis); 1879, 57, 68; 1879, 236.— Braun, 1890a, 412, 415, 442, 484, 511, 516, 517, 523, 530, 531; 1893a, 890; 1896b, 7.—Cerf., 1894, 947; 1898b, 362.—Gamb., 1896a, 73.—Goto, 1893a, 798; 1894a, 229.—Hoyle, 1890, 539 (only one species, myliobatis) (in Myliobatis aquila).—Jackson, 1888, 653.—Mont., 1888a, 13, 34, 66, 88, 98; 1891, 108; 1892, Oct. 7, 213 (g. of Monocotylidæ); 1903, 335 (subf. Monocotylinæ; f. Monocotylidæ); 1905, 69.—Pratt, 1900a, 646, 650, 656, fig. 21.—St.-Remy, 1898, 523, 541.—Stoss., 1898, 9.

ijimæ Goto, 1894a, 230–232 (in Trygon pastinaca; Aug., Hiroshima.)—Braun, 1896a, 1346.—Cerf., 1895m, 130; 1896, 539, 540; 1898b, 338, 341, 347, 352.—

St.-Remy, 1898, 541.

myliobatis Tasch., 1878, 574 (in Myliobatis aquila; Naples).—Braun, 1890a, 418, 488, 494, 531, 548, 551.—Cerf., 1894, 948.—Hoyle, 1890, 539.—Linst., 1889.—Par. & Perugia, 1890, 6.—Par., 1894, no. 726, 1022, 1059, 1060.—Pratt, 1900a, 656, 657, fig. 21.—Sons., 1890, 173 (in My. aq.).—Stoss., 1898, 9.

MONOCOTYLEA Dies., 1850a, 288, 290, 331, 431 (tribus II of Bdellidea Blainv.), 649 (tribus I of Myzelmintha); 1858, 312, 329.—Braun, 1890a, 515.—Goldb., 1855, 17, 20.—Mol., 1858, 128; 1858, 288.—Mont., 1888, 83, 84.—Stiles & Hass.,

1898a, 90, 91.—Tasch., 1879, 233, 234.—Weinland, 1859, 280.

MONOCOTYLIDÆ Tasch., 1879, 235, 234.—weimand, 1609, 260.

MONOCOTYLIDÆ Tasch., 1879, 235, 236, 238; 1879, 68.—Braun, 1890a, 516, 517, 523, 526, 530; 1893a, 890; 1896b, 7.—Cerf., 1894, 946, 947, 948; 1898b, 361, 362; 1899a, 411.—Goto, 1899, 291.—Hoyle, 1890, 539 (includes: Calicotyle, Pseudocotyle, Monocotyle).—Jackson, 1888, 653 (includes: Calicotyle, Pseudocotyle, Monocotyle).—Mont., 1888, 15, 34, 86, 88, 97, 108; 1891, 108, 127; 1892, Oct. 7, 197, 213 (f. of Eterocotylea) (raised to fam. from subf. by Leuck.; contains: Pseudocotyle, Callycotyle, Monocotyle); 1903, 336 (includes subf.: Monocotylinæ (g. Monocotyle), Pseudocotylinæ (g. Pseudocotyle=Microbothrium), Calycotylinæ (g. Calicotyle), Anisocotylinæ (g. Anoplodiscus, Merizocotyle, Lophocotyle, Dionchus); 1905, 69, 70.—Pratt, 1900a, 646, 649 (includes: Monocotyle, Calicotyle, Lophocotyle, Dionchus, Merizocotyle, (includes: Monocotyle, Calicotyle, Lophocotyle, Dionchus, Merizocotyle, Microbothrium, Pseudocotyle).—St.-Remy, 1891, Dec., 480–481; 1891, Oct., 600.—Stoss., 1898, 8.

MONOCOTYLIDES St.-Remy, 1891, 225-227 (nervous system of); 1891, 480-481; 1891, 600; 1892, 1 Nov., 45-52, 2 figs. (anatomy of).

MONOCOTYLINÆ Gamb., 1896a, 73.—Mont., 1903, 335; 1905, 69.—St.-Remy, 1898, 522, 540.

MONOGENA Mont., 1888, 85, 95, 96 (for Monogenea).

MONOGENÆA Haswell, 1892a, 457, 458, 460; 1893e, 127, 145 (for Monogenea).

MONOGENEA Ben., 1852 (Monogénèse), see Brand., 1891d. 7, 9; 1894a, 305.—
Braun, 1883a, 58; 1890a, 407, 473, 516, 517, 520, 522; 1891d, 421; 1893a, 888; 1893b, 187, 188; 1895b, 131, 136.—Carus, 1863, 477.—Cerf., 1894, 947.—
Dieckhoff, 1891, 245–276, 1 pl.—Gamb., 1896a, 73.—Goto, 1893a, 801.—
Haswell, 1892a, 457, 458, 460 (Monogenæa); 1893e, 127, 145 (Monogenæa); 1892, 150.—Hoyle, 1890, 539 (includes: Tristomeæ, Polystomeæ).—Kath., 1894a, 144.—Kholodk., 1899a, 148–149.—Knoch, 1894a, 11.—Kotoid, 1899, 183.—Looss, 1892a, 116; 1893b, 819.—Mont., 1888, 85, 95, 96 (Monogena); 1903, 334.—Odhn., 1902, 42, 43, 44; 1905, 370.—Pratt, 1900a, 645, 646.—Tasch., 1879, 234. Tasch., 1879, 234.

MONOGENETICA Haswell, 1893e, 144, 145.

MONORCHEIDES Odhn., 1905, 319–320 (m. diplorchis).

diplorchis Odhn., 1905, 318-320, pl. 4, fig. 1 (in Lumprenus medius; off Kings Bay, west side of Spitzbergen.)

MONORCHEIDINÆ Odhn., 1905, 320.

(MONORCHIS) Mont., 1893, 149, subg. of Dist., 150, 151 [type by absolute tautonymy D. monorchis] [not Monorchis Bastian; not Monorchis a Clerc., 1902, cestode].—Braun, 1893a, 894.—Looss, 1902i, 115–122; 1902k; 1903n; 1905h.—Luehe, 1900, 489.—Odhn., 1905, 319, 320.—Stiles & Hass., 1898a, 91, 98 (type by absolute tautonymy monorchis).

pachysomum (Eysenhardt, 1829) Mont., 1893, 151.—Looss, 1902i, 199, type of

Haplosplanchnus.

monorchis (Stoss., 1390) Mont., 1893, 151.

a Cestode species: cirrosa, crassirostris, dujardini, filum, hirsutum, penetrans, pseudofilum.

MONORCHIS (Mont., 1893), type monorchis.—Looss, 1902i; 1902k; 1903n; 1905h. monorchis (Stoss., 1890) Looss, 1902i, 117, fig. 1 (in Cantharus orbicularis, Oblata melanura; Triest).

parvus Looss, 1902i, 118, fig. 2 (in Sargus annularis, S. rondeletii; Triest).

MONOSICHYA Braun, 1890a, 401. See Monossichya.

MONOSSICHYA Cosmovici, 1887, supergeneric (includes Monostomum).—Mont., 1888a, 84, see Monosichya.

MONOSTAMA Mégnin, 1890c, 88, 89, for Monostoma.

MONOSTOMA a Zed., 1800a, 16, 147–160 (Festucaria Schrank, renamed, hence type TOMA ^a Zed., 1800a, 16, 147–160 (Festucaria Schrank, renamed, hence type anatis).—Andral, 1829, 617.—Baillet, 1866b, 107.—Bellingham, 1844a, 335–336.—Ben., 1858a, 1861a, 69; 1870, 363; 1870c, 142.—Ben. & Hesse, 1864, 61.—Blainv., 1828a, 582.—E. Bl., 1847, 303–304.—R. Bl., 1888a, 541, 542.—de Bonis, 1882, 103.—Brand., 1888a, 12; 1892, 504–511 (revision).—Braun, 1883a, 52, 59; 1890a, 514, 515; 1892a, 681, 696, 710, 722, 748, 768, 769, 770, 772; 1893a, 872, 879, 884, 886, 887, 890, 893, 894, 895, 896, 913, 914, 918; 1895b, 121, 128, 134, 137, 154; 1901, 561.—Bremser, 1824, 132.—Burm., 1837, 530.—Carus, 1863, 479.—Cerf., 1894, 946; 1898b, 356 (sp. in Cygnus musicus).—Cohn, 1904, 235, 237.—Cosmovici, 1887a, 128, 129.—Crep., 1829, 1, 49 (Monostomum); 1839, 285.—Dav., 1877, 73.—Deslongchamps, 1824ee, 551, 555.—Dies.. 1834a. 1231: 1850a. 287, 319–331 (syns. Cucullanus Mueller 555.—Dies., 1834a, 1231; 1850a, 287, 319-331 (syns. Cucullanus Mueller ["Cuculanus"], Festuc. Schrank, Fasc. Gœze, Amphist. Rud., Dist. Zed., Monost. Zed.), 400 (syn. of Amphist.), 411 (syn. of Notocotyle), 414 (syn. of Monost. Zed.), 400 (syn. of Amphist.), 411 (syn. of Notocotyle), 414 (syn. of Aspidogaster Baer), 609 (of Numan; syn. of Pentast.); 1855a, 380 (cf. Cheilost.), 384; 1855, 62; 1858e, 312, 324–329.—Duj., 1845a, 342–343.—Dunglison, 1893, 1174.—Eichwald, 1829a, 249.—Eiss, 1838, 23.—Fischer, 1840, 156.—Fischder., 1902a, 6.—Gamb., 1896a, 73.—Goldb., 1855a, 17, 27.—Gunther, 1858, 205.—Hahn & Lefèvre, 1884, 806 (of Rud.).—L'Herminier, 1826, 10.—Hoyle, 1890, 535, 539 (type mutabile).—Jackson, 1888, 644, 654.—Jægers., 1901, 979.—Joy, 1835a, 504.—Kholodk., 1898, 33, 34; 1899a, 153.—Kolenati, 1857, 11.—Kuech., 1855, 180.—Lamarck, 1816, 185–186.—Lamouroux, 1822a, 194.—Leuck., 1863a, 61, 66, 451, 524, 632–633; 1879, 78; 1886d, 34, 59, 64.—Linst., 1901, 2.—Looss, 1885b, 56; 1896b, 147; 1899b, 658; 1901, 192, 193 (ocreatum=Dist. lorum Meln.; verrucosum to Notocotyle; bombynæ=M. ellipticum=Dist. variegatum; mutabile=type of Cyclocelum; prismaticum=type by Dist. variegatum; mutabile=type of Cyclocelum; prismaticum=type by elimination); 1902m, 562, 564, 566, 568 (prismaticum type after Looss, 1901, but a distome after Mont., 1892; mutabile type of Cyclocelum Brand. in 1892; type of Monost. by elimination after Luehe, 1901; see also Looss, 1902m, 1892; type of Monost. by elimination after Luche, 1901; see also Looss, 1902m, 707; mutabile might be type, Looss, 1902m, 719), 700, 701, 702, 703, 707, 710, 719, 720, 721, 728, 729, 730, 746, 812, 813, 842.—Luche, 1901, 174, 175; 1901, 481.—Mayer, 1841, 4.—Moniez, [1899a], 159.—Mont., 1888, 7, 11, 64, 69 (Monostumum), 71, 73, 83, 84, 93, 106; 1892c, 514–534, figs. 1–22 (sp. in Box salpa); 1892d, 23 pp., 22 figs.; 1892, 31; 1892, Oct. 7, 183, 189, 199 (syn. of Aspidogaster) (of Rud.), 214 (g. of Monostomide); 1892, 709 (Monostonum); 1893, 15, 25, 27, 105, 115.—Moul., 1856a, 12, 15.—Nord., 1840, 614, 622–623.—Olfers, 1816, 22, 48.—Par., 1887, 327.—Pratt, 1900a, 645; 1902a, 890.—Rail., 1891, 26.—Ratzel, 1868, in 19 pp.—Rud., 1809a, 5, 20–21, 36–37, 325, pl. 12, fig. 5; 1819a, 82, 340, 583.—Schneidemuehl, 1896, 295.—Schneider, 1866, 334.—Sieb., 1854, 20, 29.—Stiles, 1902s, 28.—Stoss., 1898, 63.—Tasch., 1879, 232, 233, 258 (syn. of Aspidogaster Baer); 1879, 611.—Villot, 1878, 19.—Vogt,

^a Monostoma represents a complicated case, so far as its type is concerned, and well represents "a ship without a rudder."

There can be no question but that Monostoma was a deliberate renaming of Festucaria 1788, the name being changed for purely subjective reasons. The fact that Zeder did not specifically mention the original species of Festucaria does not seem to bear upon the question, for he gave "die mir bekannten Arten," and he refers to the original reference of Festucaria; thus the case is similar to Tænia Linn., 1758a, renamed Alyselminthus Zed., 1800. In our opinion the type must be the same as that of Festucaria, and this has been fixed by elimination to Festucaria anatis. The question whether or not this is a species inquirenda can not, so far as we see, come up at present. A deliberate renaming deserves to be treated in only one way and, logically, we see

only one way to treat it. Hoyle (in 1890, probably 1888, possibly earlier) designated mutabile as type of Monostoma. Looss (1901) by elimination designated prismaticum as type.

Had Monostoma not been a case of renaming, we should accept Hoyle's designation.

1878, 9, 10.—Wagener, 1854, 10-11; 1883, 122.—Wallenstedt, 1847, 7.—Walter, 1892, 11 July, 248-250 (in intestine of turtle).—Wedl, 1855, 380, pl. 2a, fig. 14 (sp.).

1815: Monostomeus Rafinesque, 1815, 151, Festucaria renamed.

1829: Monostomum Crep., 1829, 1, 49, for Monostoma.

1888: Monostumum Mont., 1888a, 69, misprint.

1891: Monostoum Brand., 1891d, 19, misprint.

1892: Monostonum Mont., 1892, 709, misprint. 1904: Monostromum Linst., 1904, 254, misprint.

acreatum Stoss., 1892, 18, misprint for ocreatum.

aculeatum Linst., 1879b, 338 (in Testudo græca; loc. not given); 1889.—Brand., 1892, 509.—Braun, 1893a, 915; 1899, 630; 1901a, 13, 14.—Looss, 1899b, 567. Luehe, 1899, 528.—Mont., 1892, 686, 687, 697, 705, 715.—Stoss., 1895, 224 (=Dist. linstowi Stoss.); 1898, 43 (in Test. gr.; Triest).

affine Leidy, 1858, 110–111 (t. h. Fiber zibethicus; U. S. A.); 1904a, 111.—Brand., 1892, 510.—Braun, 1893a, 875, 915; 1893d, 466 (in F. zib.); 1901e, 344.—Dies., 1859c, 425.—Mont., 1892, 685, 696, 697, 698, 703, 711.

album Kuhl & van Hasselt, 1822a; 1824a, 311 (in Chelonia midas; Iles des Cocctiers).—Braun, 1899b, 721, 723-724; 1901a, 38, 44, 45-48, 52.—Crep., 1846, 146.—Dies., 1850a, 325 (=M. trigonocephalum Rud.).—Looss, 1899b, 667, 756, 762 (type of Cricocephalus); 1902m, 531.—Shipley, 1900, 532.

alveatum Mehlis, in Crep., 1846, 142 (in Anas fusca, A. glacialis), 143 (in A. leucophthalma, A. mollissima, A. penelope), 144 (in Cygnus musicus).— Brand., 1892, 508.—Braun, 1893a, 879.—Cobbold, 1860a, 40.—Cohn. 1904, 229 (of Much., 1898) (syn. of M. alveiforme Cohn), 230.—Crep., 1846, 141, 142–144; 1851, 291.—Dies., 1850a, 331 (in Anas fusca, A. glacialis, A. marila, A. mulissima, A. musicus, A. penelope); 1858e, 328 (in Anas anser ferus).—Looss, 1899b, 662.—Mont., 1892, 38, 39, 41 (to Notocotyle); 1892, 706, 709.—Much., 1898, 19 (in Fuligula marila; Pillau); 1898, 31, 101–102, fig. 3.— Reported also for Anas bernicla, A. cinereus.

alveiforme Cohn, 1904, 230 (alveatum Mehlis of Mueh., 1898, renamed).

amiuri Staff., 1900, 402-403, fig. 2 (in swim bladder of Amiurus nebulosus); 1904, May 3, 495 (Canada).

angustum Schlotthauber, 1860, 129 (int. of Scolopax gallinago).

arcuatum Brand., 1892b, 507 (to Cyclocœlum) (in aquatic birds), 508.—Braun, 1893a, 915; 1899, 467.—Looss, 1899b, 660, 661.—Stoss., 1902, 5, 6, 21, to Cyclocœlum (syns. Monost. mutabile of Sieb., 1835, 49, pl. 1; Dies., 1858, 325; Dav., 1877a, lxxiii; Mueh., 1898, 31) (in Anas clangula; Berlin, supposed to be based on Brandes' orig. material) (also in Anas dom.; Berlin). armatum Mayer, 1841a, 4, Echinorhynchus renamed.

asperum Nitzsch, in Crep., 1849a, 71 (in Anas fuligula); in Giebel, 1857, 265 (in Mergus albellus, Anas clangula).—Brand., 1892, 507.—Braun, 1893a, 871.

asperum Vaillant, 1863, 347-348 (in Siren lacertina).—Mont., 1892, 715.

attenuatum Rud., 1809a, 328–329 (t. h. Scolopax gallinago, ceca; Greifswald) (to Monost. (Monost.)); 1819a, 84.—Baillet, 1866b, 107.—Bellingham, 1844a, 336.— Brand., 392, 508.—Braun, 1891, 90; 1891d, 434 (in Anas boschas); 1893a, 874, 879, 916.—Cobbold, 1860a, 40.—Crep., 1839, 285; 1846, 141–142, 144–145; 1849, 1, 71; 1851, 1, 290.—Dav., 1877a, lxxiii.—Dies., 1850a, 322–323; 1858e, 325 (in Anas tadorna, A. penelope, A. albifrons, A. marila).—Duj., 1845a, 350.—Hass., 1896a, 3 (syn. of Notocotyle verrucosum (Frælich)).—Looss, 1896, 192; 1899b, 662, 663.—Luehe, 1898, 625.—Mol., 1859, 824–825 (in Anas clangula, A. clypeata, A. ferina, A. fuligula, A. fusca, A. musicus, Anser dom., Mergus merganser, M. serrator).—Mont., 1892, 38, 39, 41 (syn. of Noctocotyle verrucosum Fredich); 1892, 706, 709.—Mueh., 1898, 31.—Nicoll, 1906, 515 (in Harelda glacialis).—Olfers, 1816, 48.—Rail., 1893a, 338 (of Mol.=M. mutabile Zed.), 340 (of Rud.=Notocotyle verrucosum).—Schlotthauber, 1860, 129.—Reported also for Anas glacialis, Anser cinereus, Cygnus musicus, Fuligula cristata, F. ferina, F. marila.

bijugum Miescher (1838a), pp. 28, pl. 1; 1838b, 154–158.—Baird, 1853a, 45.—Ben., 1858a, 1861a, 179, 188, 198, 200, 201.—Brand., 1891b, 265.—Dies., 1850a, 321 (syn. of M. faba Bremser).—Nord., 1840, 616, 625 (syn. M. faba).—

Sieb., 1839, 160–162.—Tasch., 1879, 608 (syn. of M. faba).

bipartitum Wedl, 1855, 378–380, 393, pl. 1a, figs. 11–13 (t. h. Thynnus vulgaris).—

Ariola, 1902, 100, 101 (of Wagener, form 2, syn. of Didymocystis reniformis), 103 (of Wedl, and of Wagener, form 1, syn. of Didymost. bipartitum, type), 105 of Wagener, form 3, syn. of Didymocistis wedli).—Braun, 1892a, 773, 660; 1893a, 894.—Darr, 1902, 661.—Dies., 1858e, 327 (in Thyn. vulg.); 1859c, 426.—Gamb., 1896a, 71.—Leuck., 1863a, 453.—Mont., 1888a, 9, 93; 1892, 714.—Par. & Perugia, 1889 or 1890, 746 (syn. of Didymozoon thynni Tasch.); 1893, 2.—Stoss., 1898, 62.—Tasch., 1879, 72; 1879, 606, 611, 612 (syn. of Didymozoon thynni Tasch.).—Wagener, 1858, 250, 252–256, pl. 10, figs. 1–10 (in Thynnus yulgaris; Niceæ). (in Thynnus vulgaris; Niceæ).

blainvillei Cobbold, 1860a, 39 (Monost. delphini Blainv., renamed) (in Delphinus dalei).—Mont., 1892, 712.

bombynæ Zed., 1800a, xvi, 151, 160 (t. h. Rana bombya, lungs; Europe); 1803a, 190.—Baird, 1853a, 53 (=Dist. variegatum).—Dies., 1850a, 322 (syn. of M. ellipticum Rud.).—Looss, 1894a, 71 (syn. of Dist. variegatum Rud.); 1901, 192.—Rud., 1809a, 333 (=M. ellipticum).

braunii Cobbold, 1860a, 43 (M. murænulæ Rud.) (in Coregonus murænula).

caouanæ Kollar, in Braun, 1901b, 23 (syn. of Enodiotrema megachondrum) (in Thalassochelys caouana=T. caretta).

capitellatum Rud., 1819a, 83, 343 (to Monost. (Monost.)) (t. h. Sparus salpa; Naples). -Barbagallo & Drago, 1903, 411 (in Box salpa; Catania).—Brand., 1892, 509.— Braun, 1891d, 421: 1892a, 765, 766, 784, 786: 1893a, 915; 1893b, 178, 179.—Carus, 1884, v. 1, 112, 122.—Cobbold, 1860a, 42 (in Box salpa, Scomber scombrus).—Creutzburg, 1890a, 21.—Dies., 1850a, 326; 1858e, 327 (in B. sal.). brus).—Creutzburg, 1890a, 21.—Dies., 1850a, 326; 1858e, 327 (in B. sal.).—Duj., 1845a. 360–361.—Fil., 1855b, 25.—Florance, 1866a. 5.—Kroyer, 1838–40a, 595 (in Scom. scom.).—Leuck., 1863, 491, fig. 166; 1879, 39, fig. 16; 69; 1886d, 30, 69, fig. 16.—Looss, 1899b, 669; 1902m, 730 (as possible type of Monost., if Rud., 1819, were taken instead of Linn., 1758).—Luehe, 1901, 59, 60.—Mont., 1892. 4–12; 1892, 16 (of Carus, 1884, 112, partim, Par., Setti, Stoss. = M. stossichianum Mont.): 1892, 685, 686, 687, 688, 689, 694, 695, 696, 697, 698, 699, 700, 702, 703, 704, 705, 716; 1893, 15, 24, 83, 84, 104, 115, 116.—Par., 1886, 5, 7; 1887, 489.—Setti, 1891, 4 (= M. stossichianum teste Mont.).—Stoss., 1883, 112, pl. 2, fig. 9 (= M. stossichianum teste Mont.); 1898, 64.—Wagener, 1857, 26, 27, 45, 101, pl. 19, fig. 5.—Will.-Suhm, 1873, 342.

Caryophyllinum (Rud., 1802) Zed., 1803a, 189; to Monost. (Hypost.) by Rud., 1809a, 323.—Baillet, 1866b, 108.—Blainv.. 1828a, [probably type of Hypost.].—Brand., 1892, 508.—Braun, 1893a, 915.—Bremser, 1824, 132, pl. 8, figs. 1-2.—Cobbold, 1858b, 156, pl. 31, figs. 2, 3 (in Gasterosteus aculeatus).—Crep., 1825a, 80; 1839, 285.—Dav., 1877a, lxxiii.—Dies.. 1850a, 328 (in Gast. acul., Gyphiæ; Anas boschas dom., Berlin).—Duj., 1845a, 360.—Gurlt, 1838, 229.—Kroyer, 1838—40a, 187 (in Gast. acul.).—Lamarck, 1816b, 186.—Mont., 1892, 717. (caryophyllum).—Nord., 1840, 623.—Olfers, 1816, 48.—Bail., 1893a, 211, 1893a. 717 (caryophyllum).—Nord., 1840, 623.—Olfers, 1816, 48.—Rail., 1893a, 339.—Risso, 1826, 262.—Rud., 1809a, 325–326, pl. 9, fig. 5; 1819a, 82.—Sieb., 1839, 169.—Verrill, 1870, 179 (of Bremser).

caryophyllum Mont., 1892, 717 (for caryophyllinum).

cercatum Ben., 1858a, 1861a, 179 [possibly lapsus for —?—].

cochleariforme Rud., 1809a, 326-327, 410 (Festuc. cyprinacea Schrank, 1790, renamed) to (Hypost.) (in Cyprinus barbus); 1819a, 82-83.—Brand., 1892, 507.—Braun, 1893a, 916.—Dies., 1850a, 329 (syns.: Festuc. cyp., Dist. punctatum Zed.) (in Barbus communis).—Duj., 1845a, 362.—Kroyer, 1846–53a, 333 (in Barbus fluviatilis Ag.).—Lamarck, 1816b, 187.—Mont., 1892, 685, 696, 698, 704, 716, 717.—Nord., 1840, 624 (syn. Festuc. cyp.).—Olfers, 1816, 48.—Stoss., 1890, 132.

conicum Zed., 1803a, 188 (Festucaria cervi Zed., 1790 renamed).—Dies., 1836, 247; 1850a, 401 (to Amphist.).—Fischder., 1901, 368 (syn. of Paramphist. cervi, type); 1902a, 11 (syn. of P. cervi); 1903h, 540.—Nitzsch, 1819, 398 (to Amphist.).—Nord., 1840, 627 (to Amphist.).—Rud., 1809a, 349 to Amphist.. 350.—Stiles, 1898a, 64.—Ward, 1895, 256 (to Amphist.), 332 (in Bos

taurus), 335 (in Ovis aries).

constrictum Dies., 1850a, 322 (t. h. Abramis brama; eye); 1855, 62, pl. 2, figs. 3–5: 1858e, 325.—R. Bl., 1888a, 542.—Brand., 1892, 511, to Diplostomulum.— Braun, 1893a, 871.—Kroyer, 1852–53a, 1223 (in Abramis brama L.).—Moniez, 1896, 154.—Mont., 1888a, 7; 1892, 716, 717.—Sramek, 1901, 108 (see Dist. retroconstrictum).

- cornu (Zed., 1800) Rud., 1819a, 85, 90, 345–346 (in Ardea cinerea, A. nycticorax).— Brand., 1892, 507.—Braun, 1893a, 916.—Dies., 1850a, 327.—Duj., 1845a, 349–350.—Mont., 1892, 706.
- cotti Linst., 1889a, 80, based on Zschokke, 1884, 204–205 (in Cottus gobio; Lake Leman), ? syn. M. marænulæ.—Braun, 1893a, 871.—Mont., 1892, 685, 716, 717.
- crenulatum Rud., 1809a, 328, to Monost. (Monost.) (t. h. Motacilla phænicurus;
 Greifswald); 1819a, 84.—Brand., 1892, 510.—Braun, 1893a, 915.—Dies., 1850a,
 327.—Duj., 1845a, 348.—Lamarck, 1816b, 187.—Mont., 1892, 713, 714 (in
 Lusciola phænicurus).—Nord., 1840, 624 (in Mot. phæn.).—Olfers, 1816, 48.—
 Stoss., 1898, 23.
- crucibulum Rud., 1819a, 83, 342–343 to (Monost.) (t. h. Muræna conger, M. cassini (M. myroides); Naples).—Dies., 1850a, 321–322; 1859c, 425, 437 (to Gasterost.).—Duj., 1845a, 363–364.—Kroyer, 1846–53a, 615 (in Anguilla conger L.).—Odhn., 1905, 305 (to Prosorhynchus).—Stoss., 1898, 60.—Tennent, 1906, 639, 640 (to Gasterost.).
- cucumerinum (Rud., 1809) Braun, 1899f, 468.—Stoss., 1902, 9, 32 (to Typhlocœlum).
 cymbium Dies., 1850a, 320 (t. h. Himantopus wilsonii; Caiçaræ, Brazil); 1855a, 63, pl. 2, figs. 1–2; 1858e, 324.—Brand., 1892, 509.—Braun, 1892a, 642, 700; 1893a, 873, 915; 1893b, 179; 1901b, 48.—Looss, 1902m, 701.—Mont., 1892, 27 (syn. of M. flavum); 1892e, 683–718, figs. 1, 2, 4, 6, 8, 9, 10, 11; 1892, 707 (syn. of M. flavum Mehlis); 1892, Oct. 7, 183; 1892f, 47 pp., 11 figs.; 1893, 19, 83, 84, 115, 116, 117.—Stoss., 1902, 4, 6, 7 (cyumbium), 27, 28 (to Hæmatotrephus).
- cyprinæ Leach in Johnston, 1865a, 35 (on Cyprina islandica; Plymouth), as syn. of Malacobdella grossa.

cyumbium Stoss., 1902, 7, for cymbium.

- delicatulum Dies., 1850a, 325 (Dist. testudinis Rud., 1819a, 121, renamed) (t. h. Emys europæa, Halichelys atra; Mus. Vicn.).—Brand., 1892, 510.—Braun, 1893a, 915; 1899, 628; 1899b, 715, 721, 722; 1901b, 54.—Mont., 1892, 685, 714.—Also reported for Emys lutraria.
- delphini Dies., 1850a, 330 (in Delphinus dalei) based on Blainv., 1825a, 141 and 1825b, 212–214 (on Delphinus sp.; Havre).—Ben., 1870, 358.—Braun, 1893a, 870.—Cobbold, 1879b, 421.—Mont., 1892, 711.—See also M. blainvillei.
- dubium Cobbold, 1858b, 156, pl. 31, figs. 4-5 (t. h. Gasterosteus spinachia); 1879b,
 463.—Brand., 1892, 511 (to Monostomulum).—Braun, 1893a, 871.—Mont.,
 1892, 717.
- dujonii Braun, 1893a, 917, for dujonis.
- dujonis Leuck., (1874), 419 (in Halicore dujong), teste Mont., 1892, 712.
- echinatum Linst., 1878, 223–224, fig. 6 (in Pandion haliaëtos); 1889.—Brand., 1892, 509.—Braun, 1892a, 570, 586; 1893a, 915.—Mont., 1892, 685, 686, 687, 694, 697, 698, 699, 702, 705, 713, 714.
- echinostomum Dies., 1850a, 326 (t. h. Cathartes aura, Sula fusca; Brazil) (includes Dist. planicolle Rud., 1819a, 686, from Pelecanus sula; Brazil); 1855, 63, pl. 2, figs. 14–16; 1858e, 327.—Brand., 1892, 506.—Braun, 1892a, 584; 1901, 567; 1902b, 28 (syn. of Anoictost. (?) planicolle).—Mont., 1888, 8, 14; 1892, 706, 710.
- elaphi (Gmelin, 1790) Zed., 1800a, xvi, 150.—Fischder., 1902a, 11 (syn. of Paramphist. cervi); 1903h, 504, 506.—Rud., 1809a, 350 (—Amphist. conicum).—Stiles, 1898a, 64.

ellicticum Mont., 1892, 715 (for ellipticum).

- ellipticum Rud., 1809a, 333 (Mon. bombynæ Zed., 1800, renamed) to (Monost.); 1819a, 84–85 (in Bufo igneus, Berlin; B. cinereus), 344–345.—Baird, 1853a, 53 (— Dist. variegatum Crep.).—Blainv., 1828a, 582.—Brand., 1892, 507.—Braun, 1893a, 876, 881, 915.—Bremser, 1824c, pl. 8, figs. 12–14.—Cobbold, 1860a, 41.—Dies., 1850a, 322 (in Bombinator igneus, Berlin; Phryne vulgaris) (syn. M. bombynæ Zed.), 355.—Duj., 1845a, 359.—Eichwald, 1829a, 249.—Looss, 1894a, 71, 72, 80 (syn. of Dist. varieg. Rud.); 1899b, 660; 1901, 192 (M. bombynæ).—Mont., 1892, 715 (ellicticum).—Nord., 1840, 625.—Olfers, 1816, 48.—Par., 1894, 168.—Schlotthauber, 1860, 129.—Sieb., 1835, 56.—Stoss., 1889, 62 (— Dist. varieg.); 1898, 35; 1902, 5.—Reported for Bufo igneus, B. vulgaris, Rana bombina, R. esculenta.
- expansum Crep., 1842, 327 (t. h. Aquila haliaëtos).—Brand., 1892, 508.—Braun, 1893a, 915.—Dies., 1850a, 321 (in Falco haliaëtos; Gryphiæ).—Duj., 1845a, 345–346.—Jægers., 1901b, 979–983, 1 fig. (to Tocotrema); 1902a, 356–357; 1903a, 1.—Mont., 1895, 685, 686, 694, 696, 697, 698, 699, 700, 703, 713, 714.

faba Bremser in Schmalz, 1831, 11-16, pl. 6, figs. 1-9 (in Parus major, Silvia sibilatrix, Motacilla boarula); 1839b, 1-8, pl. 1, figs. 1-2.—Baird, 1853, 45.— Brand., 1892, 509.—Braun, 1892a, 567, 642, 747, 751, 784; 1893a, 877, 879, 894, 915; 1893d, 468.—Cobbold, 1860a, 38, to Wedlia.—Crep., 1839b, 1-8, pl. 1, figs. 1-2; 1839a, 285.—Dav., 1877a, lxxiii.—Dies., 1850a, 320-321 (syns. M. bijugum Miescher, Globularia Rolando) (in Fringilla spinus, F. canaria, F. domestica, Motacilla boarula, Parus major, Sturnus vulgaris, Sylvia sibilatrix, S. trochilus).—Duj., 1845a, 346-348.—Leuck., 1863, 453.—Linst., 1904, 254 (Monostromum).—Looss, 1893b, 810.—Mont., 1888a, 9, 18, 93; 1892, 697, 699, 704, 713, 714.—Mueh., 1898, 31.—Nord., 1840, 625 (syn. M. bijugum).—Par., 1887, 327-329, pl. 6, fig., 36.—Rail., 1898, Oct., 628-629 (in Garrulus glanda-1887, 327-329, pl. 6, fig. 36.—Rail., 1898, Oct., 628-629 (in Garrulus glandarius).—Sieb., 1839, 160, 161.—Tasch., 1879, 608 (syn. M. bijugum Miescher).— Reported also for Cyanocitta cristata, Emberiza cirlus, Ficedula sibilatrix, F. trochilus L., Saxicola enanthe, Motacilla alba, Passer domesticus, Sylvia sp.

filarinum (Ben., 1858) Dies., 1859c, 426-427, as sp. inq.—Mont., 1893, 137.—

Reported for Sciæna umbra.

filicolle Rud., 1819a, 85-86, 347-348 (t. h. Brama raji (Sparus raji); Naples) to (Monost.).—Ariola, 1906, 184; 1906, v. 30, 185-186.—Ben., 1858a, 1861a, 104, 105 (to Dist.).—Braun, 1892a, 572.—Cobbold, 1860a, 31 (type of Köllikeria).—Dies., 1850a, 359 (syn. of Dist. okenii).—Duj., 1845a, 361.—Kroyer, 1838-40a, 219, 594 (in Pagellus centrodontus Cuv., Brama raji Bl.).—Mont., 1893, 150.— Stiles & Hass., 1898a, 91, 98.—Tasch., 1879, 608.

filigerum Rud.—Risso, 1826, 262 (de la castagnolle).

filum Duj., 1845a, 362 (t. h. Scomber scombrus).—Ben., 1870c, 140, 141,142.— Braun, 1892a, 660, 784, 786.—Dies., 1850a, 327; 1858e, 328 (in Exocœtus exsiliens).—Kroyer, 1838–40a, 595 (in Sc. sc.).—Leuck., 1863a, 490.—Mont., Par., 1894, 168.—Par. & Perugia, 1894, 121–122 (in Ex. evolans, E. exiliens).—Par., 1894, 168.—Par. & Perugia, 1893, 1, 2, 3, of Wagener (syn. of Didymozoon exocœti).—Wagener, 1854, 10; 1857, 25, 52.—Reported also for Brama

flavum Mehlis, 1831, 172 (t. h. Anas mollissima, A. fusca, A. marila, A. fuligula).— Baillet, 1866b, 96, 107.—Ben., 1858a, 1861a, 171.—Brand., 1892, 507.— Braun, 1892a, 587, 673, 764, 768, 784, 785, 786, 788, 805; 1893a, 858, 866, 873, 876, 915; 1895b, 17; 1899, 467; 1902b, 20.—Cohn, 1904, 229 (syns.: Typhlocœlum Stoss., Cyclocœlum Brand.).—Crep., 1837, 314, 324.—Dies., 1850a, 324; 1858d, 245; 1858e, 325–326 (adult in Anas mollissima, A. fusca, A. marila, 1858d, 245; 1858e, 325–326 (adult in Anas mollissima, A. fusca, A. marila, A. fuliginosa, Mergus albellus, M. serrator, young in Fringilla dom, larva in Planorbis corneus).—Duj., 1845a, 355.—Gamb., 1896a, 72.—Leuck., 1863a, 491, 515.—Levin., 1881a, 57.—Linst., 1873, 1 (young, Glenocerc. flava); 1903, 279.—Looss, 1899b, 660.—Magalhāes, 1899, 259, 260.—Mont., 1888a, 73, 76; 1892, 17–28; 1892, 683, 685, 686, 687, 696, 697–698, 699, 700, 704, 706, 707–708, 709–710, 711, 713, figs. 3, 7 (syn. M. cymbium Dies.) (in Anas fuliginosa, A. fusca, A. marila, A. mollissima, Himantopus wilsonii, Mergus albellus, M. serrator).—Moul., 1856a, 23, 207, 208.—Much., 1898, 31.—Nord., 1832a, 85, 93; 1840, 617.—Par., 1896, 2.—Schneidemuchl, 1896, 303.—Sieb., 1835, 66, 82.—Ssinitzin, 1905, 158–159; 1906, 687.—Stoss., 1902, 2, 5, 6, 7, 9, 18 (of Par. syn. of Cyclocælium robustum Stoss.), 28, 30, 31 (of Mehlis to Typhlocælum), 33 (of Maghalhāes, syn. of Typhlocælum sp.).—Also reported for Fuligula marila. (of Maghalhães, syn. of Typhlocœlum sp.).—Also reported for Fuligula marila, Grus cinerea.

foliaceum Rud., 1819a, 83, 340-342 (t. h. Accipenser sturio; Arimini) to (Monost.).— Blainv., 1828a, 582.—Braun, 1889i, 440; 1891d, 421; 1894a, 1147.—Bremser, 1824a, 132; 1824c, pl. 8, figs. 3–7.—Dies., 1836, 240; 1850a, 319–320 (in Accipenser sturio, A. glaber, A. stellatus); 1858e, 324 (in Acipenser sturio, A. nasus); 1859c, 425 (syn. Amphilina foliacea Wagener).—Duj., 1845a, 364.-Grimm, 1871, 499–502.—Kroyer, 1852–53a, 778 (in Acip. stu.).—Macé, 1882, 64.—Mol., 1858, 128; 1861, 197.—Mont., 1892, 2.—Nord., 1840, 625.—Salensky, 1874, 16 Sept., 291–292, pls. 28–32.—Villot, 1876, 1345; 1878, 16.—Wedl, 1855, 380–382, 394, pl. 2a, fig. 15; 1855, 399–400, pl. 1b, fig. 6.—Also reported for Acipenser guldenstädtii.

galeatum Rud., 1819a, 86, 349-350 (t. h. Centronotus glaucus; Naples) to (Monost.).—Braun, 1893a, 915.—Dies., 1850a, 327.— Duj., 1845a, 362.—Mont., 1892, 685, 686, 697, 698, 699, 702, 703, 704, 716.—Stoss., 1887, 90; 1898, 62.—Reported also for Lichia amia, L. glauca.

gemellatum Mont., 1892, 716, for gemellum.

- gemellum Steenstrup, 1860, 113 (in Sphyræna baracuda).
- geminum Bremser, in Schmalz, 1831, 13 (for M. faba Bremser).—Baird, 1853a, 45 (= M. faba).
- gibbum Mehlis in Crep., 1846, 137 (t. h. Fulica atra).—Also reported for Gallinula chloropus.
- gracile Rud., 1809a, 326 (t. h. Salmo eperlanus) to (Hypost.); 1819a, 82.—Brand., 1892, 508.—Dies., 1850a, 328–329.—Duj., 1845a, 363.—Kroyer, 1846–53a, 20 (in Osmerus eperlanus L.).—Lamarck, 1816b, 186–187.—Mont., 1892, 717.—Nord., 1840a, 623–624.—Olfers, 1816, 48.—Stoss., 1902, 5.
- gurltii Cobbold, 1860a, 42 (in Lacerta agilis) (includes: M. sp. Gurlt, M. lacerta Dies., Dithyridium lacertæ Valenc., D. lacertæ viridis Rud., D. lacertæ muralis Rud., Petrathyrus obesus Crep., Piestocystis dithyridium Dies.).
- hepaticum suis Willach, 1893, 40–42; 1893, 26 Sept., 438; 1893, Mar., 124; 1894 (IV, 2), 874.—Braun, 1894g, 128–129 (= Cysticercus tenuicollis); 1894h, 755.—Stiles, 1898a, 28, 96.
- himantopodis Rud., 1819a, 87 (in Charadrius himantopus; Cat. Mus. Vien.).—Dies., 1850a, 323 (syn. of M. mutabile Zed.).—Mont., 1892, 707.—Stoss., 1902, 1.
- hippocrepis Dies., 1850a, 324 (t. h. Hydrochærus capybara; Brazil); 1855, 63, pl. 2,
 figs. 6-9; 1858e, 326.—Brand., 1892, 508.—Braun, 1892a, 568, 584, 709; 1893a,
 874, 915; 1901b, 344-346, pl. 19, fig. 10.—Looss, 1902m, 610.—Mont., 1888,
 54; 1892, 685, 686, 694, 695, 696, 697, 698, 700, 703, 704, 711.—Also reported for Cavia capybara.
- histrix Mol., 1858, 128 (t. h. Pelophylax esculentus; Patavii); 1861, 197–198, pl. 1,
 fig. 12.—Braun, 1892a, 583 (hystrix); 1893a, 881, 916.—Dies., 1858e, 328 (in Pel. esc.).—Mont., 1892, 685, 697, 698, 699, 700, 702, 705, 715, 716 (hystrix).—Par., 1894, 168.
- holostomoides Mehlis in Crep., 1846, 138 (t. h. Colymbus cristatus).—Brand., 1892, 509.—Braun, 1893a, 915.—Cohn, 1904, 230.
- hyalinum Schlotthauber, 1860, 129 (in Machetes pugnax).
- hystrix Brand., 1892, 506 (in Rana esculenta), for histrix.
- idi Rud., 1819a, 87 to (?Hypost.) (t. h. Cyprinus idus).—Dics., 1850a, 414 (syn. of Aspidogaster limacoides Dies.).—Mont., 1892, Oct. 7, 202 (syn. of Asp. lim.).
- ignotum Nicoll, 1906, 514 (in Hæmatopus ostralegus), sp. inq.
- impudens Crep., 1846, 149 (t. h. Squalus griseus).
- incommodum Leidy, 1856, 43 (t. h. Alligator mississippiensis; Florida); 1891a 414,
 (to Dist.) 1904a, 85.—Brand., 1892, 510.—Braun, 1893, 872, 915.—Cobbold,
 1860a, 42.—Dies., 1858e, 329 (in All. miss.).—Mont., 1892, 714, 715.—Pavesi,
 1881, 294.—Will.-Suhm, 1870, 11.
- isabellinum Ratzel, (1868), 153, t. h. Gadus æglefinus.—Linst., 1878a, 223 (=Rhipidocotyle gracilescens (Rud.)).—Stoss., 1898, 61.
- kuhni Cobbold, 1860a, 39 (M. leporis Kuhn, renamed) (in Lepus cuniculus).—Rail., 1893a, 339 (= M. leporis Kuhn).—See Cysticercus pisiformis.
- lacertae Dies., 1850a, 331 (t. h. Lacerta agilis) (syn. Dist. ammon); for M. sp. Gurlt, 1838, 229; 1858e, 328 (syn. Tetrathyrus obesus; cf. Piestocystis dithyridium Dies., 1850a, 495).—Crep., 1851, 1, 192.
- lacteum Jægers... 1896a, 165, 167–177, pl. 9, figs. 1–9 (in Cottus scorpius); 1896b, 179–180; 1898, 15; 1900, 736.—Braun, 1899b, 724; 1901b, 47.—Looss, 1899b, 671.—Maclaren, 1904, 583.—Ward, 1901, 180.—Type of Galactosomum 1899.
- lanceolatum Wedl, 1858, 251–252, pl. 1, figs. 15–17 (t. h. Himantopus rubropterus.—Brand., 1892, 508.—Braun, 1892a, 586, 673, 764, 766, 784, 786; 1893a, 876, 915.—Dies., 1858e, 325 (in Him. melanopterus) 707 (syn. of M. mutabile Zeder).—Linst., 1878a, 132.—Mont., 1892, 26; 1892, 691, 706.—Par., 1896, 2.—Stoss., 1902, 2, 4, 5, 23 (to Hæmatotrephus).
- Stoss, 180-, 2, 4, 5, 25 (to Hamacottephus).

 lentis Gescheidt, 1833, 421, 445 (in Homo).—Assenova, 1899, 29.—E. Bl., 1846, 342.—R. Bl., 1888a, 542, 543; 1895, 729.—de Bonis, 1882, 180.—Braun, 1883a, 59; 1893a, 870; 1895b, 155; 1903, 3 ed., 151.—Cobbold, 1876, 211.—Dav., 1877a, 1xxiii, 820, 822.—Dechambre, 1875a, 196 (of Nord.).—Dies., 1850a, 329 (in Homo); 1858e, 328.—Dunglison, 1893a, 821, 1174; 1895, 21 ed., 821, 1174.—Eiss, 1838, 23.—Gamb., 1896, 63.—Guenther, 1858, 205.—Hoyle, 1890, 538.—Huber, 1896, 501.—Ijima, 1889b, 122.—Kholodk., 1898, 34; 1899a,

153.—Kuech., 1855, 180–182.—Kuech. & Zuern, 1882, 285.—Leuck., 1863, 526, 633–634.—Moniez, 1896, 86, 152, 153.—Mont., 1892, 713 (of Nord.).—Mosler & Peiper, 1894, 185.—Rayer, 1843, 114, 116, 149.—Stiles, 1898a, 48; 1902, 25, 27, 28; 1905, 54.—Swart, 1862, 34.—Vogt, 1878, 10; 1878, 13.—Wagner, 1876, 122; 1883, 122.—Ward, 1895, 328 (in Homo); 1903, 866 (syn. of Agamodist. ophthalmobium Dies.).—Weinland, 1859, 280.—Wood & Fitz, 1897, 335.

leporis Kuhn, 1829, 464, pl. 11, figs. 6-7 (t. h. Lepus cuniculus).—Baillet, 1866b, 107.—Brand., 1892, 510.—Cobbold, 1860a, 39 (renamed kuhni); 1879b, 318.—Dav., 1877a, lxxiii.—Dies., 1850a, 330.—Mont., 1892, 42; 1892, 712, 713.—Rail., 1890, 132–133; 1893a, 216, 339 (=Cysticercus pisiformis); 1893, 339.—

Stiles, 1902s, 27; 1906, 42.

liguloideum Dies., 1850a, 320 (t. h. Vastres cuvieri; Borbæ, Brazil).—Brand., 1892,
506.—Braun, 1892a, 567, 586; 1893a, 916; 1894a, 1147.—Dies., 1850a, 320;
1855, 62, pl. 1, figs. 25-29; 1858e, 324.—Mont., 1892, 2 (to Amphilina); 1892,
716 (to Amphilina).

limacoides Dies., 1835a, 421 (=Aspidogaster limacoides).

lineare Rud., 1819a, 83–84, 343–344 (t. h. Tringa vanellus) to (Monost.).—Baillet, 1866b, 107.—Blainv., 1828a, 582.—Brand., 1892, 508.—Bremser, 1824, pl. 8, figs. 8–9.—Crep., 1839, 285.—Dies., 1850a, 411 (syn. of Notocotyle triseriale Dies.).—Duj., 1845a, 349.—Hass., 1896a, 3 (syn. of Noto. verrucosum Frœlich).—Rail., 1893, 340.—Nord., 1840, 625.—Sieb., 1835, 50.

loliginis (delle Chiaje, 1841) Par., 1894, 169 (in Loligo sp.; Naples).

tucaneum Brand., 1902, 511, for lucanica.

lucania Leidy, 1904a, 143, for lucanica.

lucanica Leidy, 1877, 200–201 (in Planorbis parvus; U. S. A.) to (Glenocerc.); 1904a, 143–144.—Brand., 1902, 511 (lucaneum).—Linst., 1889a, 122 (lucanicum).

lucanicum Linst., 1889a, 122 (see lucanica).—Mont., 1892, 717.

macrorchis Brand., 1892, 508 (in marine turtles).—Braun, 1893a, 915; 1899b, 715, 722; 1901b, 38, 54, 58.—Looss, 1899b, 756.

macrostomum Rud., 1809a, 337–338 (t. h. Larus cinerarius; Greifswald), to (Monost.); 1819a, 86.—Brand., 1892, 510.—Braun, 1893a, 915.—Dies., 1850a, 330.—Duj., 1845a, 358.—Mont., 1892, 706, 709.—Olfers, 1816, 48.

macrurum Schlotthauber, 1860, 129 (in Corvus glandarius).

marænulæ Rud., 1809a, 339–340 (t. h. Salmo marænula; Europe) to (Hypost.);
 1819a, 87.—Brand., 1892, 510 (to Monostomulum).—Braun, 1893a, 871.—
 Dies., 1850a, 329.—Mont., 1892, 717 (marenulæ).—Also reported for Coregonus albula.

marenulæ Mont., 1892, 717 (for marænulæ).

marilæ Rud., 1819a, 87 (in Anas marila).—Dies., 1850a, 411 (marillæ, syn. of Notocotyle triseriale).

marillæ Dies., 1850a, 411 for marilæ, 1819.

micropterygis (Richiardi, 1902) Ariola, 1902, 103 (syn. of Didymost. bipartitum Wedl).

microstomum Crep., 1829, 1, 49–50, pl. 1, figs. 10–11 (t. h. Fulica atra; Greifswald); 1837, 314.—Ben., 1858a, 1861a, 70 (syn. of Monost. mutabile).—Braun, 1892a, 658.—Dies., 1850a, 323 (syn. of M. mutab.).—Nord., 1832a, 85.—Rail., 1893a, 338 (= M. mutab.).—Sieb., 1835, 50.—Stoss., 1902, 2, 13 (syn. of Cyclocelum mutabile).

minutissimum Stoss., 1896, 130 (in Anas boschas; Doberdò); 1898, 63.

molæ Rud., 1819a, 87, 350–351 (t. h. Orthagoriscus mola; Naples).—Dies., 1850a, 359 (syn. of Dist. okenii).—Mont., 1893, 137.

molle Leidy, 1856, 43 (t. h. Sternotherus odoratus; U. S. A.); 1904a, 86.—Brand.,
1892, 510.—Braun, 1893a, 876, 915; 1901b, 54.—Dies., 1858e, 328 (in Ster. od.).—Mont., 1892, 715; 1896, 165.—Stiles & Hass., 1894e, 414, to Dist. (Polyorchis); 1894h, 162; 1895a, 737.

murænulæ Cobbold, 1860a, 43, for marænulæ = M. braunii Cobbold.

mutabile Zed., 1800, xvi, 150, 154-155 (t. h. Fulica chloropus; Germany); 1803, labile Zed., 1800, xvi, 150, 154–155 (t. h. Fulica chloropus; Germany); 1803, 189, pl. 3, fig. 1.—Aitken, 1866, 838; 1872, 204.—Baillet, 1866b, 91, 107.—Ben., 1858a, 1861a, 66, 68, 69–77, 79, 99, 171, 172, 179, 193, 202, 203, 204, 206, 211, 213, 214, 215, 223 (syn. M. microstomum), 256, 336, pl. 12, figs. 1–20.—Biehringer, 1884, 7.—de Bonis, 1882, 102.—Brand., 1892, 507, 508.—Braun, 1890a, 510; 1892, 51–52; 1892a, 584, 587, 590, 627, 642, 648, 658, 661, 663, 673, 674, 678, 715, 727, 761, 768, 769, 776, 780, 781, 784, 785, 786, 787, 805, 813; 1893a, 818, 820, 876, 879, 880, 883, 915; 1893b, 176, 179, 187; 1898d, 468; 1894a, 1166; 1895b, 17; 1896a, 1255; 1899, 467; 1901e, 346.—Crep., 1837, 314, 323, 324, 325.—Darr, 1902, 675.—Desmonceaux, 1868, 21.—Dies., 1836, 248; 1850a, 323–324 (syns.: M. vanelli Rud., M. himantopodis Rud., Dist. calidris Rud., M. microstomum Crep.): 1859a, 382; 1858e, 325 (in Anas anser): 1859c, 425 M. microstomum Crep.); 1855a, 382; 1858e, 325 (in Anas anser); 1859c, 425 1893a, 338 (syns.: M. microstomum Crep., M. attenuatum Mol. not Rud.); 1898, Oct., 627–628 (in geese).—Rud., 1809a, 333–334 to (Monost.); 1819a, 85.—Schneidemuehl, 1896, 303.—Sieb., 1835, 49–83, pl. 1, figs. 1–9 (in Anas anser dom., Fulica atra, Gallinula chloropus, Grus cinerea, Rallus aquaticus); 1850, 671, 672; 1854, 21, 22, 23.—Spengel, 1892.—Steenstrup, 1842, 40, 49, 56.—Stein, 1882, 1.—Stoss., 1889, 184; 1901, 92 (4) (in Himantopus candidus; Roma); 1902, 1–40, pls. 1–9; 1902, v. 9 (13), 406–407; 1905 (vii), 62.—Thomson, 1855, 188.—Verrill, 1870, 179.—Vogt, 1878, fig. 30.—Wagener, 1857, 22, 24, 26, 45, 52, 67.—Zernecke, 1895, 61.—Ziegler, 1905, 36, 37 (in Gallinula chloropus).—Also reported for Anser cinereus dom., Crax alector, Fuligula marila. marila.

mutabile Zed.—Stoss., 1902, 13, to Cyclocælum.

mutabile of Ben., 1858a, 69, pl. 12; of Dies., 1859c, 425; of Par., 167; and of Stoss., 1891, v. 13, 111.—Stoss., 1902, 15 (syn. of Cyclocœlum ovopunctatum).

mutabile of Sieb., 1835, 49; of Dies., 1858e, 325; of Dav., 1877a, lxxiii; and of Mueh., 1898, 31.—Stoss., 1902, 20 (syn. of Cyclocœlum arcuatum).

nematoides Crep., 1846, 129 (in Falco albicilla).

nephriticum Mehlis in Crep., 1846, 138 (t. h. Colymbus arcticus).—Brand., 1892,
 509.—Braun, 1893a, 877, 915; 1893d, 467.—Cohn, 1904, 230 (to Eucotyle), 237.

nephrocephalum Dies., 1858e, 327 (M. renicapite renamed) (in Sphargis coriacea; America).—Braun, 1899, 627; 1901b, 52.—Mont., 1892, 715.

nigropunctatum Linst., 1883, 310, pl. 9, fig. 52 (in a bird "Akatza;" Turkestan).— Brand., 1892, 507.—Braun, 1892a, 784, 786 (in birds; Turkestan); 1893a, 915.— Mont., 1892, 26; 1892, 698, 706, 707 (syn. of M. mutabile).—Stoss., 1902, 5, 6.

noctulæ Cobbold, 1860a, 39 (M. vespertilionis Rud., renamed) (in Vespertilio noctula).--Mont., 1892, 712.

obscurum Leidy, 1887, 24 (in Megalops thrissoides; Coll. Army Med. Mus., Wash.); 1904a, 197.—Brand., 1892, 510.—Braun, 1893a, 874, 915.—Mont., 1892, 716, 717.

ocreatum (Gœze, 1782) Zed., 1800a, xvi, 150, 152–154; 1803a, 189.—Bellingham, 1844a, 337.—Blainv., 1828a, 582.—Bremser, 1824c, pl. 8, figs. 10–11.—Cobbold, 1860a, 39 (includes Dist. lorum Duj., Fasc. ocreata Gœze not Rud., Cucullanus ocreatus Schrank, C. talpæ Mueller); 1879b, 296.—Crep., 1839a,

285.—Dies., 1850a, 326 (syns.: Fasc. ocreata Gœze, Cucullanus talpæ Mueller, C. ocreatus Schrank, Dist. lorum Duj.); 1858e, 328 (in Talpa europæa; Ireland).—Duj., 1845a, 344.—Kuech., 1855, 181.—Lamarck, 1816b, 187.—Looss, 1901, 192 (= Dist. lorum Meln.).—Mont., 1888a, 7.—Nord., 1840, 624 (syns.: Fasc. ocreata Gœze, Cucullanus ocreatus Gmelin).—Olfers, 1816, 48.—Rud., 1809a, 329-331 to (Monost.); 1819a, 84.—Schlotthauber, 1860, 129.—Stoss., 1892, 17 (syn. of Dist. lorum Duj.), 18 (acreatum) (syn. Dist. lorum Duj.).—Walter, 1866, 65.

octopodis delle Chiaje, teste in Mont., 1892, 717, 718.—Par., 1894, 169 (in Octopus

vulgaris; Naples).

oculobium Cohn, 1902d, 712 (in Vanellus melanogast.; Greifswald Coll.).—Engler, 1904, 186.—Fuhrmann, 1904, 59, 61.

orbicolare Sons., 1891, 262, for orbiculare.

orbiculare Rud., 1819a, 83, 342 (t. h. Sparus salpa; Naples) to (Monost.).—Barbagallo & Drago, 1903, 411 (in Box salpa, Oblata melanura; Catania).—Brand., 1892, 509.—Braun, 1892a, 567, 568, 603, 635, 640, 650, 651; 1893a, 915.—Dies., 1850a, 320.—Duj., 1845a, 360.—Lint., 1898, 541–542, pl. 54, figs. 2–5 (in Lobotes surinamensis; Woods Hole); 1901, 416, 457.—Luehe, 1901d, 49–60, figs. 1–5; 1901e, 235–236; 1901f, 421.—Mont., 1888a, 7, 16, 41, 42, 43; 1892, 685, 687, 689, 690, 694, 697, 698, 699, 700, 701, 703, 716.—Par., 1887, v. 30, 15; 1894, 592.—Sons., 1891, 262 (orbicolare in Box salpa).—Stoss., 1883, 111; 1898, 65 (in B. salpa; Triest).

ornatum Leidy, 1856, 43 (t. h. Rana pipiens).—Brand., 1892, 510.—Braun, 1893a, 877, 915.—Dies., 1858e, 326 (in R. pip.; Philadelphia).—Mont., 1892, 685,

697, 698, 715.—Staff., 1902, 724.

ovatum Mol., 1859, 822–824, pl. 2, figs. 2–4 (in Gallinula crex; Padua).—Brand., 1892, 508.—Braun, 1893a, 916; 1893b, 185.—Looss, 1899b, 662, 663.—Mont., 1892, 38, 40–41 (syn. of Notocotyle verrucosum); 1892, 706, 709.—Odhn., 1905, 368.—Stoss., 1902, 5.—Also reported for Crex pratensis.

pandum Braun, 1901a, 48-50, fig. 17 (in Thalassochelys caretta; Naples).

petasatum Deslongchamps, 1824ee, 511 (t. h. Hæmatopus ostralegus; Caen).—
Brand., 1892, 505, 509.—Braun, 1892a, 568; 1893a, 874, 915.—Dies., 1850a, 330.—Duj., 1845a, 350–351.—Looss, 1902m, 610.—Mont., 1888, 7; 1892, 706, 709.—Villot, 1878, 18–20, pl. 5, fig. 1 (in Strepsilas interpres).

pileatum (Rud., 1802) Zed., 1803, 188 (in Sterna hirundo).—Dies., 1850a, 314 (to Holost.).—Olfers, 1816, 48.—Rud., 1809a, 338–339; 1819a, 90 (to Amphist.).

pingue Mehlis in Crep., 1846, 138 (t. h. Colymbus cristatus).—Brand., 1892,
 509.—Braun, 1893a, 877, 915; 1893d, 467.—Cohn, 1904, 230, 232, 233 (to Renicola), 235 (syn. of Taphrogonymus holostomoides).

plicatum Crep., 1829a, 878–880, pl. 52, figs. 9–11 (t. h. Balæna rostrata; Rügen).—
Ben., 1870, 357.—Brand., 1892, 509.—Braun, 1892a, 605.—Cobbold, 1879b, 421.—Dies., 1850a, 324.—Duj., 1845a, 344.—Jægers., 1891b, 32 pp., figs. 1–16 (anatomy); 1892a, 572–573 (see Ogmogaster plicatus).—Mont., 1892, 696, 704, 713; 1893, 203.—Odhn., 1905, 366 (to Ogmogaster).—Also reported for Balæna mysticetus.

præmorsum Nord., 1832, 55–56, 96 (t. h. Cyprinus brama).—Brand., 1892, 511 (to Monostomulum).—Braun, 1893a, 871, 915.—Dies., 1850a, 322 (in Abramis brama; Berlin).—Duj., 1845a, 363.—Kroyer, 1846–53a, 388 (in Abr. br. L.).—

Mont., 1892, 685, 696, 697, 698, 716.—Walter, 1866, 65.

prismaticum Zed., 1800a, xvi, 150, 151–152 (t. h. Corvus frugilegus; Europe).— Brand., 1892, 510.—Braun, 1893a, 876, 915.—Dies., 1850a, 328.—Duj., 1845a, 349.—Looss, 1901, 192, 193 (type of Monost. by elimination; see, however, mutabile and p. 308); 1902m, 700, 701, 702, 703, 707, 717, 718, 719, 720, 721, 729, 730.—Mont., 1892, 685, 699, 713, 714.—Luehe, 1901, 174.—Olfers, 1816, 48.—Rud., 1809a, 334–335 to (Monost.); 1819a, 85.

proteus Brand., 1891, 19, 22, 23; 1892, 567, 570, 571, pl. 22, figs. 5, 6 (in Chelonia viridis); 1892, 508.—Bettend.; 1897a, 11; 1897, 315.—Brand., 1891d, 19, 22, 23.—Braun, 1893a, 915 (in Chelone midas); 1893b, 176, 177, 185 (in Chelonia viridis); 1899b, 715, 722; 1901b, 38.—Darr, 1902, 673.—Looss, 1894a, 186, 204; 1899b, 661, 662, 668, 869, 756, 767-770, to Baris as type; 1902m, 619, 626, 632, 636, 647, 653, 669, 678, 685 (of Walter, 1893, 197, pl. 10, figs. 13, 18, 19, as syn. of Octangium sagitta), 694 (of Walter, 1903, 196, figs. 22-23 to Deuterobaris).—Luehe, 1901, 58.—Mont., 1893, 213, 214.—Ofenheim, 1900, 148, 167.—Shipley, 1900, 533, 539.—Walter, 1892, 248, 249, 250; 1893, 196, 197, pl. 10, figs. 13, 18, 19, 22-23.

pseudamphistomum Crep., 1846a, 146 (t. h. Chelonia mydas).

pumilio Looss, 1896b, 154–158, pl. 10, figs. 101–106 (in Pelecanus onocrotalus; Cairo); 1899b, 670 (type of Haplorchis), 753.

renicapite Leidy, 1856b, 43 (t. h. Sphargis coriacea); 1904a, 86.—Brand., 1892b,
510 (in Dermatochelys coriacea).—Braun, 1893a, 915; 1899, 627-629; 1899b,
715, 721, 722; 1901b, 38, 52-53, fig. 25.—Dies., 1858e, 327 (renamed nephrocephalum) (in Sphargis coriacea; America).—Mont., 1892, 685, 711, 714, 715.

reticulare Ben., 1859, 84-85, pl. 2, figs. 7-10 (t. h. Chelonia midas); 1858a, 1861a, 193.—Brand., 1892, 508.—Braun, 1892a, 651; 1893a, 916; 1893b, 185 (in Ch. viridis); 1899b, 715, 721, 722, 725; 1901b, 38, 54.—Jackson, 1888, 647.—Looss, 1894a, 186, 202, 204; 1899b, 756, 763, 765, 766, 767, 769; 1902m, 632, 643, 661, 662, 666, 688 (of Walter, 1893, 193, pl. 10, figs. 5, 6, 8a, syn. of Polyangium linguatula), 690, 691 (to Microscaphidium), 720.—Mont., 1892, 714, 715.—Shipley, 1900, 533, 539.—Walter, 1892, 248, 249.

rhombi lævis Dies., 1858e, 328 (in Rhombus lævis; Triest), based on Wedl, 1855, 380, 394, pl. 2, 14.—Braun, 1893a, 871.—Mont., 1892, 717 (rombi-lævis).

rombi-lævis Mont., 1892, 717 (for rhombi-lævis).

rubrum Kuhl & van Hasselt, 1822a, 113; 1824a, 311 (in Chelonia midas; Hes des Cocotiers).—Braun, 1899b, 721, 723, 724; 1901b, 38, 50-52, figs. 26, 28.—Crep., 1846, 146.—Dies., 1850a, 325 (syn. of M. trigonocephalum).—Looss, 1899b, 667, 756; 1902m, 555.—Shipley, 1900, 532.

sacidiornicola Stoss., 1902, 34 (for sarcidiornicola).

sarcidiornicola Mégnin, 1890c, 87-90, fig. 2 (in Sarcidiornis melanota; Madagascar); 1890, 685–687, fig. —Braun, 1899, 468.—Mont., 1892, 28–31; 1892, 685, 686, 687, 697, 704, 706, 708, 709, 710, 713 (in Sar. mel.).—Stoss., 1902, 9, 33, 34 (sacidiornicola) (to Typhlocœlum).

semifuscum Olss., 1876, 28 (in Sula bassana; Lund).—Brand., 1892, 505, 509.— Braun, 1893a, 916 (in S. bas.).—Mont., 1892, 706, 710.

sepiolæ delle Chiaje, teste Crep., 1846a, 158.—Mont., 1892, 717, 718.—Par., 1894,

settenii Numan, (1840), 358 (eye of horse).—Anacker, 1890m, 506.—R. Bl., 1888a, 542.—R. Bl. & Rail., 1891a, 26-28 (cestrid larva).—Braun, 1889a, 337; 1891d, 424.—Dies., 1850a, 617 (to Pentast.).—Moniez, 1896, 154.—Mont., 1892, 713.—Rail., 1895, 772.—Sieb., 1842, 299.—Stiles, 1902s, 28 (= larval dipteron).—Ward, 1895, 338 (in Equus caballus).

spathulatum Dies., 1859c, 426 (for spatulatum).—Mont., 1892, 685, 697, 698, 716. spatulatum Leidy, 1859, 111 (in fish).—Dies., 1859c, 426 (spathulatum) (in American fish).—Mont., 1892, 685, 697, 698, 716.

species Brand., 1891d, 19 (Monostoum).

species Braun, 1891e, 8.—Mont., 1892, 713.

species Braun, 1901b, 50 (in Chelone mydas).

species Kroyer, 1846-53a, 462; 1852-53a, 1224 (in Leuciscus idus L.).

species Lint., 1901, 416, 439; 1905, 335, 356, 360, 367, figs. 217, 226-229 (in Fundulus majalis, Menidia menidia, Orthopristis chrysopterus, Trachinotus carolinus).

species Stoss., 1902, 34–35 (in Grus cinerea).

species Stoss., 1902, 35 (in Mergus albus).

species Stoss., 1902, 36 (in Machetes pugnax; Berlin).

species Stoss., 1902, 36 (in Fulica atra; Seeburg).

species Stoss., 1902, 36 (in Grus cinerea; Luckenwalde).

species Piesbergen, 1886, 356.—Mont., 1892, 716 (in Cobitis barbatula).

species Villot, —.—Mont., 1892, 706 (in Strepsilas interpres).—See Linst., 1878a, 133.

spinosisimum Mont., 1892, 689 (for spinosissimum).

702, 705, 716.—Type of Centroderma 1901.

spirale Dies., 1850a, 325 (t. h. Hypsilophus tuberculatus, Podocnemis tracaxa, Chelonsidis tuberculatus; Brazil); 1855, 63, pl. 2, figs. 10–13; 1858e, 326 (in Hyp. tub.).—Brand., 1892, 507.—Braun, 1892a, 567, 569, 709; 1899, 631; 1901b, 54–55.—Cobbold, 1860a, 41.—Mont., 1888, 7, 8; 1892, 35; 1892, 685, 696, 697, 698, 714, 715.—Stoss., 1895, 232 (to Dist.).

squamula (Rud., 1819) Dies., 1850a, 321 (in Mustela putorius; Jan., Tolosæ).— Brand., 1892, 506.—Cobbold, 1860a, 38.—Stoss., 1892, 20 (to Dist.).

stossichianum Mont., 1892, 12, 19, figs. 5, 7, 11, 15, 18 (includes M. capitellatum of Stoss., 1883, 2, pl. 2, fig. 9; Carus, 1884, 122, pars; Parona, 1886, 5, 7, nota 2; 1887, 489; Setti, 1891, 4) (in Box salpa; Italy "del nostro golfo"): 1892, 685, 686, 687, 688, 689, 694, 695, 696, 697, 698, 699, 700, 702, 703, 705, 716; 1893, 15, 16, 17, 24, 35, 83, 84, 115, 116.—Brand., 1892, 507, 509.—Braun, 1893b, 178, 179.—Looss, 1899b, 669.—Luehe, 1901, 58, 59, 60.—Stoss., 1898, 63–64 (in Box salpa; Triest).

sulcatum Rud., 1809a, 337 (t. h. Rana pipa, intestine) to (Monost.); 1819a, 86.—
Brand., 1892, 510.—Braun, 1893a, 916 (in Pipa americana).—Dies., 1850a, 325–326.—Duj., 1845a, 359.—Mont., 1892, 715, 716.—Olfers, 1816, 48.

tenuicolle Rud., 1819a, 85, 346–347, 577, pl. 2, figs. 1–4 (t. h. Lampris guttatus; Groningæ) to (Monost.).—Ben., 1858a, 1861a, 111, 199, 200; 1870, 87.—Blainv.. 1828, 582 (tenuicollis).—Dies., 1850a, 359 (syn. of Dist. affine Dies.) (in Lam. gutt.); 1859c, 430.—Duj., 1845a, 361–362.—Mont., 1888a, 9; 1893, 150.—Schmalz, 1831, 16, pl. 6, figs. 10–12 (in Lam. gutt.).

testudinis M. S. Vienna Coll. in Braun, 1899, 630 (= Dist. scyphocephalum Braun).

tethyæ delle Chiaje teste Mont., 1892, 717.

thethydis delle Chiaje, teste E. Bl., 1847a, 309.

thetycola (delle Chiaje), teste Pag., 1862, 298.

todari delle Chiaje see Mont., 1892, 717, 718.—Par., 1894, 168 (in Ommastrephes todarus; Naples).

totari delle Chiaje.—E. Bl., 1847a, 309, see todari.

trigonocephalum Rud., 1809a, 336–337 (t. h. Testudo mydas; Europe) to (Monost.), includes Planaria mydæ Braun MS.; 1819a, 86, 349.—Bellingham, 1844a, 337.—Ben., 1858a, 1861a, 193.—Brand., 1892, 508.—Braun, 1891d, 424 (in Thalassochelys corticata; 1892a, 568; 1893a, 916; 1899b, 715, 721, 722, 723, 725; 1899, 627, 628; 1901b, 38–45 (originals in stomach of Chelone mydas), 45 (pars of Rud. 1819a, Dies., 1850a, Shipley, 1900, syn. of Monost. album), 46, 48, 49, 50, 51 (pars of Dies., 1850a, Shipley, 1900, syn. of M. rubrum), 52, fig. 29; 1901e, 346.—Dies., 1850a, 325 (syns. Planaria midæ Braun, M. rubrum et album Kuhl & Hasselt); 1858e, 327 (in Chelone (Eretmochelys) imbricata).—Duj., 1845a, 358–359.—Looss, 1894a, 204; 1899b, 666, 756 type of Pronocephalus [see obliquus]; 1901, 566, 567, 568; 1902m, 413, 527 (of Braun, 1901, 38, syn. of Pronocephalus obliquus), 529, 531, 548 (to Pleurogonius), 549 (original species contained 2 forms) (of Shipley, 1900, 532, pl. 54, figs. 1–7, syn. of Cricocephalus delitescens), 550, 551, 552, 556, 557, 558 (of Ben., 1859, 81. pl. 2, fig. 5, and Walter, 1893, 191, fig. 1, pl. 10, syn. of Pleurogonius longiusculus Looss), 561, 562, 563, 564, 580, 582 (= Pleurog. long. Looss), 584, 591, 595, 596, 713, 714, 716,—Mont., 1892, 685, 686, 687, 689, 694, 695, 696, 697, 698, 702, 703, 709, 711, 714, 715, fig. 5; 1893, 213.—Moul., 1856a, 23.—Olfers, 1816, 48.—Shipley, 1900, 532–540, fig. A. B., pl. 54, figs. 1–7.—Stoss., 1891, 111.—Walter, 1892, 248; 1893, 30 May, in 189–235, pls. 10–12.—Reported also for Halichelys atra.

tringæ Brand., 1892b, 507 to Cyclocœlum, 508.—Braun, 1893a, 876, 916; 1899,
 467.—Looss, 1899b, 660.—Stoss., 1902, 26, fig. 26 (to Hæmatotrephus) (in

Tringa variabilis; Tor, Peninsula of Sinai).

triseriale (Dies., 1839) Dav., 1860, xlviii; 1877a, lxxiii.—Baillet, 1866b, 107 (syn. of M. verrucosum).—Hoyle, 1890, 539.—Verrill, 1870, 179.

vanelli Rud., 1819a, 87, 350 (t. h. Tringa vanellus; Mus. Vien.).—Dies., 1850a, 323 (syn. of M. mutabile Zed.).—Sieb., 1835, 50 (in Vanellus cristatus).

variabile (Nitzsch, 1857) Dav., 1860, xlviii; 1877, lxxiii.—Verrill, 1870, 179.

ventricolum Rud., 1809a, 334, misprint for ventricosum.

ventricosum (Rud., 1802) Rud., 1809a (ventricolum, p. 334), 335–336, 339; to (Monost.); 1819a, 86 (in Motacilla luscinia; Gryphiæ).—Brand., 1892, 509.—Braun, 1893a, 876, 916.—Dies., 1850a, 328 (syn. Festucaria vent.).—Duj., 1845a, 348–349.—Mont., 1892, 713, 714.—Olfers, 1816, 48.

verrucosum (Frœlich, 1789) Zed., 1800a, xvi, 150, 155-159.—Anacker, 1887c, 513.—Baillet, 1866b, 107 (syn. M. triseriale).—Baird, 1853a, 45.—Bellingham, 1844, 336.—Ben., 1858a, 1861a, 72, 77-81, 179, 189, 190, 193, 197 (syns. Fasc. ver., F. anseris, Festucaria pedata, Notocotyle triseriale, Notocotylus triserialis) (in Anas tadorna).—Ben., 1859, 77-80.—E. Bl., 1847, 304-308, pl. 13, fig. 2.—Brand., 1892, 508.—Braun, 1891d, 421; 1892a, 709, 762, 764; 1893a, 874, 891; 1893b, 177, 185; 1893d, 468; 1901b, 39.—Cobbold, 1872b, 90; 1877, 238; 1879b, 41, 440.—Crep., 1839, 285; 1846, 141.—Dies., 1839, 234; 1850a, 411 (syn. of Notocotyle triseriale Dies.); 1858e, 369 (syn. of Notocotyle triserialis Dies.) Notocotyle triseriale Dies.); 1858e, 369 (syn. of Notocotyle triserialis Dies.) (in Anas anser dom. et ferus, Hæmatopus ostralegus, Fulica atra); 1859c, 437 (syn. of Notocotyle triserialis) (in Anas tadorna, Planorbis, Lymnæus; Belgia).—Duj., 1845a, 355-358, pl. 8, fig. B.—Hass., 1896a, 3 (to Notocotyle).— Hoyle, 1890, 539 (= Notocotyle triseriale).—Jackson, 1888, 642.—Lamarck, 1816b, 187.—Levinsen, 1881a, 78 (in Somateria mollissima).—Looss, 1896b, 146-153, 192-197, 228, pl. 10, figs. 94-100, pl. 14, figs. 146-151 (in Canard dom.; Alexandria) (syns.: Notocotyle triseriale, N. verrucosum); 1899b, 661, 662, Alexandria) (syns.: Notocotyle triseriale, N. verrucosum); 1899b, 661, 662, 663, 664; 1901, 192 (taken by Dies, as representative of Notocotyle).—Mol., 1859, 823.—Mont., 1888, 67; 1892, 26, 27, 40 (to Notocotyle); 1892, 685.—Much., 1898, 31, 32.—Nord., 1840, 602 (syn. of Nocotylus triserialis), 624 (syn. Fasc. verr. Fr.).—Odhn., 1905, 366, 367, 368, 369, 370 (to Catatropis) (of Looss, 1896, syn. of Notocotyle ægyptiaca Odhn.).—Olfers, 1816, 48.—Rail., 1893a, 340 (to Notocotyle).—Risso, 1826, 262.—Rud., 1809a, 331–333 to (Monost.) (includes Fasc. anseris Gmelin, Festucaria pedata Schrank); 1819a, 84, 344.—Schnejdemuchl, 1896, 303.—Sieb., 1835, 56, 60 (in Fulica atra, Gallinula porzana, Phasianus gallus, Rallus aquaticus).—Sons., 1897, 252.—Verrill, 1870, 179.—Wedl, 1857, 248–250, pl. 2, figs. 9–14.—Wolffhuegel, 1900, 9, 55, 56, 57.—Will.-Suhm, 1870, 7.—Reported also for Anas anser, A. bernicla, A. boschas fera, A. marila, A. mollissima, A. querquedula, A. segetum, A. boschas fera, A. marila, A. mollissima, A. querquedula, A. segetum, Cygnus olor, Harelda glacialis, Machetes pugnax, Mergus merganser, Recurvirostra avocetta, Rhynchaspis clypeata.

vespertilionis Rud., 1819a, 87 (t. h. Vespertilio noctula; Cat. Ent. Vien.).— Brand., 1892, 510.—Braun, 1893a, 916.—Dies., 1850a, 329–330 (in Vesp. noc.).—Kolenati, 1857, 11.—Mont., 1892, 712.—Also reported for Vespertilio

lasiopterus, Vesperugo noctula.

vinal-edwardsii Lint., 1901, 416, 470, figs. 373, 374, 375, 376; 1905, 329, 335, 379, 410, figs. 220-221 (in Opsanus tau, Orthopristis chrysopterus; Beaufort,

North Carolina).

viviparæ Linst., 1877, 185, pl. 13, fig. 16 (in Paludina vivipara).—Mont., 1892, 717 wedlii Cobbold, 1860a, 43 (=M. rhombi lævis Wedl, renamed) (in Rhombus lævis); 1879b, 463.

zederi Rud., 1805a, 44 [evidently not as specific combination, but as Monostoma Zed.].

(MONOSTOMA) Rud., 1809a, typical subg. of Monost. in Rud., 1809a, 328–339 "ore antico, sive terminali." As Rud. excludes Festucaria anatis from M. (Monost.), his typical subg. is to be construed as distinct from Monost. Zed. Festucaria renamed, by which it is, however, antedated.

attenuatum Rud., 1809a, 328-329 (t. h. Scolopax gallinago, cæca; Griefswald). See also under Monost.

capitellatum Rud., 1819a, 83, 343 (t. h. Sparus salpa; Naples). See also under Monost.

cornu (Zed., 1800) Rud., 1819a, 85. See also under Monost.

crenulatum Rud., 1809a, 328 (t. h. Motacilla phænicurus, intest.; Greifswald). See also under Monost.

crucibulum Rud., 1819a, 83, 342-343 (t. h. Muræna conger, M. cassini (M. myroides); Naples) (to Gasterost. by Gervais & Ben., 1859) (to Dist. (Crossodera) by Duj. 1845a, 435). See also under Monost.

ellipticum Rud., 1809a, 333 (M. bombynæ Zed., 1800, renamed). See also under Monost.

filicolle Rud., 1819a, 85-86, 347-348 (t. h. Brama raji; Naples). See also under

foliaceum Rud., 1819a, 83, 340-342 (t. h. Accipenser sturio; Arimini). See also under Monost.

galeatum Rud., 1819a, 86, 349–350 (t. h. Centronotus glaucus; Naples) (to Gasterost., by Stoss., 1898, 62). See also under Monost.

lineare Rud., 1819a, 83–84, 343–344 (t. h. Tringa vanellus). See also under Monost.
macrostomum Rud., 1809a, 337–338 (t. h. Larus cinerarius, intest.). See also under Monost.

mutabile (Zed., 1800) Rud., 1809a, 333–334. See also under Monost.

ocreatum (Gœze, 1782) Rud., 1809a, 329-331. See also under Monost.

orbiculare Rud., 1819a, 83, 342 (t. h. Sparus salpa; Naples). See also under Monost. pileatum (Rud., 1802) Rud., 1809a, 338–339 (to Amphist., by Rud., 1819a, 90). See also under Monost.

prismaticum (Zed., 1800) Rud., 1809a, 334–335. See also under Monost. sulcatum Rud., 1809a, 337 (t. h. Rana pipa). See also under Monost.

tenuicolle Rud., 1819a, 85, 346–347, 577, pl. 2, figs. 1–4 (t. h. Lampris guttatus). See also under Monost.

trigonocephalum Rud., 1809a, 336–337 (t. h. Testudo mydas, stomach; Europe). See also under Monost.

ventricosa (Rud., 1802) Rud., 1809a, 335-336. See also under Monost.

verrucosum (Frœlich, 1789) Rud., 1809a, 331-333. See also under Monost.

MONOSTOMATA Zed., 1800a, 150, plural for Monostoma.—Dowker, 1882a, 11.

MONOSTOMATIDÆ Gamb., 1896a, 73.

MONOSTOMEA Mont.—Olss., 1893.

MONOSTOMEÆ Mont., 1888, 7, 8, 9, 11, 12, 14, 15, 16, 18, 34, 36, 37, 41, 47, 48, 51, 52, 53, 54, 56, 90, 92, 102, 105, 108; 1892, 689.—Brand., 1888a, 49.—Braun, 1893a, 886, 887.—Par. & Perugia, 1890, 9.

MONOSTOMEN, German for Monostomata.—Stiles, 1902, 25, 26.

MONOSTOMES Dechambre, 1875a, 196, French for plural of Monostoma.

MONOSTOMEUS Rafinesque, 1815, 151 (new name for Festucaria Schrank, hence type F. anatis; apparently intended for Monostoma as it is attributed to Zed.

[MONOSTOMIA Rafinesque, 1815, 150, order of Epistomia.]

MONOSTOMIDA Kolenati, 1857, 11.

MONOSTOMIDÆ Cobbold, 1877f, 326; 1879b, 4.—R. Bl., 1895, 729.—Brand., 1890a, 576; 1892, 505.—Braun, 1883a, 58; 1893a, 887, 890, 895, 900, 914; 1895b, 137.—Hoyle, 1890, 539 (includes Monostomum, Monocotyle).—Jackson, 1888, 642, 654 (Monostomum).—Kholodk., 1898, 25, 33.—Looss, 1899b, 541, 543, 658, 659, 752; 1901b. 192, 193; 1902m, 524, 700 (validity of fam. name), 701, 702, 707, 731, 838.—Luehe, 1901, 174; 1901, 488.—Mont., 1888, 7, 10, 18, 21, 52, 60, 66, 67, 93, 105; 1892, Oct. 7, 214 (fam. of suborder Malacocotylea); 1892, 713.—Mueh., 1898, 31.—Pratt, 1902, 890, 909 (includes Microscaphidiinæ, Pronocephalinæ, Haplorchidinæ).—Schneidemuehl, 1896, 295, 296, 303.—Stoss., 1898, 63; 1902, 11.

MONOSTOMIDEA Ben.—Carus, 1863, 479.

MONOSTOMIDEN, German name.—Brand., 1892b, 7 Oct., 504-511.

MONOSTOMIDES R. Bl., 1888a, 541, 542, French for Monostomidæ.

MONOSTOMULUM Brand., 1892b, 510 [collective group for immature forms; requires no type].—Stiles, 1902s, 25, 28.

asperum (Vaillant, 1863) Brand., 1892b, 510 [cf. asperum 1857], (in Siren lacertina, sub cute in caps.).

delphini (Blainv., 1825) Brand., 1892b, 510 (in Delphinus dalei, in adipe folliculo inclusum).

dubium (Cobbold, 1858) Brand., 1892b, 511 (in Gasterosteus spinachia, ovarialperitoneum in caps.).

lentis (Gescheidt, 1833) Brand., 1892b, 510.—Stiles, 1902s, 24–35, pl. 3, figs. 2–5 (in eye of man); 1903u, 223.

lucaneum Brand., 1892b, 511. See also (Glenocerc.) lucanicum.

marænulæ (Rud., 1809) Brand., 1892b, 510 (in Coregonus marænula, ad ventric. in caps.).

præmorsum (Nord., 1832) Brand., 1892b, 511 (in Abramis brama, in regione branchiarum).

MONOSTOMULUM—Continued.

rhombi lævis (Wedl, 1855) Brand., 1892b, 511.

settenii (Naumann, 1840) Brand., 1892b, 511.

viviparæ (Linst., 1877) Brand., 1892b, 511 (in Paludina vivipara).

MONOSTOMUM Crep., 1829, v. 1, 49, for Monostoma Zed., 1800.

MONOSTROMUM Linst., 1904p, 254, misprint for Monostomum.

MONOSTUMUM Mont., 1888, 69, misprint for Monostomum.

MUSALIA Shipley & Hornell, 1904, 93–95 (m. herdmani), pl. 3, fig. 51, pl. 4, figs. 58, 59, 65.

herdmani Shipley & Hornell, 1904, 78, 90, 93–95, pl. 3, fig. 51, pl. 4, figs. 58, 59, 65 (in Margaritifera vulgaris; Ceylon).

MUTTUA Shipley & Hornell, 1904, 90-93 (m. margaritiferæ).

margaritiferæ Shipley & Hornell, 1904, 78, 90–93, pl. 3, figs. 53–57 (in Margaritifera vulgaris; Ceylon).

NEMATHOBOTHRIUM Mont., 1892, Oct. 7, 214 (g. of Didymozoonidæ) (for Nematobothrium).

NEMATOBOTHRIUM Ben., 1858a, 1861a, 11, 107–111, 200, 344, 345 (m. filarina; also by virtual tautonymy).—Braun, 1892a, 567, 569, 574, 577, 665, 696, 698, 699, 700, 707, 711, 727; 1893a, 878, 879, 886, 887, 890, 895, 914, 918; 1895b, 126, 136.—Gamb., 1896, 4, 73.—Hoyle, 1890, 539.—Jackson, 1888, 654.—Maclaren, 1904, 612, 613.—Moniez, 1891, 184–187.—Mont., 1888, 7, 18, 34, 52, 84, 93, 106; 1892, 6; 1892, Oct. 7, 214 (g. of Didymozoonidæ) (Nemathobothrium); 1893, 137.—Pratt, 1902, 890, 909.—Schneidemuehl, 1896, 295.—Tasch., 1879, 607, 614; 1879, 72.

benedeni (Mont., 1893) Maclaren, 1904b, 613, to (Didymozoon).

filarina Ben., 1858a, 1861a, 108–109, pl. 13. figs. 1–12 (t. h. Sciæna aquila) (filarinum in descr. of plates); 1870c, 136–143, pl. 8, figs. 1–9 (embryonic form of); 1870, 32.—Braun, 1892a, 586, 784; 1893a, 914.—Cobbold, 1879b, 462.—Dies., 1859c, 426 (to Monost.).—Gamb., 1896a, 55, fig. 22A.—Jackson, 1888, 654.—Maclaren, 1904b, 602, 603, 606, 612, 613.—Mont., 1888, 9; 1893, 137.

filarinum Ben., 1858, 1861a, descr. of plates, see filarina.

guernei Moniez, 1891, 184–187 (in Thynnus alalonga; Europe).—Braun, 1891d, 423; 1893a, 914.—Maclaren, 1904b, 603, 613.

molæ Maclaren, 1904b, 573, 602–613, pl. 21, figs. 17, 19, 21, pl. 22, figs. 23–33 (in Orthagoriscus mola) (Nemathobothrium); 1904, June 9, 443–444; 1905, Jan. 31, 20–29; 1905, June, 317; 1905, Dec., 703.—Ziegler, 1905, 37.

txnioides (Mont., 1888) Maclaren, 1904b, 613, to (Didymozoon).

NEOPHASIS Staff., 1904, May 3, 485–486 (m. pusilla, νέος=new, φάσις=appearance). pusilla Staff., 1904, 485–486 (in Anarrhichas lupus; Canada).

(NEPHROCEPHALA) Dies., 1858d, 260-264, subg. of Cerc. (Hormocerc.), renamed, hence type probably echinatoides. [See, however, infra, p. 385.]

echinata (Sieb., 1837) Dies., 1858d, 260–262 (syn. Cerc. (Hormocerc.) echinata Sieb.) (in Paludina vivipara, Planorbis corneus, Lymnæus stagnalis), larva of Dist. echinatum Zed.); 1858e, 344 (to Dist.).

cchinatoides (Fil., 1854) Dies., 1858d, 262–263 (syns. Cerc. (Hormocerc.) echinatoides Fil., Cerc. echinifera La Valette, Dist. echiniferum Pag.) (in Paludina achatina, P. vivipara); 1858e, 346 (syn. of Dist. echiniferum La Valette).

megacotyla Dies., 1858d, 263 (syn. Dist. echinatoides Pag. nec Fil.) (in Anodonta cygnea; Heidelberg).

spinifera (La Valette, 1855) Dies., 1858d, 263–264 (in Planorbis corneus; Berlin).

NEPHROCEPHALÆ Dies., 1858d, 260.

NEPHROCEPHALUS Odhn., 1902, 27–29 (m. sessilis), not Nephrocephala, a 1858; 1902, 42.—Pratt, 1902, 889, 907.

sessilis Odhn., 1902, 27–29 (in Nilkrokodil).

NEURONAIA Goodsir, (1844) (m. monroi).—Braun, 1889a, 341 (Gasterost.); 1893a, 844.

lampetræ Gulliver [1870, 849–850], 1872, 103, 425 (in Planer's lamprey; Stour River, Canterbury.—Brown, 1899a, 490 (in lamprey).

^a In our opinion (Nephrocephala, 1858) does not preoccupy Nephrocephalus, 1902. 8588—No. 37—08——21

NEURONAIA—Continued.

monroi Goodsir, (1844) (in Gadus morrhua).—Brown, 1899a, 490 (munroi).—Gulliver, 1872, 103, 425 (monroii).

monroii Goodsir teste Gulliver, 1872, 103, 425.

munroi Brown, 1899a, 490, for monroi.

NEURONAINA Brown, 1899a, 490 (see Neuronaia).

NITSCHIA Haswell, 1892a, 458 (for Nitzschia).

NITZCHIA Ben., 1858a, 1861a, 11, 38, for Nitzschia.

hippoglossii Ben., 1858a, 1861a, 21.

NITZSCHIA Baer, 1826a, 125 (m. elegans for Hirudo sturionis Abildg, renamed elegans) [not Nitzschia Denny, 1842, insect]; 1827b,675-676, pl. 32, figs. 1-6.— Ben., 1858a, 1861a, 11, 38 (Nitzchia).—Ben. & Hesse, 1864, 64, 65, 66, 67-68.— Blainv., 1828a,567-568.—R.Bl., 1888, 130.—R. Brand., 1891d, 9.—Braun, 1890a, 411, 412, 414, 423, 425, 435, 436, 440, 450, 451, 466, 474, 484, 485, 511, 515, 516, 517, 519, 523, 526, 527; 1890e, 597; 1891d, 422; 1893a, 889.—Burm., 1837a, 530; 1856a, 251.—Dies., 1850a, 290, 425-426 (syns. Hirudo Abildg., Trist. Nitzsch); 1858e, 313, 363 (mentions only elegans).—Gamb., 1896, 73.—Goldb., 1855, 19.—Haswell, 1892a, 458 (Nitschia).—Johnston, 1865, 30; 1865, 33.—Massa, 1906, 43, 48.—Mont., 1888, 10, 13, 83, 86, 87, 97; 1891, 100, 105, 107, 126; 1892, Oct. 7, 213 (g. of Tristominæ); 1893, 8, 210; 1899, 98, 99; 1903, 335 (subf. Ancyrocotylinæ; f. Tristomidæ); 1904, 117, 122.—Nord., 1832a, 60.—Pratt, 1900, 646, 648, 658.—Tasch., 1878, 564, 566 (syn. of Trist.); 1879, 233.

[dubius see Kellogg, 1876c, 544, arthropod.]

elegans Baer, 1826a, 125–126 (Hirudo sturionis Abildg., renamed); on gills of Stör; 1827b, 660–678, pl. 32, figs. 1–6 (includes Trist. elongata, 1826).—Abildg. (1797), 135, pl. 3, figs. 3–5; 1794b, 55, pl. 6, fig. 1 a. b. c.—Ben. & Hesse, 1864,, 67–68 (syns. Hirudo sturionis; Phylline, Trist. elongatum, T. sturionis).—Blainv., 1828a, 568.—E. Bl., 1847, 323.—Braun, 1889h, 433–434 (syn. of Trist. elongatum Nitzsch); 1889k, 612.—Crep., 1846, 149.—Cuv., ——Dies., 1850a, 426 (syns. Hirudo sturionis Abildg., Trist. elongatum Nitzsch, Capsala elongata Nord., Trist. sturionis Cuv. (in Accipenser sturio, A. guldenstædtii, A. acutirostris; Scotia); 1858e, 363.—Duj., 1845a, 323 (syn. of Trist. elongatum).—Johnston, 1865, 33–34, 299.—Leidy, 1887, 24.—Linst., 1903, 280.—Lint., 1898c, 508; 1901, 408, 414, 435.—Mont., 1904, 117; 1905, 117.—Moquin-Tandon, 1846a, 394.—Mueh., 1898, 11, 17.—[Nitzsch, 1826, 150–151.]—Nord., 1832a, 66; 1840, 602 (syn. of Capsala elongata).—Oken, —, 371.—Tasch., 1878, 563, 564, 565, 568 (syn. of Trist. elongatum Nitzsch); 1879, 56.

elongata (Nitzsch, 1826) Johnston, 1865, 34.—Braun, 1890a, 411, 419, 422, 424, 427, 428, 430 (elongatum), 431, 434, 437, 440, 445, 449, 450, 453, 456, 464, 472, 479, 489, 494, 497, 512, 527 (in Acipenser sturio), 547, 550; 1890e, 597, to (Trist.); 1892a, 667.—Gamb., 1896a, 56.—Kath., 1894a, 154.—Lint., 1901, 408, 414, 435 (in Acipenser sturio; Woods Hole, Mass.).—Mont., 1891, 106, pl. 6, fig. 32.—Pratt, 1900, 655, fig. 4; 657, 659.

hippoglossi (Mueller, 1776) Tasch., 1878, 568 (to Trist.) (for hippoglossii).

hippoglossii Ben., 1858a, 1861a, 21 (to Epibdella).—Tasch., 1878, 568 (hippoglossi) (to Trist.).

[linearis, a diatom.]

[palea, a diatom.]

papillosa Lint., 1898c, 508-509, pl. 40, figs. 1-6 (in Gadus callarias) [type U. S. N. M. 4874]; 1901, 414, 476 (in Gadus callarias; Woods Hole, Mass.).—Mont., 1904, 117 (to Lintonia); 1905, 70; 1905, 117 (to Lintonia as type).—Pratt, 1900, 659.

[pulicaria, a louse on the swift, Chætura pelagica.]

[sigmoidea, a diatom.]

sturionis (Abildg., 1794) Kroyer, 1852–53a, 777 (in Acipenser sturio L.).

[subtilis, a diatom.]

NOCOTYLUS Nord., 1840, 602 (for Notocotylus).

NOTOCOTYLE Dies., 1850a, 288, 411, for Notocotylus 1839 (m. triseriale) (syns. Fasc. Freelich, Festucaria Schrank, Monost. Zed., Notocotylus Dies.); 1858e, 314, 369; 1859c, 437–438.—Brand., 1891d, 22; 1892, 506, 508.—Braun, 1890a, 515; 1893a, 887, 890, 891, 893, 895, 914, 916, 918; 1893b, 179, 185; 1893d, 466; 1901b, 51, 52; 1901e, 346.—Cohn, 1904, 230.—Gamb., 1896a, 73.—Goldb.,

NOTOCOTYLE—Continued.

1855, 17.—Hoyle, 1890, 539 (N. triseriale Dies.—Monost. verrucosum).—Jackson, 1888, 642 (cf. Monost. verrucosum).—Looss, 1894a 131, 204; 1896b. 147, 8801, 1885, 661-664, 665, 667; 1901, 192 (cf. Monost. verrucosum); 1902m, 546, 602, 610, 612, 639, 701.—Luehe, 1901, 174.—Mont., 1888, 84, 93, 106; 1892m, 24-46, pl. 1. figs. 1-19; —— 126; 1892, Oct. 7, 182, 183, 214 (gen., cf. Monostomidæ); 1892; 685, 686, 687, 688, 689, 690, 691, 692, 693, 694, 695, 696, 697, 698, 699, 700, 702, 703, 704, 705, 706, 708, 709, 713, 714; 1893, 15, 24, 27, 28, 52, 83, 84, 91, 105, 108, 144, 205; 1896, 151.—Odhm., 1905, 367, 368, 370.—Ofenheim, 1900, 156, 182. Sciptific 1806, 111, 120, 121, (ip. birdest Worschau). Steep 1900, 156, 183.—Ssinitzin, 1896, in 1-20, 1 pl. (in birds at Warschau).—Stoss., 1902, 5.—Tasch., 1879, 234.

ægyptiaca Odhn., 1905, 370 (Monest. verrucesum of Loss, 1896, renamed).

alveatum (Mehlis, 1845) Mont., 1892, 39, 41, pl. 1, fig. 8 (in Anas penelope, Fuligula marila, Harelda glacialis, Oidemia fusca, Somateria mollissima).—Braun, 1893a, 916; 1893b, 185.

discrialis Ssinitzin (1896) (in Rallus aquaticus).

proteus (Brand., 1891) Mont., 1892, 703; 1892, 30.

triseriale (Dies., 1839) Dies., 1850a, 411-412 (syns. Fasc. verrucosa Frælich, F. anseris Gmelin, Festuc. pedata Schrank, Monost. verrucosum Zed., M. marillæ Rud., M. lineare Rud.); 1858e, 369 (in Anas anser dom. et ferus, marillæ Rud., M. lineare Rud.); 1858e, 369 (in Anas anser dom, et ferus, Hæmatopus ostralegus, Gryphiæ: Fulica atra); 1859c, 437 (syn. Monost. verr.) (in Anas tadorna, Planorbis. Lymnæus: Belgium).—Ben., 1858a, 1861a, 78 (syn. of Monost. verr.).—Brand., 1891d, 22 (triserialis).—Braun, 1892a, 581; 1893a. 916; 1893b, 177.—Hass., 1896a. 3 (syn. of N. verr. (Frælich)).—Hoyle, 1890, 539 (= Monost. verr.).—Kowal., 1895g, 41 (1).—Landois, 1882, 23 (triserialis).—Looss, 1894a, 204; 1896b. 146 (syn. of Monost. verr.).—Mont., 1892, 26, 40 (syn. of N. verr. Frælich).—Odhn., 1905, 367 (syn. of Catatropis verrucosa, type of Catat.); 1905, 370.—Rail., 1893a, 340 (= N. verr.).—Also reported for Anas querquedula, Anser albifrons, A. cinereus, A. cinereus dom., A. segetum, Fuligula ferina, Harelda glacialis).

verrocosum Mont., 1892, 40 (for verrucosum).

verrucosum (Frœlich, 1789) Mont., 1892, 28, 29, 30, 39, 40 (verrocosum), 41, pl. 1, figs. 1-7, 9-16 (syns. Monost. verr. Zed., M. attenuatum Rud., M. ovatum Mol., Notocotyle triseriale Dies.) (in Anas bewikii, A. boschas dom., A. boschas fer., A. penelope, A. querquedula, Anser albifrons. A. cinereus ferus. A. cinereus dom., A. leucopsis, A. segetum, Ascolopax gallinago. Cairina moschata, Crex pratensis. Cygnus musicus, Fulica atra. Fuligula cristata, F. ferina, F. marila, Gallinula chloropus. Gallus gallinaceus. Glaucio clangula. Hæmatopus ostralegus, Harelda glacialis, Mergus merganser, M. serrator, Oidemia fusca, Ortygometra porzana, Rallus aquaticus, Rhynchaspis clypeata. Somateria mollissima, Tadorna vulpanser, Vanellus cristatus): 1892, 689, 690, 691, 703, 709: 1893, 24, 51.—Braun, 1893a, 916: 1893b, 185.—Kowal., 1896d, 254 (4) (in Anas boschas, A. crecca, Fuligula marila: Dublany).—Hass. 1896a, 3 (syns. Fasc. verr. (Frœlich), Monost. verr. (Frœlich), M. attenuatum Rud., M. lineare Rud., Notocotylus triserialis Dies., Notocotyle triseriale (Dies.)) (in Gallus dom.).—Looss, 1896b, 146 (syn. of Monost. verr. Fræ.): 1899b, 661, 662, 663, 664, 770: 1902m, 444; (verrucosa Cerc. imbricata Looss), 807.—Odhn., 1902, 63 (verucosa).—Olss., 1893, 12–13.—Rail., 1893a, 340.—Stoss., 1901, 92 (4) (in Rallus aquaticus; Triest).

verucosa Odhn., 1902, 63 (for verrucosum).

NOTOCOTYLUS Dies., 1839a, 234 (m. triserialis).—Braun, 1893a, 884.—Cobbold, 1877, 237, 238; 1877f, 326; 1879b, 360.—Mont., 1892, 26 (syn. of Notocotyle).-Moul., 1856a, 16.—Nord., 1840, 601-602 (Nocotylus).—Pratt, 1902, 890, 910.

triserialis Dies., 1839a, 234–235 (syns. Fasc. verrucosa Frœlich. F. anseris, Fest. pedata Schrank, Monost. verrucosum Zed.) (in Anas anser dom., A. anser, A. querquedula, A. boschas dom., A. marila, A. penelope, A. albifrons, Cygnus bewikii, Fulica atra. Gallinula porzana, Phasianus gallus, Rallus aquaticus: Europe), pl. 15. figs. 23–25; 1850a, 411, to Notocotyle; 1858e, 369 (in Anas anser dom. et ferus, Hæmatopus ostralegus, Greifswald; Fulica atra).—Ben., 1858a, 1861a, 78 (syn. of Monost. verrucosum).—Crep., 1849a, 68.—Hass., 1896a, 3 (syn. of N. verr. (Frælich)).—Nord., 1840, 602 (Nocotylus) (syns. Fasc. verr., F. anseris. Monost. verr.).—Odhn., 1905, 366 (syn. of Catatropis verrucosa (Frœlich)).—Rail., 1893a, 340.

OCBOTRYUM Moul., 1856a, 10, misprint for Octobothrium.

OCHETOSOMA Braun, 1901g, 944 (m. monstruosum); 1902b, 64, 65, 67.—Pratt, 1902, 888, 902.

monstruosum Braun, 1901g, 944 (in Corone venustissima; West Indies); 1902b, 65.

OCREATA Gœze, 1782a, 41, 182–183, a "Gattung" of Fasciola Gœze, from a Maulwurf. OCTANGIINÆ Looss, 1902m, 699.

OCTANGIUM Looss, 1902m, 433, 633, 634, 637, 642, 645, 649, 650, 652, 653, 657, 660, 668, 670, 674, 675, 682, 684–685, 686, 687, 698, 699, 824 (tod. sagitta), family Angiodictyidæ.

hasta Looss, 1902m, 686-687, 690, 697, pl. 29, figs. 128-132, 133b (in Chelone mydas: apparently Egyptian coast).

sagitta (Looss, 1899) Looss, 1902m, 416, 633, 636, 685-686 (syns. Monost. proteus ohne Geschlechtsproducten, of Walter, 1893, 197, pl. 10, figs. 13, 18, 19, etc., Microscaphidium sagitta Looss, 1899), 687, 697, pl. 29, figs. 122-127, 133a, pl. 30, figs. 143, 144.

OCTOBOHTRINÆ Mont., 1903, 336, misprint for Octobothrinæ (f. Octocotylidæ).

OCTOBOTHRIDÆ Mont., 1888a, 86, 88, for Octobothriidæ.

OCTOBOTHRIDEA Tasch.—Olss., 1893, 4, subf. to contain Octobothrium, Diplozoon, Diporpa.

OCTOBOTHRII E. Bl., 1847, 330–331 (for Onchobothriens of Duj., 1845a, 312) (tribe of Trematoda, contains Axine, Diplozoon, Diporpa, Octobothrium, Polyst.).

OCTOBOTHRIIDÆ Tasch., 1879, 235, 236, 238 (syn. Octocotylidæ Ben.); 1879, 68 (syn. Octocotylidæ Ben.).—Braun, 1890a, 516, 517, 533, 538.—Hoyle, 1890, 539 (Octobothrium, Anthocotyle, Phyllocotyle, Platycotyle, Pleurocotyle, Diplozoon, Hexacotyle, Plectanocotyle).—Jackson, 1888, 654 (Octobothrium, Diplozoon).—Mont., 1888a, 86 (Octobothridæ), 88.

OCTOBOTHRINÆ Mont., 1903, 336.

OCTOBOTHRIUM F. S. Leuck., 1827a, 24 (m. lanceolatum); 1828, 18 (nec Dies., 1850a, Cestode) (m. lanceolatum); 1842a, 18, 29 (syns. Mazocraes, Octost.).—
Ben., 1856e (Octobothrium du Merlan); 1858a, 1861a, 11, 44-45, 57, 193, 197, 345; 1868, 22.—Ben. & Hesse, 1864, 61, 102.—Braun, 1889k, 620, 621; 1890a, 445, 451, 469, 471, 477, 480, 489, 490, 515, 516, 517, 518, 522, 523, 534; 1891d, 422; 1895a, 1204; 1900a, 1654, 1657, 1671, 1672, 1679, 1684, 1705, 1726.—Burm., 1837a, 530.—Carus, 1863, 477-478.—Cerf., 1895b, 916, 920, 921; 1895m, 139; 1896, 516, 546; 1899a, 391.—Cobbold, 1879b, 35, 41.—Crep., 1838, 84; 1839, 291.—Dies., 1850a, 417 (of Nord., syn., of Diclidophora), 419 (of Nord., syn. of Cyclocotyle), 421 (syn. of Octocotyle Dies.), 423 (syn. of Discocotyle Dies.), 480, 603 (Dies., n. g., cestode, m. rostellatum); 1863, 284.—Duj., 1845a, 312-313, 314-315, O. (Cyclocotyle).—Gamb., 1896a, 56, 73.—Goto, 1891a, 161, 174, 178, 183, 184; 1891c, 103; 1893a, 798.—Haswell, 1892b, 149; 1893e, 114.—Hoyle, 1890, 539 (of Nord.).—Ijima, 1884c, 638.—Jackson, 1888, 654.—Johnston, 1865a, 30, 31.—Kath., 1894a, 148.—Kroyer, 1843-45a, 166 (in Lota molva Cuv.).—R. Leuck., 1863a, 450, 615.—Looss, 1892, 72, 73.—Mayer, 1841a, 25, 33.—Mont., 1888, 83, 84, 88, 89; 1893, 111; 1903, 336 (syns. Glossocotyle [+Ophicotyle]) (subf. Octoboltrinæ).—Nord., 1832a, 69, 76-77; 1840, 599.—Par. & Perugia, 1896, in 135-138; 1896, 4 pp.; —, 653.—Pratt, 1900, 646, 652, 656, 666, fig. 33.—St., Remy., 1898, 523, 544-545.—Schneider, 1866, 334.—Scott, 1901, in 120-153; 1901, 145.—Tasch., 1879, 232, 233, 238, 239, 240, 241, 242, 243; 1879, 248 (of Grube) (syn. of Pleurocotyle Gerv. & Ben.); 1879, 35, 59, 69.

1856: Ochotryum Moul., 1856a, 10, misprint.

alosa Leuck., (1828).—Cerf., 1895h, 918, 920, 921, 922; 1895m, 140; 1896, 516, 517, 547.—Duj., 1845a, 313 (syn. of O. lanccolatum) (in Clupea alosa; Rennes).—Mayer, 1841a, 19, pl. 3, figs. 1–8.

alosæ (Ben. & Hesse, 1863) Braun, 1890a, 414, 418, 534, O. (Glossocotyle), 548 (Brest), 550 (in Alosa vulgaris).—Tasch., 1879, 244 (syn. Glossocotyle alosæ) (in Alosa vulgaris).—St.-Řemy, 1898, 545.

alosæ (Hermann, 1782) E. Bl., ——.—Ben. & Hesse, 1858a, 1861a, 46 (syn. of O. lanceolatum).—Scott, 1901, 8; 1901, 145, pl. 8, fig. 21 (in Clupea alosa, C. finta); 1905, 118.—Tasch., 1879, 244 (syn. of O. lanceolatum Leuck.).

alos x of Mayer, 1841a, 19, see lance olatum alos x.—Baird, 1853a, 40 (= O. lance olatum).

OCTOBOTHRIUM-Continued.

- arcuatum (Sons., 1890) Braun, 1890a, 534 (=Vallisia striata Par. & Perugia), 551 (in Lichia amia).
- bellones (Otto, 1823) Tasch., 1879, 245–246 (syn. Cyclocotyle bellones Otto) (in Bellone acus).—St.-Remy, 1898, 555.
- belones (Otto, 1823) Nord., 1840, 600 (for bellones) (syn. Cyclocotyla bellones Otto).—Dies., 1850a, 419 (to Cyclocotyle).
- bramæ Par. & Perugia, 1896, 1 (in Brama rayi; Genova).—St.-Remy, 1898, 548 (to Octocotyle).
- chrysophryi (Ben. & Hesse, 1863) Braun, 1890a, 418, 535, 548, 550.—Tasch., 1879, 247 (syn. Choricotyle chr. Ben. & Hesse) (in Chrysophrys aurata).
- denticulatum Olss., 1876, 10, pl. 1, figs. 13–17 (in Gadus virens).—Braun, 1890a, 418, 499, 514, 534, 548, 550.—Cerf., 1895h, 914, 921 (in Gadus carbonarius; Skagerrack); 1896, 510–511, 516.—Lint., 1900, 269, 283, 286, pl. 33, figs. 6–10; 1901, 408, 414, 474 (in Pollachius virens; Woods Hole, Mass.).—Par., 1899, 4; 1902, 3 (in Gadus minutus; Elba).—St.-Remy, 1898, 549 (to Dactylocotyle.)—Tasch., 1879, 246 (in Gadus virens).
- digitatum Rathke, (1843), 242–244, pl. 12, figs. 13–15 (in Hippoglossus gigas; Norway).—Ben., 1858a, 1861a, 50, 51.—Cerf., 1895h, 917 (syn. of O. palmatum); 1896, 513.—Dies., 1850a, 418 (syn. of Diclidophora palmata); 1858e, 384 (syn. of Dicl. pal.); 1859c, 443, to Octoplectanum.—Johnston, 1865, 31.—Tasch., 1879, 246 (syn. of Octob. pal.).
- [dipsacum Lint., 1897a, 806, a cestode.]
- esmarkii Scott, 1901, 147, pl. 8, fig. 22 (in Gadus esmarkii; Shetland); 1905, 118. fintæ (Ben. & Hesse, 1863) Tasch., 1879, 244 (type of Ophicotyle Ben. & Hesse) (in
- Alosa finta).—Braun, 1890a, 414, 418, 452, 534, 548, 550.—St.-Remy, 1898, 546. harengi (Ben. & Hesse, 1863) Tasch., 1879, 244 (in Clupea harengus).—Braun, 1890a, 414, 418, 452, 534, 548, 550.—St.-Remy, 1898, 545.—Scott, 1901, 145–146 (in Clupea harengus); 1905, 118.
- heterocotyle (Ben., 1870) Tasch., 1879, 245 (in Clupea sprattus).—St.-Remy, 1898, 555
- hirudinaceum Bartels, 1834, 61 (t. h. Salmo lavaretus; St. Petersburg).—Crep., 1839, 291.—Duj., 1845a, 315 (to Cyclocotyle).—Leuck., 1842, 30.—St.-Remy, 1898, 555.
- hirundinaceum Dies., 1850a, 424, for hirudinaceum (to Discocotyle) (in Coregonus wartmanni).
- lanceolatum Leuck., 1827a, 24 (in Clupea alosa; Germany); 1828, 18, pl. 1, fig. 7a-b (in Alosa vulgaris); 1842, 29.—Ben., 1858a, 1861a (of Duj.), 45-49, 50, 51, 53, 168, 169, 170, 172, 176, 186, 189, 190, 196, pl. 5, figs. 1-18 (syns. Mazocraes alosæ Hermann, Octob. alosæ Leuck., Octost. alosæ Kuhn, Octocotyle lanceolata Dies.); 1868, 5; 1868, 34, figs. 11-12.—Ben. & Hesse, 1864, 101.—Braun, 1890a, 407, 414, 419, 429, 431, 438, 439, 442, 448, 452, 453, 476, 483, 485, 489, 490, 494, 511, 534, 548, 550; 1891a, 52, 54.—Cobbold, 1872b, 90; 1879b, 41.—Crep., 1838, 84; 1839, 291.—Dies., 1850a, 422 (to Octocotyle); 1859c.—Duj., 1845a, 313, pl. 8, fig. F.—Fraip., 1880a, 400; 1880c, 416, 433; 1883a, xxxvi.—Jackson, 1888, 646, 649.—Kerbert, 1881a, 556.—Kroyer, 1846-53a, 218 (in Alosa finta Cuv.).—Looss, 1894a, 234.—Mont., 1893, 112.—Nord., 1832a, 58, 76, 77 (syns. Octost. alosæ Kuhn, Maz. alosæ Herm.); 1840, 599 (syns. Maz. alosæ Herm., Octost. alosæ Kuhn).—Par., 1896, 1.—St.-Remy, 1898, 545.—Tasch., 1879, 241, 242, 244 (syns. Maz. alosæ Herm., Octost. alosæ Kuhn, Detob. alosæ Kuhn.) E. Bl., Octocotyle lanceolata Dies., Octoplectanum lanceolatum Dies.) (in Alosa vulgaris).
- lanceolatum alosæ Mayer, 1841a, 19–25, 4 (lanceolatum), pl. 3, figs. 1–9, Hermann's species.
- leptogaster Leuck., 1830a, 612 (t. h. Chimæra monstrosa; Europe); 1842a, 22-23, 30, pl. 1, fig. 5, pl. 2, figs. 2a-b.—Braun, 1890a, 418, 514.—Dies., 1850a, 424 (to Discocotyle).—Duj., 1845a, 314-315 to (Cyclocotyle).—St.-Remy, 1898, 546.—Tasch., 1879, 245 (syns. Discocotyle lept. Dies., Placoplectanum lept. Dies.) (in Chim. monst.).—Wagener, 1852, 544.
- luscæ (Ben. & Hesse, 1863) Tasch., 1879, 247 (in Morrhua lusca).—Braun, 1890a, 418, 535, 551.

OCTOBOTHRIUM-Continued.

merlangi (Kuhn, 1830) Nord., 1832a, 76, 78–80, 81, pl. 7, figs. 1–5 (in Gadus merlangus), 115, pl. 7, figs. 1–5; 1840, 599.—Ben., 1856a, —; 1856e, —; 1857n; 1858a, 1861a, 49–52 (syn. Diclidophora longicollis Dies.) (in Gad. merl.).—Braun, 1889h, 621; 1891a, 52; 1890a, 410, 415, 419, 424, 452, 453, 534, 546, 548, 549, 550, 551; 1893b, 179.—Cerf., 1895h, 915–916 [920], 921 (syns. Octob. platygaster Leuck., Dicl. longic. Dies., Octoplectanum longic. Dies., Octob. squillarum Par. & Perugia) (in Gad. merl.); 1896, 511, 516.—Crep., 1838b, 84; 1839a, 291.—Dies., 1850a, 417 (renamed Diclidophora longicollis); 1858e, 384 (syn. of Dicl. longic. Dies.); 1859c, 443.—Duj., 1845a, 314 to (Cyclocotyle).—Gamb., 1896a, 56, fig. 23.—Scott, 1901, 146–147 (in Gad. merl.); 1901, 343 (in Gad. merl.; Irish Sea); 1905, 118 (Scott, 1895, pl. 4).—Tasch., 1879, 241, 242, 245 (syns. Dicl. longic. Dies., Octoplectanum longic. Dies.) (in Merlangus communis, Cymothoa œstroides).—Reported also for Box salpa, Boops sp., Bopyrus squillarum.

minus Olss., 1876, 10 (in Gadus melanostomus).—Braun, 1890a, 418, 514, 534, 548, 550.—Cerf., 1895h, 922; 1896, 517; 1898a, 302 (in Gad. mel.).—Tasch., 1879,

246 (in Gad. mel.).

morrhuæ (Ben. & Hesse, 1863) Tasch., 1879, 246 (in Gadus morrhua).—Braun, 1890a, 419, 535, 548, 550.

palmatum Leuck., 1830a, 612 (t. h. Gadus molva; Germany); 1842a, 24–28, 30, pl. 1, fig. 4 (in Gad. mol.).—Ben. & Hesse, 1864, 107 (to Pterocotyle).—Braun, 1890a, 419, 492, 494, 497, 514, 535, 548, 551.—Cerf., 1895h, 916–917, 921 (syns.: Octodactylus [type] inhærens Dalyell, Octob. digitatum Rathke, Diclidophora pal. Dies., Pterocotyle pal. Ben. & Hesse, Octoplectanum pal. Dies.) (in Gad. mol.); 1896d, 513, 516.—Dies., 1850a, 418 (to Diclidophora).—Duj., 1845a, 314 to (Cyclocotyle).—Johnston, 1865a, 31.—Kroyer, 1838—40a, 608 (in Lota molva L.).—Tasch., 1879, 241. 242, 246 (syns.: O. digitatum Rathke, Dicl. pal. Dies., Octoplectanum pal. Dies., Octodactylus inhærens Dalyell, Pterocotyle pal. Ben. & Hesse) (in Molva vulgaris, Hippoglossus gigas).—Reported also for Gadus merlangus.

phycidis (Par. & Perugia, 1889) Braun, 1890a, 418, 535, 548, 551 (in Phycis

blennoides).

pilchardi (Ben. & Hesse, 1863) Tasch., 1879, 244 (in Clupea pilchardus).—Braun, 1890a, 414, 418, 534, 548, 550.—St.-Remy, 1898, 546.

platygaster Leuck., 1842a, 30 (in Gadus merlangus).—Cerf., 1895h, 915 (syn. of O. merlangi); 1896a, 511.—Duj., 1845a, 314 (syn. of O. merlangi).

pollachii (Ben. & Hesse, 1863) Tasch., 1879, 246–247 (in Merlangus pollachius).— Braun, 1890a, 415, 418, 452, 535, 548, 551.—Par. & Perugia, 1891, 18, to (Dactylocotyle).

[rostellatum Dies., 1850a, 603 (syns. Tænia erythrini Fabricius, Halysis octolobata Zed., Tænia octolobata Rud.) (in Sebastes norvegicus); 1863, 284 (misprint Onchobothrium rostellatum Dies.).—Braun, 1895a, 204.—Kroyer, 1852–53a,

595?, 1261.—Luehe, 1899, 542.—Zschokke, 1903, 27, 28.]

sagittatum Leuck., 1842a, 49-57. pl. 2, fig. 5a-k (in Salmo fario).—Braun, 1890a, 518, 534, 548, 551.—Dies., 1850a, 424 (to Discocotyle as type); 1858e, 384 (syn. of Diclidophora palmata); 1859c, 443 (syns. Octoplectanum longicolle, O. palmatum).—Duj., 1845a, 314.—Hofer, 1903e, 38-39, 1 fig. (pernicious anemia).—Pratt, 1900a, 656, 657, 660 (key), fig. 33.—St.-Remy, 1898, 555.—Tasch., 1879, 241, 243-244 (syns. Cyclocotyle lanceolata, Discocotyle sag., Placoplectanum sag.) (in Salmo fario).—Also reported for Coregonus lavaretus, Salmo trutta.

scombri (Kuhn, 1829) Nord., 1832a, 76, 77–78 (in Scomber scomber); 1840, 599.—
Ben. & Hesse, 1864, 100 (of Grube to Pleurocotyle).—Braun, 1890a, 410, 418, 452, 514, 534, 548, 552.—Burm., 1837, 530.—Crep., 1838b, 84; 1839a, 291.—
Dies., 1850a, 422 (syn. of Octocotyle tunicata); 1858e, 385 (of Grube, renamed Grubea cochlear, type of Grub.).—Duj., 1845a, 313, pl. 8, fig. e.—Grube, 1855a, 137–140, pl. 6, figs. 1–3 (on Scomber scomber), see also Tetracotyle scombri.—Leuck., 1842a, 30.—Scott, 1901a, 344 (in Mackerel; Manx coast); 1901, 146, pl. 8, fig. 20 (in Scomber scombrus); 1905, 118.—Stoss., 1898, 12.—Tasch., 1878, 574, 575, 576 (of Grube, to Pleurocotyle); 1879, 241, 242, 243 (syn. Octocotyle truncata, Octoplectanum trunc., Octocotyle sc.) (in Sc. scombrus, S. colias; Naples). 248 (of Grube) (to Pleurocotyle).

smaris Ijima, in Goto, 1894a, 207, see sub Diclidophora.—Cerf., 1895m, 140; 1896,

547.

OCTOBOTHRIUM—Continued.

species Ijima, 1884c, 637.

species Braun, 1890a, 472.

squillarum (Par. & Perugia, 1889) Cerf., 1895h, 916, syn. of Octob. merlangi (Kuhn); 1896, 512.—St.-Remy, 1898, 555 (syn. of Dactylocotyle merlangi, teste Parona).

taschenbergii (Par. & Perugia, 1898) Braun, 1890a, 535, 548, 552 (in Sargus rondeletii).

thunninæ (Par. & Perugia, 1898) Braun, 1890a, 419, 534, 548, 552 (in Thynnus thunnina).

OCTOBOTHRYUM Fraip., 1883a, 36, for Octobothrium.

OCTOCOLYTE Mont., 1888a, 57 (for Octocotyle).

OCTOCOTYLE Dies., 1850a, 289, 421–422 (includes lanceolata [type by inclusion] and truncata); 1858e, 382–393, renamed Octoplectanum, (syns. Mazocraes, Octobothrium, Octostoma).—Ben. & Hesse, 1864, 96, 97.—Braun, 1890a, 477, 516, 517, 518, 522, 546; 1893a, 890.—Burm., 1856a, 251.—Cerf., 1895h, 918, 920; 1895m, 139; 1896, 514, 515, 516, 547; 1899a, 391.—Cunningham, 1897a, 279.—Goldb., 1855a, 19.—Goto, 1893a, 798; 1894a, 201.—Mont., 1888a, 7, 8, 11, 59, 66, 86, 89, 99; 1892, Oct. 7, 213 (g. of Octocotylinæ) (contains as subg.: Vallisia, Anthocotyle, Diplozoon, Hexacotyle, Phyllocotyle, Pletanocotyle, Platycotyle, Pleurocotyle); 1903, 336 (subf. Octocotylinæ).—Pratt, 1900, 646, 652 (key), 656, fig. 34.—St.-Remy, 1898, 523, 545, 546.—Stoss., 1898, 12.—Tasch., 1879, 239, 241, 242.—See Mazocraes 1782.

1858: Octoplectanum Dies., 1858e, 382 (Octocotyle renamed, hence same type).

1888a: Octocolyte Mont., 1888a, 57, misprint.

1894: Octocotyte: Par., 1894, 594, misprint.

appendiculata (Kuhn, 1829) Sons., 1890, 176.

arcuata Sons., 1890, 112–113 (on Lichia amia; [Mus. Pisa]); 1890, 137–139 (syn. of striata Par. & Perugia).—Par. & Par., 1891, 17.

bramæ (Par. & Perugia, 1896) St.-Remy, 1898, 547-548 (in Brama rayi).

emarginata (Olss., 1876) Sons., 1890, 176.

harengi Ben. & Hesse, 1863, 1864, 98–99, pl. 9, figs. 1–10 (in Clupea harengus).— Braun, 1890a, 498.—Mont., 1888a, 13.—Tasch., 1879, 244 (to Octobothrium).

lanceolata (Leuck., 1828) Dies., 1850a, 422 (includes Mazocraes alose Hermann, Octobothrium lanceolatum Leuck., Octostoma alose), in Alosa vulgaris; 1858e, 383 (to Octoplectanum); 1859c, 442 (to Octoplectanum).—Baird, 1853a, 40.—Ben., 1858a, 1861a, 44, 46 (to Octobothrium).—Cerf., 1895m, 139; 1896, 547.—Hausmann, 1897b, 4, 7, 20, 23 (in Alosa vulgaris).—Mont., 1888a, 13, 34; 1888, 26 (lanceolatus); 1893, 111.—Tasch., 1879, 244 (to Octobothrium).

lanceolatus Mont., 1888a, 26, for lanceolata.

leptogaster (Leuck., 1830, 1842) Par. & Perugia, 1890, 6, to (Octobothrium); 1892,
 2-5 (87-90), pl. 3, figs. 7-8.—Par., 1894, 135 (in Chimæra monstrosa; Nizza).—
 Wagener, 1852, 544.

major Goto, 1894a, 203–205, pl. 9, figs. 1–6 (on gills of Scomber colias; Misaki); 1899, 274, 275, 276.—Cerf., 1895m, 140 (in Sc. col.); 1896, 547.—St.-Remy, 1898, 546–547.

majus St.-Remy, 1898, 547, misprint for major.

merlangi (Kuhn, 1829) Mont., 1888a, 11, 26; 1890, 420; 1893, 111.—Par., 1894, no. 727; 1894, no. 1060.

minor Goto, 1894a, 205–206; pl. 9, figs. 7–13 (on gills of Scomber colias; Japan); 1899, 274, 275, 276.—Cerf., 1895m, 140 (in Sc. col.); 1896, 547.—St.-Remy, 1898, 547.

pilchardei Mont., 1888a, 66, misprint for pilchardi.

pilchardi Ben. & Hesse, 1863, 1864a, 99, pl. 9, figs. 29–35 (on Clupea pilchardus).— Braun, 1890a, 498.—Mont., 1888a, 13, 66 (pilchardei).—Tasch., 1879, 244 (to Octobothrium).

scombri (Kuhn, 1829) Ben. & Hesse, 1864a, 97–98 (on maquereau).—Cerf., 1895m,
140 (in Scomber scombrus); 1896, 547.—Goto, 1899a, 274–276.—Kroyer,
1838–40a, 595 (in Sc. sc.).—Mont., 1890, 421.—Par. & Perugia, 1889, 742, fig. 3
(in Sc. sc.; Genova); 1890, 742, fig. 3; 1890, 6.—Par., 1894, 503.—Pratt, 1900,
656, 657, 660, fig. 34.—St.-Remy, 1898, 546.—Staff., 1904, 482 (in Sc. sc.;
Canada).—Stoss., 1892, 66; 1898, 12.—Tasch., 1879, 243 (to Octobothrium).—
Also reported for Sc. colias.

OCTOCOTYLE-Continued.

species Mont., 1888a, 57, 58.

striata (Par. & Perugia, 1890) Sons., 1890, 137–139 (syn. O. arcuata Sons.) (in Seriola dumerilii, Pisa; Lichia amia, Pisa and Triest); 1891, 9. März, 87–88, to (Vallisia).

thunninæ Par. & Perugia, 1889, 742–743, fig. 4 (in Thynnus thunnina; Genova); 1890, 742–743, fig. 4 (in Th. thunnina; Genova); 1890, 6.—Par., 1894, 594 (Octocotyle).—St.-Remy, 1898, 557.

thynni Par. & Perugia, 1891, 19 [lapsus for thunninæ?].

truncata Dies., 1850a, 422 (Octostoma scombri, Kuhn, 1829, renamed) (t. h. Scomber scombrus; Rhedoni); 1858e, 383 (to Octoplectanum).—Cerf., 1895m, 139, [140]; 1896, 547.—Stoss., 1898, 12.—Tasch., 1879, 243 (to Octobothrium).

OCTOCOTYLIDES Cerf., 1900, 436–438.

OCTOCOTYLINÆ Braun, 1893a, 890.—Gamb., 1896, 73.—Mont., 1892, Oct. 7, 213 (subf. of Polystomidæ): 1903, Dec., 336 (f. Octocotylidæ).—Pratt, 1900, 651 (includes: Octocotyle, Octobothrium, Dactylocotyle, Diclidophora, Anthocotyle, Vallisia, Diplozoön, Phyllocotyle, Hexacotyle, Plettanocotyle, Platycotyle, Pleurocotyle).—St.-Remy, 1898, 523, 544.—Scott, T., 1901, 141.

OCTOCOTYTE Par., 1894, 594, misprint for Octocotyle.

thunninæ (Par. & Perugia, 1889) Par., 1894, 594.

OCTODACTYLUS Dalyell, 1853a, 262 (m. inhærens).—Braun, 1890a, 518.—Goto, 1893a, 798.—Tasch., 1879, 239.

inhærens Dalyell, 1853a, 262–263, pl. 36, figs. 1–2 (on the ling).—Ben. & Hesse, 1864a, 107 (syn. of Pterocotyle palmata Ben. & Hesse).—Braun, 1889a, 348 (is Octobothrium).—Cerf., 1895h, 917 (syn. of Octobothrium pal.); 1896, 513.—Johnston, 1865a, 31.—Tasch., 1879, 246 (syn. of Octob. pal. Leuck.).—Reported for Gadus morrhua.

OCTOPLECTANUM Dies., 1858e, 315, 382–383 (Octocotyle, 1850, renamed, hence lanceolata=alose (Mazocraes), type by renaming and inclusion); 1859, 442.—Braun, 1890a, 518.—Tasch., 1879, 239.—See Mazocraes.

affine Lint., 1898c, 511–512, pl. 40, figs. 10–13, pl. 41, figs. 1–5 (in Paralichthys dentatus; Woods Hole, Mass.).

harengi (Ben. & Hesse, 1863) Linst., 1889, 95, in Clupea harengus.

heterocotyle (Ben., 1870) Linst., 1885, 252–253, pl. 15, fig. 30; 1889, 96, in Clupea sprattus Bl.

lanceolatum (Leuck., 1828) Dies., 1858e, 383 (in Alausa vulgaris); 1859, 442–443 (includes Mazocraes alosæ Hermann, Octobothrium lanc., Octost. alosæ Kuhn, Octocotyle lanc.,).—Tasch., 1879, 244 (to Octobothrium).—Type by renaming and inclusion.

longicolle (Dies., 1850) Dies., 1859c, 443.—Cerf., 1895h, 915 (syn. of Octobothrium merlangi); 1896, 512.—Tasch., 1879, 245 (syn. of Octob. merl. (Kuhn)).—Reported for Merlangus pollachius, M. vulgaris, Morrhua barbata.

palmatum (Leuck., 1830) Dies., 1859c, 443.—Cerf., 1895h, 917 (to Octobothrium); 1896, 513.—Tasch., 1879, 246 (to Octob.).—Reported for Hippoglossus gigas, Lota molya.

pilchardi (Ben. & Hesse, 1863) Linst., 1889, 95 (in Clupea pilchardus).

truncatum (Dies., 1850) Dies., 1858e, 383 (in Scomber scombrus; Rhedoni); 1859c, 443–444.—Stoss., 1898, 12.—Tasch., 1879, 243 (syn. of Octobothrium scombri (Kuhn)).

OCTOSTOMA Otto, 1823, 302, as syn. of Cyclocotyla [not Octostoma syn. of Polyst.].

OCTOSTOMA Kuhn, 1829b, 358–363 (alosæ=lanceolata=alosæ (Mazocraes), type by inclusion, also type by first species rule).—Braun, 1890a, 518.—Cerf., 1899a, 349.—Mayer, 1841a, 20.—Tasch., 1879, 239.

alosæ Kuhn, 1829b, 358-361 (in Clupea alosa), pl. 17, figs. 1-3.—Ben., 1858a, 1861a, 44, 46 (syn. of Octobothrium lanceolatum).—Cerf., 1899a, 348, 349.—Duj., 1845a, 313 (=Octob. lanc.).—Nord., 1832a, 76, 77, 82 (syn. of Octob. lanc.).—Tasch., 1879, 241, 244 (syn. of Octob. lanc.).

heterocotyle Ben., 1870, 67 (in Clupea sprattus).—Tasch., 1879, 245 (to Octobothrium).

merlangi Kuhn, (1829b) (t. h. Merlangus communis).—Ben., 1858a, 1861a, 44, 49, 168, 169, 172, 196 (to Octobothrium).—Cerf., 1895h, 915 (to Octob.); 1896, 511; 1899a, 349.—Crep., 1839a, 291.—Duj., 1845a, 314 (to Octob.).—Nord., 1832a, 78–80, 82 (to Octob.) (in Gadus merlangus); 1840, 599 (to Octob.).—Scott, 1901, 343 (to Octob.).—Tasch., 1879, 245 (to Octob.).

scombri Kuhn, 1829b, 361–362, pl. 17, figs. 4–5 (t. h. Scomber scomber).—Cerf.,
1899a, 348, 349.—Duj., 1845a, 313 (to Octob.).—Nord., 1832a, 77–78 (to Octob.); 1840, 599 (to Octob.).—Scott, 1901, 344 (to Octob.).—Stoss., 1898,
12 (in Sc. scombrus; Triest).—Tasch., 1879, 243 (to Octob.).

OGMOGASTER Jægers., 1891, 131 (m. plicatus); 1891b; 1892a.—Brand., 1892, 506.-Braun, 1892a, 605, 609, 641, 664, 671, 676, 677, 683, 684, 685, 699,704, 708, 709, 711, 715, 717, 718, 721, 725, 726, 729, 737; 1893a, 887, 890, 893, 895, 914, 916, 918; 1893b, 179.—Gamb., 1896a, 73.—Lander, 1904a, 5.—Looss, 1899b, 664–665; 1902m, 610.—Mont., 1892, Oct. 7, 214 (g. of Monostomidæ); 1893, 685, 686, 687, 688, 689, 690, 691, 692, 693, 694, 695, 696, 697, 698, 699, 701, 702, 703, 704, 705, 713; 1893, 28, 2144, 202, 205. 702, 703, 704, 705, 713; 1893, 28, 83, 144, 203, 205.—Ofenheim, 1900, 156.—Pratt, 1902, 890, 910 (key).—Rossbach, 1906, 374.

plicatus (Crep., 1827) Jægers., 1891, 129, 132, 134; 1891b, 32 pp., figs. 1-6; 1892a, 572–573.—Bettend., 1897, 319; 1897a, 15.—Braun, 1892a, 575, 600, 603, 607, 608, 609, 638, 640, 641, 647, 682, 688, 712, 717, 718, 719, 724, 731, 733, 765; 1893a, 917 (in Balænoptera musculus, B. borealis); 1893b, 177 (plicata), 186; 1901e, 344.—Darr, 1902, 657.—Looss, 1899b, 665.—Mont., 1892, 685, 689; 1893, 203.—Odhn., 1905, 366 (Monost. plicatum).

OISTOSOMUM Odhn., 1902, 26-27 (m. caduceus); 1902, 42.—Pratt, 1902, 888, 899. caduceus Odhn., 1902, 26-27 (in Krokodil; Sudan).

OLIGOCOTYLIDÆ Par. & Perugia, 1890, subf.—Braun, 1890a, 546.—Cerf., 1899a, 365, 452.

OLYOCOTYLEA Mont., 1903, 335-336 ("Sezione" contains: Tristomidæ Ben., 1858, Monocotylidæ Tasch., 1879).

OMPHALOMETRA Looss, 1899b, 562 (m. flexuosa) (ὁ ὀμφαλικός=Nabel or Mitte; $\dot{\eta}$ μήτρα=uterus); 1902m, 839.—Braun, 1901b, 33.—Pratt, 1902, 887, 896.— Stoss., 1901, 93 (5).

flexuosa (Rud., 1809) Looss, 1899b, 562.—Stoss., 1901, 93 (5).

OMPHALOMETRINÆ Looss, 1899b, 543, 562; 1902m, 839.—Luehe, 1900, 490.— Pratt, 1902, 887, 896 (includes: Omphalometra, Cathæmasia).—Stoss., 1901, 93 (5).

ONCHOCOTILINÆ Cerf., 1899a, 396 (for Onchocotylinæ).

ONCHOCOTYLE Dies., 1850a, 289, 419 (m. appendiculata); 1858e, 314, 370–371; 1859e, 438.—Ben., 1853b, 59–68, figs. 1–11; 1858a, 1861a, 11, 54, 345.—Ben. & Hesse, 1864, 84.—Brand., 1891d, 21; 1894a, 308.—Braun, 1890a, 413, 415, 434, 437, 438, 444, 451, 471, 473, 474, 475, 484, 490, 491, 511, 515, 516, 517, 523, 538, 539, 546; 1893a, 890.—Carus, 1863, 478.—Cerf., 1899a, 345, 346, 347, 351, 359, 365, 368, 369, 371, 402, 411, 434, 452.—Chatin, 1880f, 591; 1881a, 311.—Gamb., 1896a, 73.—Goldb., 1855a, 19.—Goto, 1893a, 798, 800; 1894a, 222-224.—Haswell, 1887a, 286.—Hoyle, 1890, 539.—Jackson, 1888, 647, 654.-Kerbert, 1881a, 544, 554, 572.—Looss, 1894a, 203.—Mont., 1888a, 11, 16, 34, 36, 38, 43, 52, 53, 56, 57, 58, 59, 60, 65, 66, 67, 84, 86, 100; 1892, 35, 36; 1892, Oct. 7, 213 (g. of Polystominæ); 1893, 118; 1903, 336 (syn. Acanthoncocotyle) (subf. Onchocotylinæ).—Pratt, 1900, 646, 651, 656, fig. 26.—St.-Remy, 1890, Nov. 1, 41–43; 1898, 558.—Scott, T., 1901, 151.—Sons., 1890, 176–177.—Stoss., 1898, 11.—Tasch., 1878, 176; 1879, 36, 44, 45, 47, 50, 51, 53, 55, 60, 62, 64, 65, 66, 69; 1879, 252, 255 (syns.: Polyst. Kuhn, Hexabothrium Nord.).— Ziegler, 1883, 556.

1899: Acanthoncotyle Cerf., 1899a, 347 (appendiculata).

ONCHOCOTYLE-Continued.

abbreviata Olss., 1876, 12, pl. 2, figs. 27–28 (in Acanthias vulgaris).—Braun, 1890a, 418, 499, 539, 548, 550.—Cerf., 1899a, 361, 368, 369, 375 (to Squalonchocotyle).—St.-Remy, 1890, 42, 43 (in Ac. vulg.).—Sons., 1890, 177.—Staff., 1904, May 3, 482 (on Squalus acanthias; Canada).—Tasch., 1879, 253 (in Ac. vulg.); 1879, 29 (in Ac. vulg.).

appendiculata (Kuhn, 1830) Dies., 1850a, 419 (includes Hexabothrium); 1858e, 370-371 (in Mustelus lævis, M. vulgaris, Scyllium catulus), 371 (pars, from Læmargus borealis, syn. of O. borealis); 1859c, 438 (of Ben., 1858a, in Must. vulg., Galeus canis).—Ben., 1858a, 1861a, 54–58, 59, 168–169, 172, 177, 197, pl. 6, figs. 1-12 (in Mustelus vulg., N. Sea and Adriatic; Galeus canis; Scillium catulus, Mediterranean; Læmargus borealis, Greenland); 1870, 5, 6, num catulus, Mediterranean; Læmargus borealis, Greenland; 1870, 5, 6, 16.—Brand., 1891d, 20.—Braun, 1889k, 622; 1890a, 408, 410, 414, 415, 419, 420, 421, 428, 429, 434, 437, 438, 442, 449, 453, 454, 457, 462, 465, 481, 489, 498, 499, 500, 512, 514, 539, 548, 550, 551, 552; 1890e, 594; 1893b, 178.—Cerf., 1899a, 350, 351, 357, 358, 359, 360, 361, 362, 364, 366, 367, 368, 369, 371, 374, 378 (in Lemargus borealis, Scyllium catulus), 374 (of Stoss., 1887, syn. of Acanthonchocotyle canicula), 376 (of Tasch., 1879, syn. of Squalonchocotyle grisea), 376 (of Ölss., 1867 and 1876, syn. of Rajonchocotyle batis), 376 (of Sons., 1890, syn. of Rajonch. prenanti).—Cobbold, 1879b, 464.—Fraip., 1880c, 416.—Gotto, 1891, 162.—Lijma, 1881c, 638.—Lackson, 1888, 647.—Lyn., 1880. 416.—Goto, 1891a, 162.—Ijima, 1884c, 638.—Jackson, 1888, 647.—Juel, 1889, 36.—Kerbert, 1881a, 533.—Kroyer, 1852–53a, 926 (in Scymnus microcephalus).—Looss, 1885b, 5, 10.—Mont., 1888a, 8, 15, 23, 26, 37, 42; 1890, 421.—Par. & Perugia, 1890, 7.—Scott, 1901, 151, pl. 8, figs. 30, 31 (in Galeorhinus galeus, Mustela vulgaris, Raja batis, R. clavata, Scylliorhynus catulus); 1901, 344 (in Raja batis; Irish Sea); 1905, 118 (see Scott, 1901, pl. 8).—Sons., 1890, 176, 177 (see also Octocotyle app.); 1891, 259–260 (syn. O. emarginata Olss.).– St.-Remy, 1890, 42, 43 (in Scyllium catulus, Mustelus vulg., M. lævis, Galeus canis, Raja batis, Hexanchus griseus, Ac. vulg.).—Stoss., 1885, 162; 1898, 11.—Tasch., 1878, 176; 1878, 566, 574; 1879, 253 (syn. Hexathyridium app. Nord.) (in Scyllium catulus, Mustelus lævis, M. vulgaris, Hexanchus griseus, Raja batis, Galeus canis); 1879, 19–20; 1879, 24–47, pl. 3, figs. 1–3, pl. 4, figs. 1, 3–6 (syns. Hexabothrium app. Kuhn, Polyst. app. Nord.).—Ziegler, 1883, 345, 557.

borcalis Sons., 1890, 177, misprint for borealis.

borealis Ben., 18531, 59-68, figs. 1-11 (in Scimnus glacialis); 1858a, 1861a, 58-59 ans Ben., 18531, 59-68, figs. 1-11 (In Scimnus glacialis); 1858a, 1861a, 58-59 (in Sc. gl.); 1870, 12.—Braun, 1890a, 408, 419, 461, 539, 548.—Cerf., 1899a, 346, 356-358, 359, 360, 361, 362, 364, 367, 368, 369, 372, 378 (in Scymnus glacialis), 374 (to Squalonchocotyle in Scymnus borealis), 376 (of Stoss., 1885, syn. of Rajonchocotyle prenanti).—Dies., 1858e, 371 (in Læmargus borealis) (syns.: O. appendiculatum Dies., Polyst. borealis Wag.); 1859c, 438.—Linst., 1889, 101 (in Læmargus borealis).—Odhn., 1905, 372 (to Squalonchocotyle). Part 1840, 78, 1850, 77, St. Porty. lonchocotyle).—Par., 1894, No. 1050.—Par. & Perugia, 1890, 7.—St.-Remy, 1890, 42, 43 (in Scymnus borealis).—Sons., 1890, 177 (borcalis).—Stoss., 1885, 162; 1890, 52; 1898, 12.—Tasch., 1879, 253 (in Sey. bor.); 1879, 29, 32 (in Scy. bor.)—Also reported for Læviraja oxyrhynchus.

canis (Cerf., 1899) Pratt, 1900a, 656, 657, fig. 26.

emarginata Olss., 1876, 11–12, pl. 2, figs. 23–26 (in Raja clavata).—Braun, 1890a, 419, 494, 499, 539, 548, 551.—Cerf., 1899a, 361, 366, 367, 368, 369, 375, 378.— St.-Remy, 1890, 42, 43 (in R. clav.).—Sons., 1890, 176, 177 (see also Octocotyle emarginata); 1891, 259, 260 (syn. of O. appendiculata Kuhn).—Tasch., 1879, 253 (in R. clav.); 1879, 29 (in R. clav.).

prenanti St.-Remy, 1890, 41-43 (in Raja oxyrhynchus; Roscoff).—Cerf., 1899a, 365, 368, 376 (to Rajonchocotyle) (in R. oxyr.).—Goto, 1894a, 223.—Sons., 1890, 177 (in R. oxyr.).

scymni ainosi Dies., 1858e, 371 based on Wagener, 1857, 72 (in Scymnus ainosi).— Cerf., 1899a, 360, 362, 377.—Linst., 1878a, 282.—St.-Remy, 1890, 41.

spinacis Goto, 1894a, 224-226, pls. 15-16, gills of Spinax sp.; at Odawara, Japan.-Cerf., 1899a, 370, 372, 376, 435 (to Squalonchocotyle).—St.-Remy, 1898, 558.

ONCHOCOTYLINÆ Cerf., 1899a, 345–478, pls. 18–21, 396 (Onchocotilinæ); 1900a, 436-438.—Mont., 1903, 336 (f. Onchocotylidæ).

ONCHOGASTER Claparède, 1863a, 13 (m. natator).

natator Claparède, 1863a, 13, pl. 5, figs. 1-4.—Braun, 1889a, 363, after Leuck., 1863, 95, is perhaps a free swimming polystome larva.

OPHICOTYLE Ben. & Hesse, 1863; 1864, 96, 101 (m. fintæ).—Braun, 1890a, 414, 477, 516, 517, 522, 546.—Cerf., 1895h, 918; 1896, 514.—Mont., 1888a, 8, 11, 86 (Ophycotyle), 89, 99 (Ophycotyle).—Tasch., 1879, 240.

1888: Ophycotyle Mont., 1888a, 8, 11, 86, 89, 99 for Ophicotyle.

alosæ Mont., 1888a, 13 (Ophycotyle).

fintæ Ben. & Hesse, 1863; 1864, 101–102, pl. 9, figs. 19–28 (in Alosa finta).—Braun, 1890a, 446.—Tasch., 1879, 244 (to Octobothrium).

OPHTHALMOPHAGUS Stoss., 1902, 8, 28–29 (m. singularis), subf. Cyclocælinæ.

singularis Stoss., 1902, 29, pl. 7, fig. 27 (in Gallinula pusilla; loc. not given).

OPHYCOTYLE Mont., 1888a, 8, 11, 86, 99, for Ophicotyle.

alosæ Mont., 1888a, 13.

OPISCORCUS Woolley, 1906, Oct. 6, 1090 (misprint for Opisthorchis).

OPISTHIOGLYPHE Looss, 1899b, 588–589, 592 (tod. endoloba) (ὀπίσθιος=hindermost; ἡ γλυφή=das Eingegrabene).—Pratt, 1902, 888, 896, 898.

endobala Ssinitzin, 1906, 685, for endoloba.

endobola Ssinitzin, 1905, 121-136; 1906, 686, 687, for endoloba.

endoloba (Duj., 1845) Looss, 1899b, 589.—Braun, 1906, 137, figs. 72–73 (in frog).—Kowal., 1902d, 27 (9).—Ssinitzin, 1905, 121–136; 1906, 685 (endobala), 686, 687 (endobola) (Cerc. gibba is larva) (in Rana esculenta, R. temporaria; Warschau).—Staff., 1905, 686–687 (Duj. of Staff., 1900, 403, renamed Glypthelmins quieta).—Stoss., 1901, 96 (8) (in Rana esculenta; Triest).—Ward, 1903, 862.

siredonis (Poir., 1886) Looss, 1899b, 589.

OPISTHODISCUS Cohn, 1904, 240 (m. diplodiscoides), 242, 243.

diplodiscoides Cohn, 1904, 240-243, figs. 6-8 (in Rana esculenta; Coll. Creplin).

OPISTHOGONIMUS Luehe, 1900aa, 555–558, 562 (m. lecithonotus=philodryadum).— Looss, 1901b, 206; 1902m, 839.—Odhn., 1902, 41.—Pratt, 1902, 888 (related to Haplometrinæ), 901.

lecithonotus Luehe, 1900aa, 555-558 (in Coluber sp., Coluber eririo, Philodryas schotti; Brazil); 1900ee, 743 (=O. philodryadum (West)).

philodryadum (West, 1895) Luehe, 1900ee, 743 (in Philodryas schottii; Brazil).

OPISTHORCHIASIS.—A term proposed by Looss, 1905, 88, to designate infection with Opisthorchis.—Acyama, 1891a.—Askanazy, 1904, 698 (in man, etiology and pathology); 1904, Nov., 229–230.—Huber, 1896, 578.—Katsurada, 1900, 495, 500–503 (in man).—Macgregor, 1877, 3–16.—See also distomatosis, hepatic, Japan.

[O. felineus; Askanazy, 1904, (19); 1905, Jan. 10, 790.]

OPISTHORCHIC Stiles, 1901h, 1539 (misprint for Opisthorchis Bl. 1895).

OPISTHORCHIDÆ Luehe, 1901, 486, for Opisthorchiidæ.

OPISTHORCHIIDE Braun, 1901b, 34.—Jægers., 1903a, 15 (Opistorchidæ).—Looss, 1902m, 839, 840 (contains Echinostominæ, Omphalometrinæ, Opisthorchiinæ, Rhopaliadinæ; Azygia; Allocreadium; Rhytidodes; Calycodes).—Luehe, 1901, 486.

OPISTHORCHIIN.E Looss, 1899b, 543, 563, 564, 565, 566; 1902m, 718, 839.—Braun, 1901b, 20.—Luehe, 1900, 490.—Pratt, 1902, 888, 895 (includes Opisthorchis, Holometra, Metorchis; related genus, Podocotyle).—Ward, 1903, 863.

OPISTHORCHIS R. Bl., 1895f, 217 (tod. felineus); 1896b, 730.—Braun,1900g,250; 1901b, 20; 1901, 564; 1901e, 314; 1902b, 5, 6, 7 (re Campula, Stiles & Hass. have stated that Campula was erroneously made syn. of Opisthorchis): 1903, 3 ed., 157 (erroneously dated 1845); 1906, 163.—Heymann, 1905, 98.—Jacoby, 1900, 7, 9, 16.—Kamensky, 1900a, 3, 4, 5, 13, 19.—Kowal., 1898g, 74; 1898h, 131, 136, 140, 142–148 (28, 33, 37, 39–45); 1898, 121–148.—Looss, 1899b, 539, 549, 551, 559, 560, 563, 564, 565, 590; 1901, 194, 199, 200, 209; 1902m, 529, 712 (Opistorchis), 775–778, 781, 790 (amphitypic), 795, 805; 1905, —.—Luehe, 1899, 529, 532, 533; 1901, 474.—Mont., 1896, 168.—Odhn., 1902, 37 (Opistorchis).—Ofenheim, 1900, 166.—Pratt, 1902, 888, 895.—Rail., 1896, 160; 1900, 212.—Stiles, 1901, 203, 204, 205.—Stiles & Hass., 1898a, 85, 91 (Opistorchis erroneously as syn. of Campula Cobb.) (type Dist. felineum).—Stoss., 1902, 580.—Ward, 1901, 180; 1903, 869.

1896: Prosthometra Looss, 1896b, 58–60 (felineus).

OPISTHORCHIS-Continued.

1896: Opistorchis Rail., 1896, Mar. 15, 160, misprint.
1901: Opisthorchic Stiles, 1901h, 1539, misprint.

1906: Opiscorcus Woolley, 1906, 1090, misprint.

albidus (Braun, 1893) Rail., 1896, 160.—Rail. & Marotel, 1898, 38 (compared with other forms).—Bossuat, 1902, v. 6 (2), 192.—Jacoby, 1900, 8 (situs inversus 16 times in 68 specimens, 23 per cent).—Kowal., 1898h, 131, 143, 147 (28, 40, 44) (in Felis catus dom.).—Luehe, 1899, 530.—Type of Metorchis 1899.

amphileuca (Looss, 1896) Kowal., 1898, 148 (45) (in Naja haje).

buski (Bl., 1888) R. Bl., 1895, 737 (syns.: Dist. buski Lankester, D. crassum Busk nec Sieb., D. rathouisi Poir.); 1895, 217; 1900, 488.—Galli-Valerio, 1898a, 146.—Manson, 1903e, 664, fig. 107.—Type of Fasciolopsis 1899.

campula (Cobbold, 1876) Looss, 1899b, 559.

choledoca (Linst., 1883) Kowal., 1898h, 148 (45) (in Anas sp.).—Kamensky, 1900a, 5. compascua Kowal., 1898, 72; 1898h, 133, 134 [30, 31], see xanthosoma compascua.

complexus (Stiles & Hass., 1894) Stiles & Hass., 1896, 155, or Rail., 1896, 160.—
 Galli-Valerio, 1898d, 146.—Kowal., 1898h, 143, 147 (40, 44) (in Felis catus dom.).

conjunctus (Cobbold, 1860) R. Bl., 1895, 217; 1895, 735; 1900, 488.—Bossuat,
1902, v. 6 (2), 191–192.—Braun, 1883, 65–66.—Clerc, 1907, 558 (Opistorchis).—
Kamensky, 1900a, 4.—Kowal., 1898h, 143, 147 (40, 44) (in Homo, Canis vulpes fulvus).—Manson, 1903e, 635 (=Dist. conjunctum of man).—Rail., 1896, 160.

conus (Crep., 1825) Stiles & Hass., 1895c, 156–158.—Rail. & Marotel, 1898, 38.
corvina (Stiles & Hass., 1894) Kowal., 1898, 143, 148 (40, 45) (in Corvus americanus, C. ossifragus).

corvinus (Stiles & Hass., 1894) Stoss., 1904, 12.

crassiuscula (Rud., 1809) Kowal., 1898g, 71, 73; 1898h, 130, 131, 143, 144, 147 (27, 28, 40, 41, 44) (in Buteo vulgaris, Archibuteo lagopus, Haliaëtos albicilla, Circus rufus, Aquila imperialis, Surnia nyctea).—Jacoby, 1899c, in 1–30; 1900, 7.—Looss, 1899b, 565, 678.—Luehe, 1899, 530.

crassiuscula janus Kowal., 1898h, 122–132, 147 [19–27, 44] figs. 15–21, 28 (in Anas boschas dom.); 1898g, 71.—Braun, 1902b, 9.—Looss, 1899b, 565, 566 (undoubtedly a Metorchis), 678.

crassiusculus Looss, 1899b, 678, see crassiuscula.

entzi Ratz, 1900, 534, in gall ducts of Ardea purpurea.

exigua (Mueh., 1898) Kowal., 1898h, 147 (44) (in Circus rufus).

felinea (Riv., 1884) Jacoby, 1899c, in 1-30.

felineus (Rivolta, 1884) R. Bl., 1895, 217; 1895, 734–735; 1900, 488.—Askanazy, 1900b, 493, 502 (in man); 1904, 689–691; 1904, Nov., 229–230.—Bossuat, 1902, v. 6 (2), 188.—Braun, 1903, 3 ed., 157 (syns.: conus Gurlt, lanceolatum Sieb., 1836 & Tright, sibiricum, tenuicolle Mueh., 1896) 159 (cases in man); 1906, 166; 1906, 163–166, figs. 95–98 (in Limnæus stagnalis, Felis domestica, dog, fox, Gulo borealis).—Engler, 1904 (X. 21), 186–188, 1 fig. (abnormal intestine of); 1905. Jan. 1, 16; 1905 (II), 57.—Galli-Valerio, 1898d, 146; 1901c, 364 (in dog).—Hollack, 1902a, 868.—Jacoby, 1900, 7, 8–9.—de Jong, 1896a, 7 (Opistorchis).—Kamensky, 1900a, 17, 19. 20, 24 (felinea).—Looss, 1899b, 675; 1905, 89 (syns.: Dist. felineum, D. sibiricum; in dogs and cats, Europe; in man by Winogradow in Tomsk, Siberia), 90; 1907, Feb. 1, 138, 139.—Stiles, 1902, 33.—Ward, 1903, 704; 1903, 864, 869 (syns.: Dist. conus Gurlt nec Crep., D. lanceolatum Sieb., 1836 & Tright, 1889 nec Mehlis, D. sibiricum Winogradoff, D. tenuicolle Mueh.), 871.

gemina (Looss, 1896) Kowal., 1898h, 129, 148 (26, 45) (in Milvus parasiticus).

geminus (Looss, 1896) Looss, 1899b, 676, 677, 678, figs. 2, 3; 1907, Feb. 1, 138, 139 (in Anas boschas fera, Circus æruginosus, Milvus ægyptius).—Braun, 1901b, 34.

interruptus Braun, 1901g, 897 (in Alcedo viridirufa, Ardea virescens; Brazil); 1902b, 5, 6, fig. 2.—Stoss., 1904, 11.

janus Kowal., 1898g, 71, 73; 1898h, 130, 131, 132, 133, 134, 137, 138, 139, 143, 144, (22, 26, 27, 28, 29, 30, 31, 34, 35, 36, 40, 41).—Jacoby, 1900, 7.—See also crassiuscula janus.

OPISTHORCHIS—Continued.

lancea (Dies., 1850) Braun, 1901, 897; 1902b, 6, 7.—Hollack, 1902a, 868.—Stoss., 1904, 11.

longissimus (Linst., 1883) Stiles & Hass., 1896, 155.—Kamensky, 1900a, 5.— Kowal., 1898g, 72; 1898h, 140, 141, 148 (37, 38, 45) (in Botaurus stellaris).— Looss, 1899b, 564, 674.—Stoss., 1904, 11.

longissimus var. corvinus (Stiles & Hassall, 1894) Looss, 1899b, 564, 675.—Hollack, 1902a, 868.—Stoss., 1904, 12.

noverca Braun, 1902, Dec. 30, 836; 1903, 3. ed., 164, fig. 112 (=Dist. conjunctum Lew. & Cunn., 1872 of man, renamed); 1906, 170-171, figs. 103-104.—Looss, 1905, 90–91, fig. 6 (syn. Dist. conj. L. & C.).—Stoss., 1904, 11.—Ward, 1903, 704; 1903, 864, 870.

oblonga (Cobbold, 1858) Kowal., 1898h, 148 (45) (in Phocæna communis, Platanista gangetica).

pianæ Galli-Valerio, 1898c, 7–8, 1 fig. (in Anas boschas; Busto Ariizio); 1898d, 145–146 (in Anas boschas); 1898e; 1898m, 923 (Opistorchis); 1901c, 364; 1903d, 59 (Opistorchis).—Kowal., 1898c, 751–752; 1898d, 751–752; 1898e, 923.—Stoss., 1899, 12.

piscicola Odhn., 1902, 152–153 (in Gymnarchus niloticus).—Stoss., 1904, 11.

poturzycensis Kowal., 1898g, 71 (var. of simulans), 72 (n. sp.), 73, 75, or 1898h, 138, 140, 141, 142, 143, 144, 148 (in Anas boschas dom.), 150, 163, figs. 23–26 (35, 36, 37, 38, 39, 40, 41, 47).—Engler, 1904, 186.—Jacoby, 1899c, in 1–30; 1900, 7.

pseudofelineus Ward, 1901, 180 (syn. D. felineum Riv. of Ward, 1895, 152–158); in 1903, 861, 862, 869.—Braun, 1903, 3 ed., 158.

simulans (Looss, 1896) Kowal., 1898g, 72; 1898h, 129, 137, 140, 141, 142, 148 (26, 37, 38, 39, 45) (in Pernis apivoris); 1902d, 23 (5) (in Anas boschas dom.).—Braun, 1901b, 34.—Looss, 1899b, 564, 566, 673–674, pl. 24, figs. 1–1b (includes var. poturzycensis).—Stoss., 1904, 11.—Also reported for Anas penelope, Circus æruginosus, Fulix cristata).

simulans var. poturzycensis Kowal., 1898g, 71 (in Anas boschas dom.); 1898h, 135–142, 148 (32–37, 45), figs. 23–26 (in Anas boschas dom.).—Looss, 1899b 565, 673, 674.

sinensis (Cobbold, 1875) R. Bl., 1895, 217; 1895, 736–737; 1900, 488; 1901b, 204, 209, 210; 1901c, 581, 586, 587, 588, 589.—Akanuma, 1894 (in Toyama Province).—Aoyama, 1891a.—Askanazy, 1904, 689; 1904 (XI), 229–230.—Bossuat, 1902, v. 6 (2), 189.—Braun, 1903, 3 ed., 161–162, figs. 109–111; 1906, 168–170, 1902, v. 6 (2), 189.—Braun, 1903, 3. ed., 161–162, figs. 109–111; 1906, 168–170, figs. 100–102.—Clerc, 1907, 558 (Opistorchis).—Kamensky, 1900a, 4.—Kowal., 1898h, 147 (44) (in Homo sapiens; Felis catus dom.).—Looss, 1899b, 564; 1901, 209; 1905, 90, fig. 5 (syns. Dist. sinense Cobb., D. spathulatum Leuck., D. hepatis innocuum, D. hepatis endemicum, D. hepatis perniciosum Bælz, D. japonicum Bl.) (Japan, Annam, Tonkin, China, Korea); 1905m, 233; 1907, Feb. 1, 136–147.—Manson, 1903, 635.—Staf., 1905, Apr. 11, 694 (in liver of Chinaman, at Montreal, 1896).—Stilles & Garrison, 1906a, Aug., 29.—Stoss., 1904, 11.—Ward, 1903, 864, 867, 869–870 (syns. Dist. sinense Cobb., D. spathulatum Leuck. nec Rud., D. hepatis endemicum Bælz, D. hepatis perniciosum Bælz, D. hepatis innocuum Bælz, D. japonicum Bl., D. endemicum Ijima); 1903, 703, 704.—Woolley, 1906, 1090 (Opiscorcus).

speciosus Stiles & Hass., 1896, 151–155, figs. 1–2 (Dist. longissimum corvinum

renamed).—See p. 74.

tenuicollis (Rud., 1819) Stiles & Hass., 1896, 155.—Braun, 1901e, 314.—Jacoby, 1900, 8.—Kamensky, 1900a, 1–23 (syn. D. felineum Riv.); 1900b, 23 pp.; 1901a, 323–324.—Kowal., 1898g, 73, 74; 1898h, 106, 129, 138, 139, 143, 144, 147 (1, 26, 35, 36, 40, 41, 44) (in Homo, Felis catus dom., Canis familiaris, Gulo borealis, Phoca barbata, Halichærus grypus).—Looss, 1899b, 564, 566, 674–678, 700.—Odhn., 1905, 339.—Ratz, 1900, 532.—Stoss., 1904, 11.

tenuicollis-felineus Looss, 1899b, 678 (see tenuicollis and felineus).

truncatus (Rud., 1819)
Rail., 1896, 160.—Bossuat, 1902, v. 6 (2), 192.—Galli-Valerio, 1898d, 146; 1901c, 364.—Jacoby, 1899c, in 1–30; 1900, 8.—de Jong, 1896a, 7 (Opistorchis).—Kamensky, 1900a, 14 (truncata).—Kowal, 1898h, 143, 147 (40, 44) (in Gulo borealis, Felis catus dom., Canis familiaris, C. vulpes, Phoca annelata, P. vitulina).—Luehe, 1899, 530.

OPISTHORCHIS-Continued.

viverrinus).—Poir., 1886, 27-29, pl. 3, figs. 1-3.—Stoss., 1892, 1893, 24; 1904, 11.

xanthosoma (Crep., 1846) Kowal., 1898g, 72; 1898h, 134, 147 (31, 44) (in Podiceps minor).

xanthosoma var. compascua Kowal., 1898g, 71 (in Anas querquedula); 1898h, 132–134, 147 (29–31, 44), figs. 22–29.—Braun, 1902b, 8 (syn. of Dist. crassius-culum).—Looss, 1899b, 565, 566 (says this is undoubtedly a Metorchis), 678.

OPISCORCUS Woolley, 1906, Oct. 6, 1090 (misprint for Opisthorchis).

OPISTHOTREMA Fischer, 1883a, 1–42 (m. cochleare).—Biehringer, 1888a, 231, 232, 233, 234.—Brand., 1892, 506.—Braun, 1892a, 569, 635, 640, 663, 664, 676, 677, 679, 683, 687, 693, 696, 699, 700, 711, 715, 716, 717, 718, 721, 725, 727, 728, 736; 1893a, 879, 886, 887, 890, 893, 895, 899, 914, 917, 918.—Darr, 1903, 657.—Gamb., 1896a, 73.—Jackson, 1888, 648.—Juel, 1889, 24.—Looss, 1885b, 56; 1894a, 180, 207; 1899b, 665.—Mont., 1888, 7, 15, 18, 32, 35, 37, 41, 42, 48, 50, 52, 53, 54, 56, 57, 58, 60, 64, 66, 67, 93, 106; 1892, Oct. 7, 214 (gen. of Monostomidæ); 1892, 685, 686, 687, 688, 689, 690, 691, 692, 693, 694, 695, 696, 697, 698, 699, 700, 701, 702, 703, 704, 713; 1892, 37, 38; 1893, 25, 83, 144.—Ofenheim, 1900, 156.—Pratt, 1902, 890, 909.

1893: Oposthotrema Crety, 1893a, 384, misprint.

1902: Opistotrema Cohn, 1902k, 53, for Opisthotrema.

cochleare Fischer, 1883a, 1–42, 1 pl., 13 figs. (in Halicore dugong; Philippines); 1884a.—Bettend.,1897, 38; 1897, 342.—Biehringer, 1888a, 230.—Brand., 1891b, 265.—Braun, 1891d, 421; 1892a, 583, 589, 591, 593, 594, 595, 602, 604, 605, 607, 610, 628, 640, 659, 688, 703, 724, 731, 733, 762, 763, 764, 765; 1893a, 877, 917; 1893d, 468; 1901e, 344.—Cohn, 1902k, 53 (Opistotrema).—Crety, 1893a, 384 (Oposthotrema).—Darr, 1902, 648.—Juel, 1889, 37.—Kath., 1894a, 143.—Kowal., 1898h, 158 (55).—Linst., 1904u, 679, 680.—Looss, 1885b, 10, 12, 17, 56.—Maclaren, 1904b, 608.—Mont., 1888a, 7, 16, 21, 23, 26, 27, 43, 60; 1892, 99, 104; 1892, 688, 689, 695, 703, 712; 1893, 16, 109.—Ssinitzin, 1904, 768 (Opistotrema).—Reported also for Manatus americanus.

pulmonale Linst., 1904u, 678-680, figs. 1-2 (in Halicore australis; Torres-Strasse,

Australia).—Odhn., 1906, 66.

OPISTORCHIDE Jægers., 1903a, 15, for Opisthorchiidæ.

OPISTORCHIS Rail., 1896, 160, see Opisthorchis.

OPOSTHOTREMA Crety, 1893a, 384, for Opisthotrema.

ORCHIDASMA Looss, 1900, Dec. 3, 602 (Anadasmus Looss nec Walsingham renamed, hence type amphiorchis), το δάσμα, die Vertheilung: 1902m, 468.—Braun, 1901a, 20, 34.—Odhn., 1905, 342.—Pratt, 1902, 888, 895.

amphiorchis (Braun, 1899) Braun, 1901b, 20–22. figs. 7, 11 (in Thalassochelys corticata; Chelone mydas; Triest).—Looss, 1901, 560, 562, 563; 1902m, 414, 416, 463–468, pl. 23, figs. 25–29, pl. 39, fig. 24.—Odhn., 1905, 342.

ORCHIPEDUM Braun, 1901g. 944 (m. tracheicola Braun); 1902b, 15, 19, 20, 22, 23.— Pratt, 1902, 888 related to Psilostominæ), 897.

tracheicola Braun, 1901g, 943–944 (in Anas fusca; Vienna, Oct.); 1902b, 19, 20, 22, 23, figs. 14, 15 (syn. Monost. flavum).

OROPHOCOTYLE Looss, 1902e, 637–644, figs. 1–4 (tod. planci); 1902f, 400–401; 1905g, 54–55.

calyptrocotyle (Mont., 1893) Looss, 1902e, 644.

divergens Looss, 1902e, 640, 643, 644, fig. 1 (in Ranzania truncata; Triest).

foliata (Lint., 1898) Looss, 1902e, 644 (syn. Dist. foliatum Lint.).

planci (Stoss., 1899) Looss, 1902e, 637–641, figs. 2–4 (in Ranzania truncata; Triest).

ORTHOSPLANCHNUS Odhn., 1905, 330, 345, 346, 347, 348 (tod. arcticus).

arcticus Odhn., 1905, 339–343, 344, 348, pl. 3, figs. 1–5 (in Phoca barbata; west coast Spitzbergen).

fraterculus Odhn., 1905, 343–344, 348, pl. 3, fig. 6 (in Phoca barbata, Odobenus rosmarus; w. coast Spitzbergen).

OSTIOLUM Pratt, 1902a, 888 (Haplometrinæ), 900 (key to); 1903, 34–37 (m. formosum Pratt).

formosum Pratt, 1903, 34–37, pl. 4, figs. 6–8 (in frogs).—Staff., 1905, Apr. 11, 687 (in lungs of American frogs and toads; probably identical with Pneumoneces medioplexus).

OTIOTREMA Setti, 1897, 4–8, figs. 1–5 (m. torosum); 1897, in 198–247.—Braun, 1902b, 23.—Fuhrmann, 1904, 61.—Looss, 1899b, 551, 644-645, 741, 742, 743, 744, 745, 1901b, 208, 209, 210; 1902e, 642; 1902m, 813.—Luehe, 1901n, 481, 482, 486.—Odhn., 1905, 366.—Ofenheim, 1900, 183.—Pratt, 1902, 889, 905.—Stiles & Hass., 1898a, 91–92, 96 (type torosum).

torosum Setti, 1897, 4-8, pl. 8, figs. 1-5 (in Squalus sp.; Massaua).—Darr, 1902, 661.—Looss, 1899b, 645, 735, 736–746, figs. 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 69 (in Squalus sp.; Sawakin).—Luehe, 1901n, 481.—Odhn., 1905,

362.—Stiles & Hass., 1898a, 91, 96.

OTODISTOMUM Staff., 1904, May 3, 482–483 (m. veliporum) (Ωτος= giant) (closely related to Azygia).

veliporum (Crep., 1837) Staff., 1904, May 3, 482-483, 488 (in Raja lævis Mit.; Canada).—Odhn., 1905, 310.

- PACHYPSOLUS Looss, 1901, 30. Oct., 558-560 (m. lunatus Looss, 1901, 558 = D. irroratum R.); 1902m, 497, 503, 504, 505.—Pratt, 1902, 888 (related to Plagiorchiinæ), 899.
 - irroratus (Rud., 1819) Looss, 1902m, 414, 416, 485–505 (includes Dist. irroratum Rud., 1819a, 393; Braun, 1899, 717; 1901, 36, figs. 27, 30, 32), 793, 871, 887, fig. A, pl. 23, figs. 37–38, pl. 32, fig. 169 (in Thalassochelys corticata, at Triest; Thalassochelys caretta in New Guinea).—Odhn., 1905, 340.

lunatus Looss, 1901l, 558-560, 564 (in Thalassochelys corticata; Triest); 1902m, 485, 486, 496, 497, 503 (syn. of P. irroratus Rud.).

PARABASCUS Looss, 1907, Mar. 5, 481–483 (tod. lepidotus).

lepidotus Looss, 1907, Mar. 5, 481–483, figs. 3a-b (in Vesperugo kuhli; Cairo, Egypt).

limatulus (Braun, 1900) [Looss, 1907, Mar. 5, 483, as probable].

semisquamosus (Braun, 1900) [Looss, 1907, Mar. 5, 483].

PARAGONIMIASIS, name of disease (see also distomatosis, pulmonary) Stiles & Hass., 1900a, 578–600.—Huber, 1896, 577.—Inoue, 1892, 13, 15; 1893a, 79–86; 1897a, 175–178 (brain); 1900, 1–23 (distomatosis pulmonum); 1900b, 515, 664, 739; 1903, 124, 125, 130.—Katsurada, 1900, 507.—Kurimoto, 1893d, 1-6, 1 fig.-739; 1903, 124, 125, 130.—Katsurada, 1900, 507.—Kurimoto, 1893d, 1-6, 1 fig.—Kurimoto & Ijima, 1892a, (718-720).—Looss, 1905, 81, 82, 83.—Manson, 1883, (Mar. 31), 532-534; 1894, 805.—Matsushima, 1895a, 10-17 (induratio hepatis).—Maxwell, 1899, 116-117, 2 figs.—Miura & Nakanishi, 1897, 20 Aug., 31-33.—Stiles, 1901b, 79-102.—Taylor, 1884, 44-54, figs. 1-5.—Winoue, 1892, Oct. 20, 13-16.—Winoue & Katsurada, 1892, Nov. 5, 15-21.—Ztsch. d. Tokio med. Gesellsch., 1892, v. 6 (6), Mar. 20, Art. 1.—In Brain: Inoue, 1897a, 175-178.—Inoue & Katsurada, 1891, (17); 1892, v. 6 (6); 1893; 1902, v. 6 (21).
—Jida, 1899, (1).—Taniguchi, 1904, Mar. 31, 237; 1904, Aug. 1, 156; 1904, Aug. 1, 16, 350-351; 1904, v. 38 (1), 100-121, 2 pls.; 1904, 318-321; 1904, Dec. 3, 983; 16, 350–351; 1904, v. 38 (1), 100–121, 2 pls.; 1904, 318–321; 1904, Dec. 3, 983; 1905, July 29, 508–509.—Yamagiwa, 1889, Sept. 20, 8; 1890, Mar. 5, 447–460, figs. 1–3; 1890, May 20,——; 1890, Sept. 5, 336–337.

-, LOCALITY: Ijima, 1893, v. 7 (16) (Yamanashi).—Katsurada, 1900d, 507 (mountainous regions).—Miura & Nakanishi, 1897a, 31–33.—Soma, 1893 (Japan).— Sons., 1884, v. 54 (7), 17–21 (Japan & Formosa).—Stiles & Hass., 1900a, 560– 611; 1900b; 1900c, 3017-3027 (U.S.); 1901a, 45 (U.S.).—Yamagiwa, 1892, Mar. 5, 446-456 (Japan).

-, PATHOLOGY: Katsurada, 1899a, 8–29.—Katsurada, Fujiro & Fujiki, 1899a, 20 June, 1–18; 5 July, 2–29.—Magaziner, 1902, Apr., 296–302 (in man and lower animals).—Taniguchi, 1893, (794–795) (eye and testicle).—Wakabayashi, 1903, Apr. 20, 117 (orbit and lid).

-, symptoms: Huber, 1896a, 577.

- -, ткелтмент: Katsurada, Fujiro & Saki, 1899a, 141–185.—Riusai, 1884, no. 307 (sulphurous acid).
- -, IN MAN: Emerson, 1904, July-Aug., 263.—Stiles & Hass., 1900a; 1902f, 360; 1904c, 22.
 - -, IN SWINE: Stiles & Hass., 1900a, 560-611.
- PARAGONIMUS Braun, 1899g, 492 (tod. westermanii) (also places here, Dist. rude Dies., Dist. compactum Cobbold); 1900h, 5, 6; 1901e, 329, 330; 1903, 3. ed., 154–155; 1906, 160.—Looss, 1900d, 605; 1902m, 813.—Luehe, 1900, 555, 557.— Pratt, 1902, 887 (related to Fasciolinæ), 894.—Stiles, 1901, 183, 185.—Taniguchi, 1904, v. 38, — (in brain); 1904 (XII, 3), 938.—Ward, 1903, 867.

PARAGONIMUS—Continued.

1899: Polysarcus Looss, 1899b, 561 (tod. westermanii).

compactus (Cobbold, 1859) Braun, 1901, 334.

rudis (Dies., 1850) Stiles & Hass., 1900a, 604-605.—Braun, 1901e, 329, 332, pl. 20, figs. 12, 15–17.

westermani (Kerbert, 1881) Looss, 1905, 81, 82, fig. 1; 83, 84, figs. 2-3; 85, 86 (syns. Dist. ringeri, D. pulmonale, D. pulmonum) (see westermanii); 1905m, 280, 282, 283.

westermanii (Kerbert, 1878) Stiles & Hass., 1900a, 560-611, figs. 24-28, pls. 23-24, figs. 1-4; 1900b, 761-762 (westermanni); 1900c, 3017-3027; 1901a, Jan. 12, 45.—Anders, 1903, 6th ed., 1245-1246.—Braun, 1901e, 331, 332, 333, 334; 1903, 3. ed., 155, figs., 101-103; 1906, 160-163, figs. 92-94 (westermani) (in man, Royal tiger, dogs, pigs, cats; N. America).—Darr, 1902, 652, 687 (westermanni).—[Kellicott, 1894a, 123 (in dog; U. S. A.).]—Looss, 1905, 81, 82, 83, 84, 85, 86, figs. 1-3 (westermanni) (syns. Dist. ringeri, D. pulmonale, D. pulmonum), 117; 1905m, 280, 282, 283 (westermanni).—Wanson, 1903, 631, 632 monum), 117; 1905m, 280, 282, 283 (westermani).—Manson, 1903, 631, 632, 633–636, figs. 96, 97 (westermanni).—Miura, 1897, 31–33.—Ricketts, 1903, in 1204–1206.—Stiles, 1901b, 79–102 (westermanii); 1902, 45, 47; 1904i, 14–18, figs. 5–16.—Stiles & Garrison, 1906a, 29.—Strong, 1901, 44–45.—Ward, 1903, 703, 704; 1903, 863, 864, 867 (syns. Dist. westermanii Kerbert, 1878; D. ringeri Cobbold, 1880; D. pulmonale Baelz, 1883; D. pulmonis K., S., & Y., 1881 Mesogonimus westermanii Rail., 1890), 868.

westermanni (Leuck., 1889) Stiles, 1900, 762 (see westermanii).

PARAMPHISTOMIDÆ Fischder., 1901a, 367–375: 1901b, 634–636; 1902a, 59 pp., 4 figs. (syn. Amphistomidæ Mont.); 1902b, 356; 1903h, 485–600, figs. a-q, pls. 20–31; 1903i; 1904, v. 20, 453–470; 1904, 278–279; 1904, 532–533; 1904, Feb. 18, 598–601; 1904, Mar. 3, 173; 1904, Mar. 10, 403; 1905, Jan. 31, 16; 1905, Jan. 16, 119; 1905, Jan., 120; 1905, July 3, 63.—Braun, 1903, 3. ed., 145, 146.—Luehe, 1901, 488.—MacCallum, 1905, 667.—Pratt, 1902, 887, 892 (includes: Paramphistominæ, Cladorchinæ).—Shipley, 1905, v. 6 (1), 4, 8.—Ward, 1903, 864 (Gastrodiscus hominia), 865 (see Digenea) (Gastrodiscus hominis), 865 (see Digenea).

PARAMPHISTOMINÆ Fischder., 1901, 367; 1902a, 10 (subf. of Paramphistomidæ); 1903h, 490, 492, 503.—MacCallum, 1905, 668.—Pratt, 1902, 887, 892 (syns. Gastrothylax, Paramphist., Stephanopharynx).—Shipley, 1905, v. 6 (1), 8

(genera: Paramphist., Stephanopharynx, Gastrothylax).

PARAMPHISTOMUM Fischder., 1901a, 367, 370, 372, 373, 374; 1902a, 7, 10-11, 24, 25, 26, 38, 43, 54 (tod. cervi); 1903h, 490, 491, 492 (key), 503; 1904, 173; 1904, 403; 1904, 453–470 (3 species); 1904 (X), 532–533.—Looss, 1902m, 438, 780, 835.—Pratt, 1902, 887, 892.—Shipley, 1905, v. 6 (1), 7, 8.

bathycothyle Fischder., 1903h, 542, for bathycotyle.

bathycotyle Fischder., 1901a, 368, 370 (in Bos kerabau); 1902a, 15, 16, 22 (in B. k.; Ceylon); 1903h, 492, 498 (in B. k.; Ceylon), 518–520, pl. 20, figs. 8–9, 542 (bathycothyle); 1904, 454, 455, 456, 457, 458.

bothriophoron (Braun, 1892) Fischder., 1901a, 370; 1902a, 21–22, 25, 29 (in Zebu; Madagascar); 1903h, 492, 496, 498, 538–541, figs. 24–28 (in Bos taurus indicus; Annanarivo, Madagascar), 544, 545, 546, 549, 550, 571.

calicophorum Fischder., 1901a, 370 (in Bos taurus; East Africa and Capland, Queensland, China); 1902a, 22–23 (in Bos tau.); 1903h, 492, 498 (in Bos tau.); East Africa, Capland, Queensland), 509 (in Ovis aries? or Bos tau.), 541, figs. e, 29–35, 549, 550, 575 (in Bos tau.; Canton. Fu-mui, China), 580; 1904,

459.—Linst., 1906, 175 (in Bos indicus).

cervi (Schrank, 1790) Fischder., 1901a, 368, 369; 1902a, 11–13, figs. 1, 14, 15, 20, 22, 26, 42 (includes Festuc. cervi Zed., 1790; Monost. elaphi Zed., 1800; M. 22, 20, 42 (includes Festuc. cervi Zed., 1790; Monost. chapin Zed., 1800; M. conicum Zed., 1803; Amphist. conicum Rud., 1809) (in Bos bubalus, B. taurus, B. urus, Cervus alces, C. dama, C. elaphus, Ovis aries); 1903h, 492, 498 (in Bos tau., B. urus=Bison europæus, B. bubalus; Egypt, ?Japan, ?N. Africa, ?Australia, ?India; Cervus elaphus, C. alces, C. capreolus, C. dama, Capra hircus; Ovis aries), 503, 504-515, fig. A, pl. 20, figs. 1–5 (syns. Fasc. hepatica Mueller, Fest. cervi Zed., Fasc. cervi Schrank, F. elaphi Gmelin, 1790; Monost. elaphi Zed., 1800; M. conicum Zed., 1803; Amphist. conicum Rud., 1809a, and of certain other authors), 530, 534, 535, 537, 539, 543, 565, 566, 590; 1904, 459, 460, 461, 462.—Braun, 1906, 142 (in Bos taurus).—Staff., 1905. Apr. 11. 693 (syn. Amphist. conicum Rud.) (in stomach of cattle: 1905, Apr. 11, 693 (syn. Amphist. conicum Rud.) (in stomach of cattle; Canada).

PARAMPHISTOMUM—Continued.

cotylophorum Fischder., 1901a, 370 (in Bos taurus, Togo; Bos zebu, German East Africa); 1902a, 23 (in B. t., Togo; B. z., German E. Africa); 1903h, 492, 499 (in Bos taurus, B. t. indicus; East Africa), 546-551, figs. f, 36-39 (in B. t., Togo, Misahöhe; B. zebu; Africa, Langenburg); 1904, 460, 463, 464, 465.

dicranocælium Fischder., 1901a, 369 (in Bos taurus indicus; Coll. Berlin Vet. School); 1902a, 18–19, 23 (in B. t. ind.); 1903h, 492, 499 (in B. t. ind.), 528–531, fig. d, pl. 21, figs. 5–17 (in B. t. ind.), 533, 534, 547; 1904, 458, 464,

465, 467.

epiclitum Fischder., 1904, 458–463, pl. 15, figs. 4–6, fig. B (in Bos taurus indicus, Buffelus indicus; Saigon, Cochin China).

explanatum (Crep., 1847) Fischder., 1904, 454-458, pl. 15, figs. 1-3, fig. A (in Bos

taurus indicus, B. zebu, Buffelus indicus).

gracile Fischder., 1901a, 368 (in Bos kerabau from Ceylon; Portax tragocamelus);
1902a, 16-17, 18-19 (in B. k., Ceylon; Portax trag.);
1903h, 492, 499 (in B. k., Ceylon, Por. trag.),
520-524, fig. B, pl. 21, figs. 10-11, 532, 535.—Linst., 1906, 175.

liorchis Fischder., 1901a, 368 (in Cervus simplicicornis, C. campestris, C. mexicanus, C. rufus, C. dichotomus, C. namby; Brazil); 1902a, 13–14 (syn. Amphist. conicum Dies. e. p.); 1903h, 492, 499, 515–518, pl. 20, figs. 6–7 (in same hosts).

microbothorium Fischder., 1902a, 21 (for microbothrium).

microbothrium Fischder., 1901a, 369 (in Antelope dorcas; Coll. Vien. and Coll. Berl. Vet. School); 1902a, 20–21 (microbothorium); 1903h, 492, 499, 535–538, pl. 22, figs. 21–23 (in Antilope dorcas), 539, 540, 541, 542.

orthocelium Fischder., 1901a, 369 (in Bos kerabau); 1902a, 17–18, 19, 20 (in Bos kerabau; Ceylon); 1903h, 492, 499, 524–528, fig. C, pl. 21, figs. 12–14 (in Bos kerabau; Ceylon, Kænigsberg i. Pr.), 529, 530, 531, 532, 534; 1904, 467.

scoliocalium Fischder., 1904, 459, 463–468, pl. 16, figs. 7–11, fig. C (in Buffelus indicus, in Cochin China and Annam; Bos taurus, at Annam).

strephocalium Fischder., 1902a, 19–20, 25, 50 (in Bos kerabau; Ceylon) for streptocelium.

streptocælium Fischder., 1901a, 369 (in Bos kerabau; from Ceylon); 1903h, 492, 499, 531-534, pl. 21, fig. 18, pl. 22, figs. 19-20 (in Bos ker.), 537, 539, 540; 1904, 465.

PARORCHIS Nicoll, 1907, 128, Zeugorchis Nicoll, 1906, not Staff., 1905, renamed, hence type acanthus.

PATAGIUM Heymann, 1905, 82, 90 (m. brachydelphium).

brachydelphium Heymann, 1905, 81, 82–89, pl. 6, figs. 1–3 (in Dermatemys mavii Gray; Coll. Kœnigsberg i. Pr.).

PECTOBOTHRIA Braun, 1893b, 188, for Pectobothrii.

PECTOBOTHRII Burm., 1837a, 530; 1856a, 243, 251.—Braun, 1890a, 515, 516; 1893b, 188 (Pectobothria).—Mont., 1888a, 83.—Tasch., 1879, 233.

PEDICELLINÆ Mont., 1888a, 88.

PEGOSOMUM Ratz, 1903, v. 1, 417–432, 1 pl. (type by present designation saginatum), Fasciolidæ.

asperum (Wright, 1879) Ratz, 1903, 423-424, 431-432 (in Botaurus minor).

saginatum (Ratz, 1898) Ratz, 1903, 419-421, 427-429, pl. 16, figs. 2, 4 (in Ardea alba; Hungary).

spiniferum Ratz, 1903, 422-423, 429-431, pl. 16, figs. 1, 3 (in Botaurus stellaris; Hungary).

PETRATHYRUS Cobbold, 1860a, 42, for Tetrathyrus.

obesus (Crep., 1851) Cobbold, 1860a, 42, renamed Monost. gurltii.

PEUDOCOTYLE Tasch., 1879, 65, apparently for Pseudocotyle.

PHACUS Nitzsch, 1827, 69, contains Cerc. pleuronectes, C. tenax.

PHANEROPSOLUS Looss, 1899b, 608–609, 611, 612, 633 (tod. sigmoideus) φανερός = visible; δψωλδς=penis); 1902m, 822, 823.—Braun, 1900, 234; 1901, 948; 1901e, 311, 313; 1901, 567.—Pratt, 1902, 889, 903, 904.

longipenis Looss, 1899b, 608, 714-715, fig. 35 (in ape, gen. et sp. undetermined; Gizeh Zool. Garden); 1902m, 823.—Braun, 1901e, 311, 313.

8588-No. 37-08-22

PHANEROPSOLUS-Continued.

micrococcus (Rud., 1819) Braun, 1901, 567; 1902b, 41, 49, 62, 64, fig. 40 (syns. Dist. mi. Rud., 1819; Dies., 1850a; D. (Brachylaimus) mi. Stoss., 1892).

orbicularis (Dies., 1850) Braun. 1901e, 348. pl. 20. fig. 14.

oviformis (Poir., 1886) Looss, 1899b, 609.

sigmoideus Looss, 1899b, 608, 712–713, 714, fig. 31 (in Passer dom. at Alexandria; Caprimulgus europæus at Cairo); 1902m, 823, 824 (probably — Dist. micrococcum Rud., see Braun).—Braun, 1901e, 311, 313; 1901, 567; 1902b, 63, 64.—Staff., 1905, Apr. 11, 693.

PHASCIOLA Wilder, 1894 (for Fasc.).—Stiles & Hass., 1898a, 89, 92 (syn. of Fasc.).
PHILOPHTHALMINÆ Looss, 1899b, 586-587; 1902m, 839.—Braun, 1902b, 31.—
Luehe, 1901, 488.—Odhn., 1905, 314.—Pratt, 1902, 888, 898 (includes; Philophthalmus, Pygorchis).—Shipley & Hornell, 1904, 95.

PHILOPHTHALMUS Looss, 1899b, 587 (tod. palpebrarum).—Braun, 1902b, 31, 32.—Ofenheim, 1900, 182.—Pratt, 1902, 888, 898.

lachrymosus and lacrymosus Braun, 1902b, 31, 37 (Dist. lucipetum Braun p. p. in Larus maculipennis; Brazil).—Looss, 1907, Mar. 5, 480.

lucipetus (Rud., 1819) Looss, 1899b. 587, 701, 702; 1907, Mar. 5, 480.—Braun, 1902b, 32. figs. 21–23 (syns. Dist. lucipetum Rud., 1819a; Brem., 1824; Dies., 1850; Braun, 1897; D. (Dicroccelium) luc. Duj., 1845; Stoss.)

nocturnus Looss. 1907, Mar. 5, 479-480, fig. 2 (in Athene noctua; Egypt).

palpebrarum Looss, 1899b, 587, 701-702, fig. 24 (in Corvus cornix at Cairo; Milvus parasiticus); 1901, 205; 1907, Mar. 5, 479, 480 (in Athene noctua; Egypt).—Braun, 1902b, 31, 32, 37.

PHYLLINE Abildg., 1790, 31 (syns. Tænia laticeps Pallas, Caryophyllus Gœze, Fasc. fimbriata Gœze).—Audouin. 1828a. 454-455.—Baer, 1827b. 674.—Baird, 1853a. 97 (=Caryophyllæus mutabilis Rud.).—Ben. 1858a, 1861a, 12. 19, 20.—Ben. & Hesse. 1864, 67 (of Oken, syn. of Nitzschia elegans).—R. Bl., 1888, 130 (of Oken, = trematode).—Braun. 1890a. 515, 518, 527.—Burm., 1856a, 251.—Dies., 1850a, 290, 426 (of Oken, syns. Hirudo Mueller, Epibdella Blainv., Trist. Rathke). 428 (of Oken, syns. of Trist.), 445 (of Oken, syn. of Malacobdella), 577 (of Abildg., syn. of Caryophyllæus), 578 (of Abildg., syn. of C. mutab.); 1858e, 313, 363 (of Oken, mentions only hippoglossi); 1859c, 437.—Fabricius, 1794, 30.—Goldb., 1855a, 20.—Johnston, 1865, 32.—Mont., 1888a, 84, 87; 1902, 138, 143; 1905, 75, 76.—Nitzsch, 1826, 150.—Odhn., 1905, 371 (of Oken, type is diodontis=Trist, maculatum).—Rud., 1809a, 24.—Stoss., 1898, 6 (of Oken).—Tasch., 1878, 563, 566 (syn. of Trist, maculatum=Capsala martinieri).

1891: Phyllinic Sons., 1891, 262, for Phylline.

bumpusii (Lint., 1900) Linst., 1903, 355.

caligi Kroyer.—Ben., 1858a, 1861a, 13 (syn. of Udonella caligarum Johnston).

coccinea (Cuvier, 1817) Schweigger. 1820, 474.—Dies., 1850, 429 (syn. of Trist. rudolphianum).—Johnston, 1865, 33.—Stoss., 1898, 5.—Tasch., 1878, 567 (cocinea) (syn. of T. molæ).

cocinea Tasch., 1878, 567 (for coccinea).

diadema (Mont., 1902) Linst., 1903, 355.

diodontis Oken, 1817, 182, 370. pl. 10, fig. 3 (based on Martinière, 1787a, 207–208, figs. 4–5, on Diodon sp.; Nolka to Monterey, Cal.).—Des., 1850a, 430 (syn. of Trist. maculatum Rud.).—Nitzsch, 1826, 150.—Odhn., 1905, 371.—Tasch., 1878, 567 (syn. of Trist. mac. Rud.).

grossa (Mueller, 1788) Johnston, 1865a, 35.—Dies., 1850a, 445 (to Malacobdella).—

Leidy, 1852, 209 (= Malacobdella grossa Blainy.).

hendorffii Linst., 1889e, 163–180, pls. 10–11 (anatomy) (on Coryphæna hippurus; Caleta buena, Chile); 1893f, 170–172.—Braun, 1890a, 421, 426.—Mont., 1891, 126.

hippoglossi (Mueller, 1776) Oken, 1815.—Ben., 1858a, 1861a, 21 (to Epibdella).—Ben. & Hesse, 1864, 69 (to Epibdella).—Dies., 1850a, 426-427 (syns. Epibdella hipp. Blainv., Hirudo hipp. Mueller, Trist. hamatum Rathke): 1858e, 363; 1859c, 437 (in Hippoglossusvulgaris; Belgium).—Johnston, 1865a, 32; —, 431, pl. 15, figs. 1-3.—Keelliker, 1849, 21.—Linst., 1903, 355.—Moq.-Tandon, —, 392.—Nord., 18—, 526.—Odhn., 1905, 370 (to Epibdella), 371.—Tasch., 1878, 564, 565, 568 (to Trist.).—Thompson, ——, 482.—Reported also for Hippoglossus gigas, H. maximus.

PHYLLINE-Continued.

monticellii Par. & Perugia, 1895, 2.—St.-Remy, 1898, 534 (to Epibdella).—Stoss., 1898, 7 (in Mugil auratus; Triest).

sciænæ (Ben., 1856) Sons., 1891, 263 (in Sciæna umbra, Umbrina cirrhosa), 262 (Phyllinic).—Linst., 1889a, 76 (syn. of Benedenia elegans).

soleæ (Ben. & Hesse, 1863) Linst., 1903, 355.

squamula (Heath, 1902) Linst., 1903, 355.

PHYLLOCOTYLE Ben. & Hesse, 1863, 1864, 96, 103 (m. gurnardi).—Braun. 1890a, 413, 477, 516, 517, 523, 534, 536, 546; 1893a, 890.—Cerf., 1895h, 918; 1896, 514.—Gamb., 1896a, 73.—Hoyle, 1890. 539.—Mont., 1888a, 7, 8, 11, 66, 67, 86, 89, 100; 1892, Oct. 7, 213 (g. of Octocotylinæ); 1903, 336 (subf. Plectanocotylinæ); 1905, 76, 77, 78.—Pratt, 1900, 651 (on gills of marine fishes), 656, fig. 29.—Scott, T., 1901, 147.—Tasch., 1879, 69; 1879, 239, 247.

gurnardi Ben. & Hesse, 1863; 1864, 103–104, pl. 10, figs. 1–7 (in Trigla gurnardus).—
Braun, 1890a, 418, 536, 548.—Mont., 1888a, 8, 13, 16; 1905, 76.—Pratt, 1900, 656, fig. 29; 657.—Scott, 1901, 147–148, pl. 8, fig. 23 (in Trigla gurnardus); 1905, 115–116, pl. 6, figs. 19–20 (in Trigla gurnardus): 1905, 116 (of Scott, 1901, 147, pl. 8, fig. 23) (syn. of Plectanocotyle lorenzii).—Tasch., 1879, 247 (in

Trigla gurnardus).

PHYLLODISTOMUM Braun, 1899g, 492 (tod. folium) (Braun also places here: D. cygnoides Zed., after Looss, 1894, 23; D. cymbiforme Rud.; D. patellare Sturg.); 1901b, 9-10.—Looss, 1900d, 605; 1901b, 202, 203, 204, 207 (Gorgoderinæ; folium, patellare); 1901l, 557, 558; 1902m, 476, 477, 478, 479, 480, 515, 795, 797, 813, 814, 824, 827, 844, 847, 856, 857, 858, 859, 860, fig. 3; 861, 862 (Gorgoderinæ).—Odhn., 1901, 64, 65, 66, 67; 1902, 37; 1902, 64, 65, 66.—Pratt, 1902, 888, 900, 901.—Stiles, 1901, 183, 185.—See also Rhopalocerca. 1899; Spathidium Looss, 1899b, 605 (folium) [not Duj., 1841].

acceptum Looss, 1901b, 203 (in bladder of Crenilabrus pavo, C. griseus at Triest); 1901d, 404–405, fig. 4; 1902m, 480, 782, 798, 844, 857.—Odhn., 1901, 66;

1902, 66.

americanum Osborn, 1903, 252–258, figs. 1–4 (in Amblystoma punctatum; Minnesota); 1903, 532–533 (in A. tigrinum Green; Minnesota).

conostomum (Olss., 1876) Looss, 1902m, 857.

cymbiforme (Rud., 1819) [Braun, 1899, 492] Luehe, 1900, 564; 1901b, 10–13, fig. 1.—

Looss, 1902m, 469 (to Plesiochorus, type).

folium (Olfers, 1816) Braun, [1899, 492:] 1901, 947; 1902b, 146.—Looss, 1901, 202; 1902m, 480, 782, 797, 798, 827, 857, 862.—Luehe, 1901, 54.—Osborn, 1903, 252, 254, 255; 1903, 533.—Ssinitzin, 1905, 75, 101, 108; 1906, 683 (near Warschau), 684 (in Cottus gobio. Dreissenia polymorpha), 685 (to Gorgoderina).—Staff., 1904, May 3, 492 (in bladder of Esox lucius; Canada).

linguale Odhn., 1902, 66 (in Gymnarchus niloticus; Omdurman, Sudan).—Looss,

1902m, 480, 857, 861.

patellare (Sturges, 1897) [Braun, 1899, 492].—Looss, 1901b, 202; 1902m, 480, 844, 857.—Osborn, 1903, 252, 254, 255; 1903, 533.

spatula Odhn., 1902, 66-67 (in Bagrus docmac, B. bayad; Omdurman, Sudan).—
 Looss, 1902m, 480, 861 (type of Catoptroides).—Osborn, 1903, 254; 1903, 533.

spatulæforme Odhn., 1902, 67-68 (in Malapterurus electricus; Omdurman, Africa).— Looss, 1902m, 480, 861 (to Catoptroides).

superbum Staff., 1904, 492 (in Ameiurus nebulosus, Perca flavescens; Canada; fish probably brought to Montreal).

unicum Odhn., 1902, 66 (in Serranus sp.; Tor in Sinai, Red Sea).—Looss, 1902m, 480, 857, 861.—Osborn, 1903, 255: 1903, 533.

PHYLLONELLA Ben. & Hesse, 1863; 1864, 65, 66, 70-71 (m. soleæ).—Braun, 1890a, 412, 414, 469, 471, 475, 490, 498, 511, 516, 517, 519, 523, 526, 527.—Gamb., 1896a, 73.—Massa, 1906, 43.—Mont. 1888a, 7, 10, 13, 20, 66, 86, 87, 97; 1892, Oct. 7, 172, 213 (syn. of Epibdella); 1899, 98; 1902, 139 (Phylonella), 140, 142; 1903, 335 (=Epibdella).—Pratt, 1900, 848.—Tasch., 1878, 566 (syn. of Trist.) (Phylonella).—Scott, T., 1901, 142.

1878: Phylonella Lorenz, 1878a, 434.

hippoglossi (Mueller, 1776) Goto, 1899, 264–269, pl. 20, figs. 1–7.—Linst., 1903, 280.—Mont., 1902, 140; 1905, 75, to (Epibdella).

PHYLLONELLA—Continued.

soleæ Ben. & Hesse, 1863; 1864, 70–71, pl. 5, figs. 1–8 (in Solea vulgaris).—Braun, 1890a, 409, 418, 424, 446, 465, 475, 512, 519, 528, 547, 552.—Cunningham, 1890a, 93–96, figs. C–D.—Goto, 1899, 266.—Lorenz, 1878a, 434.—Mont., 1892, 106, 115, 124, 125, 126 (to Epibdella); 1902, 139 (to Epibdella).—Pratt, 1900, 655, 657, fig. 6.—Scott, 1901, 142–143, pl. 8, fig. 17 (in S. v.; Clyde River); 1901, 344 (in Solea vulgaris; Irish Sea); 1905, 118 (in Scott, 1901, pl. 8).—Tasch., 1878, 564, 568 (Phylonella) (to Trist.).

PHYLONELLA Lorenz, 1878a, 434, for Phyllonella.

PINTNERIA Poche, 1907, Jan. 4, 125. Hoploderma Cohn, 1893, not Michælis, renamed, hence type mesoccelium.

PLACOPLECTANUM Dies., 1858e, 315, 384 (Discocotyle, 1850, renamed, hence type sagittatum; also type by first species rule and because it is the only positive species).—Braun, 1890a, 518.—Tasch., 1879, 239.

hirundinaceum (Bartels, 1834) Dies., 1858e, 384, to (Discocotyle).

leptogaster (Leuck., 1830) Dies., 1858e, 384 to (Discocotyle).—Tasch., 1879, 245 (to Octobothrium).

sagittatum (Leuck., 1842) Dies., 1858e, 384.—Tasch., 1879, 244 (to Octobothrium).— Linst., 1879 (reported for Catostomus teres).

PLACUNELLA Ben. & Hesse, 1863; 1864, 65, 66, 71-74 (pini type by page prece-ASILIA Ben. & Hesse, 1803, 1804, 03, 00, 71–74 (plin type by page precedence).—Braun, 1890a, 411, 412, 415, 469, 498, 511, 516, 517, 519, 523, 526, 528; 1891d, 422.—Gamb., 1896a, 73.—Massa, 1903, 252; 1906, 43, 44, 51 (syn. of Trochopus), 58 (Ptacunella).—Mont., 1888a, 7, 10, 11, 13, 52, 66, 86, 87, 97; 1891, 104, 105, 107, 111; 1899, 98; 1903, 335 (—Trochopus).—Par. & Perugia, 1890, 13.—Par. & Mont., 1902, Dec., 46.—Pratt, 1900, 646, 648.—St.-Remy, 1898, 534.—Scott, T., 1901, 143.—Stoss., 1898, 7–8.—Tasch., 1878, 566 (syn. of Trick Chr.) of Trist. Cuv.).

1906: Ptacunella Massa, 1906, 58, misprint.

exacantha Mont., 1891, 104, for hexacantha.

hexacantha Par. & Perugia, 1889, 740-741, fig. 1 (in Serranus gigas; Genova); 1890, 740–741, fig. 1: 1890, 5/, 1894, 594.—Braun, 1890a, 418, 528, 547, 552.—Massa, 1906, 47, 59 (to Trochopus).—Mont., 1891, 1892g, 104, pl. 6, fig. 31 (exacantha).—Par., 1899, 3: 1902, 2 (in S. g.; Elba).

pini Ben. & Hesse, 1863: 1864, 72, 73, pl. 5, figs. 9-18 (in Trigla pini).—Braun, 1890a, 418, 465, 528, 547, 552.—Par. & Perugia, 1889, 740; 1890, 740; 1890, 5; 1899, 3.—Massa, 1906, 57 (to Trochopus), 64 (of Scott, 1901, 344) (syn. of Trochopus diplacanthus Massa).—Mont., 1888a, 11.—Par., 1902. 2 (in Trigla hirundo; Elba).—Pratt, 1900, 655, 657, fig. 8.—Scott, 1901, 344–345, 1 fig. (in Trigla hirundo; Irish Sea); 1901, 143 (in Trigla pini).—Tasch., 1878, 564-685 (to Trist.).

rhombi Ben. & Hesse, 1863; 1864, 73-74, pl. 6, figs. 1-7 (in Rhombus maximus).— Braun, 1890a, 412, 419, 465, 528, 547, 551.—Massa, 1906, 58 (to Trochopus).— Mont., 1888a, 11.—Tasch., 1878, 564, 568 (to Trist.).

vallei Par. & Perugia, 1895, (3) (in Naucrates ductor; Genova).—Looss, 1902m, 746.— Massa, 1903, 252; 1906, 44.—St.-Remy, 1898, 534.—Stoss., 1898, 8 (in Naucrates ductor; Trist.).—Type of Ancyrocotyle 1903.

PLACUNELLÆ Par. & Perugia, 1889, 740; 1890, 740.

PLAGIOPELTINÆ Mont., 1903, 336 (subf. of Octocotylidæ).

PLAGIOPELTIS Dies., 1850a, 289, 416–417 (m. duplicata); 1858e, 314, 368–369.—

Ben. & Hesse, 1864, 84.—Braun, 1890a, 518, 546.—Cerf., 1895h. 920; 1896.

515.—Goldb., 1855a, 19.—Mont., 1903, 336 (syn. of Hexacotyle)—Tasch., 1879, 249 (syn. of Hexacotyle Blainv.).—See Hexacotyla.

duplicata (Rud., 1819) Dies., 1850a, 417 (includes Polyst. thynni Delaroche,

1811; Hexacotyla thynni Blainv.); 1858e, 369 (in Thynnus brachypterus; Balearic Isles, Vindobonæ).—Tasch., 1879, 250 (syn. of Hexacotyle thynni (Delaroche)).

thynni (Delaroche, 1811) Braun, 1891d, 421.

PLAGIOPORUS Staff., 1904, May 3, 493–494 (m. serotinus); πλάγιος, oblique; πόρος, pore.

serotinus Staff., 1904, May 3, 493-494 (in Moxostoma macrolepidotum L. S.; Canada).

- PLAGIORCHIDÆ Luehe, 1901, 173; 1901. 486, 487.—Looss, 1902m, 839, 840 (includes: Lepodermatinæ Looss, Brachycœliinæ Looss, Pleurogenetinæ Looss, also Prymnoprion, Haplometra, Pneumonœces (=Hæmatolæchus), Saphedera (=Macrodera), Opisthogonimus Luehe, Anchitrema, Dist. mutabile, Dicrocœliinæ).
- PLAGIORCHIINÆ Pratt, 1902, 888, 898 (includes: Opisthioglyphe, Plagiorchis; related genera: Glossidium, Enodiotrema, Pachypsolus).

PLAGIORCHINE Luehe, 1901. 173.

PLAGIORCHIS Luehe. 1899k, 531, 532, 533, 534 (tld. D. lima): 1900. 558; 1901. 487 (lima designated type).—Braun, 1901b, 56, 58; 1901i, 56, 58; 1901. 563, 564, 567, 568; 1902b, 37, 40, 41, 42, 43, 46, 47, 49, 50, 53, 54, 94, 125 (syn. Lepoderma Looss, 1899).—Looss, 1900, 607; 1901b, 207; 1902m, 813, 824; 1907, Mar. 5, 483.—Pratt, 1902a, 888, 899.—Staff, 1905, Apr. 11, 691.—Stoss., 1904, Feb. 11, 1–2, 1 pl., 1 fig.; 1905, 31 Jan., 24.—(See also Dist. cirratum, D. lima, mentulatum, and probably D. erraticum, D. ramlianum, D. reniforme (=unicum[=renifera]), D. horridum Leidy, possibly didelphydis).

asperus Stoss., 1904, 1-2, 1 pl., 1 fig. (in intest. ten. of Plecotus auritus: Grisi-

gnana, Istria).

cirratus (Rud., 1802) Luehe, 1899, 530.—Braun. 1901, 564, 566; 1902b, 43, 46, 47, 50 (syns.: Fasc. cirrhata Rud., 1802, 66; Dist. cirr. Rud., 1809a, 376; 1819a, 100; Dies., 1850a, 350; Mueh., 1896, 262; Dist. (Brachylaimus) cirratum Duj., 1845a, 413; Stoss., 1892, 11).—Staff., 1905, Apr. 11, 693.

didelphidis (Par., 1896) Stoss., 1904, table (in Didelphis marsupialis; Paraguay).

elegans (Rud., 1802) Braun, 1902b, 38, 42, 43, 46, 47, 50 (syns.: Fasc. elegans Rud., 1802, 65; Dist. elegans Rud., 1809a, 375; 1819a, 100; Crep., 1829, 59; Dies., 1850a, 349; Dist. (Brachylaimus) elegans Stoss., 1892, 11, as syn. of D. cirratum; Duj., 1845a, 414; D. erraticum Linst., 1894, 335; D. elegans Rud. =D. cirratum Rud., Mueh., 1896; 589).—Stoss., 1904, table (in Uccelli insettivori diversi; Europe).

horridus (Leidy, 1850) Stoss., 1904, table (in Boa, India: Python, America).

[lima (Rud., 1809) Luehe, 1899k, 930-931.]

maculosus (Rud., 1802) Braun. 1901. 943; 1902b, 45, 46, 47, 54, 93, 94, figs. 27, 28 (syns.: Fasc. hirundinis Freelich?, 1791, 75; Dist. hirundinis Zed.?, 1800a, 169; Fasc. masculosa Rud., 1802, 67; Dist. maculosum Rud., 1809a, 374; Duj., 1845a, 412; Dies., 1850a, 349; D. maculosum =P. maculosum Braun, 1901, 566; D. (Dicroccelium) maculosum Olss., 1876, 14; D. (Brachylaimus) maculosum Stoss., 1892, 11; D. crassum Olss., 1876, 25 nec Siebold).— Kowal., 1902d, 27 [9] (maculosa); 1904, 25 [10] (in Hirundo rustica; Dublany).

mentulatus (Rud., 1819) Stoss., 1904, table (in Tropidonotus, Lacerta, Podarcis;

Europe).

- nanus (Rud., 1802) Braun, 1901, 567; 1902b, 47, 50, figs. 29, 30 (syns.: Fasc. nana Rud., 1802, 68; Dist. nanum Rud., 1809a, 376; 1819a, 101; Duj., 1845a, 446; Dies., 1850a, 350; Stoss., 1892, 41).—Stoss., 1904, table (in Glareola, Scolopax: Greifswald).
- permixtus Braun, 1901, 943 (in Hirundo rustica; Coll. Vienna); 1902b, 46, 54, figs. 33, 34.—Stoss., 1904, table (in rondine; Vienna).
- ramlianus (Looss, 1896) Stoss., 1904, table (in Cameleonte: Egitto).—Type of Lepoderma, 1899.

sauromates (Poir., 1886) Stoss., 1904, table (in Elaphis).

triangularis (Dies., 1850) Braun, 1901, 568; 1902b, 51, figs. 32, 33 (syns: Dist. triangulare Dies., 1850a, 351; Stoss., 1892, 44; D. (Brachylaimus) meropis Par., 1896, 5; Megacetes triangularis Looss, 1899, 725).—Stoss., 1904, table (in Merops apiaster; Vienna).

respertilionis (Mueller, 1784) Braun, 1900, 217–220, 228, 230, pl. 10, figs. 1–2.— Staff., 1905, April 11, 693 (syn. Dist. lima; in Vespertilio subtilis Say; Can-

ada).—Stoss., 1904. 1. table (in Chirotteri diversi; Europa centrale).

vitellatus (Linst., 1875) Braun, 1901, 943 (in Actitis hypoleucus); 1902b, 50, 51, fig. 31 (syns. Dist. vitellatum Linst., 1875, 189; D. (Brachylaimus) vitellatum Stoss., 1892, 12).—Stoss., 1904, table (in Actitis hypoleucus; Ratzeburg).

PLANARIA Mueller. 1776 (worm); 1780, 210; Gœze, 1782a (trematode); Brown. 1827 (mollusk); Lea, 1833 (mollusk). [The early literature on Planaria is rather confused, containing references to turbellaria, trematodes, and mollusks. The name is now confined to the fresh-water turbellarian group Planaria.

PLANARIA—Continued.

nariidæ. The list of species given below contains all the specific (trematode and others) names which we have thus far had occasion to index. We have not worked out the type species, except for Geze, 1782.]

 Dies., 1850a, 412 (syn. of Polyst.).—Tasch., 1879, 251 (syn. of - of Braun, -Polyst.).

- of Geze, 1782a, 41, 168-179, a genus or "Geschlecht" containing among others Fasc. hepatica, hence takes this as type; Goze divides Planaria into 4 Fasc. hepatica, hence takes this as type; Gueze divides Frankia into 4 ("Gattungen": Latiuscula, Teres seu Cylindrica, Alata seu Dubia, Subclavata.—Abildg., 1790a, 36 (syn. of Dist.); 1794, 58.—Brand., 1888a, 8.—Braun, 1893a, 883, 884, 893, 894, 902, 908.—Crep., 1837, 309.—Dies., 1850a, 307 (syn. of Hemist.), 312 (syn. of Holost.), 318 (syn. of Diplodiscus), 331 (of authors, syn. of Dist.), 388 (of Gœze, 1782a, 177, pl. 16, syn. of Dist. ferox).—Lamouroux, 1824a, 559 (syn. of Dist.).—Nitzsch, 1819, 397.—Raficastical 1811 (syn. of Monoctomers Park). Bud. 1809a, 22.—Stiles 1901 nesque, 1815, 151 (syn. of Monostomeus Raf.).—Rud., 1809a, 22.—Stiles, 1901, 180.—Stiles & Hass., 1898a, 88, 92 (syn. of Fasc.).

of Linn.—Bosc, 1802a, v. 1, 248–254.

fresh water.—Knapper, 1865a, 39 pp., 2 pls.; 1866a, 271–272. - land.—Kennel, 1878a, 26-29 (Dorpat).—Mosley, 1874, 132 (Ceylon).

- marine.—Collinwood, 1876a, 83–98, pls. 17–19 (31 species).—Gœtte, 1878a,

75-76.—Keferstein, 1869a, 3-38, pls. 1-3 (St. Malo).—Kirk, 1882, 267-268.— Monti, 1900, 1-16, 2 pls. (regeneration).—Quatrefages, 1845, 129-184.

miscellaneous.—Agassiz, 1866, 306-309.—Baer, 1826a, 126; 1827b, 1828d, 183-187 (of Duges).—Bardeen, 1902, 262-288, 12 figs. (embryology, regenerative development); 1903, 1-20, 18 figs. (heteromorphosis).—Bory de St. Vincent, 1828b, 11.—Calkins, 1901b, 12 (of Agassiz).—Curtis, 1905, July 4, 169-175, 2 figs. (location of pharynx in embryo); 1905, 855-856.—Cuvier, 1817, 43.—Dalyell, 1814a.—Dugès, 1828a, 139–183, pls. 4–5 (planariées); 1830a, 169–183; 1830b, 72–90, figs. 1–26; 1833b, 619–625, figs. 1–26.—Fabricius, 1794, 20; (1798a), 52–66, pl. 2.—Faraday, 1832a, 723–724; 1833a, 183–185; 1834a, 994.—Fischer, 1799, 96.—Geddes, 1880a, 51–58; 1880b, 99–100.—Gerstfeldt, 1859a, 261–263.— Girard, 1850a, 398–402; 1850b, 300–308; 1851a; 1851b; 1854a (from Carolina).— 1883c, 238.—Kælliker, 1846a, 291–295, figs. 1–13.—Lamarck, 1801a, 330.— Lankester, 1890, 835–836.—Lawson, 1861a; 1861b.—Leidy, 1847e, 252 (2 sp.); 1848a,78–79.—Leuck., 1863,154.—Morgan, 1904, 502–505.—Pallas, 1781a, 59.— Rathke, 1799, 82; 1799, 147, pl. 2, figs. 7–8.—Ryder, 1882, 48–51, figs. 1–10 (parasitic on Limulus).—Sabussow, 1904, 1–28.—Schneider, 1866, 11.—Slawikowski, 1819, 56.—Thienemann, 1906, 499–504.

alata Gœze, 1782a, 176–177, pl. 14, figs. 11–13 (t. h. the fox).—Brand., 1888a, 60 (to Hemist.).—Dies., 1850a, 307–308 (to Hemist.).—Nitzsch, 1819, 399 (to Holost.).—Rud., 1809a, 402 (to Dist.).—Ward, 1895, 341 (to Hemist; in Canis familiaris).

alpina (Dana).—Collin, 1891b, 177–180; 1891c, 177–180.—Borelli, 1905, 4 pp.—Brand., 1900, 303–304.—Fredericq, 1905, 199–200; 1905 (X. 18), 597 (in Belgium).

angulata (Mueller, 1774) Fabricius, 1798, 53–58, pl. 11, figs. 1–7.—Bosc, 1802a, v. 1, 256.—Herbst, 1787a, 34.

ardex (Gmelin, 1790) Rud., 1809a, 432 (see Dist. ardex stellaris Rud.).

atomata Bosc, 1802a, v. 1, 260.

auriculata Bose, 1802a, v. 1, 261.

badia Rathke, 1799, 147, pl. 3, figs. 9, a-b.

bicornis Bosc, 1802a, v. 1, 257.

bilis Braun, 1790a, 61, pl. 3, figs. 4-5 (in Falco melanaëtus; Germany); 1792, 61, pl. 3, figs. 4–5—Braun, 1901f, 561.—Dies., 1850a, 376 (syn. of Dist. crassiusculum).—Gmelin, 1790a, 3054 (to Fasc.).—Rud., 1809a, 408 (=Dist. crassiusculum Rud.).

brunnea (Mueller, 1774) — —, 1776.—Fabricius, 1798, 64-66, pl. 11, figs. 13-15.— Bosc, 1802a, v. 1, 255 (brunea).—Herbst, 1787a, 33.

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PLANARIA-Continued.
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candida (Mueller, 1774) — —, 1776.—Fabricius, 1798, 62-64, pl. 11, figs. 11-12.— Bosc, 1802a, v. 1, 262.—Herbst, 1787a, 35.

capitala Bosc, 1802a, v. 1, 261 (for capitata?), in Baltic Sea.

capitata (Mueller, 1774) — —, 1776.—Herbst, 1787a, 35.

carnea Rathke, 1799, 83, 147, pl. 3, figs. 10 a-b.

caudata (Mueller, 1774) Bosc, 1802a, v. 1, 261.—Herbst, 1787a, 35.

cavatica Fries, — ___, 151-153.

ciliata (Mueller, 1774) — —, 1776.—Bosc, 1802a, v. 1, 255.—Herbst, 1787a, 34.

cornuta Mueller, 1776a, 221.—Bosc, 1802a, v. 1, 260.—Herbst, 1787a, 34.

crenata (Mueller, 1774) — —, 1776.—Bosc, 1802a, v. 1, 259.—Herbst, 1787a, 34.

cylindrica Goze, 1782, 173: P. teres s. cylindrica, 174 (syn. of cylindraceum) (in frogs).—Dies., 1850a, 368 (syn. of Dist. cylindraceum).—Looss, 1894a, 64 (syn. of Dist. cylindraceum).—Rud., 1809a, 393.

dubia Gœze, 1782a, 177 (=P. alata).

ehrenbergii Focke, 1836a, 191-206, pl. 17, figs. 1-19.

festæ Borelli, 1898, 6 pp., 2 figs., var. albolineata.

filaria Bosc, 1802a, v. 1, 261.

flaccida (Mueller, 1774) — —, 1776.—Bosc, 1802a, v. 1, 256.—Herbst, 1787a, 34.

fulva Bosc, 1802a, v. 1, 257–258.

fusca Fabricius, 1798, 58 (syn. Fasc. angulata Mueller); 1799, 151.—Bosc, 1802a, v. 1, 258.

fuscescens Fabricius, 1798, 58-62, pl. 11, figs. 8-10; 1799, 151.

gesserensis Bosc, 1802a, v. 1, 262.

glauca (Mueller, 1774) Herbst, 1787a, 43.—Bosc, 1802a, v. 1, 258.

gonocephala Ackermann, 1905, 137-139, 4 figs.

grisea Bosc, 1802a, v. 1, 257.

grossa (Mueller, 1774) — —, 1776.—Bosc, 1802a, v. 1, 260.—Herbst, 1787a, v. 1, 35.

gulo (Mueller, 1774) — —, 1776.—Bosc, 1802a, v. 1, 255.—Herbst, 1787a, 34.

helluo (Mueller, 1774) Herbst, 1787a, 35.—Bosc, 1802a, v. 1, 259.

hirudo Johnston, 1846a, 437, pl. 15, fig. 3.—Dies., 1850a, 473.

ignorata Raspail, 1902, 119-123.

inquilina Graff, 1904, 457 (in Schnecken).

insignis Graff, 1904, 457 (in Schnecken).

intestinalis Mueller.—Abildg., 1790, 34 (syn. of Fasc.).

lactea (Mueller, 1774) Herbst, 1787a, 34.—Baer, 1826a, 126.—Bosc, 1802a, v.1, 258–259.—Nord., 1832a, 69.—Tasch., 1879, 36.

lagena Braun, 1788a, 257, pl. 10, figs. 1–3.—Dies., 1850a, 380 (syn. of Dist. nodulosum).—Gmelin, 1790a, 3057 (to Fasc.).—Looss, 1894a, 33 (syn. of D. nod.).—Rud., 1809a, 410, 412 (syn. of D. nod.).

latiuscula Gœze, 1782a, 169, 171.—Anacker, 1892c, 94.—Dies., 1850a, 333 (syn. of Dist. lanceolatum), 385 (syn. of D. echinocephalum Rud.).—Dunglison, 1893, 875.—Macé, 1882, 25.—Rud., 1809a, 353 (=Fasc. hepatica), 429 (=Dist. falconis milvi Rud.).—Stiles & Hass., 1898a, 88.—Stiles, 1898a, 29.—Ward, 1895, 246 (syn. of Fasc. hep.).

laurentiana Borelli, 1897, 4 pp., 1 fig.

limuli Graff (1879), 202-205.

linearis (Mueller, 1774) Herbst, 1787a, 35.—Bosc, 1802a, v. 1, 260.—Rathke, 1799, 83, 147, pl. 3, fig. 11.

lineata (Mueller, 1774) —, 1776.—Bosc, 1802a, v. 1, 258.—Herbst, 1787a, 34.

lingua Bosc, 1802a, v. 1, 262.

littoralis Herbst, 1787a, 35.

lucii (Mueller, 1776) Gœze. 1782a, 172, pl. 14, fig. 3.—Ben.. 1858a. 1861a, 100
(syn. of D. teret.).—Dies., 1850a, 358 (syn. of Dist. tereticolle).—Looss, 1894a, 5 (syn. of D. teret.).—Rud., 1809a, 380.—Sramek, 1901, 105 (syn. of D. teret.).

lugubris Morgan, 1901, 179-212, 14 figs; 1902, 132-139, 24 figs.

PLANARIA—Continued.

maculata Leidy, 1847e, 252; 1851b (free; Philadelphia); 1904a, 11, 54 (maculta).—
Bardeen, 1901, 351–352 (regeneration).—Curtis, 1900, 56–59, 9 figs. (anat., development of reproductive organs); 1901, 357–359 (asexual reprod.); 1902, 515–559, 11 pls. (biol., reprod. organs).—Morgan, 1898–1900, 364–397, 41 figs.; 1904, 683–695, 1 fig.—Thacher, 1902, v. 36, 633–641; 1903, 24 Feb., 115–116.

maculta Leidy, 1904a, 54, for maculata.

marmorata Bosc, 1802a, v. 1, 262 (free form).

marmorosa (Mueller, 1774) Herbst, 1787a, 35.

melis [Gœze, 1782a, 176, pl. 14, figs. 9-10, Planaria in Dachs, March].—Dies., 1850a, 381 (syn. of Dist. trigonocephalum).—Rud., 1809a, 415.

midx (=mydx renamed?) Dies., 1850a, 325 (syn. of Monost. trigonocephalum).—Braun, 1901b, 42.

montenigrina Mrazek, 1904a, 43 pp., 2 pls., 2 figs.; Montenegro.

mutabilis Eichwald, —, 78, pl. 9, fig. 16.—Dies., 1850a, 649.

mydx Braun in Rud., 1809a, 336 (=Monost. (Monost.) trigonocephalum Rud., 1809).

nigra (Mueller, 1774) Herbst, 1787a, 33.—Bosc, 1802a, v. 1, 255.—Chiaje, 1837a, 14.

notulata Bosc, 1802a, v. 1, 254-255, pl. 8, figs. 7-8.

obscura (Mueller, 1774) ——, 1776.—Bosc, 1802a, v. 1, 259.—Herbst, 1787a, 35.

operculata Herbst, 1787a, 36.—Bosc, 1802a, v. 1, 256.

polychroa Tasch., 1879, 36.

punctata (Mueller, 1774) Herbst, 1787a, 34 [6], 35.—Bosc, 1802a, v. 1, 255.

pusilla Braun, 1790a, 63-65, pl. 3, figs. 6-7 (in Erinaceus europæus; Europe).— Dies., 1850a, 360 (to Dist.).—Gmelin, 1790a, 3055 (to Fasc.).—Rudolphi, 1809a, 384.

pusio Eichwald, —, 79, pl. 9, fig. 17.—Dies., 1850a, 648.

putorii Gmelin, 1790a, 3053.—Dies., 1850a, 381 (syn. of Dist. trigonocephalum).—
 [Gœze, 1782a, 175, pl. 14, figs. 7-8].—Nord., 1840, 621 (syn. of Fasc. trigonocephala).—Rud., 1809a, 415.

quadrangulata Bosc, 1802a, v. 1, 257.

radiata (Mueller, 1774) —, 1776.—Bosc, 1802a, v. 1, 260.—Herbst, 1787a, 35. rosea (Mueller, 1774) Herbst, 1787a, 34.—Bosc, 1802a, v. 1, 256.—Lamarck, 1801a, 330.

rostrata (Mueller, 1774) ——, 1776.—Bosc, 1802a, v. 1, 259.—Herbst, 1787a, 35. rubra (Mueller, 1774) ——, 1776.—Bosc, 1802a, v. 1, 256.—Herbst, 1787a, 34. rutilans Bosc, 1802a, v. 1, 258.

schlosseri Graff, 1904, 457 (in Botryllus schlosseri).

simplicissima Morgan, 1904, 385-393, 20 figs.—Stevens, 1904, 208-220, 4 pls.

simplissima Curtis, 1900a, 10 Apr., 447–466, pls. 31–32, figs. 1–14 (reproductive system); 1900b.

stagnalis (Mueller, 1774) ——, 1776.—Bosc, 1802a, v. 1, 255.—Herbst, 1787a, 33. striata Herbst, 1787a, 34.

strigata (Mueller, 1774) ——, 1776.—Bosc, 1802a, v. 1, 260.—Herbst, 1787a, 35.

strigis (Schrank, 1788).—Fischder., 1901, 367; 1902a, 7 (=Holost. macrocephalum), type of Strigea; 1903h, 490.

subclavata Geeze, 1782a, 178–179, pl. 15, figs. 2–3 (t. h. Rana, intestine; Europe) (restricted from Fasc. subclavata, ore sessile, Pallas).—Ben., 1858a, 1861a, 81, 82 (to Amphist.).—Dies., 1850a, 318 (to Diplodiscus).—Nitzsch. 1819, 398 (to Amphist.).—Nord., 1840, 627 (to Amphist.).—Olfers, 1816, 45.—Rud., 1809a, 348 (to Amphist.).

subulata Herbst, 1787a, 36.—Bosc, 1802a, v. 1, 257.

tentaculata (Mueller, 1774) ——, 1776.—Baer, 1826a, 126.—Bosc, 1802a, v. 1, 259.—Herbst, 1787a, 34.

teres seu cylindrica Gœze, 1782a, 173.—Linst., 1905, 191.—Looss, 1899b, 680.

teres duplici poro Gœze, 1782a, 173.—Baird, 1853a, 55 (=Dist. trigonocephalum).

PLANARIA-Continued.

teres poro simplici Geeze, 1782a, 173. 174, pl. 14, figs. 4-6.—Baird, 1853a, 47, 55 (=Holost. macrocephalum).—Dies., 1850a, 312 (syn, of Holost. variabile), 383 (syn. of D. echinatum).—Rud., 1809a, 340, 418.

terrestris (Mueller, 1774) —, 1776.—Bosc, 1802a, v. 1, 261.—Carrière, 1879a, 29 Dec., 668.—Herbst, 1787a, 35.—Jenyns, 1869a, 25.—Lubbock, 1868a, 193–195 (in England).

tetragona (Mueller, 1774) —, 1776.—Bosc, 1802a, v. 1, 261.—Herbst, 1787a, 35.—Mueller, —, v. 3, 42 (=Fasc. tetragona) (free form).

torva (Mueller, 1774) Herbst, 1787a, 34.—Baer, 1826a, 126.—Ben., 1858a, 1861a, 198.—Bosc, 1802a, v. 1, 259, pl. 8, fig. 9.—Flexner, 1898, 337–346, 1 pl. (regeneration of nervous syst.).—Mont., 1888a, 44.

tremellaris (Mueller, 1774) ——, 1776.—Herbst, 1787a, 35.

trimellaris Bosc, 1802a, v. 1, 262.

truncata Bosc, 1802a, v. 1, 262.

truncata Leidy, 1851 (free form; Newark, Del.); 1904a, 54.

uncinulata Braun, 1790a, 58-61, pl. 3, figs. 1-3 (in Rana esculenta; Germany).—
 Dies., 1850a, 412 (syn. of Polyst. integerrimum).—Gmelin, 1790a, 3056 (to Fasc.).—Rud., 1809a, 451.—Stoss., 1898, 10.

unionicola Graff, 1904, 457 (in Muscheln).

relellæ Graff, 1904, 456.

ventricosa Bosc, 1802a, v. 1, 257.

vespertilionis (Mueller, 1784).—Dies., 1850a, 387 (of Gœze, 1782a, 171, pl. 14, f. 1-2, syn. of Distomum lima).—Kolenati, 1857, 12.—Rud., 1809a, 427.

viridata Bosc, 1802a, v. 1, 258.

viridis Bosc, 1802a, v. 1, 256.—Herbst, 1787a, 34.

PLANARIADÆ Mell, 1903, 191-236, 3 pls., 4 figs. (of madagassischen Subregion).

PLANARIÆ Fabricius, 1799, 151.—Blainv., 1828a, 577 (fam.).—Darwin, 1844a, 14 Oct., 241–251, pl. 5, figs. 1–4.—Faraday, 1832a. 11 Feb., 723–724; 1833a, Jan., 183–185; 1834a, 994.—de Férussac, 1821a, 90–92. pl. 116 (sp. in Brazil).— Girard, 1850a, 398–402 (embryology); 1851a, 258–273; 1851b, Jan., 41–53.— Houghton, 1870a, Sept., 255–257, figs. 1–2 (from Borneo).

PLANARIIDÆ Pease, 1860, 37-38 (of Sandwich Islands).

PLATYASPIS Mont., 1892, Oct. 7, 205 (m. lenoiri).—Braun, 1893a, 888, 890, 891, 894, 896, 897, 917; 1893b, 188.—Gamb., 1896a, 73.—Kofoid, 1899a, 181, 182, 183, 184.—Looss, 1902m, 428, 429.—Nickerson, 1902, 613, 615.—Pratt, 1902, 887, 891.—Stoss., 1899, 4.

anodontæ Osborn, 1898, 416 (in Unio luteolus, Anodonta; Lake Chautauqua, N. Y.).—Kofoid, 1899a, 179–186 (identity with Cotylaspis insignis); 1899b.

lenoiri (Poir., 1886) Mont., 1892, Oct. 7, 205–206 (in Tetrathyra vaillanti; Senegal).—Braun, 1893a, 897.—Looss, 1902m, 418, 428, 429, 783.—Nickerson. 1902, 613.

PLATYCOTYLE Ben. & Hesse, 1863; 1864, 96, 108 (m. gurnardi).—Braun, 1890a, 413, 498, 516, 517, 523, 534, 537, 546; 1893a, 890.—Cerf., 1895h, 918; 1896. 514.—Gamb., 1896a, 73.—Hoyle, 1890, 539.—Mont., 1888a, 8, 11, 86, 89. 100.—Pratt, 1900, 646, 651, 656, fig. 27.—Tasch., 1879, 69; 1879, 239, 248.

guanardi Tasch., 1879, 248, for gurnardi.

gurnardi Ben. & Hesse, 1863, 108; 1864, 108-109, pl. 11, figs. 14-15 (in Trigla gurnardus).—Braun, 1890a, 418; 469, 537, 548, 552.—Pratt, 1900, 656, 657, fig. 27.—Tasch., 1879, 248 (guanardi, in Trigla gu[r]nardus).

PLECTANOCOTYLE Dies., 1850a, 289, 420-421, 425 (m. elliptica).—Braun, 1890a, 413, 415, 516, 517, 523, 534, 537, 546; 1893a, 890.—Gamb., 1896a, 73.—Goldb., 1855, 19.—Hoyle, 1890, 539 (on gills of Labrax nucronatus).—Mont., 1888a, 11, 89, 100; 1892, Oct. 7, 213 (Octocotylinæ); 1899c, 1045-1053, figs. 1-12; 1903, 336 (subf. Plectanocotylinæ); 1905, 76-78.—Pratt, 1900, 646, 651, 656, fig. 30; 661 (on gills of marine fishes).—Tasch., 1879, 69 (of Ben. & Hesse); 1879, 239, 250 (syn. Plectanophorus).

1858: Plectanophorus Dies., 1858, 315, 382 (m. ellipticus).

PLECTANOCOTYLE—Continued.

elliptica Dies., 1850a, 421 (t. h. Labrax mucronatus; loc. not given); 18—, 69, pl. 1, figs. 4–9; 1858e, 382 (to Plectanophorus).—Braun, 1890a, 408, 537, 548, 551.—Mont., 1905, 76.—Pratt, 1900, 656, fig. 30; 657, 661.—Tasch., 1879, 250 (in Labrax mucronatus).

lorenzii Mont., 1899, 1, 1 pl.; 1899c, 1045–1053, figs. 1–12 (on Trigla sp.; Rovigno); 1905, 76, 78.—Scott, 1905, 116 (syn. Phyllocotyle gurnardi) (in Trigla sp.).

nordmanni (Dies., 1850) Braun, 1890a, 418.—Type of Encotyllabe.

PLECTANOCOTYLINÆ Mont., 1903, 336 (f. Hexacotylidæ).

PLECTANOPHORA Dies., 1858.—Ben. & Hesse, 1864, 120.—Mont., 1888a, 84.

PLECTANOPHORUS Dies., 1858e, 315, 382 (m. ellipticus; Plectanocotyle renamed); 1859c, 443.—Tasch., 1879, 250 (syn. of Plectanocotyle Dies.).

ellipticus (Dies., 1850) Dies., 1858e, 382 (in Labrax mucronatus), type of Plectanocotyle 1850.—Tasch., 1879, 250 (to Plectanocotyle).

PLECTOBOTHRII Nord., 1840, 596.

PLEORCHIS Rail., 1896, Mar. 15, 160 (for Polyorchis Stoss., 1892 [not Agassiz, 1862, cœlenterate]; type polyorchis).—Pratt, 1902, 887 (related to Fasciolinæ), 893.—Stiles & Hass., 1898a, 92, 97 (syns.: Dist. (Polyorchis) Stoss., Polyorchis Stoss.) (type polyorchis).—Stoss., 1898, 30.

cygnoides (Zed., 1800) Stoss., 1898, 31 (in Bombinator igneus; Triest).

mollis (Leidy, 1856) Stiles, 1896, 213.

polyorchis (Stoss., 1889) Stiles, 1896, 205.—Stoss., 1898, 30-31 (in Corvina nigra;

urocotyle Par., 1899, 6, 1 fig. (in Scorpæna scrofa; Portoferrajo); 1902, 5 (in Scorpæna porcus; Portoferrajo; S. scrofa; Elba).—Braun, 1902b, 23.—Odhn., 1905, 364.

PLESIOCHORUS Looss, 1901b, 205 (m. cymbiformis); ὁ πλησιόχωρος, Grenznachbar; 1901, 557, 558; 1902m, 478, 479, 480, 481, 482, 483, 484, 485, 526, 814, 844, 857 (Anaporrhutinæ), 858, 859, 860, 861, 863, fig. iv.—Odhn., 1901, 65, 67; 1902, 65, 67.—Pratt, 1902, 888, 901.

cymbiformis (Rud., 1819) Looss, 1901b, [205], 207, 209; 1901, 557, 558; 1902m, 414, 415, 417, 469–476 (includes Dist. cymbiformis Rud., 1819, 371; Sons., 1893, 2; Stoss., 1895, 38, pl. 4, fig. 1; Braun, 1899, 729; Looss, 1899b, 605; Phyllodist. cymbiforme (Rud.) Braun, 1901, 1, pl. 1, fig. 1), 479, 480, 682, 788, 791 (amphitypie very common), 813, 814, 844, 857, 863, 870, pl. 23, figs. 30–36.—Odhn., 1902, 68.—Reported for Chelone mydas, Thalassochelys corticata.

PLEUROCOTYLE a Gerv. & Ben., 1859b, 194 (based upon Grube, 1855a, 137) (m. scombri).—Ben. & Hesse, 1864, 96.—Brand., 1898a, [17] 209.—Braun, 1890a, 413, 414, 462, 477, 516, 517, 523, 534, 535, 546; 1893a, 890.—Cerf., 1895h, 918; 1896, 514.—Dies., 1859c, 444 (syn. of Grubea Dies.).—Gamb., 1896, 73.— Hoyle, 1890, 539.—Mont., 1888a, 8, 11, 48, 84, 89, 99; 1892, Oct. 7, 213 (subg. in Octocotylinæ); 1903, 336 (Pleurocotylinæ).—Pratt, 1900, 646, 651, 656, fig. 28.—Tasch., 1879, 69; 1879, 239, 248 (syns.: Octobothrium? Grube, Tetracotyle Grube, Grubea Dies.).—Ziegler, 1883, 552.—See Grubea.

cotyle Grube, Grubes Dies.).—Zieglet, 1835, 352.—See Grubea.

scombri (of Grube, 1855) [Gerv. & Ben., 1859b, 194, does not use combination, based on Troschel's Arch., 1855, 137].—Ben. & Hesse, 1863, 100; 1864, 100 (syns.: Octobothrium scombri Grube, Grubea scombri) (in Maquereau).—Braun, 1890a, 407, 414, 418, 462, 518, 535, 548, 552.—Knoch, 1894a, 11, 12.—Lang, 1880.—Mont., 1888a, 48, 49.—Par., 1894, 549, 595, 671, 680; 1902, 3 (in Scomber colias, S. scomber, S. sp.; Portoferrajo).—Par. & Perugia, 1890, 7.—Pratt, 1900, 656, 657, fig. 28.—Scott, 1901, 147.—Tasch., 1878, 575–577; 1879, 35; 1879, 248–249 (syns.: Octobothrium sc. Nord., Tetracotyle sc. Grube, Grubea cochlear Dies.) (in Scomber scombrus, S. colias: Naples). cochlear Dies.) (in Scomber scombrus, S. colias; Naples).

PLEUROCOTYLIDÆ Mont., 1903, 336 (subf. Pleurocotylinæ, g. Pleurocotyle).

PLEUROCOTYLINÆ Mont., 1903, 336 (f. Pleurocotylidæ).

PLEUROGENES Looss, 1896b, 97 (tod. claviger) $(\pi \lambda \varepsilon \tilde{v} \rho \alpha = \text{side}; \gamma \varepsilon \nu \dot{\alpha} \omega = \text{procreate});$ 1899b, 614, 615, 616-617, 622, 623; 1901, 194; 1902m, 820, 838.—Luehe, 1899, 536; 1900, 509.—Pratt, 1902, 889, 902.—Staff., 1905, Apr. 11, 683, 684.—Stiles, 1901, [183] (Pleuronectes, lapsus calami), 185.—Stoss., 1899, 7, 10.

^a Gervais & Ben. use both Pleurocotyle and Pleurocotylus, but they evidently intended the former as a French vernacular name, the latter as the Latin scientific name. Later authors apparently overlooked this fact.

PLEUROGENES-Continued.

arcanum (Nickerson, 1900) Pratt, 1902, 959.—Klein, 1905, 13 (arcana); 1905, 71 (arcanus).—Staff., 1905, 683 (type of Loxogenes).

betencourti (Mont., 1893) Stoss., 1899, 10 (in Scyllium canicula, S. stellare; Boulogne).—Looss, 1899b, 622.

brusinæ (Stoss., 1889, 25) Stoss., 1899, 10 (in Oblata melanura; Triest).—Looss, 1899, 622.—Type of Diphtherostomum, 1904.

claviger (Rud., 1819) Looss, 1899b, 569, 617; 1902m, 787, 820.—Klein, 1905, 11, 13, 17; 1905, 69.—Kowal., 1902d, [9] 27 (syns.: Dist. cl. Rud., D. neglectum Linst.); 1904, [7] 22.—Luehe, 1901, 171; 1901, 57.—Odhn., 1900, 17.—Ssinitzin, 1905, 146.—Staff., 1905, Apr. 11, 684 (type).

confusus (Looss, 1894) Klein, 1905, 68; 1905, 10, 11, 12, 13, 17.

gastroporus Luehe, 1901p, 166–171 (in Rana cyanophlyctis Schneider; India).— Klein, 1905, 10, 11, 13, 14; 1905, 68, 69.—Staff., 1905, Apr. 11, 684.

glaviger Ssinitzin, 1906, 687 (for claviger).

medians (Olss., 1876) Klein, 1905, 10, 11, 12, 13, 14, 17, 18; 1905, 68.—Looss, 1899b, 617; 1902m, 820, 824.—Luehe, 1901, 57; 1901, 171.—Odhn., 1900, 17.—Ssinitzin, 1905, 145–146; 1906, 687 (in Agrion and beetle larvæ; Warschau). Staff., 1905, Apr. 11, 683 (syn. of Loxogenes arcanum), 684, 685.—Stoss., 1900, 7–8, fig. 12 (in Rana esculenta; Triest); 1900, 17.

sphæricus Klein, 1905, 68-72, pl. 5, figs. 4-5 (in Rana hexadactyla; Kœnigsberg, from India); 1905, 10-14, pl. 1, fig. 5.

tacapense ("Sons., 1894" of Looss, 1896) Stoss., 1899, 10 (= Prosotocus tener [tacapense was a misdetermination]) (in camaleonte; Tunisia; Alessandria).

tacapensis (Sons., 1894) Looss, 1899b, 622.

tener (Looss, 1898) Looss, 1898, 461; 1899b, 622.—Klein, 1905, 10, 11, 12, 13; 1905, 68.

PLEUROGENETINÆ Looss, 1899b, 615, 623; 1902m, 839.—Luehe, 1900, 561; 1901, 171.—Odhn., 1902, 38, 42; 1902, 40.—Pratt, 1902, 889, 902 (includes: Prosotocus, Pleurogenes, Gymnophallus, Lepidophyllum).—Stoss., 1904, 198.—Ward, 1901, 185.

PLEUROGONINÆ Braun, 1900a, 1664, 1674.

PLEUROGONIUS Looss, 19011, 567, 568, 569 (tod. longiusculus); 1902m, 417, 557, 564, 569, 570 (type longiusculus), 572, 579, 583, 588, 589, 591, 594, 595, 596, 597, 599, 603, 609, 610, 612–616.—Pratt, 1902, 890, 910.

bilobus Looss, 1901l, 569 (in Chelone mydas; Egypt); 1902m, 567–568, pl. 27, fig. 100; 569, 878, pl. 27, fig. 100.

linearis Looss, 19011, 618 (in Chelone mydas; Egypt); 1902m, 565–567, 569, 878, pl. 27, fig. 99.

longiusculns Looss, 1902m, 582, misprint.

longiusculus Looss, 1901l, 568-569, 618 (in Chelone mydas; Egypt) (582 longiusculus); 1902m. 527, 558-561 (syn. Monost. trigonocephalum R. of Ben., 1859, 81, pl. 2, fig. 5; Walter, 1893, 191), pl. 27, figs. 94-98, pl. 32, figs. 176, 177; 562, 563, 564, 565, 566, 569, 570 (type of Pleurogonius), 588, 589, 591, 592, 596, 617, 788.

minutissimus Looss, 1901l, 7 Nov., 618-619 (in Chelone mydas; Egypt); 1902m, 568-569, pl. 27, fig. 101; 879.

trigonocephalus (Rud., 1809) Looss, 1901l, 567–568, 620 (in Thalassochelys corticata); 1902m, 416, 417, 548–558 (syns.: Monost. trigonocephalum Rud., 1809a, 336; 1819a, 349; Braun, 1901a, 38, pl. 2, fig. 29), pl. 26, figs. 75–78; 569, 570, 575, 588, 813, 876.—Cohn, 1904, 237.

PLEURONECTES Stiles, 1901, 183, lapsus calami for Pleurogenes.

PNEUMONŒCES Looss, 1902m, 732, 780, 839 (Hæmatolæchus Looss renamed) (type variegatus).—Klein, 1905, 64.—Staff., 1905, Apr. 11, 687 (syn. Hæmatolæchus).

1899: Hæmatolechus Looss, 1899b, 600 (tod. variegatus) [not Hæmatolecha Stål, 1874, hemipteron].

asper (Looss, 1899) Klein, 1905, 64; 1905, 6.

breviplexus (Staff., 1902) Staff., 1905, 687 (in lungs of American toads and frogs).— Klein, 1905, 3, 5, 6; 1905, 61. PNEUMONŒCES—Continued.

capyristes Klein, 1905, 60-65, pl. 5, figs. 1-2 (in Rana hexadactyla; from India); 1905, 2-7, pl. 1, figs. 1, 2.

complexus Seely, 1906, 249-252, figs. 1-2 (in Rana pipiens; North Carolina).

longiplexus (Staff., 1902) Staff., 1905, 681; 1905, Apr. 11, 687 (American frogs and toads).—Klein, 1905, 6; 1905, 64,—Seely, 1906, 249.

medioplexus (Staff., 1902) Staff., 1905, Apr. 11, 687 (syn. Ostiolum formosum) in American frogs and toads).—Klein, 1905, 3, 6; 1905, 61.—Seely, 1906, 252.

similiplexus (Staff., 1902) Staff., 1905, Apr. 11, 687 (in American frogs and toads).— Klein, 1905, 6; 1905, 64.—Seely, 1906, 252.

similis (Looss, 1899) Looss, 1902m, 762.—Klein, 1905, 2, 3, 4, 5, 6; 1905, 61.

variegatus (Rud., 1819) Looss, 1902m, 806.—Engler, 1904, 186.—Klein, 1905, 1, 2, 3, 4, 5, 6; 1905, 59.

varioplexus (Staff., 1902) Staff., 1905, Apr. 11, 687 (in American frogs and toads).— Klein, 1905, 3, 6; 1905, 61.

PODOCOTYLE (Duj., 1845) (tld. angulatum = atomon), see also (Podocotyle).—
R. Bl., 1891, 609.—Braun, 1893a, 885, 886, 890, 909, 910; 1900h, 3.—Fil.,
1857c, 9.—Jægers., 1901, 982.—Looss, 1899b, 528, 535, 538, 542, 647; 1902m,
721, 757, 764, 770, 771, 772, 827 (type angulatum).—Luehe, 1900u, 487-492;
1900v, 792; 1900, 562; 1901g, 42.—Mont., 1888a, 92, 105; 1892, Oct. 7, 214
(gen. of Distominæ); 1893, 150, 151, 152, 153, 155, 167, 168.—Odhn., 1905,
326 (type atomon).—Pratt, 1902, 888 (related to Opisthorchiinæ), 896.—
Sons., 1890, 140.—Stiles, 1901, 193, 196.—Stiles & Hass., 1898a, 92-93, 97
(syns.: ? Schisturus Rud., Dist. (Podocotyle) Duj.), type angulatum.—
Stoss., 1892, 4; 1898, 24; 1902, 582.

angulata (Duj., 1845) Jægers., 1901, 982, see atomon.

atomon (Rud., 1802) Odhn., 1905, 320–326, pl. 2. figs. 9–10 (syns.: Fasc. at., Dist. at., D. angulatum, D. simplex Rud. of Olss., Allocreadium at.), type of genus.—Nicoll. 1907, 68, 69, 70, 71, 73–77, pl. 1, figs. 1–2 (syns.: Psilost. redactum. Dist. simplex) (in Centronotus grumellus, Cottus bubalis, C. scorpius, Gasterosteus aculeatus, Gobius ruthensparri, Liparis montagui, Motella mustella, Zoarces viviparus).

contortum (Rud., 1819) Stoss., 1898, 24–25 (in Orthogoriscus mola; Triest) [type of Accacœlium].—Barbagallo & Drago, 1903, 409 (in Orthag. mola; Catania).

fractum (Rud., 1819) Stoss., 1898, 26 (in Box salpa; Triest).—Barbagallo & Drago, 1903, 409 (in B. s.; Catania).—Osborn, 1903, 316.—Par., 1902, 4 (in B. s.; Portoferrajo).

furcatum (Bremser, 1819) Stoss, 1898, 26 (in Mullus barbatus, M. surmuletus, Solea vulgaris; Triest); 1899, 5.—Barbagallo & Drago, 1903, 409 (in Mullus surmuletus, Solea vulgaris; Catania).—Fischder., 1903h, 548.—Jægers., 1901, 982.—Looss, 1902m, 771.—Luehe, 1900, 490, 491 (furcata); 1901, 475.

macrocotyle (Dies., 1858) Stoss., 1898, 25 (in Orthagoriscus mola; Triest).—Barbagallo & Drago, 1903, 409 (in Or. mola; Catania).

olssoniOdhn., 1905, 326, 327, new name for Dist. simplex of Olss. (in Gadus melanostomus, Lumprenus maculatus; West Coast of Sweden).

pachysomum (Eysenhardt, 1829) Stoss., 1898, 27 (in Mugil; Triest).—Barbagallo & Drago, 1903, 409 (in Mugil cephalus; Catania).

pedicellatum (Stoss., 1887) Stoss., 1898, 25 (in Chrysophrys aurata; Triest).—Barbagallo & Drago, 1903, 409 (in Chrys. aur.; Catania).

planci Stoss., 1899, 5, fig. 9 (in Ranzania truncata; Triest).—Looss 1902m, 637.

reflexa (Crep., 1825) Odhn., 1905, 324, 326–327 (in Cyclopterus lumpus: West Coast of Sweden).

retroflexum (Mol., 1861) Stoss., 1898, 26–27 (in Belone acus; Triest).—Barbagallo & Drago, 1903, 409 (in Bel. ac., Exocœtus volitans; Catania).

species of Mueller, 1897, 23-24, 26, pl. 3, fig. 8 (in Numenius arquatus).

(PODOCOTYLE) Duj., 1845a, 388, 401 (tld. angulatum).

angulatum Duj., 1845a, 386, 401–402, designated type of subg. by Stiles & Hass., 1898a, 93, (t. h. Anguilla vulgaris in the Morbidan).

fractum (Rud., 1819) Stoss., 1886, 49 (to Podocotyle by Stoss., 1898, 26).

furcatum (Bremser, 1819) Duj., 1845a, 402 (to Podocotyle by Stoss. 1898, 26).

(PODOCOTYLE)—Continued.

gibbosum (Rud., 1802) Duj. 1845a, 402.

macrocotyle (Dies., 1858) Stoss., 1898, 25.

pachysomum (Eysenhardt, 1829) Stoss., 1886, 57 (to Podocotyle by Stoss., 1898, 27). perlatum (Nord., 1832) Duj., 1845a, 387, 401 (to D. (Echinost.) by Stoss., 1886, 62) (type of Asymphylodora Looss, 1899b, 598).

retroflexum (Mol., 1861) Stoss., 1886, 60 (to Podocotyle by Stoss., 1898, 26). unicum (Mol., 1859) Stoss., 1886, 58.

POLISTOMEÆ Mont., 1888a, 13, 34, 42 (see Polystomeæ).

POLISTOMUM Sons., 1889, 283, for Polystomum.

POLYANGIUM Looss, 1902m, 584, 633, 634, 639, 642, 648, 652, 658, 667, 668, 675, 681, 684, 687–688 (tod. linguatula), 689, 698, 699.

linguatula (Looss, 1899) Looss, 1902m, 632, 688, 689, 697, 698, 885, pl. 30, fig. 145-150.

POLYCOTYLA Blainy., 1828a, 569-570 (family name).—Johnston, 1865a, 31.— Nord., 1840, 596.

POLYCOTYLE Will.-Suhm, 1870, 9 (m. ornata); 1871, 183.—Brand., 1888a, 33, 34, 35, 37, 58, 61; 1890a, 562, 563, 564, 567, 584.—Braun, 1892a, 581, 698, 713, 715, 717, 718, 723, 732; 1893a, 879, 881, 886, 887, 890, 892, 895, 899, 900, 901, 917; 1895b, 121, 136.—Gamb., 1896, 73.—Mont., 1888a, 10, 12, 15, 54, 90, 91, 104; 1892, Oct. 7, 196, 197, 214 (Diplostominæ).—Pratt, 1902, 890.—Schneidemuehl, 1896, 295 (Polykotyle).—Tasch., 1879, 256.—Wolf, 1903, 616, 617.

1896: Polykotyle Schneidemuehl, 1896, 295, for Polycotyle.

ornata Will.-Suhm, 1870, 9-11 (in Alligator lucius; Charleston); 1871, 183-185, pl. 11, fig. 1.—Brand., 1888a, 15, 58 (in All. luc.); 1890a, 584–585.—Braun, 1892a, 581, 733; 1893a, 901.—Pavesi, 1881, 294.—Poir., 1886, 339–345, pl. 18, fig. 7, pl. 19, fig. 1, 3, 5, 6, pl. 20, fig. 5 (in All. luc.).—Reported also for Alligator mississippiensis.

POLYCOTYLEA Dies., 1850a, 288 (tribe), 289, 408 (tribe III of Trematoda, subo. II), 416 (tribe I of Bdellidea, subo. III); 1858e, 314 (fam. of Trematoda cotylophora), 367.—Brand., 1888a, 15.—Braun, 1890a, 515.—Cerf., 1899a, 351.—Goldb., 1855a, 17, 18.—Mont., 1888a, 83, 84; 1903, 336 (section, contains: Polystomidæ Ben., 1858; Octocotylidæ Ben. & Hesse, 1863).—Tasch., 1879, 233, 234.

POLYCOTYLE.E Dies., 1839a, 234.—Cobbold, 1877, 238.

POLYCOTYLIDÆ Cobbold, 1877f, 326.—Braun, 1893a, 886.—Mont., 1888a, 104.

POLYCOTYLINE Mont., 1892, Oct. 7, 214 (subf. of Holostomidæ).—Braun, 1893a, 890, 892.

POLYKOTYLE Schneidemuehl, 1896, 295, for Polycotyle.

POLYORCHIS (Stoss., 1888) [nec Polyorchis Agassiz], renamed Pleorchis 1896, (type by absolute tautonymy polyorchis); 1892, 4; 1898, 30.—Braun, 1893a, 885, 886, 894, 909, 911; 1895b, 138; 1901b, 54.—Looss, 1894a, 174; 1896b, 59; 1902m, 846.—Mont., 1893, 148, 149, 150, 151, 154.—Ofenheim, 1900, 160.—Rail., 1896, 160 (=Pleorchis).—Sons., 1890, 134, 135.—Stiles & Hass., 1898a, 92 (syn. of Pleorchis).

formosum (Sons., 1890) Stoss., 1892, 143 (in Grus cinerea; Pisa).

molle (Leidy, 1856) Mont., 1896, 166.

polyorchis (Stoss., 1889) Mont., 1896, 166.

(POLYORCHIS) Stoss., (1888), subg. of Dist., [type polyorchis].

cygnoides (Zed., 1800) Stoss, 1889, 60.

molle (Leidy, 1856) Stiles & Hass., 1894h, 162-163, fig. 2; 1895a, 737-741, pl. 3, figs. 1–3.

ragazzii Setti, 1897, 8 (type of Synccelium Looss, 1899).

POLYSARCUS Looss, 1899b, 561 (tod. westermanni) [nec Polysarcus Fieb., 1853, orthopteron; Sauss, 1859, orthopteron; Lefèvre, 1876, coleopteron; Polysarca Schin, 1866, dipteron]; 1900, 605.—Braun, 1900h, 6; 1901e, 329 (= Paragonimus).—Luehe, 1900, 557.—Stiles, 1901, 189.—See Paragonimus.

westermanni (Leuck., 1889) Looss, 1899b, 561.

POLYSSICHIA Cosmovici, 1887 (contains Aspidogaster, Diplozoon, Diporpa, Gyrodactylus, Octobothrium, Polyst.).-Mont., 1888a, 84.

POLYSTOMA Zed., 1800a, xviii, 199-203 (nec Polyst. Steph. Col., 1835). includes Linguatula serrata Freelich, Hexathyridium pinguicola Treutler, 1793) (type was clearly intended to be Planaria uncinulata = Polyst. ranæ; name Polyst. was in MS. in 1788 based on this species, see Zed., 1803a, 223).—Andral, 1829d, 617.—Audouin, 1828b, 193.—Baer, 1826a, 126.—Ben., 1858a, 1861a, 11 (Polystomum —Ben. & Hesse, 1864, 61.—E. Bl., 1847, 331.—R. Bl., 1888a, 312 (Polystomes).—Brand.. 1891d. 9.—Braun. 1883a. 50; 1889k. 620, 621; 1890a. 413, 416, 424, 433, 435, 451, 454, 470, 479, 480, 481, 483, 487, 490, 491, 1890a. 413, 416. 424. 433, 435, 451, 454, 470, 479, 480, 481, 483, 487, 490, 491, 492, 493, 496, 498, 504, 511, 514, 515, 516, 517, 522, 523, 538, 546; 1890b, 125, 127, 128; 1890e, 597; 1891d, 421; 1892a, 753; 1893a, 890, 891.—Brem., 1824, 135.—Burm., 1837a, 530; 1856a, 251.—Carus, 1863, 478.—Ceri., 1899a, 348, 350, 351, 359, 371.—delle Chiaje, 1833, 13; 1835b, 76-90; 1837a, 5-19 Polistoma in blood of man; 1837b, 245-246; 1843b, 51.—Cobbold, 1866, 7; 1877, 238; 1879b, 41.—Cosmovici, 1887a, 129.—Crep., 1839, 290.—Cuvier, 1817, 42.—Deslongchamps, 1824mm, 648-650; 1828d, 192-193.—Dies., 1839, 234; 1850a, 288, 409 (syn. of Hexathyridium , 412-413 includes; Plan, Braun, Linguatula Freelich, Fasc, Gmelin, Hexathyridium Blainy, 1, 416 (of Delaroche, syn. of Plagiopeltis), 419 (of Kuhn, syn. of Onchocotyle), 420 (of delle Chiaje, syn. of Plagiopeltis . 419 of Kuhn. syn. of Onchocotyle). 420 of delle Chiaje, syn. of Solenocotyle). 421 (of Duj., syn. of Diclibothrium), 609 (of Rud., pars syn. of Pentast.): 1858e, 314, 371-372 (only species integerrimum). pars syn. of Pentast. 1: 1858e, 314, 371–372 only species integerrimum).—Duj., 1845a, 318.—Eichwald, 1829a, 249.—Eiss. 1838, 21.—Fischer, 1840, 59.—Gamb.. 1896a, 57, 59, 73.—Goldb., 1855, 17.—Goldschmidt, 1902e, 874.—Goto, 1891a, 174, 183, 185, 187.—Hahn, 1900a, 174–175.—Haswell, 1892a, 458; 1892b, 149; 1893e, 112, 144.—Hémont, 1827, 9.—L'Herminier, 1826, 10.—Hoyle, 1890, 535–537, 539 (two species; best known P. integerrimum; Hexathyridium is probably a synonym.—Jackson, 1888, 644, 646, 647, 654.—Joy, 1835a, 504.—Juel, 1889, 33.—Kath., 1894a, 130, 148.—Lamarck, 1816b, 175–176.—Leuck., 1863a, 450, 527.—Looss, 1892a, 72, 73, 95; 1893b, 810.—Mayer, 1841a, 20.—Mlinarich, 1832, 14.—Mont., 1888a, 8, 9, 11, 13, 14, 29, 37, 38, 40, 42, 44, 53 [Polystomum, 54, 56, 57, 58, 59, 60, 63, 64, 66, 67, 70, 83, 84, 86, 89, 100; 1892, 109; 1892. Oct. 7, 186, 213 (gen. of Polystominæ); 1893, 110; 1903, 336 [subf. Polystominæ].—Moul., 1856a, 10, 11.—Nord., 1832a, 60, 69; 1840, 544, 592, 593, 596–597.—Olfers, 1816, 22, 41.—Pag., 1857, 52.—Pintner 1903. 336 | subf. Polystominæ | — Moul., 1856a, 10, 11. — Nord., 1832a, 60, 69; 1840. 544. 592. 593, 596-597. — Olfers, 1816, 22, 41. — Pag., 1857, 52. — Pintner, 1891, 727. — Pratt. 1900. 645. 646. 647, 650. 656. 659. fig. 23. — Rafinesque, 1815. 151 | syn. of Hexathyridia | — P.ud., 1809a, 6, 22-23, 38, 441, pl. 6, figs. 1-6; 1819a, 125. 435-436, 584. — St.-Remy. 1898. 558. — Schneider, 1866. 334. — Slawikowski, 1819. 70. — Stoss., 1898a, 10, 11. — Tasch., 1879. 43, 50, 58. 62, 64, 69; 1879, 232, 233, 251-252 (syns.; Hexathyridium Blainy., Planaria Braun, Linguatula Frelich, Fasc. Gmelin). — Wagener, 1857, 50, 72. — Wallenstedt, 1847, 7. — Zeller, 1876, 23 June, 238-274. pls. 17-18.

-: Polystomum.

1837: Polistoma delle Chiaje, 1837a, 5-19, for Polystoma.

1888: Polystomnm Mont., 1888a, 53, misprint.

of Kuhn, 1829c; 1830a; 1830b, 185-187 (in Squalus catulus).—Tasch., 1879, 252 syn. of Onchocotyle Dies.

of Duj., 1845a, 319.—Tasch., 1879, 254 (syn. of Diplobothrium Leuck.).

Delaroche, 1811, 271-272 [n. g.?] type species thynni).—Audouin, 1828b, 193 (?syn. of Polyst. duplicatum).—Blainv.. 1824a. 514 ("M. de Lamarck a adopté ce genre de Delaroche; il est établi sur un animal de la famille des sangsues. suivant nous "), 515. - Rafinesque, 1815, 151 renamed Hexost.).-Tasch., 1879. 249 syn. of Hexacotyle Blainv. .—See Polyst. thynni, p. 353.

appendiculatum Kuhn, 1829c., 460–463, pl. 11. figs. 1–3 (t. h. Squalus catulus); 1829., 342.—Ben., 1858a, 1861a, 54, 55 (to Onchocotyle), 191, 192.—Bl., 1847, 336 of Nord. .—Cerf., 1899a. 349, 350. 351, 352–356, 359, 373, 375, 378 in Mustelus lævis. M. vulgaris. Scyllium catulus). 434.—Cobbold. 1872b. 90: 1879b, 41.—Crep., 1838, 84; 1839, 290.—Dies., 1850a, 419 (to Onchocotyle as type); 1858e. 371 to Onchocotyle .—Duj., 1845a, 321.—Goto, 1894a, 222 Japan).— Nord., 1832a, 80–82, pl. 5, figs, 6–7 in Squalus catulus); 1840, 601 (to Hexabothrium).—Scott. A., 1901, 344 (to Onchocotyle).—Tasch., 1879, 27, 28 (to Onchocotyle); 1879, 253.—Thaer, 1850, 602–630, pl. 20–22, figs. 1–45; 1851, 34 pp., pls. 1–3.

armatum Leuck.. 1835 Duj.. 1845a, 319 to Polyst. (Hexacotyle)).—E. Bl.. 1847, 335.—Dies., 1850a. 421 to Diclibothrium .—Tasch., 1879, 254 to Diplobothrium .—Type of Diclibothrium 1835.

borealis Ben., 1853) Wagener, 1857, 72.—Ceri., 1899a, 359, 377.—Dies., 1858e, 371 (to Onchocotyle).—St.-Remy, 1890, 41.

POLYSTOMA—Continued.

coronatum Leidy, 1888, 127 (in terrapin; eastern U. S. A.); 1904a, 213.—Braun, 1890a, 511, 538, 548, 552 (in Cistudo carolina).—Pratt, 1900, 659.

denticulatum (Rud., 1805) Rud., 1808a, 180; 1809a, 447–449, pl. 12, fig. 7 (to Polyst. (Pentast.)) (in Capra americana, C. hircus).—R. Bl., 1890a, 264.—Dies., 1850a, 615 (to Pentast.).—Olfers, 1816, 42.

duplicatum Rud., 1819a, 125, 438–439, pl. 2, fig. 6 (P. thynni Delaroche, 1811, renamed) (in Scomber thynnus; Balearic Isles).—Audouin, 1828, 193.—Crep., 1839, 290.—Deslongchamps, 1828d, 193.—Dies., 1850a, 417 (to Plagiopeltis as type) in Thynnus brachypterus.—Duj., 1845a, 318–319 to Polyst. (Hexacotyle).—Kroyer, 1838–40a, 251, 596 (in Thynnus vulgaris Cuv.).—Nord., 1832a, 62; 1840, 598 (syn. of P. thynni).—Tasch., 1879, 250 (syn. of Hexacotyle thynni).

hassalli Goto, 1895, 352 (in Kinosternon pennsylvanicum; Maryland); 1899, 276–278, pl. 21, figs. 16, 17.—Pratt, 1900, 656, fig. 23; 657, 659.—St.-Remy, 1898, 558.

integerrimum (Frœlich, 1791) Rud., 1808a, xxv, pl. 6, figs. 1–6; 1809a, 451–455, pl. 6, figs. 1–6, to P. (Hexastoma) (includes: Planaria uncinulata Braun, 1792; Linguatula integerrima, 1791; Fasc. uncinulata, 1790, Polystoma ranæ, 1800; Linguatula integerrimum); 1819a, 125 (in Rana temporaria; Gryphiæ, Jun.; R. esculenta, Bufo variabilis).—Baer, 1826a, 126; 1827a, 680-689, pl. 32, figs. 7-9.—Ben., 1858a, 1861a, 54, 169, 171, 172, 196; 1870c, 140.—Ben. & Hesse, 1864, 84–87, 1 fig. (integerrinum).—Bettend., 1897a, 4; 1897, 308.— E. Bl., 1847, 331–334, pl. 14, fig. 3.—Braun, 1883a, 41, 44, 48, 49, 51, 56, 71, E. Bl., 1847, 331–334, pl. 14, fig. 3.—Braun, 1883a, 41, 44, 48, 49, 51, 56, 71, fig. 10; 1889k, 621; 1890a, 407, 408, 410, 415, 419, 421, 424, 426, 428, 430, 433, 436, 439, 442, 445, 449, 453, 454, 455, 456, 458, 461, 464, 465, 468, 471, 472, 475, 480, 482, 483, 487, 490, 491, 492, 493, 494, 495, 497, 499, 500, 501, 503, 511, 512, 538, 548, 552; 1890d, 568; 1890e, 596; 1891a, 52, 53, 54; 1892a, 753; 1894a, 1160; 1895b, 125, 130, fig. 42; 1906, 131, fig. 66 (in frog).—Bremser, 1824e, pl. 10, fig. 26.—Civinini, 1842.—Cobbold, 1858, 162; 1879b, 452.—Crep., 1837, 322; 1839, 290.—Deslongchamps, 1828a, 193.—Dies., 1850a, 412-413 (syns.: Plan. uncinulata Braun, Linguatula integ., Fasc. uncinata Gmelin, Polyst. ranæ Zed., Hexathyridium integ. Blainv.); 1858e, 372 (only species) (in Rana platyrhinus).—Duj., 1845a, 320.—Eichwald, 1829a, 249.—Fraip., 1880a, 400; 1880c, 433; 1883a, 36.—Gamb., 1896a, 55, 58-59, figs. 24, 26.—Goldschmidt, 1902a, 397-444, pls. 22-24, figs. 1-40 (embryology); 1902b, July, 398-399; 1902c, 180-189, figs. 1-11 (development); 1902d, 736; 1902b, July, 398–399; 1902c, 180–189, figs. 1–11 (development); 1902d, 736; 1902e, 874; 1905, July, 64–65.—Goto, 1891a, 184.—Halpin, 1901a, 291–363, pls. 10–14 (embryology); 1902a, 395–398; 1904a, 444.—Hoyle, 1890, 537–539, fig. 3 B.—Ljima, 1884c, 635–639.—Jackson, 1888, 643, 644, 645, 646, 648–649, 650 (in Rana temporaria, R. esculanta, Bufo viridis).—Janicki, 1903a, 241 ff.— Juel, 1889, 14.—Kath., 1894a, 132, 141, 142, 146, 152.—Kerbert, 1881a, 535, 554, 556, 572.—Kholodk., 1899a, 148, fig. 175.—Knoch, 1862, 101.—Kowal., 1895, 372–390, pl. 8, figs. 11–13; 1896d (2), 252 (in Rana fusca; Du-Kowal., 1895, 372–390, pl. 8, figs. 11–13; 1896d (2), 252 (in Kana fusca; Dublany); 1904, (9), 24 (in Rana temporaria; Dublany).—Lejtenyi, 1881a, 3.—Leuck., 1879, 26, 52; 1886d, 19, 40.—Linst., 1903, 279.—Looss, 1885b, 17, 18, 19, 21, 24.—Macé, 1882a, 26, 29, 78.—Mayer, 1841a, 26, 34.—Mont., 1888a, 10, 18, 39, 44, 58, 59, 70; 1892, Oct. 7, 186; 1893, 110, 111.—Mueh., 1898, 11, 17–18.—Nord., 1832a, 62, 79; 1840, 546, 601.—Olfers, 1816, 42.—Pag., 1857, 47–49, pl. 6, figs. 10–13 (in braunen Frösche).—Par., 1894, 471.—Pégot, 1900, 162–164.—Pintner, 1891, 727, 728.—Polonio—.—Sieb., 1835, 70.—Sons., 1893, 187 (in Bufo viridis Laur.).—Stieda, 1870, 675; 1871, Jan., 660, 678, pl. 15, figs. 1–12, —Stoss, 1889, 29; 1898, 10–11, —Tasch, 1879, 33. 660–678, pl. 15, figs. 1–12.—Stoss., 1889, 29; 1898, 10–11.—Tasch., 1879, 33, 35, 43, 55, 56, 58, 60, 61; 1879, 252 (in Rana temporaria, R. esculenta, Bufo 55, 45, 50, 50, 58, 60, 61, 1613, 252 (iii Kana temporana, R. eschena, bulo viridis).—Walter, 1866, 65.—Will.-Suhm, 1871, 181–185; 1872, 22 March, 29–39, pl.—Zeller, 1872, v. 22 (1), 1–24, pls. 1–2; 1872, 25–28, pl. 3, figs. 1–12; 1872, Sept., 99–112; 1873, 51–55.—Woodland, 1904, 404–405, 1 fig.—Reported also for Rana fusca, Hyla viridis.

integerrinum Ben. & Hesse, 1864, 84–87, 1 fig. (see integerrinum).—Pégot, 1900, 162–164.

loliginis delle Chiaje, 1823, pl. 92, fig. 2 (in Loligo vulgaris; Naples).—Ben. & Hesse, 1864, 84.—Braun, 1890a, 518.—Dies., 1850a, 420 (syn. of Solenocotyle chiajei, type of Solen.).—Mont., 1888a, 89.—Tasch., 1879, 250.

midas Kuhl & van Hasselt, (1824).—Baird, 1853a, 41 (=P. ocellatum Rud.).—Nord., 1840, 600.

POLYSTOMA—Continued.

mydæ Kuhl & van Hasselt, (1822), 113: 1824a, 310.—Crep., 1846, 146.—Dies., 1850a, 413 (syn. of P. ocellatum Rud.).—Tasch., 1879, 252 (syn. of P. ocellatum).—Reported for Chelonia mydas.

oblongnum Mont., 1888a, 57 (for oblongum).

oblongum R. Wright, 1879, 12–15, pl. 1, figs. 9–11 (in Aromochelys (Sternothærus) odoratus; Toronto).—Braun, 1890a, 511, 538, 548, 552.—Goto, 1899, 277, pl. 21, fig. 18.—Leidy, 1888, 127.—Mont., 1888a, 57 (oblongnum).—Pratt, 1900, 659.—Staff., 1900, 405, 414; 1902, 483 (in Chelydra serpentina); 1905, Apr. 11, 690 (in Chrysemys picta; Chel. serp.; Canada).—Reported for Sternothærus odoratus.

1890a, 413 (syns. P. mydæ, Hexacotyle oc.).—Dul., 1840a, 319–320.—Gamb., 1896, 59.—Giebel, 1857, 266.—Goto, 1891a, 183.—Ijíma, 1884c, 636, 638.—Jackson, 1888, 649, 650.—Leidy, 1888, 127.—Looss, 1885b, 21, 22, 24, pl. 23, fig. 19.—Macé, 1882a, 26.—Mont., 1888a, 18, 58; 1892, Oct. 7, 186; 1893, 110, 111.—Nord., 1840, 600 (syn. of Hexacotyle oc.).—Par., 1894.—Sons., 1889, 283 (Polistomum); 1893, 183 (in Emys lutraria).—Stoss., 1890, 51.—(Tasch., 1879, 252 (syns. P. mydæ, Hexacotyle oc.) (in Emys europæa, Halichelys atra).—Reported also for Cistudo europæa.

pinguicola (Treutler, 1793) Zed., 1800a, 203 [type of Hexathyridium 1793].—Boele, guacota (Treutler, 1793) Zed., 1800a, 203 [type of Hexathyridium 1793].—Boele, 1828a, 33.—Bremser, 1819a, 233, pl. 4, figs. 15–17; 1824, 135, 272–280, pl. 9, fig. 2.—delle Chiaje, 1833, 13–14, 117–118, pl. 2, fig. 14.—Cobbold, 1866, 7; 1876, 211.—Crep., 1839, 290.—Deslongchamps, 1828d, 193.—Dies., 1850a, 410 (to Hexathyridium).—Duj., 1845a, 319.—Dunglison, 1893, 821, 900.—Eichwald, 1829a, 249.—Eiss, 1838, 21.—Hémont, 1827, 17.—L'Herminier, 1826, 14.—Joy, 1835a, 504, 519.—Kuech., 1855, 464.—Lundborg, 1817, 8.—Milnarich, 1832, 14.—Nord., 1840, 594 (to Linguatula), 601.—Olfers, 1816, 42.—Prunèyre, 1823, 7.—Puettner, 1837, 4.—Rud., 1809a, 455–456, to P. (Hexast.); 1819a, 125, 437–438.—Sjöbeck, 1830, 10.—Slawikowski, 1819, 23, 25, 70, 141. 25, 70, 141.

pinguiculum Joy, 1835a, 505, for pinguicola.

proboscideum Rud., 1814a, 106–107 (Echinorhynchus crotali Humboldt, 1808, renamed), to P. (Pentast.).—Dies., 1850a, 612 (to Pentast.).—Leidy, 1852, 97.—Nord., 1840, 645 (syn. of Porocephalus crotali, type of Poroc.).—Olfers, 1816, 42. Now an arachnoid.

ranz Zed., 1800a, xviii, 203, 204–205, pl. 4, figs. 1–3 (in Rana temporaria; bladder, Europe) (includes Plan. uncinulata Braun, 1790, from Rana esculenta; Europe); 1803a.—Dies., 1850a, 412 (syn. of P. integerrimum).—Rud., 1809a, 451, 452 (=P. integerrimum).—Stoss., 1898, 10.

sanguicola delle Chiaje, 1833, 14 (venarum, renamed), 118; 1834, ——; 1837a, 5-19.—Cobbold, 1876, 211.—Dav., 1877a, 334.—Dies., 1850a, 410 (syn. of Hexathyridium venarum).—Dolley, 1894a, 1006.—Dunglison, 1893, 821, 900.— Marcacci, 1843a.

sanguineum delle Chiaje, 1837b, 245 (based on 1837a) (in Homo).—Crep., 1839, 290.—Eiss, 1838, 21.—Leuck., 1863a, 585.—Moniez, 1896, 108.—Sieb., 1839,

164.

seymni ainosi Dies., 1858e, 371 (to Onchocotyle) based on Wagener, v. 13, 72.— Cerf., 1899a, 360.

serratum (Freelich, 1889) Zed., 1800a, 203 [type of Linguatula, 1789]; 1803a, 230.— Blainv., 1824a, 514 (type of tétragule).—Dies., 1850a, 616 (to Pentast.).—Nord., 1840, 594 (to Linguatula).—Olfers, 1816, 42.—Risso, 1826, 263.—Rud., 1809a, 449-451 to P. (Pentast.); 1819a, 124 (=Pentast. serratum).—Reported for Lepus timidus.

tænioidea Nord., 1840, 595, for tænioides.

tænioides Rud., 1809a, 441-447, pl. 12, figs. 8-12 (Tænia rhinaria Pilger, 1802, renamed), to P. (Pentast.) (in Equus cab., Canis fam.; France); 1810a, 256; 1814a, 107.—Anacker, 1890m, 506.—Blainv., 1824a, 513 (type of "prionoderme").—R. Bl., —,261.—Braun, 1903, 3. ed., 337 (syn. of Linguatula rhinaria).—Dies., 1850a, 610 (to Pentast.).—Nord., 1840, 595 (tænioidea, to Linguatula).—Olfers, 1816, 42. POLYSTOMA—Continued.

thynni Delaroche, 1811, 271–272, pl. 2, fig. 3 (in Scomber thynnus; Maroque).—Blainv., 1828a, 571 (type of Hexacotyla).—E. Bl., 1847, 335.—Crep., 1838, 84.—Dies., 1850a, 417 (syn. of Plagiopeltis duplicata, type of Pl.).—Lamarck, 1816b, 176.—Nord., 1832a, 62; 1840, 597–598 (syns.: Hexacotyle thynni, P. duplicatum), 600.—Risso, 1826, 263.—Tasch., 1879, 250 (syn. of Hexaco-

uncinatum Macé, (1880a).—Braun, 1890a, 511, 538, 548, 552.—Stoss., 1898, 10 (in

Rana temporaria; France).

uncinulatum Macé.—Braun, 1890a, 418, for uncinatum.

venarum (Treutler, 1793) Zed., 1803a, 231.—delle Chiaje, (1834a) 1833, 14–15, 118–119, pl. 2, fig. 15.—Cobbold, 1866, 7.—Crep., 1839, 290.—Dies., 1850a, 410.—Dunglison, 1893, 821, 900.—Hémont, 1827, 16.—Moniez, 1896, 108.—Olfers, 1816, 42.—Rud., 1809a, 456–457; 1819a, 136.—Slawikowski, 1819, 24, 70.—Reported for Homo.

POLYSTOMA (HEXACOTYLE) Duj., 1845a, 318-319.

armatum (Leuck., 1835) Duj., 1845a, 319.—Type of Diclibothrium, 1835.

duplicatum (Rud., 1819) Duj., 1845a, 318-319 (to Plagiopeltis as type by Dies., 1850a, 417).

POLYSTOMA (HEXAST.).

integerrimum (Freelich, 1791) Rud., 1809a, 451-455. pinguicola (Treutler, 1793) Rud., 1809a, 455-456.

POLYSTOMA (PENTASTOMA).

denticulatum (Rud., 1805) Rud., 1809a, 447-449, pl. 13, fig. 7 (to Pentast. by Rud., 1819a, 124, 423).

proboscideum Rud., 1814a, 106-107 (type of Porocephalus; Echinorhynchus crotali Humboldt, 1808, renamed) (to Pentast. by Rud., 1819a, 124).

serratum (Freelich, 1789) Rud., 1809a, 449-451 (to Pentast. by Rud., 1819a, 124).

tænioides Rud., 1809a, 441–447, pl. 11, figs. 8–12 (in Canis familiaris; France) (Tænia rhinaria Pilger, 1802, renamed) (to Pentast. by Rud., 1819a, 123).

POLYSTOMATA Zed., 1800a, 203, plural of Polystoma.

POLYSTOMATIDÆ Gamb., 1896a, 53, 55, 73.—Scott, 1901, 141, 145.

POLYSTOMATINÆ Gamb., 1896a, 73.

POLYSTOMEA Ben.—Braun, 1890a, 516.—Olss., 1893, 4.

POLYSTOMEA Ben.—Braun, 1890a, 516.—Olss., 1893, 4.

POLYSTOMEÆ Leuck.—Braun, 1883a, 40, 58; 1890a, 516, 517, 523, 524, 532; 1890b, 127.—Cerf., 1894, 948; 1895e, 523 (of Tasch.); 1895h, 914, 919; 1896, 500, 509, 515; 1898b, 361; 1899a, 365, 452 (of Tasch.).—Haswell, 1892a, 457.—Hoyle, 1890, 539 (includes Octobothriidæ, Polystomidæ, Microcotylidæ, Gyrodactylidæ).—Jackson, 1888, 642, 644, 646, 654 (includes same as Hoyle).—Maclaren, 1904, 598.—Mont., 1888a, 7, 10, 11, 13, 15, 16, 18, 20, 26, 27, 28, 31, 32, 34, 36, 37, 38, 41, 47, 48, 50, 53, 54, 56, 66, 67, 70, 84, 86, 87, 88, 90, 96, 98, 107, 108; 1888a, 16, 34, 42 (Polistomeæ); 1891, 108.—Par.,—Par. & Perugia, 1890, 6; 1890, 225–242; 1891, Mar. 7, 319.—Schneidemuehl, 1896, 296.—Sons., 1890, 175.—Tasch., 1879, 33, 56, 57, 68; 1879, 234, 235, 236–237, 238. 236-237, 238.

1888: Polistomeæ Mont., 1888a, 16, for Polystomeæ.

POLYSTOMES R. Bl., 1888a, 312, French word, see Polystoma.

POLYSTOMID.E Cunningham, 1884a, June 12, 399.—Hoyle, 1890, 539 (includes Polyst., Onchocotyle, Erpocotyle, Diplobothrium).—Jackson, 1888, 654 (includes Polyst., Onchocotyle).—Pratt, 1900, 646 (of Tasch.) (includes Polystominæ, Octocotylinæ, Microcotylinæ).—Scott, T., 1901, 141.

POLYSTOMIDÆ Braun, 1890a, 511, 516, 517, 523, 533, 538; 1893a, 890.—Carus, 1863, 477.—Cobbold, 1879b, 4.—Cunningham, 1884, 399; 1887a, 279.— Leuck., 1886d, 117.—Mont., 1888a, 8, 11, 13, 15, 16, 18, 20, 30, 34, 37, 52, 66, 70, 86, 89, 91, 100, 108; 1888a, 16, 88 (Polistomidæ), 89 (Polystomidæs); 1892, Oct. 7, 198, 213 (f. of Eterocotylea, containing subf. Polystominæ Ben., Octocotylinæ Ben. & Hesse, Microcotylinæ Tasch.); 1903, 336 of Ben. & Hesse, 1858 (subf. Polystominæ (g. Polystomum)).—Par. & Per., 1889, 742; 1890, 7.—Poir., 1886, 345.—St.-Remy, 1898, 522, 523, 544.—Schneidemuehl, 1896, 296.—Stoss., 1898, 10.—Tasch., 1879, 69; 1879, 235, 237, 251.

1888: Polistomidæ Mont., 1888a, 16, 88, for Polystomidæ.

1888: Polystomidaes Mont., 1888a, 89, typographical error.

8588-No. 37-08-23

POLYSTOMIDEA Mont., 1888a, 84.

POLYSTOMIDES Mont., 1888a, 86.—Tasch., 1879, 235.—R. Bl., 1888a, 541 (embraced in subo. Polystomiens).

POLYSTOMIENS [French] R. Bl., 1888a, 541 (suborder, includes: Tristomidés, Polystomidés, Gyrodactylidés).

POLYSTOMINÆ Pratt, 1900, 646, 650 (of Ben.) (includes Polyst., Erpocotyle, Onchocotyle, Diplobothrium, Sphyranura).

POLYSTOMINE Ben.—Braun, 1893a, 890.—Mont., 1892, Oct. 7, 213 (subf. of Polystomidæ); 1903, 336 (new subf. of Polystomidæ).—St.-Remy, 1898, 558.

POLYSTOMIS Crep., 1838, dative of Polyst.

POLYSTOMNM Mont., 1888a, 53, misprint for Polystomum.

PROBOLITREMA Looss, 1902m, 855, 857 (in subf. Anaporrhutinæ), 858, 859, 860, 863 (diagnosis), fig. 6 (tod. ricchiardii).

capense Looss, 1902m, 855, 857, 863, "Anaporrhutum ricchiardii Lopez" of Ofenh., renamed.

ricchiardii (Lopez, 1888) Looss, 1902m, 855, 857, 863 (ricchardii, misprint for ricchiardii).

PROGONUS Looss, 1899b, 643 (tod. mülleri) (ὁ πρόγονος=ancestor) [not Progona Berg, 1882, insect]; 1900, 602, 603; 1901, 208; 1902m, 732 (renamed Genarches).—Fuhrmann, 1904, 61.—Luehe, 1901, 481, 486.—Pratt, 1902, 889, 905.—Stiles, 1901, 189.

mülleri (Levin., 1881) Looss, 1899b, 643; 1902m, 732, type of Genarches, 1902.— Luehe, 1901, 481.

PRONOCEPHACUS Braun, 1901b, 50, lapsus for Pronocephalus.

PRONOCEPHALIDÆ Looss, 1902m, 527, 580-617, 843 (includes: Charaxicephalus Looss, type robustus; Pyelosomum Looss, type cochlear; Cricocephalus Looss, type albus; Epibathra Looss, type crassa; Adenogaster Looss, type serialis; Pronocephalus Looss, type obliquus; Glyphicephalus Looss, type solidus; Pleurogonius Looss, type longiusculus), 611-612 (key to genera), 611 diagnosis.

PRONOCEPHALINÆ Looss, 1899b, 665, 668; 1902m, 841.—Braun, 1901b, 50, 53.— Pratt, 1902, 890, 909 (includes: Pronocephalus, Pleurogonius, Glyphicephalus, Adenogaster, Cricocephalus, Pyelosomum; related genus: Charaxi-

cephalus).

PRONOCEPHALUS Looss, 1899b, 551, 666 (\acute{o} $\pi\rho\acute{o}\nu$ =Vorsprung) (tod. "trigonocephalus Rud." [=obliquus, 1901]); 1901, 209; 1901, 567, 620; 1902m, 455, 531, 546, 551, 552, 570, 576, 583, 586, 594, 595, 596, 598, 599, 600, 603, 604, 609, 612, 615-616.—Braun, 1901b, 41.—Pratt, 1902, 890, 909.

1901: Pronocephacus Braun, 1901b, 50, misprint.

obliquus Looss, 1901, 30. Okt., 566-567 ("trigonocephalus (Rud.)" of Looss, 1899b, 666, 756, figs. 84-86); 1902m, 527-532, pl. 25, figs. 55-56; pl. 26, fig. 70; pl. 32, fig. 175; 616 (includes "Monost. trigonocephalum" of Braun, 1901, 40).

"trigonocephalus (Rud., 1809)" Looss, 1899b, 666, 756-759, 760, 761, 762, figs. 84, 85, 86; 1901, 566 (renamed obliquus); 1902m, 527 (syn. of P. obliquus), 549 (of Rudolphi to Pleurogonius), see Looss, 1901, 567.—Braun, 1901b, 38, 40.—Shipley, 1900, 533 (Rud.).

PRONOPHARYNX Cohn, 1904, 239-240 (tod. nematoides).

nematoides Cohn, 1904, 238-240, fig. 5 (in Aquila albicilla; Coll. Creplin).

PRONOPYGE Looss, 1899b, 641–642 (ὁ πρών = Vorsprung; ἡ πύνη = der Hintere) (tod. ocreata); 1902m, 839.—Luehe, 1901, 481, 485.—Pratt, 1902, 889, 906.

ocreata (Rud., 1802) Looss, 1899b, 641–642.—Luehe, 1901, 400, 481.—See Dist. caroline.

PROSORHYNCHUS Odhn., 1905, 296, 297-305 (tod. squamatus).

aculeatus Odhn., 1905, 297, 302, 305 (syns. Gasterost. crucibulum Ben., 1870; G. armatum Olss., 1876) (in Conger vulgaris; Mediterranean, Belgium, Sweden).

crucibulum (Rud., 1819) Odhn., 1905, 297, 305 (syns. Monost. crucibulum Rud.= Gasterost. armatum Mol.=G. crucibulum Olss.) (in Conger sp.; Mittelmeer, Sweden). PROSORHYNCHUS—Continued.

squamatus Odhn., 1905, 297–304, 305, pl. 2, figs. 1–5 (syn. Gasterost. armatum Mol. of Levinsen) (in int. of Cottus scorpius; Belgium).—Nicoll, 1907, 70-71 (in Cottus bubalis, Liparis montagui).

PROSOSTOMATA Odhn., 1905, 305, suborder of Digenea to include all except the Gasterostomata (Gasterostomum, Prosorhynchus).

PROSOTOCUS Looss, 1899b, 616, 623 (πρόσω=cephalad; ό τόκος=das Gebären) (tod. confusus).—Pratt, 1902, 889, 902, 904.—Stiles, 1901, 197.

confusus (Looss, 1894) Looss, 1899b, 616.—Ssinitzin, 1905, 144-145; 1906, 687 (in Aeschna, Cordulia, and a beetle larva; Warschau).—Staff., 1905, Apr. 11, 684,

tener (Looss, 1898) Looss, 1899b, 616.—Staff., 1905, Apr. 11, 684.

PROSTHOGOMINUS Jahresb. ü. d. Leist. a. d. Geb. d. Veterinär-Med., v. 21 (1901) 1902, 248 (misprint for Prosthogonimus).

PROSTHOGONIMUS Luehe, 1899k, 539 (tod. ovatus); 1900, 555; 1901, 487.—Braun, 1901f, 561; 1901i, 56; 1902b, 67, 68, 69, 80, 85, 96 (syn. Prymnoprion Looss).-Looss, 1900, 608; 1902m, 834, 835.—Pratt, 1902, 889, 901.—Stiles, 1901, 183, 185.

1899: Prymnoprion Looss, 1899b, 628 (tod. ovatum).

1902: Prosthogominus Jahresb. ü. d. Leist. a. d. Geb. d. Vet.-Med., v. 21 (1901), 1902, 248, misprint.

1903: Prostogonimus Markow, 1903, 287–294, 295–297, figs. 1-7; 1903b, Aug. 14, 538; 1905b, July, 54.

anatinus Markow, 1902, 1903a, 287–298 (Prostogonimus), pl. 1, figs. 1-7 (in Anas boschas dom.); 1903b, 538.

cuneatus (Rud., 1809) Braun, 1901, 15-16, 17, fig. 2; 1901, 258-259; 1902b, 75-79, 80, 81, 82, figs. 44–45 (syns. Fasc. ovata Rud. (part), Dist. ovatum Rud. of Wedl, 1858; of Linst., 1878; of Auct.; D. cuneatum Rud.; Prymnoprion anceps of Looss, in Machetes pugnax; P. ovatus of Looss in Passer domesticus).—Wolffhuegel, 1906, 21 Nov., 21–25 (in hen's egg).—Reported for Cygnus musicus, Corvus cornix, C. corone, Anas clangula, Fulica atra, Fringilla cœlebs, Grus cinerea, Otis tarda, Pavo cristatus.

japonicus Braun, 1901, 17, fig. 3 (in hen's egg; Yedo, Japan); 1901, 258; 1902b, 81, fig. 46.

ovatus (Rud., 1803) [Luehe, 1899k, 539] Braun, 1901, 13–15, 17, fig. 1; 1901, 258; 1902b, 17, 69, 75, 76, 77, 78, 81, 83, 84, fig. 43 (syns. Dist. ovatus Rud.; Fasc. ov. Rud.).—Kowal., 1902d, [8] 26.—Markow, 1902, 292, 297 (Prostogonimus).—Reported for Larus canus, Anas glacialis, Pica caudata, Corvus cornix, Sturnus vulgaris.—T. h. Corvus frugilegus.

pellucidus (Linst., 1873) [Luehe, 1899k] Braun, 1901, 16–17, 18; 1901, 258–259; 1902b, 79, 80, 81, 82, fig. 45a (in Numenius arquatus, Gallus domesticus).

rarus Braun, 1901, 17–19, fig. 4 (in Fulica atra, Coll. Berlin; Anas boschas); 1901, 258, 259 (in Anas clypeata); 1902b, 22, 83, 84, 85, figs. 47, 48 (labelled Dist. ovatum in Berlin Coll. nos. 1615, 1423).

PROSTHOMETRA Looss, 1896b, 58-60 (felineus type by inclusion), no specific combinations made, but following species included: Dist. geminum, simulans, amphileucum, complexum, choledochum, longissimum, crassiusculum, xanthosomum, felineum; 1901, 194.—Stiles, 1901, 183, 185.

PROSTOGONIMUS Markow, 1903, 287, for Prosthogonimus.

PROTENTERON Staff., 1904, May 3, 494 (m. diaphanum) (πρῶτος=first; ἔντερον= intestine).

diaphanum Staff., 1904, May 3, 494 (t. h. Ambloplites rupestris; Canada).

PRYMNOPRION Looss, 1899b, 628–629 (πρυμνός=the undermost part; ὁ πρίων = the saw) (tod. ovatus); 1900, 608; 1902m, 839.—Braun, 1901i, 56; 1902b, 67, 68 (syn. of Prosthogonimus).—Ofenheim, 1900, 182.

 anceps Looss, 1899b, 629, 722–723, fig. 41 (in Machetes pugnax; Apr., Marg, Egypt).—Braun, 1901, 13, 16; 1902b, 68, 69, 78 (Looss from M. pug.; syn. of Prosthogonimus cuneatus).

ovatus (Rud., 1803) Looss, 1899b, 629, 720-722, 723, figs. 39-40 (in Passer domesticus; Alexandria and Cairo).—Braun, 1901, 16; 1902b, 73, 78 (Looss, from P. dom.; syn. of Prosthogonimus cuneatus).

pellucidus (Linst., 1873) Looss, 1899b, 629, 723.

PSEUDAXINE Par. & Perugia, 1890, 11-12, 19 (m. trachuri) (Microcotylidæ).—Braun, 1890a, 523, 540, 542, 546; 1893a, 890.—Gamb., 1896a, 73.—Mont., 1903, 336 (subf. Axininæ).—Pratt, 1900, 646, 653 (key), 657, fig. 41.

1879: Peudocotyle Tasch., 1879, 65, misprint.

trachuri Par. & Perugia, 1890, 11–12, pl. 14, figs. 10–13 (in Caranx trachurus;
Genoa).—Braun, 1890a, 542, 549, 550 (in C. tr.; Mittelmeer, Genua).—Par.,
1894, 595.—Pratt, 1900, 657, fig. 41.

PSEUDOCODYLE Tasch., 1879, 49, misprint for Pseudocotyle.

PSEUDOCODYLE Tasch., 1879, 49, misprint for Fseudocotyle.

PSEUDOCOTYLE Ben. & Hesse, 1865a, 4th appendix, 11–18 (m. squatinæ), Tristomidæ.—Ben., 1868, 28.—Brand., 1891d, 20.—Braun, 1890a, 411, 442, 451, 453, 455, 478, 482, 483, 484, 491, 492, 498, 511, 516, 517, 518, 523, 530; 1891d, 422; 1893a, 890; 1896b, 7.—Cerf., 1898b, 347, 356, 362.—Fraip., 1880c, 445; 1881b, 28.—Gamb., 1896a, 73.—Haswell, 1892b, 150; 1893e, 112, 114.—Hoyle, 1890, 539.—Ijima, 1884c, 638.—Jackson, 1888, 644, 646, 647, 653.—Kerbert, 1881a, 544, 572.—Mont., 1888a, 10, 13, 16, 20, 34, 37, 42, 43, 52, 53, 55, 56, 57, 58, 59, 60, 65, 66, 67, 88, 98; 1892, 108, 127, 128, 129; 1892, Oct. 7, 213 (gen. of Monocotylidæ); 1893, 118; 1905, 69, 70 (=Microbothrium Olss.); 1903, 336 (subf. Pseudocotylinæ; f. Monocotylidæ).—Pratt, 1900, 646, 649 (key), 655, fig. 15.—St. Remy, 1891, 213, 214, 222.—Tasch., 1878, 176; 1878, 573; 1879, 65 (Peudocotyle), 49 (Pseudocodyle), 44, 49, 50, 54, 55, 58, 60, 62, 64, 65, 66, 68; 1879, 236 (syn. Microbothrium Olss.). 68; 1879, 236 (syn. Microbothrium Olss.).

1879: Pseudocodyle Tasch., 1879, 49, misprint. 1879: Peudocotyle Tasch., 1879, 65, misprint.

apiculatum (Olss., 1868) Braun, 1890a, 530, 547, 550.—Mont., 1893, 206.—Staff., 1904, May 3, 482 (in Squalus acanthias; Canada).—Type of Microbothrium, 1869.—Reported also for Acanthias vulgaris.

fragile (Olss., 1868) Braun, 1890a, 530, 547, 551 (in Raja batis; Nordl. Eismeer).— Staff., 1904, May 3, 482 (syn. of Micropharynx parasitica Jægers., type of Mic).

minor Mont., 1888a, 16, 52, 60, 66 (in Scyllium); 1890, 191, fig. 4; 1891, 1892, 116, 127, 128; 1905, 70.—Braun, 1890a, 530, 547, 552 (in Sc. canicula; Mittelmeer, Naples).—Type of Leptocotyle 1905.

squalinæ Mont., 1892, 127, for squatinæ.

squatinæ Ben. & Hesse, 1865a, 4th appendix, 11-18, figs. 1-7 (in Squatina angelus; Ulme Ben. & Hesse, 1859a, 4th appendix, 11–18, ligs. 1–7 (in Squatina angelus; Ostende).—Ben., 1870, 13.—Brand., 1891d, 20.—Braun, 1889k, 622; 1890a, 420, 421, 424, 445, 449, 487, 530, 547, 552 (in Squat. ang.; North Sea, Mittelmeer, Naples); 1893b, 178.—Cerf., 1898b, 341.—Fraip., 1880c, 442.—Goto, 1891a, 162.—Ijima, 1884c, 638.—Juel, 1889, 36.—Kerbert, 1881a, 533.—Looss, 1885b, 5, 10.—Mont., 1888a, 7, 16, 19, 20, 23, 26, 32, 37, 39, 45, 52, 53, 66, 88; 1890, 420; 1892, 116, 127 (squalinæ), 128.—Par., 1894.—Pratt, 1900, 655, 657, fig. 15.—Tasch., 1878, 176; 1878, 573; 1879, 20, 1879, 48–55, 57, pl.3, fig. 2 (in Squat. ang.).—Ziegler, 1883, 545.

PSEUDOCOTYLINÆ Mont., 1903, 336 (f. Monocotylidæ); 1905, 69, 78.

PSEUDODISCUS Sons., 1895, 8 (for hawkesi, collinsi, ornatum), also written as subg.; 1895, 184, 185, 186; 1896, 310.—Fischder., 1903h, 489.—Piana & Stazzi, 1900,

cobboldi Montgomery, 1906, Feb. 12, 21 (in ponies; India).

hawkesi (Cobbold, 1875) Sons. [1895, 5], 1896, 310.—Piana & Stazzi, 1900, 519 [=Amphist. hawkesi].

ornatum (Cobbold, 1882) [Sons., 1895, 5].

(PSEUDODISCUS) as subg. of Amphist.—Sons., 1895, 187.

collinsi Cobbold.—Sons., 1895, 9, fig. 2; 1895, 187, fig. 2.

haukesi Cobbold of Sons., 1895, 9, fig. 1.

hawkesi Cobbold, Sons., 1895, 187, fig. 1.

PSILOSTOMIDE Looss, 1900, 604.

PSILOSTOMINÆ Looss, 1900, 604.—Pratt, 1902, 888, 896 (includes: Psilost., Crepidostomum, Rhytidodes, Allocreadium, Calycodes, Azygia, Helicometra, Cotylotretus; related genera: Ptychogonimus, Orchipedum).

PSILOSTOMUM Looss, 1899b, 573–574, 578, 579 ($\psi \lambda \delta \epsilon = \text{kahl}$) (tod. platyurum); 1900, 603, 604.—Braun, 1900, 232; 1901a, 33; 1901i, 56; 1901, 944; 1902b, 11, 15, 18, 19, 26, 147, 148.—Luehe, 1900, 489.—Pratt, 1902, 888, 897.—Stiles, 1901, 189.

PSILOSTOMUM-Continued.

brevicolle (Crep., 1829) Braun, 1902b, 12–14, fig. 9 (syns.: Dist. brevicolle Crep., 1829, 24; Duj., 1845a, 445; Mueller, 1897, 19; D. (Dicrocœlium) brevicolle Stoss., 1892, 35).

oxyurum (Crep., 1825) Braun, 1902b, 14, fig. 10.

platyurum (Mueh., 1896) Looss, 1899b, 574.

redactum Nicoll, 1906, 515, 525–527, pl. 13, figs. 9–10 (in Gasterosteus aculeatus; Scotland); 1907, 70, 73 (syn. of Podocotyle atomon).

simillimum (Mueh., 1898) Looss, 1899b, 574.

spiculigerum (Mueh., 1898) Looss, 1899b, 574.—Braun, 1902b, 15, 16, 155.

PTACUNELLA Massa, 1906, 58, for Placunella.

PTEROCOTYLE Ben. & Hesse, 1863, 1864, 96, 106 (palmatum=inhærens, type by inclusion; also type because it is only species figured).—Braun, 1890a, 477, 498, 516, 517, 522, 546.—Cerf., 1895h, 918, 920; 1896, 514, 515, 516.—Mont., 1888a, 7, 11, 16, 36, 66, 86, 89, 99; 1903, 336 (syn. of Dactycotyle).—Scott, T., 1901, 149.—Tasch., 1879, 240.

morrhuæ Ben. & Hesse, 1863; 1864, 106–107 (in Gadus morrhua).—Braun, 1890a, 492.—Cerf., 1895h, 922; 1896, 517; 1898a, 302.—Scott, 1901, 149–150, pl. 8, figs. 25-26 (in Gadus callarias, G. merlangus); 1905, 118 (in Scott, 1901, pl. 8).

Tasch., 1879, 246 (syn. of Octobothrium morrhuæ).

palmata (Leuck., 1830) Ben. & Hesse, 1863, 1864, 107–108, pl. 11, figs. 1–13 (syns. Octobothrium pal. Leuck., Octodactylus inhærens Dalyell) (in Gadus molva).—Cerf., 1895h, 917 (syn. of Octob. pal.); 1896, 513.—Scott, 1901, 149, pl. 8, fig. 27 (in Molva molva); 1905, 118 (in Scott, 1901, pl. 8).—Tasch., 1879, 246 (to Octob.).

PTERONELLA Ben. & Hesse, 1863; 1864, 94-95 (m. molvæ), fam. Udonellidés.— Braun, 1890a, 410, 446, 511, 516, 517, 523, 531, 532; 1893a, 890.—Gamb., 1896a, 73.—Mont., 1888a, 10, 86, 88, 98; 1892, Oct. 7, 213 (gen. of Udonellinæ); 1903, 336 (subf. Udonellinæ).—Pratt, 1900, 646, 649 (key), 655, fig. 14.

molvæ Ben. & Hesse, 1863; 1864, 94-95, pl. 8, figs. 20-23 (in Lota molva).—Braun, 1890a, 409, 418, 532, 548, 550, 551 (in Lota molva; Atlantic Ocean).—Linst., 1889a.—Pratt, 1900, 655, 657, fig. 14.—Tasch., 1878, 573 (to Udonella).

PTYCHOGONIMUS Luehe, 1900, 489 (m. megastomus).—Looss, 1901, 206; 1902m, 829, 830.—Pratt, 1902, 888 (related to Psilostominæ), 897, 900.

megastomus (Rud., 1819) Luehe, 1900, 489; 1901, 483.—Fischder., 1903h, 548.

PYCNOPORUS Looss, 1899b, 551, 610-611, 612, 618, 619 (πυκνός=firm, thick, strong, because of acetabulum) (tod. heteroporus); 1901, 199, 200.—Pratt, 1902, 889, 903.—Staff., 1903, 828.—Stiles, 1901, 197.

"acetabularis Looss, 1896," for acetabulatus Looss, 1899, in Braun, 1900, 388.

acetabulatus Looss, 1899b, 611, 717-719, fig. 36 (in Vesperugo kuhli; Egypt); 1901, 205; 1907, Mar. 5, 481 (in Vesp. k.; Cairo, Egypt), 488.—Braun, 1900, 227, 228.

heteroporus (Duj., 1845a) Looss, 1899b, 611.—Braun, 1900, 227, 228.—Staff., 1905, Apr. 11, 692-693.

inversus Looss, 1907, Mar. 5, 486–487, figs. 6 a-b (in Vesperugo kuhli; Cairo, Egypt).

PYELOSOMUM Looss, 1899b, 667 (ἡ πύελος=Trog, Wanne, Mulde) (m. cochlear); 1902m, 581, 582, 584, 586, 590, 593, 597, 600, 601, 602, 609, 611, 612, 613 (diagnosis), 614.—Braun, 1901b, 50.—Pratt, 1902, 890, 910.

cochlear Looss, 1899b, 667, 773-774, fig. 83 (in Chelonia mydas; Egyptian coast); 1902m, 416, 578–580, pl. 27, figs. 102–104; pl. 32, fig. 181; 593, 613.—Braun, 1901a, 53.

PYGIDIOPSIS Looss, 1907, 488 (m. genata), Heterophyiden.

genata Looss, 1907, Mar. 5, 488-490, figs. 7 a-c (in Pelecanus onocrotalus; Cairo, Egypt).

PYGORCHIS Looss, 1899b, 587 (m. affixus) (ή πυγή=der Steiss).—Braun, 1902b, 31, 37.—Ofenheim, 1900, 182.—Pratt, 1902, 888, 898.

affixus Looss, 1899b, 587, 596, 702-703, fig. 25 (in Corvus cornix; Marg, Galiub, Gizeh, Cairo; Falco tinnunculus; Galiub; Circus æruginosus; Nil, Adeleninsel; Recurvirostra avocetta).—Braun, 1901, 948; 1902b, 147.

RAJONCHOCOTYLE Ceri., 1899a, 347, 373, 420, 440, 445, 446, 455–456 (type batis, designated by Cerf. in correspondence with Stiles, Jan. 21, 1907).—Mont., 1903, 336 subf. Onchocotyline.

alba Cerf., 1899a, 377, 381, 383, 407, 408, 413, 417, 420, 433, 437, 438, 444, 446, 450, 463-464, pl. 18, figs. 5, 6, 8, pl. 19, fig. 9, pl. 20, figs. 1, 2, 3, 6, 7, pl. 21, figs. 1, 4, 8, 11, 13, 14, 15 in Raja alba Lacep. .

batis Cerf., 1899a, 376, 381, 383, 442, 451, 462-463, pl. 19, figs. 11, 12, pl. 21, fig. 12 syn. Onchocotyle appendiculata Olss., 1867 and 1876) (in Raja batis).

prenanti (St.-Remy, 1890), Cerf., 1899a, 377, 381, 383, 447, 464, pl. 18, figs. 10, 11, pl. 19, fig. 10, pl. 20, fig. 4 (syns. Onchocotyle appendiculata Kuhn of Sons., O. borealis Ben. of Stoss., 1885, O. prenanti St.-Remy, 1890) in Raja oxyrhynchus).

REDIA Fil., 1837a, 336-337 (m. gracilis); 1854a, 6.—R. Bl., 1888a, 542, 551, 553, 554, 555, 556, 557, 558, 559, 604, 645.—Braun, 1892a, 775; 1893a, 884.—Burm., 1856a, 245.—Dies., 1850a, 287, 301; 1855a, 379 (cf. sporocerca), 380, 383, 393; 1858d, 240 (cf. sporotherium).—Goldb., 1855a, 16.—Hoyle, 1890, 535.—Jackson, 1888, 644, 651, 652, 653.—Pag., 1857, 10.—Wagener, 1866, in 145-150, figs. 1-11.-Ward, 1903, 863, 865.

gracilis Fil., 1837a, 336-337, figs. 6-7 in Planorbis nitidus: Italy); 1854a, pl. 1, fig. 4.—Dies., 1850a, 301; 1855a, 394 (syn. of Diplocotyle mutabilis).— Moul., 1856a, 95 (syn. of Cerc. diesingii), 106, 107 (syn. of Cerc. amphistomi subclavati).—Pag., 1857, 25.—Par., 1894, 165.

RENICOLA Cohn, 1904, 235 (tod. pinguis).

pinguis Mehlis, 1846 Cohn, 1904, 232-235, figs. 2-3 (syn. Monost, pingue Mehlis, 1843; Braun, 1893a, 915, in Podiceps cristatus; Brand., 1892, 504-511).

RENIFER Pratt, 1902, 888, 899; 1903, 25 tld. ellipticus .

ellipticus Pratt, 1903, 25-28, 29, 34, pl. 4, fig. 1 (in Heterodon platyrhinus: North America .—Staff., 1905. Apr. 11. 691.

clongatus Pratt, 1903, 25, 28-30, pl. 4, fig. 2 (in Heterodon platyrhinus; North America .—Seely, 1906, 253, Staff., 1905, Apr. 11, 691 thinks this a Lechriorchis.

solitarius Looss, 1899 Pratt, 1903, 25.

cariabilis (Leidy, 1856) Pratt, 1903, 25, 31-34, pl. 4, fig. 5,

zschokkei (Volz. 1899) Pratt, 1903, 25, 28.

RENIFERINE Pratt, 1902, 888 contains Styphlodora, Ochetosoma, Renifer, Oistosomum. Astiotrema).

RHIPIDOCOTYLE Dies., 1858e, 313, 361–362 (type probably gracilescens).—Hausmann, 1897b, 34.—Mont., 1888, 92.—Odhn., 1905, 296.—Ziegler, 1883, 538.

grascilescens (Rud., 1819) Dies., 1858e, 361 in Lophius piscatorius: Apr., Triest; Mar., Pisa .—Linst., 1878a, 223 (includes Monost, isabellinum Ratzell).— Stoss., 1898, 61.—Tennent, 1906, 638.—Reported also for Lota molva, Merlangus vulgaris. Gadus æglefinus, G. melanostomus.

minima Wagener, 1852 Dies., 1858e, 361-362 in Trigla microlepidota .- Linst., 1878a, 227.—Tennent, 1906, 638 minimum.

RHOPALIADÆ Looss, 1899b. 543.

RHOPALIADINE Braun, 1901.—Looss, 1902m, 839.

RHOPALIAS Stiles & Hass., 1898a, 82, 93, 96 = Rhopalophorus Dies., 1850a [not Ropalophorus Westwood, 1840, hymenopteron; not Rhopalophorus Agassiz. 1846, Rhopalophora Serv., 1834] renamed | tod. coronatus | Braun, 1900, 28: 1900d, 27-29: 1901e, 318, 320, 324, 326-329 (syn. Rhopalophorus : 1903, v. 2, 23.—Looss, 1899b. 542. 581.—Pratt. 1902, 888 | related to Echinostominæ), 894.

baculifer Braun, 1900d, 28 in Didelphys palmata; Brazil; 1901e, 325-326, pl. 19, fig. 1.

coronatus Rud., 1819 Stiles & Hass., 1898a, 93.—Braun, 1900, 28, 29: 1901e, 320-323, 324, 326, 328, pls. 19, 20, figs. 2, 4, 11 (syns. Dist. cor. Rud., Rhopalophorus cor. Dies., Echinost, cor. Stoss).—Reported for Didelphys cancrivorus, D. nudicaudata, D. palmata. D. quica, D. virginiana.

horridus (Dies., 1850a, 400) Stiles & Hass., 1898a, 93.—Braun, 1901e, 323, 325, pl. 19, fig. 3 syns. Dist. coronatum; Rhopalophorus-horr. Dies. .- Reported

for Didelphys nudicaudats. D. philander.

RHOPALOCERA Fil., 1854a. 6 for Rhopalocerca).—Burm., 1856a, 250.

RHOPALOCERCA Dies., 1850a, 286, 293 (m. tardigrada=Dist. duplicatum renamed); 1855a, 379, 384; 1858d, 270.—Burm., 1856a, 250 (Rhopalocera).—Fil., 1854a, 6 (Rhopalocera).—Goldb., 1855a, 15.—Moul., 1856a, 121, 123.

tardigrada Dies., 1850a, 293–294 (Dist. duplicatum Baer, 1827b, renamed); 1855a, 378, 380, 384 (in Anodonta ventricosa, A. anatina); 1858d, 241, 242, 271–272 (syns. Dist. dup. Baer, Cerc. dup. Moul.) (in Anodonta ventricosa; Regiomontii; A. anatina, A. cygnea).—Jacobson, ——, 301, pl. 8, figs. 1–4 (syn. Dist. dup.).—Leidy, (1858) 1859, 110; 1877, 202.—Looss, 1894a, 63.—Pag., 1857, 6.—Žiegler, 1883, 540.

RHOPALOPHORUS Dies., 1850a, 288, 400 (not Ropalophorus Westwood, 1840; not Rhopalophorus Agassiz, 1846, for Ropalophorus, 1840; not Rhopalophora Serv., 1834) (tld. coronatus); 1855, 172; 1858e, 312, 357.—Braun, 1892a, 568, 569, 576, 595; 1893a, 879, 880, 886, 890, 893, 895, 908, 911, 918; 1900, 28; 1900h, 1855. 3; 1901e, 318, 319.—Carus, 1863, 479.—Gamb., 1896a, 73.—Goldb., 1855a, 17; 1899b, 536, 538, 542.—Mont., 1888a, 8, 11, 92, 105; 1892, Oct. 7, 214 (gen. of Distominæ); 1893, 82, 153, 154, 155.—Stiles, 1901, 172.—Stiles & Hass., 1898a, 93 (renamed Rhopalias, type coronatus).

coronatus (Rud., 1819a) Dies., 1850a, 400; 1855, 172–173, pl. 1, figs. 6–11; 1858e, 357.—Braun, 1893a, 911 (in Didelphys cancrivorus, D. myosurus, D. palmata, D. quica; Brazil); 1901e, 319, 320, 323, 324, 325.—Cobbold, 1879b, 432.—Kaiser, 1893a, 93.—Mont., 1893, 83.

horridus Dies., 1850a, 400 (in Didelphys myosurus, D. philander; Brazil); 1855, 173, pl. 1, figs. 12–16; 1858e, 357.—Braun, 1892a, 583; 1893, 911 (in Did. my., D. phil.; Brazil); 1901e, 319, 320, 323, 324.—Cobbold, 1879b, 432.—Mont., 1893, 83.—Stiles & Hass., 1898a, 93.

RHYTIDODES Looss, 1901l, 565 (tod. gelatinosus); 1902m, 451 (diagnosis), 839.—Odhn., 1905, 296.—Pratt, 1902, 888, 897.

gelatinosus (Rud., 1819) Looss, 19011, 563-565 (in Thalassochelys corticata Looss; Egypt); 1902m, 414, 416, 441, 445-456 (includes: Dist. gelat. Rud., 1819a, 386; Sons., 1890, 43; 1893; Stoss., 1898, 43; Braun, 1899, 716; 1901, 9, figs. 6, 12, pl. 1, fig. 9, pl. 2; Looss, 1899b, 579), 458, 460, 462, 463, 870, pl. 22, figs. 19-24.—Heymann, 1905, 82, 83, 87.

SACCOCŒLIUM Looss, 1902h, 134, 135 (tod. obesum).

obesum Looss, 1902h, 135, 140–141, 142, figs. 9–11 (in Mugil auratus, M. cephalus, M. chelo; Triest).

tensum Looss, 1902h, 141–142, figs. 12–13 (in Mugil chelo; Triest).

SAPHEDERA Looss, 1902m, 732, 839 (Macrodera, 1899, renamed) (type naja).— Staff., 1905, Apr. 11, 691.

1899: Macrodera Looss, 1899b, 603 (tod. naja) [not Macroderes ante 1882].

naja (Rud., 1819) Looss, 1902m, 732.

SCAPHANOCEPHALUS Jægers., 1903a, 1-16 (m. expansus), σκαπάνη=Spaten; κεφαλή=head.

expansus (Crep., 1842) Jægers., 1903a, 1-16, pl. 1, figs. 1-5 (in Pandion haliaëtus; Tor on Red Sea); 1904a, 279; 1904, 16 pp., 1 pl., 3 figs., Distomidæ.

SCAPHIOSTOMUM Braun, 1901g, 897 (m. illatabile); 1902b, 129, 133, 134.—Pratt, 1902, 889, 907 (key).

illatabile Braun, 1901g, 897 (in Falco nitidus Lath.; Brazil); 1902b, 134–136, fig. 80.

SCHISOSTOMA Colloridi, 1891a, 854, for Schistosoma.

SCHISTOMOSUM Cummins & Dupaquier, 1907, Jan., 496, for Schistosoma.

SCHISTOSOMA Weinland, 1858 [prior to Sept. 30], 58, 87 (m. hematobium) (not Schistosoma Brady, 1877, arachn.).—R. Bl., 1895, 730 (Schistosomum).—

Braun, 1893a, 894, 912; 1901, 562; 1901, 947; 1902b (Schistosomum), 140, 142, 144; 1903, 3 ed., 168; 1906, 176 (syns. Gynæcophorus, Bilharzia, Thecosoma).—Catto, 1904, 1499.—Christopher, 1905, Aug. 15, 259 (a peculiar egg of).—Christopher & Stephens, 1905, Nov. 11, 1289 (peculiar egg); 1905, 2341; 1905, Nov. 30, 1944; 1905, Aug. 26, 609.—Cobbold, 1879b, 39; 1885a, 498 (syn. of Bilharzia). Callavidi, 1891a, 854 (Schiscotoma). Days 1902, 669. Huber of Bilharzia).—Colloridi, 1891a, 854 (Schisostoma).—Darr, 1902, 660.—Huber, 1896, 580 (syn. of Bilharzia hæmatobia).—Katsurada, 1904b, 1237-1249 (Japanese); 1904c, 298–305 (Japanese); 1904, 160; 1904e (schistosomiasis in Japan); 1904g, Oct. 31, 126–134; Nov. 30, 135–148 (schistosomiasis in Japan); 1905a, Jan., 14-15.—Leuck., 1863a, 617.—Looss, 1895, 3; 1899b, 543, 656, 657, 658-659.—Moniez, 1896, 154-155.—Montgomery, 1906, 15-46, 2 pls.; 1906, Feb.

SCHISTOSOMA-Continued.

12, 18, 21 (Schistosumum).—Mont., 1896, 163.—Poche, 1907, 126.—Pratt, 1902, 889, 907.—Scheube, 1905, 150–155; 1905, 29 Aug., 1701.—Simon, 1897, 99.—Stiles, 1898a, 23, 27, 58, 63; 1903, 77.—Stiles & Hass., 1898a, 90, 93–94, 95, 98 (syns. Gynæcophorus, Bilharzia, Thecosoma) (type Dist. hæmotobium).— Ward, 1903, 224; 1903, 871–872.

1858: Gynæcophorus Dies., 1858, 356 (hæmatobius) later than Oct. 21, 1858.

1859: Bilharzia Cobbold, 1859, 364 (hæmatobia).

1860: Thecosoma Moguin-Tandon, 1860, 342 (hæmatobium).

1877: Bilhartzia Sons., 1877, 652 for Bilharzia.

1891: Schisostoma Colloridi, 1891a, 854, misprint.

1895: Schistosomum R. Bl., 1895, 40, for Schistosoma.

1905: Schistostoma Schwarz, 1905, 31 Mar., 236, misprint.

1906: Schistosumum Montgomery, 1906, Feb. 12, 21, misprint.

1907: Schistomosum Cummins & Dupaquier, 1907, Jan., 496.

bomfordi Montgomery, 1906, 143-147, pl. 1, figs. 1-4, pl. 2, fig. 1 (in Bos indicus; India).

bovis (Sons., 1876) R. Bl., 1895b, 101–104, fig. 7.—Braun, 1902b, 143 (syn. Bilharzia crassa).—Gomy, 1897a, 377.—Looss, 1905m, 281.—Moniez, 1896, 154.—Montgomery, 1906, Feb. 12, 17 (in cattle; Egypt; in sheep, Sicily), 18, 19, 43, 44; 1906, 138 (syns. Bilh. cr., B. bovis).—Rail., 1899, 788.—Stiles, 1898a, 23, 58, 60, 61, 62, 140, figs. 45, 46, 47.

28, 60, 61, 62, 140, ligs. 49, 46, 47.

cattoi R. Bl., in Catto [1904, Sept., 17, 663; 1904, Oct. 8, 710; 1904, Sept. 29, 1480; 1904, Nov., 15; 1904, Nov. 19, 1411; 1904, Nov. 26, 1499;] 1905, Jan. 7, 70–73, 9 figs. (in Homo); 1905, Jan. 7, 11–13, figs. 1–9 (in Homo; Prov. of Fukien, China); 1905, Jan. 28, 202; 1905, Jan. 19, 114; 1905, Mar. 1, 70–74; 1905, Mar. 31, 236; 1905, 179–189; 1906, Jan. 5, 617.—Brit. Med. J., Lond., 1905, v. 1, 27–28.—Crimp, 1905, Jan. 1, 67–68 (pathology); 1906, Apr. 22, 219.—Looss, 1905, 94; 1905m, 280.—J. Trop. Med., Lond., 1905, Apr. 1, 105.—Montgomery, 1906, Feb. 12, 18 (syn. of S. japonicum), 19.—Scheube, 1905, Apr., 150–155.—Stiles, 1905q, 821–823 (syn. of japonicum); 1905s, 1809; 1905t, 827.

crassum (Sons., 1877) Looss, 1899b, 657, 658.

 hæmatobium (Bilharz, 1852) Weinland, 1858, 87; 1859, 281.—R. Bl., 1888a, 636
 (syn. of Bilharzia hæmat.); 1895, 740-744, fig. 80; 1895, 40-101, figs. 2-6;
 1900, 488.—Bourel-Roncier, 1888a, 101ff.—Braun, 1902b, 144; 1906, 176-181, 1900, 488.—Bourel-Roncier, 1888a, 101ff.—Braun, 1902b, 144; 1906, 176–181, figs. 109–113 (syns. Dist. hæm., D. capense), 285.—Catto, 1904, 73; 1905, 71, 72; 1905, Jan. 7, 12.—Darr. 1902, 678.—Higgins, 1906, Mar. 24, 881–882 (in Canal Zone).—Katsurada, 1904, Dec., 148, 149, 150, 153, 154, 155, 156, 157.—Kowal., 1904, (10), 25 (in Homo; Kair).—Letulle, 1905e, 329–439, pls. 1–2, figs. 1–16; 1905c, 607–609.—Looss, 1896b, 158; 1899b, 657, 658, 751, 752; 1901, 27; 1905m, 281, 282, 284.—Moniez, 1896, 86, 154, 155–173, fig. 29; 1896, 86, to (Bilharzia).—Mont., 1896, 162, to (Bilharzia).—Montgomery, 1906, 139; 1906, Feb. 12, 16, 18, 31, 36, 40, 41, 43.—Rail., 1899, 787.—Roger, 1901, 94, 95.—Shaw, 1901, 1027.—Shipley, 1905, v. 6 (1), 4.—Stiles, 1898a, 23, 57, 58, 59, 60, 137, 138, 140, figs. 41, 42, 43, 44, 48; 1902, 40; 1902, 204; 1903, 8, 84.—Ward, 1895, 253 (syn. of Gynæcophorus hæmatobius), 328 (in Homo); 1903, 407; 1903, 863, 864, 872 (syns. Dist. hæm., Gynæcophorus hæm. Bilh. 1903, 407; 1903, 863, 864, 872 (syns. Dist. hæm., Gynæcophorus hæm., Bilh. hæm., Thecosoma hæm., Bilh. capensis).—Yamagiwa, 1905, v. 6 (3), 2 pls.

undicum Montgomery, 1906, 139; 1906, Feb. 12, 44, 45 (in donkey; India).

Montgomery, 1906, 183; 1906, Feb. 12, 44, 45 (In donkey; India).

*japonicum** Katsurada, 1904e, Aug. 3, 21 (in Felis catus dom. and Homo; Japan);

1904f, Dec., 147–160, pl. 7, figs. 1–10 (in Felis domestica; Homo; Japan,

Prov. of Yamanishi); 1904b, 1237–1249; 1904c, 298–305; 1904d, 1–22; 1904g,

126–134, 135–148; 1905a, 14–15; 1905, Jan. 31, 236; 1905, Apr. 1, 108–111.—

Beyer, 1905, Sept. 10, 578–579 (case in Asia); 1905, 4 pp.—Braun, 1906,

181–186, figs. 114–117 (syn. Sch. cattoi).—[Catto, 1904, Sept. 19, 1411.]—

Inouye, 1903, 131–132.—[Kurimoto, 1893, 20 Nov.]—Logan, 1905, v. 19,

243–245 (Hunan Province); 1906, Oct. 1, 294–296, figs. 1–3.—Looss, 1905m,

280–285; 1905n, 1362; 1906, Feb., 31; 1906, May 1, 132.—Montgomery, 1906,

140; 1906, Feb. 12, 18 (man and cat), 19, 40, 43, 44 (syn. S. cattoi).—Scheube. 250-253, 1905h, 1502; 1906, Feb., 31; 1906, May 1, 152.—Montgothery, 1906, 140; 1906, Feb. 12, 18 (man and cat), 19, 40, 43, 44 (syn. S. cattoi).—Scheube, 1905, Apr., 150-155.—Stiles, 1905q, 821-823 (syn. cattoi); 1905r, 7 pp.; 1905s, 1809; 1905t, 827; 1905bb, 854 (in Philippines); 1905, v. 39 (3), 280-285.—Woolley, 1906, 83-89, 3 pls., figs. 1-5 (in Philippines); 1906, Feb., 32-35; 1906, Apr. 21, 260; 1906, July 16, 530-531.

SCHISTOSOMA—Continued.

kowalewskii (Par. & Ariola, 1896) Rail., 1899, 788, to Bilharziella by Looss, 1899b. magnum (Cobbold, 1859) Rail., 1899, 788.—Looss, 1899b, 658.

mansoni Sambon, 1907, June, 365–366 (in Homo; Africa).

polonicum (Kowal., 1895) Rail., 1898, 412; 1899, 788.—Looss, 1899b, 658, type of Bilharziella.—Reported for Anas boschas, A. boschas dom., A. acuta, A. crecca, A. querquedula, Ardea cinerea, Mergus albellus.

[reflexum a term in teratology; it has no status in nomenclature.]

spindalis Montgomery, 1906, 147-150, pl. 2, fig. 2 (in Bos indicus; India).

SCHISTOSOMIASIS Toyama, 1905 (X, 26), 1739, medical term, name of the disease.— Miura, 1905a; 1906a, 46.—See also Bilharziosis.

SCHISTOSOMIDÆ Looss, 1899b, 542, 543, 659 (type Schistosoma: contains also Kællikeria, Bilharziella).—Braun, 1903, 3. ed., 168; 1906, 176.—Catto, 1905, (I, 7), 13.—Luehe, 1901, 488.—Montgomery, 1906, Feb. 12, 18, 19.—Pratt, 1902, 889, 907 (genera: Schistosoma, Bilharziella, Kællikeria).—Shipley, 1905, v. 6 (1), 4.—Ward, 1903, 864, 865.

SCHISTOSOMINÆ Stiles & Hass., 1898a, 90, 94, 95, 98.—Looss, 1901, 196.—Luehe, 1901, 175.—Stiles, 1898a, 22, 23, 58.

SCHISTOSOMUM R. Bl., 1895, 40, for Schistosoma, q. v.

SCHISTOSTOMA Schwarz, 1905, 31 Mar., 236, misprint for Schistosoma.

SCHISTOSUMUM Montgomery, 1906, Feb. 12, 21 (for Schistosoma).

SCHISTURUS Rud., 1809a, 8, 31 (m. paradoxus), 32; 1810a, 257–258.—Audouin, 1829, 408.—Blainv., 1824a, 513 "genre fort douteux établi par M. Rudolphi sur un animal incomplètement décrit par Redi''.—Deslongchamps, 1824uu, 2014. 674; 1829b, 238.—Dies.. 1850a, 331 (sym. Distomum).—Looss, 1899b, 527, 528; 1902m. 721, 764, 765. 778.—Luehe, 1900, 492.—Stiles, 1901, 195. 196.—Stiles & Hass., 1898a, 92, 94 (?sym. of Podocotyle).

paradoxvs Rud., 1810a, 257–258, pl. 12, fig. 4 (in Tetrodon mola; Europe); 1819a,
118 (syn. of Dist. nigroflavum).—Audouin, 1829, 408.—Dies., 1850a, 394 (syn. of D. nig., in Orthragoriscus mola; Naples).—Looss, 1899b, 528; 1902m, 721,
764.—Luehe, 1900, 492.—Stiles, 1901, 196.—Stiles & Hass., 1898a, 92, 93.

(SCHIZOCERCA) subg. of Cercaria.—Dies., 1858d, 264-265.

dichotoma (Mueller, 1855) Dies., 1858d, 265 (free; Nice).

fissicaudà (La Valette, 1855) Dies., 1858d, 265 (in Lymnæus stagnalis; Berlin). gracilis (La Valette, 1855) Dies., 1858d, 264–265 (in Planorbis corneus; Berlin).

SINISTROPORUS Staff., 1904, May 3, 484-485 (tld. simplex designated in letter from Staff.); sinister, left; $\pi \acute{o}\rho o\varsigma = \text{pore}$.

productus Staff., 1904, May 3, 485 (in Hemitripterus americanus; Canada) (productus, lengthened).

simplex (Rud., 1809) Staff., 1904, 484-485 (in Acanthocottus scorpius, Gasterosteus aculeatus, Hemitripterus americanus, Phycis chuss, Salmo salar, Scomber scombrus, Sebastes marinus).

SODALIS Kowal., 1902d, 27 (m. spathulatus), 28 (10).

spathulatus (Rud., 1819) Kowal., 1902, 27 (9).

SOLENOCERCA Dies., 1855a, 380, 383 "Der abgeworfene Sporenschwanz ist zuweilen von einem Schlauche oder einer Röhre durchgezogen (Solenocerca, Redia Filippi)."

SOLENOCOTYLE Dies., 1850a, 289, 420 (m. chiajei); 1858e, 314, 374.—Ben. & Hesse, 1864, 84.—Braun, 1890a, 518.—Goldb., 1855a, 19.—Mont., 1888a, 89.

chiajæ Tasch., 1879, 251, for chiajei.

chiajeæ Mont., 1888a, 89, for chiajei.

chiajei Dies., 1850a, 420 (Polyst. loliginis Chiaje, 1823, renamed) (in Loligo vulgaris; Naples); 1858e, 374.—Mont., 1888a, 89 (chiajeæ) Tasch., 1879, 251

SPATHIDIUM Looss, 1899b, 605 (ἡ δπάθη = spade) (tod. folium) [not Spathidium Duj., 1841a]; 1900, 605; 1901b, 202, 222 (Phyllodist.); 1902m, 476.—Braun, 1901b, 9.—Odhn., 1902, 65.—Osborn, 1903, 257 (syn. of Phyllodist. Braun,

cymbiforme (Rud., 1819) Looss, 1899b, 605.—Type of Plesiochorus 1901.

folium (Olfers, 1816) Looss, 1899b, 605.—See also Rhopalocerca.

patellare (Sturges, 1897) Looss, 1899b, 605.

SPBYRANURA Mont., 1888a, 11 (for Sphyranura).

SPECIES.—Looss, 1902, 779-794, variation of and conception of, among trematodes.

SPELOTREMA Jægers., 1901b, Dec. 31, 982 (tod. pygmæum); 1903a, 14, 15.—Looss, 1902m, 706, 824 (type pygmæum).

claviforme (Brand., 1888) Nicoll, 1907, 247, 248, 249, 254–256 (in Aegialitis hiaticula, Pelidna (Tringa) alpina).

excellens Nicoll, 1907, 247, 248-251, 252, 253 (syn. S. simile Jægers.) (in Larus argentatus).

feriatum Nicoll, 1907, 247, 248, 251–253 (in Aegialitis hiaticula, Hæmatopus ostralegus, Pelidna (Tringa) alpina, Totanus calidris, Vanellus vanellus).

pygmæum (Levin., 1881) Looss, 1902m, 785, 786, 809, 854.—Nicoll, 1907, 247, 248, 249, 250, 251, 253, 254, 255, 256 (in Oidemia fusca, O. nigra).—Odhn., 1905, 314–318, figs. 1, 2 (syns. Dist. pyg., Levinsenia pyg.) (in Larus sp.; west coast of Sweden).

simile (Jægers., 1900) Looss, 1902m, 706, 786, [809], 854.—Nicoll, 1906, 522 (similis) (to Levinsenia).—Odhn., 1905, 315, 316, 317, fig. 2b.

similis Nicoll, 1906, 522 (to Levinsenia), for simile.

SPH.EROSTOMA Rud., 1809a, 38, 39 (type by virtual tautonymy globiporum) [nec Sperosoma Kœler: nec Sphærodoma Keyes).—Darr, 1902, 655, 661.—Looss, 1899b, 527, 571, 595, 646, 647, 648-649; 1902m, 757, 758, 769, 830, 831.—Stiles, 1901, 167, 179, 188, 191, 192, 193, 194, 195, 200.—Stiles & Hass., 1898a, 94-95, 97 (type globiporum—bramæ).—Stoss., 1902, 582.

1902: Sphærostomum Looss, 1899b, 648, for Sphærostoma.

globiporum (Rud., 1802) Looss, 1899b, 595, 649; 1902m, 654, 747, 765.—Ssinitzin, 1905, 113–121; 1906, 685 (larva is Cerc. micrura).—Stiles, 1901, 194.

SPHÆROSTOMUM Looss, 1899b, 648-649 (for Sphærostoma Rud.).

SPHYRAMURA Mont., 1888a, 36 (for Sphyranura).

SPHYRANNRA Mont., 1888a, 49 (for Sphyranura).

SPHYRANURA R. Wright, 1879, 15–20, pl. 1, figs. 12–14, 54–75 (m. osleri).—Bettend., 1897a, 8, 38; 1897, 312, 342.—Brand., 1891d, 15; 1898a, 214 [22].—Braun, 1889k, 622; 1890a, 413, 415, 433, 434, 435, 437, 440, 442, 445, 451, 453, 457, 462, 463, 466, 469, 470, 481, 482, 483, 487, 491, 492, 511, 517, 523, 538, 540, 546; 1893a, 890.—Gamb., 1896a, 56, 73.—Goto, 1891c, 103.—Haswell, 1892a, 458, 460; 1892b, 149; 1893e, 112, 113, 114, 144, 145.—Hoyle, 1890, 539.—Lander, 1904a, 16, 17.—Looss, 1894, 23, 24; 1894a, 136, 203.—Mont., 1888a, 10, 11 (Spbyranura), 15, 16, 23, 34, 36 (Sphyranura), 37, 46, 47, 48, 49, 50, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 64, 65, 66, 67, 89, 100; 1892, Oct. 7, 213 (gen. of Polystominæ); 1893, 18, 39, 41, 111, 118; 1903, 336 (subf. Dicotylinæ).—Pratt, 1900, 646, 651, 656, 659, fig. 22 (Spyranura).

1888: Spbyranura Mont., 1888a, 11, misprint.

1888: Sphyrannra Mont., 1888a, 49, misprint.

1888: Sphyramura Mont., 1888a, 36, misprint.

osleri R. Wright, 1879, 66–71, pl. 1, figs. 13–14, in Necturus (Mesobranchus) lateralis; ? Canada.—Braun, 1890a, 410, 418, 422, 424, 425, 430, 449, 454, 456, 468, 490, 540, 548, 552; 1891a, 53.—Crety, 1893a, 384.—Goto, 1891a, 160.—Haswell, 1892b, 150, 151; 1893e, 97, 99, 115.—Kowal., 1898h, 158 (55).—Lander, 1904a, 16.—Looss, 1894, 21; 1894a, 234.—Mont., 1888a, 8, 13 (oslerii), 20 (Spyranura), 25, 26, 27, 29, 32, 34, 35, 37, 39, 41, 44, 47, 48, 49 (Sphyrannra), 56; 1891, 1892, 99, 104; 1893, 16, 39, 112.—Pratt, 1900, 656, 657, 659 (key) (Spyranura), fig. 22.—Staff., 1900, 405; 1905, 682 (in Necturus maculatus Raf.; Canada). Wright & Macallum, 1887, 1–48, pls. 1–48.

oslerii Mont., 1888a, 8, 13, for osleri.

SPOROCERCA Dies., 1855a, 379–380, "Der abgeworfene Schwanz der Cercarieen ist somit ein Sporenbehälter, welcher von Baer als Sporocystis, von Siebold als Keimschlauch, von Steenstrup als Amme, von Beneden als Scolex and von Filippi als Redia beschrieben und abgebildet wurde. Ich werde mich in der Folge dafür des Collectivnamens Sporenschwanz (Sporocerca) bedienen."

SPOROCYSTIDES Baer.—Dies., 1858d, 240, 242.

SPOROCYSTIS Baer.—Braun, 1892a, 775.—Burm., 1856a, 245.—Dies., 1855a, 379 (cf. sporocerca); 1858d, 270 (cf. sporonema).—Fil., 1854a, 6.—Pag., 1857, 10.—Wagener, 1866, in 145–150, figs. 1–11.

SPOROCYSTOPHORA Fil., 1856.

SPOROCYSTOPHORÆ Fil., ——.—Dies., 1858d, 240, 242.

SPORONEMA Dies., 1855a, 380, "Bei allen zwei schwänzigen [Cercaria] hingegen wächst die Spitze des Sporenschwanzes in eine fadenförmige unverästelte oder verästelte Röhre aus, welche stellenweise in Knoten oder Kugeln anschwillt, die neue Brut in allen Graden der Entwickelung einschliesst, und endlich auch selbst vom Sporenschwanze abfällt (Sporonema)"; 1858d, 270 (cf. Sporocystis).

SPORONEMATA Dies., 1858d, 240.—See Sporonema.

SPOROTHERIUM Dies., 1858d, 240 (i. e. Redia Fil.).

SPYRANURA Mont., 1888a, 20, for Sphyranura.

SQUALONCHOCOTYLE Cerf., 1899a, 347, 373, 420, 440, 445, 446, 454–455, 456 (type borealis, designated by Cerf. in correspondence with Stiles, Jan. 21, 1907).—Mont., 1903, 336 (subf. Onchocotyline).

abbreviata (Olss., 1876) Cerf., 1899a, 375, 380, 382, 460, pl. 19, fig. 3 (syn. Oncho-

cotyle abbreviata Olss.) (in Acanthias vulgaris; Roscoff).

borealis (Ben., 1853) Cerf., 1899a, 374, 379–380, 382, 421, 422, 423, 424, 427, 428, 431, 433, 434, 436, 443, 444, 446, 456–457, pl. 18, fig. 13, pl. 19, fig. 7, pl. 20, figs. 13, 14, pl. 21, fig. 16 (syn. Onchocotyle borealis Ben.) (in Scymnus borealis; Liége).—Odhn., 1905, 372 (syn. Onchoc. bor. Ben.).

canis Cerf., 1899a, 375, 380, 382, 398, 406, 410, 433, 443, 447, 450, 458–459, pl. 18, figs. 1, 2, 7, 12, pl. 19, fig. 2, pl. 20, fig. 12, pl. 21, figs. 2, 6, 10 (in Galeus canis;

Roscoff).

grisea Cerf., 1899a, 376, 381, 383, 461, pl. 19, fig. 8 (syn. Onchocotyle appendiculata Tasch., 1879) (in Hexanchus griseus).

spinacis (Goto, 1894) Cerf., 1899a, 376, 380–381, 383, 404, 417, 419, 424, 460–461, pl. 19, fig. 4 (syn. Onchocotyle spinacis Goto) (in Spinax sp.).

vulgaris Cerf., 1899a, 375, 379, 382, 403, 406, 409, 419, 427, 437, 438, 444, 446, 450, 457-458, pl. 18, figs. 3, 4, 9, pl. 19, fig. 1, pl. 20, figs. 5, 8-11, pl. 21, figs. 3, 5, 7, 9 (syns. Polyst. appendiculatum of Thaer, 1850; Onchocotyle emarginata of Sons., 1890) (in Mustela vulgaris).

STEGANODERMA Staff., 1904, May 3, 486–487 (m. formosum) (στεγανός=covered). formosum Staff., 1904, May 3, 486–487 (t. h. Hippoglossus hippoglossus; Canada.)

(formosus, well formed).

STENAKRON Staff., 1904, May 3, 487 (m. vetustum) (στενός=narrow; ἄκρον=end).

retustum Staff., 1904, May 3, 487 (in Hippoglossus hippoglossus; Hemitripterus americanus), based on Linton, 1901, 485, pl. 32, fig. 359, pl. 33, figs. 360–362, in Limanda ferruginea; Woods Hole) (vetustus old).

STENOCOLLUM Staff., 1904, May 3, 487 (m. fragile); στενός, narrow; collum, neck.

fragile (Lint., 1900) Staff., 1904, 487 (in Mola mola).

STEPHANOCHASMUS Looss, 1900, Dec. 3, 603 (=Stephanostomum Looss, 1899 [not Stephanostoma Danielson & Koren] renamed, hence type cesticillus); 1901b, 199; 1901e, 595, 597-605, 628, 629, 634, 654-661; 1901i; 1902g; 1902m, 441, 780, 808, 838.—Braun, 1901a, 34; 1902b, 30.—Odhn., 1902, 37; 1902, 155, 159; 1905, 331.—Pratt, 1902, 888, 895.

1899: Stephanostomum Looss, 1899b, 576 (tod. cesticillus), not Stephanostoma

Danielson, 1880, worm.

baccutus Nicoll, 1907, 72, 80–83, pl. 2, figs. 5–7 (in Hippoglossus vulgaris; Scotland). bicoronatus (Stoss., 1883) Looss, 1901e, 599–601, 604, fig. 2 (in Umbrina cirrhosa, Corvina nigra).

caducus Looss, 1901e, 603-604, fig. 4 (in Gadus minutus, Lophius piscatorius;

apparently Triest).—Nicoll, 1907, 81.

cesticillus (Mol., 1858) Looss, 1901e, 598-599, 601, 604, fig. 1 (in Lophius piscatorius).—Nicoll, 1907, 80, 81, 82, 83.

histrix (Dies., 1850, for hystrix Duj., 1845) Staff., 1904, May 3, 485 (on fins of Pseudopleuronectes americanus Walb., and in cysts free in stomach of Lophius piscatorius containing 2 partially digested Limanda ferruginea).

minutus Looss, 1901e, 604-605 (in Uranoscopus scaber; apparently Triest).—

Nicoll, 1907, 83.

pristis (Delongchamps, 1824) Looss, 1901e, 601–603, 604–605, figs. 3, 5 (in Gadus euxinus, G. minutus, Motella vulgaris).—Nicoll, 1907, 80.

sobrinus (Levin., 1881) Staff., 1904, May 3, 485 (in rectum of Hemitripterus americanus, Cryptacanthodes americanus, Lycodes sp.; Canada.)—Odhn., 1905, 331 (syn. Dist. sobrinum).

STEPHANOPHARYNX Fischder., 1901, 370 (m. compactus); 1902a, 24; 1903h, 492 (Paramphistominæ), 551.—Looss, 1902m, 835.—Pratt, 1902, 887, 892.— Shipley, 1905, v. 6 (1), 8.

compactus Fischder., 1901, 370 (in Bos taurus); 1902a, 24-26, fig. 2 (in Bos taurus;

Africa); 1903h, 493, 496, 566, 567, 584.

STEPHANOPRORA Odhn., 1902, 22-24 (m. ornata), fig. 1.—Pratt, 1902, 888, 895

ornata Odhn., 1902, 22–24, fig. 1 (in Nilkrokodil).

STEPHANOSTOMUM Looss, 1899b, 576–577, 579, 582, 583, 590, 596 (tod. cesticillus) [not Stephanostoma Danielsen, 1880, Vermes), ὁ στέφανος, Stirnkrone, Kranz; 1900, 603 (renamed Stephanochasmus); 1901, 595.—Braun, 1901b, 34; 1901i, 55.—Luehe, 1900, 489.—Rud., 1801a, 58.—Stiles, 1901, 189.

cesticillus (Mol., 1858) Looss, 1899b, 576, 582, 696–698, figs. 21–22 (in Lophius piscatorius; Naples).

hystrix (Duj. of Olss.) Looss, 1899b, 576, 582.

lydæ (Stoss., 1896) Looss, 1899b, 576, 582.—Type of Dihemistephanus 1901.

pristis (Deslongchamps, 1824) Looss, 1899b, 576, 582, 697.

sobrinum (Levin., 1881) Looss, 1899b, 576, 582.

STERELMINTHA Owen, 1835, 390.—Aitken, 1872, 146, 178–207; 1874, 57.—Baird, 1853a, 39 (pars=Trematoda).—Dunglison, 1893, 820, 1039.—Mont., 1888a, 84.—Sieb., 1839, 153.—Tasch., 1879, 234.

STERINGOPHORUS Odhn., 1905, 309 (tod. furciger).

furciger (Olss., 1868) Odhn., 1905, 301, 305–310, pl. 2, figs. 6-7 (in Pleuronectes limanda and Drepanopsetta platessoides; on Swedish west coast; Cottus scorpius, Gymnocanthus ventralis, Lycodes pallidus; East Greenland).-Nicoll, 1907, 72, 183 (in Pleuronectes limanda).

STERRHURINAE Looss, 1907, 599-600, subf. of Hemiuridæ, contains: 1. Sterrhurus, 2. Lecithochirium, 3. Synaptobothrium, 4. Plerurus.

STERRHURUS Looss, 1907, 600 (tod. musculus).

fusiformis (Luehe, 1901) Looss, 1907, 602 (in Conger conger).

grandiporus (Rud., 1819) Looss, 1907, 601-602 (in Muræna helena; Naples).

imocavus Looss, 1907, 601 (in Thypnus sp. (? thunnina); Alexandria, Egypt).

musculus Looss, 1907, 600-601 (in Anguilla vulgaris, Dentex vulgaris; Triest; also in Acipenser sturio, Gadus euxinus, Gobius jozo. Labrax lupus, Lichia amia, Lophius piscatorius, Ophidium barbatum. Pagellus erythrinus, Rhombus maximus, Šerranus cabrilla, Trachinus draco; Triest).

STICHOCOLYLE Mont., 1892, Oct. 7, 176, for Stichocotyle.

STICHOCOTOLYLE Mont., 1888a, 9, for Stichocotyle.

nephropis (Cunningham, 1884) Mont., 1888a, 9, for Stichocotyle nephropis.

609, 610, 612, 614, 615, 616, 617.—Odhn., 1902, 44, 45.—Pratt, 1902, 887, 891.

1888: Stichocotolyle Mont., 1888a, 9, misprint. 1892: Stichocolyle Mont., 1892, 176, misprint.

nephropis Cunningham, 1884a, 399 (in Nephrops norvegicus; Firth of Forth); 1887a, 273–280, pl. 39, figs. 1–6.—Braun, 1892a, 571.—Jackson, 1888, 642.—Jægers., 1899, 202.—Mont., 1888a, 9 (Stichocotolyle), 12, 26, 38, 39, 80, 89; 1893, 36, 48. Nickerson, 1895, 447–480, pls. 29–31 (in American lobster); 1895, 634–635; 1902, 614, 617 (in Raja sp.; Europe, N. America).—Odhn., 1898, 509–513 (sexual form).—Rossbach, 1906, 374.

(STICHORCHIS) Fischder., 1901, 373 (tod. giganteus), subg. of Cladorchis: 1902a,

41 (type giganteum).—Looss, 1902m, 836.—MacCallum, 1905b, 668.

giganteus (Dies., 1836d) Fischder., 1901, 373; 1902a, 41–42, 43 (in Dicotyles albirostris, D. torquatus).

subtriquetrus (Rud., 1814) Fischder., 1901, 373–374; 1902a, 35, 42–43 (syns. Amphist. subtriquetrum Rud., Dist. amphistomoides Boj.) (in Castor fiber); 1903h, 567.

STICHORCHIS (Fischder., 1901) Looss, 1902m, 439.

subtriquetrus (Rud., 1814) Looss, 1902m, 439.

STICTODORA Looss, 1899b, 671-672 (m. sawakinensis), στικτός, punctuate; ή δορά, skin; 1902m, 442.—Pratt, 1902, 890, 910.

sawakinensis Looss, 1899b, 672,754–755, fig. 90 (in Laurus sp.; Sawakin, Egypt, Jan.).

STOMYLOTREMA Looss, 1900, Dec. 3, 602 (=Stomylus Looss, 1899 [not Fæhr., 1871] renamed) (type singulare=perpastum).—Braun, 1901, 260; 1901, 896; 1902b, 85, 86, 91 (syn. Stomylus) (type S. perpastum Braun=singulare Mol. of Looss).—Odhn., 1902, 38.

1902: Stromplotrema Zool. Rec., 1902 (1901), v. 38, 53, Vermes, misprint.

bijugum Braun, 1901g, 896 (in Himantopus melanopterus Mey.; Brazil); 1902b, 91, fig. 53.

fastosum Braun, 1901g, 896 (in Caprimulgus sp., Squatarola helvetica; Brazil); 1902b, 90, fig. 52.

perpastum Braun, 1902b, 86, 88 (syn. Stomylus singularis of Looss, not Dist. singulare Mol.).

pietum (Crep., 1837) Braun, 1901, 896; 1902b, 86 (syns. Dist. pictum Crep., 1837, 313, 316; D. singulare Mol., 1861, 200).

[singulare (Mol., 1861, of Looss)].

tagax Braun, 1901g, 896 (in Hirundo versicolor Natt.; Brazil); 1902b, 90, fig. 51.

vicarium Braun, 1901, 896 (in Ibis cœrulescens Vieill.; Brazil); 1902b, 89, 90, fig. 1.

STOMYLUS Looss, 1899b, 629–630 (m. singularis) (στωμύλος=Der ein gutes Mundwerk hat) [nec Stomylus Fåhræus, 1871, coleopteron]; 1900, Dec. 3, 602 (renamed Stomylotrema).—Braun, 1902b, 85, 86, fig. 49 (syn. of Stomylotrema).

singularis (Mol., 1861, of Looss) Looss, 1899b, 596, 630, 723-725, figs. 42-43 (in Glareola prantincola; Dachschur).—Braun, 1901, 260; 1901, 896; 1902b, 86, 88 (syns. Stomylotrema perpastum, not Dist. singulare Mol.).

STRIGEA Abildg., 1790a, 37, pl. 5, a-c (m. strigis), mentions only Geze, 1782a, 174, pl. 14, figs. 4–6 (=strigis).—Baird, 1853 (=Holost. macrocephalum).—Brand., 1888a, 9.—Braun, 1893a, 884, 894, 902.—Cuv., 1817, 41.—Deslongchamps, 1824yy, 701; 1829e, 677.—Dies., 1850a, 307 (syn. Hemist. Dies.), 312 (syn. of Holost. variabile).—Fischder., 1901, 367; 1902a, 6 (renamed Amphistoma by Rud.), 7 (m. strigis Gœze, 1782a,—Amphist. macrocephalum Rud., 1809, —Holost. macrocephalum Nitzsch, 1819) ("if again made valid—Holost., while Amphist. must receive a new name"); 1903h, 487, 490.—Linst., 1905, 191.—Looss, 1902m, 438, 439, 746, 756.—Luehe, 1901, 175.—Rud., 1801a, 59; 1809a, 21.—Shipley, 1905, v. 6 (1), 7 (type Holost. macr.).

1801: Amphistoma Rud., 1801a, 50 (Strigea renamed).

1819: Holostomum Nitzsch, 1819, 399-401, type by inclusion variabile=strigis. anatis tadornæ Viborg, 1795, 196.—Dies., 1850a, 313 (syn. of Holost. erraticum).— Rud., 1809a, 352 (to Amphist.).

candida Mueller (or Abildg.), 1806a, v. 4, 32, pl. 143, fig. C, 1-2.—Dies., 1850a, 313 (syn. of Holost. erraticum).—Olfers, 1816, 48.—Rud., 1814a, 100 (renamed

Amphist. isostomum).

cervi (Schrank, 1790) Rail., 1893a, 376.—Stiles, 1898a, 64.

falconis palumbi Viborg, 1795, 243.—Baird, 1853a, 47 (=Holost. macrocephalum).—Dies., 1850a, 309 (syn. of Hemist. spathula).—Rud., 1809a, 352 (to Amphist.).

longicollis (Duj. [Rud., 1819]) Luehe, 1905, 215 (syns. Echinorhynchus gazæ. Gmelin, Zed.; E. ardeæ albæ Rud.; E. sp. Mueller).

STROMPLOTREMA Zool. Rec., 1902 (1901), v. 38, 53 Vermes, for Stomylotrema.

STYPHLODORA Looss, 1899b, 592, 593, 594, 633 (tod. serrata) $(\sigma \tau \nu \phi \lambda \delta \dot{\phi} = \text{rough}; \dot{\eta} \delta o \rho \dot{\alpha} = \text{skin}); 1901, 560; 1902m, 505, 506, 507 (diagnosis), 512, 514, 821.— (cohn, 1902, 882.—Odhn., 1902, 27, 40, 42.—Pratt, 1902, 888, 899.$

1902: Styphlodera Looss, 1902m, 872, misprint.

serrata Looss, 1899b, 592, 707-708, 709, fig. 28 (in Varanus niloticus; Zool. Inst., Leipzig).—Luehe, 1900, 561.

solitaria Looss, 1899b, 592, 708-709, fig. 29 (in Thalassochelys corticata; Abukir); 1902m, 505, 506–507, 511, 517, 872, pl. 24, fig. 40.—Luehe, 1900, 560, 561.—Pratt, 1903, 25 (Styphlodera), 33, to (Renifer).

SUBCLAVATA Gœze, 1782, 41, 169, 178-179 (a "Klasse" of Plan. Gœze, 1782, containing Fasc. subclavata Pallas).

- SYNAPTOBOTHRIUM Linst., 1904, 254 (m. copulans).—Odhn., 1906, 59-66, figs. 1-2 (syn. of Lecithochirium Luehe, 1901, 473).
 - caudiporum (Rud., 1819) Looss, 1907, 604 (in Zeus faber; Triest; also in Caranx trachurus, Lophius piscatorius, Platessa passer, Rhombus lævis, R. maximus, Scomber colias; Triest).
 - copulans Linst., 1904, 254, figs. 1–4 (in Arnoglosus laterna; ?Louvain); 1906, 751.—Odhn., 1906, Apr. 3, 59–66, figs. 1–2 (to Lecithochirium).
- SYNCŒLIIN.E Looss, 1899b, 544, 642, 645, 646; 1901, 208 (includes: Syncælium, Progonus, Otiotrema).—Fuhrmann, 1904, 61 (Syncoliinæ).—Odhn., 1905, 366.—Pratt, 1902, 889, 905 (includes: Progonus, Syncælium, Otiotrema; related genera Halipegus, Accacælium, Eurycælum).

1904: Syncoliinæ Fuhrmann, 1904, 61, for Syncœliinæ.

- SYNCŒLIUM Looss, 1899b, 643-644, 645, 646 or 647, 741, 742, 743 (m. ragazzii); 1901, 208, 209, 210; 1902m, 642, 813.—Braun, 1902b, 23.—Fuhrmann, 1904, 61.—Luehe, 1901, 481, 482, 486.—Odhn., 1905, 366.—Ofenheim, 1900, 160.—Pratt, 1902, 889, 905.
 - ragazzii (Setti, 1897) Looss, 1899b, 644, 731-736, 737, 738, 743, 746, figs. 50, 62, 63-68 (in Lamna sp.; Sawakin).—Darr, 1902, 661.—Luehe, 1901, 481.

SYNCOLIINÆ, see Syncœliinæ.

- TAPHROGONIMUS Cohn, 1904, 235-237 (m. holostomoides).
 - holostomoides (Mehlis, 1846) Cohn, 1904, 235–237 (syn. Monostomum pingue Crep., 1843; Braun, 1893a, 915; Brand., 1892, 504–511).—Reported for Podiceps cristatus.
- (TAXORCHIS) Fischder., 1901a, 373 (tod. schistocotyle); 1902a, 39–40; 1903h, 498.—Looss, 1902m, 836 (as genus).
 - schistocotyle Fischder., 1901a. 373 (in Dicotyles torquatus); 1902a, 40–41 (syn. Amphist. giganteum Dies., e. p.).
- TELORCHIINÆ Looss, 1899b, 569.—Braun, 1901b, 20.—Pratt, 1902, 888, 895 (includes Telorchis, Orchidasmus; related genus Deropristis).
- TELORCHIS Looss, 1899b, 566–569, 614 (tod. linstowi=? aculeatus): τηλοῦ,τῆλε, in der Ferne; ὄρχις, testes; 1900, 608.—Braun, 1900, 234.—Stiles, 1901, 183, 185.—See also D. ercolanii, D. poirieri, D. nematoides, D. arrectum.
- TELORCHIS Luehe, 1899, 529, 530 (tod. clava): 1900, 556, 566; 1901, 488.—Braun, 1900, 390; 1901b, 13, 14, 17, 19, 20, 21, 26, 30, 34; 1901i, 56, 58.—Looss, 1901, 207; 1902m, 824, 831, 832, 834, 839.—Pratt, 1902, 888, 895.
 - aculeatus (Linst., 1879) Braun, 1901b. 14–17 (syn. Dist. linstowi Stoss.), 19, fig. 4.—Stoss., 1904, 4, 5, 6, 8 (in Testudo græca; Albania).
 - arrectus (Duj. of Mol., 1859) Stoss., 1904, 9 (in Podarcis muralis; Italy).
 - augustus Staff., 1905, Apr. 11, 690 (in Chrysemys picta; Canada), misprint for angustum (Dist.), 1900.
 - bifurcus (Braun, 1899) Braun, 1901, 18–19, 30, fig. 2.—Heymann, 1905, 95 to (Cercorchis).—Stoss., 1904, 9 (Testuggini d'acqua dolce; Brazil).
 - clava (Dies., 1850) Luehe, [1899, 529;] 1900, 566.—Looss, 1900, 608; 1901, 207; 1902m, 831.—Odhn., 1900, 17.—Stoss., 1904, 9 (in Eunectus scytale; Brazil).
 - ercolanii (Mont., 1893) Braun, 1901b, 16.—Rizzo, 1902, 28 (syn. Dist. monticelli) (in Tropidonotus natrix; Catania).—Stoss., 1904, 4, 5-6 (syn. T. nematoides) (syn. Dist. ercolanii) (in Trop. nat., T. viperinus; Italy).
 - *linstowi* (Stoss., 1890) Looss, 1899b, 566 (type of Telorchis Looss, not Luche).—Braun, 1901b, 14 (syn. of aculeatus).—Luche, 1900, 566 to (Cercorchis, type).—Stoss., 1904, 3, 4.
 - nematoides (Mueh., 1898) [Luehe, 1899, 529.—] Braun, 1901b, 16.—Looss, 1899b, 567.—Odhn., 1900, 17.—Stoss., 1904, 4, 6 (syn. T. ercolanii) (in Tropidonotus natrix; Germany).
 - parvus Braun, 1901b, 19–20, fig. 3 (in Testudo orbicularis; Vien. Mus.).—Heymann, 1905, 95, to (Cercorchis).—Stoss., 1904, 9 (in Cistudo lutaria).
 - pleroticus (Braun, 1901) Braun, 1901b, 17–18, fig. 5.—Heymann, 1905, 95, to (Cercorchis).—Stoss., 1904, 9 (in Testuggini d'acqua dolce; Brazil).
 - poirieri (Stoss., 1895) Odhn., 1902, 31.—Heymann, 1905, 95, to (Cercorchis).—Stoss., 1904, 3–5, fig. 2 (syns. Dist. gelatinosum Rud. of Poir., 1885, D. poirieri) (in Cistudo lutaria; Italy and France; in Emys orbicularis; Sassari, Sardegna).
 - solivagus Odhn., 1902, 29–32, fig. 2 (in Clemmys caspica).—Stoss., 1904, 9 (in Cl. casp.; Caucasia).

TELORCHIS (CERCORCHIS) Heymann, 1905, 94, sp. (in Dermatemys mavii).

TEMNACEPHALE Cosmovici, 1887a, 127 (for Temnocephala).

TEMNOCEPHALA E. Bl., 1849, 51-52 (m. chilensis).—Bettend., 1897a, 8; 1897, DCEPHALA E. B1., 1849, 51–52 (m. chilensis).—Bettend., 1897a, 8; 1897, 312.—R. Bl., 1888, 137, 138 (m. chilensis) (on écrevisse; Chili), g. of Branchiobdellides.—Brand., 1891d, 13, 24, 27, 28, 29.—Braun, 1889k, 620, 621; 1890a, 408, 412, 422, 424, 425, 429, 430, 435, 436, 437, 440, 442, 444, 445, 447, 451, 456, 458, 461, 465, 466, 468, 469, 470, 471, 472, 475, 479, 481, 482, 485, 490, 491, 497, 499, 510, 511, 512, 517, 520, 521, 522, 523, 525, 1890b, 84–90, 125–128; 1890e, 595; 1893a, 889; 1893b, 179; 1895b, 25; 1906, 129.—Buttel-Reepen, 1900a, 590.—Chilton, 1888, 252; 1889a, 252–253 (on crayfish in New Zealand).—Cosmovici, 1887a, 127 (Temmacephale).—Darr, 1902, 649, 678.—Gamb., 1896a, 4, 56, 73.—Graff, 1903.—Haswell, 1887a, 279–302, pls, 20–22; Zealand).—Cosmovici, 1887a, 127 (Temnacephale).—Darr, 1902, 649, 678.—Gamb., 1896a, 4, 56, 73.—Graff, 1903.—Haswell, 1887a, 279–302, pls. 20–22; 1888a, 50–51; 1888, 279–302; 1892a, 457ff; 1892b, 149–151; 1892c, 486; 1892d, 360–362 (integument); 1893, 455–460; 1893c, 342; 1893d, 477; 1893e, 93ff; 1893f, 153, 154, 155, 156; 1894, 93–152.—Kathariner, 1894a, 132, 134.—Looss, 1894a, 219; 1895, 36.—Maclaren, 1904, 582, 601.—Mont., 1888a, 10, 15, 17, 18, 22, 27, 33, 34, 35, 36, 38, 39, 42, 43, 44, 45, 47, 48, 49, 50 (Temuocephala), 51, 52, 53, 54, 56, 57, 58, 64, 66, 67, 86, 88, 98, 110; 1889b, 4pp., 3 figs.; 1891, 1892, 100, 108, 109, 110, 128, 129,; 1892, Oct. 7, 186, 213 cg. of Temnocephalidæ); 1893, 8, 9, 114, 211; 1899, 72–122 (107 Temoncephala); 1905f, 21–24, 2 figs. (in Sesarma gracilipes; New Guinea); 1905h, 402–403.—Plate, —, 187–191.—Pratt, 1900a, 646, 647 (key), 658.—Roewer, 1906, 207, fig. 4.—St.-Remy, 1898, 522, 524–525.—Schuberg, 1895, 180.—Semper, 1872, 304.—Vayssiere, 1892, July 4, 64–65; 23 pp., 1 pl. (on Astacoides madagascariensis); 1894, 16 1892, July 4, 64–65; 23 pp., 1 pl. (on Astacoides madagascariensis); 1894, 16 Mar., 389.—Wacke, 1902, 34 pp.; 1903, 12 May, in 1–116, pls. 1–9, figs. 1–75; 1903, Aug., 2481; 1903, 17 Oct., 17–18; 1904, 17 May, 281; 1905, July, 64.— Weber, 1889, 1-20, 3 pls.

1887: Temnacephale Cosmovici, 1887a, 127 (for Temnocephala).

1888: Temuocephala Mont., 1888a, 50 (for Temnocephala).

1889: Temoncephala Mont., 1889, 107, misprint.

aurantiaca Haswell, 1900, 431, 433, pl. 22, fig. 3 (in Astacopsis sp.).

axenos Mont., 1898, ——; 1899, 83, 84, 101, 102, 103, 104, 112, 114, 120, pl. 3, figs. 9, 10 (host not known; Brazil).—Cerf., 1899a, 448.

bifasciata Haswell.—Mont., 1889, 2.

brevicornis Mont., 1889, 1-4, figs. 1-3 (in Hydromedusa maximiliani, Hydraspis radiolata; Brazil (syn. Pentadion emydum Kroyer MS.); 1892, 110; 1893, 8, 114; 1899, 72–127, 2 pls.; 1899, 72–122, pl. 3, figs. 1–8, 11–18, pl. 4, figs. 19–33 (in Hy. draspis gibba; Brazil; Hydromedusa maximiliani, H. tectifera); 1903, 2.—Brand., 1891d, 21, 24, 25.—Braun, 1890a, 525, 547, 552.—Cerf., 1899a, 448.—Haswell, 1892d, 361; 1893e, 94, 103, 104, 105, 112, 141 (in Hydrom. maxim., Brazil; Hydropsis radiolata); 1900, 433, 434.—St.-Remy, 1898, 528.

cæca Haswell, 1900b, 432, 433, pl. 22, fig. 4 (in Phreatoicopsis n. sp.; Victoria).

cecca Haswell, 1900b, 432, 433, pl. 22, fig. 4 (in Phreatoloopsis n. sp.; Victoria).

chilensis (Moquin-Tandon) E. Bl., 1849, 51–52, pl. 2, fig. 6 ("en las branquias de los cangregos de Chile").—Braun, 1890a, 499, 525, 547, 549; 1891d, 421.—Cerf., 1899a, 448.—Haswell, 1887a, 279, 284 (of Gay); 1893e, 96, 131, 140 (in Æglea; Chile).—Mason, 1875a, 336–337.—Mont., 1888a, 88 (of Blainv.); 1889, 2; 1889q, 125–133, figs. 1–13; 1889r, 9 pp., 13 figs.; 1891, 1892, 128, 129; 1890a, 500–501; 1891g, 44–45; 1892, Oct. 7, 186; 1893, 114; 1899, 72, 73, 76, 78, 79, 80, 81, 83, 84, 85, 93, 101, 102, 107, 108, 109, 111–113 (chilinsis), 114, 120 (in Æglea lævis; Chile) (of Moquin-Tandon); 1899, 125–133, figs. 1–13; 1899, 9 pp.; 1903, 2.—Philippi, 1870, 35–40.—Plate, 1894, 686–687; —, 529–531; 1897, 213.—St.-Remy, 1898, 525.—Wacke, 1903, May 12, in 1–116, 9 pls., 14 figs.; 1903, 17 Oct., 17–18; 1905, July, 64.—Wood, 1875, 336–337.

chilinsis Mont., 1899, 113, for chilensis

chilinsis Mont., 1899, 113, for chilensis.

comes Haswell, 1893e, 96, 98, 100, 123, 134, 138, pl. 13, figs. 15, 16 (in Astacopsis serratus); 1900, 433.—Mont., 1899, 83, 110, 114, 121 (in Ast. serr.; Australia, N. S. Wales).—St.-Remy, 1898, 525.

dendyi Haswell, 1893e, 96, 97, 98, 99, 100, 102, 105, 111, 115, 116, 122, 123, 124, 125, 126, 130, 135–136, 144, pl. 10, figs. 8, 12, 17, 18, pl. 11, figs. 5, 6, pl. 12, figs. 7, 11, pl. 13, figs. 1–11 (in Astacopsis bicarinatus); 1900, 433.—Mont., 1899, 83, 84, 93, 110, 114, 121 (in Ast. bic.; Australia, N. S. Wales, Victoria).— St.-Remy, 1898, 527.

digitata Mont., 1903a, 309 (in Palemonetes argentinus); 1903, 3; 1905g, July, 64.

TEMNOCEPHALA—Continued.

engæi Haswell, 1893e, 97, 124, 139–140, pl 13, fig. 20 (in Engeus fossor; Gippsland).—Mont., 1899, 83, 114, 121 (in Engeus fossor; Australia, Gippsland) (engei).—St.-Remy, 1898, 528.

engei Mont., 1899, 83, 114, 121 (for engæi).

433, pl. 22, fig. 5.—Brand., 1891d, 24.—Braun, 1890a, 409, 422, 465, 525, 547, 549.—Mont., 1888a, 53, 88; 1891, 1892, 129; 1893, 114, 1899, 83, 92, 93, 97, 99, 102, 103, 110, 114, 120 (in Ast. serr.; Australia, N. S. Wales).—St.-Remy, 1898, 525, 527.

fasciatus Haswell, 1887a, 286, for fasciata.

jheringhii Mont., 1899, 79, 83, 84, 85, 109, 111, 112, 114, 120 (for jheringii).

jheringii Haswell, 1893e, 96, 100, 102. 112, 113, 121, 130, 137–138, 140, pl. 13, fig. 19, pl. 15, fig. 2 (in Ampullaria; Brazil).—Cerf., 1899a, 448.—Mont., 1899, 79, 83, 84, 85, 109, 111, 112, 114, 120 (jheringhii) (in Ampullaria sp.; Brazil).—St.-Remy, 1898, 527-528.

madagascariensis Vayssière (1892), 64-65; 1894.—Braun, 1893b, 183.—Cerf., 1899a, 449.—Haswell, 1893e, 141 (in Astacoides madagascariensis).—Mont., 1899, 76, 79, 82, 83, 91, 93, 101, 103, 114, 115–116.—St.-Remy, 1898, 528–529 (on

Ast. mad.; Madagascar).

mexicana Vayssière, 1898, 227-235, 1 pl. (on Cambarus digneti); 1898, 17-25, pl. 11.—Cerř., 1899a, 448.—Mont., 1899, 79, 83, 85, 101, 103, 106, 114, 120 (in Camb. dig.; Mexico.—Pratt, 1900a, 655, 657, 658 (key), fig. 1.

microdactyla Mont., 1903b, 1-3; 1905b, Jan. 31, 22 (in Dilocarcinus septemdentatus; Matto Grosso).

minor Haswell, 1887a, 284, 285, 296, pl. 20, fig. 4, pl. 21, fig. 8, pl. 22, fig. 9 (in Astacopsis bicarinatus; N. S. W.); 1888a, 50 (in Ast. bicar.; N. S. W.); 1893e, 95, 96, 98, 99, 100, 101, 102, 109, 110, 116, 121, 123, 126, 131, 134, 135, 136, 137, 139, 141, 142, 144, pl. 12, figs. 2, 8, 9, pl. 15, fig. 1 (in Asta. bicari.); 1900, 433.—Braun, 1890a, 525, 547, 549.—Mont., 1888a, 53, 88; 1889, 3; 1899, 83, 93, 99, 101, 110, 114, 120 (in Ast. bicar.; Australia, N. S. Wales, Victoria).—St.-Remy, 1898, 526-527, 528, 529.

novæ zealandiæ Haswell, 1888a, 50, for novæ zelandiæ.

 novx-zelandix Haswell, 1887a, 284, 293, 296, 298, pl. 22, figs. 10, 19 (in Paranephrops setosus; New Zealand); 1888a, 50 (in Para. setosus; N. Zea.) (novæ zealandiæ); 1892a, 459; 1893e, 96, 97, 98, 99, 100, 109, 110, 115, 116, 121, 130, 131, 138–139, 145, pl. 10, figs. 4, 7, 11, pl. 11, fig. 1, pl. 13, figs. 17, 18, pl. 14, figs. 2, 3 (in Para, neozealanicus, P. planifrons; New Zealand); 1900, 433.— Braun, 1890a, 525, 547, 550.—Gamb., 1896a, 54, figs. 20, 21.—Maclaren, 1904, 582, 583, 596.—Mont., 1888a, 98; 1889, 3; 1899, 83, 86, 92, 101, 103, 105, 109, 110, 114, 121 (in Para. novæ-zelandicus, P. setosus; New Zealand).—St.-Remy, 1898, 526 (novæ-zealandiæ).—Wacke, 1903, 12 May, 1–116; 1903, 17 Oct., 17–18; 1905, July, 64.—Ziegler, 1905, 40, fig. 4 (after Wacke, 1903).

quadricornis Haswell, 1887a, 284, 296, pl. 20, fig. 3, pl. 22, fig. 8 (in Astacopsis franklinii; Tasmania); 1888a, 50 (in Asta. frank.; Tasmania); 1893e, 96, 98, 100, 101, 102, 109, 110, 120, 121, 124, 131, 136–137, pl. 12, fig. 10, pl. 14, fig. 4 (in Asta. frank.; Tasm.); 1900, 433.—Braun, 1890a, 409, 525, 547, 549.—Mont., 1888a, 10, 53, 88; 1889, 3; 1899, 83, 101, 114, 117, 120 (in Asta. frank.; Australia, Tasmania).—St.-Remy, 1898, 526.

semperi Weber, 1889, -—.—Braun, 1890a, 461, 466, 499, 525, 547, 550; 1890b, 126; 1898a, 1567.—Cerf., 1899a, 449.—Haswell, 1892a, 458; 1893e, 96, 97, 98, 99, 102, 112, 113, 115, 119, 122, 123, 128, 129, 130, 131, 138, 140 (in Philippines); 1900, 433.—Mont., 1891, 1892, 129 (semperii); 1893, 114; 1899, 73, 76, 77, 78, 79, 82, 83, 87, 91, 96, 99, 102, 103, 107, 108, 109, 112, 114, 121 (in Telphusa sp.; Filippine, Sonda).—St.-Remy, 1898, 528.

semperii Mont., 1892, 129, for semperi.

tasmanica Haswell, 1900, 430–431, 433, pl. 22, figs. 1–2 (in Astacopsis tasmanicus). tumbesiana Wacke, 1903, 12 May, 1-116; 1903, 17 Oct., 17-18; 1905, July, 64.

TEMNOCEPHALÆ Mont., 1888a, 19, 88, 96, for Temnocephaleæ.

TEMNOCEPHALEÆ Haswell, 1892a, 457–460; 1893b, 191; 1893e, 93–152 (monograph); 1893f, 153, 154, 155, 157; 1894b, 256; 1900, 430–435, 1 pl. (Temnocephalæ).—Braun, 1890a, 517, 520, 523, 524.—Cerf., 1894, 946, 948; 1898b, 361.—Mont., 1888a, 19, 88, 96 (Temnocephalæ), 7, 9, 10, 11, 13, 15, 16, 17, 18, 19, 20 (Tenmocephalæ), 22, 23, 26, 27, 28, 29, 31, 34, 36, 41 (Temnocepholæ), 42, 47, 52, 55, 59, 64, 67, 86, 88, 98, 108, 110.

TEMNOCEPHALIDÆ Haswell, 1888.—Braun, 1890a, 524; 1893a, 889.—Gamb., 1896, 53–55, 73.—Mont., 1888a, 25, 27; 1892, Oct. 7, 213 (fam. of Eterocotylea; contains Temnocephala); 1899, 116–122.—Pratt, 1900a, 646 (Temnocephalinæ, Activadestymellinm) 647 (keyr). St. Partt, 1898, 522, 524.

Actinodactynellinæ) 647 (key).—St.-Remy, 1898, 522, 524.

TEMNOCEPHALINÆ Mont., 1899, 118, 119.—Pratt, 1900a, 646 (Temnocephala, Craspedella, Dactylocephala), 647 (key).

TEMNOCEPHALOIDEA Mont., 1905, 403.

TEMNOCEPHOLEÆ Mont., 1888a, 41 (for Temnocephaleæ).

TENMOCEPHALEÆ Mont., 1888a, 20 (for Temnocephaleæ).

TERES SEU CYLINDRICA Goze, 1782a, 41, 169, 173–176 (a "Klasse" of Planaria Goze containing two "Gattungen": 1. Gattung, simpliciporo, der Entenplattwurm, Weideeulewurm; 2. Gattung, duplici poro, Iltis-Plattwurm, Dachs-Plattwurm).

TERGESTIA Stoss., 1899, 11, 16 (type probably acanthocephala).—Braun, 1901b, 34.—Looss, 1899b, 580.—Pratt, 1902a, 888, 898.

acanthocephala (Stoss., 1887) Stoss., 1899, 16 (in Belone acus).

laticollis (Rud., 1819) Stoss., 1899, 16 (in Trachurus trachurus).

TESTUCARIA Rud., 1805a, 44, for Festucaria.

TETRACOTILE Erc., 1881e, 48-54; 1882a, 284-290, for Tetracotyle.

TETRACOTYLE Fil. [1854a, 23 (tétracotyle);] 1855, 351; [tld. typica]; 1855b, 11; 1857c, 15-21, 32, pl. 2, figs. 24-31.—Brand., 1888a, 13, 14, 15, 41, 51, 52; 1890a, 571, 574, 575, 577, 578; 1892, 511.—Braun, 1892a, 658, 744, 792, 793, 794, 795; 1893a, 843, 844; 1893b, 187; 1894, 166; 1894k, 680, 681, 682; 1895b, 132, 136; 1900a, 1680.—Claparède, 1858a, 99-105, pl. 8; 1859c, 92-96.—Dies., 1855a, 381, 388; 1858e, 313, 365 (mentions only 1 form, typica), 366.—Erc., 1881e, 48-54; 1882a, 284-290 (Tetracotile).—Gamb., 1896a, 64.—Hausmann, 1897b, 4, 7, 20, 22, 36, pl. 1, figs. 9-10 (sp. in Cobitis barbatula).—Hoyle, 1890, 539 (larva of Holost.).—Jackson, 1888, 652 (non-sexual form of Holost.), 653, 654.—Luehe, 1899, 525.—Mont., 1888a, 71, 91, 92, 94; 1891, 110.—Moul., 1856a, 94, 151, 224-225, 234, 274, pl. 5, fig. 1, pl. 7, fig. 19.—Pag., 1857, 53; 1857, 1, 245-246.—Stiles, 1901, 173, 174.—Tasch., 1878, 248 (of Grube, syn. Pleurocotyle Gerv. & Ben.).—Villot, 1898, 542.

acerınæ cernuæ Claparède (1857).—Dies., 1858e, 367 (syn. of T. echinata) (in

Leuciscus idus, Acerina cernua).

colubri Linst., 1877, 192, pl. 14, fig. 22 (in Coluber natrix, Vipera berus).—Brand., 1888a, 42, 44, 45, 54; 1890a, 572, 574, 581.—Braun, 1892a, 793, 796; 1893a, 870, 901; 1894, 166–167; 1894k, 682.—Muehl., 1898, 16.—Sons., 1893, 185 (in Tropidonotus natrix Wagl.).—Villot, 1898, 542 (larva of Holost. variabile Nitzsch).—Reported also for Pelias berus.

cristallina Sons., 1893, 188, for crystallina.

crystallina (Rud., 1819) Linst., 1877b, 194–197; 1878a (Dist. crystallinum Rud., 1819, ex parte).—Braun, 1892a, 796; 1893a, 871.—Sons., 1893, 188 (cristallina) (in Rana temporaria).

cuticola (Nord., 1832) Kowal., 1902d, 23 (5) (in Scardinius erythrophthalmus); 1904, 25 (10).

cyprini idi Moul., 1856, 233.—Dies., 1858e, 366 (syn. of T. typica).

echmata Dies., 1858e, 367 (new name for Distoma cyprini idi (peritonei), see Duj.; and T. acerinæ cernuæ Claparède; in Leuciscus idus; Remi; Acerina cernua; Geneva).—Braun, 1892a, 796.—Reported also for Idus melanotus.

fatorii Linst., 1876, 1-2, fig. 2 (in Feetorius putorius).—Braun, 1892a, 796.

lenticola (Linst., 1878) Braun, 1892a, 600.

limnæi Pag., 1857, 32–33, pl. 3, figs. 15–18 (t. h. Lymnæus stagnalis).—Dies., 1858e, 366 (syn. of T. typica).

musculicola (Waldenberg, 1860) Braun, 1894, 167.

8588—No. 37—08——24

TETRACOTYLE-Continued.

ovata Linst., 1877b, 192–193, pl. 14, fig. 24.—Braun, 1892a, 600, 797; 1894, 166, 167; 1894k, 681, larva of Holost. variegatum, 682.—Mueh., 1898, 16.—Villot, 1898, 542 (larva of Holost. variegatum).—Reported for Abramis brama, Acerina cernua, Blicca bjærkna.

percæ Mont., 1888a, 7, 30.

percæ fluviatilis Moul., 1856, 230–234, pl. 7, figs. 11–14.—Braun, 1892a, 600, 796;
 1893a, 870.—Linst., 1877b, 192, pl. 14, fig. 23.—Dies., 1858c, 366 (syn. of T. typica).—Piana, 1898, 10 pp.

petromyzontis Brown, 1899a, 489-498, pl. 39, figs. 1-11 (in Ammocœtes).

scombri Grube, 1855a, 140, for ? Octobothrium scombri Nord. of Grube, 1855a (on Scomber scomber).—Tasch., 1878, 575 (to Pleurocotyle); 1879, 248.

soricis Linst., 1877b, 191 (in Sorex vulgaris).—Braun, 1892a, 797.—Reported for Sorex tetragonurus.

typica Dies., 1858e, 366–367 (new name for Dist. tarda, Tetracotyle Fil., T. cyprini idi, T. percæ fluviatilis, T. lymnæi) (reported for Mollusca; in sporotheriis Cercariæ echinatæ (Lymnæi stagnalis), C. armatæ, C. echinatoidis (Paludinæ achatinæ et viviparæ), C. vesiculosæ, Malleoli furcati, Lymnæus stagnalis, Planorbis corneus, Paludina vivipara, Perca fluviatilis, Leuciscus idus, L. dobula, Cyprinus carpio, Anas boschas fera).—Brand., 1888a, 17, 42; 1890a, 571.—Braun, 1892a, 797; 1893a, 844; 1894, 166.—Kowal., 1894, 2.—Leidy, 1891a, 416 (in Lymnæa catascopium, Physa heterostropha).—Linst., 1877, 193; 1894b, 328–332, figs. 1–8.—Mont., 1888a, 71.—Par., 1894, 166.—Reported also for Idus melanotus.

TETRACOTYLIDÆ Mont., 1892, 107, 108.—Braun, 1895b, 174.

TETRACOTYLINÆ Mont., 1892, 107.

TETRAONCHINÆ Mont., 1903, 336 (raised from subf. to fam. rank); 1905, 80.

TETRAONCHUS Dies., 1858e, 314, 379–380 (type probably monenteron by elimination; agrees with first species rule).—Ben. & Hesse, 1864, 121.—Braun, 1889k, 622; 1890a, 412, 413, 417, 465, 468, 511, 517, 518, 523, 542, 544; 1893a, 890.—Gamb., 1896a, 73.—Goto, 1893a, 798.—Hoyle, 1890, 539.—Maclaren, 1904, 574, 598, 599, 600, 601.—Mont., 1888a, 10, 13, 84, 86, 101; 1889, 113–116; 1891, 127–129; 1892, Oct. 7, 213 (gen. of Gyrodactylinæ); 1903, 336 (syns.: Ancyrocephalus, Amphibdella, Dactylodiscus; subf. Gyrodactylinæ); 1905, 79, 80.—Pratt, 1900, 646, 654 (key to), 657, fig. 46.—St.-Remy, 1898, 524, 567.—Stoss., 1898, 17.—Tasch., 1879, 69; 1879, 263 (syns.: Dactylogyrus Wagener, Gyrodactylus Wedl).

benedeni St.-Remy, 1898, 566, 567, for van benedenii.

borealis (Olss., 1893) Mont., 1905, 79.

cruciatus (Wedl, 1857) Dies., 1858e, 381 (in Cobitis fossilis).—Braun, 1890a, 545, 549, 550.—Linst., 1878a.—Mont., 1889, 115, 116 (in Cob. foss.).—Pavesi, 1881, 616.—Tasch., 1879, 264 (syn. Gyrodactylus cruciatus Wedl) (in Cob. foss.).

monenteron (Wagener, 1857) Dies., 1858e, 380 (in Esox lucius).—Braun, 1890a, 520, 545, 549, 550.—Mont., 1889, 115, 116 (in E. luc.).—Olss., 1893, 7.—Par. & Per., 1890, 9.—Pratt, 1900a, 657, fig. 46.—Tasch., 1879, 263 (syns.: Dactylogyrus mon. Wagener, Gyrodactylus cochlea Wedl) (in E. luc.).

torpedinis (Chatin, 1874) Mont., 1890, 193; 1891, 1892, 109, 111; 1905, 79.

unguicolatus Mont., 1888a, 90 to (Dactylogyrus), for unguiculatus.

unguiculatus (Wagener, 1857) Dies., 1858e, 380-381 (in Perca fluviatilis, Lucioperca sandra).—Braun, 1890a, 545, 549, 551.—Hausmann, 1897b, 4, 7, 20, 23 (in Luciop. sand.).—Linst., 1878a.—Mont., 1888a, 90 (unguicolatus) to (Dactylogyrus); 1889, 113, 114, 115, 116 (syn. Ancyrocephalus paradoxus Crep.) (in Luc. san., Perc. fluv.); 1891, 1892, 109, 127, 129, pl. 6, figs. 40, 41.—Staff., 1905, 687 (in Eupomotis gibbosus; Ambloplites rupestris).—Tasch., 1879, 238, to (Dactylogyrus); 1879, 263-264 (syns.: Dact. ung., Gyrodactylus crassiusculus, Ancyrocephalus paradoxus) (in Perca fluv., Luc. sandra).

van benedenii Par. & Per., 1890, 4, 12, 96–97 (8–9) (in Mugil auratus).—Braun, 1890a, 545, 549, 551.—Mont., 1905, 79.—Par., 1896, 2 (v. benedeni).—St.-Remy, 1898, 566, 567 (benedeni).—Stoss., 1898, 17 (in Mugil auratus; Triest).

TETRASTOMA delle Chiaje, 1833, 13 (type renale).—Andral, 1829d, 617.—Braun, 1893a, 884, 891, 892, 894; 1900a, 1681.—Burm., 1856a, 251.—Crep., 1841, 82.-Dav., 1877a, lxxx.—Dies., 1850a, 408, 597 (of Forbes & Goodsir, syn. of Scolex Mueller); 1858e, 314, 367.—Goldb., 1855, 17.—Mont., 1888a, 89.

1850: Tetrastomum Dies., 1850a, 408, for Tetrastoma.

playfairii Forbes & Goodsir, (1839); 1840, 370; 1842, 370; 1846, 160.—Dies., 1850a,
 599 (syn. of Scolex acalepharum Sars).—Mont., 1888a, 94.

renale delle Chiaje, 1833, 13, 116-117, pl. 2, fig. 13 (in Homo; Naples).—Aitken, 1866, 804, 841; 1872, 146, 206; 1874, 58.—Almeida Couto, 1872, 26.—R. Bl., 1888a, 597.—de Bonis, 1882, 180.—Braun, 1895b, 155.—Cobbold, 1866, 7; 1876, 211; 1879b, 36.—Crep., 1841, 82; 1846, 129.—Dav., 1877a, 296–297.—Dies., 1850a, 408 (in Homo; Naples); 1858e, 367.—Dunglison, 1893, 821, 1084.—Hackley, 1886, 519.—Kholodk., 1898, 26.—Leuck., 1863a, 527.—Moniez, 1896, 86, 152, 154.—Mont., 1888a, 89.—Tasch., 1879, 250.—Verrill, 1870, 171.—Weinland, 1859, 282.

TETRATHYRUS Crep., 1851b, 292 (m. obesus), Dithyridium and Piestocystis renamed.—See Cestoda.

1860: Petrathyrus Cobbold, 1860a, 42, misprint.

obesus Crep., 1851b, 292 (syns.: Dithyridium lacertæ, Monost. lacertæ, Piestocystis dithyridium) (in Lacerta agilis).—Dies., 1858e, 329 (syn. of Monost. lacertæ Gurlt, est Piestocystis dithyridium).

THAUMATOCOTYLE Scott, 1904, 278-279 (m. concinna).

concinna T. Scott, 1904, 278-279, pl. 17, fig. 15 (in Trygon pastinaca; Dornoch Firth); 1905, 118.

THECOSOMA Moq.-Tandon, 1860a, 342 (type hæmatobium) (=Schistosoma 1858, renamed because of schistosoma, a term in teratology).—Braun, 1903, 3 ed., 168 (syn. of Schistosomum).—Cobbold, 1879b, 39; 1885a, 498 (syn. of Bilharzia).—Huber, 1896a, 580 (syn. of Bilharzia).—Leuck., 1863a, 617.—Montgomery, 1906, Jan., 18 (Thexosoma); 1906, Feb. 12.—Simon, 1897, 99.—Stiles & Hass., 1898a, 94, 95 (syn. of Schistosoma).—See Schistosoma.

1906: Thexosoma Montgomery, 1906, Jan., 18, for Thecosoma.

 hæmatobium (Bilharz, 1852) Moq.-Tandon, 1860a, 342.—R. Bl., 1888a, 636 (syn. Bilharzia hæm.).—Stiles, 1898a, 58.—Ward, 1895, 253 (to Gynæcophorus; hæmatomuim, misprint); 1895, 328 (in Homo); 1903, 872 (syn. Schistosoma hæm.).

THEXOSOMA Montgomery, 1906, Jan., 18, misprint for Thecosoma.

TISTOMUM Tasch., 1879, 65 (for Tristomum).

TOCOTREMA Looss, 1899b, 585, 586, 619 (tod. lingua) (ὁ τόκος, birth; τὸ τρῆμα, hole, sucker); 1900, 608; 1902m, 833, 835.—Braun, 1901i, 56.—Jægers., 1900, 736; 1901, 981; 1903a, 13, 14.—Luehe, 1900, 557.—Odhn., 1900, 21, 22.

concavum (Crep., 1825) Looss, 1899b, 586.—Jægers., 1903a, 3, 4, 5, 11, 13.—Kowal., 1902d, 26 (8).—Type of Cryptocotyle 1899.

expansum (Crep., 1842) Jægers., 1901b, 979–983, 1 fig.; 1902a, 356–357; 1903a, 1.—
 Looss, 1902m, 706.—Odhn., 1902, 45.—Type of Scaphanocephalus 1903.

jejunum Nicoll, 1907, 248, 257-259 (in Totanus calidris).

lingua (Crep., 1825) Looss, 1899b, 586.—Jægers., 1900, 736; 1901, 979, 981; 1903a, 2, 3, 4, 5, 6, 8, 9, 10, 11, 13.—Kowal., 1902d, 27 (9).—Nicoll, 1906, 514, 519 (in Larus argentatus).—Odhn., 1902, 45.

muehlingi (Jægers., 1898) Looss, 1899b, 585.

TOCOTREMINÆ Jægers., 1903a, 14.

TRACHOPUS Ben., 1858a, 1861a, 11, for Trochopus.

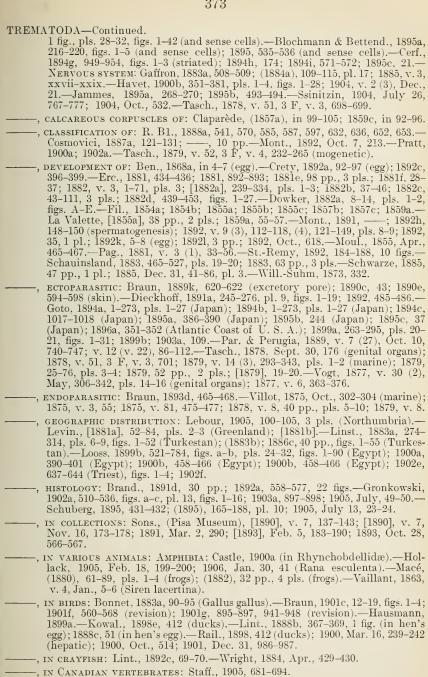
TREMATODA [order of worms] Rud., 1808a, 199; 1809a, 5-6, 20-23, 325-457; 1819a, 82-126, 337-439, 674-688.—Aitken, 1866, 804, 836-842.—Anacker, 1891a, 127; 1892c, 94.—Andral, 1829c, 506, 512; 1829d, 617, 624, 633.—[Assenova, 1899, 82-83.]—Ballowitz, 1900, 442-448.—Bellingham, 1844, 335.—Ben. & Hesse, 1862a; 1864a-c; 1865a (marine).—Bettend., 1897b, 307-358, pls. 28-32, figs. 1-42 (muscles and sense cells).—Biehringer, 1884a, 26 pp. (anat. de-valor), 1884b, 1-28, pl. 1, figs. 1-28; 1889a, 230-235, 1889a, 648-655.—B. Bl velop.); 1884b, 1–28, pl. 1, figs. 1–28; 1888a, 230–235; 1889a, 648–655.—R. Bl., 1886a, 313, 338.—Blochmann & Bettend., 1895a, 216–220, figs. 1–5.—Bos, 1894, 240.—Brand., 1892a, 558–577 (finer structure), pl. 22, 22 figs.—Braun, 1883a, 36, 37; 1889a, 209–400, figs. 1–5, pls. 6–8; 1889k, 620–622; 1890a, 401–560, pls. 9-17; 1890c, 43 (position of excretory pore); 1892a, 561-816, pls. 18-31, figs. 1-23; 1893a, 817-926, pls. 32-34, figs. 24-36; 1895b, 5, 6, 120; 1896b, 7 pp.,

TREMATODA -- Continued.

1 pl., 8 figs.; 1897d, 109–111; 1901d, 258–260.—Bremser, 1824, 131–132.— Bronguiart, (1880a).—Buetschli, 1879a, 588–589, 1 fig.—Burm., 1837a, 528; 1856a, 205, 239–252.—Carus, 1863, 476–477.—Cerf., 190— -.-Cobbold. 1862a, 24–33, 1 pl., figs. 1–7; 1879b, 4, 14–56.—Cohn, 1902d, 712–718, figs, 1–9; 1902e, 842–843; 1902h, 877–882, 5 figs.; 1903, Apr. 28, 223; 1903, May 30, 34–42; 1903, June 18, 155; 1904, May 17, 278; 1905, July, 56–57; 1905, July, 63.—Cosmovici, 1887a, 121–131.—Crep., 1825, 80–81; 1837a, 309; 1839a, 277.— Deslongchamps, 1824fff, 743.—Dies., 1850a, 287-288 (subo. of Myzelmintha), 304–414, 472–473; 1855c, 59–70, pls. 1–3 (19 species); 1855d, 163; 1856c; 1858e, 307–390 (type of Myzelmintha aprocta).—Dunglison, 1893a, 1174.—Eichhorst, 1901a, 301.—Eiss, 1838a, 20.—Fischer, 1840, 156.—Gerv., 1873, 51–55.—Giard, 1903, 1225–1226 (in pearl formation).—Goldb., [1855a], 16.—Graeffe, 1860a, 47 (Tremadota).—Gurlt, 1831, 369.—Hahn & Lefèvre, 1884a, 515–549.—Hoyle, 1888a, 535–540, figs. 1-4; (1888b); 1890, 49, 54, 535–540.—Huber, 1894a, 283–287 (bibliography, clinical); 1896, 502, 574.—Ijima, 1889, 97.—Jackson, 1888, 642, 643, 653, 654, 658.—Johnstone, 1907, 170, 177-192.—Joy, 1835a, 504.—Kerbert, 1878a, 271–273; 1881a, 529–578, pls. 26–27, figs. 1–21.—Kholodk., 1898, 2, 23; 1899a, 146–153, 158.—Kolenati, 1857, 11.—Kuech., 1852, 149.—Lampert, 1898a, lxxxii-lxxxiii.—Lankester, 1882b, 227-331, 2 figs.—Lawson, 1861a, 216-223 (affinities); 1861b.—Leuck., 1830a, 612; 1842, 5; 1863a, 448; 1879, 123; 1886, 93, (affinities); 1861b.—Leuck., 1830a, 612; 1842, 5; 1863a, 448; 1879, 123; 1886, 93, 104.—Linst., 1873i, 51–55; (1877a), 13–14; 1879b, 317; 1883b, in 623–663; [1883b].—Loos, 1892a, 72.—Luche, 1901, 488.—Maclaren, 1904, 573–618, 3 pls., figs. 1–33 (2 species); 1905, Jan. 31, 20–21; 1905, June, 317.—Manson, 1901, 538–544.—Mayer, 1841, 4, 23, 25.—Milner, 1858, 9.—Mlinarich, 1832, 13.—Mol., 1858, 127; 1861, 191.—Mont., 1888, 83, 84; 1893, 427.—Much., 1896, 588–590; 1896; 1897, 243–279, pls. 16–19, figs. 1–17.—Nicoll, 1906, v. 17, 513–527, 2 pls.; 1906, Aug., 445.—Nitzsch, 1819, 397.—Nord., 1840, 596.—Oppenheim, 1900, 181.—Pag., 1857, 56 pp. (larvæ); 1859, 42–46; 1889a, 1–208.—Poir., (1885), 465–624, pls. 23–34; 1885; ——, 25–29.—Pratt, 1900a, 645–662, figs. 1–50 (monogenetic, keys); 1900b; 1900, Sept. 7, 371–372; 1902a, 887–910, 953–979, figs. 1–130 (digenetic, keys); 1902; 1905, July, 63.—Ramsey, 1879.—Rawitz, 1898, v. 3, 1409–1410.—Roger, 1901, 94.—Rolleston, 1870, 140, 141, 142, 143.—Schneider, 1866, 13, 325, 337.—Scott, T., 1901, 141.—Sieb., 1850, 668.—Sons., 1893, May 16, 496, 500; 1893, Dec. 2, 742; 1894, Oct. 26, 756.—Staff., 1900, v. 13 (5), 399–414, pl. 26, figs. 1–6.—Steenstrup, 1860, v. 1 (1–6), 112– Sons., 1893, May 16, 496, 500; 1893, Dec. 2, 742; 1894, Oct. 26, 756.—Staff., 1900, v. 13 (5), 399-414, pl. 26, figs. 1-6.—Steenstrup, 1860, v. 1 (1-6), 112-113.—Stiles, 1896, 206; 1898a, 21, 22, 27.—Tasch., 1879, 232, 233, 234.—Tschudi, 1837, 19.—Tyson, 1903, 3 ed., 1180-1181.—Vaillant, 1879, 108 pp., 55 figs.—Verrill, 1870, 165-166, 173, 175, 176, 177, 178, 179, 213.—Villot, 1882, 505-508.—Wagener, 1883, 120-122.—Wallenstedt, 1847, 6.—Ward, 1895, 238; 1903, 860-873, figs. 4772-4795; 1905, v. 12 (1-2), Jan., 24.—Weinland, 1859, 280.—Wernicke, 1892, v. 15 (6), June, 337-347, 1 pl., figs. 1-22.—Will.-Suhm, 1870, 29 pp.. pls. 11-13; 1871, v. 21 (2), June 15, 175-203, pls. 11-13; 1873, v. 2, 51-55.—Zeller, 1872, 1-24; 1872, 168-180; 1873, v. 2, 51-55.—Ziegler, 1895, 549-552, figs. 420-425.—Zuern, 1882, 113, 202.

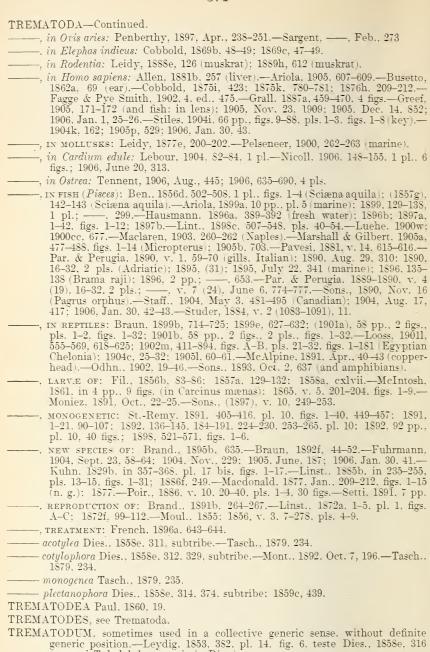
—, ANATOMY OF: Biehringer, 1884a, 26 pp. (and development); 1884b, 1-28 (and development); 1888a, 230-235 (and development); 1889a, 648-655 (and development).—Goto, 1891c, 103-104.—Looss, 1893a, 10-34; 1893c, 637.—Macé, 1881, Feb. 21, 420-421; 1884, Apr., 354-355 (segmental organ).—Mont., 1888, 130 pp.; 1888, July 19, 120-122.—Pachinger, 1888, 18 pp., pls. 2 (and physiology).—Walter, 1858, 269-297, pls. 11-13 (and histology); 1893, Oct., 636-

development); 1888a, 230–235 (and development); 1889a, 648–655 (and development).—Goto, 1891c, 103–104.—Looss, 1893a, 10–34; 1893c, 637.—Macé, 1881, Feb. 21, 420–421; 1884, Apr., 354–355 (segmental organ).—Mont., 1888, 130 pp.; 1888, July 19, 120–122.—Pachinger, 1888, 18 pp., pls. 2 (and physiology).—Walter, 1858, 269–297, pls. 11–13 (and histology); 1893, Oct., 636–637.—Wedl, 1857, v. 5 (26), 241–278, pls. 1-4; 1858, 242.—CIRCULATORY APPARATUS (see also excretory): Ben., 1852a, 573–589, figs. 1–2; 1852b, 23–33; 1852c, Sept. 22, 305.—Villot, 1876, v. 82 (23), June 5, 1344–1346; 1882, v. 5 (121), Sept. 25, 505–508.—EPITHELIUM: Blochmann, (1896a), 16 pp., 2 pls.; 1896b, 821–823.—Hein, 1904, 546–585, 3 pls.; 1904, Dec., 655; 1905, Jan. 31, 17–18; 1905, 350–352.—Maclaren, 1903, June 15, 516–524, figs. 1–6; 1904, May 17, 279–280.—Excretory system (see also circulatory): Ben., 1882b, 14–18, 2 figs. (and body cavity).—Buetschli, 1879a, 588–589, 1 fig.—Bugge, 1902, 177–234, pls. 21–24; 1905, (July 3), 25–26.—Fraip., 1880a, 397–402; 1880b, 106–107, 265–270; 1880c, 415–456, 1 fig., pls. 18–19; (1881a), vii–x, xxii–xxiv; 1881b, 1–40, pls. 1–2, figs. 1–2; 1881c, 602–604; 1883a, xxxi–xlii.—Kampmann, 1894a, 443–462, pls. 19–20, figs. 1–23; 1894b; 1895a, 843–844.—GENITAL SYSTEM: Darr.—Linst., 1873e, 95–108, pl. 5, figs. 1–6; 1873f, 231–232.—Looss, 1893b, 808–819, figs. 1–4.—Stieda, (1871), (1) 7, 31–40.—Muscullature: Bettend., 1897a, 54 pp., 1 fig., 5 pls., figs. 1–42 (and sense cells); 1897b, 307–358,



-, in Bos taurus: Giard & Billet, 1892a, 613-615 (Tonkin); 1893a, 245 (Tonkin). Hedley, 1881a, 374–375 (lungs); 1881b, 399–400; 1881c. 27–28.—Littlewood, 1887a, 546.—Stiles, 1898a, 161 pp., 124 figs. (and sheep and swine). —, in Equus caballus: Adams, 1876, 764-765 (liver).

-, IN MAMMALS: Braun, 1901e, 311-348. -, in Castor fiber: Bojanus, 1817b, 270–277. -, in Chiroptera: Braun, (1900b), 217-236, pl. 10.



(syn. of Tylodelphys craniaria Dies.).

cancri locustæ (Rud., 1810) Linst., 1878a, 315 (in Palæmon locusta).

ranæ Valentin, 1843, 91, teste Dies., 1850a, 472.

salmonis lavareti Linst., 1878a, 266 (in Coregonus wartmanni; Greenland), based on Fabricius. ——, 269.

talpæ cœcæ Dies., 1850a, 472 (in Talpa cœca).—Linst., 1878a, 18 (in Talpa cœca).
wachniæ (Rud., 1819) Linst., 1878a, 238 (in Gadus wachnia), based on Tilesius, 1810, 472.

TREMATOIDEA delle Chiaje, 1825a, 12.—Almeida Couto, 1872, 4, 7, 19.—Eichwald, 1829a, 247.—Guenther, 1858, 203-204.—Kuech., 1855, 7, 179.—Swart, 1862,

TRICHODA Mueller.—Nitzsch. 1827. 69, contains Cerc. setifera.

TRICOTYLEA Dies., 1850a. 290, 425, subtribe of Polycotylea to contain Nitzschia. Phylline, Udonella, Encotyllabe, Trochopus, Tristomum; 1858e, 313, 362.— Brand., 1888a, 15.—Braun, 1890a, 515.—Goldb., 1855, 19.—Mont., 1888a, 84.—Tasch., 1878, 564, 565; 1879, 233.

TRIPOS Bory St. Vincent, 1823b, 356, genus of Cercariées.

TRISSICHYA Cosmovici. 1887a contains Epibdella. Tristomum, Udonella.— Mont., 1888a, 84.

TRISTOMA Cuv., 1817, 42-43 m. coccineum .—Baer, 1826a, 125, 126; 1827, 675,— Bell, 1891a, 534-545 (n. sp. from Histiophorus brevirostris).—Ben., 1858a, 1861a, 11, 38, 168.—Ben. & Hesse, 1864, 61, 65, 66, 76-77.—Bettend., 1897a, 17; 1897, 321.—Bl., 1847, 321.—Brand., 1894a, 308.—Braun, 1889k, 621; 1890a, 411, 412, 415, 420, 427, 430, 450, 451, 453, 454, 455, 461, 463, 464, 465, 468, 471, 473, 474, 479, 481, 482, 483, 484, 490, 491, 511, 515, 516, 517, 518, 519, 523, 526, 527, 528; 1891d, 422; 1893a, 889.—Bremser, 1824, 134.—Burm., 519, 523, 526, 527, 528; 1891d, 422; 1893a, 889.—Bremser, 1824, 134.—Burm., 1837, 530; 1856a, 251.—Carus, 1863, 477.—Ceri., 1899a, 422, 435.—Cobbold, 1877, 238; 1879b, 5, 41.—Cosmovici, 1887a, 128.—Crep., 1837, 323; 1839, 289, 291.—Deslongchamps, 1824mmm, 754; 1830h, 392.—Dies., 1835, 10; 1836a, 1-16, pl. 1 (monograph); 1836b; 1838, 77-89; 1839a, 234; 1850a, 290, 428-431 (syns, Capsala Bosc, Phylline Oken), 425 (of Nitzsch, syn. of Nitzschia), 426 (of Rathke, syn. of Phylline), 427 (of Nord., syn. of Encotyllabe Dies.), 428 (of Dies., syn. of Trochopus); 1858e, 313, 365 (syns, Capsala, Phylline).—Duj., 1845a, 321-322.—Eichwald, 1829a, 249.—Fischer, 1840a, 158.—Gamb., 1896a, 73.—Goldb, 1855a, 20.—Goto, 1891a, 161, 184; 1893a, 798, 799; 1894a 1896a. 73.—Goldb., 1855a. 20.—Goto. 1891a. 161. 184: 1893a. 798. 799: 1894a, 237.—Haswell. 1887a. 286. 291: 1892a. 459; 1892b, 150: 1893e. 114. 118. 122. 144.—Hémont. 1827. 9.—L'Herminier. 1826. 10.—Hoyle. 1890. 535. 539 (all parasitic on fishes).—Ijima. 1884a. 638.—Jackson. 1888. 642. 643. 644. 646. 647. 648. 653.—Johnston. 1865a. 33.—Jov. 1835a. 504.—Kerbert. 1881a. 544. 554, 572.—Leuck., 1863a, 457, 461, 462, 463, 464, 465.—Looss, 1892, 72; 1894a. 145, 146.—Massa, 1906, 43, 45, 48; 1906, 51 (of Tasch., syn. of Trochopus Dies.).—Mont., 1888a, 10, 11, 13, 23, 26, 30, 32, 37, 42, 45, 50, 53, 54, 56, 57, 58. 59. 60, 65. 83. 84. 86, 87. 97: 1889, 117, 118: 1891, 1892, 101. 103, 104, 105, 106. 58, 59, 60, 65, 83, 84, 86, 87, 97; 1889, 117, 118; 1891, 1892, 101, 103, 104, 105, 106, 107, 108, 111, 116, 119, 120, 123; 1892, Oct. 7, 180, 213 [gen. of Tristominæ]; 1893, 75; 1899, 96, 97, 98, 99, 100, 101; 1903, 335 [fam. Tristominæ]; —Moul., 1856a, 10.—Nitzsch, 1826, 150 [also Tristominæ].—Nord., 1832a, 60.—Par. & Perugia, 1890, 13.—Pratt, 1900, 646, 649, 655, 658, fig. 9.—Rud., 1819a, 123, 427–428, 584.—St. Remy, 1891, 1072–1074 [genital organs]; 1892, 702; 1898, 534.—Schneider, 1866, 334.—Scott, T., 1901, 144.—Setti, 1899, 117–125, figs. 1–3, 1899; 71–84 [revision]; 1899, 7 pp.; 1899, 5 pp.; 1899, 9 pp., 3 figs.—Stoss., 1898, 5.—Tasch., 1878, 176; 1878, 563, 564, 565, 566 [syns, Capsala, Phylline, Hirudo, sp. Abildg., Nitzschia Baer]; 1879, 30, 32, 33, 34, 36, 37, 41, 43, 44, 50, 57, 58, 60, 62, 64, 65 [Tistomum], 68, 70, 71; 1879, 233, 236, 238.—Wallenstedt, 1847, 7.—Ziegler, 1883, 552.

1826: Tristomum Nitzsch., 1826, 150, for Tristoma.

1879: Tistomum Tasch., 1879, 65, misprint.

1888: Tristomun Mont., 1888a, 28, misprint.

1899: Tristonum Setti, 1899, 84, misprint.

aculeatum Couch, ——.—St.-Remy, 1898, 535 (=T. molæ).—Setti, 1898, 311 (syn, of rudolphianum).—Tasch., 1878, 570.

biparasiticum Goto. 1894a. 251-253 (in Parapetalus, Thynnus albacora: Misaki, Japan: 1899. 272.—Mont., 1899. 86, 89.—Setti, 1898. 311: 1899. 80: 1899, 124.—St.-Remy. 1898. 539.

blanchardi Tasch.. 1878. 567, for blanchardii.

blanchardii Dies., 1850a. 430 (in Squalus sp.: New Zealand) (T. squali Bl., renamed).—Mont., 1891, 1892, 101, 103, 123.—Setti, 1898, 311 (blanchardi).—Tasch., 1878, 564, 567 (blanchardi).

 cephala Risso, 1826, 262–263 (t. h. Tetraodon luna: Europe .—Dies., 1850a, 431.—
 Kroyer, 1852–53a, 745 (cephalo) in Orthagoriscus mola).—Linst., 1878a.—
 Setti, 1898, 311.—St.-Remy, 1898, 535 (=T. molæ).—Tasch., 1878, 564 (cephalo).

TRISTOMA—Continued.

cephalo Kroyer, 1852-53a, 745.—Tasch., 1878, 564.—For cephala.

coccineum Cuvier, 1817, 42-43, pl. 15, fig. 10, pl. 36 tis, figs. 1-3 (in la mole, le xiphias, etc.).—Bell, 1891a, 534, 535.—Ben., 1858a, 1861a, 37, 189.—Ben. & Hesse, 1864, 61, 76, 77 (coccinea).—Blainv., 1828a, 569 (to Capsala).—E. Bl., 1847, 322–325, pl. 14, fig. 2.—Braun, 1890a, 407, 420, 427, 458, 459, 487, 529, 547, 552; 1891d, 422.—Bremser, 1824, 134; 1824c, pl. 10, figs. 12-13.—Burm., 1837a, 530.—Costa, 1846a, 54.—Crep., 1837, 323; 1839, 289.—Dies., 1836a, 11–12, pl. 1, figs. 1–13; 1850a, 429 (of Cuv., includes T. coccineum of Cuv., Guérin, Gray, Costa, 1846, Bl.; Capsala coccinea Blainv., Moquin-Tandon; T. integrum Dies.) (on Xiphias gladius); 1850a, 429 (of Rud., 1819a, 123, 427, pl. 1, figs. 7-8, renamed T. rudolphianum; includes also T. coccineum of Brem., Nitzsch, Dies., Duj., Yarrell, Cuv., pl. 36 bis, figs. 2-3; Phylline or Brem., Nitzsch, Dies., Dul., Yarrell, Cuv., pl. 36 bis, figs. 2–3; Phylline coccinea Schweigger, Capsala coccinea Blainv., Nord.; Trist. molæ Bl.) (in Orthagoriscus mola).—Duj., 1845a, 322–323.—Goto, 1899, 272.—Gray, —, pl. 9, fig. 10.—Johnston, 1865a, 33.—Juel, 1889, 37.—Kerbert, 1881a, 533.—Kælliker, 1849, 21, 26.—Kroyer, 1838–40a, 597 (in Xiphias gladius).—Lang, 1880.—Leuck., 1863a, 450, 455.—Looss, 1885b, 5, 10.—Linst., 1903, 279.—Lint., 1898, 509–510, pl. 40, fig. 9; 1900, 278; 1901, 414, 448.—Lænnberg, 1891, 76.—Mont., 1888a, 13, 28, 52; 1891, 1892, 101, 102, 103, 110, 113, 114, 116, 123, 124; 1893, 4–5.—Nitzsch, 1826, 150.—Nord., 1840, 602 (syn. of Capsala sanguinea).—Par., 1887.—Par. & Per., 1889; 1890, 5.—Pratt, 1900, 655, fig. 9; 657, 658.—Risso, 1826, 262.—Rud., 1819a, 123, 428–430.—St.-Remv, 1898, 534–536. 658.—Risso, 1826, 262.—Rud., 1819a, 123, 428-430.—St.-Remy, 1898, 534, 536, 538.—Scott, T., 1901, 144.—Setti, 1898, 311, 313; 1899, 71, 72, 73, 74, 75, 76, 77, 78, 83; 1899, 124.—Sons., 1890, 173 (in Xiphias gladius, Tetrapturus belone, Orthagoriscus mola; Sicilia, Nizza, Napoli).—Staff., 1904, May 3, 482 (on Xiphias gladius; Canada).—Stoss., 1898, 5.—Tasch., 1878, 564, 567, 569, 570, 571, 572 (syn. Capsala coccinea) (in Orthag. mola, Xiphias gladius); 1878, 176; 1879, 59.—Wagener, 1857, 72.—Yarrell, ——, 468.—Ziegler, 1883, 545.

cornutum Verrill, 1875, 40, on Tetrapturus albidus; Block Island.—Pratt, 1900, 658.—St.-Remy, 1898, 536 (on Tet. alb.; North America).—Setti, 1898, 311,

312; 1899, 125.

elegans (Baer, 1826) Ben. & Hesse, 1864, 64, 77, of the sturgeon.

elongatum Nitzsch, 1826, 150–151 (on Stör).—Ben., 1858a, 1861a, 20.—Ben. & Hesse, 1864, 67 (syn. of Nitzschia elegans); 1864, 70.—Braun, 1889, 758; 1889h, 19 Aug., 433–434; 1889k, 621, 622 (syn. Nitzschia elegans Baer).—Crep., 1846, 149.—Dies., 1835, 12–14; 1836a, 12; 1850a, 426 (syn. of Nitzschia elegans).—Duj., 1845a, 323.—Kælliker, 1849, 21.—Nord., 1840, 602 (to Capsala).—Tasch., 1878, 563, 564, 567–568 (in Accipenser sturio); 1879, 56.—Reported also for Acipenser guldenstædtii.

excavatum Nord. in Dies., 1850a, 428 (syn. of Encotyllabe nordmanni).

foliaceum Goto, 1894a, 248 (host unknown).—Mont., 1899, 86.—St.-Remy, 1898, 538.—Setti, 1898, 311, 313; 1899, 119, 124; 1899, 79, 80.

hamatum Rathke, 1843, 238–242, pl. 12, figs. 9-11 (syns. Hirudo hippoglossi Mueller, Phylline hippoglossi Oken, Ertopdella hippoglossi Blainv.; in Pleuronectes hippoglossus: Europe).—Ben., 1858a, 1861a, 19, 21 (syn. of Epibdella hippoglossi).—Ben. & Hesse, 1864, 68, 69 (syn. of Epibdella).—Dies., 1850a, 427 (syn. of Phylline hippoglossi).—Johnston, 1865a, 32.—Kælliker, 1849, 21.—Odhn., 1905, 370 (syn. of Epibd. hipp.).—Tasch., 1879, 568 (syn. of T. hippoglossi).—Reported for Pleuronectes hippoglossus.

hippoglossi Oken, see Tasch., 1878, 568; 1879, 59, to (Epibdella).

histiophori Bell, 1891a, 534–535 (in Histiophorus brevirostris).—Goto, 1894a, 238; 1899a, 271 (syn. of T. leve Verrill).—Mont., 1899, 86.—St.-Remy, 1898, 535 (=T. læve).—Setti, 1898, 311, 312, 313; 1899, 79, 83.

integrum Dies., 1850a, 429, MS. (syn. of T. coccineum).—Setti, 1898, 311.

interruptum Mont., 1891, 1892g, 101, 105, 116, 122, 123, pl. 12, figs. 17-19 (in Thynnus brachypterus; Naples?); 1899, 86, 97.—Braun, 1891d, 422.—Goto, 1894a, 238.—St.-Remy, 1898, 535.—Setti, 1898, 311, 312; 1899, 117-119, 124, fig. 1; 1899, 80, 81, 84.—Reported also for Thynnus thynnus.

læve Verrill, 1875, 40, in Tetrapturus albidus; Block Island.—Goto, 1899a, 271–273 (leve, syns. histiophori and ovale), pl. 20, figs. 10–12.—Linst., 1906, 176 (=T. ovale Goto).—Lint., 1898, 509, pl. 40, figs. 7–8; 1901, 414, 445.—Mont., 1899, 86; 1899, 108.—Pratt, 1900, 658.—St.-Remy, 1898, 535–536, 539 (on Tet. alb.; North America).—Setti, 1898, 311; 1899, 79, 83; 1899, 124.—Reported for Gymnosarda pelamys, Tetrapturus albidus, Histiophorus orientalis, Cymbrium sp.

TRISTOMA-Continued.

læve var. armata Goto, 1899a, 273 (læve of Verrill, and histiophori of Bell).

læve var. inermis Goto, 1899a, 273 (T. ovale renamed).

leve Goto, 1899a, 291, for læve.

levinsenii Setti, 1898, 311, 312; 1899, 120–121, 124, fig. 2, for levinsenii.

levinsenii Mont., 1891, 101, 122, 123, pl. 6, fig. 21 (in Thynnus sp.).—Braun, 1891d,
422.—Goto, 1894a, 238.—Setti, 1898, 311, 312; 1899, 120–121, 124, fig. 2
(levinseni).

maculatum Rud., 1819a, 123, 430—431, pl. 1, figs. 9–10 (includes Capsala martinieri Bosc, Phylline diodontis Oken) (in Diodon).—Ben. & Hesse, 1864, 76.—Braun, 1890a, 529, 547, 550 (in Diodon sp.; California).—Dies., 1835, 10–11; 1836a, 10; 1850a, 430 (syns. Caps. mart., Phylline diodontis, Caps. maculata).—Duj., 1845a, 322.—Eichwald, 1829a, 249.—Mont., 1891, 101, 103, 123.—Nitzsch, 1826, 150.—Nord., 1840, 602 (to Caps.).—Odhn., 1905, 371.—Pratt, 1900, 658.—Setti, 1898, 311; 1899, 84; 1899, 125.—Tasch., 1878, 564, 567 (in Diodon).

megacotyle Linst., 1906, 176, pl. 1, figs. 19-20 (in Histiophorus sp.; Beruwala).

molæ Bl., E. 1847a, 326–327 (t. h. Orthagoriscus mola).—Ben. & Hesse, 1864, 66, 76, 77–79.—Bettend., 1897a, 34, 35; 1897, 338, 339.—Brand., 1898a, (12, 14, 18), 204, 206, 210.—Braun, 1883a, 43; 1889k, 621; 1890a, 419, 423, 424, 425, 427, 428, 429, 431, 432, 445, 449, 450, 451, 455, 456, 458, 460, 465, 483, 512, 529, 547, 551 (in Orthag. mola; Atlantic Ocean (England), Mittelmeer (Nizza, Naples, Palmero, Triest)); 1892a, 617.—Cerf., 1898b, 345.—Dies., 1850a, 429 (of Bl., syn. of T. rudolphianum).—Fischer, 1883a, 22.—Goto, 1891a, 161; 1899, 272.—Haswell, 1892a, 459; 1893e, 121, 144, 145.—Heath, 1902, 127, 128, 131, 132.—Hoyle, 1890, 537.—Jackson, 1888, 644, 650.—Johnston, 1865a, 33.—Kath., 1894a, 138.—Knoch, 1894, 5, 11, 17.—Lint., 1901, 408, 414, 466.—Lœnnberg, 1891, 76.—Looss, 1885b, 12; 1894a, 146, 154.—Maclaren, 1904, 585, 589, 590, 608.—Mont., 1888a, 28 (Tristomun); 1888a, 7, 10, 13, 16, 17, 19, 23, 28, 29, 33, 50, 51, 52; 1890, 418; 1891, 101, 104, 107, 116, 118, 119, 120, 122, 123, pl. 5, fig. 4; 1892, Oct. 7, 172; 1893, 5–7; 1893, 77; 1899, 97, 98.—Par. & Perugia, 1889, 741, fig. 2 (in Mola aspera); 1890, 5.—Par., 1894, 549, 677, 680.—Poir., 1885, 138, 139, 147.—Pratt, 1900a, 658.—St.-Remy, 1898, 534, 535, 537.—Scott, 1901, pl. 8).—Setti, 1898, 309, 311; 1899, 124; 1899, 80, 81, 83.—Sons., 1890, 173 (in Orthag. mola); 1891, 265.—Staff., 1904, May 3, 482 (on Mola mola; Canada).—Stoss., 1891, 111; 1896, 191; 1898, 5-6.—Tasch., 1878, 564, 567; 1879, 232–265; 1880, 12 Jan., 17–18.

nordmanni (Dies., 1850) Tasch., 1878, 568 (syn. Encotyllabe nordmanni).

nozawæ Goto, 1894a, 249–251 (in Thynnus sibi; Osatsube (Hokkaido)).—Mont.,
 1899, 86.—Setti, 1898, 311; 1899, 80, 84 (nozawai); 1899, 122, 123, 124.—
 St.-Remy, 1898, 538–539 (nozawai).

nozawai St.-Remy, 1898, 538, for nozawæ.

onchidiocotyle Setti, 1899, 121–123, fig. 3 (in "tonno" at Portoferraio).—Par., 1899, 3; 1902, 3 (in Thynnus thynnus; Elba).

ovale Goto, 1894a, 241–244 (in Histiophorus orientalis, H. sp.; Misaki); 1899a, 271 (syn. of T. læve).—Mont., 1899, 86; 1899, 108.—St.-Remy, 1898, 535 (=T. læve).—Setti, 1898, 311; 1899, 79, 83.

ovata (Goto, 1894) Heath, 1902a, 132, apparently for Epibdella ovata.

pagelli (Ben. & Hesse, 1863) Tasch., 1878, 569 (syn. Encotyllabe pagelli).

papillatum Ben., 1858a, 1861a, 38 (refers to Kælliker, 1849, 21, pl. 2).—Mont, 1893, 12.

papillorum Juel, 1889, 14, misprint.

papillosum Dies., 1836a, 313-316, pl. 17, figs. 13-16 (t. h. Xiphias gladius); 1850a, 430-431 (syn. Capsala papillosa) (in Orthagoriscus mola; Naples, Tetrapturus belone, Messina; Xiphias gladius; Panormi).—Ben., 1858a, 1861a, 169, 175, 176.—Ben. & Hesse, 1864, 66, 77.—Brand., 1891d, 14, 20.—Braun, 1883a, 45; 1889k, 621, 622; 1890a, 409, 419, 420, 424, 427, 430, 437, 438, 441, 444, 445, 456, 458, 487, 529, 547, 552; 1890b, 125; 1891d, 422; 1893b, 177.—Crety, 1893a, 384.—Duj., 1845a, 323.—Goto, 1894a, 237.—Grube, —, 49.—Juel, 1889, 37.—Kœlliker, 1846, 469; 1847a, 469; 1849b, 21-27, pl. 2, figs. 1-4 (anatomy).—Kerbert, 1881a, 533.—Kroyer, 1852-53a, 745 (in Orthag. mola).—Linst., 1903, 280.—Looss, 1885b, 5, 10.—Mont., 1888a, 7, 16; 1889, 118; 1891,

TRISTOMA—Continued.

99, 101, 102, 103, 105, 110, 113, 114, 116, 119, 123, 124, pl. 5, figs. 1-3; 1893, 79.—Nord., 1840, 602 (to Capsala).—Par., 1889, 2-3; 1894, 543, 680, 698; 1902, 2 (in Xiphias gladius; Elba).—Par. & Per., 1890, 5.—Scott, T., 1901, 144.—Setti, 1898, 311, 312; 1899, 71, 72, 73, 74, 75, 76, 77, 83; 1899, 124.—Shipley, 1898, 354.—Sons., 1890, 173; 1891, 265 (in Orthag. mola).—Stoss., 1885, 162; 1898, 6.—Tasch., 1878, 701; 1878, 564, 567, 569, 570, 572 (syn. Caps. pap.); 1878, 176; 1879, 57, 59.—Wagener, 1857, 72.—Ziegler, 1883, 545.—Reported also for Carcharias glaucus.

pelamydis Tasch., 1878, 176; 1878, 569, 570 (in Pelamys sarda; Naples).—Braun, 1890a, 418, 487, 494, 497, 499, 519, 529, 547, 551.—Mont., 1888a, 86, 87; 1889, 117; 1891, 101, 105, 116, 123, pl. 6, fig. 20; 1899, 97.—Par., 1894, 680; 1902, 2 (in P. sarda; Elba).—Par. & Per., 1890, 5.—Setti, 1898, 311; 1899, 80, 81, 82; 1899, 124.—Sons., 1890, 173 (in P. sarda; Pisa).—Stoss., 1898, 6.

perugiai Setti, 1898, 7 pp., 1 fig. (on Tetrapturus belone Raf.; Spezia); 1898, 308-313, 1 fig.; 1899, 77, 79; 1899, 124.—Mont., 1899, 86.—St.-Remy, 1898, 539—

pini (Ben. & Hesse, 1863) Tasch., 1878, 568.—Massa, 1906, 56 (to Trochopus). rhombi (Ben. & Hesse, 1863) Tasch., 1878, 568.—Massa, 1906, 58 (to Trochopus).

rotundum Goto, 1894a, 245-248, pl. 24, figs. 6-9 (in Xiphias gladius; Misaki). St.-Remy, 1898, 537–538.—Setti, 1898, 311, 313; 1899, 77, 78, 83 (syn. of T.

rudolphianum Dies., 1850a, 429 (t. h. Orthagoriscus mola; Anglia) (new name for Capsala sanguinea Blainv., T. molæ Blanchard, 1847, T. coccineum of Rud., Phylline coccinea Schweigger).—Johnston, 1865a, 33.—Kroyer, 1838–40a, 597; 1852–53a, 745 (in Orthag. mola).—Lint., 1898, 510; 1900, 281; 1901, 408, 414, 466.—Setti, 1898, 311 (syns. aculeatum Couch, cephala Risso, molæ Bl.); 1899, 84.—Stoss., 1898, 5.—Tasch., 1878, 564, 567.—Reported also for Mola mola.

scianæ Ben., Tasch., 1878, 568.

sinuatum Goto, 1894a, 239–241 (in Histiophorus sp.; Misaki); 1899a, 272.—Mont., 1899, 89; 1899, 108 (spelled also sinuatuun, 109).—St.-Remy, 1898, 536–537, 539.—Setti, 1898, 309, 311; 1899, 124.

sinuatuun Mont., 1899, 109, misprint.

soleæ (Ben. & Hesse, 1863) Tasch., 1878, 568; 1879, 59 to (Phylonella).

squali E. Bl., 1847, 327–328 (t. h. Squalus; New Zealand).—Ben. & Hesse, 1864, 76.—Braun, 1890a, 529, 547, 552.—Cuvier, —, pl. 36, figs. 3, 3a.—Dies., 1850a, 430 (syn. of T. blanchardii).—Setti, 1898, 311; 1899, 84; 1899, 125.— Tasch., 1878, 564, 567.

sturionis (Abildg., 1794) Cuvier, 18—, pl. 36, figs. 4-4a or Bl., 1847, 329-330.-Ben. & Hesse, 1864, 67 (of Bl. syn. of Nitzschia elegans).—Dies., 1850a, 426 (syn. of Nitzschia elegans).—Johnston, 1865a, 34.—Tasch., 1878, 568 (of Bl.

syn. of T. elongatum).

tubiporum Dies., 1836a, 14-15, pl. 1, figs. 14-16 (in Trigla hirundo); 1850a, 428 (renamed Trochopus longipes as type of Troch.).—Ben. & Hesse, 1864, 74, 75, 76, 77 (to Troch.).—Duj., 1845a, 323.—Massa, 1906, 43; 1906, 53, 54 (to Troch.).—Nord., 1840, 602 (to Capsala).—Sons., 1891, 261.—Stoss., 1898, 7 (in Trigla hirundo; Triest).—Tasch., 1878, 563, 564, 568 (in Trig. hir.).

uncinatum Mont., 1889h, 117-118, pl. 4, figs. 1-7 (in ?Pleuronectes; Coll. Leuck.); 1889i, 516; 1891, 101, 116, 123; 1893.—Braun, 1890a, 487, 497, 529, 547, 551

(in Hippoglossus sp.).—Goto, 1894a, 237.—Setti, 1898; 1899, 124.

TRISTOMATIDÆ Gamb., 1896a, 53, 55, 73.—Scott, T., 1901, 141, 142.

TRISTOMATINÆ Gamb., 1896a, 73.

TRISTOME [French] Blainv., 1824a, 515 ("Nouveau genre que M. Bosc avait nommé capsule pour un animal parasite sur les branchies des poissons, que je regarde

de la famille des sangsues").

TRISTOMEÆ Tasch., 1879, 56, 57, 67; 1879, 235, 238.—Braun, 1883a, 58; 1890a, 516, 517, 523, 524, 525; 1890b, 127.—Cerf., 1894, 946, 948; 1895h, 914, 919; 1898b, 36.—Haswell, 1892a, 457.—Jackson, 1888, 644, 653 (includes: Tristomidæ, Monocotylidæ, Udonellidæ).—Juel, 1889, 14.—Hoyle, 1890, 539 (includes: Tristomidæ, Monocotylidæ, Udonellidæ).—Mont., 1888a, 15, 86 (Ttistomeæ); 1888a, 7, 10, 11, 12, 13, 15, 17, 18, 20, 21, 26, 27, 31, 32, 34, 36, 37, 42, 47, 48, 49, 50, 53, 54, 56, 67, 70, 86, 87, 88, 96, 108, 110; 1891, 108.—Par. & Per., 1890, 5.—Sons., 1890, 175.

1888: Ttistomeæ Mont., 1888a, 15, 86, misprint.

TRISTOMIDÆ Cobbold, 1877f, 326; 1879b, 4.—Braun, 1890a, 516, 517, 519, 523, 526, 538; 1893a, 889; 1896b, 7.—Cerf., 1894, 946, 948; 1894k, 936–948, figs. 41 figs.; 1892, Jan. 15, 99–134, 1 fig., pls. 5–6, figs. 1–41; 1892, Oct. 7 (fam. of Eterocotylea; contains Tristominæ, Encotyllabinæ, Udonellinæ); 1893, 21; 1899, 77; 113; 1903, 335 (contains subf. Tristominæ (g. Tristomum, Trochopus [=Placunella]; 2. Acanthocotylinæ (g. Acanthocotyle); 3. Ancyrocotylinæ (g. Ancyrocotylinæ (g. Ancyrocotyle, Epibdella [=Phylonella], Nitzschia); 4. Encotyllabinæ (g. Encotyllabe); 1905, 68.—Muehl., 1898, 17.—Pratt, 1900, 646 (includes: Tristominæ, Encotyllabinæ, Udonellinæ), 648.—St.-Remy, 1891, 55 pp., 2 pls.; 1891, 11 May, 1072–1074 (genital organs); 1892, 30 May, 702–703; 1892, 1–55; 1892, 2 Oct., 615–616; 1898, 522, 532.—Schneidemuehl, 1896, 296.—Scott, 1901, 141.—Stoss., 1898, 5.—Tasch., 1879, (of Ben.) 68; 1879, 235.

TRISTOMIDEA Carus, 1863, 477.—Mont., 1888a, 84.

TRISTOMIDES Tasch., 1879, 235.—E. Bl., 1888a, 541 (embraced in Polystomiens).— Mont., 1888a, 86.

TRISTOMIDI Mont., 1892g, 99-134, figs. 1-41, for Tristomidæ.

TRISTOMII E. Bl., 1847, 321.

TRISTOMINÆ Braun, 1893a, 889.—Mont., 1892, Oct. 7, 213 (subf. of Tristomidæ, contains Tristomum, Nitzschia, Epibdella, Trochopus, Acanthocotyle); 1899, 81, 113; 1903, 335 (f. Tristomidæ).—Pratt, 1900, 646, 648 (includes: Tristoma, Nitzschia, Epibdella, Phyllonella, Trochopus, Acanthocotyle, Placunella).— St.-Remy, 1898, 532.

TRISTOMUM, see Tristoma.

TRISTOMUN Mont., 1888a, 28 (for Tristomum).

TRISTONUM Setti, 1899, 84, for Tristomum.

TROCHOPUS Dies., 1850a, 290, 428 (m. longipes) [not Carpenter, 1898, insect]; 1858e, 313, 365.—Ben., 1858a, 1861a, 38.—Ben. & Hesse, 1864, 66, 74.—Braun, 1890a, 411, 412, 415, 451, 465, 469, 511, 515, 516, 517, 519, 523, 526, 528; 1891d, 1890a, 411, 412, 413, 401, 403, 401, 410, 410, 410, 417, 418, 422; 1893a, 889.—Burm., 1856a, 251.—Gamb., 1896a, 73.—Goldb., 1855a, 20.—Massa, 1903, 252–255; 1906, 43–71 (syns.: Capsala Nord., Trist. Tasch., Placunella Ben. & Hesse).—Mont., 1888a, 7, 10, 11, 13, 52, 66, 86, 87, 97; 1891, 105, 107, 111, 124, 125; 1892, Oct. 7, 213 (gen. of Tristominæ); 1899, 98; 1903, 335—Placunella (subf. Tristominæ; f. Tristomidæ).—Pratt, 1900, 646, 648 (key).—Scott, 1901, 143; 1901, in 120–153, 2 pls.—St.-Remy, 1898, 534.—Sons., 1891, 260–261.—Stoss., 1898, 7.—Tasch., 1878, 566 (syn. of Tristomum).

differens Sons., 1891, 261 (in Cantharus lineatus Mont.).—Braun, 1891d, 422.-Massa, 1903, 255 (in C. lin.); 1906, 44, 46, 53, 60–61, 65, pl. 2, figs. 24, 25, 26

(syn. T. longipes Sons., 1890) (in C. lin.; Pisa).

diplacanthus Massa, 1903, 254 (in Trigla hirundo); 1906, 45, 46, 53, 56, 64, 65, 66, pl. 2. figs. 3, 4, pl. 3, fig. 33 (syn. Placunella pini Scott, 1901) (in T. hir.; Scotland).

exachanthus Par. & Perugia, 1889.—Massa, 1903, 255 (in Serranus gigas).—For hexacanthus.

heteracanthus Massa, 1903, see heterachanthus.

heterachanthus Massa, 1903, 252, 254 (in Trigla corax); 1906, 46 (heteracanthus), 48, 51, 53, 55, 57, 62, pl. 2, figs. 8, 9, 10, 11, pl. 3, figs. 27, 31, 37, 38, 39, 40, 41 (syns.: T. longipes Sons., T. tubiporus Sons.) (in Trig. co.; Naples).

hexacanthus (Par. & Perugia, 1889) Massa, 1906, 45, 46, 47, 53, 59-60, 65, pl. 2, figs. 15, 16, 17, pl. 3, figs. 28, 36 (in Serranus gigas; Genova).

 lineatus Scott, 1901, 143–144, pl. 8, fig. 18 (in Trigla lineata; Clyde); 1905, 118.
 Massa, 1903, 254 (in Trigla lineata); 1906, 45, 46, 52, 61, pl. 2, figs. 5, 6, 7, pl. 3, figs. 32, 35 (in Trigla lineata; Scotland).

longipes Dies., 1850a, 428 (= Trist. tubiporum 1835, renamed) (in Trigla hirundo); 1858e, 365.—Ben. & Hesse, 1863, 75; 1864, 75 (syn. of T. tubiporus).—Kroyer, 1838—40a, 582 (in Trig. hir.).—Massa, 1906, 53, 55 (syn. of T. tubiporus Dies.), 60 (of Sons., 1890) (syn. of T. differens Sons.), 62 (syn. of T. heteracanthus Massa).—Mont., 1891, 123, 124.—Sons., 1891, 260, 261.—Stoss., 1898, 7 (in Trig. hir. Trig.). Tasch., 1875, 564, 565, 568 (syn. of T. tubiporum) Trig. hir.; Triest).—Tasch., 1878, 564, 565, 568 (syn. of T. tubiporum).

TROCHOPUS—Continued.

micracanthus Massa, 1903, see micrachanthus.

micrachanthus Massa, 1903, 255 (in Trigla hirundo); 1906, 46, 53, 56, 63, 65, 66, pl. 2, figs. 21, 22, 23, pl. 3, fig. 29 (in Trigla hirundo; Genova).

microcanthus Massa, 1906, 66, for micracanthus.

onchacanthus Massa, 1906, 44, 46, 53, 65–66, pl. 2, figs. 12, 13, 14, pl. 3, fig. 30 (host unknown; Triest).

pini (Ben. & Hesse, 1863) Massa, 1903, 254 (in Trigla pini); 1906, 46, 53, 55, 56–58, 63, 65, 66, pl. 2, fig. 2 (syns.: Placunella pini Ben. & Hesse, Trist. pini Tasch., Troch. tubiporus Sons.) (in Trig. pini, T. hirundo, T. corax; Naples).

rhombi (Ben. & Hesse, 1863) Massa, 1903, 255 (in Rhombus maximus); 1906, 45, 46, 53, 58–59, 65, pl. 2, fig. 1 (syns.: Placunella rhombi Ben. & Hesse, Trist.

rhombi Tasch.) (in Rh. max.; Brest).

tubiporus (Dies., 1835) Ben. & Hesse, 1864, 66, 75–76, pl. 6, figs. 8–13 (syns.: Trist. tubiporum, T. longipes) (in Trigla hirundo).—Braun, 1890a, 418, 424, 488, 528, 547, 552.—Massa, 1903, 253, 254 (in Trig. hir.); 1906, 44, 46, 53–55 [56, 57 (of Sons., 1901, 200, syn. of T. pini Ben. & Hesse)], 60, [62 (of Sons., 1891, 260, syn. of T. heteracanthus)], 65, 66, pl. 2, figs. 18, 19, 20, pl. 3, fig. 34 (syns.: Capsala tubipora Nord., Trist. tubiporum Dies.; Troch. longipes Dies.).—Mont., 1888a, 66; 1891, 104, 124, pl. 5, fig. 11, pl. 6, figs. 12–16.—Par. & Perugia, 1890, 5.—Pratt, 1900, 655, 657, fig. 7.—St.-Remy, 1898, 535.—Scott, A., 1901, 345; 1901, 143 (in Trigla hirundo).—Sons., 1891, 260, 261.—Stoss., 1898, 7.—Reported also for Cantharus lineatus.

TTISTOMEE Mont., 1888a, 15, 86, misprint for Tristomeæ.

TURBINILLA Bory St.-Vincent, 1823b, 356, genus of Cercariées.

TYLODELPHIS Pag., 1857, 38, pl. 4, fig. 7.—Mont., 1888a, 84.—Villot, 1898, 542 (for Tylodelphys).

TYLODELPHUS Kroyer, 1838-40a, 579.—Linst., 1878a, 290, for Tylodelphys.

TYLODELPHYS Dies., 1850a, 287, 304–305 (type clavata); 1858e, 311, 315–316.— Brand., 1888a, 12, 15, 52; 1890a, 578.—Braun, 1894, 166; 1895b, 132.— Goldb., 1855a, 16.—Mont., 1888a, 84 (Tylodelphis), 91.—Olss., 1893, 8.— Pag., 1857, 46.—Villot, 1898, 542 (Tylodelphis).

1888: Tylodelphis Mont., 1888a, 84, for Tylodelphys.

clavata (Nord., 1832) Dies., 1850a, 305 (in Acerina vulgaris, Perca fluviatilis, Lucioperca sandra, Esox lucius; Rhedoni); 1858e, 316.—Braun, 1892a, 796.— Krover, 1838–40a, 579 (in Acerina cernua).—Olss., 1893, 8.

craniaria Dies., 1858e, 316, based on Leydig, 1853, 382, pl. 14, fig. 6 (in Cobitis fossilis).—Braun, 1892a, 796.—Pavesi, 1881, 616.—To Diplost. by Cobbold, 1860.

petromyci fluviatilis Linst., 1878a, 290, for petromyzonis fluv.

petromyzi fluviatilis Braun, 1892a, 796.

petromyzonis fluviatilis Dies., 1858e, 316 (in Petromyzon fluviatilis) (syns.: Diplost. petromyzoni fluviatilis Mueller, ? Diplost. rachiæum Mueller).

rachidis Hannover, 1864a, 3.

rhachixa (Henle, 1833) Braun, 1892a, 796 (see rhachidis Dies.).

rhachidis Dies., 1850a, 305 (=Dubium ranarum Rud., Diplost. rhachiæum Henle, renamed) (in Rana esculenta, R. temporaria); 1858e, 316 (in Pelophylax esculentus).—Braun, 1893a, 871.—[Caldani, 1794, 312–318, pl. 7, figs. 7–8.]—Hannover, 1864a, 3 (rachidis).—Pag., 1857, 38, pl. 4, fig. 7 (in Rana esculentus). lenta).—[Rud., 1810a, 268 (Dubium ranarum).]

TYPHLOCŒLUM Stoss., 1902, 9, 30–32 (tod. flavum); 1902, 33 (syn. Monost. flavum) (in Anas boschas brasiliana).—Cohn, 1904, 229, 231 (syn. of Monost. flavum),

232.

cucumerinum (Rud., 1809) Stoss., 1902, 32–33, pl. 8, fig. 28 (syns.: Dist. cuc. Rud., Monost. cuc.) (in Avis riparia).

flavum (Mehlis, 1831) Stoss., 1902, 30–32, pl. 9, figs. 31, 32 (syn. Monost. fl.) (in Fuligula marila; Rossitten).—Cohn, 1904, 230–232, fig. 1

sarcidiornicola (Mégnin, 1890) Stoss., 1902, 33-34 (syn. Monost. sarc.) (in Sarcidiornis melanota; Madagascar).

UDONELLA Johnston, 1835a, 497 (m. caligorum); 1865a, 30, 34.—Ben., 1858a, 1861a, 11, 12, 190, 207, 210, 297.—Brand., 1894a, 308.—Braun, 1890a, 451, 452, 465, 468, 477, 483, 484, 490, 498, 511, 515, 516, 517, 518, 522, 523, 531, 532; 1893a, 468, 477, 483, 484, 490, 498, 511, 515, 516, 517, 518, 522, 523, 531, 532; 1893a, 890.—Burm., 1856a, 251.—Carus, 1863, 477.—Chatin, 1880f, 591; 1881a, 311.—Cobbold, 1879b, 463.—Dies., 1850a, 290, 427 (syns.: Hirudo? Kroyer, Amphibothrium Frey & Leuck.); 1858e, 313, 362–363 (m. caligorum); 1859c, 437.—Gamb., 1896a, 73.—Goldb., 1855a, 20.—Hoyle, 1890, 539 (type caligorum).—Jackson, 1888, 644, 653.—Leuck., 1863a, 489; 1879,139; 1886d, 107.—Mont., 1888a, 10, 52, 70 (Udonetla), 84, 86, 88, 98; 1892, Oct. 7, 186, 213 (gen. of Udonellinæ); 1903, 336 (subf. Udonellinæ); 1905, 65.—Par. & Perugia, 1890, 14.—Pratt, 1900, 646, 649 (on Caligus and Anchorella), 655, fig. 12.—Scott, T., 1901, 144.—Stoss., 1898, 8.—Tasch., 1878, 572; 1879, 236, 238; 1879, 56, 59, 68, 73.—Wagener, 1857, 25, 26.

1888: Udonetla Mont., 1888, 70.

caligarum Tasch., 1878, 564, 565, 572-573, for caligorum.

caligi (Kroyer—) Ben., 1858a, 1861a, 189.

caligorum Johnston, 1835a, 497, fig. 45 a-c (on Caligus on Hippoglossus vulgaris); 1865a, 34, 299–300, fig. 49.—Ben., 1858a, 1861a, 12, 13–18, pl. 1, figs. 1–15 (syns.: Phylline caligi Kroyer, Amphibothrium kroyeri) (in Pleuronectus hippoglossus, Gadus morrhua, Trigla gurnardus), 260.—Braun, 1890a, 418 (caligarum), 468, 485, 500, 512, 532, 548, 550, 551 (in Caligus sp.; North Sea; (cangarum), 408, 405, 500, 512, 532, 548, 550, 551 (in Cangus sp.; North Sea; Atlantic Ocean).—Cobbold, 1879b, 484.—Dies., 1850a, 427 (syns.: Hirudo Kroyer, Amphibothrium kroyeri Frey & Leuck.); 1858e, 363; 1859c, 437 (in Caligos hippoglossi vulgaris et gadi morrhuæ; Belgia).—Gamb., 1896a, 55, fig. 22B.—Grube, 1851, 116, 150.—Hoyle, 1890, 539 (on Caligus which is on Hippoglossus vulgaris).—Jackson, 1888, 646, 650 (on Caligus).—Moul., 1856a, 11.—Pag., 1857, 11.—Pratt, 1900, 655, fig. 12, 657.—Quatrefages, 1854, 23.—Scott, 1901, 144 (in Caligus curtus); 1905, 118; 1901, pl. 8.—Staff., 1904, May 3, 482 (on Caligus on Gadus callarias; Canada).—Tasch., 1878, 564, 565, 572-573 (coligarum) 572-573 (caligarum).

hirundinis (Ben. & Hesse, 1863) Tasch., 1878, 573 (syn. Echinella hirundinis).

lupi Ben. & Hesse, 1863, 92, pl. 8, figs. 11-14 (in Labrax lupus).—Braun, 1890a, 408, 417, 427, 482, 532, 548, 550, 551 (in L. lup.; Atlantic Ocean).—Mont., 1890, 420.—Stoss., 1898, 8.—Tasch., 1878, 573.

merlucii Ben. & Hesse, 1863, 93; 1864, 93 (in Merlucius vulgaris).—Braun, 1890a, 417, 532, 548, 550, 551 (in M. vulg.; Atlantic Ocean).—Tasch., 1878, 573.

molvæ (Ben. & Hesse, 1863) Tasch., 1878, 573.

pollachii Ben. & Hesse, 1863, 90–91, pl. 8, figs. 1–8; 1864, 90–91, pl. 8, figs. 1–8 (in Merlangus pollachius).—Braun, 1890a, 408, 410, 417, 446, 510, 532, 548, 550, 551 (in M. poll.; Atlantic Ocean).—Gamb., 1896a, 58, fig. 25B.—Mont., 1888a, 7, 10, 34.—Tasch., 1878, 573.

sciænæ Ben. & Hesse, 1863; 1864, 93, pl. 8, figs. 15–16 (in Sciæna aquila).—Braun, 1890a, 465, 532, 548, 549, 552 (in S. aq.; Atlantic Ocean).—Tasch., 1878, 573.

triglæ Ben. & Hesse, 1863; 1864, 92, pl. 8, figs. 9–10 (in trigles).—Braun, 1890a, 417, 446, 532, 548, 550, 552 (in Trigla sp.; Atlantic Ocean).—Tasch., 1878,

UDONELLIDÆ Tasch., 1879, 235 (Udonellides), 236; 1879, 68.—Braun, 1890a. 516, 517, 523, 526, 531; 1896b, 7.—Cerf., 1894, 946, 947, 948; 1898b, 361, 362.— Haswell, 1893e, 125.—Hoyle, 1890, 539 (Udonella).—Jackson, 1888, 653 (Udonella).—Mont., 1888a. 7, 10, 13, 18, 34, 36, 66, 67, 86 (Udonellides), 88, 98, 108, 110; 1891, 108; 1903, 336 (subf. Undonellinæ (g. Udonella, Echinella, Pteronella).—Scott, 1901, 141.

UDONELLIDES Tasch., 1879, 235 (see Udonellidæ).—Mont., 1888a, 86.

UDONELLINÆ Mont., 1892, Oct. 7, 213 (subf. of Tristomidæ); 1903, 336 (f. Udonellidæ).—Braun, 1893a, 890.—Pratt, 1900, 646 (includes: Udonella, Echinella, Pteronella), 649.—Scott, T., 1900, 141, 144.

UDONETLA Mont., 1888a, 70, for Udonella.

UNICOLA Rafinesque, 1815, 151, genus of Fasciolaria, nomen nudum; to contain species of Fasciola, but these are not mentioned. The name has therefore no status at present.

UROCENTRUM Nitzsch, 1827, 68 (m. Cerc. turbo).

UROGONIMINÆ Looss, 1899b, 655.—Pratt, 1902, 889, 906 (includes Urorygma, Urogonimus, Urotocus, Urotrema).

UROGONIMUS Mont., 1888a, 15, 92, 104 (tod. macrostomus); 1892, Oct. 7, 214 (gen. of Distominæ; 1893, 82, 154, 155, 157, 162, 164, 165; 1896, 167,—Braun, 1892a, 696, 699, 715, 727, 736; 1893a, 885, 886, 890, 893, 909, 911; 1895b, 138; 1900h, 3; 1900, 390; 1900, 234, 1901, 942, 943; 1902b, 136, 140,—Darr, 1902, 655,—Looss, 1894a, 173, 174; 1896b, 139; 1899b, 536, 538, 542, 551, 648, 653-654, 655, 665, 749, 750,—Luehe, 1899, 532; 1900, 555,—Moniez, 1896a, 90,—Nickerson, 1902, 606,—Ofenheim, 1900, 183,—Poche, 1907, 125,—Pratt, 1902a, 889, 906,—Stiles, 1901, 176, 194,—Stiles & Hass., 1898a, 95, 96 (syn. Leucochloridium), type Fasc, macrostoma,—Stoss., 1892, 4, 5; 1898, 23,—See also Leucochloridium 1835.

cercatum, see cercatus.

cercatus (Mont., 1893) Mont., 1893, 40, 42, 43, 83, 95, 102, 157 (cercatum and cercatus, 162-166, pl. 5, figs. 64-65; 1896, 167.—Looss, 1894a, 174 (cercatum); 1896b, 139, 140; 1899b, 654, 750.—Mueh., 1898, 16 (footnote); 1898, 99.

insignis Looss, 1899b, 596, 654, 748–750. figs. 49, 71 (in Fulica atra: Marg, Egypt). macrostomum (Rud., 1803) Mont., 1893, 27, 67, 68, 76, 83, 95, 102, 107, 157.

macrostomus (Rud., 1803) Mont., 1892. Oct. 7, 187 (macrostomum); 1893. 27, 67, 68, 76, 83, 95, 102, 106, 107, 157, 162, 164 (macrostomum and macrostomus); 1896, 167.—Braun. 1901. 562, 564, 565, 567, 568; 1902b, 40 (in Fringilla cœlebs), 42 (in Fringilla linaria, Parus palustris, of Vienna Col., Nos. 361 and 466), 124, 125 (macrostomum), 136, 139 (syns.; Dist. elegans pars, D. caudale of Mueller in Coracias garrula, D. holostomum Rud., 1819, (Iadoccelium hol. Stoss., 1892) (in Rallus aquaticus, Gallinula chloropus, G. porzana).—Jacoby, 1900, 20.—Kowal, 1902d, 28 (10): 1904, 25 (10) (in Turtur aurita; Dublany).—Looss, 1896b, 139 (to Dist.); 1899b, 654, 750, 761.—Muehl., 1898, 16, 17, footnote): 1898, 22, 99, 101.—Stoss., 1896, 127, fig. 1; 1898, 23–24 (in Numenius tenuirostris; Monialcone).—Reported also for Passer domesticus.

rossittensis Mueh., 1898, 16-17 (in Turdus pilaris; East Prussia; 1898b, 4, 22, 100-101, fig. 15 at Rossitten).—Jacoby, 1900, 20.—Looss, 1899b, 654, 750, type of

Urotocus.

URORYGMA Braun, 1901g. 942 (m. nanodes); 1902b, 138.—Pratt, 1902, 889, 904 (key to).

nanodes Braun, 1901g, 942-943 (in Falco nitidus; Brazil; 1902b, 138-140, figs. 81, 82.

UROTOCUS Looss, 1899b, 654-656 (tod. m. rossittensis): 1902m, 839.—Braun, 1900, 234; 1900, 390; 1902b, 140.—Luehe, 1901, 488.—Ofenheim, 1900, 183.—Pratt, 1902, 889, 907.

rossittensis (Mueh., 1898) Looss, 1899b, 655.

UROTREMA Braun. 1900, 390–391 (m. scabridum); 1902b, 140.—Looss, 1902m, 839.— Luehe, 1901, 488.—Pratt. 1902, 889, 901.

scabridum Braun, 1900, 390–391 (in Fledermaus): 1900b, 234–236, pl. 10, figs. 5, 10 Noctilio macropus. Molossus. Phyllostoma: Brazil).

VALLISIA Par. & Per., 1890. 21 (m. striata); 1890. 18 (striata), Octocotylidæ.—
Braun, 1890a, 407, 451, 453, 469, 481, 484, 519, 523, 534, 536, 546; 1893a, 890.—
Ceri., 1895h, 920; 1896, 515.—Gamb., 1896a, 73 (Vallisnia).—Mont., 1892. Oct.
7, 213 (subg. in Octocotylinæ); 1903, 336 (subi. Octobohtrinæ).—Pratt, 1900, 646, 651 (key., 656, fig. 32.—Stoss., 1898, 13.

striata Par. & Per. 1890, 21 (in Lichia amia: Triest); 1890, 7; 1891, Jan. 19, 17–19.—Braun. 1890a. 534, 536, 548, 551 (in Lichia amia; Mittelmeer, Triest).—Par., 1894, 594, 671.—Pratt. 1900, 656, 657, fig. 32.—Sons., 1890, 138 (to Octocotyle).—Stoss., 1898, 13–14.

VALLISNIA Gamb., 1896a. 73, for Vallisia.

VERTUMNUS Otto, 1823, 294-300 (m. thetidicola), syn. of Phænicurus.

thetidicola Otto, 1823, 294–300, pl. 41, figs. a-i (in Thetis fimbria) (syn. of Phonicurus varius Rud.).—Braun, 1889a, 326.—Krohn, 1842a, 418–423.

VIRGULINA Bory St.-Vincent, 1823b, 356, genus of Cercarieés.

WEDLIA Cobbold, 1860a, 37-38 (biparrita Wedl, from Thynnus vulgaris, was clearly intended as type).—Braun, 1893a, 894 (faba apparently taken as type by elimination).—Mont., 1888a, 9, 93: 1892, 714.—Tasch., 1879, 607.

1878: Didymozoon Tasch., 1878, 716.

1884: Weidlia Scudder, 1884, 335, misprint.

WEDLIA—Continued.

1902: Didymostoma Ariola, 1902, 103 (bipartitum).

bipartita (Wedl, 1855) Cobbold, 1860a, 38 (in Thynnus vulgaris).—Mont., 1893,

150.—Type of Didymostoma 1902.

faba (Bremser, 1831) Cobbold, 1860a, 38 (syns.: Monost. faba Bremser, Duj., Crep., Dies.; M. bijugum Miescher, Sieb.; Globularia Orlando) (in Fringilla canaria, F. domestica, F. spinus, Parus major, Sylvia sibilatrix, S. trochilus, Motacilla boarula).

WEIDLIA Scudder, 1884, 335, misprint for Wedlia.

XENODISTOMUM Staff., 1904, May 3, 483 ($\xi \dot{\epsilon} \nu o \varsigma = \text{stranger}$) (m. melanocystis).

melanocystis Staff., 1904, May 3, 483 (t. h. Lophius piscatorius; Canada) (μέλας, black; κύστις, bladder).—Odhn., 1905, 310.

ZEUGORCHIS Staff., 1905, Apr. 11, 691–692 (ζεῦγος, pair; ὄρχις, testicle) (m. æquatus).

xquatus Staff., 1905, Apr. 11, 691-692 (= Dist. sp. Staff., 1902, 482) (in Eutenia sirtalis; Canada).

ZEUGORCHIS Nicoll, 1906, 514 (m. acanthus); 1907, 128, renamed Parorchis.

acanthus Nicoll, 1906, 514, 519-522, pl. 12, figs. 4-5, pl. 13, figs. 6-7 (in Larus argentatus).

(XIPHIDIOCERCARIA) Dies., 1855a, 388–390 (subg. of Cercaria); 1858d, 253, renamed C. (Acanthocephala).

1858: C. (Acanthocephala) Dies., 1858d, 253-255.

armata (Sieb., 1837) Dies., 1855a, 388; 1858d, 251, to C. (Acanthocephala).—See endoloba, type of Opisthioglyphe 1899.

qibba (Fil., 1854) Dies., 1855a, 389; 1858d, 257, to C. (Acanthocephala).

macrocerca (Fil., 1855) Dies., 1855a, 389; 1858d, 255, to C. (Acanthocephala).— See cygnoides, type of Gorgodera.

microcotyla (Fil., 1854) Dies., 1855a, 390; 1858d, 253, to C. (Acanthocephala).— See tetracystis, type of Cystagora.

vesiculifera Dies., 1855a, 389; 1858d, 254, to C. (Acanthocephala).

vesiculosa (Fil., 1854) Dies., 1855a, 389 (syn. of Cerc. vesiculifera).

ZOOGONINÆ Odhn., 1902, 63-64.—Goldschmidt, 1902e, 870.—Pratt, 1902, 889, 906 (key) (Zoögonus, Zoögonoides).

ZOOGONOIDES Odhn., 1902, 61, 64 (tod. viviparus).—Goldschmidt, 1902e, 870.—Pratt, 1902, 889, 906 (key).

viviparus (Olss., 1868) Odhn., 1902, 62–63, 64, fig. 2.—Goldschmidt, 1902e, 870.— Nicoll, 1907, 72, 83-84, pl. 2, fig. 8, pl. 3, fig. 9 (in Pleuronectes limanda, P. platessa, Rhombus maximus).

ZOOGONUS Looss, 1901d, 439 (tod. mirus indicated by "n. g. n. sp.").—Luehe, 1901, 176.—Odhn., 1902, 58-63, 64.—Pratt, 1902, 889, 906 (key).

mirus Looss, 1901, 439–442, fig. 6 (in Labrus merula; Triest).—Goldschmidt, 1902e, 870–876, figs. 1–6 (in Labrus merula); 1903a, 535–536; 1905, July, 56; 1905, 607–654, pls. 36–38, figs. 1–42.—Nicoll, 1907, 83.—Odhn., 1902, 58, 59, 60, 61, 64.—Rœwer, 1906, 220, 221.

rubellus (Olss., 1868) Odhn., 1902, 59-61, 63, fig. 1.—Goldschmidt, 1902e, 870, 872, 873.

viviparus (Olss., 1868) Looss, 1901, 439, 440, 441, 442.—Type of Zoogonoides 1902. ZOOSPERMA Bory St.-Vincent, 1823b, 356.



ADDENDA.

Since the manuscript of this bulletin left our hands, a number of additional references to trematodes have come into our possession. Only such references as involve new names and new combinations will be inserted in the addenda.

Particular attention is invited to an important change in the International Code relative to the collective groups, namely:

ART. 8.—Recommendation: Certain biological groups which have been proposed distinctly as collective groups, not as systematic units, may be treated for convenience as if they were genera, but they require no type species. Examples: Agamodistomum, Amphistomulum, Agamofilaria, Agamomermis, Sparganum.

In the present bulletin, types were designated for certain of these groups before the foregoing article was adopted. Such designation becomes null and void under this article.

(385

858S-No. 37-08-25

ADDITIONAL SPECIFIC NAMES.

acanthus Nicoll, 1906. [See supra, p. 10.] Add: 1907: Parorchis.

affinis Looss, 1907, 158, t. h. Anguilla chrysypa, Osmerus mordax; U. S. A.—1907: Brachyphallus. albida Braun, 1893. [See supra, p. 12.] Add: 1898: Campula. angulatum Linst., 1907, 202, t. h. Lucioperca sandra; Wolga.—1907: Phyllodist. annuligerum Nord., 1832. [See supra, p. 14.] Add: 1904: Diplost. appendiculata Pelseneer, 1906, 167, t. h. Natica alderi; Boulogne-sur-Mer.-1906: Cerc. baccutus Nicoll, 1907, 72, t. h. Hippoglossus vulgaris; Scotland.—1907: Stephanochasmus. bacillare Mol., 1859. [See supra, p. 16.] Add: 1907: Ophecona, type. baculatum Lint., 1907, 119, for baculum (Gasterost.). bathycothyle Fischder., 1903h, 542, for bathycotyle (Paramphist.). bovium Nakahama, (1883b).—1883: Amphist. branchialis Willem, 1906, 599, t. h. Raie bouclée; Belgium.—1906: Acanthocotyle. brevicauda Pelseneer, 1906, 167, t. h. Littorina rudis; Wimereux.—1906: Cerc breviductus Looss, 1907, 596, t. h. Pelamys sarda; Atlantic Ocean.—1907 Dinurus. burki Rathelot, 1892a, 14, for buskii (Dist.). buski Rail., 1898, 172, for buskii Lankester.—1898: Campula. caducum Looss, 1907, 615, t. h. Umbrina cirrhosa.—1907: Pristisomum. calculus Looss, 1907, 610, t. h. Larus argentatus, L. ridibundus; Triest.—1907 Pachytrema. caligarum Ben., 1858a, 1861a, 12, for caligorum (Udonella). cignoides Desmonceaux, 1868a, 21, for cygnoides (Dist.). columbæ Mazzanti, 1889. [See supra, p. 24.] Add: 1890: Mesogonimus. commutatum Dies., 1850. [See supra, p. 24.] Add: 1898: Clinost. complexa Stiles & Hass., 1894. [See supra, p. 25.] Add: 1898: Campula. conchicala Nord., 1833b, 281, for conchicola (Aspidogaster). conchilega Braun, 1883a, 41, for conchicola (Aspidogaster). conjuncta Cobbold, 1860. [See supra, p. 25.] Add: 1898: Campula. conus Crep., 1825. [See supra, p. 26.] Add: 1898: Campula. cornu Dies., 1839. [See supra, p. 26.] Add: 1907: Diplodiscus. cornuta Mueller, 1776a, 221.—1776: Plan. cotti Zschokke, 1884, 6, t. h. Cottus gobio.—1884: Sporocystis. crenatum Mol., 1859. [See supra, p. 27.] Add: 1907: Lecithocladium. crispata Pelseneer, 1906, 171, t. h. Natica alderi; Boulogne.—1906: Cerc. eristatum Rud., 1819. [See supra, p. 27.] Add: 1907: Lecithocladium. cuculus Thoss, 1897; 1, t. h. Larus ridibundus; Kiel.—1897: Holost. cygnoides ranæ Wagener, 1857, 19.-1857: Dist.

cylindricus Dies., 1836. [See supra, p. 28.] Add: 1907: Pseudocladorchis

cylindriaceum Daday, 1907, 560, for cylindraceum (Dist.).

. type.

- dentalii Pelseneer, 1906, 170, t. h. Dentalium tarentinum.—1906: Cerc.
- didactyla Mrazek, 1907, 1, t. h. Atyaëphyra desmarestii; Plavnica.—1907; Scutariella.
- dipsilis Nicoll, 1907, 247, t. h. Oidemia fusca, O. nigra; Great Britain.—1907: Gymnophallus.
- dorsale Freund, 1907, 718, for chordale (Amphist.).
- egyptiaca Pease, 1901b, 7, for ægyptiaca (Fasc.).
- emasculans Pelseneer, 1906, 166, t. h. Littorina rudis; Wimereux.—1906: Cerc. endorfii Par. & Perugia, 1895, 2, for hendorfii (Phylline).
- cxcellens Nicoll, 1907, 247, t. h. Larus argentatus; Great Britain.—1907: Spelotrema.
- fenestratum Lint., 1907, 111, t. h. Lycodontis moringa; Bermuda.—1907: Dist.
- feriatum Nicoll, 1907, 247, t. h. Ægialitis hiaticula, Hæmatopus ostralegus, Pelidna (Tringa) alpina, Totanus calidris, Vanellus vanellus; Great Britain.—1907: Spelotrema.
- ferrum-equinum Dies., 1836. [See supra, p. 35.] Add: 1907: Microrchis.
- flum Looss, 1907, 606, t. h. Talpa europæa; Leipzig.—1907: Ityogonimus.
- furcata Mueller, 1786a, 299, free form.—1786: Vorticella. 1816: Furcularia.
- fusiforme Luehe, 1901. [See supra, p. 37.] Add: 1907: Sterrhurus.
- galeatus Looss, 1907, 165, t. h. Mugil auratus; Coast of Egypt.—1907: Lecithaster.
- gazzettæ Arch. f. Naturg., 1901, v. 2 (8), 188, for garzettæ (Echinost.).
- giardi Pelseneer, 1906, 170 t. h. Buccinum undatum; Boulogne.—1906: Cerc.
- glaviger Ssinitzin, 1906, 687, for claviger (Pleurogenes).
- gracilius Pelseneer, 1906, 176, for grascilescens (Gasterost.).
- grandiporum Rud., 1819. [See supra, p. 39.] Add: 1907: Sterrhurus.
- gratiosum Nicoll, 1907, 247, t. h. Pelidna (Tringa) alpina; Great Britain.—1907:
 Maritrema.
- gravidum Looss, 1907, 603, t. h. Anguilla vulgaris; ?Triest.—1907: Lecithochirium.
- gulosum Lint., 1901. [See supra, p. 39.] Add: 1907: Lecithocladium.
- gyrinus Lint., 1907, 107, t. h. Lactophrys tricornis, L. trigonus; Bermuda.—1907; Dist.
- hamatodium Dewitz, 1892b, 106, for hamatobium (Dist.).
- haswelli Mont., (1898), 1899, 122, see blanchardi 1893.—1899: Actinodactynella (type).
- hepaticum hominis Cobbold, 1884g, 976.—1884: Dist. See Fasc. hep.
- heterophyes Sieb., 1853. [See supra, p. 41.] Add: 1898: Clinost. 1904: Paragonimus.
- hamatobium Rathelot, 1892a, 14, 15, for hamatobium (Dist.).
- humile Nicoll, 1907, 247, t. h. Totanus calidris; Great Britain.—1907: Maritrema.
- imocavus Looss, 1907, 601, t. h. Thynnus sp. (?thunnina); Alexandria, Egypt.—1907: Sterrhurus,
- inermis Goto, 1899a, 273, see læve var. inermis (Trist.).
- inversum Looss, 1907, 615, t. h. Chrysophrys aurata; Triest.—1907: Placotrema.
- jacksoni Cobbold, 1869a, 80, for jacksonii (Fasc.).
- jaksoni Stazzi, 1900, 3, 4, 5, for jacksonii (Fasc.).
- jejunum Nicoll, 1907, 248, t. h. Totanus calidris; Great Britain.—1907: Tocotrema.
- kordatum see cordatum.
- laguncula Looss, 1907, 69, t. h. [fish]; Triest.—1907: Aponurus, type.
- lamelliforme Lint., 1907, 108, t. h. Balistes carolinensis, Lactophrys tricornis, L. trigonus; Bermuda.—1907; Dist.

lapidus Looss, 1907, 69, t. h. fish. [See p. —.]—1907: Ectenurus.

laurentina Borelli, 1897, 1.—1897: Plan.

lenori Mont., 1896, 167, for lenoiri (Cephalogonimus).

lepidum Nicoll, 1907, 247, t. h. Larus argentatus; Great Britain.—1907; Maritrema.

lepidus Looss, 1907, 597, t. h. Lichia amia.—1907: Ectenurus, type.

leptosomum Rœwer, 1906, 185, 186, for leptostomum (Dist.).

levenseni Lint., 1907, 110, t. h. Epinephelus maculosus, E. striatus; Bermuda.—1907; Dist.

longisinus Looss, 1907, 596, t. h. Coryphæna hippuris; Red Sea, Aden.—1907; Dinurus.

lophocerca Fil., 1857. [See supra, p. 49.] Add: 1907: Monost.

lugubris Morgan, 1901, 179.—1901: Plan.

lunatus Dies., 1836. [See supra, p. 49.] Add: 1907: Chiorchis.

macrophallos Linst., 1875. [See supra, p. 50.] Add: 1907: Levinseniella.

macrostomus Daday, 1907, 506, t. h. Salmo pacupa, S. sp.; South America.—1907: Pseudocladorchis.

mamillaris Gærtner in Pallas, 1774, 20.—1774: Distomus.

marcnzelleri Daday, 1907, 473, t. h. Salmo sp.; South America.—1907: Diplodiscus.

margaritarum Dubois, 1901. [See supra, p. 51.] Add: 1907: Gymnophallus. marmorata Bosc, 1802a, v. 1, 262, for marmorosa?.—1802: Plan.

m'connelli McConnell, 1878a, 406, for macconnelli (Dist.).

mcgacotyle Dies., 1836. [See supra, p. 52.] Add: 1907: Microrchis, type.

megalocotyle Walter, 1893, 19, 24, for megacotyle (Amphist.).

mclangi Lebour, 1907, 442, for merlangi (Octobothrium).

migoccra Lebour, 1906, 7, for myocerca (Cerc.).

minor Ben., 1858, 98, see armata minor (Cerc.).

montenegrina Mrazek, 1904a, 1; Montenegro.—1904: Plan.

monticelli Mont., 1905, 74, for monticellii (Acanthocotyle).

muehlingi Nicoll, 1907, 257, 259, for mühlingi (Dist.).

musculus Looss, 1907, 600, t. h. Anguilla vulgaris, Dentex vulgaris; Triest.—1907: Sterrhurus, type.

myocercoides Pelseneer, 1906, 162, t. h. Syndosmya alba; Boulogne-sur-Mer.—1906; Cerc.

nanus Stiles & Goldberger, 1908, 23. t. h. Francolinus subtorquatus; Africa.—1908: Agamodist.

nephrodorchis Daday, 1907, 501, t. h. Salmo pacu, S. pacupa; South America.— 1907: Pseudocladorchis.

nigrotineta Pelseneer, 1906, 166, t. h. Syndosmya alba; Boulogne-sur-Mer.—1906: Cerc.

novæ zealandiæ Haswell, 1888, 50, for novæ zelandiæ (Temnocephala).

obtusicauda Pelseneer, 1906, 182, for obtusicaudata (Cerc.).

obtusicaudata Pelseneer, 1906, 167, t. h. Natica alderi; Boulogne.—1906: Cercocysta Lebour, 1907, 439, t. h. Paludestrina stagnalis; Northumberland Coast.—1907: Cerc.

opaca Ward, 1894. [See supra, p. 59.] Add: 1907: Levinseniella.

ophtalmobium Rathelot, 1892a, 14, for ophthalmobium (Dist.).

paguri Nord., 1833b, 379.—1833: Polyst.

pankreatikum Katsurada & Saito, ——, Hyg. Centralbl., v. 3, Oct., 279, for pan creaticum (Dist.).

papillatus Daday, 1907, 520, t. h. Cholossoma brachypoma; Paraguay.—1907 Chiorchis.

paronæ Mont., 1907, 3, t. h. Crenilabrus pavo; Genoa.—1907: Encotyllabe parvirenalis Pelseneer, 1906, 165, t. h. Natica alderi; Boulogne.—1906: Cerc pedicillatum Luehe, 1900u, 487, for pedicellatum (Dist.).

pellucida Jægers. 1907, 134, t. h. Anas boschas fera, Fuligula fuligula; West Coast of Sweden.—1907: Levinseniella.

petromyzi fluviatilis Dies., 1858e, 316, for peteromyzi fluviatilis (Diplost.).

philippineusis Stiles & Goldberger, 1908, 23, t. h. Bos sp.; Manila, P. I.—1908: Homalogaster.

pirum Lebour, 1907, 439, t. h. Paludestrina stagnalis; Northumberland Coast.— 1907: Cerc.

polymastus Schneidemuehl, 1896, 303, for polymastos (Gastrodiskus).

preistis Mola, 1907, 39, for pristis (Dist.).

propinqua Jægers., 1907, 135, t. h. Charadrius hiaticula, Hæmatopus ostralegus; West Coast of Sweden.—1907: Levinseniella.

pumex Looss, 1907, 615, t. h. Caranx trachurus.—1907: Pristisomum.

[pupula Bory St.-Vincent, 1825b, 252.—1825; Histrionella.]

pussilum Harz, 1881c, 3, for pusillum (Dist.).

putorii Schrank,? or Gmelin, 1790. [See supra, p. 66.] Add: 1898: Echinost.
quadrangulatum Daday, 1907, 470, t. h. Salmo pacu; Cuyaba.—1907: Dist.

raggazzii Janicki, 1907, 719, for ragazzii (Syncœlium).

rajazzii Ariola, 1899, 135, for ragazzii (Dist.).

ranerium Nardo, 1833a, 523, for raynerianum (Dist.).

rathousii Mont., 1893, 33, for rathouisi (Dist.).

ropaloïdes Olss., 1867a, 19, 48, for ropaloides (Amphist.).

rugosa Dies., 1850a, 408, t. h. Antilope pyarga; Port Natal.—1850: Gyrocotyle, type.

rugosus Looss, 1907, 591, t. h. Clupea pilchardus, C. sardina, Rhombus maximus; Triest.—1907: Hemiurus.

sanguicola delle Chiaje, 1825a, 15.—1825: Hexathyridium.

scienæ Fraip., 1880c, 442, for sciænæ (Epibdella).

scleroporum Rud. of Brand., 1891d, 19, for Crep.?—1891: Amphist.

semifuscum Looss, 1907, 607, t. h. Circetus [=Circetus?] gallicus; Genoa.— 1907: Platynosum, type.

seminis Owen, 18351, 394.—1835: Cerc.

serrata Bory St.-Vincent, 1825a, 84, t. h. "infusion de foin."—1825: Furcocerca, siluris glanidis Hofer, 1904a, 169, for siluri glanidis (Dactylogyrus).

simile Jægers., 1900. [See supra, p. 72.] Add: 1906: Dist.

simplicissima Morgan, 1904, 385.—1904: Plan.

sinensis Cobbold, 1875. [See supra, p. 72.] Add: 1898; Campula.

stellatus Looss, 1907, 165, t. h. Belone acus, Dentex vulgaris, Mæna vulgaris; Triest.—1907: Lecithaster.

stossichi Looss, 1907, 69, for stossichii Mont.—1907: Aphanurus (type).

subtenue Lint., 1907, 106, t. h. Calamus calamus, Harpe rufa, Iridio bivittatus, Lachnolaimus maximus; Bermuda.—1907: Dist.

syndosmyæ Pelseneer, 1906, 172, t. h. Syndosmya alba; Boulogne-sur-Mer.—1906; Cerc.

tenuicollis Rud., 1819. [See supra, p. 77.] Add: 1898: Campula.

tomex Lint., 1907, 112, t. h. Epinephelus striatus; Bermuda.—1907: Dist.

tornatum Rud., 1819. [See supra, p. 78.] Add: 1907: Dinurus (type).

triangulæ Linst., 1878, 72, for triangulare (Dist.).

trilobata Bory St.-Vincent, 1825a, 84, t. h. "infusions d'écorce de chêne."—1825: Furcocerca.

trimellaris Bosc, 1802a, v. 1, 262.—1802: Plan.

trulla Lint., 1907, 109, t. h. Ocyurus chrysurus; Bermuda.—1907: Dist.

ubiquita Lebour, 1907, 439, t. h. Littorina obtusata, L. rudis, Paludestrina stagnalis; Northumberland Coast.—1907: Cerc.

urna Grube & Wagener, 1852, 543, t. h. Chimera monstrosa.—1852: Amphiptyches (type).—1896: Gyrocotyle. [Now in the Cestodaria.]

vallei Mont., 1907, 3, t. h. Chrysophrys aurata; Triest.—1907: Encotyllabe.

- vaullegeardi Pelseneer, 1906, 175, t. h. Trochus cinerarius; Wimereux.—1906; Cerc.
- virgula Looss, 1907, 111, t. h. Cepola rubescens, Engraulis encrasicholus; Triest.—1907: Aphanurus.
- volgensis Linst., 1907, 201, t. h. Lucioperca sandra; Wolga.—1907: Ptychogonimus.
- watsoni Conyngham, 1904. [See supra, p. 83.] Add: 1907: Gastrodiscus.
- westermanni Rail., 1898, 173. for westermanii Kerbert, 1878. [See supra, p. 83.] Add: 1898: Clinost.

ADDITIONAL GENERIC AND SPECIFIC BIBLIOGRAPHY.

ACANTHOCOTYLE. [See above, p. 85.]

branchialis Willem, 1906, 599-607, figs. 1-10 (in Raie bouclée; Belgium).

AGAMODISTOMUM. [See above, p. 87.]

nanus Stiles & Goldberger, 1908, 30–33, figs. 45–64 (in Francolinus subtorquatus; Benguella, West Africa).

AMPHIPTYCHES Grube & Wagener, 1852, 543 (m. urna).

urna Grube & Wagener, 1852, 543-554 (in Chimæra monstrosa).

AMPHISTOMA. [See above, p. 89.]

bovium Nakahama, (1883b).—Teste Surg.-Gen. Cat.

dorsale Freund, 1907, 718, for chordale.

megalocotyle Walter, 1893, 19, 24, for megacotyle.

ropaloïdes Olss., 1867a, 19, 48, for ropaloides.

APHANURUS Looss, 1907, May 14, 591 (tod. stossichi), Hemiurinæ, Hemiuridæ; 1907, 69, 74, 78, 98, 107–109, 160.

stossichi (Mont., 1891) Looss, 1907, 591 (in Box boops, Caranx trachurus, Clupea pilchardus (t. h.), C. sardina, Lichia amia, Mæna vulgaris; Naples, Triest, t. l.); 1907, 69, 107, 109-111. 159, pl. 8. figs. 7, 8 (in Clupea aurita).

virgula Looss, 1907, May 14, 592 (in Cepola rubescens, Engraulis encrasichelus; Triest); 1907, 111, 159, pl. 9, figs. 15, 16.

APONURUS Looss, 1907, May 14, 607-608 (tod. laguncula), Lecithasterinæ, Hemiuridæ; 1907, 69, 74, 78, 99, 101, 111, 166-168.

laguncula Looss, 1907, May 14, 608 (in Belone acus, Lichia amia, Engraulis encrasicholus, Mullus barbatus, Cadus euxinus, Merlucius esculentus, Trachinus draco; Triest); 1907, 69, 110, 167, 169, pl. 13, figs. 53, 54, pl. 15, figs. 77, 78 (in Mæna vulgaris; Triest).

ASPIDOGASTER. [See above, p. 103.]

conchicala Nord., 1833b, 281, for conchicola.

BRACHILAIMUS Neumann, 1897f, 674, misprint for Brachylaimus. [See above, p. 116.]

BRACHYPHALLUS. [See above, p. 116.]

affinis Looss, 1907, 158-159, B. crenatus Rud. of Lander, 1904, renamed (in Anguilla chrysypa, Osmerus mordax; U. S. A.).

CALLYCOTYLE Mont., 1892a, 213, for Calicotyle. [See above, p. 119.]

CAMPULA. [See above, p. 120.]

albida (Braun, 1893) Rail., 1898, 172 (in cat and dog).

buski (Lankester, 1857) Rail., 1898, 172.

complexa (Stiles & Hass., 1894) Rail., 1898, 172 (in cat).

conjuncta (Cobbold, 1860) Rail., 1898, 172 (in Canidés).

conjuncta (of McConnell, 1875) Rail., 1898, 172 (in man).

conus (Crep., 1825) Rail., 1898, 172 (syn. Amphist. truncatum).—Reported for Phoca grænlandica, P. vitulina.

sinensis (Cobbold, 1875) Rail., 1898, 172.

tenuicollis (Rud., 1819) Rail., 1898, 172 (syns. D. felineum, D. tenuicolle) (in cat and dog, Phoca).

(CEPHALOGONIMUS). [See above, p. 121.]

species Staff., 1902, 481.

CERCAREA Cobbold, 1873c, 14, for Cercaria. [See p. 121.]

CERCARIA. [See above, p. 121.]

appendiculata Pelseneer, 1906, 167, 174–175, 179, pl. 12, figs. 46–52 (in Natica alderi; Boulogne-sur-Mer).

brevicauda Pelseneer, 1906, 167, 179, 184, pl. 9, figs. 10, 11 (in Littorina rudis; Wimereux).

crispata Pelseneer, 1906, 171–172, 179, pl. 11, figs. 39–40 (in Natica alderi; Boulogne).

dentalii Pelseneer, 1906, 170, 171, 181, pl. 11, figs. 34, 35 (in Dentalium tarentinum).

emasculans Pelseneer, 1906, 166, 179, pl. 9, fig. 12 (in Littorina rudis; Wimereux).

giardi Pelseneer, 1906, 170-171, pl. 11, figs. 36-38 (in Buccinum undatum; Boulogne).

migocera Lebour, 1906, 7, for myocerca (in Scrobicularia tenuis).

minor Ben., 1858, 98, see armata minor.

myocercoides Pelseneer, 1906, 162–163, 179, pl. 8, figs. 1–2 (in Syndosmya alba; Boulogne-sur-Mer).

nigrotincta Pelseneer, 1906, 166, 179, pl. 9, fig. 9 (in Syndosmya alba; Boulogne-sur-Mer).

obtusicauda Pelseneer, 1906, 182, ?for obtusicaudata.

obtusicaudata Pelseneer, 1906, 167-168, 179, pl. 9, figs. 16-19, pl. 12, fig. 44 (in Natica alderi; Boulogne).

oocysta Lebour, 1907, 439, 440, 445–446, pl. 10, figs. E, F, G, (in Paludestrina stagnalis; Northumberland Coast).—Nicoll, 1907, 269.

parvirenalis Pelseneer, 1906, 165–166, 179, pl. 8, figs. 7, 8 (in Natica alderi; Boulogne).

pirum Lebour, 1907, 439, 440, 446–447, pl. 9, figs. F, G, (in Paludestrina stagnalis; Northumberland Coast).—Nicoll, 1907, 269.

seminis Owen, 18351, 394.

species Lebour, 1906, 7 (in Donex vittatus, Tellina tenuis; Alnmouth Land).

species Lint., 1905d, 333, 401 (in Monacanthus hispidus; Beaufort, N. C.). syndosmyæ Pelseneer, 1906, 172–173, 179, 185, pls. 10, 11, figs. 20, 21, 23 in Syndosmya alba; Boulogne-sur-Mer).—Giard, 1907, 419 (in Syndosmya alba).

ubiquita Lebour, 1907, 439, 440, 444–445, pl. 9, figs. D. E. (in Littorina obtusata, L. rudis, Paludestrina stagnalis; Northumberland Coast).

vaullegeardi Pelseneer, 1906, 175–176, 179, 180, pl. 11, figs. 41–43, 45 (in Trochus cinerarius; Wimereux).

CHELONELLA Ben. & Hesse, 1863. S0.—Mont., 1907, 6, syn. of Encotyllabe. QHIORCHIS. [See above, p. 135.]

lunatus (Dies., 1836) Daday, 1907, 472.

papillatus Daday, 1907, 520-525, 534, 535, 536, 537, 538, 541, 542, 543, 545, 546, 547, 548, 550, 551, 552, 554, 555, 556, 557, 558, 561, 562, 564, 566, 567, 568, 570, 571, 572, 579, 581, 582, pl. 26, figs. 16, 17, 21, 24, 25, pl. 27, figs. 3, 4 (t. h. Colossoma brachypoma; Paraguay; also in Doras murica, Salmo pacu).

CLINOSTOMUM. [See above, p. 137.]

commutatum (Dies., 1850) Rail., 1898, 173 (in pigeonneaux, poulets).

heterophycs (Sieb., 1853) Rail., 1898, 173 (in chien, homme).

westermanni (Kerbert, 1871) Rail., 1898, 173 (in chat, chien, homme).

DACTYLOCOTYLE. [See above, p. 144.]

species Lint., 1905d, 333, 352, pl. 20, fig 151 (in Brevoortia tyrannus: Beaufort, N. C.).

DACTYLOGYRUS. [See above, p. 145.]

siluris glanidis Hofer, 1904a, 169, for siluri glanidis.

species Olss., 1883a, 53 (in Coregonus lavaretus).

DIACROCŒLIUM Neumann, 1897f, 673, misprint for Dicrocœlium.

DINURINÆ Looss, 1907, May 14, 592-593 (subf. of Hemiuridæ, contains: Dinurus, Ectenurus); 1907, 69, 99, 111-112, 125.

DINURUS Looss, 1907, May 14, 593 (tod. tornatus) [nec Dinoura Ashmead]; Dinurinæ, Hemiuridæ; 1907, 69, 99, 112-117, 123, 131

barbatus (Cohn, 1902) Looss, 1907, 594–596 (in Coryphæna equisetis, C. hippurus, Pelamys sarda); 1907, 118, 121, pl. 8, fig. 11, pl. 9, figs. 19, 20, 21.

breviductus Looss, 1907, May 14, 596 (t. h. Pelamys sarda; Atlantic Ocean; also in Coryphæna hippurus, at Beaufort, N. C.); 1907, 118–119, 120, 121, 122, pl. 8, fig. 12, pl. 10, figs. 22, 23.

longisinus Looss, 1907, May 14, 596 (in Coryphæna hippurus; Red Sea, near Aden); 1907, 86, 119-123, pl. 8, fig. 13, pl. 10, figs. 24, 25.

tornatus (Rud., 1819) Looss, 1907, 593-594 (in Coryphena equisetis, C. hippurus, (t. h.) Pelamys sarda; Beaufort, N. C., Atlantic Ocean); 1907, 69, 112, 117, 120, 121, 122, pl. 8, figs. 9, 10; pl. 9, figs. 17, 18.

DIPLODISCUS. [See above, p. 155.]

cornu (Dies., 1839) Daday, 1907, 478–481, 527, 530, 533, 545, 546, 547, 549, 550, 551, 552, 554, 565, 568, 576, 581, pl. 24, figs. 11–15 (in Cataphractus vaca [= Doras] vaca; Rio Branco).

marenzelleri Daday, 1907, 473–478, 481, 527–530, 531, 532, 533, 546, 547, 548, 549, 550, 551, 552, 554, 564, 565, 568, 572, 576, 581, pl. 24, figs. 4–10 (in Salmo sp.; loc. South America, same as Amphist. oxycephalum).

DIPLOSTOMIASIS Hofer, 1904a, 138ff, refers to infection with Diplostomum. DIPLOSTOMUM. [See above, p. 156.]

annuligerum (Nordm., 1832) Hofer, 1904a, 295, 296, fig. 189 (in Perca fluviatilis).

petromyzi fluviatilis Dies., 1858e, 316, for peteromyzi fluv.

DISTOMA. [See above, p. 159.]

betencourti Mont., 1892b, 127, pl. 8, fig. 12.

burki Rathelot, 1892a, 14 for buskii.

cignoides Desmonceaux, 1868a, 21, for cygnoides.

cylindriaceum Daday, 1907, 560, for cylindraceum.

fenestratum Lint., 1907, 111-112, pl. 12, figs. 86-91 (in Lycodontis moringa; Bermuda); includes Dist. sp. Lint., 1904, 373, figs. 213-214.

folium Olfers, 1816. [See supra, p. 190.]—Zschokke, 1884, 11, 50–52, pl. 2, fig. 11 (in Cottus gobio, Salmo umbla, Thymallus vulgaris, Trutta variabilis); 1896, 773, 774, 776, 780, 783, 820 (in Trutta fario, Cottus gobio).

gyrinus Lint., 1907, 107-108, pl. 10, figs. 72-74 (in Lactophrys tricornis, L. trigonus; Bermuda).

hæmatodium Dewitz, 1892b, 106, for hæmatobium.

hæmatobium Rathelot, 1892a, 14, 15, for hæmatobium.

lamelliforme Lint., 1907, 108-109 (in Balistes carolinensis, Lactophrys tricornis, L. trigonus; Bermuda).

levenseni Lint., 1907, 110, pl. 12, figs. 80-83 (in Epinephelus maculosis, E. striatus; Bermuda).

muehlingi Nicoll, 1907, 257, 259, for mühlingi.

noverca (Braun, 1902) Verdun, 1907, 274.

ophtalmobium Rathelot, 1892a, 14, for ophthalmobium.

pankreatikum Katsurada & Saito, Hyg. Centralbl., v. 3, Oct., 279, for pancreaticum.

preistis Mola, 1907, 39, for pristis.

quadrangulatum Daday, 1907, 470-472, pl. 24, figs. 1-3 (in Salmo pacu=Mycetes bidens; Cuyaba).

rajazzii Ariola, 1899, 135 to (Polyorchis), for ragazzii.

ranerium Nardo, 1833a, 523, for raynerianum.

similis (Jægers., 1900) Lebour, 1906, 6.

DISTOMA—Continued.

- species Hausmann, 1899a, 448, 450 (in Gallus dom.).
- species Hausmann, 1899a, 449 (in Corvus frugilegus.
- species Schreder, Hofer, 1904a, 285 (in Esox lucius).
- species Lebour, 1905, 1-3, figs. A, C (in Cardium edule).
- species Lebour, 1907, 102-104, pl. 7 (in Cardium edule; England).
- species Lebour, 1907, 105-106, pl. 8, figs. C-D (in Patella vulgata).
- species Lebour, 1907, 104-105, pl. 8, figs. A, B (in Purpura lapillus; England).
- species Lint., 1905d, 335, 389, pl. 23, figs. 168, 169, 170 (in Bairdiella chrysura; Beaufort, N. C.).
- species Lint., 1905d, 335, 403, pl. 29, fig. 208 (in Chilomycterus schæpfi; Beaufort, N. C.).
- species Lint., 1905d, 335, 374 (in Coryphæna equisetis; Beaufort, N. C.).
- species Lint., 1905d, 335, 373, pl. 30, figs. 213, 214 (in Coryphæna hippurus; Beaufort, N. C.).
- species Lint., 1905d, 335, 385 (in Cynoscion regalis; Beaufort, N. C.).
- species Lint., 1905d, 335, 350, pl. 29, fig. 209 (in Galeichthys milberti; Beaufort, N. C.).
- species Lint., 1905d, 335, 382, pl. 24, fig. 179 (in Lagodon rhomboides; Beaufort, N. C.).
- species Lint., 1905d, 335, 393 (in Leiostomus xanthurus; Beaufort, N. C.).
- species Lint., 1905d, 335, 415 (in Lophopsetta maculata; Beaufort, N. C.).
- species Lint., 1905d, 335, 360 (in Menidia menidia; Beaufort, N. C.).
- species Lint., 1905d, 335, 397 (in Micropogon undulatus; Beaufort, N. C.).
- species Lint., 1905d, 335, 410, pl. 22, fig. 167, pl. 29, fig. 205, pl. 30, fig. 215 (in Opsanus tau; Beaufort, N. C.).
- species Lint., 1905d, 335, 413, 414 (in Paralichthys albiguttus; Beaufort, N. C.).
- species Lint., 1905d, 335, 404, pl. 21, fig. 157 (in Prionotus scitulus: Beaufort, N. C.).
- species Lint., 1905d, 335, 349, pl. 30, fig. 210 (in Pteroplatea maclura; Beaufort, N. C.).
- species Lint., 1905d, 335, 372, pl. 23, figs. 171, 172 (in Rachycentron canadus; Beaufort, N. C.).
- species Lint., 1905d, 335, 364, pl. 29, figs. 206, 207 (in Seriola lalandi; Beaufort, N. C.).
- species Lint., 1905d, 335, 359 (in Siphostoma fuscum; Beaufort, N. C.).
- species Lint., 1905d, 335, 402, pl. 22, fig. 165 (in Spheroides maculatus; Beaufort, N. C.).
- species Lint., 1905d, 335, 361, pl. 30, fig. 211 (in Sphyræna borealis; Beaufort, N. C.).
- species Lint., 1905d, 335, 353, pl. 22, fig. 166 (in Stolephorus brownii; Beaufort, N. C.).
- species Lint., 1905d, 335, 416, pl. 22, figs. 161, 162, 163, 164, pl. 30, fig. 212 (in Symphurus plagiusa; Beaufort, N. C.).
- species Lint., 1905d, 335, 366, pl. 29, fig. 204 (in Trachinotus carolinus; Beaufort, N. C.).
- species Lint., 1907, 114, pl. 7, fig. 57 (in Angelichthys ciliaris; Bermuda).
- species Lint., 1907, 117, pl. 12, fig. 84 (in Balistes carolinensis; Bermuda).
- species Lint. 1907, 116, pl. 10, fig. 70 (in Bodianus fulvus punctatus; Bermuda).
- species Lint., 1907, 115-116, pl. 10, fig. 69 (in Chætodon sp.; Bermuda).
- species Lint., 1907, 117, pl. 13, fig. 85 (in Paranthias furcifer; Bermuda).
- species Lint., 1907, 118 (in Salarichthys textilis; Bermuda).
- species Lint., 1907, 114, pl. 8, figs. 59-60 to (Lecithocladium) (in Seriola dumerili; Bermuda).

DISTOMA—Continued.

species Lint., 1907, 113, pl. 7, figs. 55-56 (in Seriola fasciata; Bermuda).

species Lint., 1907, 116, pl. 10, fig. 70 (in Sphryæna sphryæna; Bermuda).

species Lint., 1907, 118 (in Teuthis corruleus; Bermuda).

species Lint., 1907, 114-115, pl. 8, fig. 61 (in Teuthis hepatus; Bermuda).

species Lint., 1907, 115, pl. 8, fig. 62 (in Tylosurus acus; Bermuda).

species Olss., 1876a, 148 (in gall bladder of Larus argentatus).

subtenue Lint., 1907, 106, pl. 9, fig. 65 (in Calamus calamus, Harpe rufa, Iridio bivittatus, Lachnolaimus maximus; Bermuda).

tomex Lint., 1907, 112-113, pl. 14, figs. 94-96 (in Epinephelus striatus; Bermuda).

trulla Lint., 1907, 109, pl. 9, fig. 79 (in Ocyurus chrysurus; Bermuda).

DUCROCŒLIUM Neumann, 1897f, 659, misprint for Dicrocœlium.

ECHINOSTOMUM. [See above, p. 246.]

gazzettæ Arch. f. Naturg., 1901, v. 2 (8), 188, for garzettæ.

putorii (Schrank,? or Gmelin, 1790) Rail., 1898, 172 (syn. Fasc. trigonoce-phala) (in dog).

species Hausmann, 1899a, 448, 449, 452 (in Corvus frugilegus). species Wolffhuegel, 1900, 35.

ECTENURUS Looss, 1907, May 14, 596-597 (tod. lepidus); Dinurine, Hemiuridæ; 1907, 69, 99, 123-124, 131, 153, 167.

lepidus Looss, 1907, 597 (t. h. Lichia amia; also in Atherina hepsetus. Caranx trachurus, Cepola rubescens, Lophius piscatorius, Mæna vulgaris, Scomber colias, Smaris alcedo, Trachypteron tænia); 1907, 69, 123, 124, pl. 10, figs. 26, 27, pl. 13, fig. 46.

ENCOTYLLABE. [See above, p. 251.]

paronæ Mont., 1907, 3, 4, 6, 7, 9-10, pl. 10, figs. 13-15 (in Crenilabrus pavo; Genova), E. sp. of Braun, Par. & Per., St. Remy.

species Lint., 1907, 103, pl. 7, figs. 49–53 (in Calamus calamus; Bermuda). vallei Mont., 1907, 3. 4, 6, 7, 10–11, pl. 10, figs. 4–9, 16 (in Chrysophrys aurata; Triest).

FASCIOLA. [See above, p. 254.]

egyptiaca Pease, 1901b, 7, for agyptiaca.

jacksoni Cobbold, 1869a, 80, for jacksonii.

jaksoni Stazzi, 1900, 3, 4, 5, for jacksonii.

magna (Bassi, 1875) Sons., 1889, 275.

species Merrem, 1781, 169–172, pl. 1, figs. 3–7 (in mouse) "Sack-Egels."—Stiles & Stev., 1905a, 10 syn. of Tænia teniæformis.—Stiles, 1906a, 43, syn. of Cysticercus fasciolaris.

GASTEROSTOMUM. [See above, p. 270.]

baculatum Lint., 1907, 119, ?for baculum.

gracilius Pelseneer, 1906, 176, for gracilescens.

species Braun, 1893a, 866 (in Belone vulgaris, Cardium edule, C. rusticum, Ostrea edulis, Rochen, Haie).

species Lint., 1907, 119 (in Mycteroperca apua; Bermuda).

species Lint., 1905d, 335, 379 (in Orthopristis chrysopterus; Beaufort, N. C.).

species Lint., 1905d, 335, 364 (in Seriola lalandi; Beaufort, N. C.).

species Olss., 1869b, 498.

species Tennent, 1906, 640 (in Scomberomorus maculatus).

species Tennent, 1906, 640, 679, 682 (in Tylosurus marinus).

species Ziegler, 1883, 539 (in Cyclopterus lumpus).

GASTRODISCUS. [See above, p. 273.]

watsoni (Conyngham, 1904) Verdun, 1907, 285, fig. 110.

GASTROSTOMUM Otto, 1896a, 122, for Gasterostomum.

GYMNOPHALLUS. [See above. p. 276.]

dipsilis Nicoll, 1907, 247, 263-265 (in Oidemia fusca, O. nigra; Great Britain).

margaritarium (Dubois, 1901) Dubois, 1907, 502-504 (in Mytilus edulis, M. gallo-provincialis).—Giard, 1907, 419.

GYRODACTYLIASIS Hofer, 1904a, 134ff, name of infection.

HEMIURUS. [See above. p. 282.]

rugosus Looss, 1907. May 14, 591 (in Clupea pilchardus, C. sardina, Rhombus maximus; Triest); 1907. 105-106, 159, pl. 7, figs. 4, 5 (syn. H. stossichi Luehe, not Apoblema st. Mont., 1891).

HEXATHYRIDIUM. [See above, p. 286.]

sanguicola delle Chiaje, 1825a, 15 (syn. of Polyst. venarum).

HOLOSTOMUM. [See above. p. 289.]

cuculus Thoss. 1897, 1-66, pls. 1-2 (in Bursa fab. of Larus ridibundus; Kiel).—Kopczynski, 1907, 642.

HOLOSTONUM Thoss. 1897, 12, for Holostomum.

HOMALOGASTER. [See above, p. 294.]

philippinensis Stiles & Goldberger, 1908; 25-30, figs, 29-44 (in Bos sp.; Manila, P. I., Phrapatoom, Siam).

ITYOGONIMUS. [See above. p. 204.]

filum Looss, 1907, 606-607, figs. 2a-b (in Talpa europæa: Leipzig).

LECITHASTER. [See above. p. 295.]

galeatus Looss, 1907, May 14, 607 (in Mugil auratus; Egyptian Coast); 1907, 165-166, pl. 15, figs. 73, 74.

stellatus Looss, 1907, May 14, 606-607 (in Belone acus, Mæna vulgaris; Triest); 1907, 165, pl. 14, fig. 60, pl. 15, figs. 75, 76 (in B. a., Dentex vulg., M. v.).

LECITHOCHIRIUM. [See above. p. 295.]

gravidum Looss, 1907, May 14, 603 (t. h. Anguilla vulgaris; also in Gobius capito, Platessa passer, Rhombus maximus; Triest); 1907, 142, 148-149, pl. 14, figs. 60, 61, 63 (in A. v., Conger conger, Corvina nigra. G. c., Hippocampus guttulatus, Labrax lupus, P. p., R. m., Syngnathus acus; Triest).

LECITHOCLADIUM. [See above, p. 296.].

crenatum (Mol., 1859) Looss, 1907, 509 (in Centrolophus pompilius), perhaps n. sp.: 1907, 134-135, 169 (in C. p.: Triest).

cristatum (Rud., 1819) Looss, 1907, 598-599 (in Dactylopterus volitans;
 Naples: Stromateus fiatola; Rimini, Triest, Naples): 1907, 133-134,
 135, 169, pl. 11, figs. 30, 31, pl. 13, figs. 44, 45 (in S. f.: Triest).

gulosum (Lint., 1901) Looss, 1907, 135 (in Rhombus (Stromateus, triacanthus; Woods Hole, U. S. A.).

LEVINSENIELLA. [See above. p. 299.]

macrophallos (Linst., 1875) Jægers., 1907, 148,

opaca (Ward, 1894) Jægers, 1907, 148.

pellucida Jægers., 1907, 134, 136, 141–144, 147–148, 150, 151, figs 6, 7 (in Anas boschas fera, Fuligula fuligula; West Coast of Sweden).

propinqua Jægers., 1907, 135-139, 140, 141, 142, 143, 144, 147-148, 150, figs.
 1, 2, 3, 4, 5 (in charadrius hiaticula, Hæmatopus ostralegus; West Coast of Sweden).

MARITREMA Nicoll, 1907, 246, 265-266 (tod. gratiosum).

gratiosum Nicoll. 1907, 247, 248, 265, 266-267, 268, 269 (in Ægialitis hiaticula, Hæmatopus ostralegus, Larus ridibundus, (t, h.) Pelidna (Tringa) alpina; Great Britain).

humile Nicoll, 1907, 247, 266, 268-269 (in Totanus calidris; Great Britain), lepidum Nicoll, 1907, 247, 266, 267-268 (in Larus argentatus; Great Britain),

MESOGONIMUS. [See above. p. 302.]

columbæ (Mazzanti, 1889) Mazzanti, 1890, 139,

MICROCOTYLE. [See above, p. 304.]

species Lint., 1905d, 335. 385 (in Cynoscion regalis; Beaufort. N. C.).

species Lint., 1905d, 335, 370, pl. 20, figs. 147-150 (in Pomatomus saltatrix; Beaufort, N. C.).

species Lint., 1907, 103-104 (in Calamus calamus, Diplodus sargus; Bermuda).

MICRORCHIS Daday, 1907, 481-482, 495, 526, 527, 528, 529, 530, 531, 533, 534, 535, 536, 538, 539, 540, 542, 545, 550, 552, 566, 567, 579, 580, 581 (type by present designation, on page precedence, megacotyle).

ferrum-equinum (Dies. 1836) Daday, 1907, 484, 485, 487, 488-494, 528, 530, 532, 533, 535, 537, 538, 539, 546, 547, 549, 553, 554, 556, 559, 561, 565, 576, 578, 581, pl. 24, figs. 27-29; pl. 25, figs. 1-6 (syn. Amphist. ferrum-equinum) (in Cataphractus corome, C. murica).

megacotyle (Dies., 1836) Daday, 1907, 482–488, 494, 527, 530, 532, 537, 546, 547, 549, 554, 561, 563, 565, 576, 578, 581, pl. 24, figs. 16–26 (syn. Amphist, megacotyle) (in Silurus palmito; Mattogrosso).

MONOSTOMUM. [See above, p. 308.]

lophocerca (Fil., 1857) Lebour, 1907, 439, 440, 443-444 (in Bythinia tentaculata, Paludestrina stagnalis; England).

species Lint, 1907, 118-119, pl. 14, figs. 92-93 (in Bathystoma striatum, Hæmulon flavolineatum; Bermuda).

species Nicoll, 1907, 247 (in Larus argentatus).

species Nicoll, 1907, 247 (in Oidemia fusca).

NITYCHIA Nord, 1833b, 384, for Nitzschia.

NITYSCHIA Nord., 1833b, 376, for Nitzschia.

OCTOBOTHRIUM. [See above, p. 324.]

melangi Lebour, 1907, 442, for merlangi.

species Olss., 1876a, 148 (in Thymallus vulgaris).

OPHECONA Looss, 1907, May 14, 616 (tod. bacillare).—Poche, 1907, Aug. 20, 103.

bacillare (Mol., 1859) Looss, 1907, May 14, 616.

PACHYTREMA Looss, 1907, 610 (tod. calculus).

calculus Looss, 1907, 610-613, figs. 3-4, Opisthorchiiden (in Larus argentatus, L. ridibundus; Triest).

PARAGOMINUS Daniels & Stanton, 1907, 343, for Paragonimus.

PARAGONIMUS. [See above, p. 335.]

heterophyes (Sieb., 1853) Neveu-Lemaire, 1904a, 73, 88, 89.

PARAPHISTOMUM Daniels & Stanton, 1907, 342, for Paramphistomum.

PARORCHIS. [See above, p. 337.]

acanthus (Nicoll, 1906) Nicoll. 1907, 247 (in Larus argentatus, L. canus).

PHYLLINE. [See above, p. 338.]

endorfii Par. & Per., 1895, 2. for hendorfii.

PHYLLODISTOMUM. [See above. p. 339.]

angulatum Linst., 1907, [2] 202. 1 fig. (in Lucioperca sandra; Wolga).

PLACOTREMA Looss, 1907, May 14, 615 (tod. inversum).

inversum Looss, 1907, May 14, 615 (in Chrysophrys aurata; Triest).

PLATYNOSUM Looss, 1907, 607 (tod. semifuscum).

semifuscum Looss, 1907, 607-608, fig. 3 (in Circetus [=Circætus?] gallicus: Genoa).

PLERURUS Looss, 1907, May 14, 604-605 (tod. digitatus), Sterrhurine, Hemiuride; 1907, 69, 78, 100, 152-153, 154.

digitatus (Looss, 1899) Looss, 1907, 605 (in Sphryæna vulgaris; Red Sea, Sawakin); 1907, 69, 152, 153-154, pl. 15, figs. 68, 69.

POLYSTOMA. [See above, p. 350.]

paguri Nord., 1833b, 379, see Bull. Soc. Philom., 1811, no. 44, 271, t. 2.

POLYSTOMIDÆS Mont., 1888a, 89, for Polystomidæ.

PRISTISOMUM Looss, 1907, May 14, 615 (tod. pumex).

caducum Looss, 1907, May 14, 615 (in Umbrina cirrhosa; loc. not given).—Poche, 1907, Aug. 20, 103.

pumex Looss, 1907, May 14, 615 (in Caranx trachurus; loc. not given).

PSEUDOCLADORCHIS Daday, 1907, 481 494-495, 526, 527, 528, 529, 530, 532, 533, 534, 536, 537, 541, 547, 549, 550, 552, 553, 554, 561, 565, 566, 567, 576, 580, 581 (type by present designation, on page precedence, cylindricus).

cylindricus (Dies., 1836) Daday, 1907, 495-501, 502, 503, 504, 506, 512, 530, 535, 538, 546, 554, 566, 579, pl. 25, figs. 7-16 (in Cataphracta murica, Salmo pacupeba, S. sp., Silurus megacephalus; Villa Maria, Brazil).

macrostomus Daday, 1907, 506-509, 530, 531, 535, 538, 546, 554, 566, pl. 26, figs. S-12 (in Salmo pacupa, S. sp.; loc. South America, same as Amphist. oxycephalum).

nephrodorchis Daday, 1907, 501–506, 508, 509, 530, 535, 538, 546, 554, 566, pl. 25, figs. 17–22; pl. 26, figs. 1–7 (in Salmo pacu, S. pacupa, S. sp.; loc. South America, same as Amphist. oxycephalum).

PTYCHOGONIMUS. [See above, p. 357.]

volgensis Linst., 1907, 201 [1] (in Lucioperca sandra; Wolga).

SCHISTOSOMUM. [See above, p. 359.]

hæmatobium japonicum (Katsurada, 1904) Verdun, 1907, 290, 311, 313 (syn. of S. cattoi).

SCUTARIELLA Mrazek, 1907, 1-6, 1 pl., figs. 1-6 (m. didactyla).

didactyla Mrazek, 1907, 1-6, pl. 1, figs. 1-6 (in Atyaëphyra desmarestii; Moraca-Flusses, Scutarisee bei Plavnica).

SPOROCYSTIS. [See above, p. 362.]

cotti Zschokke, 1884, 6, 11, 56-58, pl. 2, fig. 13 (in Cottus gobio).

STERRURHINÆ Looss, 1907, 99, for Sterrhurinæ, see p. 364.

STROMYLOTREMA Poche, 1907, Aug. 20, 105, for Stomylotrema, see p. 365.

SYNCŒLIUM. [See above, p. 366.]

raggazzii Janicki, 1907, 719, for ragazzii.

TRISTOMA. [See above, p. 375.]

inermis Goto, 1899a, 273, see læve var. inermis.

UDONELLA. [See above, p. 381.]

caligarum Ben., 1858a, 1861a, 12, 13–18, pl. 1, figs. 1–15, for caligorum.

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Public Health and Marine-Hospital Service of the United States

Walter Wyman, Surgeon-General

HYGIENIC LABORATORY.—BULLETIN No. 38

M. J. ROSENAU, Director June, 1907

THE INFLUENCE OF ANTITOXIN UPON POST-DIPHTHERITIC PARALYSIS

By

M. J. ROSENAU

and

JOHN F. ANDERSON



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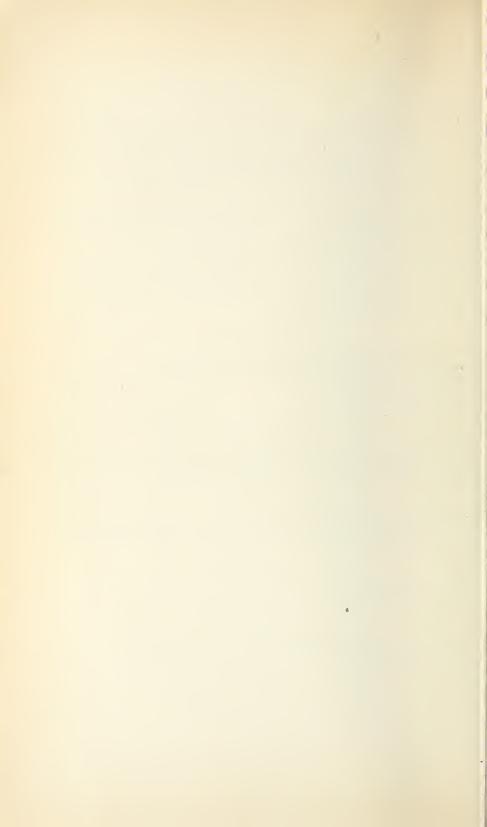
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CONTENTS.

| | Page |
|--|------|
| Introduction | 7 |
| Post-diphtheritic paralysis in man | 11 |
| The relation of post-diphtheritic paralysis to antitoxin | 12 |
| Post-diphtheritic paralysis in the guinea pig. | 14 |
| Can antitoxin influence diphtheritic paralysis after the paralysis has | |
| appeared? | 16 |
| Can antitoxin influence diphtheritic paralysis when injected shortly before | |
| the paralysis develops? | 20 |
| How long after infection can antitoxin influence paralysis? | 20 |
| Can antitoxin, given before infection as a prophylactic, prevent post-diphthe- | |
| ritic paralysis? | 30 |
| Summary and conclusions | 31 |
| | |



THE INFLUENCE OF ANTITOXIN UPON POST-DIPHTHERITIC PARALYSIS."

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INTRODUCTION.

Diphtheria, under various names, can be traced back to remote antiquity; but the specific contagious disease which we now call diphtheria was not differentiated until 1821, when Brétonneau be clearly described its clinical features and gave to it its present name. However, a definite relation between faucial attacks and subsequent paralysis was demonstrated almost one hundred years before.

Chomel, dealing with the epidemic of Paris in 1743, Ghisi, with that at Cremona in 1749, and Samuel Bard with that of New York in 1789, described cases of paralysis following sore throat. It is a surprising fact that nearly a century elapsed before the remarkable observations of these three writers were confirmed by subsequent investigators.

Brétonneau, in 1855, recorded an instance of post-diphtheritic paralysis in the case of Horpin, a surgeon of the hospital at Tours.

a Manuscript submitted for publication June 14, 1907.

b Brétonneau: (a) Communication to the French Academy, 1821. (b) Des inflammations spéciales des tissus muqueux, et en particulier, de la diphtérite, ou inflammation pelliculaire. Paris, Crevot, 1826, 540 p., illus. 8°.

^c These historical data are taken largely from J. D. Rolleston's excellent "Clinical observations on diphtheritic paralysis," The Practitioner, London, vol. 73, 1904, pp. 597–623, 794–824.

d Ghisi: Lettere mediche. Cremona, 1749.

^e Bard, Samuel: An inquiry into the nature, cause and cure of the angina suffocativa, etc. Trans. Phil. Soc., Phila., 1789.

f Fifth memoir on diphtheria, 1855, New Sydenham Society, p. 182.

The first monograph on post-diphtheritic paralysis was by Maingault, of Paris, in 1854, which appeared as an inaugural thesis entitled "De la paralysie du voile du palais à la suite d'angine." Maingault's more important work, "Sur les paralysies diphtéritiques," appeared in 1860. Between 1854 and 1860 several isolated cases of diphtheritic palsies had been described by observers, especially by Trousseau, who by this time had collected 90 cases, 29 of which were in children, with a mortality of 12.

The first histological investigations as to the nature of diphtheritic paralysis were made by Charcot and Vulpian in the case of a woman who had died with palatal paralysis. They showed that the motor nerves alone were affected, and consisted of tubules entirely devoid of medullary substance. The neurilemma contained numerous granular cells, mostly elliptical in shape and in some cases nucleated. A few of the muscle fibers were fatty. The sensory nerves showed no signs of alteration.

In 1883 Klebs ^b found very peculiar and striking bacteria constantly present in the pseudo-membranes in the throats of those dying of true epidemic diphtheria. The next year Löffler ^c isolated these organisms in pure culture and produced more or less characteristic pseudo-membranes by inoculating the cultures upon the mucous membranes of susceptible animals, frequently causing characteristic lesions with death.

In 1888 Roux and Yersin ^d published their notable "contributions to the study of diphtheria." They were the first to prove that paralysis is a common sequel in the lower animals after recovery from experimental diphtheria. They observed the palsies in pigeons and rabbits following pharyngeal and tracheal inoculation. They also observed the same sequel in rabbits recovering from intravenous inoculations. Roux and Yersin concluded that the existence of these palsies following the inoculation of the Klebs-Löffler bacillus completes the resemblance between the experimental and the natural disease and establishes the specific rôle of this bacillus.

The introduction of the antitoxin treatment in 1894, followed by an increase in the number of palsies observed, has led to the publication of several monographs and experimental work dealing with the subject.

a Comp. rend. Soc. biol., 1862.

b Klebs: Congrès de Wiesbaden, 1883. Arch. f. exper. Path., I and IX. Also, "Verhandlungen des Congresses für innere Med.," 1883.

^c Löffler: Untersuchungen über die Bedeutung der Mikroorganismen für die Entstehung der Diphtherie biem Menschen. Mitt. a. d. kais. Gesundheitsamte, Bd. 2, 1884.

^d Roux, E., and Yersin, A.: Contribution à l'étude de la diphtérie. Ann. de l'Inst. Pasteur, vol. 2, 1888, p. 629.

The school founded by Ehrlich a in 1897, based on his scholarly work upon the constituents of the diphtheria poisons, and methods of standardizing the antitoxic value of the curative serum, made much use of the post-diphtheritic paralyses produced experimentally in the guinea pig. Ehrlich believes that at least two primary poisons are produced by the growth and multiplication of the diphtheria bacillus in nutrient broth; the one, toxin, produces acute death within about four days; the other, toxon, is incapable of causing acute death, but is responsible for the late palsies. In accordance with Ehrlich's view, post-diphtheritic paralysis is a toxon poisoning.

Ehrlich b has recently (1906) given a forcible defense of his classical paper entitled "The constituents of diphtheria toxin." On account of its importance in connection with our work, a brief summary of Ehrlich's views bearing upon diphtheritic paralysis or toxon poisoning is given practically in his own words, as follows:

We are evidently dealing with a primary secretory product of the diphtheria bacilli, the "toxon." The toxon possesses the same haptophore group as the toxin, but a weaker affinity for the antitoxin. The main difference is in the toxophore group, for even when given in large doses the toxon does not produce death, but only paralyses, which develop after a long incubation of fourteen days or more.

The presence of an independent poison (toxon) was inferred by Ehrlich from the curves shown in the spectra, which are based upon the assumption that 1 immunity unit contains 200 combining units.

The independent existence of the toxons is further corroborated by the fact that the toxon zone varies enormously in different specimens of poison. In one it may amount to about one-fifth of the toxin portion; in another he has seen equal parts of toxon and toxin. Dreyer and Madsen in fact have recently described a poison which contained three times as much toxon as toxin. According to our present experiences, therefore, the amount of toxon calculated on the toxin can vary from 0 per cent to 300 per cent.

This still left undecided whether the toxon is a primary bacillary secretion or secondary modification of the toxin. A study of the development of one poison (poison V) finally gave Ehrlich a clue to this. (Poison V has been described in the Deut. med. Woch., 1898.)

This particular toxine remained constant, as far as the L+ dose was concerned, but the L⁰ dose increased considerably, from 0.125 to 0.21. Ehrlich explains this phenomenon by assuming that the toxin portion remained absolutely unchanged, as indicated by the constancy of the L+ dose. On the other hand, the toxon portion, which is expressed by the difference between the L+ and the L⁰ dose, disappeared. This eliminates the possibility of the transformation of toxin into toxon.

It is difficult to say, a priori, what becomes of the toxon which has disappeared. Ehrlich assumes that we are dealing with a formation of an analogue of toxoid, viz, a substance which he terms toxonoid. This he conceives to be a toxon in which the toxophore group has become modified.

a Ehrlich, P.: Die Wertbemessung des Diphtherieheilserums und deren theoretische Grundlagen. Klin. Jahrb., Jena, v. 6 (2), 1897, pp. 299–326.

^{——:} Ueber die Constitution des Diphtheriegiftes. Deut. med. Woch., Leipzig, v. 24 (38), 1898, pp. 597-600.

b Ehrlich, Paul: Studies on immunity. XXXVII, The constituents of diphtheria toxin. New York, John Wiley & Sons, 1906, p. 481.

²⁷⁴⁸⁻No. 38-07-2

Another fundamental difference—one which in Ehrlich's opinion argues in favor of the individuality of toxin and toxon—consists in the different action of the two constituents. The action of diphtheria toxin, as is well known, is such that the animals die with lesions of hydrothorax, ascites, congestion of the suprarenals, necrosis of the skin. Somewhat smaller doses kill guinea pigs in from six to seven days, the L. D.—no longer produce death, but regularly cause necroses, which are surrounded by an extensive area of total loss of hair. Small fractions of the fatal dose always produce emaciation of the animals. In contrast to this, the toxon—i. e., a serumpoison mixture in which only the toxin fraction is completely neutralized—never kills animals acutely, even in high doses. The inflammatory properties may be entirely absent in small doses, while in large doses they are present to only a slight degree. The ædema disappears completely in the course of a few days, there are no necroses, and the loss of hair, if it occurs at all, is only partial. On the other hand, the paralyses are very characteristic, and these appear at any time between the fourteenth and twentieth days, depending upon the dose, usually in the third week. Frequently the animals do not show even a trace of local reaction, and maintain their weight; then suddenly they are attacked with the paralyses and may die from these within a few days. Ehrlich has never seen such a result in animals inoculated with a pure diphtheria poison. Now and then a guinea pig was observed which showed these paralytic phenomena. It was usually one that had received a considerable fraction of the L. D. Invariably it showed extensive necroses, was generally very sick from the beginning, and had suffered considerable loss of weight. In view of the slight amount of toxon which was found in these poisons, such animals were evidently supersensitive to the toxon.

Ehrlich believes that the diphtheria poison contains at least three different varieties of poison and that these possess different affinities and different actions. These poisons are:

- 1. Toxin, possessing the highest affinity, kills rabbits and guinea pigs acutely, but is more toxic for the former.
 - 2. Toxon, killing rabbits acutely and guinea pigs with symptoms of paralysis.
 - 3. Toxonoid, producing paralyses in rabbits; nontoxic for guinea pigs.

The fact that all three poisons act more strongly on rabbits than on guinea pigs is explained by the absolute higher susceptibility of the former.

Upon the constituents of diphtheria toxin Ehrlich sums up his views as follows:

- 1. The diphtheria bacillus produces several kinds of poison, especially toxins and toxons.
 - 2. The affinity of diphtheria toxin to the antitoxin is very great.
- 3. The deviations from a straight line as they manifest themselves in the graphic representation of the neutralization of the poison can not be explained by the assumption of a single poison possessing a weak affinity. They are rather the expression of the fact that the poison bouillon contains admixtures of various kinds of substances of a toxoid character.
- 4. The varied affinity of the toxoids can not be explained by the assumption that a simple toxin when transformed into toxoid suffers a change in affinity either positively or negatively. Rather does this indicate that the toxic bouillon contains, preformed, various toxins of different affinities.
 - 5. There is no change in the haptophore group in the formation of toxoid.
- 6. The absolute number of combining units contained in the immune unit or in the L⁰ dose of poison is 200.

POST-DIPHTHERITIC PARALYSIS IN MAN.

Paralysis probably follows in one-fourth of all cases of diphtheria that recover. The palsies are frequently so slight in degree and extent that they may readily be overlooked. Of 50,851 cases observed from 1895 to 1902 in the hospitals of the Metropolitan Asylums Board 19.54 per cent developed post-diphtheritic paralysis.

Post-diphtheritic paralysis follows both mild and severe cases; even fatal paralysis may follow an ambulatory case.^a Both sexes are equally affected. While the palsies follow the attacks of diphtheria in all ages, clinicians differ radically as to the relative frequency of the sequel at different ages.

The paralysis may affect the heart, palate, ciliary muscle (loss of accommodation), ocular muscles (strabismus), the pharynx, lips, diaphragm, or any of the skeletal muscles. It is sometimes localized,

sometimes general.

The paralysis may more properly be described as a palsy or paresis, as the loss of power is seldom complete. It usually appears about the second or third week, sometimes as late as the eighth week. According to Rolleston, early paralysis indicates a profound intoxication of the nervous system and is a bad prognosis, early involvement of the palate being often associated with cardiac paralysis. This corresponds precisely with our observations upon the guinea pig. Paralysis appearing early in the guinea pig—between the thirteenth and eighteenth days following infection—is almost invariably severe and fatal; paralysis appearing later is commonly benign.

The tendency of diphtheritic palsy is to recover. When death occurs it is usually within the first three weeks, and is commonly due to cardiac paralysis. The figures indicate that from 2 to 4 per cent of the total number of cases of diphtheria die as a result of post-

diphtheritic paralysis.

A precocious form of palatal palsy occurring about the fifth day of the disease or earlier has been described. Deguy^b believes these early palsies of the palate to result from the local effect of inflammatory reactions other than diphtheria. The presence of large numbers of diplococci, both in the leucocytes and in the thrombosed capillaries of the part, adds weight to the belief that the action is a diplococcemia superadded to a diphtheritic intoxication.

Rolleston believes that precocious palatal palsy in diphtheria is almost invariably associated with malignant forms, as is shown by

b Rev. neurology and psychia., vol. 4, Sept., 1906, p. 614.

a This corresponds to our work upon guinea pigs. Mixtures containing large doses of toxin partially saturated with antitoxin, when injected into guinea pigs. may cause little or no acute symptoms, but there may be enough uncombined "toxon" to cause severe or fatal post-diphtheritic paralysis.

the high mortality, the association of other grave symptoms during the acute stage, and subsequent more frequent development of paralysis in convalescence in the cases in which it occurs. It resembles the ordinary forms of diphtheritic palsy in its tendency to be frequently incomplete and by its higher incidence among young persons. It is, as a rule, of much longer duration than the palatal affection which occurs at a later date.

THE RELATION OF POST-DIPHTHERITIC PARALYSIS TO ANTI-TOXIN.

The statement is frequently made that paralysis is even more common in the cases treated with antitoxin than under former methods of treatment.

Welch^a, 1895, thinks this doubtful; if true, it may be attributed to the survival of a larger proportion of the cases.

McCollom^b, 1905, drawing from his rich experience with diphtheria at the Boston City Hospital, states that, although paralysis occurs after the severe cases, it has not been so frequent as it would have been in an equal number of cases treated without antitoxin.

Marfan^c, at the Hôpital des Enfants Malades de Paris, where over 1,000 young diphtheria patients are treated annually, is of the opinion that since the introduction of antidiphtheric serum paralysis has become much less frequent. It seems, however, that the cases are not kept under observation long enough at this hospital to warrant such a strong conclusion.

Variot,^d 1898, dismisses the subject briefly by saying that paralyses are not rarer after serum treatment than before, but present the same localization, the same clinical characters, and the same duration as before.

Weill and Deguy^e distinguish two kinds of paralysis: The first including paralysis of the palate and larynx, which they admit is still very frequent, but according to them is less due to diphtheritic intoxication than to local inflammation. The second group includes paralysis of the limbs, eyes, and viscera, which is due to the action of the diphtheria toxine and is much less frequently met with since the introduction of antitoxin.

Rolleston believes that the most satisfactory answer to this question is given by the statistics published annually by the Metropolitan Asylums Board. The yearly admission to its fever hospitals of

^a Welch, W. H.: The treatment of diphtheria by antitoxin. Bull. Johns Hopkins Hosp., July-Aug., 1895, vol. 6, Nos. 52 and 53, p. 1119.

^b McCollom, J. H.: Boston med. and surg. journ., vol. 152, 1905, pp. 621-632.

^cClinique Infantile, May 15, 1904.

^dLa diphtérie et la sérumthérapie, 1898.

e Traitement de la diphtérie, 1902.

diphtheria patients is considerable and each case is, as a rule, kept long enough for paralysis to develop:

| Year. | Number admitted. | | Percentage of paralysis cases. |
|-------|------------------|--------|----------------------------------|
| 1893 | 2,848 | 14.2. | Pre-antitoxin year. |
| 1894 | 3,666 | 13.1. | Only a few cases injected. |
| 1895 | 3,635 | 20.4. | General employment of antitoxin. |
| 1896 | 4,508 | 20.5. | |
| 1897 | 5,637 | 20.55. | |
| 1898 | 6, 566 | 19.42. | |
| 1899 | 7,066 | 20.0. | |
| 1900 | 7,195 | 18.50. | |
| 1901 | 6,926 | 15.0. | |
| 1902 | 6, 534 | 17.07. | |
| 1903 | 5,072 | 17. 1. | |
| 1904 | 4,687 | 14. 8. | |
| 1905 | 4,148 | 12. 4. | |
| 1906 | 5, 218 | 10. 9. | • |

It is plain from the above figures that while the percentage of paralysis during the years following the introduction of antitoxin is higher than in 1893 and 1894, in recent years a decrease is evident; this is probably due to larger dosage and earlier administration of serum. The medical superintendents of these hospitals pointed out in 1895 that there is more than one explanation of the increase first noted. In the first place, cases that before would have died in the acute stage now survive and suffer those palsies which before were found only in the less severe cases. Secondly, the new treatment aroused fresh interest in diphtheria and stimulated more careful clinical observation.

Rolleston expresses the emphatic opinion that antitoxic treatment does not make paralysis more liable to ensue. He states that the early administration of antitoxin makes paralysis less likely to occur, especially in the severe form. He gives the following figures in support of his views:

| Day of the disease. | Paralysis cases. | Percent- age. | Severe forms only. | Percent-age. |
|---------------------|------------------|------------------|--------------------|--------------|
| First day | 1 | 5.5 | 0 | 0.0 |
| Second day | 16 | 15.09 | 4 | 3.7 |
| Third day | 28 | 18.7 | 8 | 5.3 |
| Fourth day | 27 | 28.7 | 12 | 12.7 |
| Fifth day | 21 | 35.0 | 5 | 8.3 |
| Sixth day | 15 | 34.9 | 10 | 23.2 |
| Seventh day | 4 | 19.4 | , 1 | 4.7 |
| Ninth day | 1 | 50.0 | 0 | |
| Eleventh day | . 1 | 50.0 | 1 | 50.0 |
| Thirtieth day | 1 | 100.0 | 1 | 50.0 |
| Total | 115 | | 41 | |

Rolleston fully appreciates and emphasizes the importance of early administration and large dosage in diminishing the incidence, especially in a severe form of subsequent paralysis. In severe cases Rolleston uses 18,000 to 24,000 units, usually repeated once, sometimes twice, by injections on following days. The mildest cases receive early from 3,000 to 12,000 units.

There is no definite proof that post-diphtheritic paralysis ever occurs in cases treated with antitoxin within the first twenty-four hours of the disease. This corresponds precisely with what we see in the guinea pig. Paralysis may follow in cases treated with serum on the second and third day of the disease, again an exact duplicate of the picture we see in the guinea pig.

This emphasizes the importance of administering this soveriegn remedy early. But if the case is not seen early, can larger doses prevent the occurrence of post-diphtheritic paralysis and save life?

It seemed to Ransom, a 1900, worth while to undertake an examination into the conditions under which diphtheritic paralysis occurs in animals and to ascertain by means of experiments what effect, if any, the antitoxin has on this very disagreeable sequel to an attack of diphtheria. As a result of his work Ransom concludes that—

1. Paralysis may certainly be expected after intoxication with not less than onefourth of the fatal dose. With doses between one-fourth and one-eighth paralyses occur but are not constant, and below one-eighth no paralysis was noticed.

2. The larger the dose of toxine the severer will be the paralysis, if the animal

survives long enough.

3. Neutralized mixtures of toxine and antitoxine, containing only about one lethal dose or less, do not appear to cause paralysis.

4. Antitoxine given fifteen to twenty-two hours after intoxication, with doses of toxine not greater than the lethal dose, exercises in large doses a mollifying influence on the subsequent paralysis. This influence is more evident on smaller doses of toxine * than on such as are but little less than the minimal fatal dose. Small doses of antitoxine have no evident effect in diminishing the paralysis.

5. Transferring these results to practice among human beings, we may expect liberal doses of antitoxine, given early in the illness, to influence favorably the subsequent paralysis, and this beneficial influence is likely to manifest itself not so much on the local paralysis (soft palate, etc.) as on such symptoms as failure of the heart. Severe cases are, however, likely to be followed by some paralysis in spite of even large doses of antitoxine.

POST-DIPHTHERITIC PARALYSIS IN THE GUINEA PIG.

Diphtheria in the guinea pig closely resembles the same disease in man. Post-diphtheritic paralysis in the guinea pig is an almost exact counterpart of the same complication in man. We are, therefore,

^a Ransom, F.: Diphtheritic paralysis and antitoxine. Journ. path. and bact., 1900 vol. 6, p. 397.

able to bring forward experimental evidence showing the relation of antitoxin to post-diphtheritic paralysis.

It is now well known that guinea pigs may develop paralysis following the injection of a sublethal dose of toxine or the injection of a toxine-antitoxin mixture in which the toxine is but partially neutralized. As will be seen by our experiments, paralysis may unerringly be produced in guinea pigs with a partially neutralized toxine. We look upon the paralysis as a "toxon" poisoning.

By no means every guinea pig that shows symptoms of postdiphtheritic paralysis dies. Recovery from this sequel in the guinea pig is frequent and depends entirely upon the amount of toxon in the toxine-antitoxin mixture. It is very easy to mix the toxine and antitoxin in such proportion that every guinea pig will develop postdiphtheritic paralysis and die. By adding a little more antitoxin or a little less toxine to the mixture the guinea pigs will develop mild paralytic symptoms from which almost all will recover.

Almost all of our work was done with partially neutralized mixtures containing sufficient toxon to produce early and malignant paralysis. As far as the guinea pig is concerned paralysis can rarely be detected before the fourteenth day. The first indications are a softness of the abdominal muscles near the site of the injection. The weakness gradually spreads to the extremities and finally to the muscles of respiration. In the guinea pig paralysis occurring early is almost always followed by death; symptoms appearing late are apt to be mild, and complete recovery follows.

We can confirm the observations of Lewis,^a in that the paralysis in the guinea pig may be slight, affecting but one limb or somewhat embarrassing the respiration; or, it may be severe and cause sudden or gradual death. It may last for two days or two weeks, or it may terminate fatally at any time. The paralysis is never permanent.

Lewis ^a recently made a study of diphtheria toxone paralysis in the guinea pig, and concludes as follows:

- 1. By grouping and analyzing the records of the guinea pigs used in a large number of routine tests of the strength of recently prepared toxins it is shown that the crude toxins, produced by the same culture of diphtheria bacillus under conditions which are identical within limits which it is possible to determine at the present time, differ greatly in the amount of toxon or paralysis-producing poison which they contain.
- 2. Using old crude toxins which have become relatively stable in composition, in mixtures with antitoxin on the present Ehrlich system, it is shown that—

(a) Guinea pigs are somewhat more susceptible to the action of

^aLewis, Paul Λ.: Diphtheria toxon paralysis in the guinea pig. Journ. med. research, vol. 15, no. 3, n. s., vol. 10, no. 3, Dec., 1906, pp. 469-482.

toxon during the winter months than during the summer. A variation probably dependent on the kind of food supplied and possibly

also on the ventilation of their quarters. .

(b) Guinea pigs of increased resistance to the "toxin" fraction of the poison produced by the diphtheria bacillus also show a wellmarked increase in resistance toward toxon. That is, they are less frequently afflicted with paralysis than are animals of normal resistance when treated with unsaturated but nonfatal toxin-antitoxin mixtures.

3. For guinea pigs of normal resistance at least the nearer the L+dose is approached, supposing the animal to survive the necessary length of time, the greater the chance of paralysis developing.

CAN ANTITOXIN INFLUENCE DIPHTHERITIC PARALYSIS AFTER THE PARALYSIS HAS APPEARED?

Antitoxic serum has been extensively used in France as a curative agent for this sequel and many cases of complete recovery have been reported following its use, after the appearance of paralytic symptoms.

Comby,^a 1904, claims never to have lost a case after its administration. Recently (August, 1906), Comby^b reports further cases illustrating the value of antidiphtheritic serum in the treatment of diphtheritic paralysis. He advocates the free use of the serum whether or not it had been previously used in the treatment of the primary disease. He used 10 to 20 cubic centimeters of the French serum according to age, repeated daily for three, four, or five days if necessary.

Rolleston, commenting on Comby's report, points out that the mortality of post-diphtheritic paralysis has always been very low and intimates that the reported cases of cure after the administration of antidiphtheritic serum would have recovered as quickly without it.

From our own experimental evidence obtained upon guinea pigs it seems to us useless to give diphtheria antitoxic serum to control the paralysis after the paralysis has appeared.

b Comby, J.: Paralysie diphtérique guérie par le sérum. Arch. de med. enf., vol.
 9, no. 8, August, 1906, pp. 480-484.

c Rolleston, J. D.: Lancet, July 7 and Sept. 24, 1996.

^a Comby, J.: Paralysies diphtherique queries par le sérum. Arch. de med. ent., Paris, vol. 7, 1904, pp. 411–417.

^{——:} Traitement des paralysies diphtheriques. Bull. et mem. soc. med. de hop. de Paris, 3 s., vol. 23, no. 21, June 21, 1906, pp. 626-631.

We injected a number of guinea pigs with 400 or 500 units of antitoxin during various stages of paralysis. These pigs had all been given toxine-antitoxin mixtures for the purpose of testing the strength of various serums bought on the open market in accordance with the law of July 1, 1902.^a

It will be seen from Table 1 that the antitoxin apparently had no influence at all to save life or ameliorate the symptoms when given during the stage of paralysis.

^a An act to regulate the sale of viruses, serums, toxins, and analogous products in the District of Columbia, to regulate interstate traffic in said articles, and for other purposes. Approved July 1, 1902.

^{2748—}No. 38—07——3

TABLE NO. 1.—GUINEA PIGS TREATED WITH ANTITOXIN WHILE PARALYZED.

B P=Beginning paralysis. M P=Mild paralysis. P=Paralysis.

400=400 units of antitoxin subcutaneously. 500=500 units of antitoxin subcutaneously. + = Death.C P=Complete paralysis.

| | | | | | | 1 | | | | | | | | | | | | | | |
|------------|--|------------|------------|------------|-----|------|-------|--------|---|------|---------|-------|----------|--------|----------|-----|-----|-----|-----|------|
| Guinea | 1 | | | | Day | afte | r inj | ection | Day after injection of the toxin-antitoxin mixture. | e to | cin-a | ntito | xin | nixt | ure. | | | | | |
| pig No. | TOXIII-ARIGIOOXIII IIUXGURG IIJGGGGG SHIDGURAIGOURIY. | 17. | 18. | 19. | 20. | 21. | -25 | 23. | 24. | 25. | 56. | 27. 2 | 28. | 29. | 30. | 31. | 32. | 33. | 34. | 35. |
| 7566 | 0.142 c. c. toxine No. 5 + 415 c. c. P. D. 07635 | | B P 400 | 2 | = | 2 | 2 | ы | 2 | + | 1 | | | | | | | | | : |
| 7534 | 0.142 e. e. toxine No. 5 + ¹ / _{28σ} e. e. P. D. 08021 | : | : | B P | 2 | + | | : | | ÷ | -:- | | | | | : | : | = | - | |
| 7494 | $0.24 e. e. toxine No. 9 + _{xb\sigma} e. e. P. D. 08516 \dots$ | : | | 1 | | -:- | - | ВР | P 400 | 2 | <u></u> | 2 | <u>-</u> | 24 | <u>~</u> | 2- | 2 | + | | |
| 7481 | 0.24 c. c. toxine No. 9 + 100 c. c. S68 D | | : | | | | - | ВР | P 400 | + | : | | | - | | | : | | | |
| 7530 | 0.142 e. e. toxine No. $5 + \frac{1}{367}$ c. e. P. D. 08021 | | : | B P 500 | 2 | + | | | | | : | : | | - | -: | : | | -:- | | - : |
| 7539 | 0.142 e. e. toxine No. $5 + {}_{4}{}_{1}{}_{0}$ c. c. P. D. 08021 | • | : | B P | 2 | 2 | + | | | | | | - | | | | - : | -:- | | |
| 7577 | 0.142 c. c. toxine No. 5 + $\frac{1}{8}$ c. c. P. D. 07635. | | B P 500 | 2 | Ъ | - | 2. | à | <u>-</u> | 2 | ь | + | - | - | + | | : | | | - 11 |
| 7528 | 0.142 e. e. toxine No. $5 + \frac{1}{4} \log e$. e. P. D. 08021 | | | M P 400 | 2 | -d | + | | | - | 1 | | | \div | - | | | | | |
| 7532 | 0.142 e. e. toxine No. $5 + \frac{1}{3}$ g. e. e. P. D. 08021 | : | : | M P | 2 | + | | | | | -:- | | - | : | - | | : | | | |
| 7533 | 0.142 c. c. toxine No. 5 + 455 c. c. P. D. 08021. | | | 400 + | i | : | | | | : | | - | | : | -:- | | - | | : | |
| 7588 | 0.142 e. e. toxine No. 5 + 1355 e. e. N. Y. B. H. 305-6 | M P 400 | ã | 4 | Ъ | 4 | + | | | ÷ | : | | - : | : | -:- | i | ÷ | : | : | |
| 7519 | 0.142 e. c. toxine No. 5 + 3½ c. c. P. D. 08021. | - | | M P 500 | Ъ | 4 | + | | | ÷ | : | | | - | -:- | | : | : | | : |

| 7526 | 0.142 c. c. toxine No. 5 + 4½ c. c. P. D. 08021 | | M P 500 | - A | 2 | Д. | + | | - | <u>:</u> | - | - | - | - | | | - | - | |
|------|--|----------|----------|----------|---|----------|----|-----|----|----------|--------------|----------|-----|---------|-----|-----|------------|---|----|
| 7544 | 0.142 e. e. toxine No. 5 + 155 e. e. M. 2100 | | M P | Ч | 러 | Ъ | д | ы | Д. | <u></u> | - H | | | | | : | | | |
| 7596 | 0.142 c. c. toxine No. 5 + 4 dg c. c. A. 245 | - 500 | | Н | 4 | ы | e, | + | | | -:- | | | | | | | | |
| 7483 | 0.24 c. c. toxine No. 9 + 15c c. c. M. 2100. | | | | | | | C P | | ٠. ا | | + | - | | | + | - | | |
| 7538 | 0.142 c. c. toxine No. 5 + 4½ c. c. P. D. 08021 | | C P 400+ | | : | | | | | | - | : | - : | | - | -: | - : | | |
| 7540 | 0.142 e. e. toxine No. 5 + 4½ e. e. P. D. 08021 | | C P | Д | Ъ | Д | + | | - | - | | - : | | | | : | | | |
| 7543 | 0.142 e. e. toxine No. $5 + \frac{1}{150}$ e. c. M. 2100 | | C P | <u>-</u> | 2 | Д. | д | e, | 4 | ы | - Н - Н | <u>.</u> | | | - | - : | | | |
| 7587 | 0.142 e. c. toxine No. $5 + \frac{1}{1800}$ c. c. N. Y. B. H. 305-6 | <u>-</u> | e, | 4 | + | <u> </u> | : | | | | | | : | | - : | -:- | | | |
| 7385 | 0.142 e. e. toxine No. 5 + 1125 e. e. N. Y. B. H. 305 | | | | : | | : | | | | - 11 | | | C P 500 | | - A | - д - д | + | |
| 7518 | 0.142 c. c. toxine No. 5 + 3.5 c. c. P. D. 08021 | | C P 500 | 4 | + | | | | | -: | - | - : | - | | - | : | | | |
| 7522 | 0.142 c. c. toxine No. 5 + 345 c. c. P. D. 08021 | | C P 500 | Д. | + | | | | | - | | | - : | | - | - | | | |
| 7575 | 0.142 e. c. toxine No. $5 + \frac{1}{54}\sigma$ e. c. P. D. 07635 | C P 500 | + | | | | : | | | | | - : | - : | | | - : | | | , |
| 7583 | 0.142 c. c. toxine No. 5 + 5½ c. c. P. D. 07635 | C P 500 | - A | + | | - | : | | | - :- | - : | - : | - : | | - : | - : | | | |
| 7585 | 0.142 c. c. toxine No. 5 + z _b ₀ c. c. P. D. 07635 | C P 500 | + | : | : | i | | | | | : | : | | | | | | | |
| | | - | | | | | | | | - | - | - | - | - | 1 | - | - | - | 16 |

CAN ANTITOXIN INFLUENCE DIPHTHERITIC PARALYSIS WHEN INJECTED SHORTLY BEFORE THE PARALYSIS DEVELOPS?

The following tests show that the administration of 400 units of antitoxin, given fifteen days after the injection of a partially neutralized dose of toxine and five to six days before paralysis appeared, can not control the appearance of the symptoms or save life. Further experimental proof of this will be found in Tables Nos. 3 and 4. Guinea pigs Nos. 7930 and 7929 are illustrations of toxon poisoning with complete recovery.

TABLE NO. 2.—THE EFFECT OF ANTITOXIN WHEN INJECTED SHORTLY BEFORE SYMPTOMS APPEAR.

| Opinea pig No. | Texine-antitexin mixture. | Paralysis. | Resut. |
|-------------------|--|--|------------------|
| 7899 | 0.24 c. c. toxine No. s- sive c. c. Steams No. 1500. | Paralysis began about 20th day. Control. | Death. 24th day. |
| 7900 | 0.24 c. c. toxine No. 9+ 11/10 c. c. Stearcs No. 1500. | Paralysis began about 20th day= 900 units antitoxin 15th day. | Death, 29d day. |
| 7360 | 0.24 c. c. toxine No. 9+24co | Paralysis began about 24th day. (Control). | Recovered. |
| 7924 | 0.24 c. c. toxine No. 9+2500 c. c. N. Y. B. H. 300. | Paralysis began about 21st day = 400 units antitoxin 15th day. | Recovered. |
| 7223 | 0.24 c. c. toxine No. 9+2566 c.c. N. Y. B. H. 310 | Paralysis began about Mith day. (Control). | Death, 25th day. |
| 720 | 0.14 c. c. toxine No. 9+ 1944 c. c. N. Y. B. H. 320. | Parslysis began about 19th day = 400 units antitoxin 15th day. | Death, 15th day. |
| 7 (A) / | 0.24 c. e. toxine No. 9+ 19kg | Paralysis began a liut Nwk day. Control . | Death, 15th day, |
| - 123 | c. c. N. Y. B. H. 300 0.14 c. c. toxine No. 9+ sylop c. c. N. Y. B. H. 200 | Paralysis began about 21st day=400 units antitoxin 16th day. | Death, 25th day. |

HOW LONG AFTER THE INFECTION CAN ANTITOXIN INFLUENCE PARALYSIS?

A number of tests were undertaken to determine this important point. The first series of guinea pigs were all given an L+ dose of toxine partially neutralized so that all of them, if untreated, as shown by the controls, would have developed paralysis between the thirteenth and fourteenth days and died on the twentieth to the twenty-fifth day.

Each guinea pig in Table No. 3 received a mixture containing 0.142 c. c. of our toxine No. 9, which is the L+ dose of this poison, plus 137 c. c. of antitoxic horse serum. The minimal lethal dose of this toxine is 0.006. Each guinea pig, therefore, received 24 minimal lethal doses. The toxine and antitoxin were mixed and allowed to

stand one hour at room temperature before being injected into the guinea pig.^a Not one of the 50 guinea pigs receiving this mixture

died acutely of toxin poisoning.

All the guinea pigs receiving an injection of 250 units of the diphtheria antitoxin at intervals of 4, 8, 12, 16, and 20 hours after infection were protected against paralysis. The two guinea pigs (Nos. 8124 and 8125) receiving 250 units of antitoxin twenty-four hours after infection developed slight paralysis on the twenty-fifth day; later than the controls. They made a complete recovery.

The guinea pigs (Nos. 8138, 8139, and 8140) which received 2,000 units of antitoxin twenty-four hours after infection developed very slight paralysis, from which they recovered. It was noticeable that the guinea pigs which received 2,000 units twenty-four hours after infection developed a somewhat milder type of paralysis than those

which received 250 units.

Further, the guinea pigs (Nos. 8130 and 8131) receiving repeated injections of 250 units beginning 24 hours after infection developed but slight paralysis.

From the first series we also learn that 1 unit of antitoxin given before or at the time of infection is sufficient to prevent the development of paralysis. On the other hand, as much as 2,000 units, given 24 hours after infection, did not prevent the development of this sequel, although it modified the severity of the palsy and saved life.

^aFor details of the technic, see Hygienic Laboratory Bulletin No. 21, "The immunity unit for standardizing diphtheria antitoxin (based on Ehrlich's normal serum); official standard prepared under the act approved July 1, 1902." M. J. Rosenau.

TABLE NO. 3.—THE BFFECT OF ANTITOXIN, GIVEN AFTER INFECTION, UPON THE DEVELOPMENT OF PARALYSES.

(FIRST SERIES.)

P. Paralysis. († 250
; 1,000; 2,000=Units of antitoxin injected subentameously,
† - Death,

[Each guinea pig received a subentaneous injection = 0.142 e. c. toxiue No. 9+ v_{1g} c. c. autitoxic horse serum (S. 1110).]

| 1 | lt. | | | | | Recovered. |
|--|--|------------|------------|------------|------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|---------------|
| | Result. | + Control. | + Control. | + Control. | + Control. | No paralysis. |
| | 28, 29, 50 31, 32 33, 34, | | | : | : | : | : | : | : | : | - : | - : | : | : | : | : | : | : | : | |
| | 88 | | : | : | : | | _: | _: | .: | | _: | | _: | | | : | : | : | _: | : |
| - | 윓 | | | : | : | | - : | | | - : | : | - : | | | | : | : | | : | |
| Days after the injection of the toxine-antitoxian mixture. | <u> </u> | | | | | | | | | | | | | | -: | : | | : | | |
| iix t | 8 | | | | : | | - : | | | : | | | | | | ; | | - : | : | |
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| 0Xİ | - 8 | | | | | | : | | -: | | | | | | : | | | | | |
| 125 | 26. 27. | | | : | | | | | | | _: | | : | | | : | _: | : | | : |
| 문공 | | | | | - : | | | | | | | | - : | | | | | | | |
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| \$ | 89 | <u>-</u> + | | = | | | - : | | | | - 1 | | | | | | i | | | |
| | - 67 | <u> </u> | _ | 2 | | | | | | - : | | | _: | - : | | : | | | | |
| tio | 51 | | _ | <u></u> | - | | | | | | : | | : | | : | : | : | | | |
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| | 48 hours after. | | | | | | | - | | : | | | | : | : | - : | : | | : | |
| | 36 lionrs after. | | | : | | | : | | : | | | | | | - | : | : | | | |
| | 24 hours after. | | | | | | | | | : | : | : | - : | - : | : | : | : | : | - | |
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| g IEXO | 12 16 20 24 36 48 10 13. 14 15. 16 17, 18, 19 20, 21, 22 after, a | | | : | | | : | : | : | : | : | | | | - | | - }- | | - :- | 250 |
| f antii | 12 nours l | | | - | | | : | : | : | - | : | : | | : | : | : | : | 250 | 520 | |
| Units of antitoxin given. | 8 10mrs 1 after, 4 | | | - | | | : | | : | | | | | : | - | 250 | 250 | : | : | |
| | hours At hours hours be- once, after, after, | | | : | | | : | : | : | | - | | : | 250 | 250 | : | : | : | : | |
| | At once. | | | | : | | | : | : | _ | - | 550 | 250 | - | : | : | : | : | : | : |
| | 24 hours be- fore. | | | | | -1 | | 220 | 250 | : | : | : | : | : | : | : | : | : | : | |
| Siq | .0 V. | 8153 | 8155 | 8156 | 8157 | 8106 | 8107 | 8108 | 8100 | 8110 | 8111 | 8112 | 8113 | 8114 | 8115 | 8116 | 8117 | 8118 | 8119 | 8120 |

| No paralysis. Recovered. No paralysis. Recovered. No paralysis a Recovered. Slight paralysis of hind logs. Recovered. Slight paralysis of hind legs. Recovered. | No paralysis. Recovered. Very slight paralysis. Recovered. Covered. Covered. Covered. | No paralysis. Recovered. No paralysis. Recovered. No paralysis. Recovered. Slight paralysis. Recovered. Slight paralysis. Recovered. Slight paralysis. Recovered. |
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Each guinea pig of the second series (Table No. 4) received the same mixture of toxine and antitoxin as those of the first series.

We learn from this series that three guinea pigs (Nos. 7948, 7949, and 7950) which received 250 units of antitoxin twenty-four hours after infection had paralysis but recovered. In two of them (Nos. 7949 and 7950) the paralysis came on later than in the controls, viz, on the nineteenth day. It is therefore plain that the antitoxin given twenty-four hours after infection had the power of modifying the paralysis and saving the life of these animals.

We learn next that 250 units of antitoxin given forty-eight hours or more after infection totally failed to modify the paralysis or save

the lives of the guinea pigs (Nos. 7951, 7952, and 7953).

The animals which received 250 units of antitoxin daily beginning twenty-four hours after infection (Nos. 7978, 7979, and 7980) developed paralysis late and benign in type. The symptoms appeared on the seventeenth, twentieth, and twenty-sixth days, respectively. All three recovered. Here again the antitoxin undoubtedly modified the severity of the paralysis and saved the lives of these pigs.

The guinea pigs which received 250 units of antitoxin daily beginning forty-eight hours after infection nicely demonstrate the importance of using the serum early. Two (Nos. 7981 and 7982) developed paralysis on the seventeenth and twentieth days, respectively, and recovered. The other one (No. 7983) developed paralysis on the fifteenth day and died despite the administration of 1,250 units of antitoxin. Therefore it is plain that antitoxin given forty-eight hours after infection and in large doses may fail to influence the paralysis or save life. However, when given as late as the fourth or fifth day after infection and in repeated doses it seemed to exert some favorable action, for it will be noticed that guinea pigs Nos. 7984 to 7992 developed symptoms on the average several days later than the controls; in one case (No. 7986) death was delayed, and in another case (No. 7988) the animal recovered.

TABLE NO. 4.—THE EFFECT OF ANTITOXIN, GIVEN AFTER INFECTION, UPON THE DEVELOPMENT OF PARALYSIS.

(SECOND SERIES.)

P=Paralysis. + c=Chloroformed.

F = Faratysis. + C = Canologomea. + = Death. 250 = Subcutaneous inje

250=Subcutaneous injection of 250 units antitoxic serum.

[Each guinea pig received a subcutaneous injection: 0.142 c. c. toxine No. 5+₁₃₀-c. c. c. antitoxic horse serum (S. 1100).]

| | Result. | | + Control + Control + Control. | Recovered. | Recovered. | Recovered. | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + |
|---|---------|-------|--------------------------------------|------------|------------|------------|---------------|--------------|----------|------|--------------|------|------|----------|--------------|----------|----------|----------|------|----------|----------|
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| | 27. | | | ы | ы | Ъ. | : | + | + | + | - | + | - | : | : | - | - | - | - | : | |
| | 26. | | | 2 | <u>_</u> | | : | : | : | 1 | - | - | - | : | : | : | : | 1 | - | ÷ | |
| | 25. | | | ם | 2 | <u></u> | : | : | : | : | - | i | : | : | : | : | - | : | ÷ | ÷ | - |
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| | 21. | 2 2 2 | | ы | Ъ | Ъ | - | - | : | : | : | : | Ъ | : | - | : | : | 2 | : | : | |
| re. | 20. | 2 2 2 | 4 4 4 | <u>a</u> | Ъ | д | +c | +c | +c | - | +c · | +c | Ы | +c | +c | +6 | +c | | +c | +c | +c |
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| kin n | 18. | 226 | 7 2 2 | 굅 | 1 | - ; | <u>~</u> | ы | Ы | Ы | 7 | 7 | 7 | д | | 4 | Ь | 7 | 7 | 2 | _ _ |
| titoz | 17. | 222 | 4 A A | 2 | : | : | 4 | 러 | <u>-</u> | Ъ | Ъ. | 7 | Ъ. | 2 | 7 | <u>-</u> | ī | | ы | 4 | Ъ |
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| ng th | 10. | | | | | : | : | | | | : | | | : | | : | | | | | |
| Days following the injection of the toxine-autitoxin mixture. | 9. | | | | : | : | - | | : | : | - | - | - | : | : | : | : | : | - | - | |
| ays fo | ∞ | : : | | 1 | : | : | : | : | : | : | - | - | i | - | - | | - | - | : | i | |
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| | 6. | | | | | | : | | | : | : | : | | | : | - : | - : | : | 250 | 250 | 250 |
| | 5. | | | | | | | | | | | | | | : | 250 | 250 | 250 | - | | |
| | 4- | | | | : | - | : | - | | - | : | - | 250 | 250 | 250 | - | : | - | | | |
| | 65 | | | | : | | : | : | : | 250 | 250 | 250 | : | - : | : | | | : | | | _ |
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| | pig No. | 7943 | 7945 7946 | 7948 | 7949 | 7950 | 7951 | 7952 | 7953 | 7954 | 7955 | 7956 | 7957 | 7958 | 7959 | 7960 | 7961 | 7962 | 7963 | 7964 | 7965 |

Table No. 4.—THE EFFECT OF ANTITOXIN, GIVEN AFTER INFECTION, UPON THE DEVELOPMENT OF PARALYSIS—Con.

| Result. | + | - + | + | + | + | + | + | + | + | + | + | + | + | Recovered. | Recovered. | Recovered. | Recovered. | Recovered. | + | + | + | + | + | Recovered. | + | + | + | + |
|---|------|------|------|------|------|------|----------|------|----------|------|------|------|-----|------------|------------|------------|------------|------------|------|------|------|-----|------|------------|------|------|------|------|
| 30. | | | : | : | : | : | : | : | : | : | : | | | 24 | 4 | 2 | 2 | 2 | : | : | 1 | + | - | 2 | | | - | - : |
| 29. | | | : | | - : | : | - 1 | 1 | : | _ : | | | | 4 | 4 | 4 | Ъ | 4 | - : | : | : | Ъ | : | 7 | - | : | : | |
| 28. | | | + | - : | : | : | - | : | : | : | | | | Д | 凸 | Д | ы | 2 | - : | - | | 4 | - | д | _: | : | - : | - |
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| 26. | 1 | | ĩ | | - 1 | | - | - : | . : | | | | | д | Ь | 4 | Д. | Ъ | : | - | : | 2 | : | 2 | | | | |
| 25. | | | Â | | | - | - : | - 1 | - : | - : | - : | | | | 凸 | Ъ | Ъ | Ъ | - | | - : | 4 | : | Ь | - | | : | : |
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| 22. | | | : | - : | 24 | Ы | : | - : | 7 | | : | | | : | 4 | Ь | Ъ | Ь | T. | 2 | Ъ | Ь | Ъ | Ь | Ь | Ъ | Ы | Ы |
| 21. | | | - : | : | Ы | Ъ | : | : | Ъ | : | : | | | : | 7 | 4 | Ы | Ъ | Ъ | Ы | Ъ | Ъ | 4 | 4 | Ъ | 러 | Ь | Ч |
| nm. | + | +0 | | : | Д | 4 | | +6 | <u>-</u> | +6 | +0 | + | 1 | | Ъ | 7 | 2 | î. | Ъ | 2 | Ъ | Ы | 2 | Ъ | 凸 | Ч | Ъ | д |
| 19. | | | : | - | Ъ | 2 | - | 2, | 24 | 4 | 러 | Д | - | : | 7 | | Ъ | : | Д | Ы | Ы | : | 7 | : | : | Ь | : | : |
| oxin 18. | 1 2 | Д | : | + | 2 | - : | + | Ь | : | Ь | Ъ | 2 | 7 | : | 4 | - | Ъ | : | 2 | : | 7 | - 1 | Ъ | : | : | Ч | - : | : |
| untit 17. | 1 2 | Ъ | : | Ъ | 2 | : | Ъ | Ч | : | Ъ | Ч | Ω | - | : | 7 | : | 4 | - | 4 | - : | 4 | - | 7 | : | | Ъ | : | : |
| ine-5 | 2 | Д | : | Ъ | : | - : | 2 | 2 | : | : | | 2 | - | - | - | - | | - | Ъ | - 1 | : | : | - 1 | - : | : | - : | - : | : |
| tox 15. | 1 2 | Д | : | Ъ | - 1 | : | Д | Ы | | : | - | Д | 4 | : | : | - ; | : | : | Ъ | : | - : | - : | - | : | : | - | - | : |
| f the | 2 | Д | : | Ъ | 1 | : | <u>-</u> | Ч | : | - | - | 2 | 4 | - | - | : | 1 | : | : | : | : | : | | - | : | : | - : | : |
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| he in 11. | | | - 1 | - } | : | | | : | : | : | : | | | _: | : | : | : | : | - | : | : | : | : | | - : | - 1 | : | - ; |
| ving t | | | | | : | | : | : | | 250 | 250 | 950 | 200 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 |
| Days following the infection of the toxine-antitoxin serum 8. 9. 10. 11. 12. 13. 14. 15. 16. 17. 18. 19. 20 | - | | | | : | : | 250 | 250 | 250 | | : | | | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 |
| Days 8. | | | | 250 | 250 | 250 | : | | | | : | | | 220 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 |
| 7. | 250 | 250 | 2.50 | : | : | | : | : | | : | : | | | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 |
| 6. | | | : | : | | | | | | | | | | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 2:50 | 220 | 250 | 250 | 250 | 250 |
| 22 | | | | | : | : | | : | | | | | | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 |
| 4- | | | : | : | : | | | - | | | | | - | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | : | : | |
| ಣ | | | : | | : | | : | | : | | : | | | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | 250 | : | | : | - | | |
| ci | | | | : | | | | | | | | | | 250 | 250 | 250 | 250 | 250 | 250 | | : | : | : | : | : | : | : | |
| - | | | : | : | | - | : | : | | | : | | | 250 | 250 | 250 | : | : | - | | : | : | : | | | : | : | |
| Guinea pig No. | 9962 | 7967 | 7968 | 6962 | 7970 | 7971 | 7972 | 7973 | 7974 | 7975 | 7976 | 7077 | | 8462 | 7979 | 7980 | 7981 | 7982 | 7983 | 7984 | 7985 | 986 | 7987 | 7988 | 6862 | 0662 | 1662 | 7992 |

In Table No. 5, in which is shown the third series of guinea pigs given very large doses of antitoxin at varying times following the subcutaneous inoculation of the toxine-antitoxin mixture, we find that only the three guinea pigs (Nos. 8183, 8184, and 8185) which received the antitoxin twenty-four hours after infection recovered. The paralysis in these three pigs was so transient and slight in degree that its existence was open to some doubt in our minds, and is therefore not recorded in the table.

We confess to some disappointment that the guinea pigs receiving such massive doses of antitoxin as 4,000 units forty-eight hours after infection became paralyzed and died. (Nos. 8186, 8187, and 8188.) In Table No. 4 it appears that two of the three guinea pigs receiving repeated doses of antitoxin beginning forty-eight hours after infection recovered, and we were led to hope that one very large dose given forty-eight hours after infection would modify the paralysis more favorably than was actually the case with the guinea pigs in Table No. 5. This emphasizes more forcibly the importance of giving antitoxin early. It should be remembered that 4,000 units of antitoxin for a guinea pig weighing 250 grams is an enormous dose and would represent about 400,000 units for a 50-pound child.

TABLE NO. 5. THE EFFECT OF ANTITOXIN, CIVEN APPER INFECTION, UPON THE DEVELOPMENT OF PARALYSIS.

(THIRD SERIES.)

P=Paradyala. 250; 1,000; 4,000=Units of antitoxin injected subcutameoutly. + = Doath.

[Ench guinea pig recelved a subertantencous injection of 0.142 c. c. toxine No. 5 1 156 c. c. muthoxic horse section (8, 1100).]

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| | 48 frontes after. | 250 1,000 4,000 4,000 4,000 1,000 1,000 |
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| | Calinea plg No. | N1780 N170 N170 N181 N182 N183 N183 N183 N183 N183 N190 N190 N191 N193 N193 N193 N194 N195 N195 N195 N195 N195 N195 N195 N195 |

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| | 152 hours after. | 4,000 4,000 4,000 4,000 | |
| | 144 nours after. | | |
| 4,000 4,000 4,000 | 128 noursh ifter. | 4,000 4,000 4,000 4,000 | |
| | 120 128 144 152 168 hours hours hours hours after. after. after. | 4,000 4,000 4,000 4,000 4,000 4,000 4,000 4,000 4,000 4,000 4,000 4,000 | |
| | 104 nours lafter. | | |
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Our results upon the guinea pig correspond with the statistics of post-diphtheritic paralysis in man following treatment with antitoxic serum. We would, however, expect antitoxin when given in massive doses to have a more favorable influence upon paralysis in man than in guinea pigs, for the reason that our experimental animals received 24 minimal lethal doses of toxine but partially neutralized with antitoxic serum. This produces an early and fatal form of paralysis. In man this malignant form but rarely follows cases that have recovered from diphtheria. If, then, we are able to modify or control this sequel and save life in the guinea pig by using antitoxin forty-eight hours after infection we could expect a like beneficial result in man to follow the use of antitoxin a longer period after the onset of illness. We gave our guinea pigs the extreme charge of toxine at once; in man the toxine is doubtless elaborated and absorbed more slowly. Therefore, the following figures are significant:

Petit^a found that in 48 cases of paralysis observed at the Hôpital des Enfants malades, following diphtheria, treated with serum, the following:

Cases treated about the 2nd day, 6.25 per cent developed paralysis. Cases treated about the 3rd day, 19 per cent developed paralysis. Cases treated about the 4th day, 24.70 per cent developed paralysis. Cases treated about the 7th day, 38.70 per cent developed paralysis. Monti^b states that of the cases treated—

About the 3rd day of the disease, 8 per cent developed paralysis. About the 4th day of the disease, 12 per cent developed paralysis. About the 5th day of the disease, 33.3 per cent developed paralysis.

About the 5th day of the disease, 55.5 per cent developed paralysis.

About the 7th day of the disease, 66.2 per cent developed paralysis. Reichsfald c says that of cases treated—

About the 2nd day of the disease, 25 per cent developed paralysis. About the 3rd day of the disease, 33 per cent developed paralysis. About the 5th day of the disease, 50 per cent developed paralysis. See also Rolleston's cases, page 13.

CAN ANTITOXIN, GIVEN BEFORE INFECTION AS A PROPHYLACTIC, PREVENT POST-DIPHTHERITIC PARALYSIS?

Although we have but limited data upon this question we are enabled to answer it affirmatively. Even so small a quantity as one unit, given to a guinea pig twenty-four hours before the injection of

^a Petit, Rene: Note sur 48 cas de paralysis diphtheretique. Rev. mens. mal. enf., vol. 15, 1897, p. 76.

b Monti., Wiener med. Woch., 1895, no. 4 and 5.

c Reichsfald. Revue hebdomadaire 1895 no. 6 (text in Russian).

a toxine-antitoxin mixture containing sufficient toxon to invariably cause paralysis in control animals, is sufficient to prevent the development of paralysis and save life (see guinea pigs Nos. 8106, 8107, 8110, and 8111, Table No. 3).

SUMMARY AND CONCLUSIONS.

Post-diphtheritic paralysis in the guinea pig is an almost exact counterpart of the same sequel in man. We are therefore able to bring forward experimental evidence showing the effect of antitoxin upon post-diphtheritic paralysis.

In the guinea pig antitoxin can not influence the diphtheritic

paralysis after the paralysis has appeared.

Antitoxin has no influence in preventing post-diphtheritic paralysis if injected shortly before the paralysis develops.

Antitoxin given twenty-four hours after the infection can save the

life of the guinea pig and greatly modify the paralysis.

Antitoxin given in a single large dose forty-eight hours after the infection did not modify the paralysis or save life. Thus, in our experiments 4,000 units failed to modify the paralysis or save the life of guinea pigs weighing about half a pound. Weight for weight this corresponds to 400,000 units for a 50 pound child.

Antitoxin given in repeated injections beginning twenty-four or forty-eight hours following infection seems to have a more favorable

effect upon the subsequent paralysis than a single injection.

A very small quantity (1 unit) of antitoxin given twenty-four hours before or at the time of infection in our experiments prevented the development of paralysis.

In man we would expect more favorable results from the use of antitoxin than our work upon the guinea pig indicates, for we were dealing with an early and malignant form of experimental post-diphtheritic paralysis. This grave variety is, fortunately, rare in man. Further, we injected the entire charge of the poison directly into the tissues of the guinea pig, while in man the toxine is doubt-less elaborated more slowly. We may therefore assume that antitoxic serum, given at a somewhat later period than in our work upon guinea pigs, would exert beneficial effects.

The fact that one unit of antitoxin prevents paralysis and saves life when administered timely, whereas 4,000 units totally fails when delayed 48 hours, emphasizes the importance of using this sovereign

remedy early.



NOTICE TO LIBRARIANS AND BIBLIOGRAPHERS CONCERNING THE SERIAL PUBLICATIONS OF THIS LABORATORY.

The Hygienic Laboratory was established in New York, at the Marine Hospital on Staten Island, August, 1887. It was transferred to Washington, with quarters in the Butler Building, June 11, 1891, and a new laboratory building, located in Washington, was authorized by act of Congress, March 3, 1901.

The following bulletins [Bulls. Nos. 1–7, 1900 to 1902, Hyg. Lab., U. S. Mar.-Hosp. Serv., Wash.] have been issued.

No. 1.—Preliminary note on the viability of the Bacillus pestis. By M. J. Rosenau.

No. 2.—Formalin disinfection of baggage without apparatus. By M. J. Rosenau.

No. 3.—Sulphur dioxid as a germicidal agent. By H. D. Geddings.

No. 4.—Viability of the Bacillus pestis. By M. J. Rosenau.

No. 5.—An investigation of a pathogienic microbe (B. typhi murium Danyz) applied to the destruction of rats. By M. J. Rosenau.

No. 6.—Disinfection against mosquitoes with formaldehyd and sulphur dioxid. By M. J. Rosenau.

No. 7.—Laboratory technique: Ring test for indol, by S. B. Grubbs and Edward Francis; Collodium sacs, by S. B. Grubbs and Edward Francis; Microphotography with simple apparatus, by H. B. Parker.

By act of Congress approved July 1, 1902, the name of the "United States Marine Hospital Service" was changed to the "Public Health and Marine-Hospital Service of the United States," and three new divisions were added to the Hygienic Laboratory.

Since the change of name of the Service the bulletins of the Hygienic Laboratory have been continued in the same numerical order, as follows:

No. 8.—Laboratory course in pathology and bacteriology. By M. J. Rosenau. (Revised addition March, 1904.)

No. 9.—Presence of tetanus in commercial gelatin. By John F. Anderson.

No. 10.—Report upon the prevalence and georgaphic distribution of hookworm disease (uncinariasis or anchylostomiasis) in the United States. By Ch. Wardell Stiles.

No. 11.—An experimental investigation of Trypanosoma lewiss. By Edward Francis.

No. 12.—The bacteriological impurities of vaccine virus; an experimental study. By M. J. Rosenau.

No. 13.—A statistical study of the intestinal parasites of 500 white male patients at the United States Government Hospital for the Insane; by Philip E. Garrison, Brayton H. Ransom, and Earle C. Stevenson. A parasitic roundworm (*Agamomermis culicis* n. g., n. sp.) in American mosquitoes (*Culex sollicitans*); by Ch. Wardell Stiles. The type species of the cestode genus *Hymenolepis*; by Ch. Wardell Stiles.

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No. 21.—The immunity unit for standardizing diphtheria antitoxin (based on Ehrlich's normal serum). Official standard prepared under the act approved July 1, 1902. By M. J. Rosenau.

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No. 27.—The limitations of formaldehyde gas as a disinfectant with special reference

to car sanitation. By Thomas B. McClintic.

No. 28.—A statistical study of the prevalence of intestinal worms in man. By Ch. Wardell Stiles and Philip E. Garrison.

No. 29.—A study of the cause of sudden death following the injection of horse serum. By M. J. Rosenau and John F. Anderson.

No. 30.—I. Maternal transmission of immunity to diphtheria toxin. II. Maternal transmission of immunity to diphtheria toxin and hypersusceptibility to horse serum in the same animal. By John F. Anderson.

No. 31.—Variations in the peroxidase activity of the blood in health and disease. By Joseph H. Kastle and Harold L. Amoss.

No. 32.—A stomach lesion in guinea pigs caused by diphtheria toxine and its bearing upon experimental gastric ulcer. By M. J. Rosenau and John F. Anderson.

No. 33.—Studies in experimental alcoholism. By Reid Hunt.

No. 34.—I. Agamofilaria georgiana n. sp., an apparently new roundworm parasite from the ankle of a negress. II. The zoological characters of the roundworm genus Filaria Mueller, 1787. III. Three new American cases of infection of man with horse-hair worms (species Paragordius varius), with summary of all cases reported to date. By Ch. Wardell Stiles.

No. 35.—Report on the origin and prevalence of typohid fever in the District of Columbia. By M. J. Rosenau, L. L. Lumsden, and Joseph H. Kastle. (Including articles contributed by Ch. Wardell Stiles, Joseph Goldberger, and A. M. Stimson.)

No. 36.—Further studies upon hypersusceptibility and immunity. By M. J. Rosenau and John F. Anderson.

No. 37.—Index-catalogue of medical and veterinary zoology. Subjects: Trematoda and trematode diseases. By Ch. Wardell Stiles and Albert Hassall.

No. 38.—The influence of antitoxin upon post-diphtheritic paralysis. By M. J. Rosenau and John F. Anderson.

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Public Health and Marine-Hospital Service of the United States

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HYGIENIC LABORATORY.—BULLETIN No. 39

M. J. ROSENAU, Director

JULY, 1907

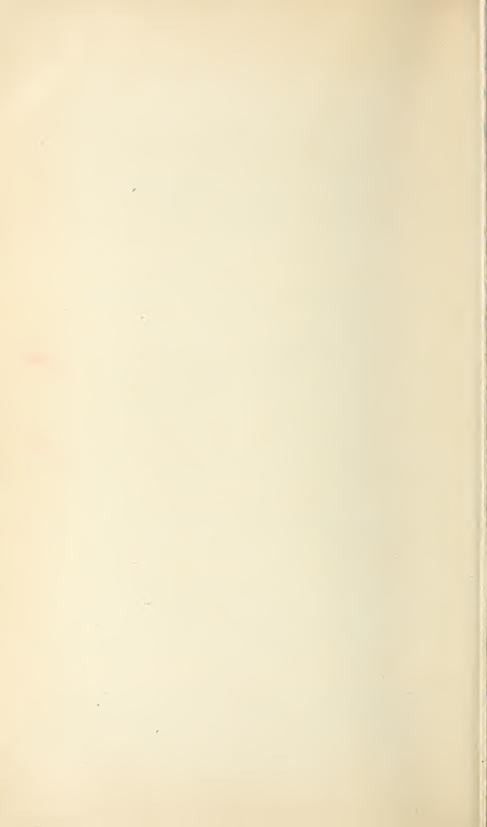
THE ANTISEPTIC AND GERMICIDAL PROPERTIES OF SOLUTIONS OF FORMALDEHYDE AND THEIR ACTION UPON TOXINES

By

JOHN F. ANDERSON



WASHINGTON
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1907



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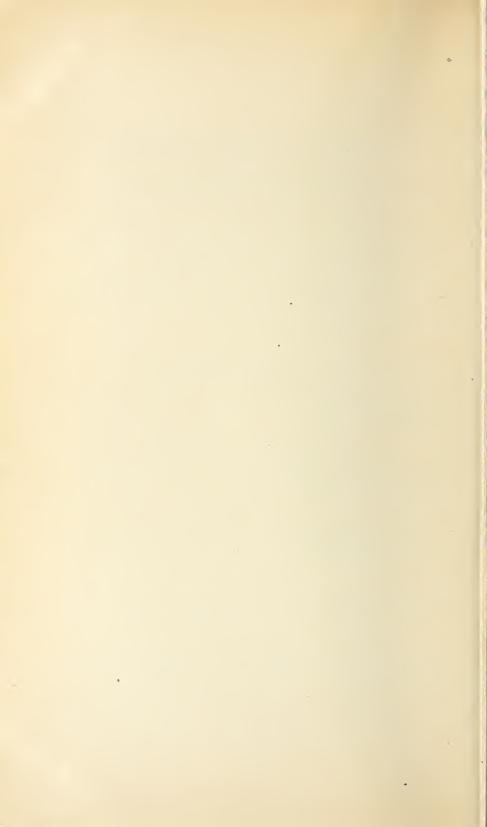
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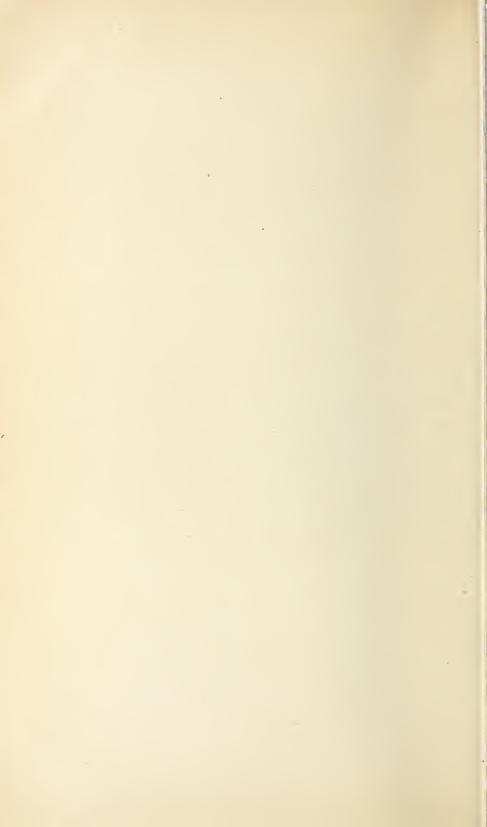
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CONTENTS.

| | Page. |
|---|-------|
| Introduction | 7 |
| Properties and uses | 8 |
| Discovery and methods of preservation | 8 |
| Tests | 10 |
| Work by previous investigators. | 12 |
| Original work | 25 |
| Antiseptic power of formalin. | 25 |
| Antiseptic power against putrefactive organisms | 25 |
| Antiseptic power against pure cultures | 26 |
| Summary of antiseptic power against pure cultures | 33 |
| Germicidal power of formalin | 35 |
| Germicidal power against pure cultures | 35 |
| Time of maximum germicidal effect | 37 |
| Action on feces. | 38 |
| Disinfection of sputum | 40 |
| Action on tetanus and diphtheria toxines | 40 |
| Summary and conclusions | 43 |
| Bibliography | 45 |



THE ANTISEPTIC AND GERMICIDAL PROPERTIES OF SOLUTIONS OF FORMALDEHYDE AND THEIR ACTION UPON TOXINES.

INTRODUCTION.

Since the discovery by Loew and Fischer, in 1886, of the germicidal action of formic aldehyde, a large amount of work has been done with the substance, both as a gas evolved by various means and in the watery solution.

The recent work by McClintic (1) seems to have established its limitations when used as a gas. The great variation in the results of various workers in regard to its action as a germicide and antiseptic in its watery solution has led me to try to determine its real value. The need of an efficient and, at the same time, a safe and comparatively cheap disinfectant is great, especially for the disinfection of the excreta of persons sick with infectious diseases. For this purpose the agent should be rapid in its action, an efficient deodorant, comparatively cheap, and safe to handle. Formic aldehyde in solution, in my opinion, comes nearer to fulfilling these conditions than any other agent we have at the present time.

In the minds of the laity, a deodorant is more or less synonymous with a germicide, but of course a deodorant may be almost valueless as a disinfectant, and, on the other hand, a disinfectant may possess but slight deodorizing properties. In the use of an agent for the disinfection of excreta, etc., it is important that the person using it understands just what is meant when told to disinfect the discharge with a 5 per cent solution. It may be understood that a 5 per cent solution of the agent is to be added to the discharge, while what is really meant is that the agent shall be in the proportion of 5 parts in 100 of the total volume under treatment. Of course it is needless to say that there must be a thorough mixture of the agent and of the material to be disinfected, and that they must remain in contact a sufficient length of time for the maximum action of the germicide to be exerted. While, for example, a 2 per cent solution might kill typhoid bacilli in thirty minutes. it might take a 1 per cent solution an hour or longer.

A number of samples of different solutions of formic aldehyde were analyzed to determine the exact percentage of the aldehyde contained

therein and quite a variation in the percentage was noted. The highest was 52 per cent, which was in a sample sent by a manufacturer, but it soon decreased to 43; the lowest was 29 per cent.

Bernard Smith (2) examined 29 samples and the lowest percentage he found was 32.45 per cent and the highest 39.11 per cent, the average of the 29 samples being approximately 37 per cent. My work was done with solutions varying from 37.2 per cent to 37.4 per cent and will be discussed in percentages of formalin, the name by which the watery solution is best known, of about 37.2 per cent formic aldehyde.

To Passed Asst. Surg. M. J. Rosenau I am much indebted for suggesting this study and for suggestions as to the work. Also, to Dr. J. H. Kastle and Asst. Surg. Norman Roberts, who very kindly collected for me most of the data on methods of preparation and tests.

PROPERTIES AND USES.

Formalin or solution of formaldehyde is an aqueous solution containing not less than 37 per cent, by weight, of absolute formaldehyde (U. S. P.). Formalin is a clear, colorless liquid with a sharp, penetrating odor, caustic taste, and is very irritating to the mucous membranes. It mixes with alcohol and water in all proportions. When subjected to low temperatures it becomes turbid, due to the formation of paraformaldehyde. It is not corrosive to the metals, except unpolished steel and iron. It combines readily with albumin, and its germicidal action is due to its combination with the cell body of the bacterium.

It is used to some extent for tanning leather. Its use as a preservative of cadavers in embalming fluids is wide. It enters into the composition of Orth's and Kaiserling's solutions, and is frequently used for fixing of specimens in microscopical work. It has been used illegally for the preservation of various food products, especially milk and cream. It is also used to some extent by farmers for the destruction of harmful fungi, such as smut on seed grain and scab on potatoes.

The gas evolved by various methods from the solution is one of our most efficient gaseous disinfectants.

According to Fischer 2 cubic centimeters of a $\frac{1}{1000}$ formalin solution per 100 grams of body weight given intraperitoneally will kill guinea pigs. In whatever way it is introduced into the body it is capable of producing lesions in the parenchymatous organs.

DISCOVERY AND METHODS OF PREPARATION.

Formic aldehyde was first obtained by Hoffman (3) in 1867. He made it by passing the vapors of methyl alcohol, laden with air, over a heated platinum spiral.

In 1875 J. Volhard (4) obtained it in the following manner: A coil of platinum wire was heated in the flame of a wood spirit lamp. The

flame was then extinguished and the slow oxidation of the alcohol was allowed to proceed. Solutions containing $\frac{1}{2}$ per cent of the aldehyde were thus prepared.

In 1881 Kabloukoff (5) prepared large amounts of the aldehyde by passing mixtures of air and methyl alcohol through a glass tube filled

with platinized asbestos and heated in a combustion furnace.

In 1884 Tollens (6) prepared the aldehyde by passing air and the vapors of methyl alcohol, heated to 55° C., over platinum foil, and concentrating the distillates by a second distillation. In this manner he obtained solutions containing 11.3 per cent of the aldehyde, and by

drying over sulphuric acid he obtained the pure compound.

O. Loew (7) in 1886 devised the following method for the preparation of the aldehyde: A current of dry air is drawn as quickly as possible through a half-liter flask half filled with methyl alcohol and then through a hard glass tube 30 cm. long, containing a cylinder of coarse copper gauze 5 cm. long, and then successively through an empty flask of 300 to 400 c. c. capacity and two flasks half filled with water. The part of the glass tube containing the copper gauze is surrounded with brass gauze and gently heated. When the alcohol vapor reaches the copper gauze the latter glows more or less according to the rate of the current of air passing over it. When the apparatus has once gotten in proper working order the process requires no further attention beyond keeping up the supply of alcohol, and the apparatus may be left running night and day. A 15 per cent to 20 per cent solution of formic aldehyde was obtained in this way. Tollens (6) in 1886 obtained still better results by passing the air through methyl alcohol that had been heated to 45°-50° C. and by substituting for platinum foil a coil of copper gauze, omitting the condenser and maintaining a constant stream of air. In this way he was able to obtain 30 per cent to 40 per cent solutions of the aldehyde with ease, and found that 30 per cent of the methyl alcohol burned was converted into the aldehyde.

In 1890 W. Eschweiler (8) employed essentially the same method as Tollens and Loew. He found that the best yield is obtained by keeping the copper gauze at such a low red heat that it only could be seen to glow in the dark. In the first receiver he obtained a 40 per cent solution of formic aldehyde, and the mixed solutions from all the receivers were found to contain from 17 per cent to 18 per cent of formic aldehyde.

In 1886 Loew and Fischer (9) discovered that formic aldehyde possesses powerful antiseptic properties. Since these earlier observations the germicidal action of formic aldehyde has been investigated by Trillat, Buchner, Aronson, Cohn, and others. As the result of these studies, formic aldehyde has come to be used on a large scale as a disinfectant and deodorizer, and the commercial methods used in the

manufacture of 40 per cent solutions of this compound are based on the methods already set forth in the above. These methods, however, are secret processes protected by chemical patents, and hence are not described in the literature.

TESTS.

The following is an outline of the most important methods now in use for the detection and determination of formaldehyde. Those most favorably mentioned in the literature are briefly described.

At the present time formaldehyde is extensively employed as an antiseptic, germicide, and food preservative. Methods therefore have been devised whereby it may be detected and determined (a) in strong solutions, such as formalin; (b) as a gas in the atmosphere of closed spaces, and (c) in admixture with various other substances, usually of an organic nature, such as milk and other foodstuffs.

The following qualitative methods are said to be the most delicate and trustworthy:

Hehner's (10) and its modifications. Bring into contact without mixing two layers, the bottom one of concentrated sulphuric acid, the top one of milk suspected of containing formaldehyde. Richmond and Boseley (11) dilute the milk with its own volume of water, which tends to prevent charring. A purple color at the zone of contact indicates formaldehyde. The casein of the milk is one of the reagents. The test can be applied to substances other than milk by the addition to the liquid to be tested of formaldehyde-free milk or certain other proteids, the best, according to Acree (12), being a globulin extracted from squash seeds. Adding to the acid a solution of ferric chloride (Lyons, 13) or solid potassium chloride (Luebert, 14) is also said to be an improvement.

Lebbin's (15). Boil a few c. c. of the liquid with 0.05 G. of resorcinol and about an equal volume of 50 per cent sodium hydroxide solution. The yellow color changes to red.

Rimini's (16). Directly applicable to foods, especially milk. To 15 c. c. of the substance are added successively 1 c. c. of dilute phenylhydrazine hydrochloride solution, a few drops of fresh sodium nitroprusside solution, and a few drops of caustic soda solution. A blue color is produced, turning finally red.

Hydrochloric acid test (51). Commercial hydrochloric acid (specific gravity 1.2) containing 2 c. c. of 10 per cent ferric chlorid per liter is used as a reagent. Add 10 c. c. of the acid reagent to an equal volume of milk in a porcelain casserole and heat slowly over the free flame nearly to boiling, holding the casserole by the handle and giving it a rotary motion while heating to break up the curd. The presence of formaldehyde is indicated by a violet coloration, varying in depth with the amount present. In the absence of formaldehyde the solution

slowly turns brown. By this test 1 part of formaldehyde in 250,000 parts of milk is readily detected before the milk sours. After souring, the limit of delicacy proves to be about 1 part in 50,000. Various aldehydes when introduced into milk give color reactions under the above treatment, but formaldehyde alone gives the violet coloration, which is perfectly distinguishable and unmistakable.

According to Leach this seems to be a most satisfactory test.

The following quantitative methods are generally considered to be the best:

Romijn's (17) cyanide method. The formaldehyde solution is added to an excess of potassium cyanide solution, and this mixture is at once poured into an excess of silver nitrate solution, which contains enough nitric acid to maintain an acid reaction throughout. The formaldehyde combines quantitatively with the cyanide to form the compound CH²O.KCN, the excess of the cyanide reacts with the silver nitrate to form silver cyanide, and the excess of the silver nitrate left in the solution is titrated with ammonium sulphocyanate.

Romijn's (18) iodometric method. To the formaldehyde solution are added 25 c. c. of decinormal iodine solution, followed by enough strong sodium hydroxide solution to make the mixture pale yellow. Allow to stand ten minutes (some authorities say longer), and then add concentrated hydrochloric acid until the reaction is slightly acid. Titrate for free iodine with sodium thiosulphate. Each two atoms of iodine consumed represents one molecule of formaldehyde.

Blank and Finkenbeiner's (19). To a carefully weighed amount of the formaldehyde solution are added normal sodium hydroxide solution, followed at once, drop by drop, by neutral hydrogen peroxide. When the reaction is completed, the excess of alkali is titrated with normal sulphuric acid. Modifications suggested by Schoorl (22).

Legler's (20). To the formaldehyde solution is added an excess of standard ammonia solution. In the gravimetric method the resulting hexamethylenetetramine is evaporated to dryness and weighed. In the volumetric the excess of ammonia is titrated with standard acid. Eschweiler (21) pointed out that when litmus or phenolphthalein is the indicator the hexamethylenetetramine itself is neutral; hence six molecules of formaldehyde need four molecules of ammonia for neutralization; but with methyl-orange, cochineal, Congo-red, or tropeolin the monoacid compound of hexamethylenetetramine is neutral; hence six molecules of formaldehyde need only three of ammonia for neutralization.

The qualitative tests of Hehner (10), Lebbin (15), and Rimini (16) are generally recommended as being about the best. It is generally best to distill the substance and to apply the tests to the distillate. It must be borne in mind that the presence of a trace of formaldehyde in food substances or elsewhere may not be the result of intentional

addition, as formaldehyde exists in the air, in smoke, and elsewhere as an incidental and often unsuspected product.

For the determination of the formaldehyde in strong solutions, the method most highly recommended is that of Blank and Finkenbeiner (19). Legler's volumetric method (20) is also in frequent use.

For the determination of the formaldehyde content of weak solu tions Romijn's (18) iodometric method is the most convenient and is trustworthy if the solution is known to be pure; other aldehydes and acetone are especially not allowable. In dilute solutions containing these or unknown substances in addition to the formaldehyde it is safer to use the cyanide method (17).

The exact determination of small amounts of formaldehyde in admixture with other organic matter is always difficult and often impossible, owing to chemical changes taking place between the organic matter and the formaldehyde or between the reagents employed for the formaldehyde and the organic matter. Colorimetric methods (23, 24, 25) furnish approximate results, and in the case of milk, at least, (26) a nearly constant fraction of the formaldehyde can be distilled over if exact methods are employed.

In quantitative determinations of formaldehyde much care is needed on account of its volatility and instability. In many of the best methods, also, unanticipated reactions are apt to vitiate the results unless directions are followed exactly; hence it is recommended by Fresenius and Gruenhut that no result should be accepted without the concordance of at least two distinct methods (27).

WORK BY PREVIOUS INVESTIGATORS.

There has been an immense amount of work done by investigators upon the antiseptic and germicidal value of formalin. The following is a brief résumé of all the principal articles.

Trillat (28) showed that the following percentages of formaldehyde are necessary to prevent putrefaction of bouillon kept at 30° C.:

1:50,000 delayed putrefaction very sensibly.

1:25,000 bouillon unaltered at the end of four days.

1:12,000 remained unaltered after several weeks.

Bouillon inoculated with bacillus anthracis was rendered sterile by 1:25,000.

The putrefaction of 10 c. c. of bouillon inoculated with 10 gtts. human caliva was sensibly delayed by 1:50,000.

1:30,000 remained unaltered at the end of fourteen days.

1:1,000 killed all the organisms in less than two hours.

Gelatine contaminated with sewage water containing 1,800,000 organisms to the c. c. showed no growth after being treated with 1:20,000.

1:1,000 killed all of the organisms after some hours exposure.

Its power of preventing fermentation in milk and wine is very marked in very small proportions, 1:4,000 in wine putting an end to fermentation.

Meat immersed and quickly withdrawn from a solution of 1:500 was preserved from putrefaction for several days.

Aronson (29) found that any proportion of formaldehyde in excess of 1:20,000 prevented the growth of B. typhosus, B. anthracis, and Staphylococcus pyogenes aureus: 1:40,000 hindered the growth, but 1:80,000 seems to have no influence.

He found that the growth of B. diphtheriæ was prevented by the application for ten seconds of a solution of formaldehyde containing 1:250; also, that twenty seconds exposure to a solution of 1:400 sterilized a culture of B. diphtheriæ.

Berlioz (30) gives the quantity of formalin to 1,000 grams required to check the growth of the organisms and the quantity of formalin to 1,000 grams which did not check their growth, as follows:

| | Checks growth. | Does not check growth. |
|---|------------------------|-------------------------------------|
| Culture from— Leucorrheal discharge. B. coli communis B. typhosus. B. anthracis | Gram. 0.03 .03 .05 .05 | Gram. 0.026 .02 .04 .04 |

Putrefaction of urine and bouillon is prevented by 0.06 gram to 1.000 grams.

Coagulation of milk is prevented by 0.2 gram to 1,000 grams.

Pieces of sterilized silk inoculated with cultures of B. anthracis and typhosus were immersed in a 1:1,000, 1:500, and 1:100 solution of formalin, then inoculated into bouillon. The bouillon became cloudy.

Blum (31) in his experiments added 10 c. c. of a bouillon culture to 90 c. c. of the formaldehyde solution, making 5 per cent of the 40 per cent solution [Formalin, A. M. S.] or 2 per cent formaldehyde.

B. chicken cholera.—Seven-day culture, 37° C., no effect after ten, seventeen, and twenty-five minutes; but no growth occurred when inoculations were made next day.

Proteus capsulatus.—Eight-day culture. After fifteen minutes the power to grow was much influenced, as a clouding of the inoculated bouillon occurred only after several days. After twenty-five minutes no living bacteria remained.

Staphylococcus pyogenes aureus.—Twenty-four hourculture. Growth much retarded by fifteen and twenty-five minutes exposure. Inoculations after thirty-five minutes remained sterile.

B. typhi abdominalis.—Not killed by exposure for five, ten, fifteen, twenty-five, and thirty-five minutes.

B. anthracis.—Bouillon culture three weeks old. No disinfection after ten, fifteen, and twenty-five minutes. Growth is, however, slower. Bouillon inoculation next day remained sterile. Twenty-five minutes exposure did not affect virulence to white mice.

Anthrax spores.—Silk threads impregnated with anthrax spores not killed by thirty-five minutes exposure. Killed by nine days.

Concludes that the disinfectant action is slow, but the antiseptic action more marked.

Carlo Ascoli (32) used slips of bibulous paper impregnated with cultures of various micro-organisms. The disinfecting agent was allowed to act for different periods, washed in sterile water, planted in bouillon, and placed in the incubator for ten days' observation. He found that 5 per cent formalin killed V. choleræ in three minutes, anthrax bacilli in fifteen minutes, anthrax spores in five hours, B. diphtheriæ in ten minutes, and Staphylococcus pyogenes aureus in thirty minutes.

Slater and Rideal (33) made experiments to determine the proportion of formaldehyde required to inhibit the growth of micro-organisms.

Formalin was added to tubes of bouillon, so that they contained formaldehyde in proportions varying from 1:1,000 to 1:20,000. These tubes were inoculated with various micro-organisms and placed in the incubator twenty-four hours. Vigorous cultures were used, either in bouillon or on agar.

| Organism. | Proportion formalde- hyde inhib- iting growth. | Proportion formalde- hyde allow- ing some growth. | Remarks. |
|--|--|---|--|
| Staphylococcus pyogenes aureus. B. typhosus. B. coli communis. B. anthracis Spirillum choleræ. B. mallei B. pyocyaneus B. prodigiosus. B. iacticus. B. butyneus Heuppe | 1: 7, 000 1: 15, 000 1: 20, 000 1: 20, 000 1: 7, 000 | 1:10,000 1:20,000 1:10,000 1:20,000 | Growth poor, 1:10,000; delayed much, 1:20,000. Very scanty. After 72 hours' incubation. Scanty growth sixth day. On third day. |

Silk threads infected by soaking in twenty-four hour cultures; strength of formaldehyde being 1 per cent.

| Organism, | Time required to kill microbes. |
|---|---------------------------------|
| Staphylococcus pyogenes aureus B. typhosus. B. coli communis B. anthracis Spirillum choleræ | Less than 15 minutes. |

Silk threads infected with twenty-four hour cultures; strength of formaldehyde 1:1,000.

| Organism. | Was not killed after— | Was killed in— |
|---|--------------------------|----------------|
| B. anthracis (no spores) Spirillum choleræ. Staphylococcus pyogenes aureus B. typhosus B. coli communis B. mallei. Putrefactive organisms | 12 hoursdododo | Do. Do. |

Experiments to determine its value as a disinfectant for soiled linen—Time of exposure to disinfectant twenty to twenty-four hours.

| Material. | 1 per cent solution. | 1:10,000 solution. |
|------------------------------|----------------------|--------------------|
| Cloths from post-mortem room | do | Sterile. |

Marcel-Arsine Mariot (34) states that proportions of formaldehyde 1:1,000 to 1:50,000 prevented putrefaction of unsterilized bouillon up to forty-eight hours, when observations ceased. The tubes were kept at 37° C.

G. Bardet (35) found that formalin in the proportion of 1:30,000 sensibly retarded the putrefaction of bouillon kept in the incubator at 30° C. and in 1:25,000 the bouillon remained unaltered at the end of four days; 1:12,000 preserved the bouillon from putrefaction for several weeks.

In sewage containing 1,800,000 organisms per c. c. all of the organisms are destroyed by formalin (proportions not stated).

Small quantities suffice to sterilize flasks containing anthrax and organisms from the saliva.

Ehrlich (36) made the following observations:

100 c. c. of milk + formaldehyde.

| | 0.02 c. c. | 0.004 e. e. | 0.002 e. c. | 0.0 с. с. |
|-------|------------|----------------------|-------------|-------------------------|
| Taste | do | Begins to coagulate. | ble. do | 0. Coagulated; sour. |

Amount of formalin in 100 &. c. of milk.

| | 0.4 c. c. | 0,2 c.c. | 0.12 e. e. | 0.08 c.c. | 0.04 e, e. | 0,0 e, e, |
|---|--------------------------------|--------------------------------|------------------------|------------------------|---------------------------|--------------|
| | 0.1 0.0. | 0.2 0.0 | | | | |
| Taste | Bad, strong itching in throat. | Bad, strong itching in throat. | Bad itching in throat. | Bad itching in throat. | Bad itching in throat. | Good. |
| Smell | | | | | Formol | 0. |
| Smell, after 24 hours. Status af- | Weak for- | mol. Weak for- mol. | mol. 0 | 0 | 0 | 0. |
| ter— | Yot coagu- | Vot coagu- | Vot coagu- | Not coagu- | Not coagu- | Regins to |
| 18 hours | lated. | lated. | lated. | lated. | lated. | coagulate. |
| | | | | | | and sour. |
| 96 hours . | do | do | do | do | do | sour. |
| | | | | | do | Do. |
| 144 hours. | do | do | do | do | Begins to co- agulate. | Do. |
| 168 hours. | do | do | do | | | Ill smelling |
| | | | | agulate. | | sour, and |
| | | | Begins to co- | | Coagulated and sour. | |

Amount of formalin in 100 c. c. of milk—Continued.

| | 0.02 e. e. | 0.004 c. c. | 0.002 c. c. | 0.0 c. c. |
|--------------------------------------|---------------------------|----------------------------------|-----------------|-----------------------------|
| | dehyde. | Less distinct for- maldehyde. | tible. | |
| Smell after24hours. Status after— | 0 | 0 | 0 | 0. |
| | | FreshBegins to coagu- | Sour and coagu- | |
| 96 hours | do | late. Coagulated and | | Do. |
| 120 hours | Begins to coagu- late. | sour. do | do | Do. |
| 144 hours | | Lumpy and sour | Lumpy and sour. | Do. |
| 168 hours | | do | do | Sour, lumpy, foul smelling. |

Sibilia (37) experimented with different percentages of formalin upon anthrax spores. His results are indicated in the table here given.

Anthrax spores in suspension used to contaminate silk threads; temperature, 37° C.

| | 1 per cent. Hours. | | 2 per cent. Hours. | | 5 per cent. Hours. | | | 10 per cent. Hours. | | | | |
|------------------------------------|-----------------------|--------|---------------------|-------|-----------------------|-----|-----|------------------------|----------|----|-------|-----|
| Minutes exposure. | | | | | | | | | | | | |
| | 3. | 5. | 20. | 3. | 5. | 20. | 3. | 5. | 20. | 3. | 5. | 20. |
| 1 2 5 5 10 20 20 30 40 50 60 60 60 | ++++++ | ++++++ | + + + | +++++ | +++++ | + | +++ | ++++ | ++++++++ | ++ | + + + | ++ |

Vanderlinden and De Buck (38) obtained the following:

Showing the results of 5 per cent formalin acting upon organisms for various periods of time.

| Organism. | Minutes action of formalin. | Result. |
|--|-----------------------------------|-------------------------------------|
| Anthrax spores B. coli communis B. diphtheriæ B. typhosus Staphylococcus pyogenes aureus Streptococcus | 15 15 30 35 | Growth. Killed. Do. Do. Do. Growth. |

Showing the results of 10 per cent formalin acting upon organisms for various periods of time.

| Organism, | Minutes action of formalin. | Result. |
|--|-----------------------------------|--------------------------------------|
| B. coli communis Typhoid Streptococcus Staphylococcus pyogenes aureus | 5 30 | Growth. Killed. Growth. Do. |

Showing the results of 10 per cent formalin acting upon pus for various periods of time, kept at room temperature.

| Pus from— | Minutes action of formalin. | Result. |
|---|-----------------------------------|------------------------------|
| Phlegmon Anthrax pustule Osteomyelitis, on sound Osteomyelitis, on sponge | 30 15 | Killed. Do. Do. Do. |

Showing results of 5 per cent formalin acting upon pus, kept at room temperature.

| Pus from— | Minutes action of formalin. | Result. |
|---|-----------------------------------|---------|
| Infected compound fracture, on sound and sponge | 30 | Growth. |

Showing results of 5 per cent formalin acting upon pus for various periods af time, kept at temperatures of 35 to 38° C.

| Pus from— | Minutes action of formalin. | Result. |
|--|-----------------------------------|----------------|
| Osteomyelitis, sound. Osteomyelitis, sponge. | 30 15 | Killed. Do. |

Showing results of 10 per cent formalin acting upon pus for various periods of time, kept at temperatures of 35 to 38° C.

| Pus from— | Minutes action of formalin. | |
|-----------------------------------|-----------------------------------|----------------|
| Phlegmon, sound. Phlegmon, sponge | 5 15 | Killed. Do. |

Rideal (39) states that 1 part of formaldehyde to 10,000 of milk the milk remained fresh without curdling for seven days.

Milk and water 10,000 parts and milk and water 100,000 parts, with 1 part of formalin, remained sweet and fresh for twelve and four days, respectively. Control soured on the third day.

One part of formaldehyde to 46.08 parts of milk, or 1 c. c. formaldehyde to 18.432 c. c. milk, keeps it fresh at least three days.

The strength of formalin used by the trade to preserve milk was found to be 1:320.

Strehl (40) used Schering's formalin. Action of spray on carpets. Various kinds of carpets treated by rubbing in virulent cultures (bouillon) of staphylococcus and anthrax, then dried and hung up. Various strengths of formalin solution then sprayed on the surface so as to moisten it evenly. After twenty-four hours pieces were placed in bouillon. Five per cent formalin did not kill staphylococci, but 10 per cent killed all bacteria.

In his original work, Walter (41) calls attention to the misunderstanding arising from the improper use of the terms formaldehyde and formalin. Aronson, Trillat, and most investigators used formaldehyde solutions; Oehmichen used formalin. The author also uses as a basis formalin (40 per cent formaldehyde solution).

Treated nutrient gelatin with different amounts of formalin, inoculated with broth cultures, made Esmarch roll tubes, and observed after a week:

| Concentration. | Anthrax spores. | Cholera. | Typhoid. | Staphylo- coccus pyogenes aureus. | Diphthe- ria. |
|--|-----------------|----------|----------|--|-------------------|
| 1:10,000 and greater. 1:20,000. 1:50,000 and 1:100,000. Control | + | ++++++ | ++++++ | - + ++ ++ | + ++ ++ |

- = no growth.

+ =small growth.

++= rich growth.

To test the bactericidal power he made up formalin solutions of various strengths in flasks, added to each agar cultures of germs, shook well, and added the mixtures to fluid gelatin from which roll tubes were made after certain exposures. He says that the antiseptic action of what formalin remained after dilution in gelatin was negligible.

SUMMARY OF RESULTS.

Staphylococcus pyogenes aureus.

- 1:2,000, growth after 2 hours' exposure, liquefaction after 13 hours to half a minute.
- 1:1,000, growth after 2 hours, liquefaction after 1 hour and less.
- 1:100, no growth after 45 minutes, growth after 30 minutes, liquefaction after 10 minutes.
 - 3:100, no growth after 2 minutes, growth after 1 minute.

Streptococcus pyogenes.

- 1:2,000, no growth after $1\frac{1}{4}$ hours, growth after 1 hour and less.
- 1:1,000, no growth after 20 minutes, growth after 15 minutes.
- 1:100, no growth after 1 minute, growth after ½ minute.

Anthrax spores (dried on silk threads).

- 1:2,000, growth after 2 hours, liquefaction after 1 hour and less.
- 1:1,000, growth after 2 hours, liquefaction after 2 minutes and less.
- 1:100, no growth after 30 minutes, growth after 20 minutes, liquefaction after 5 minutes and less.
- 3:100, no growth after 15 minutes, growth after 10 minutes, liquefaction after 5 minutes and less.

B. typhosus.

- 1:1,000, no growth after $1\frac{3}{4}$ hours, growth after $1\frac{1}{2}$ hours and less.
- 1:100, no growth after 20 minutes, growth after 15 minutes and less.
- 3:100, no growth after 1 minute, growth after ½ minute.

Cholera.

1:1,000, no growth after 1 hour, growth in 45 minutes, liquefaction in 2 minutes and less.

1:100, no growth after 5 minutes, growth after 2 minutes, liquefaction after $\frac{1}{2}$ minute.

3:100, no growth after $\frac{1}{2}$ minute.

Diphtheria.

1:1,000, no growth after 1½ hours, growth after 1 hour and less.

1:100, no growth after 15 minutes, growth after 10 minutes and less.

3:100, no growth after ½ minute.

Putrefying blood (50 c. c. + 49 c. c. aq. dist. - 1 c. c. formalin).

1:100, no growth after 1\frac{1}{4} hours, growth after 1 hour, liquefaction after 5 minutes.

Formalin did not work well in disinfecting instruments. Three per cent was efficient in disinfecting those artificially infected.

To determine the action of formalin on larger articles pieces of cloth, a uniform collar, etc., were soaked in cultures of staphylococcus pyogenes aureus, dried, and sprayed with formalin solutions of various strengths. One part was wrapped rather tightly in a towel and the other placed under an air-tight bell jar. After six hours the articles were sterile. The amounts used were 3, 5, 10, and 100 per cent, the results being the same.

In the next experiment large cloths of linen were soaked in staphylococcus bouillon cultures. One was strewn with formalin powder which contained 20 per cent formalin and tied up fairly tight in watertight tissue. After twenty-four hours some colonies grew on gelatin, but after forty-eight and seventy-two hours none grew; molds grew on all three.

A series of experiments showed that while neither formalin 3 per cent or absolute alcohol alone killed anthrax spores dried on silk threads, a mixture of alcohol and formalin 3 per cent killed them on threads moistened in sterile water after 8 minutes' exposure.

Also, as regards staphylococcus pyogenes aureus-

Absolute alcohol, growth after 3 minutes, none after 4 minutes.

50 per cent alcohol, growth after 10 minutes and less.

3 per cent formalin, no growth in 1 minute.

3 per cent formalin in absolute alcohol, no growth in 1 minute.

3 per cent formalin in 50 per cent alcohol, no growth in 1 minute.

2 per cent formalin in 50 per cent alcohol, no growth in 1 minute.

1 per cent formalin in 50 per cent alcohol, no growth in 4 minutes, growth after 3 minutes.

1 per cent formalin, growth after 10 minutes.

Walter got the following results with artificially infected stools (cholera and B: coli):

1 per cent, cholera killed in 5 minutes. B. coli lived after 60 minutes and less.

3 per cent, cholera killed. B. coli killed after 30 minutes, lived after 10 minutes.

5 per cent, cholera killed. B. coli killed after 30 minutes, lived after 10 minutes. 10 per cent, cholera killed. B. coli killed after 10 minutes, lived after 5 minutes.

His conclusions concerning liquid formalin were as follows:

Concentrations of 1:10,000 make further growth of anthrax, cholera, typhoid, staphylococcus pyogenes aureus, and diphtheria impossible.

One per cent solutions kill pathogenic germ cultures in one hour. In dilute alcoholic solutions the action is more intense.

Three per cent solutions render the hands sterile.

Articles may be disinfected by spraying with formalin solutions and inclosing air-tight.

Feces are rendered germ free in ten minutes by 10 per cent solutions. Hammer and Feitler (42) noted that formaldehyde in gas form or solution is more active against anthrax and its spores than against various other bacteria. This has given rise to the idea that its bactericidal efficiency was greater than it really was, because anthrax and its spores were assumed to be very resistant, and their destruction led to the assumption that other bacteria would be similarly influenced. Attention is not called to this in the literature. Only Gruber notes how easily anthrax is killed. He says that anthrax spores placed in cattle cars and exposed twenty-four hours to the vapor developed by spraying the inner walls with 10 to 20 per cent solution of formalin were killed.

Bouillon cultures were exposed to various strengths of solution for various times and then carried over to agar (diphtheria to blood serum) and growth observed one week.

Spore-free anthrax was always killed by 1 per cent formalin after five minutes, while cholera, diphtheria, green pus, staphylococcus, and typhoid were only killed after one hour.

Anthrax spores were killed by 1 per cent in two hours, by 0.4 per cent in one hour, and 0.5 per cent in ten minutes.

Spores of other bacteria were killed as follows:

Mycoides, 0.5 per cent, only after one hour.

Subtilis, 0.5 per cent, only after twenty-four hours.

Potato bacteria, 0.5 per cent, only after twenty-four hours.

Burring (43), to determine the germicidal properties of formaldehyde, made tests with spores of *B. anthracis* and *Staphylococcus* pyogenes aureus. Bouillon tubes with formaldehyde in the following proportions were prepared 1:3,000, 1:2,000, 1:800, 1:500, 1:300, and 1:200 and inoculated with virulent cultures of above-named organisms, but showed no growth in any case.

Thresh and Lowden (44) observed no sterilizing effect whatever with spray which did not thoroughly wet the surface with the disinfectants used.

Young, active cultures in bouillon of diphtheria, typhoid, pyocyaneus bacilli, and micrococcus prodigiosus were distributed by means of pipettes on surfaces, as wood, whitewash, wall paper; controls always

made. The surfaces of wood and whitewash were scraped deeply in making cultivations. Excluding the experiments which failed by reason of the presence of *B. subtilis*, 5 per cent solution of formalin killed all organisms on wood and wall paper, and failed only with *B. pyocyaneus* on whitewash. With 1 per cent the only failure was with *Streptococcus pyogenes albus* on whitewash; with 2 per cent every surface was absolutely sterilized.

The results of the experiments are:

1. That for spraying to be efficient every portion of the surface infected must be thoroughly moistened with the disinfectant.

2. That whitewashed surfaces require particular attention, being by far more difficult to disinfect than wood or paper. Solutions con-

taining under 2 per cent formalin are not reliable.

Kokubo (45) compared lysol or septoform and formalin soap containing 10 per cent and 25 per cent formalin with carbolic acid. He concluded that both formalin preparations were much more destructive toward anthrax than carbolic acid, for the spores lived in a 3 per cent carbolic acid solution for twenty-five days, while a 50 per cent solution of formalin soap killed in twenty-five minutes; but upon typhoid, streptococcus, and staphylococcus the carbolic acid solution was slightly more active.

Price (46) made a study of the influence of preservatives upon digestive enzymes, undertaken with the object of determining the minimum amount of formaldehyde, boric acid, borax, and salicylic acid required to preserve milk for forty-eight hours, the effect of the several preservatives upon the digestibility of the milk being subsequently determined by feeding the treated milk to calves.

A number of experiments were also carried on to determine the minimum amount of formaldehyde that could be added to milk without affecting the action of certain enzymes in vitro, and, in addition, the effect of formaldehyde upon some of the more common bacteria was studied.

His conclusions were:

- 1. Formaldehyde added to milk, 1:20,000, preserves it for forty hours.
- 2. Formaldehyde added to milk, 1:10,000, does not interfere with the digestibility when fed to calves.
- 3. Milk preserved with formaldehyde fed to calves for long periods; calves remained healthy and gained in weight.
- 4. Formaldehyde added to milk, 1:2,500 or less, has no effect on the activity of the fresh enzymes, rennet, pepsin, pancreatin, and steapsin in vitro.
- 5. Formaldehyde plus starch, 1:2,500 or less, has no effect on the conversion of starch by ptyalin and amylopsin in vitro.

6. Formaldehyde with milk, 1:20,000, has no effect on the activity of the enzyme galactose in vitro.

7. Formaldehyde and milk, 1:2,000, prevents the development of the common bacteria; 1:1,560 kills them in twenty-four hours.

Southard (47) found that 1:1,000,000 solution of formalin will prevent development of bacteria, while 1:75,000 is germicidal. A 5 per cent solution gives better results than 2.5 per cent carbolic acid or 1:500 solution of corrosive sublimate.

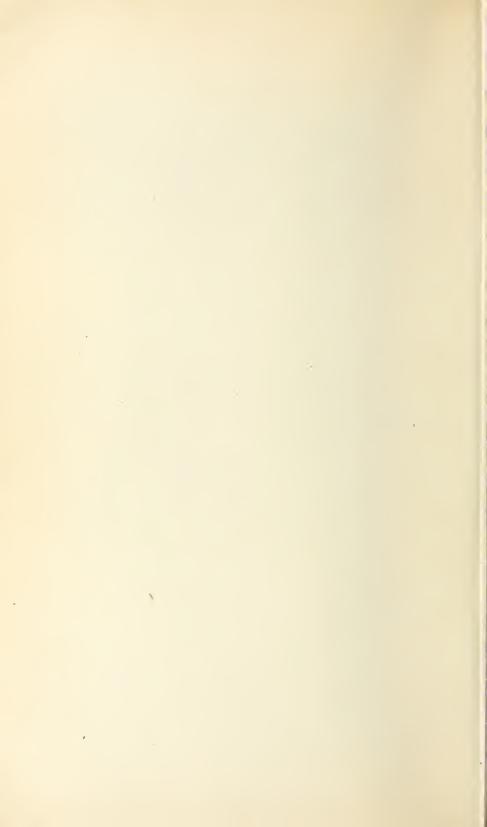
Levy (48) gives a critical review of literature and methods. He gives 269 references to literature. Following is a tabulated statement of the results of various investigators:

 $Growth\ inhibited\ in\ formal dehyde\ dilutions.$

| Author. | Staphylo- coccus pyogenes aureus. | B. typho- | B. an- thracis. | B. coli commu- nis. | B. chole- ræ. | Observations. |
|---|--|-----------|--------------------|---------------------------|------------------|---|
| Slater and Rideal. Loëw and Walter. Aronson. Trillat and Berlioz | 1:10,000 1:20,000 1:20,000 | 1:20,000 | 1:10,000 | 1:33,000 | | Weakened growth at 1:14,000. No certain action at 1:8,000. Minimal growth. |
| Abel F. Blum Pottevin. Rosenberg | | 1:7,000 | | | | 1:3,000. 1:6,250 to 1:14,300 according to number of germs. 1:10,000 "Holzin." |

The germs are killed as follows:

| Observer. | Organism. | Time. | Solution. | Result. |
|--------------------|---|-------------|---|------------------------------------|
| Slater and Rideal | Staphylococcus | 55 minutes. | | Dried on silk thread. |
| | Typhoid | 45 minutes | dehyde. | De |
| | Anthrax | | do | Do. Do. |
| | Colon | 35 minutes. | do | Do. |
| | | | do | Do. |
| | Staphylococcus | 24 hours | 1 per thousand | Do. |
| | | | do | Do. |
| | Anthrax | | do | Do. |
| | Colon | | do | Do. |
| | Cholera | | do | Do. |
| Aronson | Diphtheria | 10 seconds. | ** to -1 | Dried on wood. |
| | Typhosus | 15 minutes. | 760 | Do. |
| Œmichen | Tubercle | do | 1 per cent | |
| Wernicke | | | ‡ per cent | Do. |
| Rosenberg | Anthrax | | 10000 | Using "sterisol" lactose |
| | | | | with 3 per cent for- maldehyde. |
| Stahl | do | hour | 10000 | |
| | Staphylococcus | | 750 | |
| Berlioz | B. coli | | 0 03 | |
| Schmitt | do | | 20000 | |
| | Staphylococcus | | 20000 | |
| Ascoli | | | 5 per cent | |
| | | | do | |
| | Anthrax | 15 minutes. | do | |
| TT | Staphylococcus | | do | X |
| Kraus | - • • • • • • • • • • • • • • • • • • • | | $\frac{1}{500}$ $\frac{1}{2000}$ \cdots | Necessary proportion for |
| Van der Linden and | Diphtheria | 15 90 min | E now comt | killing. |
| Buck. | Dipitineria | utes. | 5 per cent | |
| Duck. | Typhosus | | do | |
| | | do | do | |
| | Streptococcus | 30 minutes | do | |
| | B. coli | do | do | |
| Blum | | | do | |
| | | | do | |
| | Anthrax | | | |
| Walter | Nearly all patho- | 30 minutes. | 1000 | |
| | genic organ- | | 1000 | |
| | isms. | | | |
| | Anthrax spores | | 1 per cent | |
| | do | 15 minutes. | 3 per cent | |
| | do | 2½ hours | do | |
| | do | 1 hour | 2 per cent | |
| Ascoli | do | 5 nours | 5 per cent | 14.150 |
| D. 11 | | | | |
| | do | | | |
| | dodo | 5 minutes | do | At 52°. Do. |



ORIGINAL WORK.

ANTISEPTIC POWER OF FORMALIN.

A series of experiments was made to determine the amount of formalin necessary to restrain the development of bacteria and molds in stable manure, garden earth, etc. For this purpose nutrient broth was placed in sterile flasks and formalin added in proportion of 1 to 7,000 down to 1 to 500. The broth was then inoculated with the various materials, kept at room temperature, and observed for fourteen days.

Table No. 1.—Nutrient bouillon containing various precentages of formalin contaminated by the addition of garden earth; kept at room temperature (37.2 per cent formic aldehyde).

| Dilution. | Growth appeared on— | | | | | |
|--------------------------------------|---------------------|------------|-------------|----------------|--|--|
| Dilution. | Second day. | Third day. | Fourth day. | Seventh day. | | |
| 1 to 7,000 | bottom | | | | | |
| 1 to 5,000 | | | | | | |
| 1 to 4,000 | do | Mord | | | | |
| 1 to 2,000 1 to 1,000 1 to 500 | | | | Mold on bottom | | |
| 1 00 000 | | | | | | |

Table No. 2.—Nutrient bouillon containing various percentages of formalin contaminated by the addition of stable manure; kept at room temperature (37.2 per cent formic aldehyde).

| | | | | | | | |
|--------------------------|---------------------|----------------|----------------|-----------------|--|--|--|
| Dillestion | Growth appeared on— | | | | | | |
| Dilution. | Second day. | Seventh day. | | | | | |
| 1 to 7,000 | Cloudy | Mold on bottom | | | | | |
| 1 to 6,000 1 to 5,000 | dodo | dodo | | | | | |
| 1 to 5,000 1 to 4,000 | | hottom | | | | | |
| 1 to 3,000 | | Mold on bottom | Mold on bottom | | | | |
| 1 to 1,000 | | | | Mold on bottom. | | | |
| 0 | | | | | | | |

Table No. 3.—Nutrient bouillon containing various percentages of formalin contaminated by the addition of bits of hay; kept at room temperature (37.2 per cent formic aldeyde).

| D11-11- | | Growth | appeared on— | |
|--|--------------|--------------|----------------|--------------|
| Dilution. | Second day. | Third day. | Fourth day. | Seventh day. |
| 1 to 7,000 | mold. | | | |
| 1 to 5,000 1 to 4,000 1 to 3,000 | Surface mold | Cloudy | | |
| 1 to 2,000 1 to 1,000 | | Surface mold | Mold on bottom | |
| 1 to 500 | | | | |

It will be seen from Table No. 1 that in the flask to which garden earth was added bacterial growth ceased in the dilution of 1 to 5,000 and molds developed on the seventh day in the dilution of 1 to 1,000.

In the flask to which stable manure was added bacterial growth occurred in the dilution of 1 to 4,000 and molds on the seventh day in 1 to 1,000.

In the flask to which were added bits of hay bacterial growths occurred in a dilution of 1 to 4,000 and molds on the fourth day in a dilution of 1 to 1,000.

It will be seen that the lowest dilution in which bacterial growth occurred was 1 to 4,000 and molds in 1 to 1,000.

ANTISEPTIC POWER AGAINST PURE CULTURES.

The above experiment having been made to determine its restraining influence for putrefactive changes, another series was made to determine its antiseptic action against pure cultures of bacteria.

For this purpose bouillon containing various proportions of formalin was distributed in sterile test tubes, and each tube was then inoculated with a small loopful of a forty-eight-hour agar culture of the particular organism, placed in the incubator at 37° C., and observed daily for fifteen days. The formalin used contained 37.2 per cent formic aldehyde.

Table No. 4.—Bouillon containing various percentages of formalin was distributed in test tubes, and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of Bacillus typhosus, kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth: - no growth.]

| | | | | | [, 2 | | | | . 8.0 | | | | | | |
|------------|------------|-------------|------------|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
| 1 to 7,000 | _ | - | - | { + scant. | }+ | + | + | + | + | + | + | + | + | + | + |
| 1 to 6,000 | _ | - | - | · – | - | | { + scant. | } + | + | + | + | + | + | + | + |
| 1 to 5,000 | _ | - | - | - | - | - | scant. | } + | + | + | + | + | + | + | + |
| 1 to 4,000 | - | - | - | | - | | { + seant. | } + | + | + | + | + | + | + | + |
| 1 to 3,000 | - | - | _ | _ | _ | | | _ | - | _ | | _ | - | - | |
| 1 to 2,000 | - | - | - | - | - | _ | - | _ | _ | - 1 | | - | - | _ | - |
| 1 to 1,000 | _ | _ | - | | - | - | - | _ | _ | _ | - | _ | - | | - |
| 1 to 500 | _ | | _ | _ | _ | _ | - | _ | | - | _ | | - | _ | _ |

Table No. 5—Bouillon containing various percentages of formalin was distributed in test tubes, and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of Staphylococcus pyogenes citreus, kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth duy. | Ninth glay. | Tenth day. | Bleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|------------|------------|-------------|------------|-------------|------------|------------|--------------|-------------|-------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 | _ | - | - | + | + | + | + | + | + | + | + | + | + | + | + |
| 1 to 6,000 | - | - | - | - | - | { scant. | } + | 1 + | + | + | + | + | + | + | + |
| 1 to 5,000 | - | | - | - | - | {scant. | } + | + | + | + | + | + | + | + | + |
| 1 to 4,000 | - | - | - | - | - | _ | - | {scant. | } + | + | + | + | + | + | + |
| 1 to 3,000 | - | - | _ | _ | - | - | _ | - | , – | - | | _ | | _ | - |
| 1 to 2,000 | - | _ | - | - | - | - | - | _ | - | | | - | - | - | _ |
| 1 to 1,000 | - | - | - | -) | - | - | - | - | - | - | - | - | - | _ | - |
| 1 to 500 | _ | <u> </u> | _ | | _ | _ | _ | | _ | | | _ | _ | | _ |

Table No. 6.—Bouillon containing various percentages of formalin was distributed in test tubes, and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of Staphylococcus pyogenes albus, kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|--|------------|-------------|------------|----------------------------|---------------|------------|--------------|-------------|------------|------------|---|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 1 to 6,000 1 to 5,000 1 to 4,000 1 to 3,000 1 to 2,000 1 to 1,000 | + | ++ | +++ | + + + + scant. | + + + + + + + | + + + + + | + + + + | + + + + + | ++++ | ++++ | +++++++++++++++++++++++++++++++++++++++ | + + + + | +++ | ++++ | +++++ |

Table No. 7.—Bouillon containing various percentages of formalin was distributed in test tubes, and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of Staphylococcus epidermidis albus, kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth: - no growth.]

Fourteenth day Thirteenth duy, Fifteenth day. Eleventh day Seventh day Twelfth day Second day, Fourth duy Eighth day Tenth day Third day Ninth day Sixth day. First day. Fifth day. Dilution. 1 to 7,000... + 1 to 6,000... + scant. 1 to 5,000... 1 to 4.000... 1 to 3,000... 1 to 2,000... 1 to 1,000... _ 1 to 500

Table No. 8.—Bouillon containing various percentages of formalin was distributed in test tubes, and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of Staphylococcus pyogenes aureus, kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|--|------------|-------------|------------|-------------|------------|------------|--------------|---|--------------------------------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 1 to 6,000 1 to 5,000 1 to 4,000 1 to 2,000 1 to 1,000 1 to 500 | | ++ | ++ | + + + | +++ | +++ | +++ | + + + + seant. - - - | + + + + - - - - | ++++ | + + + | + + + + | ++++ | + + + | + + + + |

Table No. 9.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. prodigiosis kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|------------|------------|-------------|------------|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 | - | + | + | + | + | + | + | + | + | + | + | + | + | + | + |
| 1 to 6,000 | - | - | - | _ | {scant. | }+ | + | . + | + | + | + | + | + | + | + |
| 1 to 5,000 | - | - | - | - | {scant. | }+ | + | + | + | + | + | + | + | + | + |
| 1 to 4,000 | _ | - | _ | - | - | - | - , | - | {scant. | } + | + | + | + | + | + |
| 1 to 3,000 | _ | - | _ | _ | _ | - | - | _ | _ | _ 3 | - | _ | _ | _ | _ |
| 1 to 2,000 | _ | | - | _ | - | - | _ | - | | - | _ | - | - | - | _ |
| 1 to 1,000 | - | - | - | - | | - | _ | - | _ | - | _ | - | - | - | - |
| 1 to 500 | | - | - | _ | - | _ | _ | _ | _ | _ | _ | _ | | _ | _ |

Table No. 10.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. acidi lactici kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|--|------------|-------------|------------|-------------|------------|-----------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 1 to 6,000 1 to 5,000 1 to 4,000 1 to 2,000 1 to 1,000 1 to 500 | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + |

Table No. 11.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. subtilis kept under observation in the incubator at 37° C. for fifteen days.

| [+ means growth; - no gro |
|---------------------------|
|---------------------------|

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|--------------------------|------------|-------------|------------|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 1 to 6,000 | _ | | ++++ | + | ++ | ++ | + + | ++ | ++ | ++ | + + | ++ | ++ | ++ | ++ |
| 1 to 5,000 | - | - | { scant. | }+ | + | + | + | + | + | + | + | + | + | + | + |
| 1 to 4,000 | - | - | _ | _ | - | _ | scant. | } + | + | + | + | + | + | + | + |
| 1 to 3,000 | _ | _ | _ | | _ | | ` - | _ | - | _ | _ | - | - | _ | - |
| 1 to 2,000 | | | _ | - | _ | _ | - | - 1 | _ | - | - | _ | - | _ | _ |
| 1 to 1,000 | - | - | _ | - | | _ | - | - | _ | - | - | _ | _ | - | - |
| 1 to 500 | - | - | - | - | - | - | _ | - | - | - | - | | - | | Trans. |

Table No. 12.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. megaterium kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|---|------------|-------------|------------|----------------------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 1 to 6,000 1 to 5,000 1 to 4,000 1 to 3,000 1 to 2,000 1 to 1,000 1 to 500 | | | | - - - - - - | {scant. | } + | + | + | + | + | + | + | + | + | + |

Table No. 13.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. pestis kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|------------|------------|-------------|------------|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 | _ | - 1 | + | + | + | + | + | + | + | + | + | + | + | + | + |
| 1 to 6,000 | - | - | - | - | - | | {scant. | + | + | + | | + | + | + | + |
| 1 to 5,000 | _ | | - 1 | | - | | | | | | - | _ | | - | |
| 1 to 4,000 | _ | - | | | | _ | | | | | | | | - | |
| 1 to 3,000 | - | - | | ~~ | | | | | | | | | | | |
| 1 to 2,000 | - | - | - | - | | | | | | | | | | | |
| 1 to 1,000 | - | - | - | _ | | | | | | - | | | | | |
| 1 to 500 | - | - | 20.00 | - | - | _ | - | - | | | _ | | - 1 | | _ |

Table No. 14.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. mallei kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|------------|------------|-------------|------------|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 | _ | _ | _ | - | _ | _ | {scant. | } + | + | + | + | + | + | + | + |
| 1 to 6,000 | - | _ | | _ | _ | _ | _ | | - | - | - | _ | | | _ |
| 1 to 5,000 | _ | _ | - | - | _ | - | _ | _ | _ | _ | - | _ | _ | _ | - |
| 1 to 4,000 | _ | - | - | _ | _ | - | _ | - | - | _ | _ | - | - | | _ |
| t to 3,000 | | - | - 1 | - | - | - | - | _ | - | _ | _ | _ | | - | |
| 1 to 2,000 | - | - | - 1 | _ | - | _ | i. — | _ | _ | _ | _ | - | | - | - |
| 1 to 1,000 | _ | - | - | _ | | _ | - | _ | - | - | _ | - | _ | - | - ` |
| 1 to 500 | _ | - | - | | _ | _ | - | - | - | | - | | _ | _ | - |

Table No. 15.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. anthracis kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth: - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|------------|------------|-------------|------------|-------------|--|------------|--------------|--|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 | - | _ | - | - | $\begin{cases} + \\ \text{scant.} \end{cases}$ | } + | + | + | + | + | + | + | + | + | + |
| 1 to 6,000 | - | _ | - | - | _ | {scant. | }+ | + | + | + | + | + | + , | + | + |
| 1 to 5,000 | - | - | _ | - | _ | _ | _ | $\begin{cases} + \\ \text{scant.} \end{cases}$ | } + | + | + | + | + | + | + |
| 1 to 4,000 | _ | _ | _ | _ | _ | - | _ | _ | _ | _ | - | - | - 1 | _ | _ |
| 1 to 3,000 | _ | _ | _ | - | _ | - | - | - | - | - | - 1 | - | - 1 | _ | - |
| 1 to 2,000 | | - | - | - | - | - | - | - | _ | _ | - | - | - | _ | _ |
| 1 to 1,000 | Parent | - | - | - | _ | - | - | - | _ | _ | _ | ~ | - 1 | _ | - |
| 1 to 500 | - | _ | _ | | · - | _ | - | _ | - | _ | _ | _ | _ | _ | _ |

Table No. 16.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. diphtheriæ kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|--|------------|-------------|------------|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 1 to 6,000 1 to 5,000 1 to 4,000 1 to 3,000 1 to 2,000 1 to 1,000 | | | | + + | ++ | + + | + + | ++ | + + | + + | + + | + + | + + | + + | ++ |

Table No. 17.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of Cholera kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth: - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|------------|------------|-------------|------------|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 | _ | _ | _ | _ | _ } | _ | { + | } + | + | + | + | + | + | + | + |
| 1 to 6,000 | | | | | | | \scant. |) | | _ | | | | | |
| 1 to 5,000 | | | | | | _ | | _ | | | | | | _ | |
| 1 to 4,000 | _ | | _ | _ | _ | _ | _ | - | _ : | - | _ | _ | _ | _ | - |
| 1 to 3,000 | _ |) — I | _ | - 1 | _ | _ | _ | - | - 1 | - | - | _ | - | _ | - |
| 1 to 2,000 | _ | - | - | | - | _ | - | _ | - | _ | - | - | _ | - | - |
| 1 to 1.000 | - | - | _ | - | - | - 1 | _ | - | _ | - | - | - | - | - | - |
| 1 to 500 | - | - | - | - | _ | - | _ | - | _ | - 1 | - | - | - | - | - |

Table No. 18.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. pyocyaneus kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.]

| First day | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day, | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|--|-----------------------|--|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7,000 — 1 to 6,000 — 1 to 5,000 — 1 to 4,300 — 1 to 2,000 — 1 to 1,000 — 1 to 500 — | + + + scant. | \\ + + \-\ \\ \-\ \-\ \-\ \-\ \-\ \-\ \- | + + + + | + + + + | + + + + | + + + + + | + + + + | + + + + + | + + + + + | +++++ | + + + + + | + + + + | + + + + | + + + + |

Table No. 19.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. dysenterix Shiga kept under observation in the incubator at 37° C. for fifteen days.

[+ means growth; - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|------------|------------|-------------|------------|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| | | | | | , | | | | | | | | | | |
| 1 to 7,000 | _ | - | - | { + scant. | + | + | + | + | + | + | + | + | + | + | + |
| 1 to 6,000 | - | _ | _ | - | { + scant. | }+ | + | + | + | + | + | + | + | + | + |
| 1 to 5,000 | - | - | _ | - | _ | | - | - | { + scant. | } + | + | + | + | + | + |
| 1 to 4,000 | - | _ | | | | - | _ | _ | - | _ | _ | _ | - 1 | - | _ |
| 1 to 3,000 | - | - | -040 | - 3 | - | - | - | - | - | - | - | _ | _ | _ | - |
| 1 to 2,000 | - | - | - | - | - | - | - | | - | _ | - | - | _ | _ | - |
| 1 to 1,000 | - | - | _ | - | _ | - | - | - | - | _ | _ | _ | _ | _ | - |
| 1 to 500 | _ | - | - | _ | _ | _ | _ | | ~ | _ | _ | - | _ | _ | _ |

Table No. 20.—Bouillon containing various percentages of formalin was distributed in test tubes, and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. coli communis kept under observation in the incubator at 37° C. for rifteen days.

| FI moone | orrowth. n | a armayersh T |
|-----------|-------------|---------------|
| 1 THEATIS | growth: - n | TO RIOW LITT |

| Dilution. | Plrst day. | Second day. | Thurd day. | Fourth day. | Fifth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fiftgenth day. |
|------------|------------|-------------|------------|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7.000 | - | - | 1 | + | + | - | + | + | + | + | + | + | + | - | ÷ |
| 1 to 6,000 | - | - | scant. | }+ | + | _ | 1 | ÷ | + | <i>†</i> | + | - | + | + | + |
| 1 to 5.000 | | | scant. | }- | - | + | - r | + | + | - | + | ÷ | + | _ | + |
| 1 to 4.000 | _ | - | - ' | _ | -, | - | | | - | { scant. | } + | + | - | + | + |
| 1 to 3.000 | | - | _ | _ | | _ | - | - | - | _ | _ \ | - | - | - | |
| 1 to 2.000 | - | _ | - | | - | - 1 | - | | - | - | - | | - | _ | - |
| 1 to 1,000 | - | - | - | - | _ | _ | | | | _ | - | | _ | - | - |
| 1 to 500 | - | - | _ | - | - | - | _ | _ | _ | _ | - | - | - | - | - |

Table No. 21.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. proteus kept under observation in the incubator at 37° °C, for fifteen days.

[+ means growth: - no growth.]

| Dilution. | First day. | Second day. | Third day. | Fourth day. | Piffth day. | Slxth day. | Seventh day. | Elghth day. | Ninth day. | Tenth day. | Bleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|------------|------------|-------------|------------|-------------|-------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7.000 | | | - | | seant. | + | + | + | 4 | 1 | _ | + | + | + | + |
| 1 to 6,000 | | _ | _ | | _ | _ | | | - > | | | | _ | - | |
| 1 to 5.000 | - | | | - | | - | - | | _ | | | - | | _ | |
| 1 to 4.000 | - | | | - | - | | - | | | - | - | - | _ | _ | - |
| 1 to 3,000 | - | - | - | | | | - | _ | | _ | _ | | - 1 | - | _ |
| 1 to 2.000 | | - | _ | | _ | - | _ | | - | - | | | - | - | - |
| 1 to 1.000 | | - | | | - | _ | _ | - | _ | - | - | | - | - | _ |
| 1 to 500 | | | | - | - | - | - | - | - | | - | - | - | - | - |
| | | | | | | | | | | | | | | | |

Table No. 22.—Bouillon containing various percentages of formalin was distributed in test tubes and each tube was inoculated with a small loopful of a forty-eight-hour agar culture of B. enteritidis kept under observation in the incubator at 37° C. for fifteen days.

[- means growth: - no growth.]

| Dilution. | Pirst day. | Second day. | Third day. | Fourth day. | FIFth day. | Sixth day. | Seventh day. | Eighth day. | Ninth day. | Tenth day. | Eleventh day. | Twelfth day. | Thirteenth day. | Fourteenth day. | Fifteenth day. |
|--------------------------|------------|-------------|--------------|-------------|------------|------------|--------------|-------------|------------|------------|---------------|--------------|-----------------|-----------------|----------------|
| 1 to 7.000 1 to 6.000 | | | + | - | + | + | + | + | + | ++ | + | ++ | ++ | ++ | + + |
| 1 to 5,000 | | - | + seant. | }+ | - | - | + | + | + | | + | - <u>†</u> | + | + | <u>-</u> |
| 1 to 4,000 | - | | _ | - | | | fscant. | } + | ŧ | + | + | + | - | + | + |
| 1 to 3.000 | | _ | | | _ | _ | _ | | _ | | _ | _ | - | - | _ |
| 1 to 2,000 | _ | | _ | - | - | | _ | - | | _ | _ | - 1 | - | _ | |
| 1 to 1,000 | | | - | - | - | _ | _ | - | - | _ | _ | _ | | | |
| 1 to 500 | | | - | - | - | - | - | - | - | - | - | - | - | - | _ |

Table No. 23.—Summary showing lowest dilution in which all growth was restrained.

| Organism. | Dilution in which all growth was re- strained. |
|--|--|
| Staphylococcus epidermidis albus Staphylococcus pyogenes albus Staphylococcus pyogenes aureus Staphylococcus pyogenes citreus Bacillus typhosus Bacillus coli communis Bacillus dysenteriæ (Shiga) Bacillus pyocyaneus Bacillus pyocyaneus Bacillus diphtheriæ Bacillus diphtheriæ Bacillus anthracis Bacillus matlei Bacillus megaterium Bacillus uspodigiosis Bacillus prodigiosis Bacillus prodigiosis Bacillus prodigiosis Bacillus prodigiosis Bacillus proteus Bacillus proteus Bacillus proteus | 1 to 3,000 1 to 3,000 1 to 3,000 1 to 3,000 1 to 4,000 1 to 6,000 1 to 6,000 1 to 6,000 1 to 6,000 1 to 6,000 1 to 5,000 1 to 5,000 1 to 5,000 1 to 3,000 |
| | |

From the above it will be seen that many of the organisms used grew in dilutions as low as 1 to 4,000; a few were restrained in a dilution of 1 to 6,000; none grew in a stronger solution than 1 to 3,000.



GERMICIDAL POWER OF FORMALIN.

The germicidal power of formalin in various strengths was studied on pure cultures, human feces, and sputum. The growth in broth tubes was principally used. Agar plates were also made at corresponding intervals in order to determine when the maximum effect of formalin is exerted.

GERMICIDAL POWER AGAINST PURE CULTURE.

The germicidal power of formalin was determined in 1, 2, 3, 5, and 10 per cent dilutions against the principal pathogenic organisms. The following method was observed in each case: For a 5 per cent strength 94 c. c. of sterile water was placed in a sterile flask and 1 c. c. of a heavy emulsion, freed from clumps, of a forty-eight-hour slant agar culture of the organism being studied was added; then 5 c. c. of formalin was added to the entire contents of the flask and well shaken. One loopful at ten-minute intervals up to one hour was planted in a full tube of nutrient bouillon and the growth noted from day to day.

The experiments were repeated in each dilution so as to check any irregularities that might occur.

Table No. 24.—One per cent formalin (37.2 per cent aldehyde).

| Organism | | Ž | linute | s. | | | Hours | | Control grew in— |
|--|----------|---|--------|---|-------------|------|-------|----|--|
| | 10. | 20. | 30. | 40. | 50. | 1. | 11. | 2. | |
| Vibrio cholera B. coli communis B. diphtheriæ B. mallei B. pyocyaneus B. dysenteriæ (Shiga) Staphylococcus pyogenes albus Staphylococcus pyogenes aureus B. typhosus | ++++++++ | +++++++++++++++++++++++++++++++++++++++ | ++++++ | + | + + + + + + | ++++ | + + + | | 24 hours. Do. Do. Do. Do. Do. Do. Do. Do. Do. Do |

From the above table it will be seen that the vibrio of cholera failed to grow after ten minutes' exposure; B. coli grew up to ninety minutes; B. diphtheriæ was killed in twenty minutes; B. mallei was killed in ten minutes; B. pyocyaneus grew up to forty minutes; B. dysenteriæ (Shiga) and Staphylococcus albus grew up to two hours; Staphylococcus pyogenes aureus and B. typhosus grew up to sixty minutes.

(35)

Table No. 25.—Two per cent formalin (37.2 per cent aldehyde).

| 0. | 20. | linute: | s. | | | Hours. | | Control |
|-----|-----|-------------------|-----|-----|-----|----------|----|-----------|
| 0. | 20 | | | | | grew in- | | |
| | | 30. | 40. | 50. | 1. | 11. | 2. | |
| - 0 | _ | _ | | | _ | | _ | 24 hours. |
| - | + | + | - | + | +s. | - | _ | Do. |
| 1 1 | - | _ | - | - | - | - | - | Do. |
| - | _ | _ | - | - | - | _ | _ | Do. |
| + | - | _ | _ | - | _ | _ | _ | Do. |
| - | -5- | + | 1 | - | +s. | - Names | _ | Do. |
| + | + | + | - | 1. | + | | _ | Do. |
| - | _ | - | - | | _ | | | Do. |
| 4 | - | - | + | - | | | _ | Do. |
| | + | + - + + + + | - | - | | | | |

This table shows that none of the bacteria tested resisted an exposure of longer than sixty minutes to a 2 per cent solution of formalin. The fact that the colon, dysentery, and typhoid bacillus and the cocci were more resistant is shown in this table.

Table No. 26.—Three per cent formalin 37.2 per cent aldehyde).

| Organism. | | 7 | linute | s. | | | | Control grew in- | |
|-------------------------------|-----|-----|--------|-----|-----|----|-----|---------------------|-----------|
| | 10. | 20. | 30. | 40. | 50. | 1. | 11. | 2. | |
| Vibrio cholera. | | | | | _ | | | _ | 24 hours. |
| B. coli communis. | _ | + | | - | | - | | _ | Do. |
| B. diphtheriæ | 4 | - | - | | | _ | - 1 | _ | Do. |
| B. mallei | - | - | | | | | | _ | Do. |
| B. pyocyaneus | | | | | | | _ | | Do. |
| B. dysenteriæ (Shiga) | _ | | + | | - | _ | | - | Do. |
| staphylococcus pyogenes albus | + | - | + | | | | | | Do. |
| taphylococcus pyogenes aureus | + | 1 | _ | | | | | - | Do. |
| B. typhosus | - | - | - | | | | | _ | Do. |

Only one of the organisms, B. dysenteriæ, tested resisted an exposure to 3 per cent formalin forty minutes; this organism, B. coli, and Staphylococcus albus again show to be more resistant than the others tested.

Table No. 27.—Fire per cent formalin (37.2 per cent aldehyde).

| | | Tir | ne of e | exposu | re to | forma | lin. | | |
|--------------------------------|-----|-----|---------|---------------------|-------|-------|------|----|-----------|
| Organism. | | 21 | | Control grew in— | | | | | |
| | 10. | 20. | 30. | 40. | 50. | 1. | 11. | 1) | |
| Vibrio cholera | _ | | | | | | | | 24 hours. |
| B. coli communis | - | | +d | _ | - | _ | _ | _ | Do. |
| B. diphtheiræ | _ | _ | | - | - | _ | | _ | Do. |
| B. mallei | - | _ | - 1 | _ | | | - | - | Do. |
| B. pyocyaneus | _ | - | - 1 | - | | _ | _ | _ | Do. |
| B. dysenteriæ Shiga | + | | + | -d | - | - | - | - | Do. |
| Staphylococcus pyogenes albus | + | - | -d | | - | _ | _ | | Do. |
| Staphylococcus pyogenes aureus | + | -d | - 1 | _ | _ | _ | _ | _ | Do. |
| B.typhosus | + | -d | | - 1 | _ | _ | _ | _ | Do. |

This table shows practically the same results as with 3 per cent formalin. It would seem that there are a few of the organisms that

are more resistant than the great mass of them which are killed off by the 3 per cent solution, but these finally succumb to a 5 per cent solution.

Table No. 28.—Ten per cent formalin (37.2 per cent aldehyde).

| | | Ti | me of | expost | ire to f | ormal | in. | | |
|---|-----------------|-------------|---------------|--------|----------|-------|------------------|------------------|---------------------|
| Organism. | Minutes. Hours. | | | | | | | | Control grew in— |
| | 10. | 20. | 30. | 40. | 50. | 1. | $1\frac{1}{2}$. | 2. | |
| Vibrio cholera B. coli communis B. diphtheriæ. B. mallei | _ | · - - | · · · · · · · | - - | | | | | 24 hours. |
| B. pyocyaneus B. dysenteriæ (Shiga) Staphylococcus pyogenes albus Staphylococcus pyogenes aureus B. typhosus. | ++ | | | | | | | - - - - | Do. Do. Do. |

Only those organisms which resisted the 5 per cent solution for ten minutes were used. Of the nonspore-bearing organisms only B. dysenteriæ and Staphylococcus albus survived an exposure of ten minutes to a 10 per cent solution of formalin.

TIME OF MAXIMUM GERMICIDAL EFFECT.

In order to study when the greatest germicidal action is exerted, pure cultures of the five most resistant organisms were exposed to a 2 per cent and a 3 per cent solution of formalin. The dilutions were made just as in the preceding experiments, but instead of making plants in broth a loopful of the exposed culture was put into a full tube of melted agar, and then poured into a petri dish which, as soon as the agar had hardened, was placed in the incubator.

The colonies were counted on the fourth day. Plants were made at five-minute intervals, except in the case of *B. pyocyaneus*, when exposed to 3 per cent formalin, and then plants were made at two-minute intervals.

Table No. 29.—Two per cent formalin (37 per cent aldehyde, 1.8 acidity).

| | Numbe | r of colonies | grown | at differe | ent inter | vals of | expos | sure. | |
|--|----------|----------------------------------|------------------|------------------|------------------|---------------------|------------------|---------------------|---------------------|
| Organism. | At once. | 5 minutes. | 10 min- utes. | 15 min- utes. | 20 min- utes. | 30 min- utes. | 40 min- utes. | 50 min- utes. | 60 min- utes. |
| B. dysenteriæ B, pyocyaneus | do | 410 290 | 22 | 19 0 | 5 0 | 3 0 | 1 | 1 | 1 |
| Staphylococcus pyo- genes albus. Staphylococcus pyo- | dodododo | 6,839 Not countable. 1,080 | 7, 200 410 | 4,800 | 2,600 | 800 | 480 2 | 0 63 0 | 0 2 |
| genes aureus. | | 1,000 | 110 | 01 | 02 | | - | | |

The above results show very plainly that in a 2 per cent solution the maximum effect is executed in the first five minutes, B. dysenteriæ and the cocci showing the greatest resistance.

Table No. 30.—Three per cent formalin (37 per cent aldehyde, 1.8 acidity).

| | Numbe | er of colonies | grown a | t differe | nt interv | als of | exposi | ure. | |
|--|----------|---|--------------------------|-------------------------|-----------------------|---------------------|----------------------|---------------------|---------------------|
| Organism. | At once. | 5 minutes. | 10 min- utes. | 15 min- utes. | 20 min- utes. | 30 min- utes. | 40 min- utes. | 50 min- utes. | 60 min- utes. |
| B. dysenteriæ. B. typhosus. Staphylococcus pyogenes albus. Staphylococcus pyogenes aureus. | do | 360 2,580 Almost in- numerable. 2,160 | 37 8 4, 320 310 | 16 3 2, 820 83 | 6 1 1,800 21 | 2 0 93 6 | 2 0 0 | 0 | |
| | | 2 minutes. | 4 min- utes. | 6 min- utes. | 8 min- utes. | 10 min- utes. | 15 min- tites. | | |
| B. pyocyaneus | do | 300 | 29 | 7 | 0 | 0 | 0 | | |

This table is especially interesting as showing that in its effect in a 3 per cent solution upon B. pyocyaneus the great mass of organisms are killed in the first two minutes.

Both tables show that the larger number of organisms are killed in the first five minutes, the more resistant ones gradually being killed off by a longer exposure.

Compare these results with Tables 25 and 26, page 36.

ACTION ON FECES.

Its germicidal action having been determined against pure cultures, experiments were done to determine its value as a disinfectant of human excreta.

Human feces with urine added was thoroughly mixed with sufficient water so that there should remain no large clumps. A watery suspension from a number of forty-eight-hour agar slants of *B. pyocyaneus* was added to the mixture and well mixed in, this being done on account of the green color it shows in culture, which was used as an index of the destruction of the nonspore-bearing organisms. To an accurately measured quantity of this mixture sufficient formalin was added so as to give a 3 per cent, 5 per cent, or 10 per cent dilution by volume of formalin. Plants were made into broth tubes and agar, which latter were at once poured into petri dishes. The growth in the broth tubes was observed for ten days; the colonies on the plates were counted on the fourth day.

Where the letter "P" appears pyocyaneus, as shown by the characteristic color, was noted.

Table No. 31.—Feces and B. pyocyaneus, 3 per cent formalin, 37.2 per cent aldehyde.

[*An anærobic spore-bearing rod.]

| Time. | Number of colonies | Days' growth in broth tubes. | | | | | | | | | | | |
|--|--------------------|------------------------------|----|----|---------|----------------|----|----|----|----|-----|--|--|
| Time. | grown. | 1. | 2. | 3. | 4. | 5. | 6. | 7. | 8. | 9. | 10. | | |
| Raw. At once. 10 minutes. 20 minutes. 40 minutes. 50 minutes. 60 minutes. 120 minutes. | 64 | +P | | + | + + + + | +* +* +* | | | | | | | |
| 120 minutes | ••••• | ••••• | | | | +* | | | | | | | |

Table No. 32.—Feces and B. pyocyaneus, 5 per cent formalin, 37.2 per cent aldehyde.

[* An anærobic spore-bearing rod.]

| Time. | Number of colonies grown. | Days' growth in broth tubes. | | | | | | | | | | | |
|--|--|------------------------------|----|----|-----|----|----|----|----|----|---------|--|--|
| Time. Number of colonies grown. | | 1. | 2. | 3. | 4. | 5. | 6. | 7. | 8. | 9. | 10. | | |
| Raw | Innumerable, PAlmost innumerable; not counted 73 | +P | | + | | | | | | | | | |
| 30 minutes 40 minutes 50 minutes | 6. 3. | | | | +++ | +* | | | | | | | |
| 60 minutes | | | | | | +* | | | | | • • • • | | |

Table No. 33.—Feces and B. pyocyaneus, 10 per cent formalin, 37.2 per cent aldehyde.

[* An anærobic spore-bearing rod.]

| * Time. Number of colonies grown. | | Days' growth in broth tubes. | | | | | | | | | |
|---|---------------------------|------------------------------|----|----|----|----|----|----|----|----|-----|
| | Number of colonies grown, | 1. | 2. | 3. | 4. | 5. | 6. | 7. | 8. | 9. | 10. |
| Raw. Innumerable, P At once Very few, "spreaders" 10 minutes 6. | | +P + | | | | | | | | | |
| 20 minutes | 3 1 | | | ++ | +* | | | | | | |

A look at the above tables shows that in all three dilutions the maximum effect is exerted within the first ten minutes. In the 10 per cent dilution the great majority of the bacteria are killed almost immediately; *B. pyocyaneus* was killed in the 3 and 5 per cent dilutions in less than ten minutes and at once in the 10 per cent.

The deodorant action of formalin is an important consideration when used for the disinfection of feces; they are almost completely deodorized at once by a 10 per cent solution; a 5 per cent solution does not completely deodorize, but renders them almost inoffensive in a few minutes. There seems to be but little difference in the deodorant action of a 3 and a 5 per cent solution.

On account of its germicidal efficiency and deodorant action formalin would seem to be one of the most useful agents for the disinfection of infected human discharges when used in a 10 per cent dilution, allowing an exposure of one hour after thorough mixing.

DISINFECTION OF SPUTUM.

In order to determine the efficiency of formalin for the destruction of tubercle bacilli in sputum the following experiment was done:

Ten cubic centimeters of sputa from an advanced case of pulmonary tuberculosis was added to 180 cubic centimeters of sterile water, and the mass of sputum broken up as much as possible. An amount of the mixture of sputum and water equal to that used to inoculate the guinea pigs after exposure to formalin was given to a guinea pig as a control. Sufficient formalin was added to the sputum to make a 5 per cent solution by volume. At intervals of fifteen minutes up to one hour a small amount was given to a guinea pig intraperitoneally, with the following result:

TABLE No. 34.

| Time. | Death in— | Post-mortem findings. |
|------------|-----------|-----------------------|
| 30 minutes | 31 days | |

The control pig died in thirty-five days with lesion of generalized tuberculosis.

The experiment was repeated with another specimen of sputum with practically the same result.

It would seem from the above that 5 per cent formalin in one hour was sufficient to render tuberculous sputum harmless to a guinea pig.

ACTION OF FORMALIN ON TETANUS AND DIPHTHERIA TOXINES.

Aronson (49) called attention to the fact that formalin, in the proportion of 1 to 500, added to diphtheria toxine, was sufficient to protect a guinea pig against 100 fatal doses of the toxine.

Burckhard (50) found that formalin, when added to tetanus toxine in the proportion of 1 to 250, was sufficient to protect a mouse against a dose of tetanus toxine that caused the death of the control in twenty-four hours. Burckhard does not seem to have determined the least fatal dose of his tetanus toxine.

Neither of them states how long the formalin was left in contact with the toxine before it was injected into the animal. My work on the action of formalin was all tested on guinea pigs, and the MLD of toxines accurately determined.

Table No. 35.—Showing the effect of different percentages of formalin on tetanus toxine, six hours' exposure.

| | Weight | Dose in MLDs. | Time exposed, in hours. | Forma- lin. | Result. | | | | |
|---------------|----------------------------|---------------------------------|-------------------------|----------------------|------------------|------------------|-----------|--|--|
| Date. | guinea pig. | | | | Death. | | Recovery. | | |
| | Pis. | | 111 110 0101 | | Days. | Hours. | | | |
| March 1, 1907 | Grams. 520 375 400 390 400 | 100 100 100 100 100 | 6 6 6 6 | Per cent. 1 2 3 4 5 | 2 3 4 7 | 1 8 4 6 | + | | |

Table No. 36.—Showing the effect of different percentages of formalin on tetanus toxine, twenty-four hours' exposure.

| | Wainba | Dose in MLDs. | Time exposed, in hours. | Forma- | Result. | | | |
|---------------|----------------------------|---------------------------------|----------------------------------|----------------------|---------|--------|-----------------|--|
| Date. | Weight guinea pig. | | | | Death. | | Recovery. | |
| | pig. | | | | Days. | Hours. | | |
| March 1, 1907 | Grams, 520 500 510 490 530 | 100 100 100 100 100 | 24 24 24 24 24 24 | Per cent. 1 2 3 4 5 | 4 14 | 21 0 | + . + . + | |

Table No. 37.—Showing the effect of 3 per cent formalin on tetanus toxine, after exposure of from one to six hours.

| | *** | Dose in MLDs. | Time exposed, in hours. | | Result. | | | | |
|---------------|--------------------------------|--|-------------------------------|---------------------------|----------------------------|------------------------------|-----------|--|--|
| Date. | Weight guinea pig. | | | Forma- lin. | Death. | | D | | |
| • | Pre. | | III HOUIS. | | Days. | Hours. | Recovery. | | |
| March 6, 1907 | Grams. 325 320 335 315 360 385 | 100 100 100 100 100 100 | 1 2 3 4 5 6 | Per cent. 3 3 3 3 3 3 3 3 | 2 4 4 5 5 6 | 11 4 16 8 0 4 | | | |

Two control guinea pigs for the above three series died in twenty-seven and twenty-eight hours, respectively.

Table No. 38.—Showing the effect of different percentages of formalin on diphtheria toxine after six hours' exposure.

| | Weight guinea pig. Dose in MLDs. | MIDa | Time exposed, in hours. | | Result. | | | | |
|---------------|-----------------------------------|--|-------------------------------|---------------------|-------------------------|------------------------|-------------------|--|--|
| Date. | | | | Forma- lin. | Death. | | Recovery. | | |
| | | III HOUIS. | | Days. | Hours. | | | | |
| March 1, 1907 | Grams. 400 390 375 400 425 | 200 200 200 200 200 200 | 6 6 6 6 6 | Per cent. 1 2 3 4 5 | 1 3 7 20 20 | 6 13 9 0 0 | Paralysis, Do. | | |

Table No. 39.—Showing the effect of different percentages of formalin on dipatheria toxine after twenty-jour hours' exposure.

| | Weight burnea pig. | | Time exposed. in hours. | Forma- lin. | Result. | | | |
|---------------|--|---------------------------------|-------------------------------|----------------|----------|--------|---------------|--|
| Date. | | Dose in MLDs. | | | Death. | | D | |
| | | | | | Days. | Hours. | Recovery. | |
| March 1, 1307 | Grants. 430 420 370 390 370 | 200 200 200 200 200 | 54 54 54 54 54 | Per cent. | 28 25 | 0 | Paralysis. Do | |

Table No. 40.—Showing the effect of 3 per cent formalin on diphtheria toxine, after exposure for various lengths of time.

| | Weight guinea pig | Dose in MLDs. | Time exposed, in hours. | | Result. | | | | |
|---------------|--|---|-------------------------------|----------------|------------------|---------------------|-----------|--|--|
| Date. | | | | Forma- lin. | Death. | | Dallamann | | |
| | | | | | Days. | Hours. | Recovery. | | |
| March 18, 195 | 420 420 420 430 435 440 | 200 200 200 200 200 200 200 | 10 00 dt 10 00 | Per cent. | He He Me Me Line | 13 15 4 17 | | | |

Two control pigs with the toxine alone died in seventeen and nineteen hours, respectively.

A study of the above tables shows that when tetanus toxine is exposed to 5 per cent formalin for six hours a guinea pig is able to withstand 100 MLD of this formalinized toxine: that 3 per cent formalin after twenty-four hours exposure protects against the toxine: that 3 per cent formalin after one hour destroys a part of the toxine, and that its action increases with the length of exposure.

Diphtheria toxine is more susceptible than tetanus toxine, as 4 per cent formalin after six hours' exposure protects a guinea pig against acute death, paralysis, however, not being prevented. One per cent, after twenty-four hours' exposure, has the same effect. Its action against the toxine increases with the length of exposure.

This action of formalin on toxine is an important property in its use as a disinfectant, for not only are the bacteria themselves destroyed, but their soluble products as well.

Attempts have been made to treat certain septicemic conditions by means of the intravenous injection of weak formalin solutions, but without much success. Perhaps more success would attend its use in localized infections where there is a nest of germs elaborating a toxine which is absorbed and distributed throughout the system. This phase of the subject, it is hoped, will be treated in a later paper.

SUMMARY AND CONCLUSIONS.

Formalin in the proportion of 1 to 5,000 was sufficient to restrain all bacterial growth in bouillon contaminated with garden earth. Molds developed in a dilution of 1 to 1,000.

In a dilution of 1 to 4,000 formalin restrains bacterial growth in bouillon contaminated with stable manure.

A dilution of 1 to 4,000 restrains bacterial growth in bouillon contaminated with wisps of hay. Molds developed in a dilution of 1 to 1,000.

Summary showing lowest dilution in which all visible growth of pure cultures was restrained.

| Organism. | Dilution in which growth was restrained. |
|---|--|
| Staphylococcus epidermidis albus Staphylococcus pyogenes albus Staphylococcus pyogenes aureus Staphylococcus pyogenes citreus Bacillus typhosus Bacillus typhosus Bacillus dysenteriæ (Shiga) Bacillus dysenteriæ (Shiga) Bacillus pyocyaneus Bacillus mallei Bacillus mallei Bacillus negaterium Bacillus megaterium Bacillus modicis Bacillus prodigiosus Bacillus prodigiosus Bacillus eriteitälis Bacillus diphtheriæ | 1 to 3,000 1 to 3,000 1 to 3,000 1 to 3,000 1 to 3,000 1 to 4,000 1 to 4,000 1 to 6,000 1 to 5,000 1 to 6,000 1 to 6,000 |

One per cent formalin (containing 37.2 per cent aldehyde) killed the vibrio of cholera in ten minutes, B. coli in ninety minutes, B. diphtheriæ in twenty minutes, B. mallei in ten minutes, B. pyocyaneus in forty minutes, B. dysenteriæ and Staphylococcus albus grew up to two hours, and Staphylococcus pyogenes aureus and B. typhosus were killed in ninety minutes.

Two per cent formalin (containing 37.2 per cent aldehyde) killed B. coli after sixty minutes' exposure, B. diphtheriæ after ten minutes, B. pyocyaneus ten minutes, B. dysenteriæ sixty minutes. Staphylococcus pyogenes albus sixty minutes, and Staphylococcus pyogenes aureus and B. typhosus after forty minutes.

Three per cent formalin (containing 37.2 per cent aldehyde) killed B. coli after thirty minutes' exposure, B. dysenteriæ forty minutes, Staphylococcus pyrogenes albus thirty minutes, Staphylococcus pyogenes aureus twenty minutes, and B. typhosus after twenty minutes.

Five per cent formalin (containing 37.2 per cent aldehyde) killed B. coli and Staphylococcus pyogenes albus after thirty minutes' exposure, B. dysenteriæ after forty minutes. Staphylococcus pyogenes aureus and B. typhosus after twenty minutes.

Ten per cent formalin (containing 37.2 per cent aldehyde) killed all the nonspore-bearing organisms tested in less than ten minutes, except B. dysenteriæ and Staphylococcus pyogenes aureus, which were killed after ten minutes.

With a 2 per cent solution of formaldehyde the maximum germicidal effect was exerted in the first five minutes.

With a 3 per cent solution this immediate effect is more apparent, and in the case of B. pyocyaneus the great mass of organisms is killed in the first two minutes.

A 3 per cent solution destroys a great majority of the nonsporebearing organisms in feces in the first ten minutes; a few, however, resisting a somewhat longer exposure; the spore-bearing organisms grew up to one hundred and twenty minutes.

Five per cent formalin acting upon feces destroyed most of the nonspore-bearing organisms within ten minutes; a few resisted forty minutes. The spore-bearing organisms resisted as long as sixty minutes.

Feces exposed to 10 per cent formalin solution were rendered practically sterile immediately; a few colonies developed in ten minutes. The spore-bearing organisms were destroyed after forty minutes.

A 10 per cent solution of formalin completely deodorized feces at once. A 3 and 5 per cent solution renders them almost odorless after a very few minutes' exposure.

On account of its germicidal efficiency and deodorant action formalin would seem to be one of the most useful agents for the disinfection of infected human discharges when used in a 10 per cent dilution, allowing an exposure of one hour after thorough mixing.

Tubercle bacilli in tuberculous sputum were killed after an exposure to 5 per cent formalin for sixty minutes. Sputum exposed for fifteen, thirty, and forty-five minutes, respectively, still contained live virulent tubercle bacilli.

When tetanus toxine is exposed to 5 per cent formalin for six hours a guinea pig is able to withstand 100 MLD of this formalinized toxine. Three per cent formalin after twenty-four hours exposure protects against the toxine; it destroys a part of the toxine in one hour, its action increasing with the length of exposure.

Diphtheria toxine is more susceptible than tetanus toxine, as 4 per cent formalin, after six hours exposure, protects a guinea pig against acute death; paralysis, however, is not prevented. One per cent, after twenty-four hours exposure, has the same effect. Its action against the toxine increases with the length of exposure.

This action of formalin on toxine is an important property in its use as a disinfectant, for not only are the bacteria themselves destroyed, but their soluble products as well.

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The Hygienic Laboratory was established in New York, at the Marine Hospital on Staten Island, August, 1887. It was transferred to Washington, with quarters in the Butler Building, June 11, 1891, and a new laboratory building, located in Washington, was authorized by act of Congress March 3, 1901.

The following bulletins [Bulls. Nos. 1-7, 1900 to 1902, Hyg. Lab., U. S. Mar.-Hosp.

Serv., Wash.] have been issued:

No. 1.—Preliminary note on the viability of the Bacillus pestis. By M. J. Rosenau.

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No. 6.—Disinfection against mosquitoes with formaldehyde and sulphur dioxid.

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No. 7.—Laboratory technique: Ring test for indol, by S. B. Grubbs and Edward Francis; Collodium sacs, by S. B. Grubbs and Edward Francis; Microphotography with simple apparatus, by H. B. Parker.

By act of Congress approved July 1, 1902, the name of the "United States Marine-Hospital Service" was changed to the "Public Health and Marine-Hospital Service of the United States," and three new divisions were added to the Hygienic Laboratory.

Since the change of name of the Service the bulletins of the Hygienic Laboratory

have been continued in the same numerical order, as follows:

No. 8.—Laboratory course in pathology and bacteriology. By M. J. Rosenau. (Revised edition March, 1904.)

No. 9.—Presence of tetanus in commercial gelatin. By John F. Anderson.

No. 10.—Report upon the prevalence and geographic distribution of hookworm disease (uncinariasis or anchylostomiasis) in the United States. By Ch. Wardell Stiles.

No. 11.—An experimental investigation of Trypanosoma lewisi. By Edward Francis.

No. 12.—The bacteriological impurities of vaccine virus; an experimental study. By M. J. Rosenau.

No. 13.—A statistical study of the intestinal parasites of 500 white male patients at the United States Government Hospital for the Insane; by Philip E. Garrison, Brayton H. Ransom, and Earle C. Stevenson. A parasitic roundworm (*Agamomermis culicis* n. g., n. sp.) in American mosquitoes (*Culex sollicitans*); by Ch. Wardell Stiles. The type species of the cestode genus *Hymenolepis*; by Ch. Wardell Stiles.

No. 14.—Spotted fever (tick fever) of the Rocky Mountains; a new disease. By

John F. Anderson.

No. 15.—Inefficiency of ferrous sulphate as an antiseptic and germicide. By Allan J. McLaughlin.

No. 16.—The antiseptic and germicidal properties of glycerin. By M. J. Rosenau.

No. 17.—Illustrated key to the trematode parasites of man. By Ch. Wardell Stiles.

No. 18.—An account of the tapeworms of the genus Hymenolepis parasitic in man, including reports of several new cases of the dwarf tapeworm $(H.\ nana)$ in the United States. By Brayton H. Ransom.

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(47)

No. 22.—Chloride of zinc as a deodorant, antiseptic, and germicide. By T. B. McClintic.

No. 23.—Changes in the Pharmacopæia of the United States of America. Eighth Decennial Revision. By Reid Hunt and Murray Galt Motter.

No. 24.—The International Code of Zoological Nomenclature as applied to medicine. By Ch. Wardell Stiles.

No. 25.—Illustrated key to the cestode parasites of man. By Ch. Wardell Stiles.

No. 26.—On the stability of the oxidases and their conduct toward various reagents. The conduct of phenolphthalein in the animal organism. A test for saccharin, and a simple method of distinguishing between cumarin and vanillin. The toxicity of ozone and other oxidizing agents to lipase. The influence of chemical constitution on the lipolytic hydrolysis of etheral salts. By J. H. Kastle.

No. 27.—The limitations of formaldehyde gas as a disinfectant with special refer-

ence to car sanitation. By Thomas B. McClintic.

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No. 31.—Variations in the peroxidase activity of the blood in health and disease. By Joseph H. Kastle and Harold L. Amoss.

No. 32.—A stomach lesion in guinea pigs caused by diphtheria toxin and its bearing upon experimental gastric ulcer. By M. J. Rosenau and John F. Anderson.

No. 33.—Studies in experimental alcoholism. By Reid Hunt.

No. 34.—I. Agamofilaria georgiana n. sp., an apparently new roundworm parasite from the ankle of a negress. II. The zoological characters of the roundworm genus Filaria Mueller, 1787. III. Three new American cases of infection of man with horse-hair worms (species Paragordius varius), with summary of all cases reported to date. By Ch. Wardell Stiles.

No. 35.—Report on the origin and prevalence of typhoid fever in the District of Columbia. By M. J. Rosenau, L. L. Lumsden, and Joseph H. Kastle (including articles contributed by Ch. Wardell Stiles, Joseph Goldberger, and A. M. Stimson).

No. 36.—Further studies upon hypersusceptibility and immunity. By M. J. Rosenau and John F. Anderson.

No. 37.—Index-catalogue of medical and veterinary zoology. Subjects: Trematoda and trematode diseases. By Ch. Wardell Stiles and Albert Hassall.

No. 38.—The influence of antitoxin upon post-diphtheritic paralysis. By M. J. Rosenau and John F. Anderson.

No. 39.—The antiseptic and germicidal properties of solutions of formaldehyde and their action upon toxines. By John F. Anderson,

In citing these bulletins, beginning with No. 8, bibliographers and authors are requested to adopt the following abbreviations: Bull. No. ——, Hyg. Lab., U. S. Pub. Health & Mar.-Hosp. Serv., Wash., pp. ——.

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TREASURY DEPARTMENT

Public Health and Marine-Hospital Service of the United States

WALTER WYMAN, Surgeon-General

HYGIENIC LABORATORY.—BULLETIN No. 40

M. J. ROSENAU, Director

May 25, 1908

- 1.—The Occurrence of a Proliferating Cestode Larva (Sparganum proliferum) in Man in Florida. By Ch. Wardell Stiles.
- 2.—A Reexamination of the Type Specimen of Filaria restiformis Leidy, 1880=Agamomermis restiformis. By Ch. Wardell Stiles.
- 3.—Observations on Two New Parasitic Trematode Worms:

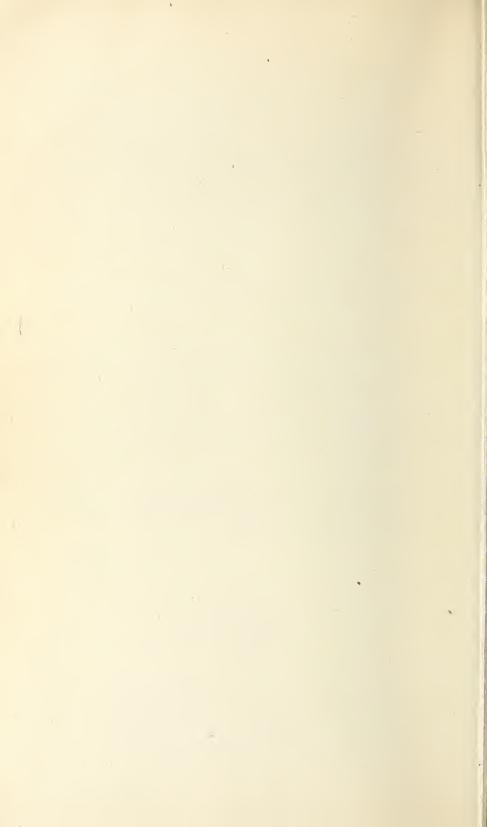
 Homalogaster philippinensis n. sp., Agamodistomum
 nanus n. sp. By Ch. Wardell Stiles and Joseph
 Goldberger.
- 4.—A Reexamination of the Original Specimen of *Tænia* saginata abietina (Weinland, 1858). By Ch. Wardell Stiles and Joseph Goldberger.



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1908



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United States Public Health and Marine-Hospital Service.

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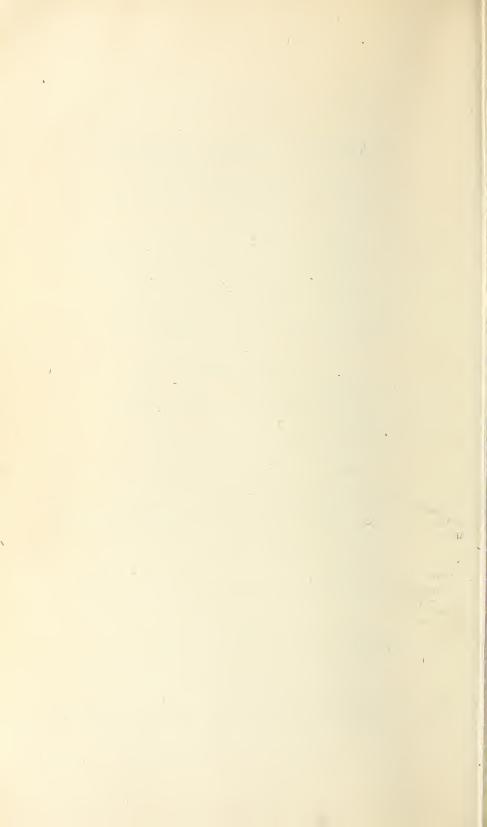
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CONTENTS.

| | Page. |
|--|-------|
| Summary | 6 |
| The occurrence of a proliferating cestode larva (Sparganum proliferum) | |
| in man in Florida | 7 |
| A reexamination of the type specimen of Filaria restiformis Leidy, 1880= | |
| Agamomermis restiformis | 19 |
| Observations on two new parasitic trematede worms: Homalogaster phil- | |
| ippinensis, Agamodistomum nanus | 23 |
| A reexamination of the original specimens of Tania saginata abietina | |
| Weinland | 35 |
| Index to zoological names | 39 |

SUMMARY.

This bulletin contains four articles on parasitic worms.

I. A very peculiar case of parasitism in man in Florida has been found by Doctor Gates, of Manatee. The worms, which live in the subcutaneous tissue and cause an acnelike swelling, have been determined as very closely related to, perhaps identical with, *Sparganum proliferum*; this parasite has been reported on only one former occasion (1905), when it was found in Japan; its life history, source of infection, prevention, and treatment are still unknown; its chief peculiarity is a reproduction in its larval stage by means of forming supernumerary heads, which may become independent and wander through the tissue.

II. Filaria restiformis is a worm described as a parasite from man. The original specimen has been found and restudied. It proves to be an immature horse hair worm and it is therefore exceedingly doubtful whether it was a parasite of man.

III. Some parasites referred to us by an official in Manila and Phrapatoom, collected from the cecum of cattle (Bos ——) prove to be a new species of trematode (Homalogaster philippinensis). Some parasites collected from the partridge (Francolinus subtorquatus) in West Africa by Dr. F. C. Wellman, and referred to us by the American Society of Tropical Medicine, prove to be an agamic encysted new species (Agamodistomum nanus) of trematode, rather closely related to the genus Clinostomum. It is impossible to state at present whether or not it is transmissible to man, but it does not agree with any form at present known for man; it is not known at present for any game or other birds in the United States.

IV. A few segments of the original specimen of *Tania abietina* have recently been found. They are in very poor condition, and this precludes any final judgment as to their exact systematic position. They appear as a dwarfed specimen of *T. saginata*, yet there are certain indications which point to a difference from that species. For the present the form may perhaps best be viewed as a possible subspecies of *T. saginata*.

All bibliographic references are taken from Stiles and Hassall, Index-Catalogue of Medical and Veterinary Zoology, Authors, Bull. 39, U. S. Bureau of Animal Industry.

All drawings were prepared by Leonard H. Wilder, artist of this laboratory.

THE OCCURRENCE OF A PROLIFERATING CESTODE LARVA (SPARGANUM PROLIFERUM) IN MAN IN FLORIDA."

By CH. WARDELL STILES, Ph. D.,

Chief of Division of Zoology, Hygienic Laboratory, U. S. Public Health and Marine-Hospital Service.

(Figs. 1 to 18.)

Medical History of the case.—In June of this year (1907), I received from Dr. H. Gates, of Manatee, Fla., specimens of worms, for identification, which he had taken from the skin of a man. The following extracts from Doctor Gates's letters give all that is known to me regarding the medical history of the case. In connection with this history, it will be well to abstract in considerable detail an account of a very similar case recently reported by Ijima (1905) in Tokyo, Japan.

Gates's case.—Inclosed find specimens of worms from human flesh. They are inclosed in sacks deep under the skin in the connective tissue. My patient has thousands of them all over the trunk; they can be seen and felt as nodules; deeper in the connective tissue in the left groin and left breast there seem to be large masses of them. I send some I obtained by cutting open the nodules and squeezing them out, and others still in the cyst as I found them.—From letter dated June 17, 1907.

In reply to your request for history, etc., of patient (J. W. M.) infected with larval tapeworm, I would state that I first discovered the condition in April of this year. I was called in to treat him for dysentery. While examining over abdomen I found a great many nodules in the skin and also in the fascia between the skin and the muscles. Deep down in the abdominal cavity I found large and small tumors, some movable and others fixed by adhesions. All the lymphatic glands on the side most infected were enlarged and in masses, as if they had formed close union with surrounding tissue.

The muscles were exempt from nodules, but soft and flabby. I opened one nodule near the surface and obtained two worms. Most of the nodules on the

^a This article forms part of a paper which, by permission of the Surgeon-General, was presented upon invitation before the Sixth International Dermatological Congress, in New York, September 9–15, 1907.

skin are about the size and shape of grains of rice. When they first appear there is an itching produced. The cyst is filled with a clear watery fluid in which is found the small worm. In a few days a cyst wall is formed, surrounding the worm, which lies in a jelly or slime like substance. After weeks or months the cyst wall becomes firm, and surrounds one or more worms. In some cysts I have found as many as three worms. In the course of a few months the cyst wall breaks and there is nothing left but, sometimes, a blue spot showing a small hemorrhage; after this is absorbed, there will be only a spot of indurated tissue to mark the place. The places that have been infected the longest appear as a mass of indurated skin and fascia closely connected so that the skin can be picked up only with the mass of worm and tissue.

Patient is now 48 years old. He came to Florida in 1872 from Minnesota and settled with his father on a point on the Manatee River, now known as Manns Point, which was accessible to fish and oysters.

When 23 years old, while hunting in the woods about 1½ miles from the coast, he noticed a small pimple on the left shoulder, which attracted his attention because of the itching. He thought the skin had been punctured by a thorn. He squeezed the lump, from which came a small flat worm about ½-inch wide and ½-inch long. One year after the first appearance on the shoulder, he noticed four or five small lumps on his chest; these he opened with a knife and he picked out the same kind of worms; these swellings also had produced an itching sensation. Patient was then living as fisherman on Sarasota Bay.

While fishing, patient's diet consisted of smoked and dried fish, raw oysters, scallops, and clams. Up to five or six years ago he was a robust, healthy specimen of manhood, but lacked energy. At present he has a tired expression, has less energy, and becomes exhausted after little exertion.

From the photographs (figs. 1-2) you will see enlargement of the left breast and shoulder; also of left groin and lumbar région. Spleen and liver are enlarged.

The infection is slight on the right side.

Patient has a wife and five children, all of whom are healthy.

There probably was another case similar to this a few years ago in this county. I have been trying to obtain a history of it, but have failed thus far. The man moved from here to California, where he died. The report was that he was eaten up with worms before he died.—Extracts from letter, August, 1907.

Japanese case.—The patient was a Japanese woman, Yae Tanaka by name, resident in Tokyo or immediate vicinity. Before her marriage to a dealer in old furniture she was a weaver, "occupations which place her decidedly in the lower class of society."

At the age of 33 years, in the spring of 1904, she visited the University Hospital at Tokyo for treatment for left inguinal hernia, entering the surgical wards of Dr. J. Kondo. This hernia was traceable to the presence of parasites in the region of the ligamentum poupartii. In addition to the hernia, she presented a peculiarly swollen condition of the integument, which bore scattered spots of acnelike appearance. This abnormal condition extended over nearly the entire body, except on the face and upper extremities; it was most prominent on the left thigh, which was greatly swollen and presented very much the appearance of elephantiasis, although the skin and underlying tissues were quite soft, so that they hung down by their own weight and could be grasped in a flaccid mass by the hand.

When 25 years old the patient had a tapeworm, the species of which is not known. The dermal affection was first felt when she was 31 years old, so that at time of entrance to hospital it was of about two years' standing. It had given no particular trouble beyond that imposed upon motion by the

swollen thigh and the itching of the skin in parts where a pimplelike hardening made its appearance; scratching with the nails, in order to allay the itching, had led to breaking the skin, from which a soft whitish mass, together with some fluid, could be pressed out. A number of resulting recent scars, especially on the breasts, were visible.

In preparations, made of skin taken from the left thigh, Ijima became convinced of the presence in the connective tissue of numerous encapsuled worms, the cestode nature of which was recognized from the calcareous corpuscles.

On each of two subsequent occasions, July 9 and 24, 1904, a very large piece of skin, with the underlying connective tissue, was excised from the left thigh, in order to relieve the patient of the superfluous tissue. Altogether several pounds in weight were removed during the patient's stay at the hospital.

When freshly excised the subcutaneous tissues presented an unusual appearance. At places several centimeters thick they were moderately rich in panniculus adiposus and extraordinarily rich in lymph; the latter swelled the connective tissue between the panniculi, giving it a slimy or gelatinous appearance and consistency; the slimy character seemed more manifest in the deeper parts; lymph exuded copiously from the cut surfaces; numerous capsules, with the contained worm, were observable as whitish objects, isolated or in clusters, in all parts of the tissues. The thickness of tissues between the surface and the underlying tissue measures 30 to 60 mm., notwithstanding the fact that the hardening process has contracted the subcutaneous connective tissue, through loss of the lymph, into dense fibrous bundles, so that it no longer bears a resemblance to what it was in the fresh state. The corium in the same piece may be said to be 3 to 6 mm. thick; it seems to be on the whole considerably thicker than in the normal state.—Abstracted from Ijima, 1905, pp. 1–21.

Further medical details are not given by Ijima, but he states that Professor Kondo will publish a report of clinical and pathological observations. I have not as yet learned of the publication of the report in question.

From the foregoing abstracts the suspicion immediately arises that in Florida we have a skin infection, hitherto unrecognized for the American continent, and similar to, perhaps identical with, an infection recognized only on one former occasion, namely, in Japan.

It is interesting to note the following comparison in the cases, without laying too much stress upon these points at present:

Both cases occurred near the eastern shore of the continents in question (Old World, between 35° and 36° N., and New World, between 27° and 28° N.); both patients lived in or near cities or towns located directly on the water, very near larger bodies of water (Pacific Ocean and Gulf of Mexico); both patients, though of different sex, were adult (23 years, male; 31 years, female) when the infection was first noticed; both patients belonged to the poorer class of society; one had a professedly fish diet, the other lives in a country where a fish diet is very common; both infections are of long standing (in one case over three years, in the other case about twenty-five years); in both cases the number of individual worms present was very great; both observers (Ijima for Japan and Gates for Florida) independently call attention to the acnelike lesion resulting from the infec-

tion; each patient is a native of the country in which the case was found, and in neither case is there any history given of the patient's

having visited the country of the other patient.

NATURE OF THE PARASITE.—A microscopic examination of the worms forwarded by Doctor Gates showed them to contain calcareous corpuscles, hence the diagnosis of cestode infection was immediately established in this case on the same basis as was the diagnosis in the Japanese case. The cestode in question is a larval form, without suckers on the head, and, as far as seen, without any primordium of genital organs. The most striking feature of the worm is its irregular shape, with tendency to proliferation by forming supernumerary heads. These characters immediately bring up for consideration the question as to whether the worm found by Gates is identical with the parasite recently reported by Ijima (1905) for Japan. As the American and the Japanese parasites are very closely related, possibly specifically identical, it will be well to follow the two in comparison. In doing this it will be advisable to abstract Ijima's paper rather liberally, more especially since it is published in a journal not generally accessible to dermatologists.

The worm capsule of Ijima's parasite.—Ijima (1905, 4–5) states that the worm capsules of various sizes occur in abundance in all parts of the subcutaneous tissues and less abundantly in the corium. They were also observed in some numbers in the intermuscular connective tissue, but not in the muscles themselves, so far as such observation could be made on parts incidentally exposed during the surgical operation. In the corium the capsule may be situated so close to the epidermis that the latter is externally raised into an acnelike prominence. On a piece of the preserved skin about 2 inches square, Ijima found at least four such prominences, which, as seen on the surface, appear smooth and less pigmented than the surrounding parts. Capsules so superficially situated might easily be ruptured

by force applied through the skin from without.

In shape the capsules are generally subspherical or ovoid. While the smallest of them are considerably less than 1 mm., others measuring 1 to 2 mm. or more are of quite common occurrence; one of the largest seen was elongate, 2.5 mm. broad by 8 mm. long; another measured 3 mm. by 6 mm. The larger capsules were found only

in the subcutaneous parts, not in the corium.

The capsular wall, consisting of a dense felt work of connective tissue fibers of the host, may reach nearly 0.33 mm. in thickness; in sections the capsules may appear not unlike a transversely cut blood vessel on account of the tough and compact looking wall; under a hand lens the inner surface of the capsule appears smooth; in some of the larger cysts the internal cavity is traversed by-branching trabeculæ; microscopically the wall either shows no special limit-

ing structure, or is lined with a deposit of granular coagulum or tissue débris.

Abundance of parasite.—In Doctor Gates's letter of June 17 he states that his "patient has thousands of" these parasites.

In the Japanese case a section of about 11 sq. cm. showed nearly 60 capsules; in the most thickly infested portions of the thigh there was one capsule to every 20 sq. mm. of cut surface, or to every 100 cub. mm. of infested tissue; this gives 1,000 capsules per 100 cub. cm. of tissue. It was estimated that there must have been considerably over 10,000 capsules in the left thigh alone.

Worms without capsules .- Scarely any of the worms Gates forwarded to me bore any remnant of the cyst. Very probably most of these specimens were originally encysted and were freed from their cyst by Doctor Gates before he forwarded them.

Comparatively young, slender worms were found by Ijima free in the connective tissue—that is, not surrounded by a capsule.— Ijima, 1905.

Movements of the worm.—Not having seen the worm alive, I can give no details regarding movements.

Ijima reports that the living worms when taken from the patient showed slow movements of extension and contraction, but effecting little or no change in position; upon cooling the worms no longer exhibited such movements; in case of worms placed in salt solution motion could be revived up to a period of four hours if the parasites were slightly warmed.

The head (narrow end) was the most motile, evaginating and invaginating at the apex, in addition to shortening and extending; the terminal, but inconstant, depression in some cases reminded the observer of a terminal sucker, such as seen in the fish bothriocephalid Cyathocephalus. In addition to a motion as if feeling about, the head started a lively peristalsis from before backward; such combined movements would aid the worm in penetrating and moving through tissues.

The broader parts of the body showed at most slow vermiform movements, with more or less constant indentation at the extreme hind end.

The head.—Ijima states that the head of his parasite is devoid of any definitely formed or permanent organ of attachment. This holds true also for the worm found by Gates. In some few specimens a slight apical depression is observed, but as the material is preserved in alcohol this might possibly be either an artifact or a depression due to sudden contraction on the part of the worm.

Encysted worms.—Gates writes that he found as many as 3 worms in one cyst. This condition is doubtless due, as Ijima explains also for his case, to the tendency of the worm to multiply by budding.

Ijima reports that the smaller capsules usually contained only a single worm; in the larger cysts, however, 2 or more worms or pieces were more frequently found; from one capsule 5 worms were obtained, and from another 7 worms.

Size and shape.—The longest specimen I have observed is 12 mm. in length. Some of the worms are simple elongated bodies, either more or less flattened, or nearly round in transverse section. The larger specimens, however, assume all manner of bizarre and irregular shapes which can not be well described. These variations in form may be reduced to a progressive but irregular formation of buds, the apex of each bud representing a structure similar to the cephalic end of the original worm: the form varies, of course, according to the number, position, contraction, etc., of the buds, and according to the contraction of the parent stock. Figures 5 to 15 will give an idea of the great variety of forms found.

According to Ijima, many of the worms are small, filiform, about 0.3 mm. in diameter. 3 mm. in length: others attain, even when moderately contracted, 12 mm. long by 2.5 mm. broad. In some specimens the body is flattened dorsoventrally, but there is no clew as to which is the dorsal and which is the ventral surface. In its simplest form the worm is plerocercuslike, or narrow at the head and broader caudad when moderately contracted, or irregularly cylindrical when strongly contracted.

This simple plerocercuslike larva, when encysted, may assume a rather complicated structure, due to its ability to form buds or supernumerary heads, especially on the lateral edges of the flattened body in younger specimens, but quite irregularly in the more complicated older forms. When the heads detach themselves they represent small independent plerocercuslike larvæ, and their method of formation explains the presence of several worms in one cyst.

The formation of heads in the manner described naturally tends to give the worms a very irregular outline; this irregularity is increased by the formation of subcuticular bodies, which Ijima interprets as food material. Ijima assumes that these young heads leave the capsule and wander through the connective tissue until they grow in size, and then in turn form a capsule of their own.

Microscopic anatomy.—According to Ijima, the cuticule of the Japanese form may attain S_{μ} in thickness; the dermal musculature consists of external circular and internal longitudinal fibers. These

statements are correct as applied to the Florida form also.

The calcareous corpuscles of the Japanese worm are described by Ijima as spherical or ellipsoidal, 7.5 to 12μ in diameter, and abundant in all parts of the parenchyma, except in the head, in which they are lacking. In the Florida form also the calcareous corpuscles are abundant; they vary in size from 8.8 to 17.6μ ; in shape they are

spherical to ellipsoidal. Thus, in reference to the size of the calcareous corpuscles, there seems to be a slight difference between the American and the Japanese parasites.

Reserve food bodies.—Ijima has described as present in the parenchyma certain bodies which he views as reserve food material, Usually they are roundish or oval, 100 to 300μ in diameter: but they may become very elongate.

In the Florida parasite similar bodies are present, but in the specimens thus far examined microscopically they do not seem to be quite so numerous or quite so large and prominent as described by Ijima for the Japanese form. It is possible, however, that this is a matter of individual variation.

Excretory system.—The parasites, as described by Ijima, contain an extensive system of anastomosing excretory vessels. In this respect the Florida form agrees with the Japanese species. Some of the canals are quite large, others are smaller, some are very small. Ijima calls attention to the absence of excretory vessels in the peripheral zone of the posterior part of the body: he also states that he was unable to find any opening.

In sections of one of the Florida parasites fine canals were found rather close to the cuticle, but because of the branched condition of the worm it is difficult to state just what portion of the body this was; it was not, however, a head. Likewise, in one case sections of a pore (fig. 18) on the surface, with a centripetally directed canal, were distinctly seen. In view of the absence of genital organs one is naturally inclined to look upon this pore as belonging to the excretory system.

The longitudinal muscles of the Japanese worm are described as well developed, and in addition there is a less strongly developed set of muscle fibers running in different directions but mainly in the transverse plane. Near the head these transverse muscles may be quite regular (some dorsoventral, others crossing these at right angles), but in thicker portions of the body they may become very irregular. This description applies in a general way to the Florida form also.

Nervous system.—Ijima noticed a pair of longitudinal, lateral nerve trunks in the cephalic portion; they seemed to unite close to the tip of the head. In several sections of the Florida worms nerves were distinguished, but details as to their topography were not studied.

Life cycle.—Experiments to raise the adult stage by feeding the Japanese parasites to cats, dogs, and pigs were negative.

As all of my own material was preserved no experiments could be undertaken. The question as to the source of infection, life cycle, etc., must be left open for the present.

Systematic position.—From the general structure, especially from the presence of calcareous corpuscles, it is clear that both the Japanese and the Florida parasites are cestodes; the absence of suckers seems to place both forms in the old family Bothriocephalidæ, now known as Dibothriocephalidæ. Further than this, the exact systematic position is not clear at present and can not well be determined until the adult stage is known.

So far as can be judged from the material thus far studied (prior to the meeting of the International Dermatological Congress in New York, September 9, 1907), the Florida form must be considered as very closely allied to, perhaps specifically identical with, the Japanese form. The only anatomical point of difference thus far brought out is a difference in size of the calcareous corpuscles; the only biological difference known is the habitat—in two widely separated localities.

Ijima points out the structural affinities between the Japanese form and the bothriocephalid larval *Sparganum* of Diesing; he refers also to the similarity between the Japanese form and "*Ligula mansoni*" (=*Sparganum mansoni*).

Sparganum is an artificial collective group of worms, distinctly proposed, not as a systematic unit, but as a collective group of larval bothriocephalid cestodes. Under the International Code of Nomenclature (1907) such names may be proposed as a matter of convenience and may be used as if they were generic names; they do not require any type species and hence do not compete with generic names under the law of priority.

Both the Japanese and the American parasite may be temporarily classified in *Sparganum*.

The Japanese worm was originally published under the two names *Plerocercoides prolifer* and *Plerocercus prolifer*, but Ijima distinctly states that he uses the names as a matter of convenience, namely, not in a taxonomic sense. Certain objections arise, however, to the use of the names *Plerocercus* and *Plerocercoides* in this connection, and on this account I transferred (1906a) the parasite to *Sparganum*.

The nomenclatural points involved are somewhat complex and it may be well to explain them in this place.

Under the original international code, the names of larval cestodes and of certain other forms were, for special reasons, exempted from the law of priority. Later (1901), contrary to the judgment of helminthologists, this exemption was done away with.

To apply the law of priority consistently to all such larval names would be almost an impossibility. There are, in fact, many names which have been proposed, not in a generic sense, but as names of admittedly artificial groups, which were used simply as a matter of

temporary convenience, and it was on account of a failure to distinguish between names of this category and names proposed for supposed genera that the exception was rescinded. In 1907, at the Boston congress, a helminthologist proposed the following, which was adopted as part of the code:

"Certain biological groups which have been proposed distinctly as collective groups, not as systematic units, may be treated for convenience as if they were genera, but they require no type species. Examples: Agamodistomum, Amphistomulum, Agamofilaria, Agamo-

mermis, Sparganum."

As will be shown below, *Plerocercus* and *Plerocercoides* also now come under this paragraph. As matters now stand it is necessary to show that a name was distinctly proposed to designate an artificial collective group in order to bring it under this provision. A name like *Cysticercus* or *Echinòcoccus*, originally proposed as generic, not distinctly to cover an admittedly artificial group, is subject, now as before, to the law of priority.

The term *Plerocercus* (πλήρης, full; κέρκος, tail) was proposed by Braun (1883a, 98) as designation for the parenchymatous cysticerci (namely those the caudal portion of which contains no fluid) as distinguished from the bladder worms, or true cysticerci; thus it is a descriptive term for a stage of development (larva) possessing certain characters, but not the designation, of a systematic unit. As examples, Braun cited a *Plerocercus* [*Dithyridium lacertæ* Val.] of lizards and a *Plerocercus* of *Tetrarhynchus*. This plerocercus of the lizards happens to figure in a true nomenclatural sense, namely in the genus—

DITHYRIDIUM Rudolphi, 1819.

1819: Dithyridium Rud., 1819a, 559 (lacertæ viridis, lacertæ muralis; Europe. Type by later absolute tautonymy Piestocystis dithyridium = Dith. lacertæ).

1850: Piestocystis Dies., 1850a, 478, 494-496 (Dithyridium 1819, renamed; includes as valid species P. crispa (Rud.), P. rugosa Dies., P. variabilis Dies., P. dithyridium Dies., [for Dithyridium of Rud., namely D. lacertæ Valenciennes, 1844]).

This genus is based upon a larval tapeworm (a plerocercus) provided with four suckers, and is classified in the family Taniidae; the type species occurs in Europe in lizards of the genus *Lacerta*.

In 1866 Baillet mentioned a parenchymatous cysticercus (namely a plerocercus) from the abdominal cavity of the cat and the dog. In 1882 or 1883. Blumberg described this form as a new species under the name Cysticercus elongatus; the latter name however was already preoccupied (cf. C. elongatus Leuck., 1842). In 1885, Railliet renamed this form Cysticercus bailleti, and in 1893a, p. 314, he classi-

fied it in the genus *Dithyridium* as *D. elongatum*. In the meantime, however, Neumann (1892a, 537-539, figs. 292-293) referred to the same parasite as "*Plerocercoides bailleti*," clearly using a Latin binominal nomenclature.

The name *Plerocercoides* as used by Neumann is traceable to Braun (1883a, 100) who used a German term ("Plerocercoiden") to designate certain larval forms which differed from the cysticercoids by having parenchymatous tails. As examples Braun cites a form which occurs in the body cavity of *Trichodectes canis*, and the young (*Gyporhynchus* of *Tænia macropeos* and *T. unilateralis*. Blanchard (1888a, 491) used a French form (*Plérocercoides*) of the word, while Neumann (1892a, 537) seems to have first used the Latin *Plerocercoides*. From Neumann's text, however, it is clear that he based his name on Braun's "Plerocercoiden;" hence Neumann's *Plerocercoides* is not a generic name but the designation of an artificial collective group; hence also the type designation (bailleti) suggested by myself in 1906 is not necessary under the new (1907) code.

As a plerocercus may be the larval form of species belonging to widely distinct families, even to different orders, it is wise not to use the combination *Plerocercus prolifer* in case a better designation is available; this point probably occurred to Ijima, for he used the combination only once. *Plerocercoides* is also open to the same objection, and in the only use of the term prior to Ijima it was used for a *Dithyridium*.

It so happens that Diesing proposed a name which is open to fewer objections. This is the

Collective Group SPARGANUM Diesing, 1850.

DIAGNOSIS.—Dibothriocephalidw: An artificial collective group to contain larval stages of bothriocephalid worms which have not reached a stage in their development that they can be determined generically.

Such groups do not require a type species.

In 1906 I placed Ijima's form in this group as

The Proliferating Japanese Tapeworm Larva—SPARGANUM PROLIFERUM (Ijima, 1905) Stiles, 1906.

Specific diagnosis.—Sparganum: Larva may attain 1 to 12 mm. in length and 2.5 mm. in breadth; head narrower and more motile than posterior end and may show an apical depression which, perhaps, serves as sucker; no true suckers or other organs of attachment present. Calcareous corpuscles spherical or ellipsoidal, 7.5 to $12\,\mu$ (Japanese worm) or 8.8 to $17.6\,\mu$ (Florida worm) in diameter, and situated in any part of body except head; irregularly distributed reserve food bodies present in older specimens, but they later undergo disintegration; genital organs not present; longitudinal muscles better developed than either dorsoventral or transverse system; transverse fibers do not

divide body into cortical and medullary layers; excretory system well developed, consisting of larger approximately longitudinal branches, with anastomoses. The larvæ possess the power of multiplying by transverse fission and of forming supernumerary heads which may become independent. Adult unknown.

Habitat.—Encysted in subcutaneous tissue and elsewhere in man.

GEOGRAPHICAL DISTRIBUTION.—Found but twice; once by Ijima in Tokyo, Japan; once by Gates in Manatee, Fla.

Whatever results may be obtained from examination of further material, which I could not study prior to the meeting of this Dermatological Congress, at the present time I do not feel justified in separating the American form specifically from the Japanese species, despite the difference in geographic distribution and the slight difference in the calcareous corpuscles. That the adult stage may eventually prove the Florida form to represent a distinct species seems entirely possible; in fact, when we consider the seeming isolation of the two cases this appears probable. At the same time, if it should eventually be shown that the infection was contracted from eating marine fish, the possibility would not be excluded that the two forms are identical, despite the wide difference in locality. In the interest of conservatism, accordingly, I classify for the present the two in the same species.

In an earlier paper (1906a, 86) I called attention to the fact that because of the remarkable reproduction of the larval stage described by Ijima a new genus would probably be justified. I hesitated somewhat to make the genus without seeing actual specimens. After examining the American specimens I am further convinced of the probability that the worm in question represents a new genus.

The proposition of a new generic name at this time presents both advantages and disadvantages. To continue to call the worm Sparganum shows that the family position is recognized, but that the adult is unknown; the worm is, however, so different from the other forms of Sparganum that it seems advisable to bring out this difference in a generic name; further, as long as a new generic name seems almost inevitable, it would appear wise to introduce it as soon as possible, in order to reduce its competition (through homonymy) in the future. On the other hand, to introduce a new combination at present does not seem absolutely necessary; its introduction would destroy the advantages we have at present in the use of the name Sparganum.

There is, I believe, a conservative method by which the advantages of both plans may be united, namely, by the introduction of a new subgeneric name. This course permits the continuation of the use of the name *Sparganum*, and at the same time brings out the fact that the worm is very different from the other forms of *Sparganum*;

further, it procures for the name any advantages in homonymy which may be gained by its proposal this year instead of later.

As such subgeneric name I propose-

GATESIUS a n. subg.

Diagnosis.—Dibothriocephalidæ,? Ligulinæ, classified temporarily in Sparganum: Adult unknown. Larva in its simplest form similar to the plerocercus of Dibothriocephalus, except for suckers, which are not indicated; possesses the property of branching and of reproducing by budding, thus forming supernumerary heads which become free from parent and assume the simple plerocercoid form. Body contains numerous calcareous corpuscles, richly developed canal system, and may contain reserve food bodies.

Type species.—Sparganum (Gatesius) proliferum (Ijima, 1905) Stiles, 1908, from connective tissue of man; Japan.

It will be noticed that from the form of this proposal the name *Gatesius* is proposed not as a collective group, similar to *Sparganum*, but as a true systematic name of subgeneric rank. When the adult form becomes known the species should be taken out of the group *Sparganum* and this subgenus raised to generic rank.

 $^{^{}a}$ Dedicated to Dr. H. Gates, of Manatee, Fla., who found the first American case.

A REEXAMINATION OF THE TYPE SPECIMEN OF FILARIA RESTIFORMIS LEIDY, 1880, = AGAMOMERMIS RESTIFORMIS.

By CH. WARDELL STILES, Ph. D.

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(Figs. 19 to 26.)

While examining some specimens in the Army Medical Museum with the late Dr. James Carroll, I found Leidy's original specimen of *Filaria restiformis*, reported as a parasite of man. As the parasite has been reported but once, and as little is known regarding its nature, I requested Major Carroll to place the specimen at my disposal for study. This he kindly did.

HISTORICAL REVIEW.—Leidy (1880c; 1904a, 157-159) received from Dr. J. J. Woodward, U. S. Army, a specimen of a worm which had been sent to the Army Medical Museum, Washington, D. C., by Dr. C. L. Garnett, of Buffalo, Putnam County, W. Va., together with the following extract from a letter:

During the winter of 1876 a man, a common laborer, aged about 50, presented himself to me for treatment, having a gleety discharge from the urethra, with a burning sensation during and after micturition. Previously he had been treated for gonorrhea, and I prescribed accordingly. The patient, not improving, applied to other practitioners. In April, 1878, he came to me with a round, vivid-red worm, 26 inches in length (the specimen you now possess), and very active in its movements, instantly coiling up like a watch-spring on being touched. Having no work on helminthology for reference, the only description I found which appeared to answer to the worm was that of Strongylus gigas, in Niemeyer, Vol. II, p. 47. The patient is an illiterate man, with no motive for deception. He informed me that he discovered the worm protruding from his penis and drew it out without pain or difficulty. He was in much agitation and alarm about the occurrence, fearing, as he said, that "there might be more behind that one." For a few days previous to its passage his urine was of a milky hue and some time subsequently of a yellow cast and slightly tinged with blood and mingled with mucus. The man is truthful, and no doubt exists in my mind or in the minds of his neighbors as to the correctness of his statements. I regret exceedingly that I did not appreciate the scientific interest of the subject, and send you the specimen in a fresh state, but the busy routine of a country practitioner's life leaves no time for the study of other than subjects of practical value in one's everyday experience.

Leidy seems to have had some slight misgivings as to whether the worm actually represented a parasite of man, but he described it as a new species, with the following diagnosis:

Body long, restiform, nearly uniformly cylindrical, smooth, shining, elastic, tough, without evident annulation other than transverse wrinkling, with the anterior extremity evenly tapering in the continuous head, the end of which is rounded and smooth or without appendages of any kind; the posterior extremity not tapering, with the caudal end incurved, bluntly rounded, without appendages and imperforate or without evident anal or genital aperture. Mouth a terminal pore without lips, papillæ, or armature of any kind. Pharynx cylindrical and opening into a straight cylindrical intestine, apparently ending in a blind pouch. Generative organs unobserved. Length of worm 26 inches, greatest thickness 1.5 mm. Width of head just behind the rounded extremity 0.375 mm., opposite the commencement of the intestine 0.625 mm., at the middle 1.5 mm., at the incurved caudal extremity 1.5 mm. Length of esophagus 1.125 mm.

Leidy adds that the worm is clearly neither a Gordius nor a Mermis.

Neither the figures nor the description as given by Leidy seem to be strongly confirmatory of the determination of *Filaria*. In fact, the description and the figure of the esophagus give rise to the question whether the worm may not belong to the *Mermithidæ*, while the striation figured for the caudal end nearly establishes this point. Authors since 1880 have referred to *F. restiformis*, usually without expressing an opinion regarding the species, but Railliet (1893a, 530) and Penel (1905, 8) consider that it was probably a spurious parasite.

Bibliography.—1880: Filaria restiformis Leidy, 1880c, 130-132, figs. 1-2 (in Homo; West Virginia); 1904a, 157-159, 278.—R. Blanchard, 1890a, 13, fig. 390 a-b; 1895, 785.—Braun, 1883a, 184; 1895b, 227; 1903, 3 ed., 275.—Ijima, 1889b, 367.—Moniez, 1896, 359.—Penel, 1905, 8.—Railliet, 1893a, 530.—Stossich, 1897, 78.—Vaullegeard, 1901, 128.—Ward, 1895, 331; 1903, 704; 1903, 212.

REEXAMINATION OF TYPE.

Condition of material.—The worm is broken into several fragments and is not suitable to a detailed study. It was in alcohol when Doctor Carroll turned it over to me. It was next transferred to alcohol and glycerin; the alcohol was evaporated, and the fragments were studied in glycerin. Very few characters can be recognized, but sufficient was seen to exclude the worm from the genus Filaria.

General contour.—By placing the various fragments together figure 21 is obtained, showing the general appearance of the worm, natural size. It will be noticed that while the body is of nearly uniform diameter, the head is distinctly attenuated.

Cuticle.—The cuticle is about 32 μ thick on the anterior portion, 48 μ on the tail, and when focused sharply is seen to be composed of several concentric layers; this point is brought out clearly at the ends of the fragments and on transverse section. Sharp focus at high power shows in addition the distinct presence of a diagonal fiber system, such as is found in the Mermithidæ, and such as was clearly seen by Leidy and figured in his original publication.

Head.—The cephalic extremity is distinctly attenuated, as observed by Leidy, and then ends rather bluntly. The terminal mouth is very small, and without lips. Directly back of the mouth there are six papillæ, which can still be observed more or less distinctly. The presence of these papillæ definitely excludes the worm from the Gordiidæ. Unfortunately it is impossible to roll the head, as it is somewhat compressed, apparently dorsoventrally. Two of the papillæ seem, however, to be anterior of the other four; it seems probable, therefore, that the former are lateral; the latter the submedian papillæ.

About 442 μ back of the mouth a structure was seen which bore some resemblance to an excretory pore with canal. If, however, the position of the papillæ is correctly interpreted, this excretorylike structure would be lateral, hence this can hardly be the excretory pore, but is more probably an artifact.

Tail.—The tail is curved ventrally and is bluntly rounded. No anus was discernible. In the body could be seen a dark cecal structure. Caudal papillæ were not observed. It would appear, accordingly, that the specimen is either a female or that it is immature.

Esophagus.—In describing the intestine Leidy says: "Pharynx cylindrical and opening into a straight cylindrical intestine, apparently ending in a blind pouch." This description immediately suggests the Mermithidæ. The only other group which would seem to come into consideration would be Dracunculus, which might be considered as a possibility on the assumption that the structures were to be interpreted similarly to Looss's 1905, fig. 39, p. 183, of Dracunculus medinensis. Such interpretation is not in harmony, however, with the diagonal fibers of the cuticle.

From the mouth a distinct cuticular tube, 17.6μ to 26.4μ in diameter, cuticle 8μ , lumen 9 to 10.4μ , can be traced for some little distance; this tube evidently represents the cuticular lining of the esophagus, and it is surrounded by a somewhat indistinct body, evidently the esophagus, about 130μ in diameter.

At a point 0.88 mm. from the anterior extremity there is seen at one side of the esophageal cuticular lining a blind sack, directed cephalad, and apparently representing the fat body reported for the *Mermithidæ*.

Genital organs.—No genital organs can be discovered, so that the conclusion is drawn that the specimen is immature.

Cross section.—Advantage was taken of the fact that the specimen is in fragments to prepare transverse sections (fig. 26) of the end of one of the pieces. A careful histological study of the slides is precluded by the condition of the material. Nevertheless, some important anatomical observations are possible.

The cuticle consists of concentric layers. A subcuticula can also be distinguished more or less clearly. The muscles are divided into six fields, separated by the six longitudinal lines, but the finer structure

of these lines can not be interpreted safely.

In view of the foregoing data it appears that Filaria restiformis should be transferred from the Filariidæ to the family Mermithidæ, as Agamomermis restiformis. Further, the idea that an error has occurred in interpreting this worm as a parasite of man seems to gain support, for it would be exceedingly difficult to explain the presence of a Mermithidæ in the bladder.

OBSERVATIONS ON TWO NEW PARASITIC TREMATODE WORMS: HOMALOGASTER PHILIPPINENSIS, AGAMODISTOMUM NANUS.

By Ch. Wardell Stiles, Ph. D., Chief of Division of Zoology, and

JOSEPH GOLDBERGER,

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Among the parasites recently sent to us for identification are two interesting trematodes which appear to be new species. One of them (*Homalogaster philippinensis*) is an amphistome from the Philippines, the other (*Agamodistomum nanus*) a distome from Africa.

Genus HOMALOGASTER Poirier, 1883.

1883: Homalogaster Poirier, 1883, 74-76, 79 (m. palonix); ὁμαλός, flat; γαστήρ, stomach; 1885, 120. For bibliography see Stiles and Hassall, 1908, Index Cat., Trematoda, Bull. 37, Hyg. Lab., U. S. P. H. & M. H. S.

Generic diagnosis.—Paramphistomidæ,? Cladorchinæ: a Body divided into a large oval, anteriorly pointed, cephalic portion containing all of the inner organs, and a caudal portion composed of the large acetabulum. Dorsum convex. Venter flat or excavate, provided with alternating longitudinal rows of large mammalike structures; of these, the papillæ near the middle of the rows (both longitudinal and transverse) are larger than those further from the middle. Pharynx with two lateral pouches; esophagus present. Genital pore ventromedian, cephalad of testicles. Testicles in anterior half of body; ovary near acetabulum; vitellaria rather well developed, lateral of ceca, extending about from esophageal bifurcation to ovary. Egg with operculum.

Habitat: Large intestine of ruminants.

Type species.—Homalogaster paloniæ Poirier, 1883.

Comparatively little work has been done on this genus. In fact, Fischæder was obliged to leave open certain questions in regard to the generic characters. With the new form here described, the genus contains three species. An effort to obtain the two known species for

 $^{^{}a}$ In a later paper we shall have occasion to discuss the subfamily position of this genus.

purpose of comparison has not been successful. The three species in question may be distinguished by the following key and diagnoses:

KEY TO THE SPECIES OF HOMALOGASTER.

- 2. Testicles, one caudad of the other: oral papillæ present; genital pore at equator of esophagus; type locality, Manila, P. I.

H. philippinensis (p. 25).

3. Testicles on same transverse plane and divided into 2 large equal lobes, simulating 4 testicles; oral papillæ present; genital pore caudad of bifurcation of intestine; type locality, Tonkin____H. poirieri (p. 24).

In view of the comparatively restricted knowledge of this genus it seems advisable to give our present data regarding the two species already described before passing to a discussion of the new form.

HOMALOGASTER PALONIÆ Poirier, 1883.

(Figs. 27 to 28.)

1883: Homalogaster paloniæ Poirier, 1883, 74-76, pl. 2, figs. 1a-b (cecum of Palonia frontalis; from Java).—For bibliography see Stiles and Hassall, 1908. Index Cat., Trematoda, Bull. 37, Hyg. Lab., U. S. P. H. & M. H. S.

Specific diagnosis.—Homalogaster (p. 23): Something over 12 mm. long; form lanceolate; anterior cephalic portion very pointed, 11 mm. long by 6 mm. in maximum breadth; caudal portion much shorter, 3 mm. in maximum breadth. Genital pore on center of a papilla 1.5 mm. from very small terminal mouth. Venter with alternating longitudinal rows of large papillæ extending from about 3 mm. from oral margin to acetabulum. Acetabulum 2 mm. in diameter. Oral papillæ are not reported. Esophagus bifurcates 3 mm. from mouth; intestinal ceca extend to end of anterior portion of body.

Male organs: Testicles small, placed obliquely in cephalic half of body between ceca; one slightly to the right of the median line and 4 mm. from oral margin: the other 2 mm. further caudad, slightly to the left of the median line.

Female organs: Ovary 9.5 mm, from oral margin; vitellogene glands ramified, along the sides of body; uterus almost without coils. Excretory pore dorsal, 12 mm, from mouth; excretory vesicle with 2 lateral lobes.—Based on Poirier, 1883, 74-76.

Habitat.—In cecum of "Palonia frontalis," from Java.

HOMALOGASTER POIRIERI Giard and Billet, 1892.

1892: Homalogaster poirieri Giard and Billet, 1892a. 615 (large intestine of cattle; 'Tonkin).—For bibliography see Stiles and Hassall, 1908, Index Cat.. Trematoda, Bull. 37. Hyg. Lab.. U. S. P. H. & M. H. S.

Specific diagnosis.—Homalogaster (p. 23): Size not given. Mouth with slender digitate papilla. Genital pore caudad of intestinal bifurcation, on elevated papilla, visible to naked eye, a little behind pharynx. Acetabulum very large. Pharynx followed by 2 long simple ceca, which extend in almost a straight line to nearly the latitude of acetabulum (?? indicating absence of esophagus). Vasa deferentia (?efferentia) short, thick, tortuous; testicles lat-

eral (apparently meaning on same transverse plane), divided into 2 large equal lobes with sinuous margins, thus giving the appearance of 4 testicles occupying the corners of a square. Uterus with numerous slings, crowded together, and crowded with eggs in different stages of development; vitellaria lateral of intestinal ceca, with grapelike follicles; vitelline canals unite in posterior part of body, above (cephalad? or dorsad? of) acetabulum in the open space; ovary and shell gland near median line.—Based on Giard and Billet, 1892a, 615.

Habitat.—Attached to mucosa of large intestine of cattle Bos taurus? or B. taurus indicus? at Tonkin.

Giard and Billet do not describe the form of the body, but state that it is different from that of *H. paloniæ*.

HOMALOGASTER PHILIPPINENSIS new species.

(Figs. 29 to 44.)

Specific diagnosis.—Homalogaster (p. 23): Body 7.5 to 9 mm. long by 4.5 to 5 mm. (or flattened to 7 mm.) broad, canoe shaped, cephalic extremity attenuated, caudal extremity rounded; when flattened, sides very convex; dorsum may show submedian longitudinal depressions. Genital pore 1 mm. from oral margin (about one-ninth to one-seventh of body length from mouth, and about at equator of esophagus) surrounded by depressed circular area bearing numerous small papillæ. Venter with alternating longitudinal rows of large papillæ extending from about 2 or 2.5 mm. from oral margin to acetabulum. Margins curved ventrally and are fairly sharp. Acetabulum about 2.5 by 2.7 to 3 mm. Esophagus extends to about one-fifth of body length from oral margin; intestinal ceca narrow, slightly tortuous, long, extending to acetabulum.

Male organs: Testicles lobate, in cephalic half of space between ceca; one caudad of the other; vesicula seminalis quite compactly coiled; pars muscularis distinctly but not highly developed; pars prostatica not prominent, may enlarge to quite a large diameter, thus resembling a "vesicula seminalis interna;" ductus ejaculatorius present; cirrus absent.

Female organs: Ovary and shell gland submedian, near end of one of the ceca, ovary cephalad of shell gland; vitellaria extend about from the bifurcation of esophagus to caudal plane of ovary; uterus with many coils, well developed, passes cephalad, dorsally of testicles, ventrally between vasa efferentia, ventrally of vas deferens, to pore; Laurer's canal opens dorsomedian at plane of termination of ceca. Genital papilla, when extruded, resembles human penis' with glans; bears on its vertex a pore, from which runs the short ductus hermaphroditicus; when retracted the papilla resembles a partially inclosed cirrus pouch. Egg oval, operculated, with small knob at opposite pole; 125 to 139μ by 67.5 to 86μ .

Excretory pore dorsomedian about at equator of acetabulum.

Habitat.—Cecum of Bos——, at Manila, P. I. (type locality), and Phrapatoom, Siam.

Type.—U. S. P. H. & M. H. S. 9580 (mounted): Cotypes 9581 and 9960.

Source of material.—This species has been sent to us on two occasions by Dr. Paul G. Woolley. The first specimens (U. S. P. H. & M. H. S. Nos. 9580 and 9581) were collected in 1904 from the cecum of two calves (Bos———————————) at Manila, P. I.; the second sending (U.

S. P. H. & M. H. S. No. 9960) was collected from the small intestine of a cow (*Bos*——), Sept. 28, 1906, at Phrapatoom, Siam. No difference has been noticed between the Philippine and the Siam specimens.

EXTERNAL CHARACTERS.

Size.—The specimens vary from 7.5 to 9.5 mm. in length, and from 4.5 to 5 mm. in maximum breadth; may flatten under pressure to 7 mm. broad.

Color.—Alcohol specimens are of a grayish buff color.

Form.—In form the fixed specimens are canoe shaped with an attenuated oral and a rounded caudal extremity. The body is divisible into a posterior somewhat cylindrical portion, forming about one-fourth of the total length and an anterior somewhat lancet-shaped portion forming the remaining three-fourths. The posterior cylindrical portion is slightly flattened on its ventral aspect on which it bears the aperture of the acetabulum. The anterior three-fourths of the body is arched dorsally and excavated ventrally. The dorsum may show two longitudinal depressions just mediad of the intestinal ceca. In some of the specimens the lateral margins have curled ventrad in a scroll-like fashion, but it is possible to flatten the specimens without breaking the margins.

Surface.—The surface of the rounded posterior portion is smooth; that of the anterior portion is smooth dorsally (without spines or papille), but ventrally it is provided with two sets of dissimilar papillæ. One of these sets consists of numerous small papillæ covering a circumscribed, slightly depressed area around the base of the genital papilla: when the genital papilla is not extruded, the genital pore is still in the center of a mammalike elevation (fig. 41), which is much larger than the circumgenital papilla just mentioned, and which is separated (in sagittal sections) about 1 mm. from the oral margin. The other set of papillæ consists of large and quite prominent elevations arranged in parallel longitudinal rows, extending from about 2 to 2.5 mm. from the oral margin to the rounded caudal portion of the body: laterally they do not attain quite the margins of the body: the papillæ near the median line are the largest, those near the lateral margins are smaller; further, the most anterior and the most posterior of the papillæ incline to a smaller size: in their arrangement the papillæ of any given longitudinal row are situated more or less regularly to correspond to the spaces between the papillæ of the next longitudinal row; as a result, they appear not only in longitudinal, but also in diagonal rows.

The venter and dorsum meet laterally in fairly sharp margins.

Genital pore.—As indicated above, the genital pore (figs. 31 and 41) is ventromedian about 1 mm. from the oral margin, namely,

about one-ninth to one-seventh of the length of the worm from the oral margin.

Acetabulum.—Measurements of the acetabulum taken from press preparations make it about 2.6 mm. in longitudinal diameter by 2.75 to 3 mm. in transverse diameter. The aperture is rather oval in outline, about 1.55 mm. in transverse by 1 mm. in longitudinal diameter.

INTERNAL ANATOMY.

DIGESTIVE TRACT.—The anterior bluntly pointed, attenuated extremity (cephalic cone) is pierced by the mouth, which is not provided with a true sucker. The oral margin is surrounded by a number of slender, digitate papillæ. The mouth gives entrance to a muscular pharvnx; the lumen of the latter is at first more or less circular in transverse section, but it rapidly becomes flattened and crescentic with the convexity of the crescent dorsad. From the horns of this crescent spring two lateral pouches and the lumen of the pharynx terminates blindly as a narrow crescentic slit connecting the two lateral pouches. From the ventral aspect and a little above the blind, slitlike termination of the lumen of the pharynx, the esophagus rises and passes caudad, forming a ventral curve in its course. At its point of origin it communicates laterally with the lumen of the pharyngeal pouches, between and a little ventrad of which it lies. The relation of the esophagus to the pharvngeal pouches and blind slit will be seen by reference to figures 32 to 36.

At about one-fifth of the total body length from the oral margin of the worm, the esophagus divides into two simple cecal tubes which pass at first laterocaudad, then in slightly tortuous line caudad to about the level of the cephalic margin of the acetabulum. Just before reaching their termination they shift a little toward the median line. One tube may be somewhat longer, and so terminate at a slightly lower level (more distal) than the other. In their course they are wavy and are separated by a distance about equal to that which separates them from the corresponding lateral margins of the worm. The pharynx, pharyngeal pouches, and esophagus are lined by a cuticle in anatomical continuation with that of the surface. At the esophageal fork this lining abruptly ceases, the intestinal tubes being lined by a nucleated epithelium.

Genital system.—Male organs: The two testicles lie in the cephalic half of the area included between the intestinal ceca, and are somewhat nearer the dorsal than the ventral surface. (Fig. 31.) One is directly caudad of the other, the two being separated by a narrow interval which is partially occupied by some coils of the uterus. The anterior testicle is somewhat smaller than the posterior and in press preparations their outlines are sinuous, but sections

show them to be so markedly indented as to amount to a very distinct lobation. A vas efferens rises from each testicle; that from the posterior testicle appears to spring from the right dorsolateral aspect and passes cephalad between the anterior testicle and the right intestinal cecum; that from the anterior testicle springs from near the left margin of its cephalic aspect, and passing cephalad, it unites with its fellow beneath the esophageal fork to form the vas deferens. In their course they both become dilated and contain masses of spermatozoa. The vas deferens winds its way cephaloventrad in close relation with the coils of the uterus, and opens to the exterior by a pore common with the opening of the female duct. During its course, several distinct portions may be more or less clearly recognized, but not with equal ease in all specimens. Most favorable for interpretation seem to be specimens with fully extruded, mushroom-like genital papilla. The centripetal portion of the vas represents the vesicula seminalis, which is coiled up into a more or less compact mass and which, when filled with spermatozoa, may present quite a large lumen. Next. centrifugally, comes the pars muscularis: this is provided with distinct muscular fibers, but is not developed to such an extent as seen in certain other amphistomes, for instance in Paramphistomum calicophorum. The duct then becomes rather suddenly enlarged, is surrounded by a less prominent layer of muscular tissue, is provided with a considerable number of cells resembling those found in the pars prostatica of other amphistomes, but much less striking in their development and arrangement, so that were it not for analogy, one would probably hesitate to interpret them as prostatic cells; the lumen of this portion may contain large masses of spermatozoa, so that in general effect the structure reminds the observer of the vesicula seminalis interna described for certain species possessing a cirrus pouch. As this portion narrows centrifugally, the prostatic-like cells at first seem to increase, then to decrease in number, until they finally disappear. Then follows a portion of the male canal, of narrower diameter, but still remaining larger than the corresponding portion of the female duct, and apparently devoid of both muscles and prostatic cells; this apparently represents the ductus ejaculatorius and opens into such a short ductus hermaphroditicus that the latter almost represents only a pore common to the male and the female canals,

Female organs: Caudad of the testicles, the interspace between the intestinal ceca contains coils of the uterus, the ovary, and the shell gland. The ovary is a somewhat globular body, very much smaller than either testicle: it is placed a little to the left of the median line, near the end of one of the intestinal ceca, and somewhat above (cephalad of) the anterior margin of the acetabulum. From its dorsoinferior (caudal) aspect springs the oviduct which curves cau-

dad to enter the shell gland; the latter is immediately beneath (caudad of) the ovary. In the shell gland the oviduct unites with the vitelloduct. Just before entering the shell gland the oviduct receives Laurer's canal, from which point the latter passes centrodorsad to open in the median line on the dorsal surface about on the transverse plane of the termination of the intestinal ceca.

From the ventral aspect of the shell gland and appearing like a continuation of a spindle shaped dilatation (ootyp) of the canal formed by the vitello- and oviduct there emerges the uterus which at once forms numerous coils. In its course cephalad the uterus passes dorsad of the testicles; a loop of it is tucked into the narrow interspace between the two testicles. Cephalad of the anterior testicle it winds its way ventrad to the genital pore, passing caudad of the arch formed by the union of the two vasa efferentia, and remaining in close relation to the ventral aspect of the male canal.

In its course the uterus contains numerous eggs and near its point of emergence from the shell gland some of the eggs are intermingled

with a mass of spermatozoa.

The vitellogene glands lie in the lateral areas, beginning at the level of the esophageal fork and extending to a plane slightly caudad of the ovary. These vitellaria are composed of scattered irregularly globular follicles confined to the inner half of these areas. Somewhat above (cephalad of) the level of the ovary the main right and left vitelloducts originate and pass mediad and slightly caudad in front (ventrally) of the intestinal tubes, to unite and form a common duct which curves around the caudal aspect of the shell gland, which it pierces to unite with the oviduct.

Genital pore.—The genital pore is situated at a point about midway from the oral margin to the esophageal fork, namely, about one-ninth to one-seventh of the body length from the oral margin. It is at the summit of a very peculiar genital papilla, which is easily subject to misinterpretation because of its different appearance in different specimens, due to the greater or less condition of extrusion or retraction. Sections with fully extruded papilla (figs. 31 and 37) form the best basis for study. When fully extruded, this papilla resembles somewhat the human penis with its glans. There is a distinctly swollen terminal portion 0.4 mm. in diameter, and 0.2 mm. long; this is followed centripetally by a constricted peduncle about 0.25 mm. in diameter and 0.15 mm. in length; its surface is smooth. At its vertex is situated the genital pore (opening of the ductus hermaphroditicus) from which the genital canals lead centripetally in quite a direct course. The parenchyma of this papilla is composed chiefly of very striking, large cells, provided with disproportionately small nuclei.

When the bulbous portion of this papilla is retracted (fig. 41) the large parenchymatic cells assume a rather deep position and might easily be interpreted as glandular cells; further, the retracting muscles arrange themselves in such a way as to give to the structure a remarkable resemblance to a partially formed cirrus pouch such as is described by Looss (1896b, p. 28, fig. 14) for *Gastrodiscus ægyptiacus*.

EGG.—The uterine egg (fig. 44) is oval in form and white in color by reflected light under a low power. With a slightly higher magnification by transmitted light it is seen to contain a dark, granular mass, with a circular area, somewhat eccentrically placed; this appears almost free of these dark granules; with still higher magnification it is noted that the eggshell is rather thin in the equatorial zone, but slightly thicker at the poles. At one pole the shell is provided with a small operculum; at the other pole, which at first appears to be somewhat less blunt, but which in reality is slightly more so, there is a small knob. The contents of the egg are inclosed in a membrane which with a high magnification does not appear to be in direct contact with the shell. The average measurements of 27 eggs were $132.2~\mu$ in length, by $74.5~\mu$ in width; the extremes were $125~\mu$ and $130~\mu$ in length and $67.5~\mu$ and $86~\mu$ in breadth.

EXCRETORY SYSTEM.—The excretory vesicle is situated dorsad of the acetabulum in the caudal rounded portion of the body. A little caudad of the cecal ends of the intestinal tubes it receives two large lateral excretory canals, which can be traced some distance cephalad. From the vesicle a canal passes caudodorsad to open in the median line of the dorsum about at the level of the center of the acetabulum. The canal is lined by a cuticle in anatomical continuation with that of the surface.

AGAMODISTOMUM NANUS new species.

(Figs. 45 to 66.)

Specific diagnosis.—Agamodistomum: Minute distomes 0.35 to 0.412 mm. long, 0.246 to 0.27 mm. broad, 0.147 mm. thick; oval, venter flat, dorsum convex, cephalic and caudal margins bluntly rounded, lateral margins convex. Apparently without spines. Oral sucker subterminal, 32 μ . Acetabulum slightly larger, 41 to 46 μ , in equatorial plane of body. Genital pore median, about midway between acetabulum and caudal margin (compare Clinostomum). Pharynx apparently present; postpharyngeal esophagus very short or wanting; bifurcation 92 μ from center of mouth; ceca fusiform extending beyond genital pore, may extend nearly to caudal margin. Primordium of cirrus pouch (?) large and prominent, extending nearly straight dorsad, between ceca; one genital primordium dorsocaudad of this, another caudad, near caudal margin. Excretory pore dorsosubterminal; primordium of excretory vesicle present; at least two excretory canals present.

Habitat.—Encysted in pectoral muscle of African partridge (*Francolinus subtorquatus*) at Benguella, West Africa.

Type.—U. S. P. H. & M. H. S., No. 9834, two worms mounted in toto on one slide; cotypes in sections. Alcohol material in U. S. National Museum and in collection of American Society of Tropical Medicine.

Source of Material.—A bottle of material collected by Doctor Wellman in Benguella, West Africa, and sent to us for determination, contained portions of the "pectoral muscles of a partridge, Francolinus subtorquatus." The muscles are beset with a number of small cysts, about 1.0 mm. long, which bear a superficial resemblance to the sarcosporidia occasionally found in birds. Upon careful teasing, as well as upon section, the cysts are seen to contain minute agamic distomes, apparently one parasite in each cyst.

Significance of infection from standpoint of food inspection.—As will be seen from the following account of the anatomy, this parasite does not correspond to any species thus far known to occur in man. Whether it would develop in man further if taken in the food can not be stated at present. From the standpoint of food inspection the infection would for the present be similar to a severe infection with sarcosporidia, namely, an infection not dangerous to man, but one which so altered the condition of the meat as to lead to its condemnation.

At the present moment this infection is not known for partridges in the United States.

EXTERNAL CHARACTERS.

MEASUREMENTS.—The parasites vary in size, attaining 0.352 to 0.416 mm. in length by 0.246 to 0.272 mm. in transverse diameter, and 0.147 mm. in dorsoventral diameter.

Color.—The unstained alcoholic specimens are of a yellowish gray color, with two longitudinal lighter fusiform spots, due to the intestinal ceca. In transmitted light the worm has the appearance of a small amount of foam, this foamy appearance being due to the cellular structure.

FORM.—The worms are oval in outline, with flat venter and convex dorsum; the oral (cephalic) and aboral (caudal) margins are bluntly rounded and of nearly the same form; the lateral longitudinal margins are convex.

Surface.—No spines could be distinguished on the cuticle.

Oral sucker.—The oral sucker is ventrosubterminal, 32.2 μ in diameter, and directed from dorsocaudal to ventrocephalad.

VENTRAL ACETABULUM.—The ventral acetabulum may be exactly in the equatorial plane, or slightly caudad of equator; it is 41 to 46 μ in diameter, with a circular aperture of 13.8 μ ; on section it is 38.8 μ in dorsoventral diameter.

Genital Pore.—Halfway between the acetabulum and the caudal 24800—Bull. 40—08—3

margin, or slightly nearer the acetabulum, is situated the genital pore, which is rather prominent and is surrounded by darkly staining tissue.

EXCRETORY PORE.—The excretory pore is dorsoterminal.

INTERNAL ANATOMY.

DIGESTIVE TRACT.—The digestive tract is the most prominently developed organ system in this stage of development. Through its entire extent its lumen is entirely occupied by a gelatinous or (?) granular mass, in which cellular structure is not distinguishable.

Extending dorsocaudad from the oral sucker the esophagus may be distinguished; at the point where the esophagus leaves this sucker there is a bulbous structure, the wall of which is composed of a single row of nucleated cells; the impression gained is that this is a pharvnx in process of development. The bifurcation (into two intestinal ceca) occurs about 56 μ caudad of the center of the oral sucker, so that the postpharyngeal portion of the esophagus, though distinct, is exceedingly short. In their course caudad the intestinal ceca diverge from the median line and at first, for a distance of about 92 u, increase gradually in diameter; then they increase rapidly in diameter, but upon nearing the region of the genital pore they again decrease, each ending in a rather sharp point somewhat caudad of the pore; in some cases they extend to a plane about halfway between the genital pore and the caudal end of the body. In transverse section the ceca are nearly circular, but the dorsoventral diameter is slightly greater than the transverse diameter (88 to 76 u in one case). Thus the ceca represent the two fusiform spots seen in the unstained specimen.

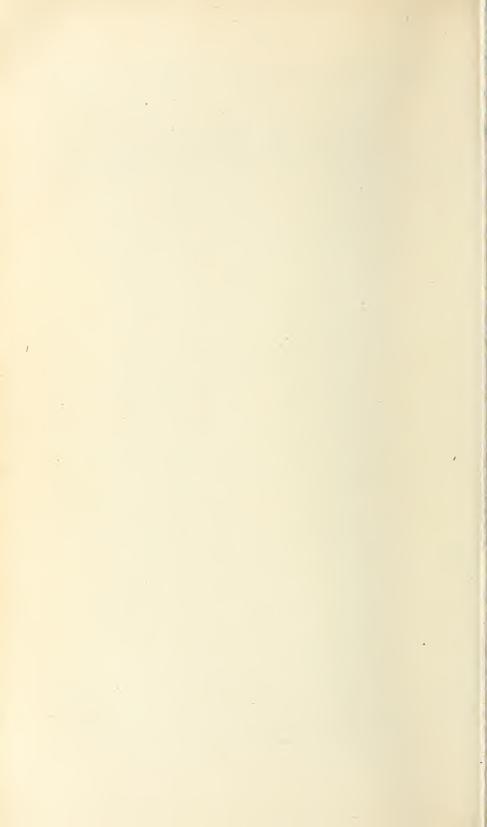
In the wall of the ceca may be seen a few very small nuclei. The ceca are somewhat nearer the dorsum than the venter.

General system.—The primordium of the genital system is present, but the arrangement of the sexual glands can not be analyzed. On section a distinct canal is seen to run almost directly dorsad, but slightly caudad, between the intestinal ceca; in one section this canal had the appearance of dividing into two canals, a male and a female duct, but this point can not be asserted without reserve. The canal is surrounded by a mass of darkly staining tissue, about 69 μ long (i. e., dorsoventrad in reference to the worm) by 46 μ broad (cephalocaudad in reference to the worm); the entire structure gives the impression of being a cirrus pouch in course of development.

Dorsocaudad of this structure is a second, much smaller, darkly staining mass of cells, and still further caudad, near the caudal margin of the worm, is still a third darkly staining mass.

EXCRETORY SYSTEM.—At the caudal end, slightly dorsad, is an invagination, surrounded by a row of cells, and apparently representing the excretory vesicle. Excretory canals are also visible on section.

Systematic Position.—From the foregoing account it will be seen that this worm is an agamic distome with genital pore about midway between the acetabulum and the caudal margin (compare for instance Clinostomum).



A REEXAMINATION OF THE ORIGINAL SPECIMEN OF TÆNIA SAGINATA ABIETINA (WEINLAND, 1858).

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Since 1858 works on zoology and on the practice of medicine make mention of a tapeworm which Weinland described as *Tænia solium* var. *abietina*. Opinions of authors have differed in regard to the exact status of the parasite in question, some writers considering that it represented a distinct species, others that it was a variety of *Tænia solium*, and still others, that it was a peculiar specimen of *Tænia saginata*.

In a recent visit to Boston one of us found the original specimen, or rather all that seems to be left of it, in the Warren Anatomical Museum, and through the courtesy of Doctor Whitney, curator of the museum, we have been able to reexamine this interesting material. In connection with a brief account of our rather unsatisfactory results it may be well to give a review of the history of the parasite in question, which now bears the U. S. P. H. & M. H. S. No. 9713, but which will be returned to the Warren Museum.

SYNONYMY.

- 1858: Tænia solium var. abietina Weinland, 1858, 43-45, 84; 1861, 4-5, 12-14, , pl. 5, figs. 18-20.
- as variety of *Tænia mediocanellata* in Leuckart, 1863, 289, fig. 80 (based upon reexamination of original material).—Diesing, 1864a, 369.
- as variety of T. saginata in Leuckart, 1880, 605-606, fig. 272.—Blanchard, 1886a, 362-363, fig. 238.
- as syn. of T. saginata in Braun, 1903, 3 ed., 224.
- as possible syn. of T. confusa in Braun, 1903, 3 ed., 229.
- as doubtful syn. of T. saginata in Stiles, 1898a, 72.
- 1864: Tania mediocanellata var. abietina (Weinland, 1858) Diesing, 1864a, 369.
- 1873: Twnia abietina (Weinland, 1858) Davaine, 1873a, 572; 1877a, l, 909, 910, 920.—Cobbold, 18—, —.—Swart, 1862, 18.
- as doubtful syn. of T. saginata in Stiles. 1898a, 72.

1880: Tænia saginata var. abietina (Weinland, 1858) Leuckart, 1880, 605-606 fig. 272.—Stiles, 1906, June, 31-32, figs. 35-38.

1885: Tænia abietina (Weinland, 1858) Guzzardi Osmundo, 1885a, 580.

HISTORICAL REVIEW: Weinland's (1858, 43-45, 84) original account of this species is as follows:

[p. 43] § 63. Under this name [Tania solium var. abietina] I will introduce a specimen of tapeworm which comes from a Chippewa Indian, at the Sault Ste. Marie, Lake Superior; it was obtained there by Professor Agassiz during his famous trip to that lake. The specimen consists of a chain [p. 44] of several feet in length, from the mature part of the worm. The head, neck, and the whole anterior half are wanting.

The most striking thing in this worm is its extreme narrowness and meagerness, while *Twnia mediocanellata*, which it resembles in the configuration of the uterus, is very broad and thick, according to Kuechenmeister. A figure of this worm, of its uterus and eggs, we intend to publish in our work on the parasites of man.

All the joints which are preserved are very thin, nearly transparent, and equally narrow, their transverse diameter being about 4 mm., and the longitudinal about 12 mm. The genital openings are very small, and without external lips; this may be owing to the very mature age of the joints in question. There is no pigment in either vagina or spermatic duct. The uterus is more regular than either in Twnia solium or in Twnia mediocanellata, yet it more resembles the latter. The middle trunk of the uterus is quite straight; the branches, about 30 in number, start from the main stem, either at right angles or at an angle of about 45°. These branches are always quite parallel, and are generally straight; but whenever they are bent, all make the same angle; they are never arborescently divided, nor furcated at the ends, with the exception of the foremost and the hindmost in each joint, which run, the former forwards, the latter backwards, both being forked and crooked. The eggs, which are extremely plenty in these joints, and which show the whole configuration of the uterus in a yellowish tint to the naked eye, are 0.033 mm. long, and 0.030 mm. broad; they are protected, first, by an outside shell (chorion) which is 0.003 mm. thick, dark in its outer layers, transparent, yellowish inwards; then follows a second shell (yolk membrane), 0.0006 mm, thick, entirely transparent. In the cavity of the egg lies the embryo, occupying about two-thirds of it, and measuring only 0.016 mm. We saw other eggs, unripe, and with one eggshell only, but very rarely.

We consider this worm merely as a variety of *Tunia solium*, and we called it varietas *abictina*, from abies, a pine tree, which the configuration of its uterus resembles.

We hope soon to get more information concerning this Indian tapeworm from our western and Canadian medical friends.

[p. 84] 2b. Tania solium, L. Varietas abietina Weinland.

Obtained by Prof. L. Agassiz from a North American (Chippewa) Indian, at Lake Superior. The specimen is preserved in the Zoological Museum. Cambridge, Mass.

Later, Weinland (1861, 4–5, 12–14, pl. 5, figs. 18–20) reverts to this subject; his account of the parasite is practically a translation of the paragraph quoted above, but he adds that the worm was collected in 1850; the eggs measure 33 by 30 to 33 μ ; the chorion is dark brown externally, lighter internally. Weinland was unable to recognize any further details.

So far as we have been able to find, Leuckart (1863, 289, fig. 80) is the only other helminthologist who has examined this parasite; he reexamined part of Weinland's original material and concluded that it represented a variety of *Tænia mediocanellata*; in his later work (1880, 605-606, fig. 272) he refers to it as *Tænia saginata* var. abietina. Not altogether in harmony with Weinland's description, Leuckart's figure of the uterus shows a number of the lateral pouches of the uterus as branched; he gives about 40 branches each side.

All later references to the parasite are either based directly upon the conclusions drawn by Weinland and by Leuckart, or are opinions

expressed on basis of the data given by these two authors.

Diesing (1864a, 369) considers abietina a variety of *T. saginata*. Blanchard (1886a, 362–363, fig. 238) considers it as "a simple variety of *T. saginata*."

Braun (1903, 3 ed., 224) thinks it a "*T. saginata*, with uterine branches somewhat more thickly set;" he also suggests (p. 229) that it may possibly be identical with *T. confusa* Ward.

Stiles (1906, June, 31-32, figs. 35-38) recognized the form provisionally as a subspecies of *Tænia saginata*. He republished Weinland's and Leuckart's illustrations of the worm.

REEXAMINATION OF ORIGINAL MATERIAL.—Segments: The specimen consists of only 5 gravid segments, which are in very poor state of preservation. At the time it came into our hands it was completely dried out and, naturally, brittle. It was soaked in very weak alcohol, which was gradually increased in strength; then it was transferred to alcohol glycerin, the alcohol was allowed to evaporate, and the worm eventually mounted in glycerin jelly. Because of its having been dried a detailed study of the anatomy is of course excluded.

The segments measure as follows:

About 12 mm. long by about 2 mm. broad.

About 12 mm. long by about 2 to 3 mm. broad.

About 12 mm. long by about 2 to 2.5 mm. broad.

About 11 mm. long by about 2.5 mm. broad.

About 11 mm. long by about 2.5 to 3.5 mm. broad.

Genital pores.—The genital pores are irregularly alternate and are caudad of the equator of the segment—more exactly, about five-eighths from the anterior margin.

Uterus.—No remnants of the genital glands can be seen, and it is scarcely possible to count the lateral branches of the uterus; these are, however, numerous.

Eggs.—The uterus is crowded with eggs, which vary in size from 31 to 37.5 by 30 to 33.9 μ . An average of 31 embryophores gave 36.27 by 32.09 μ ; the usual length (21 cases) was 37.5, the most frequent breadth (11 cases) 31.9 μ .

We have obtained or compiled the following measurements for the embryophores of the tænias of man:

T. solium, almost round, 31 to 36 μ .

T. saginata, ovoid, 35 to 40 by 20 to 30 μ .

T. confusa, 39 by 30 μ .

T. africana, 31 to 39 by 33.8 μ .

T. abietina, 31 to 37.5 by 30 to 33.9 μ .

From these measurements it will be seen that the eggs of *T. abietina* do not agree exactly with any of the species mentioned.

Calcareous corpuscles.—The calcareous corpuscles are exceedingly numerous, round to oval in shape, varying considerably in size, and attaining 17.6 μ in diameter. Comparing these with the measurements given for other forms, we have the following table:

Tænia solium, up to 12μ , sparse.

T. saginata, up to 18 μ , plentiful.

T. confusa, up to 11 μ , sparse.

T. africana, 10.4 by 16.9 μ , sparse.

T. hominis, very numerous.

T. abictina, up to 17.6 μ , very numerous.

The data given above are scarcely sufficient to enable one to express a positive opinion upon the systematic value of T. abietina. They seem, however, to be too meager to fully justify its separation from T. saginata, and almost too much to fully justify viewing it as a typical T. saginata.

Were these fragments sent to us for determination, without knowledge of their origin, we should conclude that they were probably a dwarfed specimen of *Tænia saginata*.

In order to obtain opinions from other persons, we have submitted the fragments to Hassall, Ransom, and Ward, requesting them to come to a conclusion before they inquired into the history of the material. Ward was disinclined to consider the segments as belonging to *T. saginata*, while Hassall and Ransom were both inclined to consider them as belonging to this species.

Under existing circumstances no exact determination can be accepted as final, but four of the five helminthologists who have reexamined the material incline to the *saginata* determination, while one is disinclined.

That the material does not agree exactly with the typical *Tænia* saginata, as we find it in this country, must be admitted, but we do not feel justified at present in recognizing it as of full specific rank.

Under existing circumstances perhaps the best solution of the matter is to accept it as a doubtful subspecies, *Tænia saginata abietina* and call attention to the worm in the hope that some new specimens may be collected which will permit of a definite opinion. We must look to the physicians in the North and West, particularly to those near Sault Ste. Marie, to find such material.



Fig. 1.



F1G. 2.



Fig. 3.



F1G. 4.

Figs. 1-4.—Four photographs of Gates's patient in Florida, showing acne-like condition and enlarged breasts, due to infection with *Sparganum proliferum*. Original; photos kindly furnished by Doctor Gates.





Fig. 5.—Sparganum proliferum, in part in a cyst. Original. $\times 10$.





Fig. 6.—Sparganum proliferum, escaped from the cyst. Original. $\times 10$.





Fig. 7.

Figs. 7-15.—Nine specimens of $Sparganum\ proliferum$, showing various forms, buds, and supernumerary heads. Original. $\times 10$.





LH-MILDER DEL

Fig. 8.



L'H WILDER DEL.

FIG. 9.



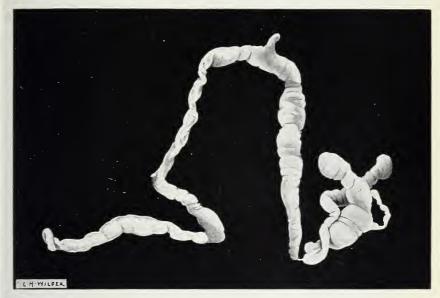


FIG. 10.



FIG. 11.





FIG. 12.

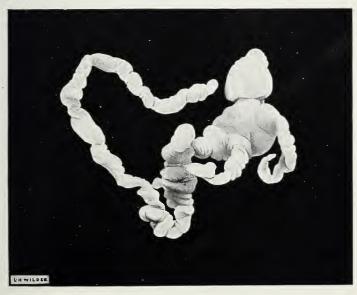


Fig. 13.





Fig. 14.



Fig. 15.



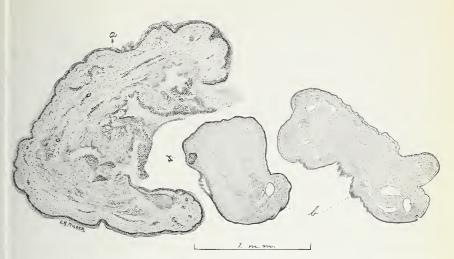


Fig. 16.—Section through a cyst (a), with the escaped $Sparganum\ proliferum\ (b);\ x,$ reserve food particle. Original,

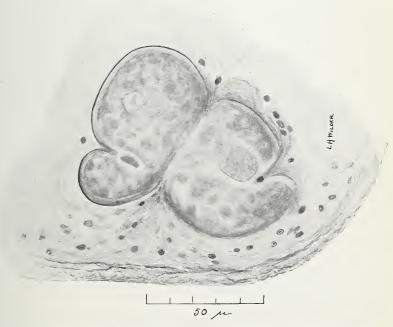


Fig. 17.—Section through a reserve food particle. (See x, fig. 16.) Enlarged. Original.





Fig. 18.—Section showing large excretory canal, smaller canals, calcareous corpuscles, and a pore. Enlarged. Original.





FIG. 21.

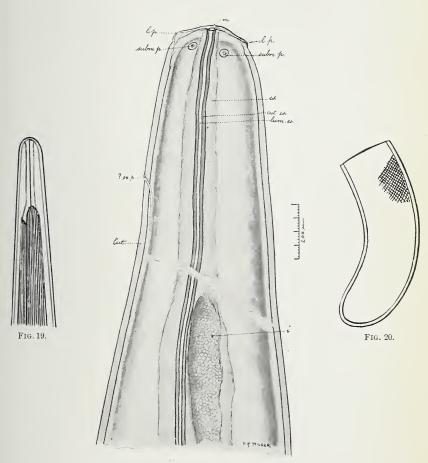


FIG. 22.

- Fig. 19.—Head of Filaria restiformis. Enlarged. Leidy's original figure.
- Fig. 20.—Tail of F. restiformis. Enlarged. Leidy's original figure.
- Fig. 21.—Figure of type specimen of F. restiformis. Natural size. Original.
- FIG. 22.—Enlarged view of head of same; the body is surrounded by a distinct cuticle (cut.); cuticle of esophagus (cut. es.); esophagus (cs.); blind intestinal sac (i.); lumen of esophagus (lum. es.); a pair of lateral papillæ (l. p.) and two submedian papillæ (sulm. p.) are visible; the mouth is terminal (m.). Original.



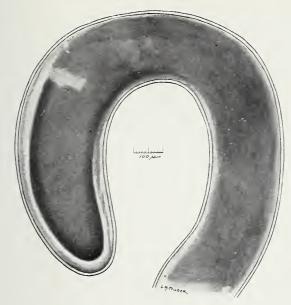


FIG. 23.

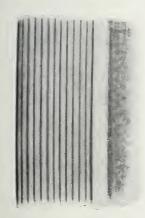


FIG. 24.



FIG. 25.

Fig. 23.—Tail of same; the dark body in the center represents the intestine. Enlarged. Original. Fig. 24.—Optical section of cuticle showing layers. Greatly enlarged. Original. Fig. 25.—Diagonal system of fibers of cuticle. Greatly enlarged. Original.



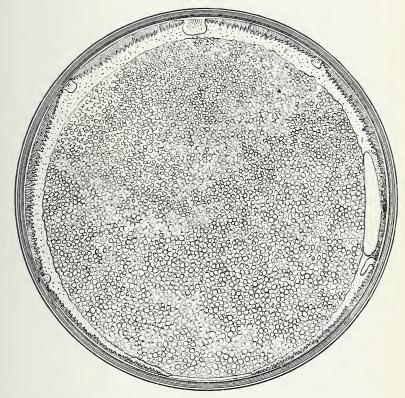
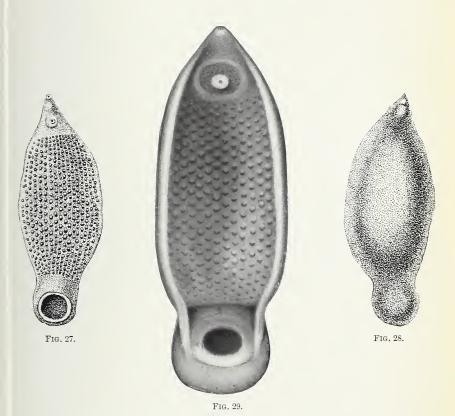


Fig. 26.—Transverse section of type specimen, showing 6 muscular fields, separated by 6 longitudinal lines; the central mass is the fat body. Greatly enlarged. Original.





Figs. 27-28.—Homalogaster paloniæ. (After Poirier 1883, figs. 1a-b.) Fig. 29.—Homalogaster philippinensis, ventral view. Enlarged. Original.



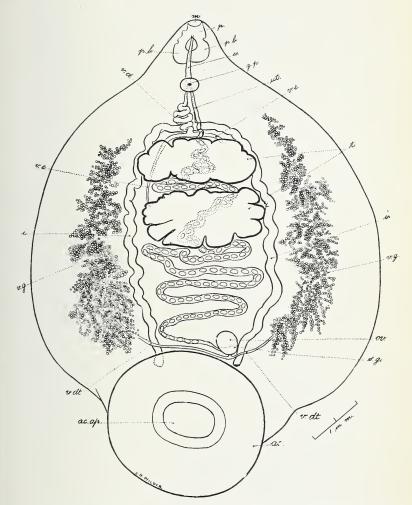


FIG. 30.—Press preparation of *Homalogaster philippinensis*, showing internal anatomy: ac., acetabulum; ac. ap., aperture of acetabulum; es., esophagus; g. p., genital pore; î., intestine; m., mouth; ov., ovary; p., pharnax; ph. b., pharyngeal pouch; s. g., shell gland; t., testicle; ut., uterus; v. d., vas deferens, v. e., vas efferens; v. g., vitellogene gland; v. dt., transverse vitello-duct. Enlarged. Original.



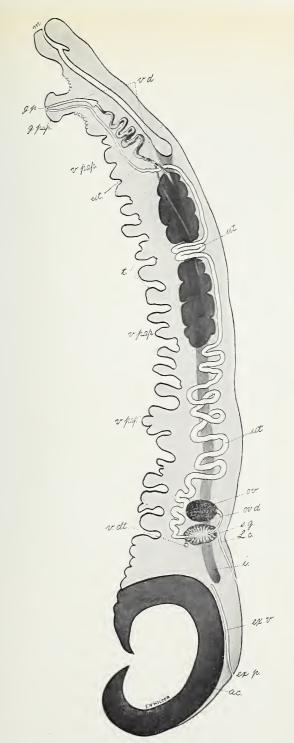


Fig. 31.—Optical, sagittal section showing internal anatomy: ϵx , p., excretory pore; ϵx , v., excretory vesicle; g. pap., genital papilla; L. c., Laurer's canal; ov. d.. oviduet; v. p., ventral papillæ. Remainder of lettering as in figure 30. Semidiagrammatic. Enlarged. Original.



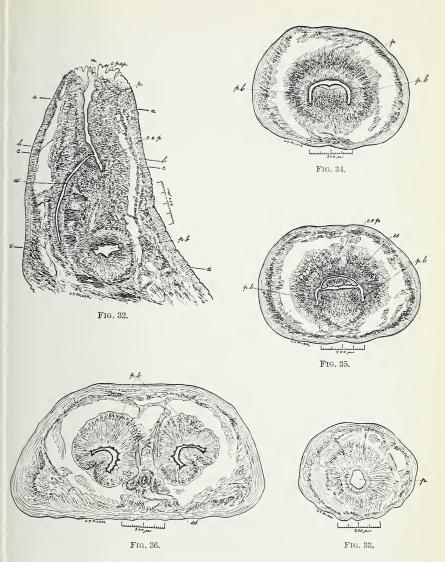


Fig. 32.—Sagittal section through oral extremity: $c.\ e.p.$, cecal extremity of pharynx; es., esophagus; m., mouth; $o.\ pap.$, oral papillæ; p., lumen of pharynx: $p.\ b.$, lumen of pharyngeal pouch: $a-a,\ b-b,\ c-c,\ d-d$, planes of section. Enlarged. Original.

Fig. 33.—Transverse section through plane a-a of figure 32, showing pharyngeal lumen (p.). Enlarged. Original.

Fig. $\overline{34}$.—Transverse section through plane b-b of figure 32, showing pharyngeal lumen (p.) and aperture of pharyngeal pouches (p. b.). Enlarged. Original.

Fig. 35.—Transverse section through plane c-c of figure 32, showing cecal extremity of pharynx $(c.\ e.\ p.)$; beginning of esophagus (es.) and aperture of pharyngeal pouches $(p.\ b.)$. Enlarged, Original.

FIG. 36.—Transverse section through plane d-d of figure 32, showing relative positions of esophagus (es.) and pharyngeal pouches (p, b.). Enlarged. Original.



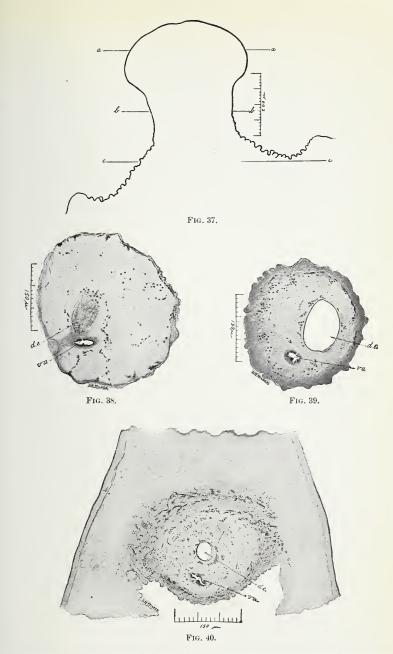


Fig. 37.—Outline of genital papilla in sagittal section, showing also the papillæ around the base; a-a, b-b, c-c, planes of section. Enlarged. Original.

Fig. 38.—Section through genital papilla in plane a-a, of figure 37. The section is not quite at right angles to the axis of this papilla. It shows the metraterm (va.) and the ductus ejaculatorius (d.e.) filled with spermatozoa. Enlarged. Original.

Fig. 39.—Section through genital papilla in plane b-b of figure 37. Lettering as figure in 38. Enlarged. Original,

Fig. 40.—Section through genital papilla in plane c-c of figure 37. Lettering as in figûre 38. Enlarged. Original.



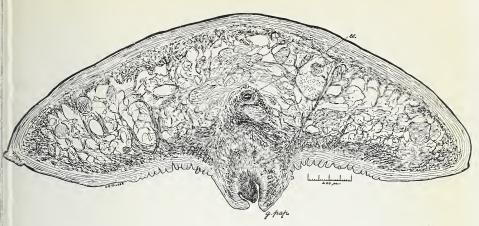


Fig. 41.—Transverse section showing retracted genital papilla (y. pap.), esophagus (es.) and the papillæ around base of genital papilla. Enlarged. Original.

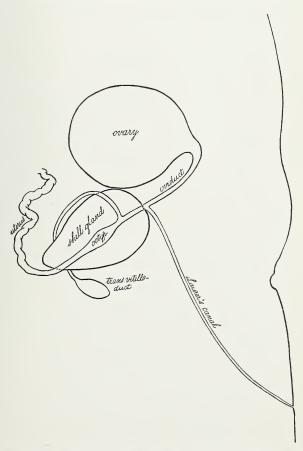
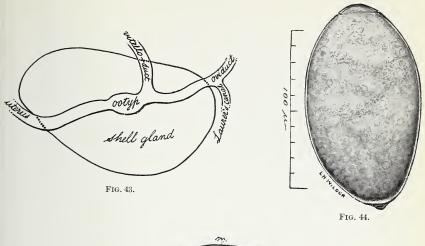


Fig. 42.—Diagram showing relation of ovary, shell gland, oviduct, ootyp, uterus, and Laurer's canal in sagittal plane. Original.





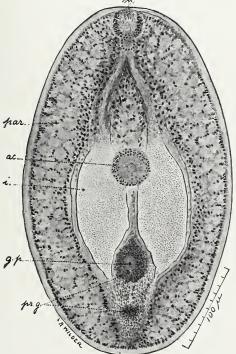


Fig. 45.

Fig. 43.—Diagram showing shell gland, ootyp, vitelloduct, uterus and Laurer's canal. Original.

Fig. 44.—Egg. Enlarged. Original.
Fig. 45.—Ventral view of Agamodistomum nanus; ac., acetabulum; g. p., genital pore; i, fusiform intestine; m., oral sucker; par., foamy parenchyma; pr. g., primordium of genital glands.



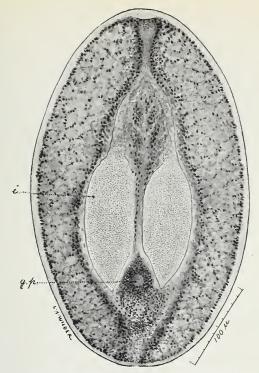


Fig. 46.—Dorsal view of Agamodistomum nanus. Lettering as in fig. 45.



Fig. 47.

Figs. 47-53.—Series of 7 consecutive sagittal sections of cyst with contained parasite, showing anatomy of parasite from its right lateral margin to a plane on the left of the median line. Note the foamlike structure of the parenchyma of the worm, the fusiform intestine (i., 50, 51, 53), the pharynx (p., 51, 52), the acetabulum (ac., 52), ventral genital pore (g. p., 52), genital primordium (pr. g., 52), and the dorsosubterminal exerctory pore (x. p., 52).



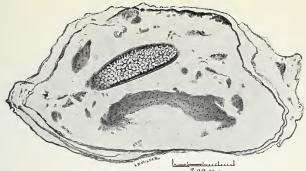
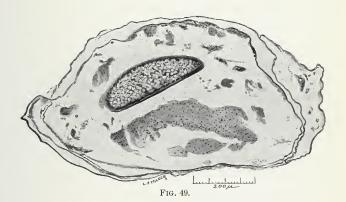


Fig. 48.



Landard Landard 200 for

Fig. 50.



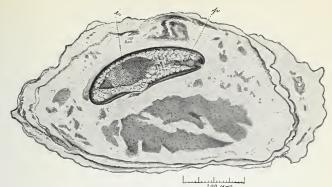


Fig. 51.

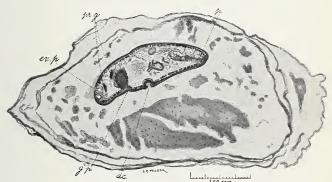


Fig. 52.

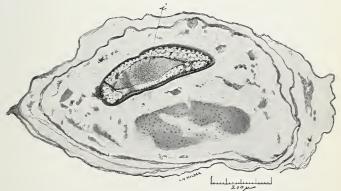
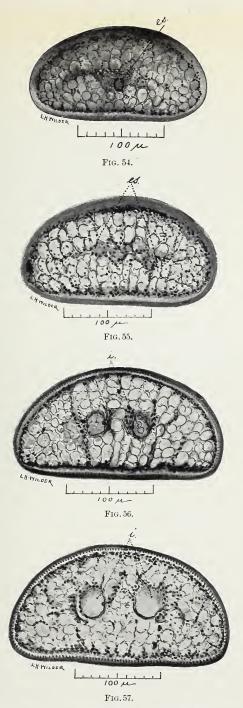


Fig. 53.





Figs. 54-66.—Series of 13 consecutive transverse sections of $Agamodistomum\ nanus$, showing the anatomy. Note the esophagus (es., 54), its bifurcation (es., 55), the intestinal ceca $(i., 56\ to\ 65)$, the ventral acetabulum $(ac., 60\ to\ 61)$, the genital primordium $(pr.\ g.)$ and pore $(g.\ p., 63\ to\ 65)$, the excretory canals $(ex.\ c., especially\ in\ 65)$.



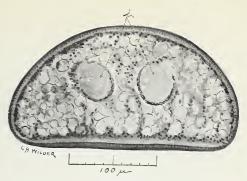
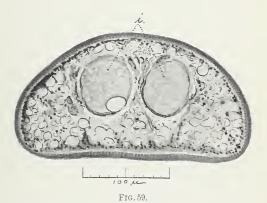


FIG. 58.



LH WILDER ac.

FIG. 60.



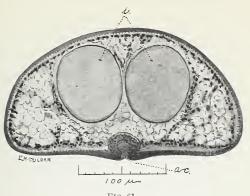


FIG. 61.

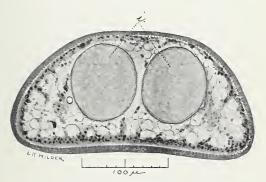


Fig. 62.

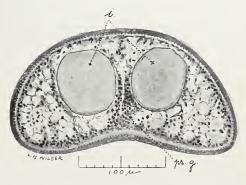


FIG. 63.



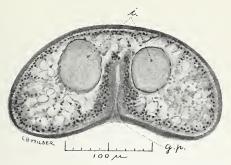
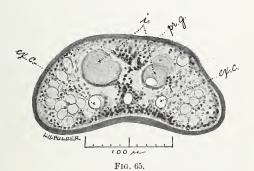


FIG. 64.



THE PRINCES TO SHARE THE PRINCES OF

Fig. 66.



INDEX TO ZOOLOGICAL NAMES.

| | Page. | | Page. |
|-----------------------------|---------------|-------------------------------------|----------|
| abietina (Tænia) | 6, 35, 36, 38 | gigas (Strongylus) | 19 |
| egyptiacus (Gastrodiscus) | 30 | Gordiidæ | 21 |
| africana (Tænia) | 38 | Gordius | 20 |
| Agamodistomum | 15, 30, 31 | Gyporphynchus | 16 |
| nanus | 6, 23, 30 | Homalogaster 23. | , 24, 25 |
| Agamofilaria | 15 | paloniæ 23 | , 24, 25 |
| Agamomermis | | philippinensis _ 6, 23 | |
| restiformis | | poirieri | 24 |
| Amphistomulum | 15 | hominis (Tænia) | 38 |
| bailleti (Cysticercus) | | Lacerta | 15 |
| (Plerocercoides) | | lacertæ (Dithyridium) | 15 |
| Bos | 6 | muralis (Dithyridium) | 15 |
| taurus | 25 | viridis (Dithyridium) | 15 |
| taurus indicus | | Ligula mansoni | 14 |
| calicophorum (Paramphistomu | m) = 28 | Ligulinæ | 18 |
| canis (Trichodectes) | | macropeos (Tænia) | 16 |
| Cladorchinæ | | mansoni (Ligula) | 14 |
| Clinostomum | | (Sparganum) | 14 |
| confusa (Tænia) | 35, 37, 38 | medinensis (Dracunculus) | 21 |
| crispa (Piestocystis) | | mediocanellata (Tania) 35 | , 36, 37 |
| Cyathocephalus | | var. abietina (Tænia). | 35 |
| Cysticercus | | Mermis | 20 |
| bailleti | | Mermithidæ 20 | , 21, 22 |
| elongatus | 15 | nanus (Agamodistomum) 6 | , 23, 30 |
| Dibothriocephalidæ | 16, 18 | Palonia frontalis | 24 |
| Dibothriocephalus | 18 | paloniæ (Homalogaster) 23 | , 24, 25 |
| dithyridium (Piestocystis) | | Paramphistomidæ | 23 |
| Dithyridium | | Paramphistomum calicophorum | 28 |
| elongatum | | philippinensis (Homalogaster). 6,23 | , 24, 25 |
| lacertæ | 15 | Piestocystis | 15 |
| (lacert* muralis) | 15 | crispa | 15 |
| (lacertæ viridis). | 15 | dithyridium | 15 |
| Dracunculus | 21 | rugosa | 15 |
| medinensis | 21 | variabilis | 15 |
| Echinococeus | 15 | Plerocercoides | , 15, 16 |
| elongatum (Dithyridium) | 16 | bailleti | 16 |
| elongatus (Cysticerus) | 15 | prolifer | 14 |
| Filaria | 20 | Plerocercus | |
| restiformis | 6, 19, 20, 22 | prolifer | 14, 16 |
| Filariidæ | 22 | poirieri (Homalogaster) | 24 |
| Francolinus subtorquatus | 6, 30 | prolifer (Plerocercoides) | 14 |
| frontalis (Palonia) | 24 | (Plerocercus) | 14, 16 |
| Gastrodiscus ægyptiacus | 30 | (Sparganum) (Gatesius)) | 18 |
| Gatesius | 18 | proliferum (Sparganum) | 6, 7, 16 |

| Page. | Page, |
|----------------------------------|----------------------------------|
| restiformis (Agamomermis) 19, 22 | Tænia hominis |
| (Filaria) 6, 19, 20, 22 | macropeos |
| rugosa (Piestocystis) | mediocanellata 35, 36, 37 |
| saginata (Tænia) | mediocanellata var. abietina 35 |
| abietina (Tænia) 35, 36, 37, 38 | saginata 6, 35, 37, 38 |
| solium (Tænia) 35, 36, 38 | saginata abietina 35, 36, 37, 38 |
| var. abietina (Tænia) 35, 36 | solium 35, 36, 38 |
| Sparganum | solium var. abietina 35, 36 |
| (Gatesius) proliferum 18 | unilateralis |
| mansoni 14 | taurus (Bos) |
| • proliferum 6, 7, 16 | indicus (Bos) |
| Strongylus gigas | Tetrarhynchus 15 |
| subtorquatus (Francolinus) 6, 30 | Trichodectes canis |
| Tænia abietina 6, 35, 36, 38 | unilateralis (Tænia) |
| africana 38 | variabilis (Piestocystis) |
| confusa | |

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The following bulletins [Bulls. Nos. 1-7, 1900 to 1902, Hyg. Lab., U. S. Mar.-Hosp. Serv., Wash.] have been issued.

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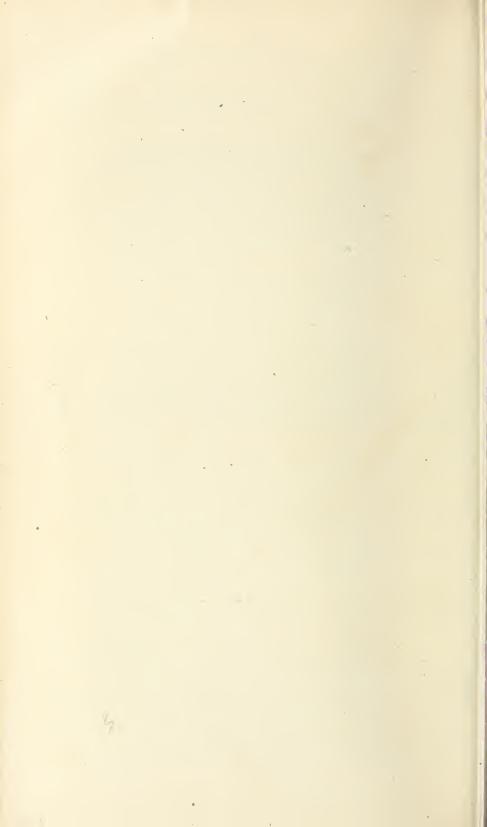
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